# 1 Running head: Indigenous plants promote urban insect diversity

# 2 Title: Indigenous plants promote insect biodiversity in urban greenspaces

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- 34 Abstract (max 350 words)

35	The contribution of urban greenspaces to support biodiversity and provide benefits for
36	people is increasingly recognised. However, ongoing management practices still
37	favour (1) vegetation oversimplification, often limiting greenspaces to lawns and tree
38	canopy rather than multi-layered vegetation that includes under and midstorey; and
39	(2) the use of nonnative plant species. These practices likely hinder the potential of
40	greenspaces to sustain indigenous biodiversity, particularly for taxa like insects, that
41	rely on plants for food and habitat. Yet, little is known about which plant species may
42	maximise positive outcomes for taxonomically and functionally diverse insect
43	communities in urban greenspaces. Additionally, while urban environments are
44	expected to experience high rates of introductions, quantitative assessments of the
45	relative occupancy of indigenous vs. introduced insect species in greenspace are rare
46	- hindering understanding of how greenspace management may promote indigenous
47	biodiversity while limiting the establishment of introduced insects. Using a
48	hierarchically replicated study design across 15 public parks, we recorded occurrence
49	data from 552 insect species on 133 plant species – differing in planting design

50	element (lawn, midstorey and tree canopy), midstorey growth form (forbs, lilioids,
51	graminoids and shrubs) and origin (nonnative, native and indigenous) - to assess: (1)
52	the relative contributions of indigenous and introduced insect species and (2) which
53	plant species sustained the highest number of indigenous insects. Our data indicates
54	that the insect community was predominately composed of indigenous rather than
55	introduced species. Our findings further highlight the core role of multi-layered
56	vegetation in sustaining high insect biodiversity in urban areas, with indigenous
57	midstorey and canopy representing key elements to maintain rich and functionally
58	diverse indigenous insect communities. Intriguingly, graminoids supported the
59	highest indigenous insect richness across all studied growth forms by plant origin
60	groups. Taken together, our study emphasise the opportunity posed by indigenous
61	understory and midstorey plants, particularly indigenous graminoids in our study area,
62	to promote indigenous insect biodiversity in urban greenspaces. Our work provides a
63	blueprint and stimulus for built-environment professionals to incorporate into their
64	practice plant species palettes that foster a larger presence of indigenous over
65	regionally native or nonnative plant species, whilst incorporating a broader mixture of
66	midstorey growth forms.
67	Key words: Bayesian inference, Greenspace design and management, Invertebrates,
68	Nature in cities, Plant-insect metanetwork, Species-specific responses, Urban ecology
69	Introduction
70	Urban greenspaces provide well-documented benefits for biodiversity and people.
71	Remnant bushland, parks, gardens, golf courses, greenroofs, pop-up parks and other
72	types of greenspace support a great diversity of microbial, fungal, plant and animal
73	species (Madre et al. 2013, Aronson et al. 2014, Baldock et al. 2015, Beninde et al.
74	2015, McGregor-Fors et al. 2016, Mata et al. 2017, Threlfall et al. 2017, Baldock et

75	al. 2019, Mata et al. 2019), and provide a diverse array of health, mental, cognitive,
76	social, cultural and spiritual benefits for people who interact with them (Keniger et al.
77	2013, Dadvand et al. 2015, Hartig and Kahn 2016, Flies et al. 2017, Maller et al.
78	2018, Lai et al. 2019, Mata et al. 2020). Hence, researchers, practitioners, built-
79	environment professionals and policymakers are increasingly working together to
80	promote the positive socio-ecological outcomes of greenspaces (Aronson et al. 2017,
81	Lepczyk et al. 2017, Nilon et al. 2017, Parris et al. 2018, Soanes et al. 2019). Further,
82	the importance of greenspaces has been recently highlighted at an international policy
83	level with the United Nations' New Urban Agenda committing to "promoting the
84	creation and maintenance of well-connected and well-distributed networks of open,
85	multipurpose, safe, inclusive, accessible, green and quality public spaces" (United
86	Nations 2017).
87	An ubiquitous practice that can hinder the potential of greenspaces to support
88	biodiversity is the oversimplification of vegetation structure (Le Roux et al. 2014,
89	Threlfall et al. 2016), which has led many greenspaces to be vegetated by only two
90	planting design elements: lawn and tree canopy (Ignatieva et al. 2015, Aronson et al.
91	2017). In contrast, greenspaces with a more complex, multi-layered vertical structure
92	- those including understorey and midstorey vegetation (henceforth midstorey for
93	brevity) – provide positive outcomes for a taxonomically and functionally diverse
94	range of taxa (Beninde et al. 2015, Mata et al. 2017, Threlfall et al. 2017, Majewska
95	and Altizer 2020). Unlike the lawn and tree canopy, the midstorey is a heterogenous
96	mix of different plant growth forms, including forbs, graminoids, lilioids and shrubs
97	amongst others. Yet, at present, the combination of plant species and growth forms
98	that maximise positive outcomes for non-plant species in the midstorey remains
99	poorly understood.

100	An additional issue that limits biodiversity in urban areas is that most non-remnant
101	greenspaces, particularly intensively manicured ones such as residential gardens and
102	public parks, are composed predominately of nonnative plant species (Threlfall et al.
103	2016). Nonnative plants are rarely well-suited to provide resources for indigenous
104	primary consumers (e.g. herbivorous insects and frugivorous birds), nor to indigenous
105	secondary and apex consumers (e.g. predatory and parasitoid insects and
106	insectivorous birds and bats) that depend on primary consumers as food resources
107	(Ballard et al. 2013, Burghardt and Tallamy 2013, Ikin et al. 2013, Salisbury et al.
108	2015, Threlfall et al. 2017). These studies highlight how management practices that
109	promote the use of nonnative plants are likely to reduce the capacity of greenspaces to
110	sustain diverse communities of indigenous biodiversity.
111	In most of the studies to date, plant origin has been treated as an aggregate plot-level
112	explanatory variable (e.g. treatment plot as either nonnative or native; plot nativeness
113	as percentage cover of native vegetation). A focus on plant species rather than plot as
114	the unit of analysis allows for a more nuanced understanding of how plant origin may
115	influence the capacity of plants to provide resources for associated consumer species.
116	Moreover, focusing on the plant species level may advance understanding of how
117	plant origin interacts with other plant-level attributes — such as planting design
118	element and growth form — to produce positive outcomes for consumer species,
119	particularly of taxa that rely extensively on plants for food and habitat resources, such
120	as insects.
121	Insects are a particularly important component of urban biodiversity (Sattler et al.
122	2011, New 2015) and the ecological functions they perform provide numerous
123	benefits to urban residents (Prather et al. 2013, Benett and Lovell 2014, Baldock et al.
124	2015), along with some disbenefits (Dunn 2010, Rust and Su 2012). Plants and

125	insects have often coevolved in close association with each other, with many insect
126	species showing high levels of specialisation (Forister et al. 2012). Plants also provide
127	food, foraging, nesting, oviposition, shelter and overwintering resources to insect
128	detritivores, predators and parasitoids; indeed, practices that promote these resources
129	in agroecosystems by fostering plant diversity and structural complexity are key
130	components of pest management strategies (Landis et al. 2000). The availability of
131	suitable host plants within greenspaces is therefore a key determinant of insect
132	diversity in urban environments (Aronson et al. 2016). However, as far as we are
133	aware no study has sought to identify specific plant species of differing origins and
134	growth forms that promote taxonomically and functionally diverse insect
135	communities in urban greenspaces.
136	Urban environments are central nodes of human-mediated dispersal networks
137	(Bullock et al. 2018) and hotspots of novel resource utilisation (Valentine et al. 2020),
138	and introduced species are therefore often prominent (Cadotte et al. 2017, Paap et al.
139	2017). The number of introduced insect species can be relatively low (Madre et al.
140	2013, Mata et al. 2017), but they often occur in high abundance and this is especially
141	the case for generalist bees and butterflies (Matteson & Langellotto 2010, Threlfall et
142	al. 2015). However, as far as we are aware, no quantitative assessments of the relative
143	contributions of indigenous and introduced species in greenspace insect communities
144	have been reported in the literature. This paucity of data hinders understanding of
145	how greenspaces may be managed to promote indigenous insect biodiversity while
146	limiting the establishment of introduced species.
147	Here we use a plant-insect metanetwork dataset collected across 15 greenspaces
148	within a densely urbanised inner-city municipality to assess: (1) the relative
149	contributions of indigenous and introduced species in insect communities of urban

150	environments, and (2) which plant species should be planted to support indigenous
151	insect species. Th project followed the science-government partnerships model (Ives
152	and Lynch 2014) – an approach that advocates for industry professionals and
153	researchers to work in close association to guarantee that theoretically interesting and
154	practically important questions are identified. This ensured that the implications of
155	our research findings could be applied as practical on-ground actions that were
156	embedded into a new business as usual at the City of Melbourne. We began by
157	determining what proportion of insect species occurring in the greenspaces are
158	indigenous, and whether indigenous insect species are more common than introduced
159	species. We then examined how the richness of indigenous insect species varies with
160	planting design element (lawn, midstorey and tree canopy), midstorey growth form
161	(forbs, lilioids, graminoids and shrubs) and plant origin groups (nonnative, native and
162	indigenous). Finally, we grouped plants species according to a combination of
163	planting design element by plant origin (Fig. 1a), and midstorey growth form by plant
164	origin (Fig. 1b) to assess how the richness of indigenous insect species vary amongst
165	these groups. We do this assessment for the whole insect community and for five key
166	functional groups: pollinators and other flower-visiting taxa, herbivores, predators,
167	parasitoids and detritivores. We also examined variation amongst the plant groups in
168	insect species composition and number of unique species.
169	Methods

170 Study design

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171 The study was conducted in the City of Melbourne, Australia. This is one of 31

172 municipalities within Greater Melbourne, a large metropolitan area spanning

approximately  $10,000 \text{ km}^2$  within the Urban Growth Boundary and home to over five

174 million people (Victorian State Revenue Office: https://www.sro.vic.gov.au/greater-

175	melbourne-map-and-urban-zones). The City of Melbourne covers 37.7 km <sup>2</sup> and
176	incorporates a central business district, transport and distribution hubs. Approximately
177	13% of the land area is covered by vegetated open space, including grassy woodlands,
178	wetlands, estuaries and a greenspace network of parks, gardens and streetscapes. The
179	municipality is home to approximately 180,000 residents and receives approximately
180	900,000 daily visitors (City of Melbourne: https://www.melbourne.vic.gov.au/).
181	Our study was conducted across 15 public parks, which varied in size across four
182	orders of magnitude $(1.1 \times 10^3 \text{ m}^2 - 1.3 \times 10^6 \text{ m}^2)$ ; Appendix 1: Table S1; Appendix 1:
183	Fig. S1a). We established a total of 130 plots across the 15 parks, with the number of
184	plots in each park (2-36), and their size (84-148 m <sup>2</sup> ), varying according to the park's
185	area (Appendix S1: Eq. S1-S3) and planting design elements and midstorey growth
186	forms present (Appendix 1: Table S1; Appendix 1: Fig. S1b). Within each plot we
187	identified all plants, totalling 133 species, genera or species complexes across the
188	study area (Appendix S2: Table S2).
189	We classified plant species by origin as indigenous (n=30), regionally native (n=9)
190	and nonnative (n=94). We define indigenous plant species – also referred to in the
191	literature as locally native – as those that are native to the local bioregions. For this
192	study, indigenous plant species are those that occurred before European settlement in
193	the Volcanic Victorian Plain and Gippsland Plain bioregions, (State of Victoria:
194	https://www.environment.vic.gov.au). Regionally native (henceforth native) are
195	species that are native to Australia but not to the local bioregions and have been
196	anthropogenically introduced. Nonnative species are those that have been introduced
197	to Australia.
198	We also classified plant species by planting design element (henceforth design
199	element) as lawn complex (n=41), midstorey (n=67) and tree canopy (n=25) (Fig. 1a).

200	Midstorey taxa were further stratified by growth form as forb (n=8), lilioid (n=13),
201	graminoid (n=8) and shrub (n=38) (Fig. 1b). We define lawn complexes (henceforth
202	lawns) as patches dominated by turf forming grasses (Poaceae) intermixed with one
203	or more small, ruderal herbaceous species. Midstorey species included broad-leaved
204	perennial and annual herbaceous plants (forbs); petaloid monocots in orders Liliales
205	and Asparagales (lilioids); grasses, sedges and rushes of typically vertical habit with
206	linear foliage and inconspicuous wind pollinated flowers (graminoids); and woody
207	perennials with multiple stems and $< 5$ m in height (shrubs). Tree canopy species
208	included single-stemmed woody plants $> 5$ m in height.
209	Insect survey
210	We sampled plant species for 12 insect groups known to dominate insect communities
211	on above-ground vegetation: ants, bees, beetles, cicadas, flies, heteropteran bugs,
212	jumping plant lice, leaf- and treehoppers, parasitoid wasps, planthoppers, sawflies and
213	stinging wasps. Samples were taken by direct observation and by sweeping with an
214	entomological net. Observation time and sweeps per plant species were standardised
215	as a proportion of the plant species' volume within the plot (Appendix S1: Eq. S4-
216	S5), with each plant species in each plot sampled three times from January (summer)
217	to late March (autumn) 2015. Sampled insect specimens were processed in the
218	laboratory and identified to species/morphospecies. We assigned these as (1) either
219	indigenous to the studied bioregions and/or native to Australia (henceforth
220	indigenous) or introduced to Australia, and (2) one or more of the following
221	functional groups: pollinators and other flower-visiting taxa (henceforth pollinators),
222	herbivores, predators, parasitoids and detritivores.
223	Data analysis

224 Estimating insect occupancy and species richness per plant species

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225	To assess the proportion of indigenous insect species occurring in the greenspaces,
226	whether they were more common than introduced ones, and how their species
227	richness varied amongst the studied single and combined plant groups, we analysed
228	our data with a three-level hierarchical metacommunity occupancy model (Kéry and
229	Royle 2016). Plant species was our unit of analysis for drawing inferences on insect
230	species occupancy, and each repeated spatial (individuals of the same plant species
231	sampled in different plots) and temporal (same individual of a given plant species
232	sampled at different times) samplings constituted the unit of detection replication. We
233	structured the model around three levels: one for species occupancy; a second for
234	species detectability; and a third to treat the occupancy and detection parameters for
235	each species as random effects (Kéry and Royle 2016). Specifically, we used a
236	variation of the model described by Mata et al. (2017), in which we specified the
237	occupancy level model as:
238	$Z_{i,j} \sim \text{Bernoulli}(\Psi_{i,j})$
239	where $\Psi_{i,j}$ is the probability that insect species <i>i</i> occurs at plant species <i>j</i> , and the
240	detection level model as:
241	$\mathbf{y}_{i,j,k} \sim \text{Bernoulli} \left( \Phi_{i,j,k} \bullet Z_{i,j} \right)$
242	where $\Phi_{i,j,k}$ is the detection probability of insect species <i>i</i> at plant species <i>j</i> at
243	spatiotemporal replicate k.
244	The occupancy and detection level linear predictors were specified on the logit-
245	probability scale as:
246	logit ( $\Psi_{i,j}$ ) = occ <sub>i</sub>
247	$logit (\Phi_{i,j,k}) = det_i$

248 where  $occ_i$  and  $det_i$  are the species-specific random effects, which were specified as:

249 occ<sub>*i*</sub> ~ Normal ( $\mu$ .occ<sub>*Int*...*Ind*,  $\tau$ .occ)</sub>

### 250 $det_i \sim Normal (\mu.det, \tau.det)$

- 251 where the metacommunity mean occupancy hyperpriors for introduced and
- indigenous insect species,  $\mu$ .occ<sub>Int</sub> and  $\mu$ .occ<sub>Ind</sub>, respectively, and the metacommunity
- 253 mean detection hyperprior  $\mu$ .det, were specified as Uniform (0, 1); and the
- 254 metacommunity precision occupancy and detection hyperpriors,  $\tau$ .occ and  $\tau$ .det,
- respectively, were specified as Gamma (0.1, 0.1).
- 256 We then use the latent occurrence matrix  $Z_{ij}$  to estimate the insect species richness
- associated with each plant species  $SR_i$  through the summation:

$$SR_j = \sum_{i=S_{ij}} Z_{ij}$$

258 where  $S_{ii}$  is a 'specificity' vector indexing the insect species to be included in each 259 plant species' estimate.  $SR_i$  is then an estimate that accounts for plant-insect 260 specificity, in which, for each plant species, the observed insect species are included 261 with probability of occurrence = 1 and a limited random sub-sample of other insect 262 species occurring in the study area are included with their 0 < Z < 1 estimated 263 probabilities of occurrence. This allowed us to work within the reasonable ecological 264 assumption that across the study area not every insect species will be associated with 265 every co-occurring plant species. We conducted these estimations for the insect 266 community as a whole but also independently for introduced and indigenous species. 267 We estimated model parameters under Bayesian inference, using Markov Chain 268 Monte Carlo (MCMC) simulations to draw samples from the parameters' posterior 269 distributions. As the species richness calculations were conducted within this 270 modelling framework, we were able to derive the insect species per plant species 271 estimates with their full associated uncertainties. This allowed us to average the

272 species richness estimates of plant species belonging to the same group, and therefore 273 obtain posterior distributions for each group that we could statistically compare. 274 Our model was implemented in JAGS (Plummer 2003) and accessed through the R 275 package *jagsUI* (Kellner 2016). We used three chains of 5,000 iterations, discarding 276 the first 500 in each chain as burn-in. We visually inspected the MCMC chains and 277 the values of the Gelman-Rubin statistic to verify acceptable convergence levels of R-278 hat < 1.1 (Gelman & Hill 2007). 279 *Community dissimilarity* 280 To determine whether the composition of insect species varied amongst the design 281 elements/growth form by plant origin groups, we reorganised the data into insect-by-282 plant species matrices – cell values summarising the number of times a given insect 283 species was sampled on a given plant species across its spatiotemporal replicates – 284 and used these to calculate amongst-group community dissimilarity. Specifically, we 285 used 1 - Jaccard similarity index as implemented in the R package vegan (Oksanen et 286 al. 2016). We further used the data to create insect species lists for each group, which 287 we partitioned into their corresponding Venn sets with the R package VennDiagram 288 (Chen 2016). This allowed us to calculate the number of unique insect species – 289 species found exclusively at a given plant group and not shared with any of the other 290 groups – belonging to each group. 291 Results 292 Our survey recorded 552 insect species, with the richest taxa being beetles (125 293 species), parasitoid wasps (121), flies (101), heteropteran bugs (61), leaf- and 294 treehoppers (40) and jumping plant lice (31) (Appendix S2: Table S1). These

- represented 154 pollinator, 299 herbivore, 231 predator, 150 parasitoid and 231
- 296 detritivore species. The most commonly occurring species was the minute brown

297 scavenger beetle Cortinicara sp. 1 (Latridiidae), an indigenous detritivore species that 298 accounted for 12% of all records. The Argentine ant Linepithema humile was the most 299 frequently occurring introduced species, accounting for approximately 3% of all 300 records. Four new species were also discovered: one ant, one heteropteran bug and 301 two jumping plant lice (Mata et al. 2015, 2016). 302 Indigenous vs introduced insect species 303 There were approximately 30 times more indigenous (534) than introduced (18) insect 304 species across the study area, with our model estimates indicating that any particular 305 plant species was associated with 19 times more indigenous than introduced insect 306 species (Fig. 2a; Appendix S2: Table S2). The mean number of introduced insect 307 species found on individual plant species varied from zero to four; whereas the mean 308 number of indigenous insect species varied from one to 109, with most plant species 309 being associated with more than ten indigenous insect species (Fig. 2b; Appendix 2: 310 Table S3; Appendix 2: Figure S1). The probability of occurrence of any particular 311 insect species at a given plant across the study area was similarly low for introduced 312 and indigenous species (Fig. 2c; Appendix 2: Table S4). The introduced insect fauna 313 was represented by species showing moderate to very low (0.6 > Pocc > 0) species-314 specific probabilities of occurrence (Fig. 2d; Appendix 2: Table S1); whereas the 315 species-specific probabilities of occurrence of indigenous insect species varied 316 widely, with a few species showing very high occupancy levels (Pocc > 0.8) and most 317 species showing low occupancy levels (Pocc < 0.4) (Fig. 2d; Appendix 2: Table S1). 318 The probability of detecting any particular insect species at a given plant across the 319 study area was similarly very low for introduced and indigenous species (Pdet < 0.03; 320 Appendix S2: Table S4), and the species-specific probabilities of detection were 321 consistently low for the large majority of insect species (Appendix S2: Table S1).

# 322 Effect of design element, growth form and plant origin

323	Our model estimates indicate that all three design elements had different levels of
324	indigenous insect species richness, with the average midstorey or tree canopy species
325	showing approximately 2.5 times more insect species than the average lawn complex
326	(Fig. 3a; Appendix S2: Table S5). Likewise, we found that the species richness of
327	indigenous insects varied amongst midstorey growth forms, with the average
328	graminoid species showing 2.9 times more insect species than the average lilioid, 2.4
329	times more than the average forb and 1.8 times more than the average shrub (Fig. 3b;
330	Appendix S2: Table S5). Our estimates further indicate marked statistical differences
331	with plant origin in the number of indigenous insect species (Appendix 2: Figure S1),
332	with the average indigenous plant species showing 2.9 and 1.6 times more insect
333	species than the average nonnative and native plant, respectively; and the average
334	native plant showing 1.9 times more insect species than the average nonnative plant
335	(Fig. 3c; Appendix S2: Table S5).
335 336	<ul><li>(Fig. 3c; Appendix S2: Table S5).</li><li><i>Combined effect of design element and plant origin</i></li></ul>
336	Combined effect of design element and plant origin
336 337	Combined effect of design element and plant origin Our model estimates indicate that all design element by plant origin groups had
336 337 338	Combined effect of design element and plant origin Our model estimates indicate that all design element by plant origin groups had different levels of indigenous insect species richness. In general, indigenous groups
336 337 338 339	Combined effect of design element and plant origin Our model estimates indicate that all design element by plant origin groups had different levels of indigenous insect species richness. In general, indigenous groups were associated with higher species richness than native and these with higher
<ul><li>336</li><li>337</li><li>338</li><li>339</li><li>340</li></ul>	Combined effect of design element and plant origin Our model estimates indicate that all design element by plant origin groups had different levels of indigenous insect species richness. In general, indigenous groups were associated with higher species richness than native and these with higher richness than nonnative. The single exception was native midstorey and nonnative
336 337 338 339 340 341	Combined effect of design element and plant origin Our model estimates indicate that all design element by plant origin groups had different levels of indigenous insect species richness. In general, indigenous groups were associated with higher species richness than native and these with higher richness than nonnative. The single exception was native midstorey and nonnative tree canopy, which showed insect species richness levels that were not statistically
<ul> <li>336</li> <li>337</li> <li>338</li> <li>339</li> <li>340</li> <li>341</li> <li>342</li> </ul>	Combined effect of design element and plant origin Our model estimates indicate that all design element by plant origin groups had different levels of indigenous insect species richness. In general, indigenous groups were associated with higher species richness than native and these with higher richness than nonnative. The single exception was native midstorey and nonnative tree canopy, which showed insect species richness levels that were not statistically different from each other (Fig. 4a; Appendix S2: Table S6). Overall, the indigenous
<ul> <li>336</li> <li>337</li> <li>338</li> <li>339</li> <li>340</li> <li>341</li> <li>342</li> <li>343</li> </ul>	Combined effect of design element and plant origin Our model estimates indicate that all design element by plant origin groups had different levels of indigenous insect species richness. In general, indigenous groups were associated with higher species richness than native and these with higher richness than nonnative. The single exception was native midstorey and nonnative tree canopy, which showed insect species richness levels that were not statistically different from each other (Fig. 4a; Appendix S2: Table S6). Overall, the indigenous midstorey was associated with the highest level of indigenous insect species richness,

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347	second highest level of insect species richness, with the average plant species in this
348	group showing 1.8 and 1.4 times more insect species than the average nonnative and
349	native tree canopy plant species, respectively (Fig. 4a; Appendix S2: Table S6).
350	Across insect functional groups, lawns showed the lowest insect species richness (Fig.
351	5a,c,e,g,i; Appendix S2: Table S6). The indigenous midstorey showed the highest
352	number of indigenous insect species; however, for predators and detritivores the
353	indigenous midstorey was not statistically different to the indigenous tree canopy
354	(Fig. 5e,i; Appendix S2: Table S6). Other departures from the general pattern were
355	observed for each functional group. The indigenous and native tree canopy groups did
356	not show different levels of indigenous pollinators or parasitoid species (Fig. 5a,g;
357	Appendix S2: Table S6), and the native tree canopy and midstorey groups did not
358	show different levels of indigenous herbivore species (Fig. 5c; Appendix S2: Table
359	S6). For predators, parasitoids and detritivores, the nonnative tree canopy group has a
360	higher species richness than the native midstorey and was not statistically different
361	than the native tree canopy – the native and nonnative midstorey groups in turn did
362	not show different levels of associated species (Fig. 5e,g,i; Appendix S2: Table S6).
363	Combined effect of growth form and plant origin
364	Our model estimates indicate that the species richness of indigenous insects varied
365	amongst the growth form by plant origin groups. In general, indigenous groups had
366	higher insect species richness than did native and these had higher insect richness than
367	nonnative (Fig. 4b; Appendix S2: Table S6). The group with the highest insect species

- 368 richness was indigenous graminoids, with the average plant species in this group
- 369 showing nearly five times more species than the average nonnative graminoid (Fig.
- 370 4b; Appendix S2: Table S6). Indigenous shrubs were associated with the second

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371 highest level of insect species richness, with the average indigenous shrub showing

372 2.1 and 1.6 times more insect species than average nonnative and native shrubs, 373 respectively (Fig. 4b; Appendix S2: Table S6). The group with the third highest insect 374 species richness was indigenous lilioids, with the average indigenous lilioid showing 375 2.5 times more insect species than the average nonnative lilioid (Fig. 4b; Appendix 376 S2: Table S6). 377 Indigenous graminoids showed the highest number of indigenous insect species 378 across all insect functional groups (Fig. 5b,d,f,h,j; Appendix S2: Table S6). In 379 general, the indigenous growth form groups were associated with the highest number 380 of indigenous insect species across all insect functional groups (Fig. 5b,d,f,h,j; 381 Appendix S2: Table S6). 382 *Community composition and unique species* 383 Indigenous insect community composition varied markedly across the design 384 elements by plant origin (Fig. 6a) and growth forms by plant origin (Fig. 6b) groups. 385 The insect composition of a few group pairs was markedly similar such as between 386 indigenous and native midstorey (5%; Fig. 6a) and between nonnative graminoids and 387 shrubs (5%; Fig. 6b). However, most group pairs showed moderate to high 388 dissimilarity (> 20%) in species composition, for example, between native and 389 nonnative tree canopy (32%; Fig. 6a) and between nonnative forbs and lilioids (40%; 390 Fig. 6b). 391 The number of unique insect species also varied substantially across groups (Fig. 6). 392 Within the design element by plant origin group, up to 33% of all indigenous insect 393 species recorded in the study were unique to the indigenous groups followed by their 394 nonnative and native counterparts, with 12% and 6% unique species, respectively 395 (Fig. 6a). From the design element perspective, 30% of all indigenous insect species 396 recorded in the study were unique to the midstorey groups, followed by their tree

397	canopy and lawn counterparts, with 19% and 2% unique species, respectively (Fig.
398	6a). Similarly, within the growth form by plant origin group, as much as 29% of all
399	indigenous insect species recorded in the midstorey were unique to the three
400	indigenous groups, followed by their nonnative and native counterparts, with 11% and
401	3% unique species, respectively (Fig. 6b). From the growth form perspective, 24% of
402	all indigenous insect species recorded in the midstorey were unique to the shrub
403	groups, followed by the graminoid, forb and lilioid groups, with 14%, 3% and 2%
404	unique species, respectively (Fig. 6b).
405	Discussion
406	Our findings demonstrate that taxonomically and functionally diverse indigenous
407	insect communities occur in greenspaces in densely urbanised inner-city
408	municipalities such as the City of Melbourne, with the potential to boost ecosystem
409	multifunctionality (Soliveres et al. 2016) and biotic resistance against the
410	establishment of introduced insects (Kennedy et al. 2002). We have shown that insect
411	communities in Melbourne greenspaces are predominately composed of indigenous
412	rather than introduced species. Our study further highlights that multi-layered,
413	structurally complex indigenous vegetation plays a core role in sustaining high insect
414	biodiversity in urban areas, with the indigenous midstorey and canopy key to
415	maintaining a rich and functionally diverse indigenous insect community within this
416	system. Within the indigenous midstorey, graminoids surprisingly support the highest
417	indigenous insect species richness across all functional groups – particularly for
418	herbivores, predators and detritivores – followed by indigenous shrubs and lilioids.
419	The indigenous midstorey also hosts the largest percentage of unique species. Taken
420	together our findings emphasise the opportunity presented by indigenous understory

421 and midstorey plants, particularly indigenous graminoids in our study area, to

422 promote indigenous insect biodiversity in urban greenspaces.

423 Greenspace insect communities are dominated by indigenous insect species 424 Our results indicate that the insect community in our study area was composed 425 predominately of indigenous species. This finding aligns with previous studies of 426 insect richness in cities across other continents (Goertzen and Suhling 2014, Sing et 427 al. 2016, Brown and Hartop 2017). Despite the expectation that urban environments 428 act as hotspots for biological invasions (Cadotte et al. 2017), particularly of nonnative 429 plants and insects (Pysek et al 2010), we found a relatively low number of introduced 430 insect species. Interestingly however, some of these introduced species, for example 431 the European honeybee *Apis mellifera*, were common in and widespread across the 432 studied greenspaces. Consequently, the probability of occurrence of introduced insect 433 species on any particular plant across the study area is similar than that of indigenous 434 species. 435 Indigenous plant species promote indigenous insect diversity

436 We found multiple threads of evidence to suggest that indigenous plant species 437 sustain the highest numbers of indigenous insect species and host the largest 438 percentage of unique species. It is generally accepted in restoration ecology that the 439 presence of indigenous plant species promotes recolonisation by indigenous insect 440 species (Moir et al. 2005, Nemec and Bragg 2008). In most cases, phytophagous taxa 441 drive this trend (Proches et al. 2008, Woodcock et al. 2009) and it is a function of the 442 host-specificity of the insect species, provided that other factors are accounted for, 443 such as the insect's power of dispersal and suitable micro-climate conditions (Moir et 444 al. 2005).

445	It follows, therefore, that indigenous plants should encourage the occurrence of
446	indigenous insects, especially for herbivores, but also for other insect functional
447	groups. Indeed, our findings distinctly show that pollinators, herbivores, predators,
448	parasitoid and detritivores reach higher levels of species richness in association with
449	indigenous plant species. Experimental studies also support this assumption (Ballard
450	et al. 2013, Burghardt and Tallamy 2013, Salisbury et al. 2015, 2017). For example,
451	using experimental plantings of tree and shrub species, Burghardt and Tallamy (2013)
452	showed that nonnative plants supported less diverse herbivorous insect communities
453	than indigenous plants. Similarly, in an early successional experiment the biomass,
454	abundance and species richness of herbivorous, predatory and parasitoid insects was
455	lower on nonnative forbs than on indigenous forbs (Ballard et al. 2013). Working
456	specifically in an urban setting, Salisbury and colleagues (2015, 2017) experimented
457	with the origin of flowering plants in garden borders, demonstrating that insects
458	across a diverse range of functional groups were less abundant on nonnative than
459	indigenous plant species. These experimental findings have been substantiated by
460	observational approaches, particularly by studies conducted within urban
461	environments. For instance, a study that included the species-specific responses of
462	bees, beetles and heteropteran bugs to vegetation attributes of gardens, parks and golf
463	courses revealed how occurrence probabilities for most insect species decreased as a
464	function of the amount of nonnative plants present in the studied greenspaces
465	(Threlfall et al. 2017).
466	Midstorey as a key planting design element
467	Our results have shown that the midstorey is a highly valuable ecological asset in
468	urban landscapes in terms of promoting insect diversity. Midstorey vegetation -
460	which in our study also included plants appoint associated with the understance

469 which in our study also included plants species associated with the understorey –

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470	harboured nearly as many indigenous insect species as canopy vegetation. Indigenous
471	midstorey plant species in particular promoted higher levels of overall insect richness,
472	a pattern that was consistent across all insect functional groups. The midstorey also
473	sustained the highest number of unique indigenous insect species. Taken together, our
474	results indicate that the under- and midstorey are underappreciated strata with great
475	potential for supporting insect biodiversity across urban environments.
476	Our findings go beyond the accepted understanding that the greater structural
477	complexity of experimental, restored or managed sites, the higher the taxonomical
478	and functional diversity of insects and other invertebrates (Murdoch et al. 1972,
479	Brown 1984, Majer et al. 2007, Woodcock et al. 2009, Gibb and Cunningham 2010,
480	Mata et al. 2017, Threlfall et al. 2017, Schuldt et al. 2019). We show that within the
481	midstorey it is the indigenous species - particularly indigenous graminoids and shrubs
482	- that distinctly outperform their native and nonnative counterparts. Our results are
483	also consistent with previous studies that have documented how insect and other
484	arthropod communities are highly stratified across forest strata (Basset et al. 2003,
485	Ulyshen 2011). It is likely that greenspace midstorey vegetation supports different
486	insect taxa due to differences in habitat structure (e.g. foliage complexity or plant
487	surface textures), microclimate (e.g. light, temperature, wind or humidity differences),
488	unique food resources or particular inter-specific interactions, as has been discussed
489	for temperate deciduous forest (Ulyshen 2011). Unpacking the causal mechanisms for
490	our results would require experimental manipulations that fell beyond the scope of
491	this study.
492	Graminoids as a key midstorey growth form
493	A striking result to emerge from the data is that graminoids sustain the highest

494 number of indigenous insect species across all growth forms and indigenous

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495	graminoids show the highest species richness of indigenous insects across all growth
496	form by plant origin groups. Three indigenous tussock grass species made particularly
497	important contributions to supporting indigenous insect biodiversity in our study area:
498	common tussock-grass Poa labillardierei, wallaby grass Rytidosperma sp. and
499	kangaroo grass Themeda triandra. Indeed, P. labillardierei had the highest number of
500	indigenous insect species across the study, with any particular tussock grass patch
501	supporting as much as 5.4 times more indigenous insect species than the most
502	speciose lawn complex and 1.7 times more indigenous insect species than the spotted
503	gum Corymbia maculata, which was the tree species sustaining the highest number of
504	indigenous insect species across the study. These results substantiate previous
505	findings stressing the relevance of indigenous tussocks and other structurally complex
506	graminoids in providing a diversity of habitat and food resources for the immature
507	and adult life stage of insects and other invertebrates (Tscharntke and Greiler 1995,
508	Morris 2000, Barratt et al. 2005).
509	Remarkably, the capacity of indigenous graminoids to support the highest levels of
510	indigenous insect species across all studied plant groups was true for all insect
511	functional groups. This finding extends previous studies reporting on the positive
512	effects of tussocks and other graminoids on specific insect functional groups -
513	predominantly on herbivores, predators and parasitoids (Dennis et al. 1998,
514	Woodcock et al. 2007, Haaland et al. 2011), but also for pollinators (Saarinen et al.
515	2005, Potts et al. 2009). Our finding that graminoids, which are predominantly
516	pollinated by wind, support more pollinators and other flower-visiting insect species
517	than do lilioids or shrubs, which are predominantly pollinated by insects, is highly
518	noteworthy and provides insight into function and value of non-floral resources for
519	insect pollinators (Roulston and Goodell 2011, Requier and Leonhardt 2020). We

520	hope that our research will serve as a base for future studies on the capacity of
521	graminoids to provide habitat and food resources for insects in urban greenspaces,
522	particularly of non-floral resources for pollinators and other flower-visiting species.
523	Beyond graminoids, our data further emphasises the contributions of other midstorey
524	growth forms. For instance, indigenous shrubs and lilioids sustained the second and
525	third highest number of indigenous insect species across the growth from by plant
526	origin groups. Indeed, our findings indicate that any particular indigenous shrub or
527	lilioid species supports the same number of indigenous insect species as any particular
528	indigenous tree species; and substantially more insect species than any particular
529	native or nonnative tree species. However, we found that the number of unique
530	indigenous insect species varied markedly between these two plant groups, with
531	indigenous shrubs showing as much as 7.4 times more unique insect species than their
532	lilioid counterparts. In general, shrubs outperformed all other midstorey growth
533	forms, with approximately one out of every four indigenous insect species across the
534	study being exclusively associated with shrub species.
535	As underlined by an increasing body of literature (Mata et al. 2017, Threlfall et al.
536	2017, Aguilera et al. 2019, Norton et al. 2019, Majewska and Altizer 2020), including
537	recent reviews (Burkman and Gardiner 2014, Aronson et al. 2017), and meta-analyses
538	(Beninde et al. 2015), the evidence we found points to the critical role that midstorey
539	growth forms – particularly indigenous plant species – play in supporting
540	taxonomically and functionally diverse indigenous insect communities in urban
541	greenspaces. This finding is not only critical for insect conservation in urban
542	environments but of direct, immediate relevance for a wide range of animals such as
543	reptiles, birds and mammals that rely on them as a primary or complementary food
544	source. Our findings therefore underscore the potential of a diverse, primarily

545 indigenous understorey and midstorey strata to increase the positive biodiversity

- 546 outcomes provided by structurally complex vegetation. As such, they support ideas
- 547 that move beyond the stagnant approach of designing urban greenspaces
- 548 predominantly or often exclusively with nonnative short turfgrass lawn and tall trees
- 549 (Ikin et al. 2015, Smith et al. 2015, Parris et al. 2018, Norton et al. 2019).
- 550 *Limitations and future research*
- 551 We are aware of some features of our study context and design that might have
- 552 influenced our results. Firstly, the small proportion of introduced insects found in this
- study may be the result of recent introductions that have not yet had enough time to
- develop into established, large populations a pattern that is common to both the
- 555 Northern (Roques et al. 2009) and Southern Hemispheres (Ward and Edney-Browne
- 556 2015). It is therefore not inconceivable that some introduced species occurring in our
- study area in small, isolated populations might have gone undetected. Indeed, a recent
- follow up study in one of the study sites revealed the occurrence of the European
- 559 firebug Pyrrhocoris apterus a Palaearctic species not previously recorded in
- 560 Australia (LM unpublished data). As with any observational study, our study design
- 561 may have introduced unintended bias because our data collection period was
- 562 purposely designed to coincide with the peak activity season for insects in our study
- area (i.e., the summer months). We believe however that this effect is negligible as farfewer insect species are more active over the colder periods.
- This research has revealed many questions in need of further investigation. While we believe that our finding, that indigenous plants sustain substantially more indigenous insect species than their regionally native and nonnative counterparts, are transferable to other urban environments worldwide, the strong relationships we found between indigenous graminoids and indigenous insects might be less transferable. While

570 graminoids, particularly tussock forming species, were a dominant growth form in our 571 study area before colonial settlement other growth forms might have been more 572 representative in other bioregions. A global study across many bioregions is needed to 573 shed light on this question. 574 While we have emphasised the critical role of indigenous plants, we have also shown 575 that indigenous insects are being found in association with a wide array of regionally 576 native and nonnative plants. Thus, our study provides considerable insights in support 577 of the idea that urban environments may facilitate novel resource utilisation 578 (Valentine et al. 2020). We recommend future experimental work on this topic to 579 clarify to what extent these associations reflect host shifting patterns unique to urban 580 environments and to fully understand the opportunities and risks provided by novel 581 urban resources. On a wider level, research is also needed to determine to what extent 582 the increases in insect biodiversity provided by complementing lawns and trees with a 583 diverse palette of midstorey growth forms, particularly indigenous species, can boost 584 ecosystem multifunctionality in urban greenspaces – as recently demonstrated for 585 meso- and macrofauna influencing soil multifunctionality (Tresch et al. 2019). The 586 prospect of being able to understand the mechanistic links between, and to quantify 587 the contributions of, increased biodiversity due to greenspace management actions 588 and greenspace multifunctionality serves as a continuous incentive to future research. 589 *Implications for greenspace design and management* 590 As our research was conducted following the science-government partnerships model 591 (Ives and Lynch 2014), our findings have now been used by the City of Melbourne to 592 provide practical guidance for designing greenspaces that meet the needs of both 593 people and nature. We share these examples of applied ecological knowledge here as 594 a demonstration of how this ecological research can inform practical actions.

595	In the first instance, our study provides a blueprint and stimulus for built-environment
596	professionals, including architects, engineers, planners and designers, to conceptualise
597	and incorporate into their practice palettes of plant species that foster a larger
598	presence of indigenous plants over regionally native or nonnative species, whilst
599	incorporating a broader mixture of midstorey growth forms. These features are
600	expected to promote taxonomically and functionally diverse indigenous insect
601	communities - even when increasing the amount of greenspace is not feasible due to
602	other pressures (Beninde et al 2015). Integrating these plant palettes into practice may
603	further allow built-environment professionals to plan and design complex plant
604	communities that support and boost indigenous biodiversity in greenspaces and that
605	will likely contribute to bring locally extinct or rare species back into urban
606	environments (Baruch et al. 2020, Mata et al. 2020).
607	Another promising pathway that can be explored by greenspace professionals
608	includes identifying locations where lawns can be converted to more complex
609	vegetation that includes indigenous plants, particularly, at least in the bioregional
610	context of our study, graminoids, shrub and lilioids. Simple strategies for
611	incorporating more complex vegetation without compromising access to lawn areas
612	include placing the plantings around the greenspace boundaries or under the canopy
613	areas of larger trees where they can also act as a subtle exclusion zone and reduce the
614	risk of injury due to falling limbs or branches. Placing the taller midstorey plantings
615	away from footpaths and other infrastructure can also help meet 'Crime Prevention
616	Through Environmental Design' principles by maintaining a line of site (Piroozfar et
617	al. 2019).
618	Making a difference – implications for policy and beyond

*Making a difference – implications for policy and beyond* 

619	Through our study we have gained considerable insights that advance knowledge of
620	plant-insect relationships in urban greenspaces. However, to bring about beneficial
621	outcomes for urban landscapes, this scientific evidence must be embedded into policy
622	and, ultimately, operationalised into practice. Indeed, our research was conducted as
623	part of 'The Little Things that Run the City' (Mata et al. 2015, 2016), and two of our
624	co-authors were working for the City of Melbourne during the formation and early
625	analysis of this research. Not unexpectedly, project findings have been contributing to
626	inform City of Melbourne decision-making and policy, including the Council's
627	'Nature in the City Strategy' (City of Melbourne 2017), which includes goals to
628	increase indigenous biodiversity and specific targets to increase plant-related
629	midstorey habitat for insects and other taxa. Project findings have also been applied to
630	develop an insect biodiversity educational portal
631	(http://biodiversity.melbourne.vic.gov.au/insects/) and a children's book (Cranney et
632	al. 2017). These non-academic outcomes highlight the value of our work and provide
633	encouragement for future partnerships between industry professionals and researchers
634	advocating for and evidencing the value of urban biodiversity.
635	Another significant approach that could be used by local governments to advance
636	practice aimed at promoting insect biodiversity in urban environments is to
637	incentivise the translation of research findings into landscape design guidelines. These
638	non-mandatory documents, which have traditionally focused on aesthetic outcomes at
639	the expense of biodiversity, can distinctly influence outcomes on the ground.
640	Ultimately however, contractors and consultants responsible for delivering capital
641	works projects will overlook these if the required plants are not readily available
642	when needed. Therefore, the crucial role played by plant nurseries should not be
643	overlooked – an industry where supply tends to influence demand, with growers often

644	limiting production to reliable, profitable and easy to grow plants. Influencing plant					
645	supply will require a dedicated engagement with the nursery industry to broaden					
646	production to include a wider range of plant species known to support insect					
647	biodiversity as those provided evidence for in this study.					
648	Finally, while government officers may apply research findings to design greenspaces					
649	capable of supporting diverse insect communities, they must synergistically					
650	encourage operational and maintenance programs that can enable these communities					
651	to thrive. On ground operational and maintenance teams, that practice greenspace					
652	management on a regular basis, have the potential to be involved in identifying					
653	opportunities and challenges for supporting insects – and the plants they are more					
654	closely associated with - which may expand beyond the obvious choices available to					
655	or envisaged by office-based decision-makers. They are, for example, uniquely					
656	positioned to readily transfer knowledge on where the optimal soil and microhabitat					
657	conditions required for focal plant species within a given greenspace are met – a					
658	necessary prerequisite for these to be able to deliver the resources needed by insects					
659	to become established and thrive.					
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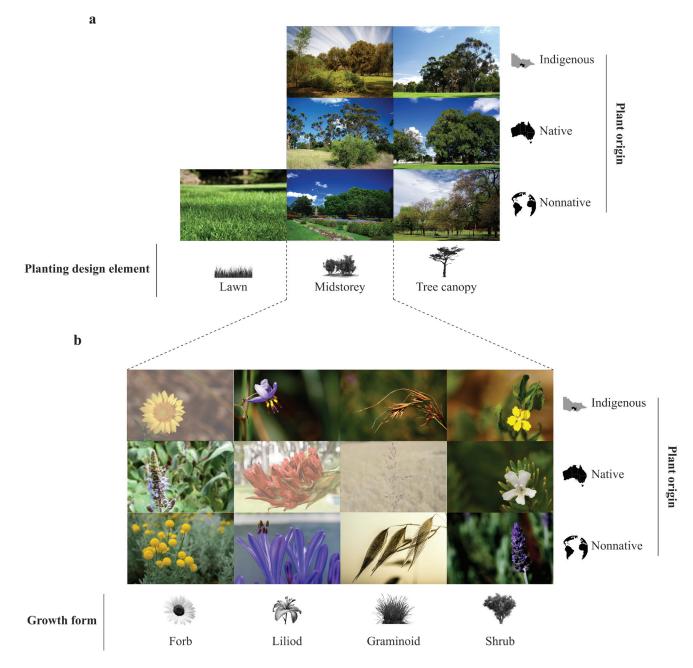
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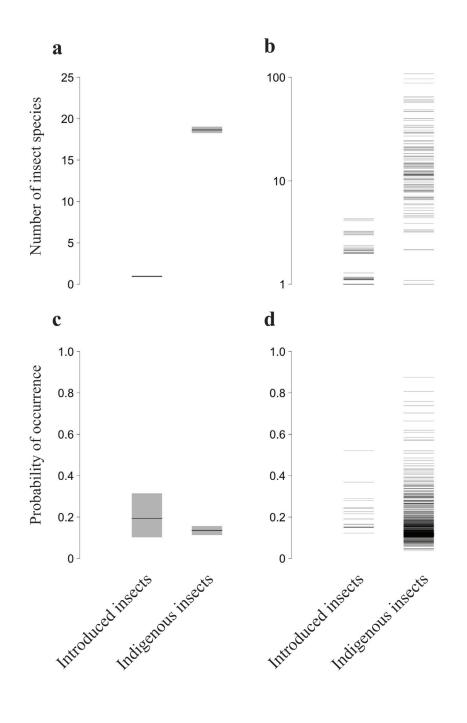
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- 918 Figure legends
- 919 Figure 1 Visual representation of the planting design element by plant origin (a) and
- 920 midstorey growth form by plant origin (b) groups that were part of the theoretical and
- 921 empirical dimensions of this study. Dimmed boxes in (b) indicate theoretical
- 922 combinations of midstorey growth form by plant origin groups that did/may not occur
- 923 in the study area or that occur but not at surveyable densities within the study's plots.
- 924 Figure 2 Estimated species richness of introduced and indigenous insect species at the
- 925 average plant species across the study (a) and at each surveyed plant species (b).
- 926 Estimated probabilities of occurrence of introduced and indigenous insect species at
- 927 the average plant species across the study (c) and at each surveyed plant species (d).
- 928 In all figures the black lines represent mean responses; the grey boxes in (a) and (c)
- 929 represent the associated statistical uncertainty (95% Credible Intervals). Y-axis in (b)
- 930 drawn in the log10 scale.
- 931 Figure 3 Estimated species richness of indigenous insects by planting design element
- 932 (a), midstorey growth form (b) and plant origin (c). Black lines represent mean
- 933 responses and grey boxes the associated statistical uncertainty (95% Credible
- 934 Intervals).
- 935 Figure 4 Estimated species richness of indigenous insect by planting design element
- 936 by plant origin (a) and midstorey growth form by plant origin (b). Black lines

937	represent mean	responses and	coloured boxes	the associated	statistical uncertainty
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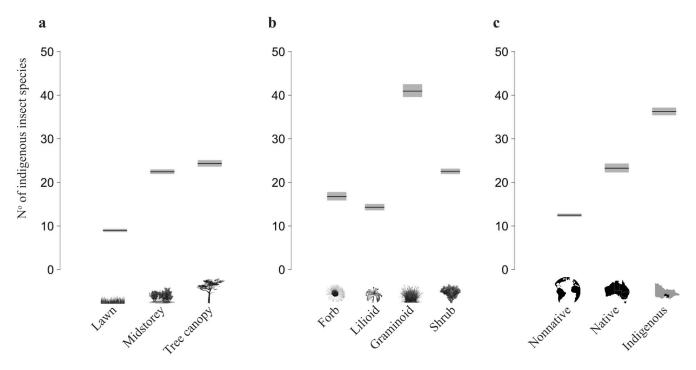
- 938 (95% Credible Intervals). For ease of interpretation plant origin has been colour coded
- 939 as yellow (nonnative), orange (native) or blue (indigenous).
- 940 Figure 5 Estimated species richness of indigenous insects by planting design element
- 941 by plant origin (a,c,e,g,i) and midstorey growth form by plant origin (b,d,f,h,j) for
- 942 pollinators (a,b), herbivores (c,d), predators (e,f), parasitoids (g,h) and detritivores
- 943 (i,j). Black lines represent mean responses and coloured boxes the associated
- 944 statistical uncertainty (95% Credible Intervals). For ease of interpretation plant origin
- has been colour coded as yellow (nonnative), orange (native) or blue (indigenous).
- 946 Figure 6 Indigenous insect community dissimilarity matrices for the planting design
- 947 element by plant origin (a) and midstorey growth form by plant origin (b) groups.
- 948 Percentages in the white cells were calculated using the Jaccard dissimilarity index,
- 949 where 0 and 1 represents minimum and maximum dissimilarly, respectively. Values
- 950 in the adjacent left columns represent the percentage of unique species observed in
- 951 each group.



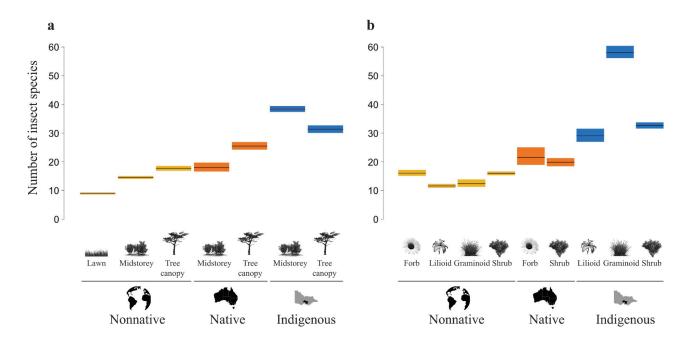
**Figure 1.** Visual representation of the planting design element by plant origin (a) and midstorey growth form by plant origin (b) groups that were part of the theoretical and empirical dimensions of this study. Dimmed boxes in (b) indicate theoretical combinations of midstorey growth form by plant origin groups that did/may not occur in the study area or that occur but not at surveyable densities within the study's plots.



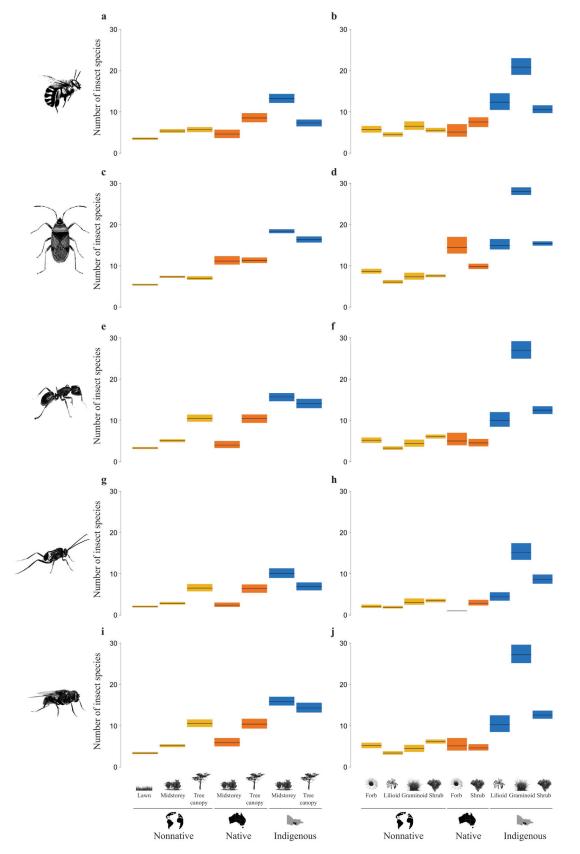
**Figure 2.** Estimated species richness of introduced and indigenous insect species at the average plant species across the study (a) and at each surveyed plant species (b). Estimated probabilities of occurrence of introduced and indigenous insect species at the average plant species across the study (c) and at each surveyed plant species (d). In all figures the black lines represent mean responses; the grey boxes in (a) and (c) represent the associated statistical uncertainty (95% Credible Intervals). Y-axis in (b) drawn in the log10 scale.



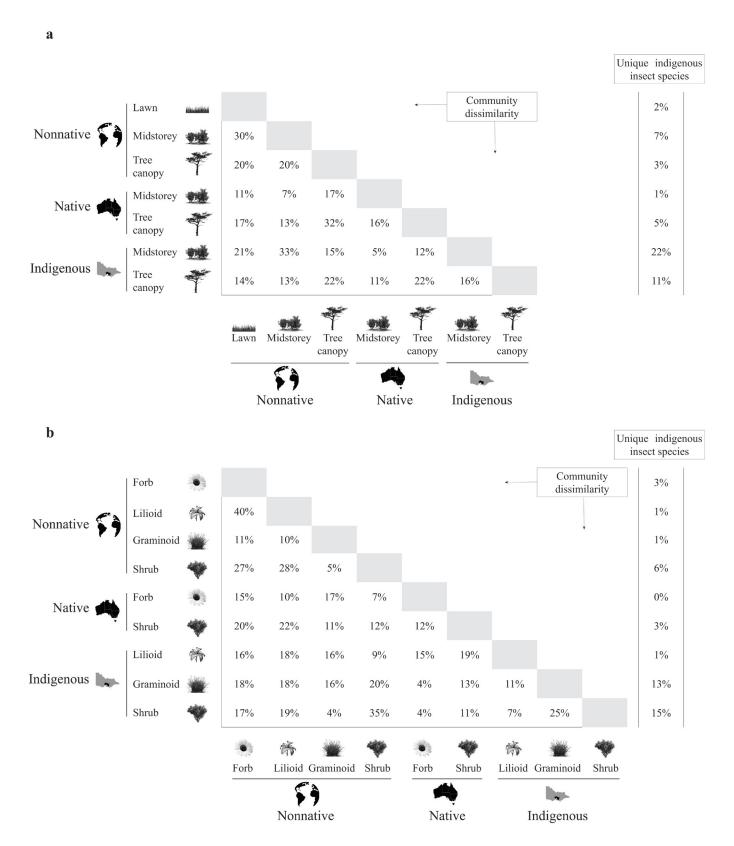
**Figure 3.** Estimated species richness of indigenous insects by planting design element (a), midstorey growth form (b) and plant origin (c). Black lines represent mean responses and grey boxes the associated statistical uncertainty (95% Credible Intervals).



**Figure 4.** Estimated species richness of indigenous insect by planting design element by plant origin (a) and midstorey growth form by plant origin (b). Black lines represent mean responses and coloured boxes the associated statistical uncertainty (95% Credible Intervals). For ease of interpretation plant origin has been colour coded as yellow (nonnative), orange (native) or blue (indigenous).



**Figure 5.** Estimated species richness of indigenous insects by planting design element by plant origin (a,c,e,g,i) and midstorey growth form by plant origin (b,d,f,h,j) for pollinators (a,b), herbivores (c,d), predators (e,f), parasitoids (g,h) and detritivores (i,j). Black lines represent mean responses and coloured boxes the associated statistical uncertainty (95% Credible Intervals). For ease of interpretation plant origin has been colour coded as yellow (nonnative), orange (native) or blue (indigenous).



**Figure 6.** Indigenous insect community dissimilarity matrices for the planting design element by plant origin (a) and midstorey growth form by plant origin (b) groups. Percentages in the white cells were calculated using the Jaccard dissimilarity index, where 0 and 1 represents minimum and maximum dissimilarly, respectively. Values in the adjacent left columns represent the percentage of unique species observed in each group.