1 **Title: The effect of resource limitation on the temperature dependence of mosquito**
2 **population fitness**

6 Data accessibility statement:

7 Data will be made publicly available at publication from the Dryad Digital Repository.

ABSTRACT

1. Laboratory-derived temperature-dependencies of life history traits are increasingly being used to make mechanistic predictions for how climatic warming will affect the abundance of disease vectors. These laboratory data are typically from populations reared on optimal resource supply, even though natural populations are expected to experience fluctuations in resource availability.

2. Using laboratory experiments and stage-structured population projection modelling, here we ask how resource limitation affects temperature-dependence of life history traits and emergent fitness of a principal arbovirus vector, *Aedes aegypti*, across a temperature range it typically experiences (22–32ºC).

3. We show that low-resource supply significantly depresses the vector's maximal population growth rate (r_{max}) across the entire temperature range and causes it to peak at a lower temperature, than under high-resource supply. This difference is driven by the fact that resource limitation significantly increases juvenile mortality, slows development, and reduces lifespan and size at maturity (which then decreases fecundity in adults). These results show that resource supply can significantly affect the temperature-dependence of population-level fitness of disease vectors by modifying the thermal responses of underlying traits.

4. Our study suggests that by ignoring resource limitation, projections of vector abundance and disease transmission based on laboratory studies are likely to substantially underestimate the effect of temperature on development time and juvenile survival, and overestimate the effect of temperature on lifespan, size and fecundity.

5. Our results provide compelling evidence for future studies to consider resource supply when making predictions about the effects of climate and habitat change on disease vectors. More generally, our results point at the need to consider the effects of resource limitation on temperature-dependence of life history traits to further advance Ecological Metabolic Theory and improve its utility for predicting the responses of holometabolous insects to climate change.

KEY WORDS: climatic warming, Ecological Metabolic Theory, environmental temperature, population-level fitness, population projection matrices, resource limitation, size-temperature rule, vector-borne disease

INTRODUCTION

The global public health burden of some vector-borne diseases has increased substantially in recent decades (Stanaway et al. 2016, World Health Organization 2014). The geographical distribution and abundance of vectors is strongly linked to transmission patterns of these diseases (Li et al. 2019, Ng et al. 2018). Climate and land-use change, coupled with rapid globalization, are thought to have contributed to shifts in both the distributions of vector species and the diseases they transmit (Brown, Medlock & Murray 2013, Kraemer et al. 2019, Lambin et al. 2010, Wilder-Smith & Gubler 2008). However, mechanistic models that account for how fundamental environmental drivers (temperature, rainfall and resource supply) may independently and interactively affect population-level vector fitness are largely absent (Mordecai et al. 2019).

Ecological Metabolic Theory (EMT) links body size and environmental temperature through metabolic rate to individual-level trait performance and population-level fitness (Brown et al., 2004, Savage et al. 2004). For example, within an ectotherm's operational thermal range, biological rates (e.g. metabolic, development, mortality) increase approximately exponentially with temperature to affect population growth in a similar way (Gillooly et al. 2001, Gillooly, Charnov & West 2002, Savage et al. 2004). While this core EMT prediction is well-supported, it has not yet been tested across varying levels of resource supply. This is because EMT models linking metabolic rates to population-level fitness assume that resource supply is not a limiting factor (Savage et al. 2004).

Temperature and resource supply may interact to significantly affect fitness and its component traits, because the energy cost of somatic maintenance, development and growth generally increases with temperature (Kooijman, 1988, 2000; Marshall et al. 2020). The ability to meet this increasing demand is influenced by resource supply from an individual's environment. If the resources available to an individual do not keep pace with these increasing maintenance and growth requirements, its growth, development, and survival should be compromised. Ultimately, these effects should negatively affect fitness, and the severity of these effects should increase with temperature.

The thermal responses of individual traits have been used extensively in the rapidly expanding body of research on the responses of disease transmission rates to climate change (Bhatt et al. 2013, Iwamura, Guzman-holst & Murray 2020, Mordecai et al. 2013, 2017; Parham & Michael 2010). The classical EMT framework has recently provided important

contributions in this area (Kirk et al. 2018, 2019; Molnár et al. 2013), including vector-borne

diseases (Mordecai et al. 2013, 2017). However, despite the potential importance of resource

supply, very little is known about how it interacts with temperature to influence vector

demographics and transmission capacity (Mordecai et al. 2019). For example, recently,

Mordecai et al. (2013, 2017; Ryan et al. 2019) used a EMT trait-based framework to predict

the temperature dependence of R_0, but all the parameterisations for the temperature

dependence of component traits come from mosquitoes provided with optimal food

quantities. Thus, the question of whether and how temperature and resource supply may

together modulate disease transmission through underlying traits remains open.

Additionally, temperature indirectly affects ectotherm fitness by determining size at maturity.

Generally, size at maturity decreases with rising temperature (the size-temperature rule,

Atkinson, 1994; Pettersen et al. 2019), which also applies to disease vectors, such as

mosquitoes (Rueda et al. 1990). For vectors specifically, female size is demographically and

epidemiologically important because it is associated with longevity, fecundity, and biting

behaviour. For example, individuals reared at warmer temperatures emerge earlier and are

smaller, shorter-living and less fecund than conspecifics reared at cooler temperatures

(Costanzo et al. 2018, Rueda et al. 1990). Reduced development time, due to increased

temperature, can increase fitness, but cooler temperatures yield larger, more fecund and

longer-living females, which are all traits that generally increase fitness (Birch 1948,

Costanzo et al. 2018, Livdahl & Sugihara 1984). Furthermore, since larger females live

longer, they are more likely to outlive a pathogen's external incubation period, and hence

have greater transmission potential than smaller individuals (Clements & Paterson 1981).

However, smaller females may bite more frequently, which can increase transmission

probability (Scott et al. 2000). All these lines of evidence underline the importance of

understanding the governing physiological mechanism(s) of the size-temperature rule and

determining how it relates to mosquito fitness and disease transmission. However, it remains

largely untested under resource limitation (Atkinson 1994, Forster et al. 2012).

Studies on the effects of resource availability on mosquito traits generally report a positive

association between food level and immature development and survival (Agnew et al. 2002,

Couret et al. 2014, Romeo Aznar et al. 2018), as well as adult body size, nutritional reserves

and fecundity (Briegel 1990, Colless & Chellapah 1960, Steinwascher 1982). Many studies

have examined how resource quantity effects fitness in interaction with larval density, as

competition for limited resources between larvae is generally believed to be a major regulator

of adult mosquito abundance (Dye 1982, 1984; Gilpin & McClelland 1979, Southwood et al.
1972). However, studies on the effects of resource supply on fitness have not generally
included temperature.

Given that wild mosquitoes are likely to experience variations of temperature and resource
availability (Arrivillaga & Barrera 2004, Barrera, Amador & Clark 2006, Walsh et al. 2011),
measuring the effects of both variables directly is important for predicting their population
dynamics. In this study, we investigated whether and how fitness and its component traits are
affected by the interaction between realistic temperature and resource supply variation and
interactions between these two factors in *Aedes aegypti*; a principal mosquito vector of
human arboviruses (e.g. dengue, yellow fever and zika; Brady & Hay 2020). We show that
these factors can both, act alone, and interact to significantly modify the responses of body
size, life history traits and fitness to temperature.

METHODS

To investigate the effects of temperature and resource supply on mosquito life history, we
employed a 3×2 factorial design comprised of three temperatures (22, 26, and 32°C) and two
resource supply levels: 0.1 (low-resource supply) and 1 mg/larva/day (high-resource supply).
These experimental temperatures span the range of average annual temperatures that this
strain of *Ae. aegypti* is likely to experience in the wild (Fort Meyer, FL; Arguez et al. 2012).
This particular low-resource supply level was chosen because previous work has
demonstrated that it is the lowest resource limitation that can be applied to this species
without resulting in almost complete juvenile mortality; a level of limitation often
experienced by wild populations (Barrera et al. 2006, Arrivillaga & Barrera 2004). The high-
resource supply level corresponds to the upper mid-range of the high resource supply levels
used in Arrivillaga & Barrera (2004) and Barrera et al. (2006), and is consistent with the
levels of resource supply commonly used in laboratory studies on this species (Couret et al.
2014, Rueda et al. 1990).

Batches of approximately 300 *Ae. aegypti* (F16-19) eggs were randomly assigned to one of
the three experimental temperatures and immersed in plastic tubs containing 150 ml of tap
water. Each tub was provided with a pinch of powdered fish food (Cichlid Gold®, Hikari,
Kyrin Food Industries Ltd., Japan) to stimulate overnight hatching. The tubs were then
submerged in water baths (Grant Instruments: JAB Academy) set at either 22, 26, or 32°C.
Water baths were situated in a 20°C climate-controlled insectary with a 12L:12D photoperiod

and 30 minutes of gradual transition of light levels to simulate sunrise and sunset. On the

following day, first instar larvae were separated into cohorts of 30 and held in tubs containing

150 ml of water. We created three replicate tubs per treatment (90 individuals/treatment).

Low-resource supply treatments were provided 3 mg of powdered fish food and high-

resource supply treatments received 30 mg. Thereafter, resource levels were adjusted daily

according to the number of living individuals in each tub prior to feeding each day such that

resource levels were maintained at an approximately constant level during the juvenile

lifespan. Water volumes were also adjusted daily in accordance with mortality to maintain

larval density (0.2 larvae $\times$ ml^{-1}).

Fitness calculation

To calculate population-level fitness, we used our data to parameterise age-structured (Leslie)

population projection matrices (Caswell 1989), which describe change in a population over

time:

$$\mathbf{N}_{t+1} = \mathbf{A}\mathbf{N}_t, \tag{1}$$

where $\mathbf{N}_t$ is a vector of the age structure at time t and $\mathbf{A}$ is the population projection matrix.

The first row of elements in $\mathbf{A}$ is populated with fecundity data. Fecundity is the number of

female offspring produced per female at age i. The sub-diagonal of $\mathbf{A}$ is populated with the

probability of survival from age i to age $i + 1$. Multiplying the transition matrix ($\mathbf{A}$; eqn. 1)

and age-structured population size vector ($\mathbf{N}_t$; eqn. 1) sequentially across time intervals yields

population dynamics. Once the stable age (stage) distribution of the abundance vector is

reached, the dominant eigenvalue of the system is the finite population rate of increase (λ)

(Caswell 1989). Then, the intrinsic rate of population growth,

$$r_{\max} = \log(\lambda).$$

This is a population's inherent capacity to reproduce, and therefore a measure of population-

level fitness (Birch 1948, Cole 1954, Savage et al. 2004). Negative $r_{\max}$ values indicate

decline and positive ones, growth. The projection matrices were built and analysed using the

`popbio` R package (R Core Team 2018, Stubben & Milligan 2007).

Parameterisation

Immature development time and immature and adult survival probabilities

Matrix survival elements (the sub-diagonal of the matrix **A**; eqn. 1) were populated with

continuous survival probabilities derived from the Kaplan-Meier survival function in the

`survival` R package (Therneau 2020, R Core Team 2018). We assumed life stage duration

(i.e. larva-to-pupa-to-adult) was the mean duration of transitioning into and out of that stage,

and a fixed age of adult emergence at the mean age of emergence. Adult survival elements

were populated with the Kaplan-Meier probabilities. Hatching-to-adult development times

were calculated by recording the day and time that egg eclosion, pupation and adult

emergence occurred for each individual. Upon pupation, mosquitoes were held in individual

falcon tubes containing 5ml of tap water. This enabled pupa-to-adult development durations

and the lifespans of individual starved adults to be recorded. In the absence of food, this

condition forces adults to metabolise the nutritional reserves accumulated during larval

development. Therefore, starved adult lifespan is a useful indicator of the carry over effects

of temperature and resource availability in the larval habitat (Agnew et al. 2002, Briegel

1990).

Fecundity (the first row of **A**; eqn. 1)

As a proxy for body size, wing length (L in eqn. 3) was measured to the nearest 0.01 mm

from the alula to the wing tip, excluding the apical fringe (van Handel & Day 1989). Wings

(one per female) were removed, mounted onto glass slides, photographed using a dissecting

microscope and then measured with ImageJ software (Schneider et al. 2012). Body size was

estimated from these wing lengths using the equation ($R^2 = 0.92$; $n = 75$, $p<0.001$; Nasci

1990):

$$M = -0.140 + 0.029L^3 \tag{3}$$

213 Here, M is dry mass (mg) at emergence and L is wing length (mm).

214 An allometric relationship ($R^2 = 0.46$; $n = 57$, $p<0.001$; Colless & Chellapah 1960) was then

215 used to estimate lifetime fecundity (F) from M:

$$\log(F) = 4.24 + 0.99 \log(M) \tag{4}$$

216 The Colless and Chellapah (1960) data are from mosquitoes reared at 26°C, so mass-

217 dependent fecundity (F) was corrected to each of our experimental temperatures (F_{T_j}'s, with

218 the index j ranging over the set of our experimental temperatures) using the equation
219 (Gillooly et al. 2001, Dell et al. 2011):

$$F_{T_j} = e^{4.24} M^{0.99} e^{-\frac{E}{k}\left[\frac{1}{T_j} - \frac{1}{299.15}\right]}. \tag{5}$$

Here, the scaling constant ($e^{4.24}$), and exponent (0.99) are from eqn. 4, E is the average
activation energy of heterotrophic respiration in eukaryotes (0.65 eV), k is Boltzmann's
constant (8.62 x 10^{-5} eV K^{-1}), 299.15 ($26^{o}C$) is the reference temperature in Kelvin (the
temperature at which the original study was conducted; Colless & Chellapah 1960), and T_j
(also in Kelvin) is the jth experimental temperature the rate is standardised to. Each F_{T_j} was
then multiplied by 0.5 (assuming a 1:1 male-to-female offspring ratio) and divided by
lifespan to obtain the temperature-specific individual daily fecundity.

Parameter sensitivity

We used the delta method to approximate 95% confidence intervals (CIs) for our fitness
calculations (Bowles et al. 2015, Caswell 1989, Skalski et al. 2007) to account for how
uncertainty in survival and fecundity estimates is propagated through to the r_{max} estimate.
This method requires the standard errors of the survival and fecundity element estimates. For
survival, we used the standard errors estimated by the Kaplan-Meier survival function. For
fecundity, we used the standard errors of the mean daily egg production rates for each
treatment. However, since the R^2 for eqn. 4 is low (0.46), as an additional sensitivity analysis,
we also calculated fitness using the upper and lower 95% CI's of the exponent (0.99 ± 0.28;
eqn. 4) for the scaling of lifetime fecundity with mass.

Elasticity analysis

Elasticities were used to quantify the proportional contributions of individual life history
traits to r_{max}. Elasticity, e_{ij}, measures the proportional effect on λ of an infinitesimal change in
an element of **A** (eqn. 1) with all other elements held constant (the partial derivative)
(Caswell et al. 1984, de Kroon et al. 1986). This partial derivative of λ with respect to each
element of **A** is, $s_{ij} = \partial\lambda/\partial a_{ij} = v_i w_j$ with the dot product $\langle \mathbf{w}, \mathbf{v} \rangle = 1$. Here, **w** is the dominant
right eigenvector (the stage distribution vector of **A**), **v** is the dominant left eigenvector (the
reproductive value vector of **A**), and a_{ij} is the $i \times j^{th}$ element of **A**. Elasticities can then be
calculated using the relationship: $e_{ij} = a_{ij}/\lambda \times s_{ij}$. Multiplying an elasticity by λ gives the
absolute contribution of its corresponding a_{ij} to λ (Caswell 1986, de Kroon et al. 1986).

Absolute contributions for juvenile survival and fecundity elements were summed and
changed proportionally to quantify the sensitivity of r_{max} to these traits.

Statistical analyses

All analyses were conducted using R (R Core Team 2018). In the first instance, to assess how
temperature, resource supply, their interaction (whether low-resource supply treatments
exhibited different thermal responses) and replicate influenced trait performance we fitted
linear regression models. Model diagnostics and Shapiro-Wilk (1965) normality tests were
used to assess data normality. Data for development, juvenile mortality rate, lifespan, wing
length and fecundity were nonlinear, positive and right skewed. Therefore, for these traits we
used full factorial generalized linear models (GLM) with gamma distributions and identity
link functions (predictor effects were considered additive) to determine the significance of
each predictor on the thermal response of each of these traits. Count data for cumulative
juvenile mortality were modeled using a Poisson GLM.

The best model in each case was obtained by dropping terms from the full model (consisting
of the trait its fixed effect predictors) sequentially using the Akaike Information Criterion
(AIC). Goodness-of-fit of the GLMs was assessed using model deviance/d.f. scores and
model residuals.

RESULTS

Except for juvenile mortality rate, all trait responses varied significantly with temperature and
resource supply, with a significant interaction between the two environmental variables
(Table 1, Fig. 1). Thus, the realised effect of temperature on trait responses was consistently
and significantly mediated by resource supply.

Development time varied significantly with the interaction between temperature and resource
supply (ANOVA; $F_{2,0.75} = 24.40$, $p<0.001$; Table 1); whereas development time decreased
both at warmer temperatures and under high-resource supply, the rate of decrease with
temperature was greater under low-resource supply than under high-resource supply. Under
low-resource supply, development time decreased by 15.48 days as temperatures increased
from 22 to 32°C, whereas under high-resource supply, it decreased by 6.38 days across this
range (Table 2, Fig. 1a).

Juvenile mortality rate significantly increased with temperature (ANOVA; $F_{1, 26.80} = 20.04$,
$p<0.001$; Table 1) and resource supply ($F_{1, 7.19} = 7.19$, $p<0.05$). Mortality rate was higher

under high-resource supply than under low-resource supply, but the resource level gradients did not differ significantly (interaction: $F_{1,\ 0.47} = 0.35$, $p>0.61$). Under high-resource supply mortality rate increased by 0.31 day^{-1} as temperatures increased from 22 to 32°C; whereas under low-resource supply it increased by 0.25 day^{-1} (Table 2, Fig 1b).

Cumulative juvenile mortality varied significantly with the interaction between temperature and resource supply (ANOVA; $\chi^2=26.39$, $p<0.001$; Table 1). Under low-resource supply, cumulative mortality decreased as temperatures increased from 22 to 26°C and increased from 26 to 32°C (Fig. 1c). Under high-resource supply, it increased linearly with temperature.

Lifespan varied significantly with the interaction between temperature and resource supply (ANOVA; $F_{2,\ 2.51} = 15.34$, $p<0.001$; Table 1). Although lifespan decreased both at warmer temperatures and under low-resource supply, the rate of decrease with temperature was greater under high-resource supply than under low-resource supply. High-resource supply lifespan decreased by 8.61 days, whereas low-resource supply lifespan decreased by 4.71 days as temperatures increased from 22 to 32°C (Table 2, Fig 1d).

The interaction between temperature and resource supply resulted in significant variation in size at maturity (wing length) between resource levels (ANOVA; $F_{1,0.03} = 4.50$, $p=0.01$; Table 1). Size decreased both at warmer temperatures and under low-resource supply, though the rate of decrease with temperature was greater under high-resource supply than under low-resource supply. Under low-resource supply, length decreased by 0.37 mm as temperatures increased from 22 to 32°C, while under high-resource supply, length decreased by 0.54 mm (Table 2, Fig 1e).

Predicted fecundity varied significantly with the temperature $\times$ resource supply (ANOVA; $F_{2,\ 2.58} = 6.99$, $p<0.01$; Table 1). However, only the variation between resource levels at 26°C was significant (Fig. 1f). Fecundity under high-resource supply increased by 7.75 eggs/female/day as temperatures increased from 22 to 32°C; whereas under low-resource supply it increased by 6.05 eggs/female/day (Table 2).

Population fitness (r_{max})

Resource limitation depressed r_{max} to negative values at all temperatures, with a unimodal relationship of r_{max} with temperature (Fig. 2, Table 2). Low-resource supply r_{max} increased from –0.11 at 22°C to –0.05 at 26°C and then decreased acutely to –0.40 at 32°C. In contrast,

under high-resource supply, r_{max} was always positive and increased monotonically with

temperature from 0.13 at 22°C to maximal growth (0.22) at 32°C.

Elasticity analysis

Juvenile survival was the most important contributor to r_{max} (Fig. 3). For example, under low-

resource supply at 32°C, a 0.25 proportional increase in juvenile survival would halve the rate

of decline from –0.40 to –0.20 (Fig. 3a). In contrast, for the same treatment, a proportional

increase of the same magnitude for fecundity would increase r_{max} from –0.40 to –0.39 (Fig.

3b). This highlights how the temperature dependence of r_{max} derives mainly from how

resource supply level impacts juvenile mortality and development, which determine the

number of reproducing individuals and the timing of reproduction, respectively. Fecundity,

on the other hand, has a relatively negligible effect on r_{max}, which suggests that the carry-over

effect of reduced size at maturity on r_{max} is relatively weak.

DISCUSSION

Our results show that the temperature dependence of population-level fitness and its

component life history traits can be significantly modulated by resource supply. When adult

lifespan and juvenile development time were expressed as rates (adult mortality and juvenile

development rate, respectively), all traits under high-resource supply met EMT expectations

by being positive and increasing nonlinearly with temperature (Fig. 1a and d) (Dell et al.

2011). This result is consistent with previous studies that have used laboratory-derived trait

responses to temperature to estimate *Ae. aegypti* fitness and R_0; these studies report similar

responses of traits to our high-resource supply responses (e.g. Iwamura et al. 2020, Ryan et

al. 2019). For example, in Mordecai et al. (2017), juvenile *Ae. aegypti* development rate

increased from approximately 0.07 day^{-1} at 22°C to 0.14 day^{-1} at 32°C. In the present study,

development rate (Fig. 1a, Table 2) increased by a similar margin (~0.08 to ~0.16 day^{-1})

across the same temperature range.

In contrast, under low-resource supply, although trait thermal responses qualitatively met

EMT predictions, they deviated markedly from the results of previous laboratory

experiments. For example, in the present study under low resource supply, juvenile

development rate was ~0.03 day^{-1} at 22°C, which is 57% less than the rate (~0.07 day^{-1}) used

at this temperature in Mordecai et al. (2017) to predict R_0. Similarly, under low-resource

supply at 32°C, we found juvenile development rate was ~0.05 day^{-1}; whereas in Mordecai et

al. (2017) it was 64% greater (~0.14 day^{-1}). Juvenile survival proportions under low-resource

supply also differed markedly from the high-resource supply proportions used in Mordecai et

al. (2017). In the present study, survival was 0.51 at 22°C, 0.64 at 26°C and 0.22 at 32°C;

whereas, in Mordecai et al. (2017) it was approximately 0.83 between 20-22°C, 0.76 at 25°C,

and 0.77 between 30-34°C.

These differences between the temperature dependence of traits under low- versus high-

resource supply resulted in a marked divergence of the temperature dependence of r_{max} (Fig.

2) between the two treatments, which was negative throughout and unimodal (declining

steeply from 26 to 32°C) under low-resource supply, and positive and increasing under high-

resource supply. This indicates that population fitness becomes increasingly constrained by

resource limitation as temperatures increase. Although resource limitation decreased juvenile

mortality rate (Fig. 1b), the elasticity analysis shows that the primary mechanism underlying

this is increased cumulative juvenile mortality, which decreased population-level fertility and,

ultimately, fitness (Figs. 1c and 3a). The effect of resource limitation on fitness was further

compounded by the increase in juvenile development time (Fig. 1a), which delayed the onset

of reproduction.

The effects of resource supply on adult traits (size at emergence, adult lifespan and fecundity)

were weak compared to its effects on juvenile traits (Fig. 3). For example, under high-

resource supply, adult lifespan and body size were higher at 32°C than at 26°C, yet fitness at

32°C was predicted to be 41% higher (Fig. 2). This is because high-resource supply and

increased temperature minimised cumulative juvenile mortality and optimised development

rate. This allowed faster recruitment at 32°C, leading to increased fitness as greater numbers

of individuals could contribute to growth through reproductive output sooner than for other

treatments. This finding is consistent with general studies of ectotherm fitness (Huey &

Berrigan 2001), including mosquitoes (Dye 1984). Therefore, projections of vector

abundance and disease transmission that assume high-resource supply are likely to

underestimate the effect of temperature on development time and juvenile survival, and

overestimate effects of temperature on lifespan and fecundity.

Cumulative juvenile mortality increased significantly with temperature, and it was generally

greater under low-resource supply (Fig. 1c, Tables 1 and 2). This is probably because somatic

maintenance cost increases as metabolic rate increases (Kooijman 1988, 2000); if the amount

of resource available cannot satisfy basal requirements, mortality will increase. This explains

why the highest level of mortality occurred at 32°C and under low-resource supply, where the energy supply-demand deficit was likely largest.

Development time at both resource supply levels decreased non-linearly with temperature, though it decreased more steeply with temperature under low-resource supply than under high-resource supply (Table 1, Fig. 1a). Qualitatively, this result supports development cost theory, which posits that while metabolic rate increases approximately exponentially with temperature (Brown et al. 2004), development cost is expected to be U-shaped (Marshall et al. 2020), ultimately resulting in a U-shaped temperature dependence of development time. Although our temperature range was not wide enough to determine whether development time is U-shaped in *Ae. aegypti*, our results suggest the relationship is, at least, L-shaped. Development time was highest at 22°C for both resource supply levels. However, high-resource supply possibly minimised development cost at 22°C, which reduced high-resource supply development time (by 67%) relative to low-resource supply at this temperature. At both resource supply levels, the steepest declines in development time occurred as temperatures increased from 22 to 26°C, likely because this change in temperature increased metabolic rate at the cost of development (thus reducing development cost).

Size at emergence decreased significantly with temperature for both resource supply levels (Table 1, Fig. 1e), consistent with the size-temperature rule (Atkinson 1994, Pettersen et al. 2019). Resource supply drove significant intercept variation in the size-temperature relationship, which reflects how high-resource supply allowed larvae to accumulate greater energy reserves and grow to become larger adults than their less well-fed conspecifics that were exposed to the same temperature. Size decreased more steeply with temperature under high-resource supply than under low-resource supply. Wing length decreased by ~0.20 mm at both low- and high resource supply as temperatures increased from 22 to 26°C (Table 2). However, as temperatures increased from 26 to 32°C, wing length decreased by ~0.17 and ~0.34 mm under low- and high-resource supply, respectively. For size at emergence, the decline with temperature under high resource supply may reflect the optimal (or approaching optimal) attainable size at each temperature.

Like size at emergence, adult lifespan decreased more acutely with temperature under high-resource supply than under low-resource supply (Table 1, Fig. 1d). This derived from the difference in lifespan between resource supply levels at 22°C. As temperatures increased from 26 to 32°C, lifespans declined to a similar extent at both resource supply levels. Under

low-resource supply, lifespan decreased by ~3.69 days, whereas under high-resource supply

it decreased by ~3.96 days (Table 2). However, as temperatures increased from 22 to 26°C,

lifespan decreased by ~1.02 days under low-resource supply and by ~4.65 days under high-

resource supply. This relatively flatter decline under low-resource supply may reflect how

larvae respond to cooler temperatures when resources are limited. Cooler temperatures

prolong development by slowing underlying rate processes (Angilletta et al. 2004). These

processes may have been slowed further by metabolic down-regulation in response to

resource limitation, which is predicted to delay development, but increase survival, as

individuals invest available energy in maintenance (Moore & Whitacre, 1972, Storey &

Storey 1990). Under high-resource supply, lifespan declined linearly, which may reflect how

higher levels of resource supply allow temperature to optimise trait responses. For lifespan,

the decline with temperature under high resource supply may reflect the optimal (or

approaching optimal) attainable lifespan at each temperature.

We did not measure the effect of temperature directly on fecundity but used the size-scaling

of this trait to estimate this effect. This is because most of the effect of resource limitation on

juveniles is expected to affect adult mosquitoes indirectly, by reducing size at emergence,

with few direct carryover effects (Briegel 1990, Steinwascher 1982). Predicted fecundity

increased nonlinearly with temperature, significantly mediated by resource supply levels (Fig

1f, Table 1). Across both resource levels, these predicted fecundity values are somewhat

lower than those reported in past studies. For example, in Mordecai et al. (2017), *Ae. aegypti*

fecundity increased from approximately 3.58 to 11.20 eggs/female/day as temperatures

increased from 22 to 32°C compared to an increase from 1.71 to 9.46 eggs/female/day under

high-resource supply and 1.79 to 7.84 eggs/female/day under low supply, across the same

temperature range. This difference likely arises from the fact that our size-scaling based

predictions ignore additional effects on fecundity, such as adult food supply. However, even

substantial underestimation of fecundity by our size-scaling predictions would not affect our

main conclusions, because predicted fitness was relatively insensitive to this trait (Fig. 3b).

Although we have not considered the temperature dependence of resource supply itself

(supply was held constant across temperatures in our experiments), in nature, the availability

of resources may in fact be temperature-dependent. This is because microbial growth rates

increase with temperature to some optimum, which may increase the concentration of food in

the environment (Craine, Fierer, & McLauchlan 2010; Cross et al. 2015, Smith et al. 2019).

For example, *Anopheles* (Chouaia et al. 2012) and *Aedes* (Souza et al. 2019) mosquitoes can

be reared exclusively on cultures of *Asaia* bacteria. The temperature dependence of resource
supply is an interesting and potentially important avenue of investigation that would further
elucidate how the interaction between temperature and resource availability affects the fitness
of mosquitoes.

Organisms experience significant resource limitation over space and time in nature. This is
particularly true for insects such as mosquitoes, which have juvenile stages restricted to
small, ephemeral aquatic habitats that are susceptible to resource fluctuations (Arrivillaga &
Barrera 2004, Barrera et al. 2006, Subra & Mouchet 1984, Walsh et al. 2011). Overall, our
results provide compelling evidence for the importance of resource availability in larval
habitats in *Ae. aegypti* and other mosquito vectors in mediating the effect of climatic
warming on their population growth and therefore, disease transmission. More generally,
considering the effect of resource availability is also likely to be important for further
advancing Ecological Metabolic Theory, and improving its utility for understanding the
responses of holometabolous insects to environmental change.

References

Agnew, P., Hide, M., Sidobre, C., & Michalakis, Y. (2002). A minimalist approach to the effects of density-dependent competition on insect life-history traits. *Ecol. Entomol.*, 27, 396–402.

Angilletta, M. J., Steury, T. D., & Sears, M. W. (2004). Temperature, growth rate, and body size in ectotherms: Fitting pieces of a life-history puzzle. *Inter. Comp. Biol.*, 44, 498–509.

Arguez, A., Durre, I., Applequist, S., Vose, R. S., Squires, M. F., Yin, X. et al. (2012). Noaa's 1981-2010 U.S. climate normals. *Bull. Am. Meteorol. Soc.*, 93, 1687–1697.

Arrivillaga, J., & Barrera, R. (2004). Food as a limiting factor for *Aedes aegypti* in water-storage containers. *J. Vector. Ecol.*, 29, 11–20.

Atkinson, D. (1994). Temperature and Organism Size—A Biological Law for Ectotherms? In: *Adv. Ecol. Res.,* 25 {[ed(s).] [Begon, M. & Fitter, A.H.]}. Academic Press, 1-58.

Barrera, R., Amador, M., & Clark, G. G. (2006). Ecological factors influencing *Aedes aegypti* (Diptera: Culicidae): Productivity in artificial containers in Salinas, Puerto Rico. *J. Med. Entomol.*, 43, 484–492.

Bhatt, S., Gething, P. W., Brady, O. J., Messina, J. P., Farlow, A. W., Moyes, C. L. et al. (2013). The global distribution and burden of dengue. *Nature*, 496, 504–507.

Birch, L. C. (1948). The intrinsic rate of natural increase of an insect population. *J. Anim. Ecol.*, 17, 15–26.

Bowles, M. L., McBride, J. L., & Bell, T. J. (2015). Long-term processes affecting restoration and viability of the federal threatened Mead's milkweed (*Asclepias meadii*). *Ecosphere*, 6, 1–22.

Brady, O. J., & Hay, S. I. (2020). The global expansion of dengue: How *Aedes aegypti* mosquitoes enabled the first pandemic arbovirus. *Annu. Rev. Entomol.*, 65, 191–208.

Briegel, H. (1990). Metabolic relationship between female body size, reserves, and fecundity of *Aedes aegypti*. *J. Insect Physiol.*, 36, 165–172.

Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M., & West, G. B. (2004). Toward a metabolic theory of ecology. *Ecology*, 85, 1771–1789.

Brown, L., Medlock, J., & Murray, V. (2013). Impact of drought on vector-borne diseases - how does one manage the risk? *Public Health*, 128, 29–37.

Caswell, H. (1986). Life cycle models for plants. *Lectures on Mathematics in the Life Sciences*, 18, 171–233.

Caswell, H. (1989). *Matrix population models construction, analysis, and interpretation*. *Natural Resource Modeling*. Sunderland, Massachusettes: Sinauer Associates.

Caswell, H., Naiman, R. J., & Morin, R. (1984). Evaluating the consequences of reproduction in complex salmonid life cycles. *Aquac. Res.*, 43, 123–134.

Chouaia, B., Rossi, P., Epis, S., Mosca, M., Ricci, I., Damiani, C. et al. (2012). Delayed larval development in *Anopheles* mosquitoes deprived of *Asaia* bacterial symbionts. *BMC Microbiol.*, 12, 1–8.

Clements, A. N., & Paterson, G. D. (1981). The analysis of mortality and survival rates in wild populations of mosquitoes. *J. Appl. Ecol.*, 18, 373–399.

Cole, L. C. (1954). The population consequences of life history phenomena. *Q. Rev. Biol.*, 29, 103–137.

Colless, D. H., & Chellapah, W. T. (1960). Effects of body weight and size of blood-meal upon egg production in *Aedes aegypti* (Linnaeus) (Diptera, Culicidae). *An. Trop. Med. Parasit.*, 54, 475–482.

Costanzo, K. S., Westby, K. M., & Medley, K. A. (2018). Genetic and environmental influences on the size-fecundity relationship in *Aedes albopictus* (Diptera: Culicidae): Impacts on population growth estimates? *PLoS ONE*, 13, 1–17.

Couret, J., Dotson, E., & Benedict, M. Q. (2014). Temperature, larval diet, and density effects on development rate and survival of *Aedes aegypti* (Diptera: Culicidae). *PLoS ONE*, 9, 1–9.

Craine, J. M., Fierer, N., & McLauchlan, K. K. (2010). Widespread coupling between the rate and temperature
sensitivity of organic matter decay. *Nat. Geosci.*, 3, 854–857.

Cross, W. F., Hood, J. M., Benstead, J. P., Huryn, A. D., & Nelson, D. (2015). Interactions between temperature
and nutrients across levels of ecological organization. *Glob. Change Biol.*, 21, 1025–1040.

de Kroon, H., Plaisier, A., van Groenendael, J., & Caswell, H. (1986). Elasticity: The relative contribution of
demographic parameters to population growth rate. *Ecology*, 67, 1427–1431.

Dell, A. L., Pawar, S., & Savage, V. M. (2011). Systematic variation in the temperature dependence of
physiological and ecological traits. *Proc. Natl. Acad. Sci. USA*, 108, 10591–10596.

Dye, C. (1982). Intraspecific competition amongst larval *Aedes aegypti*: Food exploitation or chemical
interference? *Ecol. Entomol.*, 7, 39–46.

Dye, C. (1984). Models for the population dynamics of the yellow fever mosquito, *Aedes aegypti. J. Anim.
Ecol.*, 53, 247–268.

Forster, J., Hirst, A. G., & Atkinson, D. (2012). Warming-induced reductions in body size are greater in aquatic
than terrestrial species. *Proc. Natl. Acad. Sci. USA*, 109, 19310–19314.

Gillooly, J. F., Brown, J. H., West, G. B., Savage, V. M., & Charnov, E. L. (2001). Effects of size and
temperature on metabolic rate. *Science*, 293, 2248–2251.

Gillooly, J. F., Charnov, E. L., & West, G. B. (2002). Effects of size and temperature on developmental time.
Nature, 417, 70–73.

Gilpin, M. E., & McClelland, G. A. (1979). Systems analysis of the yellow fever mosquito *Aedes aegypti.
Fortschr. Zool.*, 25, 355–388.

Huey, R. B., & Berrigan, D. (2001). Temperature, demography, and ectotherm fitness. *Am. Nat.*, 158, 204–210.

Iwamura, T., Guzman-holst, A., & Murray, K. A. (2020). Accelerating invasion potential of disease vector
Aedes aegypti under climate change. *Nat. Commun.*, 11, 1–10.

Kirk, D., Jones, N., Peacock, S., Phillips, J., Molnár, P. K., Krkošek, M. et al. (2018). Empirical evidence that
metabolic theory describes the temperature dependency of within-host parasite dynamics. *PLoS Biol.*, 16,
1–20.

Kirk, D., Luijckx, P., Stanic, A., & Krkošek, M. (2019). Predicting the thermal and allometric dependencies of
disease transmission via the metabolic theory of ecology. *Am. Nat.*, 193, 661–676.

Kooijman, S. A. L. M. (1988). The von Bertalanffy growth rate as a function of physiological parameters: a
comparative analysis. In: *Mathematical Ecology* {[ed(s).] [Hallam, T.G., Gross, L.J. & Levin, S.A.]}.
World Scientific, 3–45.

Kooijman, S. A. L. M. (2000). *Dynamic energy and mass budgets in biological systems*. Cambridge: Cambridge
University Press.

Kraemer, M. U. G., Reiner, R. C., Brady, O. J., Messina, J. P., Gilbert, M., Pigott, D. M. et al. (2019). Past and
future spread of the arbovirus vectors *Aedes aegypti* and *Aedes albopictus. Nat. Microbiol.*, 4, 854–863.

Lambin, E. F., Tran, A., Vanwambeke, S. O., Linard, C., & Soti, V. (2010). Pathogenic landscapes: Interactions
between land, people, disease vectors, and their animal hosts. *Int. J. Health Geogr.*, 9, 1–13.

Li, R., Xu, L., Bjørnstad, O. N., Liu, K., Song, T., Chen, A. et al. (2019). Climate-driven variation in mosquito
density predicts the spatiotemporal dynamics of dengue. *Proc. Natl. Acad. Sci. USA*, 116, 3624–3629.

Livdahl, T. P., & Sugihara, G. (1984). Non-linear interactions of populations and the importance of estimating
per capita rates of change. *J. Anim. Ecol.*, 53, 573–580.

Marshall, D. J., Pettersen, A. K., Bode, M., & White, C. R. (2020). Developmental cost theory predicts thermal
environment and vulnerability to global warming. *Nat. Ecol. Evol.*, 4, 406–411.

Molnár, P. K., Kutz, S. J., Hoar, B. M., & Dobson, A. P. (2013). Metabolic approaches to understanding climate
change impacts on seasonal host-macroparasite dynamics. *Ecol. Lett.*, 16, 9–21.

Moore, C. G., & Whitacre, D. M. (1972). Competition in mosquitoes. 2. Production of *Aedes aegypti* larval
growth retardant at various densities and nutrition levels. *Ann. Entomol. Soc. Am.*, 65, 915–918.

Mordecai, E. A., Caldwell, J. M., Grossman, M. K., Lippi, C. A., Johnson, L. R., Neira, M. et al. (2019).
Thermal biology of mosquito-borne disease. *Ecol. Lett.*, 22, 1–19.

Mordecai, E. A., Cohen, J. M., Evans, M. V., Gudapati, P., Johnson, L. R., Lippi, C. A. et al. (2017). Detecting
the impact of temperature on transmission of Zika, dengue, and chikungunya using mechanistic models.
PLOS Negl. Trop. Dis., 11, 1–18.

Mordecai, E. A., Paaijmans, K. P., Johnson, L. R., Balzer, C., Ben-Horin, T., de Moor et al. (2013). Optimal
temperature for malaria transmission is dramatically lower than previously predicted. *Ecol. Lett.*, 16, 22–
30.

Nasci, R. S. (1990). Relationship of wing length to adult dry weight in several mosquito species (Diptera:
Culicidae). *J. Med. Entomol.*, 27, 716–719.

Ng, K.-C., Chaves, L., Tsai, K.-H., Chuang, T.-W., Ng, K.-C., Chaves, L. F. et al. (2018). Increased adult *Aedes
aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae) abundance in a dengue transmission hotspot,
compared to a coldspot, within Kaohsiung City, Taiwan. *Insects*, 9, 98.

Parham, P. E., & Michael, E. (2010). Modeling the effects of weather and climate change on malaria
transmission. *Environ. Health Perspect.*, 118, 620–626.

Pettersen, A. K., White, C. R., Bryson-Richardson, R. J., & Marshall, D. J. (2019). Linking life-history theory
and metabolic theory explains the offspring size-temperature relationship. *Ecol. Lett.*, 22, 518–526.

R Core Team. (2018). R: A language and environment for statistical computing. R Foundation for Statistical
Computing, Vienna, Austria. Availble at: http://www.r-project.org/

Romeo Aznar, V., Alem, I., De Majo, M. S., Byttebier, B., Solari, H. G., & Fischer, S. (2018). Effects of
scarcity and excess of larval food on life history traits of *Aedes aegypti* (Diptera: Culicidae). *J. Vector
Ecol.*, 43, 117–124.

Rueda, L. M., Patel, K. J., Axtell, R. C., & Stinner, R. E. (1990). Temperature-dependent development and
survival rates of *Culex quinquefasciatus* and *Aedes aegypti* (Diptera: Culicidae). *J. Med. Entomol.*, 27,
892–898.

Ryan, S. J., Carlson, C. J., Mordecai, E. A., & Johnson, L. R. (2019). Global expansion and redistribution of
Aedes-borne virus transmission risk with climate change. *PLOS Negl. Trop. Dis.*, 13, 1–20.

Savage, V. M., Gilloly, J. F., Brown, J. H., Charnov, E. L., Gillooly, J. F., & West, G. B. (2004). Effects of body
size and temperature on population growth. *Am. Nat.*, 163, 429–441.

Schneider, C. A., Rasband, W. S., & Eliceiri, K. W. (2012). NIH Image to ImageJ: 25 years of image analysis.
Nat. Methods, 9, 671–675.

Scott, T. W., Amerasinghe, P. H., Morrison, A. C., Lorenz, L. H., Clark, G. G., Strickman, D. et al. (2000).
Longitudinal studies of *Aedes aegypti* (Diptera: Culicidae) in Thailand and Puerto Rico: Blood feeding
frequency. *J. Med. Entomol.*, 37, 89–101.

Shapiro, S. S., & Wilk, M. B. (1965). An Analysis of Variance test for normality (Complete Samples).
Biometrika, 52, 591–611.

Skalski, J. R., Millspaugh, J. J., Dillingham, P., & Buchanan, R. B. (2007). Calculating the variance of the finite
rate of population change from a matrix model in Mathematica. *Environ. Modell. Softw.*, 22, 359–364.

Smith, T. P., Thomas, T. J. H., Garcia-Carreras, B., Sal, S., Yvon-Durocher, G., Bell, T., & Pawar, S. (2019).
Community-level respiration of prokaryotic microbes may rise with global warming. *Nat. Commun.*, 10,
5124.

Southwood, T. R., Murdie, G., Yasuno, M., Tonn, R. J., & Reader, P. M. (1972). Studies on the life budget of
Aedes aegypti in Wat Samphaya, Bangkok, Thailand. *Bull. Org. Mond. Sante Bull. Wld Hlth Org.*, 46,
211–226.

Souza, R. S., Virginio, F., Riback, T. I. S., Suesdek, L., Barufi, J. B., & Genta, F. A. (2019). Microorganism-
based larval diets affect mosquito development, size and nutritional reserves in the yellow fever mosquito
Aedes aegypti (Diptera: Culicidae). *Front. Physiol.*, 10, 1–24.

Stanaway, J. D., Shepard, D. S., Undurraga, E. A., Halasa, Y. A., Coffeng, L. E., Brady, O. J. et al. (2016). The
global burden of dengue: An analysis from the Global Burden of Disease Study 2013. *Lancet Infect. Dis.*,
16, 712–723.

Steinwascher, K. (1982). Relationship between pupal mass and adult survivorship and fecundity for *Aedes
aegypti*. *Environ. Entomol.*, 11, 150–153.

Storey, K. ., & Storey, J. M. (1990). Metabolic rate depression and biochemical adaptation in anaerobiosis,
hibernation and estivation. *Q. Rev. Biol.*, 65, 145–174.

Stubben, C., & Milligan, B. (2007). Estimating and analyzing demographic models using the popbio package in
R. *J. Stat. Softw.*, 22, 1–23.

Subra, R., & Mouchet, J. (1984). The regulation of preimaginal populations of *Aedes aegypti* (L.) (Diptera:
Culicidae) on the Kenya coast. II. Food as a main regulatory factor. *An. Trop. Med. Parasit.*, 78, 63–70.

Therneau, T. M. (2020). A package for survival analyis in R. Available at: https://cran.r-
project.org/package=survival

van Handel, E., & Day, J. F. (1989). Correlation between wing length and protein content of mosquitoes. *J. Am.
Mosquito Contr.*, 5, 180–182.

Walsh, R. K., Facchinelli, L., Ramsey, J. M., Bond, J. G., & Gould, F. (2011). Assessing the impact of density
dependence in field populations of *Aedes aegypti*. *J. Vector Ecol.*, 36, 300–307.

Wilder-Smith, A., & Gubler, D. J. (2008). Geographic expansion of dengue: The impact of international travel.
Med. Clin. North Am., 92, 1377–1390.

World Health Organization (2014). A global brief on vector-borne diseases. *World Health Organization*,
WHO/DCO/WH, 1–56.

Trait	Predictor	χ^2	df	*F* value	p-value
Cumulative juvenile mortality	**Temperature**	**131.54**	**2**	**-**	**<0.001 *****
$R^2 = 0.54$	**RS**	**7.92**	**1**	**-**	**<0.01 ****
	Temperature: RS	**26.39**	**2**	**-**	**<0.001 *****
Juvenile mortality rate	**Temperature**	**28.06**	**2**	**9.61**	**<0.001 *****
$R^2 = 0.23$	**RS**	**8.45**	**1**	**5.79**	**0.02 ***
	Temperature: RS	1.43	2	0.49	0.61
	Residuals	157.67	108		
Development time	**Temperature**	**9.78**	**2**	**319.08**	**<0.001 *****
$R^2 = 0.96$	**RS**	**50.43**	**1**	**3292.46**	**<0.001 *****
	Temperature: RS	**0.75**	**2**	**24.40**	**<0.001 *****
	Residuals	2.30	150		
Lifespan	**Temperature**	**34.23**	**2**	**209.76**	**<0.001 *****
$R^2 = 0.73$	**RS**	**2.83**	**1**	**34.57**	**<0.001 *****
	Temperature: RS	**2.51**	**2**	**15.34**	**<0.001 *****
	Residuals	12.30	150		
Body size	**Temperature**	**0.66**	**2**	**112.92**	**<0.001 *****
$R^2 = 0.78$	**RS**	**0.92**	**1**	**315.53**	**<0.001 *****
	Temperature: RS	**0.03**	**2**	**4.50**	**0.01 ***
	Residuals	0.42	143		
Fecundity rate	**Temperature**	**64.48**	**2**	**175.04**	**<0.001 *****
$R^2 = 0.79$	**RS**	**0.84**	**1**	**4.56**	**0.03 ***
	Temperature: RS	**2.58**	**2**	**6.99**	**<0.01 ****
	Residuals	26.34	143		

626 **Table 1 | Type II Analysis of Variance results from GLMs fitted to the responses of life history**

627 **traits to temperature and resource levels.** Significant effects are shown in boldface type. * $\Rightarrow$

628 $p<0.05$; ** $\Rightarrow p<0.01$; *** $\Rightarrow p<0.001$. Each pseudo R^2 (= residual deviance/null deviance)

629 approximates how much variance the model was able to capture.

Trait	Temperature (°C)	Mean ± 95% Confidence intervals	
		Low-resource supply	High-resource supply
Cumulative juvenile mortality (number of individuals)	22	10.60 ± 3.19	4.50 ± 2.05
	26	7.58 ± 3.21	11.40 ± 4.17
	32	22.13 ± 4.88	17.00 ± 2.48
Juvenile mortality rate (day^{-1})	22	0.10 ± 0.06	0.16 ± 0.08
	26	0.13 ± 0.13	0.33 ± 0.10
	32	0.34 ± 0.04	0.47 ± 0.04
Development time (days)	22	36.48 ± 1.24	12.19 ± 0.39
	26	27.86 ± 1.51	8.53 ± 0.25
	32	21.00 ± 4.73	5.81 ± 0.19
Lifespan (days)	22	6.61 ± 0.95	11.35 ± 0.69
	26	5.59 ± 0.70	6.70 ± 0.45
	32	1.90 ± 0.53	2.74 ± 0.39
Wing length (mm)	22	2.65 ± 0.10	3.13 ± 0.03
	26	2.45 ± 0.07	2.93 ± 0.04
	32	2.28 ± 0.10	2.59 ± 0.03
Fecundity (eggs/female/day)	22	1.79 ± 0.57	1.71 ± 0.11
	26	1.97 ± 0.24	3.16 ± 0.21
	32	7.84 ± 3.42	9.49 ± 2.01
Population-level fitness (r_{max})	22	–0.11 ± 0.02	0.13 ± 0.02
	26	–0.05 ± 0.03	0.15 ± 0.03
	32	–0.40 ± 0.05	0.22 ± 0.04

Table 2 | Comparison of the effect of resource supply on the temperature dependence of fitness and its component traits. For individual traits, the significance of the pairwise comparisons between resource supply levels are shown in Fig. 1. The numbers of individuals that survived to adulthood (*n*) in each treatment were: 22°C under low-resource supply *n* = 23, under high-resource supply *n* = 37; 26°C under low-resource supply *n* = 29, under high-resource supply *n* = 30; 32°C under low-resource supply *n* = 10, under high-resource supply *n* = 27.

Temperature (°C), Resource level (mg/larva/day)

Fig. 1 | The combined effect of temperature and resource supply level on *Ae. aegypti* life history

traits. Fecundity data were estimated using eqns. 3, 4 and 5. p-values denote significance levels of the

pairwise comparisons of the means for the resource supply levels at each temperature (GLM). $* \Rightarrow p$

value<0.05; $*** \Rightarrow p$ value<0.001; ns. $\Rightarrow$ nonsignificant. The resulting ANOVAs of the GLMs for

each trait are presented in Table 1. The interaction between temperature and resource supply was

significant in all cases except for juvenile mortality.

Fig 2 | Population-level *Ae. aegypti* fitness (r_{max}) by resource supply (0.1 (low) or 1 mg/larva/day

(high)) across temperatures. Fitness estimates for each treatment, with 95% confidence intervals

(CI). The three data points for each treatment represent r_{max} estimated using the 95% CI bounds

(± 0.28) of the exponent for the scaling of lifetime fecundity with mass (eqn. 4). The lightest greyscale

hue estimates derive from the lower 95% CI (0.99–0.28), the midrange hue estimates with trend lines

derive from the slope (0.99) and the darkest hue derive from the upper 95% CI (0.99+0.28).

Fig. 3 | Sensitivity of r_{max} to proportional changes in juvenile survival (a) and fecundity (b) for

each *Ae. aegypti* population. Juvenile survival was the most important contributor to r_{max}, as

relatively small changes in the summed matrix elements for this trait would result in substantial

changes in r_{max}. Sensitivity of r_{max} to fecundity was much weaker compared to sensitivity to juvenile

traits. Dashed vertical lines in a and b denote r_{max} at zero as represented by the midrange grey hue

estimates in Fig. 2. Open symbols denote high-resource supply (1 mg/larva/day); closed symbols

denote low-resource supply (0.1 mg/larva/day).