Title: Comparing diversification rates in lakes, rivers, and the sea

Short title: Diversification in aquatic habitats

Elizabeth Christina Miller[1]

[1]School of Aquatic and Fishery Sciences, University of Washington, Seattle, Washington 98105, U.S.A.

Correspondence to: emille3@uw.edu

Author contributions: E.C.M. conceived the study, collected data, performed analyses, interpreted the results, and wrote the manuscript.

Acknowledgements: I thank John Wiens and Luke Tornabene for helpful discussion related to this manuscript. I was supported by an NSF Postdoctoral Fellowship (DBI-1906574).

Data Accessibility Statement: All data (including habitats for each species) and R code needed to replicate analyses are in the Dryad package associated with this manuscript. The temporary link to the repository during peer review is: XXXXXXX.

TITLE:

Comparing diversification rates in lakes, rivers, and the sea

ABSTRACT:

The diversity of species inhabiting freshwater relative to marine habitats is striking, given that

freshwater habitats encompass <1% of Earth's water. The most commonly proposed explanation

for this pattern is that freshwater habitats are more fragmented than marine habitats, allowing

more opportunities for allopatric speciation and thus increased diversification rates in freshwater.

However, speciation may be generally faster in sympatry than in allopatry, as illustrated by

lacustrine radiations such as African cichlids. Differences between rivers and lakes may be

important to consider when comparing diversification broadly among freshwater and marine

lineages. Here I compared diversification rates of teleost fishes in marine, riverine and lacustrine

habitats. I found that lakes had faster speciation and net diversification rates than other aquatic

habitats. However, most freshwater diversity arose in rivers. Surprisingly, riverine and marine

habitats had similar rates of net diversification. Biogeographic models suggest that lacustrine

habitats are evolutionary unstable, explaining the dearth of lacustrine species in spite of their

rapid diversification. Collectively, these results suggest that diversification rate differences are

unlikely to explain the freshwater paradox. Instead, this pattern may be attributable to the

comparable amount of time spent in riverine and marine habitats over the 200-million-year

history of teleosts.

KEY WORDS:

Freshwater paradox, rivers, lakes, marine fishes, freshwater fishes, diversification rates

INTRODUCTION:

The great biodiversity of freshwater habitats is striking. Habitable freshwater makes up 0.009% of the Earth's water volume but contain at least 40% of ray-finned fish species (Cohen 1970; Horn 1972), a pattern called the "freshwater paradox" (Tedesco et al. 2017a). The diversity of freshwater relative to marine fishes is thought to be due to the greater fragmentation and isolation of freshwater habitats, creating more opportunities for geographic population subdivision than in the ocean (Cohen 1970; Horn 1972; May 1994; Grosberg et al. 2012). For this reason, speciation and net diversification rates are expected to be higher in freshwater than in marine lineages (e.g., Tedesco et al. 2017a).

Two innovations have allowed researchers to rigorously test longstanding ideas about the origin of biodiversity: the expansion in size and taxonomic scope of molecular phylogenies (Rabosky et al. 2013; Rabosky et al. 2018), and the development of increasingly sophisticated approaches to estimate diversification rates (Maddison et al. 2007; Rabosky 2014). The hypothesis that freshwater fishes have faster diversification rates than marine fishes has been tested in at least 10 studies over the past decade, with mixed results (Table 1). Most studies focusing on a single clade of fishes spanning both marine and freshwater biomes have found little difference in net diversification rates (speciation minus extinction rates; see references in Table 1). This could perhaps be due to low power to detect diversification rate differences within small clades (Davis et al. 2013), or because speciation and extinction rates are both higher in freshwater leading to small differences in net diversification (Bloom et al. 2013). Note that higher turnover in freshwater compared to marine habitats cannot explain the freshwater paradox, because species richness will not change when speciation and extinction rates are about equal (see also Meseguer and Condamine 2020). Studies including all ray-finned fishes have

found no difference in diversification rates (Carrete Vega and Wiens 2012), faster rates in marine

habitats (Betancur-R et al. 2015), faster rates in freshwater (Tedesco et al. 2017a), or faster rates

in only some freshwater clades (Rabosky 2020). The only study to my knowledge that

exclusively used the fossil record to test this hypothesis found that the accumulation of family

diversity has slowed over time in marine but not freshwater fishes (Guinot and Cavin 2015).

However, familial diversity is lower overall in freshwater, so it is unknown if freshwater

diversity will also plateau if it ever reaches a similar magnitude as the ocean (Figure 1 of Guinot

and Cavin 2015). Studies of organisms other than ray-finned fishes are also collectively unclear

as to whether marine, freshwater or terrestrial habitats have the fastest diversification rates

(Table 1). In sum, whether freshwater or marine fishes have faster diversification rates could be

influenced by choice of methods, focal clade, or the breadth of clades included in the study. This

sensitivity could exist because the difference in rates between biomes is actually narrower than

expected (and therefore hard to detect), or because the habitat with the faster rate differs among

clades.

Both 'freshwater' and 'marine' are broad categories that encompass diverse habitats and

lineages. Diversification rates are known to vary among marine fishes: for example,

diversification is faster on coral reefs than in other marine habitats (Santini et al. 2013; Tedesco

et al. 2017a). I am not aware of similar studies in fishes comparing diversification rates among

different freshwater habitats. An examination of diversification-rate variation among freshwater

fish clades (Rabosky et al. 2013; Seehausen and Wagner 2014) shows that some species-rich

clades do not have fast rates (e.g. cyprinids), while some groups with fast rates are not species-

rich (e.g. sticklebacks). To what extent can these patterns be explained by differences in habitat?

The most commonly proposed mechanism for faster freshwater diversification rates, that

freshwater habitats give more opportunities for population subdivision than marine habitats,

implies that allopatric speciation has generated much of the world's freshwater diversity (Lynch

1989). This may be the case, as major river systems such as the Amazon are among the most

diverse freshwater habitats (Lévêque et al. 2008; Albert et al. 2020; Miller and Román-Palacios

2021). The fragmentation of rivers has been positively related to endemism and cladogenesis in

fishes (Dias et al. 2013). However, lakes are also known for their diverse faunas (Lévêque et al.

2008). Ancient lakes such as the African rift lakes and Lake Baikal contain species flocks, or

monophyletic groups that underwent rapid speciation within a single lake (Cohen 1995;

Cristescu et al. 2010; Wagner et al. 2012; Seehausen 2015). While allopatric speciation is also

common within and among lakes (Parenti 1984; Seehausen and van Alphen 1999; Rico and

Turner 2002; Hrbek et al. 2002; Martin et al. 2015), intralacustrine radiations represent some of

the best-supported cases of sympatric speciation across the Tree of Life (Schluter and McPhail

1993; Bolnick and Fitzpatrick 2007; Seehausen and Wagner 2014; Kautt et al. 2020). The

relative contribution of speciation in rivers versus lakes to freshwater diversity overall is an open

question (Seehausen and Wagner 2014).

A related question is the pace of speciation inherent to allopatric versus sympatric

speciation modes. Speciation is thought to proceed slowly in allopatry because genetic

differences between isolated populations accumulate passively (genetic drift) or through gradual

local adaptation. Reproductive isolation may be achieved faster in sympatry than in allopatry

because disruptive selection may actively select against hybridization (Bush 1975; McCune and

Lovejoy 1998; Crow et al. 2010; Germain et al. 2020). McCune and Lovejoy (1998) showed that

sympatric lacustrine sister-species had fewer genetic differences between them than allopatric

riverine sister-species, suggesting that reproductive isolation was achieved faster between

sympatric sister-species. However, later studies in other fishes found that speciation can also be

rapid in allopatry in both lacustrine and riverine habitats (Hrbek et al. 2002; Near and Bernard

2004). It is unclear what generalities can be drawn from these studies limited to a few well-

studied fish clades.

There are at least two reasons why we may expect differences in net diversification rates

in fishes found in riverine versus lacustrine habitats. First, these habitats may be differently

conducive to alternative speciation modes which differ in their time to completion. Allopatric

speciation seems to be the dominant mode in rivers because rivers become fragmented and

change course through time. Evidence for this statement comes from the allopatric distribution of

closely related species in river systems around the world (Near and Bernard 2004; Dias et al.

2013; Albert et al. 2020; Sholihah et al. 2021). A few cases of sympatric or parapatric speciation

within rivers are well-supported (mormyrid fishes, Sullivan et al. 2002; African *Teleogramma*

cichlids, Alter et al. 2017; Neotropical *Crenicichla* cichlids, Burress et al. 2018), but in general

sympatric speciation appears to be less common in rivers than in lakes (Seehausen and Wagner

2014; Seehausen 2015). If speciation rates are faster in sympatry than in allopatry, then we may

expect lacustrine fishes to have faster speciation and net diversification rates than riverine fishes

(McCune and Lovejoy 1998).

Second, rivers and lakes may differ in ecological opportunity (Yoder et al. 2010). Early

literature posited that lakes should have faster speciation rates for ecological and structural

reasons. Briggs (1966) and Lowe-McConnell (1969) thought that rates of speciation should

generally be faster in peripheral habitats than in centers of species diversity, with both authors

comparing rivers and lakes. Lowe-McConnell (1969) and later Seehausen (2015) also argued

that there may be fewer available niches in rivers: since rivers are locally unstable, riverine fishes

are more likely to be opportunistic feeders. The more stable conditions of lakes allow for trophic

specialization. In addition, since lakes tend to be wider and deeper than most rivers, they offer

three dimensions to partition habitats: onshore-offshore, shallow-deep, and along the shoreline

(Seehausen 2015). This dimensionality may encourage divergent selection, promote species

coexistence and allow for faster speciation (Germain et al. 2020).

Differences in diversification rates among freshwater habitats may be a crucial missing

piece to interpreting comparisons between freshwater and marine diversification. Specifically, an

alternative hypothesis to explain the freshwater paradox is that freshwater diversity generally

accumulated over long periods of time rather than at a very fast rate (Wiens 2012; Carrete Vega

and Wiens 2012; Miller and Román-Palacios 2021). This could still be true if most freshwater

species arose through fine-scale allopatric speciation as predicted (e.g., Grosberg et al. 2012). If

so, the higher diversification rates sometimes observed in freshwater relative to marine fishes

(Table 1) may actually be driven by rapid speciation within lakes, not allopatric speciation in

rivers. This would call into question the most commonly proposed mechanism for why

diversification rates seem to differ between biomes.

In this study, I compared rates of speciation, extinction, and net diversification between

marine and freshwater teleost fishes. However, unlike past studies, I compared these rates in

riverine and lacustrine habitats separately rather than treating freshwater as a single category. I

found that diversification rates in riverine and marine fishes are actually similar on average.

However, diversification rates in lakes are much faster than in other aquatic habitats. These

results show that more nuance is needed when comparing marine and freshwater diversification

patterns.

MATERIAL AND METHODS:

Data acquisition:

In this study I used the largest available time-calibrated phylogeny of ray-finned fishes

for all analyses (Rabosky et al. 2018). This maximum likelihood phylogeny was constructed by

aggregating genetic data from many past studies, and contains 11,638 species (36.9% of

described actinopterygian fishes). I limited the present study to teleost fishes, a group containing

99.8% of ray-finned fishes, because living non-teleost clades are much older with many extinct

members (Betancur-R et al. 2015) and it may not be appropriate to compare their diversification

rates with teleosts. I removed 51 tips that were duplicate or unresolved species and 48 tips

belonging to the four living non-teleost orders (Polypteriformes, Acipenseriformes,

Lepisosteiformes, and Amiiformes), leaving phylogenetic data for 11,539 species.

To assign each species in the phylogeny to aquatic habitat categories I used descriptions

of geographic range and habitat in FishBase (Froese and Pauley 2019), the IUCN Red List

version 6.2 (2020), and a recent compilation of habitat in fishes (Corush 2019). I identified

species that were marine, freshwater, diadromous, or estuarine/brackish. Freshwater species were

further categorized as riverine, lacustrine, or occurring in both lakes and rivers. Some species

were found in additional freshwater habitats including swamps, ponds, temporary pools, caves,

and springs. I found that diversification rates in these habitats were similar to those in rivers

(details in Extended Text 1; Figure S1), perhaps because these habitats tend to be created or

maintained by fluvial action. Species found in these other habitats were simultaneously found in

rivers much more often than in lakes (Table S1). For these reasons, I used the species'

distribution in rivers and/or lakes as the sole basis for its habitat categorization. This approach

was not applicable to the (very few) species that were endemic to these other habitats (Table S1).

Rather than exclude these species or include many additional habitat categories in my analyses, I

coded these species as riverine.

Of the 11,539 species in the phylogeny, I identified 4,984 as exclusively marine (44.4%),

631 as brackish, estuarine or euryhaline (5.6%), 263 as diadromous (2.3%), 3,985 as riverine

(35.5%), 510 as lacustrine (4.5%), and 854 in both rivers and lakes (7.6%). I removed the

remaining 312 species with unclear habitat affinities from analyses, leaving 11,227 species.

Comparing diversification rates among habitats:

I compared diversification rates among habitat types using tip-associated rates (Title and

Rabosky 2019) calculated for each species by Rabosky et al. (2018). Rates associated with each

species were downloaded using the R package fishtree version 0.2.0 (Chang et al. 2019). I

compared seven alternative rate types: (1) the DR statistic (Jetz et al. 2012) which was calculated

from the phylogeny of ray-finned fishes with all missing species imputed on the phylogeny, (2)

speciation calculated by Bayesian Analysis of Macroevolutionary Mixtures (BAMM; Rabosky

2014) under a time-varying model on the phylogeny with genetic data only, (3) speciation

calculated by BAMM under a time-constant model, (4) extinction calculated by BAMM under a

time-varying model, (5) extinction calculated under a time-constant model, (6) BAMM net

diversification rates (speciation minus extinction rates) under a time-variable model, and (7)

BAMM net diversification rates under a time-constant model. Note that tip rates were not

available for 172 species (Chang et al. 2019). Analysis settings used to obtain these rates are

detailed in the original study (Rabosky et al. 2018). The DR statistic for any given species is

estimated from the number of branches (i.e. splitting events) between the species and the root of

the tree and the length of those branches (Jetz et al. 2012). This type of rate estimate resembles

speciation rates more closely than net diversification rates (Title and Rabosky 2019). BAMM

uses a model of diversification to simulate a posterior distribution of rate shift configurations, in

which taxa in the same rate shift cohort have identical tip rates. While -SSE models can also be

used to compare diversification rates among habitats (Table 1), I chose to use rates calculated

using methods agnostic to traits (DR, BAMM). This is because it is straightforward to compare

rates among several habitat categories at once using these trait-agnostic rates, while -SSE models

become very computationally intensive when more than two traits are considered especially at

this broad phylogenetic scale.

I statistically compared the mean tip rates of habitats using phylogenetic ANOVA

(Garland et al. 1993) implemented in phytools version 0.6–44 (Revell 2012). The *phylANOVA*

function in phytools implements a simulation-based post hoc test to compare pairwise

differences in means, and also uses a Holm-Bonferroni correction for multiple comparisons. I

made a total of four habitat comparisons with and without cichlids, using each of the seven rate

types (see above), for a total of 56 comparisons. First, I compared tip-associated diversification

rates between species in two habitats, as classically done (Table 1): exclusively marine and

exclusively freshwater (excluding diadromous or estuarine species). Second, I compared rates

among six habitat categories: riverine, lacustrine, both riverine and lacustrine, marine, estuarine,

and diadromous. Third, I compared four habitats: exclusively marine, riverine, lacustrine, and

both riverine and lacustrine. I excluded diadromous and euryhaline species from this comparison

because the habitat where speciation occurred may be unclear when species interact with both

freshwater and marine habitats. Fourth, I compared only freshwater habitat types (three

categories: riverine, both riverine and lacustrine, and lacustrine). Finally, I repeated these four

habitat comparisons after removing the family Cichlidae. This was to test if cichlids were driving

any differences in rates observed among habitats. Lacustrine cichlids possess the fastest

diversification rates among fishes (Burress and Tan 2017; McGee et al. 2020) and so may have

an overwhelming influence on the results.

Alternative divergence times:

The age of clades can potentially affect diversification rate estimates by shortening or

lengthening branch lengths within the clade. Several new divergence time estimates are available

for fishes using phylogenomic techniques. However, the dense sampling of the Rabosky et al.

(2018) phylogeny is needed to estimate tip-associated diversification rates. To test whether

alternative divergence estimates have cascading effects on diversification rate differences among

habitats, I used the "congruification" approach of Eastman et al. (2013). This approach uses a

reference phylogeny, which is a tree containing few exemplar tips representing higher taxa, to

time-calibrate a target phylogeny with shared higher taxa but denser species-level sampling.

Thus, the congruification approach attempts to join the efforts of systematists using alternative

approaches to build and date trees (few exemplar taxa with many base pairs per species versus

broad sampling with less data per species).

I used the genomic phylogenies of Hughes et al. (2018) for teleosts and Alfaro et al.

(2018) for Acanthomorpha (spiny-rayed fishes) as alternative target phylogenies. Notably, Alfaro

et al. (2018) found that most major clades within acanthomorphs diverged near the K-Pg

boundary (66 mya), more recently than in Hughes et al. (2018) or Rabosky et al. (2018). I

downloaded the undated maximum likelihood phylogeny of fishes (branch lengths scaled by

molecular substitutions) from the Fish Tree of Life website

(https://fishtreeoflife.org/downloads/). I implemented the congruification using the function

congruify.phylo in the R package geiger version 2.0.7 (Pennell et al. 2014) which uses TreePL

(Smith & O'Meara 2012) to time-calibrate the tree. This procedure resulted in two rescaled trees:

a phylogeny of teleosts with identical sampling as Rabosky et al. (2018) but dates similar to

Hughes et al. (2018), and a phylogeny of acanthomorphs with identical sampling as Rabosky et

al. (2018) but dates similar to Alfaro et al. (2018).

I calculated the DR statistic using each rescaled tree and compared diversification rates

among habitat categories as detailed above. This set of analyses added 16 comparisons (4 habitat

combinations, with and without cichlids, using each re-scaled phylogeny) to the 56 performed

using the original phylogeny for a total of 72 comparisons among three phylogenies.

Biogeographic model fitting:

I found that global freshwater biodiversity was mostly found in rivers, with much lower

richness distributed among lakes (Fig. 1). Species richness among habitats can be explained by

three processes: in-situ speciation, extinction, and dispersal (transitions) among habitats

(Ricklefs 1987; Wiens 2012). To understand why there are much fewer species in lakes than in

rivers, I used biogeographic models to trace the history in each habitat along the phylogeny

(Rabosky et al. 2018). I fit alternative biogeographic models implemented in the program

BioGeoBEARS version 1.1.2 (Matzke 2014). I summarize my approach to model fitting here and

give more details in Extended Text 2. Note that these analyses are similar to those in Miller and

Román-Palacios (2021). The goal of that study was to compare colonization patterns and

diversification rates among the freshwater faunas of biogeographic regions, but did not

distinguish between lakes and rivers within these regions.

I modeled dispersal among 13 habitat-region combinations. I included six biogeographic

regions following Leroy et al. (2019), who used clustering algorithms to identify a

bioregionalization scheme based on shared freshwater fish diversity (region names also follow

this study). The six regions were: (1) Nearctic, (2) Neotropical, (3) Palearctic, (4) Ethiopian, (5)

Sino-Oriental, and (6) Australia. Note that Miller and Román-Palacios (2021) used the earlier

regionalization of Tedesco et al. (2017b) which has minor differences in boundaries. Species in

these six regions were further assigned to riverine and/or lacustrine habitats, giving 12 region-

habitat combinations (e.g. Neotropical rivers and Neotropical lakes). Marine and estuarine

species were coded as belonging to a 13th "marine" region. I restricted the root of the teleost

phylogeny to the marine state in accordance with the fossil record of fishes (Betancur-R et al.

2015). Following prior biogeographic studies in freshwater systems (Toussaint et al. 2017; Miller

and Román-Palacios 2021), I restricted dispersal probabilities in accordance with changing

regional connectivity through time (Table S11). These dispersal constraints were common to all

models.

I also compared the fit of six model variants with modified transition matrices in order to

test hypotheses about the role of lakes in dispersal. In variant 1, dispersal among biogeographic

regions was unrelated to habitat. For example, dispersal from the Neotropical to the Nearctic

region could occur as easily within the riverine as the lacustrine state, and marine-to-lacustrine

transitions could occur as easily as marine-to-riverine transitions. In variant 2, the probability of

transition between marine and lacustrine habitats was reduced (multiplied by a scalar value of

0.05). This model would be preferred if most transitions from marine to freshwater habitats

occurred via rivers. In variant 3, the probability of dispersal among biogeographic regions was

much lower via lakes than via rivers (i.e., all regional dispersal probabilities within the lacustrine

state were multiplied by 0.05). For example, dispersal from Neotropical to Nearctic rivers was

more likely than dispersal from Neotropical to Nearctic lakes. This model would be preferred if

most biogeographic dispersal occurred within rivers. In variant 4, restrictions on lacustrine

dispersal were relaxed such that lacustrine dispersal occurred at the same probability as riverine

dispersal across adjacent regions or short marine barriers, but lacustrine dispersal was

downweighed across large overland or overseas barriers (multiplied by 0.05). For example,

dispersal from Neotropical to Nearctic rivers was equally likely as from Neotropical to Nearctic

lakes; however, dispersal from the Neotropical rivers to Ethiopian rivers was more likely than

the equivalent through lakes. Variant 5 combined marine-lake constraints with strict constraints

on biogeographic dispersal through lakes. Variant 6 combined marine-lake constraints with

relaxed constraints on dispersal through lakes.

In total, I fit 36 alternative biogeographic models (Table 2). These were six models with

varying dispersal matrices for each of six base model types (DEC, DEC+J, DIVALIKE,

DIVALIKE+J, BAYAREALIKE, and BAYAREALIKE+J). Models with +J denote the addition

of a jump parameter allowing cladogenetic dispersal; otherwise, an assumption of models is that

all dispersal occurs anagenetically (further differences among models are described in Matzke

2014). The best-fit model was chosen by comparing Akaike weights (Burnham and Anderson

2002).

Biogeographic patterns:

In order to identify independent dispersal events and visualize uncertainty in the ancestral

reconstruction, I simulated 100 biogeographic stochastic maps (Dupin et al. 2017) using the best-

fit model. Each individual simulation is a realized history that is possible given the model and

data, including the time and location on the branches for biogeographic events. Averaging over

all of these simulations will approximate the ancestral state probabilities calculated by the model.

301   To understand why there are fewer species in lakes than rivers, I used biogeographic

stochastic maps to answer the following questions. First, what is the rate of transition into and

out of lakes relative to other habitats? To obtain habitat transition rates, I counted the number of

transitions to marine, riverine only, lacustrine only, and riverine+lacustrine states (combining

these counts among regions for each habitat type). The count of events alone can be a misleading

measure of the transition rate. As an example, if most freshwater lineages occur in rivers then

there may be more transitions from rivers to lakes simply by chance. To convert these counts

into a transition rate, for each transition type I divided the number of transitions from the source

habitat to the receiving habitat by the sum of branch lengths reconstructed in the source habitat.

For example, to calculate the transition rate from marine to riverine I divided the count of these

transitions by the sum of branch lengths reconstructed in the marine state. These counts vary

slightly among stochastic maps, so I did this for all 100 simulations individually and then took

the mean of the 100 rate estimates.

314   Second, what is the contribution of individual transition events to overall freshwater

diversity, and are there replicated patterns shared by habitats across biogeographic regions?

More specifically, for each lineage representing an independent transition to a habitat I counted

the number of descendants of that transition (species richness; note that this count only included

species remaining in that habitat by the present), the mean tip-associated diversification rate

among the descendants (DR), and the age of the lineage (time of the transition). Then, I took the

mean of these values among lineages found in each habitat and biogeographic region. Again, the

estimated location of habitat transitions on the phylogeny varies slightly among stochastic maps,

so I did this for all 100 simulations. Note that the richness of independent habitat transitions was

inferred by counting tips in the phylogeny descended from these transitions, because I could not

assign unsampled species in the phylogeny to individual biogeographic events. I assessed the

potential sensitivity of these results to biased sampling in Extended Text 3.

Finally, I visualized the accumulation of diversity through time in each freshwater habitat

and region using the lineage-through-space-and-time approach of Skeels (2019) implemented

using the R package ltstR version 0.1.0. This approach is analogous to a lineage-through-time

plot but uses the output of biogeographic stochastic mapping to separate temporal patterns of

species accumulation by region.

RESULTS

Diversification rates in aquatic habitats:

The results of 56 rate-habitat comparisons using the phylogeny of Rabosky et al. (2018)

are summarized in Table 3 and given in full in Tables S2–S8. When comparing only two habitat

categories, net diversification rates in marine and freshwater teleosts were broadly overlapping

but slightly higher in freshwater (mean net diversification rate=0.10 species/million years in

marine and 0.17 in freshwater under a BAMM time-varying model; Fig. 1). This difference was

not significant using phylogenetic ANOVA no matter whether speciation, extinction or net

diversification rates were used, the method to calculate these rates, or whether the family

Cichlidae was included (Table 3).

When comparing six habitat categories, lacustrine fishes had much faster speciation,

extinction and net diversification rates than other aquatic habitats (mean net diversification

rate=0.63 under a BAMM time-varying model; Fig. 1). Surprisingly, rates in marine and riverine

habitats were similar (mean net rate=0.10 in marine and 0.12 in riverine habitats). Species found

in both rivers and lakes diversified at rates more similar to rivers than to lakes (mean net

rate=0.15). Estuarine and diadromous species had slightly lower net diversification rates than

other habitats (mean net rate=0.07 in estuaries and 0.08 in diadromous species).

Comparisons of net diversification rates among six habitat categories using phylogenetic

ANOVA were significant when including all teleosts (BAMM time-varying model: P=0.047;

BAMM time-constant model: P=0.041). Differences in speciation and extinction rates were not

significant, nor were net diversification rates after excluding Cichlidae (Table 3). Still, lakes

were significantly different from the other five habitats in speciation and net diversification rates

in all pairwise post hoc tests, even when the ANOVAs were not globally significant. This was

true using all methods to calculate rates and whether or not cichlids were included (Table 3).

Comparing four habitat categories (excluding estuarine and diadromous species) resulted in the

same pattern as comparing six: phylogenetic ANOVAs were not globally significant, but lakes

were significantly different in speciation and net diversification rates from the other three

habitats in pairwise post hoc tests. When comparing just the three freshwater habitats (riverine,

lacustrine, and both riverine and lacustrine), phylogenetic ANOVAs became globally significant

in speciation, extinction and net diversification rates. This was driven by faster rates in lakes

compared to rivers or rivers+lakes, no matter the rate method or exclusion of cichlids. Among all

56 comparisons, pairwise tests were never significant between other habitat categories except for

one other case (diadromous species had faster extinction rates than estuarine or marine species;

Tables S3, S6).

Taken together, these results suggest that: (1) lakes have faster speciation and net

diversification rates than other freshwater habitats (and potentially faster extinction, though this

was not significant), (2) rapid lacustrine diversification rates are not limited to cichlids, and (3)

marine rates are similar to riverine rates. As a consequence of the latter result, comparing just

two categories (freshwater and marine) can mask the signal of rapid lacustrine diversification

since riverine species are much more common than lacustrine species (Fig. 1). The similarity

between marine and riverine diversification rates also explains why phylogenetic ANOVAs were

not significant when comparing four categories but were significant when comparing the three

freshwater categories (Table 3).

These patterns held when using phylogenies re-dated in reference to phylogenomic

hypotheses (Alfaro et al. 2018; Hughes et al. 2018). The results of the 16 additional comparisons

using re-dated trees are summarized in Fig. 2 and shown in full in Tables S9–S10.

Unsurprisingly, re-scaling the phylogeny of Acanthomorpha using the Alfaro et al. (2018)

hypothesis resulted in slightly faster rates for all habitats because the younger divergence time

estimates shortened branch lengths across the entire phylogeny. Still, diversification rates (DR

statistic) were generally similar before and after time-calibrating the Rabosky et al. (2018) tree

based on either genomic phylogeny (Fig. 2; Fig. S2). Lacustrine species still had the fastest

diversification rates among aquatic habitats as shown by pairwise post hoc tests.

Habitat transitions and the formation of freshwater diversity:

The best-fitting biogeographic model (BAYAREA+J with dispersal matrix variant 6;

Table 2) constrained marine-lacustrine transitions to be less probable than marine-riverine

transitions. This model also applied some constraints on regional dispersal while in the lacustrine

state. Rates of cladogenetic ("jump") dispersal were six times higher than rates of anagenetic

dispersal (j=0.0078 events/million years versus d=0.0013; Table 2), suggesting that founder

events are an important means of dispersal and speciation among freshwater habitats and regions.

In general, marine and riverine habitats were reconstructed in deep ancestral nodes, while

the lacustrine state was concentrated towards more recent nodes (Fig. 3A). The rate of transition

from marine to riverine habitats was seven times higher than the rate from marine to lacustrine

(0.0007 versus 0.0001 events per million years; Fig. 3B). The transition rate from the joint

river+lake state into lakes alone was over twice as high as the rate of direct dispersal from rivers

to lakes (0.0031 versus 0.0013). Thus, the most typical means of becoming a lacustrine fish was

to first transition from marine to riverine habitats, then from rivers alone to both rivers and lakes,

then finally becoming endemic to lakes.

Transition rates out of lakes were higher than transition rates into lakes (Fig. 3B). The

rate of transition from lakes alone back to rivers+lakes was about seven times higher than the

reverse (0.0223 events/million years versus 0.0031). The direct dispersal rate from lakes to rivers

was over three times as high as the reverse (0.0048 versus 0.0013). This suggests that of the few

lineages that become endemic to lakes, many will eventually enter rivers again.

I compared four metrics characterizing lineages associated with habitat transitions: the

number of independent habitat transitions, mean richness per habitat transition, mean timing of

habitat transitions, and mean diversification rate (DR). Differences in these metrics among

rivers, lakes, and rivers+lakes were generally shared across the six biogeographic regions (Fig.

3D). Transitions into the rivers+lakes state were much more common than transitions into rivers

or lakes alone for four of six regions (all but Neotropical and Australian regions). Transitions

into lakes were the least common in five of six regions (all but Sino-Oriental).

412   Mean species richness per habitat transition was highest for the riverine state in five of

413 six regions (Fig. 3D). The mean richness of transitions into lakes was low in all but the Ethiopian

414 region, where richness-per-transition was similar between rivers and lakes. Note that these

415 richness differences cannot be attributed to sampling biases by habitat: lacustrine species were

416 better sampled in the phylogeny than riverine species (Extended Text 3). In all six regions,

417 lineages transitioning into the rivers+lakes state were usually limited to single species,

418 suggesting that this habitat is a transitional state between rivers and lakes.

419   In all six regions, riverine transitions were older on average than transitions into the

420 rivers+lakes or lacustrine states (Fig. 3D).

421   The mean diversification rate (DR) of lacustrine lineages was higher than that of the other

422 habitat states in all regions. The difference between lacustrine and riverine diversification rates

423 was large in the Ethiopian, Australian, Nearctic and Palearctic regions, and smaller in the

424 Neotropical and Sino-Oriental regions (Fig. 3D).

425   The temporal patterns of lineage accumulation in rivers and lakes worldwide reflect these

426 differences in transition rates, transition timing, and diversification rates with habitat (Fig. 3C).

427 All six regions have much greater diversity contained in rivers than lakes, even though average

428 diversification rates were always higher in lakes (Fig. 3C). Freshwater diversity has been highest

429 overall in Neotropical and Sino-Oriental rivers for the past 75 million years, reinforcing the role

430 of time for explaining these mega-diverse faunas (Miller and Román-Palacios 2021). Only the

431 Ethiopian region has lacustrine diversity rivalling total riverine diversity in some other regions,

432 which is due to the exceptionally fast diversification rates in African lakes (Fig. 3D). Note that

433 time and diversification-rate patterns characterizing biogeographic regions themselves were

434 consistent with those found by Miller and Román-Palacios (2021).

Taken together, these biogeographic results (Table 2; Figure 3) suggest that most global

freshwater diversity is riverine in spite of faster lacustrine diversification rates for the following

reasons. First, most lineages transitioning to freshwater from marine biomes enter rivers first.

Second, transitions from rivers to lakes are relatively infrequent. Third, lineages leave lakes more

often than they enter lakes. For these reasons, riverine lineages have had greater opportunity to

build diversity over long periods of time than lacustrine lineages.

DISCUSSION

Why do freshwater habitats have higher species richness than expected given their total

volume on Earth, especially compared to the ocean (Cohen 1970; Horn 1972)? Many past studies

compared speciation and/or net diversification rates between freshwater and marine organisms in

order to answer this question, but collectively have found mixed results (Table 1). In this study, I

compared diversification rates in marine versus freshwater teleost fishes while separating

freshwater species by riverine and lacustrine habitats. Contrary to expectations, marine and

riverine fishes have similar net diversification rates on average. Lacustrine fishes have much

faster diversification rates than all other aquatic habitats yet make up a small fraction of overall

freshwater diversity. Therefore, I suggest that diversification rate differences are not sufficient to

explain the freshwater paradox, and a more nuanced explanation is needed.

It is generally expected that freshwater fishes should have faster speciation and net

diversification rates than marine fishes because freshwater habitats are more fragmented, so there

are presumably more opportunities for speciation in freshwater than in the ocean (Grosberg et al.

2012; Tedesco et al. 2017a). This explanation implies that most freshwater diversity arose

through allopatric speciation, an implication supported by the geographic distributions of closely

related freshwater fish species (Lynch 1989; Hrbek et al. 2002; Near and Benard 2004; Dias et

al. 2013; Albert et al. 2020; Sholihah et al. 2021). One potential problem with this explanation is

that allopatric speciation is thought to proceed more slowly than sympatric speciation (McCune

and Lovejoy 1998; Crow et al. 2010). Most freshwater diversity arose in rivers (Figs. 1, 3),

which suggests that most freshwater species are indeed the product of allopatric speciation

(assuming that this is the most common speciation mode in rivers). However, speciation and net

diversification rates of riverine fishes were not much faster than those of marine fishes on

average (Fig. 1). This finding seems to contradict the leading hypothesis for why freshwater

fishes should have faster speciation rates than marine fishes. Still, this hypothesis is supported in

part: allopatric speciation *sensu lato* appears to be important for explaining high freshwater

species richness, but not necessarily fast rates of allopatric speciation. The fine-scale

fragmentation of rivers might allow allopatric populations to complete the speciation process

without being outcompeted by congeners or reabsorbed through hybridization (Germain et al.

2020), thus allowing the accumulation of many species over a relatively small spatial scale

compared to other habitats. The density of speciation events in space can be decoupled from the

frequency of events over time (Boucher et al. 2020).

How then can we explain the freshwater paradox, if not by faster diversification rates in

freshwater versus the ocean? The results of biogeographic reconstructions (Fig. 3) suggest that

explanations invoking geologic time could be more appropriate (Stephens and Wiens 2003;

Wiens 2012). Diversity has been accumulating for comparably long periods of time in modern

marine and riverine lineages of teleosts (over 100 million years; Fig. 3A). Given this amount of

time, faster speciation is not necessary to explain high richness in freshwater. In fact, it seems

that speciation is about equally as frequent in freshwater and the ocean (Fig. 1). This similarity in

time-for-speciation, as well as the similarity in average net diversification rates, is consistent

with the roughly similar species richness in riverine and marine habitats today (Fig. 1). Put

another way, high freshwater diversity might be explained by the early colonization of

freshwater in the history of teleosts, regular opportunities for speciation because of the

continuous fragmentation of rivers, and protection from extinction allowing diversity to

accumulate over long time scales. This time-based explanation for the freshwater paradox has

also been suggested by past studies (Carrete Vega and Wiens 2012, Seehausen and Wagner

2014, Betancur-R et al. 2015; see also Miller and Wiens 2017, Miller and Román-Palacios

2021), but seems to have been given less attention from researchers than diversification rates

(Table 1).

How do we reconcile the observation that speciation rates do not differ between riverine

and marine habitats with evidence to the contrary from past studies? Manel et al. (2020) found

that freshwater fishes have higher intraspecific genetic diversity than marine fishes. This

presumably reflects the greater fragmentation of freshwater habitats, which could in principle

lead to faster speciation rates. However, the authors found that genetic diversity was weakly

related to local species richness (Correlation coefficient=0.20 in marine and 0.36 in freshwater,

Manel et al. 2020). Population subdivision may not have a straightforward relationship with

speciation rates since there may be distinct (and sometimes competing) factors driving

population isolation, reproductive isolation, persistence and coexistence of incipient species

(Harvey et al. 2019; Germain et al. 2020). For example, allopatric speciation creates daughter

species with smaller population sizes than the parent species, which leads to greater risk of

stochastic extinction (Moen and Morlon 2014). Further, species richness is itself only partially

related to diversification rates and also depends on historical factors (Ricklefs 1987; Wiens

2012). Spatial patterns of species richness are more closely related to dispersal frequency and

timing than to diversification rates in both the ocean (Miller et al. 2018) and freshwater (Miller

and Román-Palacios 2021). Therefore, the links between population subdivision, speciation, and

species richness can break down or be influenced by confounding factors.

Some macroevolutionary studies did find faster speciation and/or net diversification rates

in freshwater (e.g., Bloom et al. 2013, Tedesco et al. 2017a, Miller et al. 2018; see Table 1). In

light of the present study, it is possible that faster freshwater rates inferred by past studies were

actually driven by lacustrine species, and not reflective of a general difference between marine

and freshwater biomes. Indeed, Rabosky (2020) found that apparently faster average freshwater

rates of speciation in actinopterygians were driven by select clades and were not broadly

characteristic of marine-to-freshwater transitions. Lacustrine cichlids in particular have the

fastest speciation rates among all fishes (Burress and Tan 2017; McGee et al. 2020). Future

studies should aim to explain the variation in diversification rates seen within freshwater fishes,

which are diverse in age and ecology.

The other side of the coin is that diversification rates in marine fishes are faster than

expected given the apparent lack of barriers to dispersal in the ocean. Some marine lineages are

known to have exceptionally fast speciation rates (Rüber and Zardoya 2005; Rabosky et al. 2018;

Siqueira et al. 2020; see also Miller and Wiens 2017 for marine amniotes). Marine fishes are not

a monolithic group, and a general expectation for low speciation rates in the ocean seems overly

simplistic. The mechanisms for speciation in the marine environment appear to be as varied as

those found in continental biomes, and speciation with gene flow may be common (Crow et al.

2010; Bowen et al. 2013). Furthermore, while there is a 7,500-fold difference in water volume

between the ocean and freshwater (Horn 1972), most marine biodiversity is found along

coastlines, not in the open ocean. In particular, coral reefs contain ~40% of marine fishes within

only 0.1% of the Earth's surface area (Cowman et al. 2017). The similarity in species richness

between marine and freshwater fishes (Fig. 1) does not seem as puzzling when considering the

distribution of species within the ocean.

The most famous examples of lacustrine radiation are cichlids in the African rift lakes

(Wagner et al. 2012; Seehausen 2015; Burress and Tan 2017; McGee et al. 2020). However,

rapid diversification in lakes was not limited to cichlids (Fig. 2) nor African lakes (Fig. 3).

Diversification rates were faster in lakes than in rivers within all six continental regions of the

world (Fig. 3D). Lacustrine radiations with elevated diversification rates other than cichlids

include: Lake Titicaca *Orestias* (mean DR=0.6 species/million years), *Coregonus* of Nearctic

(DR=2.67) and Palearctic (2.09) postglacial lakes, Lake Baikal sculpins (0.71), Lake Tana

Labeobarbus (4.08), and Sulawesi telmatherinid rainbowfishes (0.31). Additional examples are

summarized in Table 4.

If diversification rates are so fast in lakes, then why do lakes contain a small proportion

of freshwater diversity (Figs. 1, 3C)? Biogeographic models suggested that this contradictory

pattern can be explained by transition rates among habitats (Fig. 3B). In the best-fitting model,

direct transitions from the ocean into lakes were constrained to be rarer than marine-riverine

transitions (Table 2). This is intuitive, as lakes are fed by rivers and do not usually interface

directly with the ocean (but see Schluter and McPhail 1992 and Hughes et al. 2020 for examples

of marine-lacustrine connections). The most common path to entering lakes is by first entering

rivers, then occurring in both rivers and lakes, then finally becoming endemic to lakes (Fig. 3B).

Since there are several evolutionary steps to becoming lacustrine, lacustrine radiations were

usually nested within riverine lineages (Fig. 3A). Therefore, lacustrine radiations had less time-

for-speciation than riverine counterparts (Fig. 3D; Stephens and Wiens 2003). In addition,

transition rates out of lakes were higher than those into lakes (Fig. 3B). This suggests that being

endemic to lakes is an evolutionarily unstable state.

These biogeographic results can be framed in light of the age of rivers versus lakes. On a

local scale, conditions in rivers change frequently and seasonally, while conditions in lakes are

more stable. However, lakes are generally younger than rivers over geologic timescales because

lakes dry out and can disappear entirely (Lowe-McConnell 1969; McCune 1987; Joyce et al.

2005; Cristescu et al. 2010). Indeed, the fossil record documents past lacustrine radiations that

went extinct and were replaced as lakes dried out and refilled (McCune 1987). Lacustrine fishes

may avoid extinction by entering rivers (Joyce et al. 2005; Seehausen and Wagner 2014),

explaining the high transition rates out of lakes. Another consideration is that rapid

diversification rates are probably short lived, and the richness of lacustrine radiations may

ultimately be constrained by the features of individual lakes (Wagner et al. 2012, 2014). It is

noteworthy that many intralacustrine radiations are limited to populations within a single species,

or pairs of species (McPhail and Schluter 1992, 1993; Hudson et al. 2007). In contrast, some

large and ancient lakes host a greater diversity of species (Cohen 1995; Wagner et al. 2014) but

these lake systems may be atypical.

It is somewhat unexpected that rivers have lower speciation rates than lakes, since the

fragmentation of rivers promotes population subdivision and endemism (Dias et al. 2013). Note

however that a speciation rate represents the number of species generated per unit of time, and

not simply the number of species generated; therefore, the occurrence of many speciation events

might not translate to a fast speciation rate if these events occurred over several million years.

McCune and Lovejoy (1998) suggested that the reason for faster speciation rates in lakes was

that sympatric speciation completes faster than allopatric speciation. My results across fishes are

consistent with McCune and Lovejoy's findings, assuming that sympatric speciation is more

common in lakes than in rivers (Bolnick and Fitzpatrick 2007; Seehausen and Wagner 2014).

Still, I did not directly identify geographic modes of speciation in this study. At the time of

writing, phylogenetic sampling across fishes is still generally below that needed to accurately

infer the speciation mode from present-day geographic ranges (Skeels and Cardillo 2019). In

addition, allopatric speciation occurring over short distances within a lake can be difficult to

distinguish from sympatric speciation without fine-scale spatial and ecological data (Rico and

Turner 2002; Seehausen and van Alphen 1999). An unanswered question is whether allopatric

speciation within and among lakes is also generally faster than allopatric speciation in rivers.

Some lacustrine radiations for which allopatry appears to be the dominant mode also have rapid

diversification rates (Table 4).

Besides speciation mode, there are ecological differences between rivers and lakes that

could explain faster lacustrine diversification rates. Compared to rivers, lakes may allow for

more ecological specialization because of their more stable conditions and greater structural

diversity (Seehausen 2015). Stronger competition and predation pressure in rivers may also

inhibit speciation (Briggs 1966; Lowe-McConnell 1969; Seehausen 2015), such that fishes

experience ecological release when they colonize lakes (Yoder et al. 2010; Burress and Tan

2017). The observation that the present-day richness of many lakes appears to be below the

lake's carrying capacity (Barbour and Brown 1974; Doenz et al. 2018; Hauffe et al. 2020)

supports the idea of high ecological opportunity in lakes.

In conclusion, I found that lacustrine fishes have faster speciation and net diversification

rates than other aquatic habitats. Still, lakes represent a small fraction of freshwater diversity

compared to rivers. Surprisingly, riverine and marine fishes had similar rates of diversification,

despite past literature predicting that these habitats had opposite effects on diversification (Horn

1972; May 1994; Grosberg et al. 2012; Tedesco et al. 2017a). While allopatric speciation is

indeed important for generating freshwater diversity, this may not be the mechanism driving

faster speciation rates between freshwater and marine habitats observed in some past studies.

Furthermore, these results support an alternative explanation for the freshwater paradox.

Speciation is roughly as frequent in freshwater and marine habitats on average, and teleosts have

occupied these habitats for comparable amounts of time over their 200-million-year history.

Future research should aim to identify ecological and evolutionary mechanisms for how riverine

habitats support long-term diversification and lineage persistence while lacustrine habitats

promote rapid diversification.

607
608

609   **REFERENCES**

611   Alfaro, M. E., Santini, F., Brock, C., Alamillo, H., Dornburg, A., Rabosky, D. L., Carnevale, G.,

612   and L. J. Harmon. 2009. Nine exceptional radiations plus high turnover explain species

613   diversity in jawed vertebrates. Proc. Nat. Acad. Sci. U.S.A. 106:13410–13414.

614   Alfaro, M. E., B. C. Faircloth, R. C. Harrington, L. Sorenson, M. Friedman, C. E. Thacker, C. H.

615   Oliveros, D. Černý, and T. J. Near. 2018. Explosive diversification of marine fishes at the

616   Cretaceous-Palaeogene boundary. Nat. Ecol. Evol. 2:688–696.

617   Albert, J. S., V. A. Tagiacollo, and F. Dagosta. 2020. Diversification of Neotropical freshwater

618   fishes. Ann. Rev. Ecol. Evol. Syst. 51:27–53.

619   Alter, S. E., J. Munshi-South, and M. L. J. Stiassny. 2017. Genomewide SNP data reveal cryptic

620   phylogeographic structure and microallopatric divergence in a rapids-adapted clade of

621   cichlids from the Congo River. Mol. Ecol. 26:1401–1419.

622   Anseaume, L. and G. G. Teugels. 1999. On the rehabilitation of the clariid catfish genus

623   *Bathyclarias* endemic to the East African Rift Lake Malawi. J. Fish. Biol. 55:405–419.

624   Barbour, C. D., and J. H. Brown. 1974. Fish species diversity in lakes. Am. Nat. 108:473–489.

625   Beaulieu, J. M., and B. C. O'Meara. 2016. Detecting hidden diversification shifts in models of

626   trait-dependent speciation and extinction. Syst. Biol. 65:583–601.

627   Betancur-R, R., G. Ortí, A. M. Stein, A. P. Marceniuk, and R. A. Pyron. 2012. Apparent signal

628   of competition limiting diversification after ecological transitions from marine to

629   freshwater habitats. Ecol. Lett. 15:822–830.

630   Betancur-R, R., G. Ortí, and R. A. Pyron. 2015. Fossil-based comparative analyses reveal ancient

631   marine ancestry erased by extinction in ray-finned fishes. Ecol. Lett. 18:441–450.

Bloom, D. D., J. T. Weir, K. R. Piller, and N. R. Lovejoy. 2013. Do freshwater fishes diversify

faster than marine fishes? A test using state-dependent diversification analyses and

molecular phylogenetics of new world silversides (Atherinopsidae). Evolution 67:2040–

2057.

Bolnick, D. I., and B. M. Fitzpatrick. 2007. Sympatric speciation: models and empirical

evidence. Annu. Rev. Ecol. Evol. S. 38:459–487.

Boucher, F. C., A.-S. Quatela, A. G. Ellis, and G. A. Verboom. 2020. Diversification rate vs.

diversification density: Decoupled consequences of plant height for diversification of

Alooidae in time and space. PLoS ONE 15:e0233597.

Bowen, B. W., L. A. Rocha, R. J. Toonen, S. A. Karl, et al. 2013. The origins of tropical marine

biodiversity. Trends Ecol. Evol. 28:359–366.

Brown, K. J., L. Rüber, R. Bills, and J. J. Day. 2010. Mastacembelid eels support Lake

Tanganyika as an evolutionary hotspot of diversification. BMC Evol. Biol. 10:188.

Briggs, J. C. 1966. Zoogeography and evolution. Evolution 20:282–289.

Burnham, K. P., and D. R. Anderson. 2002. Model selection and multimodel inference: a

practical information-theoretic approach. Springer, New York.

Burress, E. D., and M. Tan. 2017. Ecological opportunity alters the timing and shape of adaptive

radiation. Evolution 71:2650–2660.

Burress, E. D., F. Alda, A. Duarte, M. Loureiro, J. W. Armbruster, and P. Chakrabarty. 2018.

Phylogenomics of pike cichlids (Cichlidae: *Crenicichla*): the rapid ecological speciation

of an incipient species flock. J. Evol. Biol. 31:14–30.

Bush, G. L. 1975. Modes of animal speciation. Annu. Rev. Ecol. Syst. 6:339–64.

Carrete Vega, G., and J. J. Wiens. 2012. Why are there so few fish in the sea? Proc. R. Soc.

Lond. B. 279:2323–2329.

Chang, J., D. J. Rabosky, S. A. Smith, and M. E. Alfaro. 2019. An R package and online

resource for macroevolutionary studies using the ray-finned fish tree of life. Methods

Ecol. Evol. 10:1118–1124.

Cohen, D. M. 1970. How many recent fishes are there? Proc. Cal. Acad. Sci. 17:341–346.

Cohen, A. S. 1995. Paleoecological approaches to the conservation biology of benthos in ancient

lakes: a case study from Lake Tanganyika. J. N. Am. Benthol. Soc. 14:654–668.

Corush, J. B. 2019. Evolutionary patterns of diadromy in fishes: more than a transitional state

between marine and freshwater. BMC Evol. Biol. 19:168.

Cowman, P. F., V. Parravicini, M. Kulbicki, and S. R. Floeter. 2017. The biogeography of

tropical reef fishes: endemism and provinciality through time. Biol. Rev. 92:2112–2130.

Cristescu, M. E., S. J. Adamowicz, J. J. Vaillant, and D. G. Haffner. 2010. Ancient lakes

revisited: from the ecology to the genetics of speciation. Mol. Ecol. 19:4837–4851.

Crow, K. D., H. Munehara, and G. Bernardi. 2010. Sympatric speciation in a genus of marine

reef fishes. Mol. Ecol. 19:2089–2105.

Day, J. J., Bills, R., and J. P. Friels. 2009. Lacustrine radiations in African *Synodontis* catfish. J.

Evol. Biol. 22:805–817.

Davis, A. M., P. J. Unmack, B. J. Pusey, J. B. Johnson, and R. G. Pearson. 2012. Marine-

freshwater transitions are associated with the evolution of dietary diversification in

terapontid grunters (Teleostei: Terapontidae). J. Evol. Biol. 25:1163–1179.

Davis, M. P., P. E. Midford, and W. Maddison. 2013. Exploring power and parameter estimation

of the BiSSE method for analyzing species diversification. BMC Evol. Biol. 13:38.

Dias, M. S., J. Cornu, T. Oberdorff, C. A. Lasso, and P.A. Tedesco. 2013. Natural fragmentation

in river networks as a driver of speciation for freshwater fishes. Ecography 36:683–689.

Doenz, C. J., D. Bittney, P. Vonlanthen, C. E. Wagner, and O. Seehausen. 2018. Rapid buildup

of sympatric species diversity in Alpine whitefish. Ecol. Evol. 8:9398–9412.

Dupin, J., N. J. Matzke, T. Särkinen, S. Knapp, R. G. Olmstead, L. Bohs, and S. D. Smith. 2017.

Bayesian estimation of the global biogeographical history of the Solanaceae. J. Biogeogr.

44:887–899.

Eastman, J. M., L. J. Harmon, and D. C. Tank. 2013. Congruification: support for time scaling

large phylogenetic trees. Methods Ecol. Evol. 4:688–691.

Fitzjohn, R. G. 2012. Diversitree: Comparative phylogenetic analyses of diversification in R.

Methods Ecol. Evol. 3:1084–1092.

Froese, R., and D. Pauly. 2019. FishBase Version 12/2019. Available at

http://www.fishbase.org/.

Garland Jr., T., A. W. Dickerman, C. M. Janis, and J. A. Jones. 1993. Phylogenetic analysis of

covariance by computer simulation. Syst. Biol. 42:265–292.

Germain, R. M., S. P. Hart, M. M. Turcotte, S. P. Otto, J. Sakarchi, J. Rolland, T. Usui, A. L.

Angert, D. Schluter, R. D. Bassar, et al. 2020. On the origin of coexisting species. Trends

Ecol. Evol. 36:284–293.

Goldberg, E. E., and B. Igić. 2012. Tempo and mode in plant breeding system evolution.

Evolution 66:3701–3709.

Goldberg, E. E., L. T. Lancaster, and R. H. Ree. 2011. Phylogenetic inference of reciprocal

effects between geographic range evolution and diversification. Syst. Biol. 60:451–465.

Grosberg, R. K., G. J. Vermeij, and P. C. Wainwright. 2012. Biodiversity in water and on land.

Curr. Biol. 22:R900–R903.

Guinot, G., and L. Cavin. 2015. Contrasting "fish" diversity dynamics between marine and

freshwater environments. Curr. Biol. 25:2314–2318.

Harvey, M. G., S. Singhal, and D. L. Rabosky. 2019. Beyond reproductive isolation:

Demographic controls on the speciation process. Ann Rev. Ecol. Evol. Syst. 50:75–95.

Hauffe, T., D. Delicado, R. S. Etienne, and L. Valente. 2020. Lake expansion elevates

equilibrium diversity via increasing colonization. J. Biogeogr. 47:1849–1860.

Herder, F., A. W. Nolte, J. Pfaender, J. Schwarzer, R. K. Hadiaty, and U. K. Schliewen. 2006.

Adaptive radiation and hybridization in Wallace's Dreamponds: evidence from sailfin

silversides in the Malili Lakes of Sulawesi. 273:2209–2217.

Horn, M. H. 1972. The amount of space available for marine and freshwater fishes. Fish. Bull.

U.S. 70:1295–1297.

Hrbek, T., F. Küçük, T. Frickey, K. N. Stölting, R.H. Wildekamp, and A. Meyer. 2002.

Molecular phylogeny and historical biogeography of the *Aphanius* (Pisces,

Cyprinodontiformes) species complex of central Anatolia, Turkey. Mol. Phylogenet.

Evol. 25:125–137.

Hudson, A. G., P. Vonlanthen, R. Müller, and O. Seehausen. 2007. Review: The geography of

speciation and adaptive radiation in coregonines. Advanc. Limnol. 60:111–146.

Hughes, L. C., G. Ortí, Y. Huang, Y. Sun, C. C. Baldwin, A. W. Thompson, D. Arcila, R.

Betancur-R., C. Li, L. Becker, et al. 2018. Comprehensive phylogeny of ray-finned fishes

(Actinopterygii) based on transcriptomic and genomic data. Proc. Nat. Acad. Sci. U.S.A.

115:6249–6254.

Hughes, L. C., Y. P. Cardoso, J. A. Sommer, R. Cifuentes, M. Cuello, G. M. Somoza, M.

González-Castro, L. R. Malabarba, V. Cussac, E. M. Habit, et al. 2020. Biogeography,

habitat transitions and hybridization in a radiation of South American silverside fishes

revealed by mitochondrial and genomic RAD data. Mol. Ecol. 29:738–751.

IUCN 2020. The IUCN Red List of Threatened Species. Version 6.2. Available at

https://www.iucnredlist.org.

Jetz, W., G. H. Thomas, J. B. Joy, K. Hartmann, and A. O. Mooers. 2012. The global diversity of

birds in space and time. Nature 491:444–448.

Joyce, D. A., D. H. Lunt, R. Bills, G. F. Turner, C. Katongo, N. Duftner, C. Sturmbauer, and O.

Seehausen. 2005. An extant cichlid fish radiation emerged in an extinct Pleistocene lake.

Nature 435:90–95.

Kautt, A. F., C. F. Kratochwil, A. Nater, G. Machado-Schiaffino, M. Olave, F. Henning, J.

Torres-Dowdall, A. Härer, C. D. Hulsey, P. Franchini, et al. 2020. Contrasting signatures

of genomic divergence during sympatric speciation. Nature 588:106–111.

Kirchhoff, K. N., T. Hauffe, B. Stelbrink, C. Albrecht, and T. Wilke. 2017. Evolutinary

bottlenecks in brackish water habutats drive the colonization of fresh water by stingrays.

J. Evol. Biol. 30:1576–1591.

Lévêque, C., T. Oberdorff, D. Paugy, M. L. J. Stiassny, and P. A. Tedesco. 2008. Global

diversity of fish (Pisces) in freshwater. Hydrobiologia 595:545–567

Leroy, B., M. S. Dias, E. Giraud, B. Hugueny, C. Jézéquel, F. Leprieur, T. Oberdorff, and P. A.

Tedesco. 2019. Global biogeographical regions of freshwater fish species. J. Biogeogr.

46:2407–2419.

Lowe-McConnell, R. H. 1969. Speciation in tropical freshwater fishes. Biol. J. Linn. Soc. 1:51–

75.

Lynch, J. D. 1989. The gauge of speciation: on the frequencies of modes of speciation. Pp. 527–

553 in D. Otte and J. A. Endler, eds. Speciation and its consequences. Sinauer Associates,

Sunderland, MA.

Maddison, W. P., and R. G. FitzJohn. 2015. The unsolved challenge to phylogenetic correlation

tests for categorical characters. Syst. Biol. 64:127–136.

Maddison, W. P., P. E. Midford, and S. P. Otto. 2007. Estimating a binary character's effect on

speciation and extinction. Syst. Biol. 56:701–710.

Magallón, S., and M. J. Sanderson. 2001. Absolute diversification rates in angiosperm clades.

Evolution 55:1762–1780.

Manel, S., P.-E. Guerin, D. Mouillot, S. Blanchet, L. Velez, C. Albouy, and L. Pellissier. 2020.

Global determinants of freshwater and marine fish genetic diversity. Nat. Commun.

11:692.

Martin, C. H., J. S. Cutler, J. P. Friel, C. D. Touokong, G. Coop, and P. C. Wainwright. 2015.

Complex histories of repeated gene flow in Cameroon crater lake cichlids cast doubt on

one of the clearest examples of sympatric speciation. Evolution 69:1406–1422.

Matzke, N. J. 2014. Model selection in historical biogeography reveals that founder-event

speciation is a crucial process in island clades. Syst. Biol. 63:951–970.

May, R. M. 1994. Biological diversity: differences between land and sea. Phil. Trans. R. Soc.

Lond. B 343:105–111.

McCune, A. R. 1987. Lakes as laboratories of evolution endemic fishes and environmental

cyclicity. Palaios 2:446–54

McCune, A. R., and N. R. Lovejoy. 1998. The relative rate of sympatric and allopatric speciation

in fishes. Pp. 172–185 in D. J. Howard and S. H. Berlocher, eds. Endless forms: species

and speciation. Oxford Univ. Press, New York, NY.

McDowall, R. M. 1998. Phylogenetic relationships and ecomorphological divergence in

sympatric and allopatric species of *Paragalaxias* (Teleostei: Galaxiidae) in high elevation

Tasmanian lakes. Environ. Biol. Fishes 53:235–257.

McGee, M. D., S. R. Borstein, J. I. Meier, D. A. Marques, S. Mwaiko, A. Taabu, M. A. Kishe, B.

O'Meara, R. Bruggmann, L. Excoffier, et al. 2020. The ecological and genomic basis of

explosive adaptive radiation. Nature 586:75–79.

Meseguer, A. S., and F. L. Condamine. 2020. Ancient tropical extinctions at high latitudes

contributed to the latitudinal diversity gradient. Evolution 74:1966–1987.

Miller, E. C., and J. J. Wiens. 2017. Extinction and time help drive the marine-terrestrial

biodiversity gradient: is the ocean a deathtrap? Ecol. Lett. 20:911–921.

Miller, E. C., and C. Román-Palacios. 2021. Evolutionary time best explains the latitudinal

diversity gradient of living freshwater fish diversity. Global Ecol. Biogeogr. 30:749–763.

Miller, E. C., K. T. Hayashi, D. Song, and J. J. Wiens. 2018. Explaining the ocean's richest

biodiversity hotspot and global patterns of fish diversity. Proc. Roy. Soc. B.

285:20181314.

Moen, D., and H. Morlon. 2014. Why does diversification slow down? Trends Ecol. Evol.

29:190–197.

Nagelkerke, L. A. J., and F. A. Sibbing. 2000. The large barbs (*Barbus* spp., Cyprinidae,

Teleostei) of Lake Tana (Ethiopia), with a description of a new species, *Barbus osseensis*.

Neth. J. Zool. 50:179–214.

Nakov, T., J. M. Beaulieu, and A. J. Alverson. 2019. Diatoms diversity and turn over faster in

freshwater than marine environments. Evolution 73:2497–2511.

Near, T. J., and M. F. Benard. 2004. Rapid allopatric speciation in logperch darters (Percidae:

Percina). Evolution 58:2798–808.

Osinov, A. G., A. L. Senchukova, N. S. Mugue, S. D. Pavlov, and I. A. Chereshnev. 2015.

Speciation and genetic divergence of three species of charr from ancient Lake

El'gygytgyn (Chukotka) and their phylogenetic relationships with other representatives

of the genus *Salvelinus*. Biol. J. Linn. Soc. 116:63–85.

Parenti, L. R. 1984. A taxonomic revision of the Andean killifish genus *Orestias*

(Cyprinodontiformes, Cyprinodontidae). B. Am. Mus. Nat. Hist. 178:107–214.

Parenti, L. R. 2008. A phylogenetic analysis and taxonomic revision of ricefishes, *Oryzias* and

relatives (Beloniformes, Adrianichthyidae). Zool. J. Linn. Soc. 154:494–610.

Pennell, M. W., J. M. Eastman, G. J. Slater, J. W. Brown, J. C. Uyeda, R. G. FitzJohn, M. E.

Alfaro, and L. J. Harmon. 2014. Geiger v2.0: an expanded suite of methods for fitting

macroevolutionary models to phylogenetic trees. 30:2216–2218.

Poulin, R. 2016. Greater diversification of freshwater than marine parasites of fish. Int. J.

Parasitol. 46:275–279.

Rabosky, D. L. 2014. Automatic detection of key innovations, rate shifts, and diversity-

dependence on phylogenetic trees. PLoS ONE 9:e89543.

Rabosky, D. L. 2020. Speciation rate and the diversity of fishes in freshwaters and the oceans. J.

Biogeogr. 47:1207–1217.

Rabosky, D. L., and H. Huang. 2016. A robust semi-parametric test for detecting trait-dependent

diversification. Syst. Biol. 65:181–193.

Rabosky, D. L., F. Santini, J. Eastman, S. A. Smith, B. Sidlauskas, J. Chang, and M. E. Alfaro.

2013. Rates of speciation and morphological evolution are correlated across the largest

vertebrate radiation. Nat. Commun. 4:1958.

Rabosky, D. L., J. Chang, P. O. Title, P. F. Cowman, L. Sallan, M. Friedman, K. Kaschner, C.

Garilao, T. J. Near, M. Coll, et al. 2018. An inverse latitudinal gradient in speciation rate

for marine fishes. Nature 559:392– 395.

Revell, L. J. 2012. phytools: an R package for phylogenetic comparative biology (and other

things). Methods Ecol. Evol. 3:217–223.

Ricklefs, R. E. 1987. Community diversity: Relative roles of local and regional processes.

Science 235:167–171.

Rico, C., and G. F. Turner. 2002. Extreme microallopatric divergence in a cichlid species from

Lake Malawi. Mol. Ecol. 11:1585–1590.

Rüber, L., and R. Zardoya. 2005. Rapid cladogenesis in marine fishes revisited. Evolution

59:1119–1127.

Santini, F., M. T. T. Nguyen, L. Sorenson, T. B. Waltzek, J. W. Lynch Alfaro, J. M. Eastman,

and M. E. Alfaro. 2013. Do habitat shifts drive diversification in teleost fishes? An

example from the pufferfishes (Tetraodontidae). J. Evol. Biol. 26:1003–1018.

Schluter, D., and J. D. McPhail. 1992. Ecological character displacement and speciation in

sticklebacks. Am. Nat. 140:85–108.

Schluter, D., and J. D. McPhail. 1993. Character displacement and replicate adaptive radiation.

Trends Ecol. Evol. 8:197–200.

Sherbakov, D. Y. 1999. Molecular phylogenetic studies on the origin of biodiversity in Lake

Baikal. Trends Ecol. Evol. 14:92–95.

Seehausen, O. 2015. Process and pattern in cichlid radiations- inferences for understanding

unusually high rates of evolutionary diversification. New Phytol. 207:304–312.

Seehausen, O., and J. J. M. van Alphen. 1999. Can sympatric speciation by disruptive sexual

selection explain rapid evolution of cichlid diversity in Lake Victoria? Ecol. Lett. 2:262–

271.

Seehausen, O., and Wagner, C. E. (2014). Speciation in freshwater fishes. Annu. Rev. Ecol.

Evol. S. 45:621–651.

Sholihah, A., E. Delrieu-Trottin, F. L. Condamine, D. Wowor, L. Rüber, L. Pouyaud, J-F.

Agnèse, and N. Hubert. 2021. Impact of Pleistocene eustatic fluctuations on evolutionary

dynamics in Southeast Asian biodiversity hotspots. Syst. Biol., in press

(https://doi.org/10.1093/sysbio/syab006).

Siqueira, A. C., R. A. Morais, D. R. Bellwood, and P. F. Cowman. 2020. Trophic innovations

fuel reef fish diversification. Nat. Commun. 11:2669.

Skeels, A. C. 2019. Lineages through space and time plots: Visualising spatial and temporal

changes in diversity. Front. Biogeogr. 11:e42954.

Skeels, A., and M. Cardillo. 2019. Reconstructing the geography of speciation from

contemporary biodiversity data. Am. Nat. 193:240–255.

Smith, S. A., and B. C. O'Meara. 2012. treePL: divergence time estimation using penalized

likelihood for large phylogenies. Bioinformatics 20:2689–2690.

Stephens, P. R., and J. J. Wiens. 2003. Explaining species richness from continents to

communities: The time-for-speciation effect. Am. Nat. 161:112–128.

Sullivan, J. P., S. Lavoué, and C. D. Hopkins. 2002. Discovery and phylogenetic analysis of a

riverine species flock of African electric fishes (Mormyridae: Teleostei). Evolution

56:597–616.

Tedesco, P. A., E. Paradis, C. Lévêque, and B. Hugueny. 2017a. Explaining global-scale

diversification patterns in actinopterygian fishes. J. Biogeogr. 44:773–783.

Tedesco, P. A., O. Beauchard, R. Bigorne, S. Blanchet, L. Buisson, L. Conti, J. Cornu, M. S.

Dias, G. Grenouillet, B. Hugueny, et al. 2017b. A global database on freshwater fish

species occurrence in drainage basins. Sci. Data 4:170141.

Title, P. O., and D. L. Rabosky. 2019. Tip rates, phylogenies, and diversification: What are we

estimating, and how good are the estimates? Methods Ecol. Evol. 10:821–834

Toussaint, E. F. A., D. Bloom, and A. E. Short. 2017. Cretaceous West Gondwana vicariance

shaped giant water scavenger beetle biogeography. J. Biogeogr. 44:1952–1965.

Vuorinen, J. A., R. A. Bodaly, J. D. Reist, and M. Luczynski. 2005. Phylogeny of five

Prosopium species with comparisons with other Coregonine fishes based on isozyme

electrophoresis. J. Fish. Biol. 53:917–927.

Wagner, C. E., L. J. Harmon, and O. Seehausen. 2012. Ecological opportunity and sexual

selection together predict adaptive radiation. Nature 487:366–369.

Wagner, C. E., L. J. Harmon, and O. Seehausen. 2014. Cichlid species-area relationships are

shaped by adaptive radiations that scale with area. Ecol. Lett. 17:583–592.

Wiens, J. J. 2012. Why biogeography matters: Historical biogeography vs. phylogeography and

community phylogenetics for inferring ecological and evolutionary processes. Front.

Biogeogr. 4:128–135.

Wiens, J. J. 2015a. Faster diversification on land than sea helps explain global biodiversity

patterns among habitats and animal phyla. Ecol. Lett. 18:1234–1241.

Wiens, J. J. 2015b. Explaining large-scale patterns of vertebrate diversity. Biol. Lett.

11:20150506.

Yoder, J. B., E. Clancey, S. Des Roches, J. M. Eastman, L. Gentry, W. Godsoe, T. J. Hagey, D.

Jochimsen, B. P. Oswald, J. Robertson, et al. 2010. Ecological opportunity and the origin

of adaptive radiations. J. Evol. Biol. 23:1581–1596.

887
888

889　**Table 1.** Studies comparing diversification rates between (A) freshwater and marine ray-finned fishes, and (B) freshwater, marine,

890　and/or terrestrial habitats using other organisms. Here I note if the study supports the hypothesis that net diversification rates are faster

891　in freshwater than other habitats. See footnotes for method references.

Study	Clade	Method	Result	Supports hypothesis?
A: Ray-finned fishes:				
Davis et al. 2012	Terapontidae (grunters)	MEDUSA[1]	Higher freshwater net diversification rates	Yes
Carrete Vega and Wiens 2012	Ray-finned fishes	Method-of-moments[2]	No difference in net diversification rates	No
Betancur-R et al. 2012	Ariidae (sea catfishes)	GeoSSE[3]	No differences in rates	No
Bloom et al. 2013	Atherinopsidae (New World silversides)	BiSSE[4]	Higher freshwater speciation and extinction rates; little difference in net diversification rates	No
Santini et al. 2013	Tetraodontidae (puffers)	MuSSE[5]	Higher marine speciation rates	No
Betancur-R et al. 2015	Ray-finned fishes	GeoSSE	Higher marine net diversification rates	No
Guinot and Cavin 2015	Ray-finned fishes	Fossil family diversity through time	Accumulation of families has slowed through time in marine but not freshwater fishes	Unclear (see text)

Tedesco et al. 2017a	Ray-finned fishes	Method-of-moments	Higher freshwater net diversification rates	Yes
Miller et al. 2018	Percomorpha (e.g. cichlids, guppies, reef fishes)	HiSSE[6]	Slightly higher freshwater speciation, extinction and net diversification rates	Yes
Rabosky 2020	Ray-finned fishes	STRAPP[7]	Higher freshwater speciation rates driven by a few clades, but not a general pattern among fishes	Potentially
B: Other organisms:				
Wiens 2015a	All animal phyla	Method-of-moments	Higher non-marine net diversification rates	Unclear-freshwater and terrestrial combined
Wiens 2015b	Vertebrates	Method-of-moments	Higher terrestrial net diversification rates compared to aquatic habitats, but no difference between non-marine and marine habitats	Unclear-freshwater combined with other habitats
Poulin 2016	Fish parasites	Species-to-genus ratios	Higher parasite species-to-genus ratios in freshwater versus marine fishes	Yes
Miller and Wiens 2017	Amniotes	HiSSE	No consistent difference between marine and non-marine rates among clades	Unclear-freshwater and terrestrial combined

| Kirchhoff et al. 2017 | Myliobatiformes (stingrays) | ClaSSE[8] | No difference in rates between freshwater and marine habitats; brackish rates were lowest | No |
| Nakov et al. 2019 | Diatoms | HiSSE | Higher freshwater speciation, extinction, and net diversification rates | Yes |

894   1: Alfaro et al. 2009; 2: Magallón and Sanderson 2001; 3: Goldberg et al. 2011; 4: Maddison et al. 2007; 5: Fitzjohn 2012; 6: Beaulieu

895   and O'Meara 2016; 7: Rabosky and Huang 2016; 8: Goldberg and Igić 2012

Table 2. Comparing the fit of alternative biogeographic models fit using BioGeoBEARS (Matzke 2014). Models differed in the constraints set on dispersal in order to test hypotheses about biogeographic connections among lakes. Constraints were set by multiplying the probability of dispersal by 0.05 (more details in Extended Text 2 and Table S11). The best-fitting model (in bold) was used in downstream analyses (Figure 3).

Marine-lake transitions	Regional dispersal via lakes	Base model	LnL	No. params	*d*	*e*	*j*	AIC	Akaike weight
No constraints	No constraints	BAYAREA	-8746.8	2	0.0015	0.0035	-	17497.6	<0.01
		BAYAREA+J	-7317.8	3	0.0010	0.0004	0.0061	14641.7	<0.01
		DEC	-7580.9	2	0.0019	0.0001	-	15165.9	<0.01
		DEC+J	-7468.5	3	0.0016	<0.0001	0.0024	14943.0	<0.01
		DIVA	-7993.1	2	0.0020	<0.0001	-	15990.1	<0.01
		DIVA+J	-7870.4	3	0.0017	<0.0001	0.0017	15746.8	<0.01
Constrained	No constraints	BAYAREA	-8526.4	2	0.0020	0.0033	-	17056.8	<0.01
		BAYAREA+J	-7084.7	3	0.0013	0.0003	0.0076	14175.3	<0.01
		DEC	-7295.1	2	0.0024	0.0001	-	14594.3	<0.01
		DEC+J	-7186.0	3	0.0021	<0.0001	0.0032	14377.9	<0.01
		DIVA	-8740.3	2	0.0031	0.0063	-	17484.5	<0.01
		DIVA+J	-7555.9	3	0.0022	<0.0001	0.0035	15117.7	<0.01

No constraints	Always constrained	BAYAREA	-8767.7	2	0.0015	0.0035	-	17539.5	<0.01
		BAYAREA+J	-7324.8	3	0.0010	0.0004	0.0063	14655.7	<0.01
		DEC	-7599.6	2	0.0019	0.0001	-	15203.2	<0.01
		DEC+J	-7485.6	3	0.0017	<0.0001	0.0020	14977.3	<0.01
		DIVA	-8007.6	2	0.0021	<0.0001	-	16019.1	<0.01
		DIVA+J	-7868.5	3	0.0017	<0.0001	0.0030	15743.1	<0.01
No constraints	Some types constrained	BAYAREA	-8746.6	2	0.0015	0.0035	-	17497.3	<0.01
		BAYAREA+J	-7312.5	3	0.0010	0.0004	0.0062	14631.0	<0.01
		DEC	-7580.0	2	0.0019	0.0001	-	15163.9	<0.01
		DEC+J	-7466.5	3	0.0016	<0.0001	0.0024	14939.0	<0.01
		DIVA	-7991.6	2	0.0020	<0.0001	-	15987.2	<0.01
		DIVA+J	-7853.1	3	0.0017	<0.0001	0.0027	15712.2	<0.01
Constrained	Always constrained	BAYAREA	-8546.5	2	0.0020	0.0033	-	17096.9	<0.01
		BAYAREA+J	-7086.3	3	0.0014	0.0003	0.0080	14178.6	<0.01
		DEC	-7318.8	2	0.0027	<0.0001	-	14641.6	<0.01
		DEC+J	-7198.6	3	0.0022	<0.0001	0.0027	14403.1	<0.01
		DIVA	-8741.0	2	0.0031	0.0062	-	17486.1	<0.01
		DIVA+J	-7564.1	3	0.0023	<0.0001	0.0037	15134.2	<0.01
Constrained	**Some types constrained**	BAYAREA	-8526.1	2	0.0020	0.0033	-	17056.2	<0.01
		BAYAREA+J	**-7077.1**	**3**	**0.0013**	**0.0003**	**0.0078**	**14160.2**	**1.00**

DEC	-7293.0	2	0.0024	0.0001	-	14590.0	<0.01
DEC+J	-7183.9	3	0.0022	<0.0001	0.0029	14373.9	<0.01
DIVA	-8732.9	2	0.0031	0.0063	-	17469.8	<0.01
DIVA+J	-7550.6	3	0.0022	<0.0001	0.0036	15107.2	<0.01

Table 3: Summary of 56 phylogenetic ANOVA and post hoc tests comparing diversification rates (seven types) among habitat categories (four habitat combinations with and without cichlids). Rates were calculated by Rabosky et al. (2018; see also Chang et al. 2019). Full results including post hoc tests are reported in Tables S2–S8. Six habitats=marine, estuarine, diadromous, riverine, lacustrine, both riverine and lacustrine. Four habitats=marine, riverine, lacustrine, both riverine and lacustrine. Three habitats=riverine, lacustrine, both riverine and lacustrine. Two habitats=marine and freshwater. Boldface=ANOVA was significant. *=Lakes were significantly different from all other habitats in pairwise post hoc tests, even if the ANOVA was not globally significant.

Rate type	All teleosts, P-value				Teleosts excluding Cichlidae, P-value			
	Six habitats	Four habitats	Three habitats	Two habitats	Six habitats	Four habitats	Three habitats	Two habitats
DR	0.093*	0.121*	**0.001***	0.653	0.464*	0.468*	**0.001***	0.789
BAMM, speciation, time-varying	0.062*	0.071*	**0.001***	0.625	0.428*	0.448*	**0.001***	0.789
BAMM, speciation, time-constant	0.059*	0.071*	**0.001***	0.537	0.418*	0.417*	**0.001***	0.747

BAMM, extinction, time-varying	0.298	0.344	0.061	0.727	0.355	0.487*	**0.001***	0.832
BAMM, extinction, time-constant	0.270	0.266	**0.038**	0.635	0.379	0.460*	**0.001***	0.748
BAMM, net diversification, time-varying	**0.047***	0.071*	**0.001***	0.624	0.555*	0.546*	**0.001***	0.811
BAMM, net diversification, time-constant	**0.041***	0.056*	**0.001***	0.597	0.562*	0.561*	**0.001***	0.793

917 **Table 4.** Examples of lacustrine radiations other than cichlids. Note that this list is not exhaustive. These radiations represent

918 independent transitions to the lacustrine state based on biogeographic reconstructions (Fig. 3). Radiations are ranked by diversification

919 rate in descending order. The mean DR represents the mean tip-associated diversification rate (species/million years) of members of

920 the lacustrine lineage identified by BioGeoBEARS (Fig. 3D). Taxonomy follows Rabosky et al. (2018). Here I note whether

921 monophyletic clade(s) are found within individual lakes (intralacustrine radiations) based on prior phylogenetic studies.

922
923

Radiation	Family	Region	Lake system	Mean DR	Intralacustrine radiation(s)?	References
Labeobarbus	Cyprinidae	Ethiopian	Lake Tana, Ethiopia	4.08 ± 2.39	yes	Nagelkerke and Sibbing 2000
Coregonus	Salmonidae	Nearctic	North American postglacial lakes	2.67 ± 1.16	no	Hudson et al. 2007
Coregonus	Salmonidae	Palearctic	Eurasian postglacial lakes	2.09 ± 2.4	some members	Hudson et al. 2007; Doenz et al. 2018
Salvelinus and *Salvethymus*	Salmonidae	Palearctic	Lake El'gygytgyn and surrounding basins, Russia	1.06 ± 0.73	no	Osinov et al. 2015
Bathyclarias	Clariidae	Ethiopian	Lake Malawi, East Africa	0.96 ± 0.04	yes	Anseaume and Teugels 2005
Lake Baikal sculpins	Cottidae	Palearctic	Lake Baikal, Russia	0.71 ± 0.4	yes	Cohen 1995; Sherbakov 1999

Orestias	Cyprinodontidae	Neotropics	Lake Titicaca and surrounding basins, Peru/Bolivia	0.6 ± 0.42	no	Parenti 1984
Synodontis	Mochokidae	Ethiopian	Lake Tanganyika, East Africa	0.54 ± 0.23	yes	Day et al. 2009
Poblana	Atherinopsidae	Neotropics	Central Mexican Plateau	0.38 ± 0.12	some members	Bloom et al. 2013
Sulawesi telmatherinid rainbowfishes	Telmatherinidae	Sino-Oriental	Malili Lake system, Sulawesi	0.31 ± 0.16	some members	Herder et al. 2006
Prosopium	Salmonidae	Nearctic	Bear Lake, Idaho/Utah, U.S.A.	0.22 ± 0.07	yes	Vuorinen et al. 1998; Hudson et al. 2007
Mastacembelus	Mastacembelidae	Ethiopian	Lake Tanganyika, East Africa	0.19 ± 0.07	yes	Brown et al. 2010
Sarcocheilichthys	Cyprinidae	Sino-Oriental	Lake Biwa, Japan	0.15 ± 0.01	yes	Hauffe et al. 2020
Oryzias	Adrianichthyidae	Sino-Oriental	Malili Lake system, Sulawesi	0.12 ± 0.08	no	Parenti 2008
Paragalaxias	Galaxiidae	Australian	Central Plateau of Tasmania	0.10 ± 0.03	no	McDowall 1998

924
925
926

 FIGURE LEGENDS

Figure 1. Diversity and diversification rates in aquatic habitats. (A) Proportion of teleost species richness in each habitat (n=11,227

species). (B) Tip-associated net diversification rates in species found exclusively in marine or freshwater habitats (n=10,177 species).

(C) Tip-associated speciation, extinction, and net diversification rates among six aquatic habitats. Boxplots summarize rates for

individual species, including medians, first and third quartiles, and outliers (outside the 95% confidence interval). Mean rates are

printed below each habitat. All rates shown here were calculated using BAMM under a time-varying model of diversification rates

(Rabosky et al. 2018; Chang et al. 2019). Results were similar using other methods of calculating diversification rates (Table 3; Table

S2–S8).

Figure 2. Diversification rate variation by habitat using alternative divergence time estimates and clade selection. All rates shown

were calculated using the DR statistic (full results in Tables S2–S10). Boxplots summarize rates for individual species, including

medians, first and third quartiles, and outliers (outside the 95% confidence interval). Left panels compare rates in species exclusive to

marine and freshwater habitats; right panels compare rates among six aquatic habitats. Top panels compare rates using all members of

the clade; right panels compare rates after excluding the family Cichlidae. For teleosts, red bars represent rates calculated from the

Rabosky et al. (2018) phylogeny redated using Hughes et al. (2018) as a reference; green bars represent rates using the original tree.

For acanthomorphs (spiny-rayed fishes), blue bars represent the Rabosky et al. (2018) phylogeny redated using Alfaro et al. (2018) as

a reference; purple bars represent rates using the original tree (Acanthomorpha alone). Fish icons from PhyloPic (http://phylopic.org/)

with credit to Lily Hughes, Tyler McCraney, and Milton Tan.

Figure 3. Results of biogeographic and habitat modelling. In panels A and B, biogeographic regions were combined to illustrate

habitat transitions overall. In C and D, habitats are shown separately for each region. (A) Ancestral habitats based on best-fit model

(Table 2). Here, light grey=marine, blue=riverine, red=lacustrine, dark grey=both riverine and lacustrine, and black=uncertain (no

habitat with probability >80%). (B) Transition rates among habitats. Black arrows are sized to scale of transition rates. Number is the

mean transition rate calculated from 100 biogeographic stochastic maps. (C) Lineages-through-space-and-time plot (Skeels 2019) of

total riverine and lacustrine diversity within six biogeographic regions. The width of lines represents the range of values observed

among 100 stochastic maps. Bioregionalization follows Leroy et al. (2019). (D) Properties of lineages representing independent

habitat transitions by region. From top to bottom: number of independent transitions into that habitat, mean number of species

descending from transitions, mean time of transitions, and mean tip-associated diversification rate (DR) of descendants. Points show

the mean and range of mean values among 100 stochastic maps.

Comparing two habitats:

Comparing six habitats:

All species

Inset right:

Excluding

diversification rate (DR)

marine   freshwater

marine   brackish   diadromous   riverine   both R+L   lacustrine

Teleostei:
- redated using Hughes et al. (2018)
- original dates

Acanthomorpha:
- redated using Alfaro et al. (2018)
- original dates