ATP dependent DNA transport within cohesin: Scc2 clamps DNA on top of engaged heads while Scc3 promotes entrapment within the SMC-kleisin ring

James E Collier[1†], Byung-Gil Lee[2†], Maurici B Roig[1], Stanislav Yatskevich[1], Naomi J Petela[1], Jean Metson[1], Menelaos Voulgaris[1], Andres Gonzalez Llamazares[2], Jan Löwe[2*] and Kim A Nasmyth[1*]

[1] Department of Biochemistry, University of Oxford, Oxford, OX1 3QU, UK

[2] MRC Laboratory of Molecular Biology, Cambridge CB2 0QH, UK

[†] These authors contributed equally to this work.

[*] Corresponding authors:

Email: ashley.nasmyth@bioch.ox.ac.uk and jyl@mrc-lmb.cam.ac.uk

SUMMARY

In addition to extruding DNA loops, cohesin entraps within its SMC-kleisin ring (S-K) individual DNAs during G1 and sister DNAs during S-phase. All three activities require related hook-shaped proteins called Scc2 and Scc3. Using thiol-specific crosslinking we provide rigorous proof of entrapment activity in vitro. Scc2 alone promotes entrapment of DNAs in the E-S and E-K compartments, between ATP-bound engaged heads and the SMC hinge and associated kleisin, respectively. This does not require ATP hydrolysis nor is it accompanied by entrapment within S-K rings, which is a slower process requiring Scc3. Cryo-EM reveals that DNAs transported into E-S/E-K compartments are “clamped” in a sub-compartment created by Scc2’s association with engaged heads whose coiled coils are folded around their elbow. We suggest that clamping may be a recurrent feature of cohesin complexes active in loop extrusion and that this conformation precedes the S-K entrapment required for sister chromatid cohesion.

INTRODUCTION

Protein complexes containing SMC and kleisin subunits organise the spatial arrangement, or topology, of DNAs in most if not all living organisms (Nasmyth, 2001; Yatskevich et al., 2019). Best characterised are the eukaryotic cohesin and condensin complexes that are thought to organise chromosomal DNAs during interphase and mitosis, respectively, by a process of loop extrusion (LE) (Golfier et al., 2020). Cohesin in addition mediates the connections between sister DNAs that hold sister chromatids together during mitosis until their disjunction at the onset of anaphase (Oliveira et al., 2010; Uhlmann et al., 1999). Many clues as to their molecular mechanisms have emerged from structural studies. All contain a pair of rod-shaped SMC proteins with a dimerisation domain, known as the hinge, at one end and an ABC-like ATPase domain at the other, separated by a ~50 nm long anti-parallel intra-molecular coiled coil (Haering et al., 2002). Their association creates V-shaped dimers whose apical ATPase head domains are interconnected by a kleisin subunit (Scc1) whose N-terminal domain forms a three-helix bundle with the coiled coil emerging from Smc3's ATPase head (Gligoris et al., 2014), called its neck, and whose C-terminal winged helical domain binds to the base (or cap) of Smc1's ATPase head to complete the ring (Haering et al., 2004).

Hinge dimerisation facilitates numerous other contacts between Smc1 and Smc3. First, their coiled coils interact with each other extensively, all the way from the hinge to the joint, a small break in the coiled coil roughly 5 nm above the heads, effectively zipping up the coiled coils (Bürmann et al., 2019; Chapard et al., 2019; Diebold-Durand et al., 2017; Soh et al., 2015). This process leads to the juxtaposition of the Smc1 and Smc3 heads, which are loosely associated under these conditions (Chapard et al., 2019; Diebold-Durand et al., 2017). Second, their coiled coils fold around an elbow, which results in an interaction between the hinge and a section of the coiled coils approximately 10 nm from the heads (Bürmann et al., 2019). Finally, the γ-phosphate of ATP bound to one ATPase head binds a signature motif on the other, resulting under appropriate conditions in engagement of the heads and a sandwiching of two molecules of ATP between them, a process that is a precondition for subsequent ATP hydrolysis (Arumugam et al., 2003; Lammens et al., 2004; Marcos-Alcalde et al., 2017). Head engagement has been proposed to disrupt coiled coil interactions, at least in the vicinity of the heads, yet the full extent of this disruption is not known.

The ATPase activities of SMC-kleisin complexes as well as all their biological functions in vivo depend on additional proteins that are recruited through their association with kleisin subunits and act by binding DNA and interacting with various SMC protein domains. In cohesin, this class of proteins consists of large hook-shaped proteins composed of HEAT repeats, known as Heat repeat containing proteins Associated With Kleisins (HAWKs) (Wells et al., 2017). Cohesin has three such HAWKs known as Scc2, Scc3, and Pds5. Scc3 is thought to be permanently bound to the complex (Tóth et al., 1999) while association of Scc2 and Pds5, whose occupancy is mutually exclusive, are more dynamic (Petela et al., 2018). Scc2 is essential for cohesin's ATPase activity (Petela et al., 2018), for its loading onto chromosomes (Ciosk et al., 2000), for maintaining cohesin's chromosomal association during G1 (Srinivasan et al., 2019), and for cohesin's ability to extrude loops in vitro (Davidson et al., 2019; Kim et al., 2019). However, Scc2 is not required to maintain cohesion during G2 or even establish cohesion during S phase from complexes previously associated with unreplicated DNAs (Srinivasan et al., 2019). Pds5 also has multiple functions. By recruiting Wapl, it promotes cohesin's dissociation from chromosomes, a process blocked by acetylation of two lysine residues on Smc3 during S phase (Beckouët et al., 2016; Chan et al., 2013, 2012). Pds5 also promotes acetylation during S phase and inhibits deacetylation during G2 and thereby protects sister chromatid cohesion which would otherwise be destroyed by Wapl-mediated release (Chan et al., 2013).

Though the mechanism by which cohesin extrudes loops remains mysterious, there is a clear and simple hypothesis as to how cohesin holds sister DNAs together, namely by entrapping them both inside the S-K ring created through the binding of a kleisin subunit to the ATPase heads of an Smc1/Smc3 heterodimer (Gruber et al., 2003; Haering et al., 2002). This model explains the key observation that cleavage of cohesin's kleisin subunit by separase, or any other site-specific protease, is sufficient to trigger sister chromatid disjunction at anaphase (Oliveira et al., 2010; Uhlmann et al., 2000). To measure such entrapment in yeast, we have substituted residues within all three interfaces that make up S-K rings by pairs of cysteine residues that can be crosslinked by the thiol-specific reagent bis-maleimidoethane (BMOE). Around 20% of cohesin complexes can be crosslinked simultaneously at all three interfaces in vivo (Gligoris et al., 2014), and in post-replicative cells this is accompanied by formation of SDS-resistant structures that hold together the sister DNAs of circular minichromosomes, called catenated dimers or CDs (Chapard et al., 2019; Gligoris et al., 2014; Srinivasan et al., 2018). Because the two DNAs associated with

CDs are not otherwise intertwined (Haering et al., 2008), they must be held together by cohesin through a topological mechanism, either by co-entrapment within a chemically circularised S-K ring or conceivably in a three-way Borromean ring containing a pair of sister DNA rings and an SDS-resistant S-K ring. Importantly, the study of numerous mutants has revealed a perfect correlation between CD formation and cohesion establishment (Srinivasan et al., 2018), suggesting that these structures are actually responsible for sister chromatid cohesion or at the very least are produced by a highly related mechanism. Using cysteine pairs that crosslink heads that are not engaged, but are otherwise closely juxtaposed (J) (Chapard et al., 2019), it has been established that sister DNAs are at least some of the time entrapped between juxtaposed heads and the kleisin associated with them, namely within a J-K sub-compartment of the S-K ring (see Fig. 1A for an overview of the compartments).

Loading of cohesin onto minichromosomes during G1 leads to a different topological interaction, namely catenation of individual circular DNAs by S-K rings (chemically circularised for detection), known as catenated monomers or CMs. Though loading of cohesin throughout the genome is normally accompanied by CM formation on minichromosomes, cohesin complexes containing a hinge with mutations within its lumen that neutralises its positive charge can load throughout the genome but cannot form either CMs or CDs (Srinivasan et al., 2018), implying that stable chromosomal association is not necessarily synonymous with entrapment of DNAs within S-K rings. That cohesin can associate with DNA in a functional manner without being topologically entrapped within S-K rings is supported by the finding that LE in vitro can be mediated by a version of human cohesin whose Smc1, Smc3, and kleisin subunits are expressed as a single polypeptide and whose hinge interface has been crosslinked by BMOE (Davidson et al., 2019). Importantly, neither of the above observations exclude the possibility that cohesin usually associates with chromosomal DNAs by entrapping a loop within its S-K ring, a type of association that has been termed pseudo-topological.

Previous reports claiming entrapment of DNA within cohesin rings in vitro used salt resistance and sensitivity to cleavage as their criteria (Murayama and Uhlmann, 2015, 2014). However, there are fundamental limitations to such experiments. Many types of association other than entrapment within a closed compartment could give rise to salt resistance and cleavage sensitivity. Equally serious, even if these criteria were indicative of

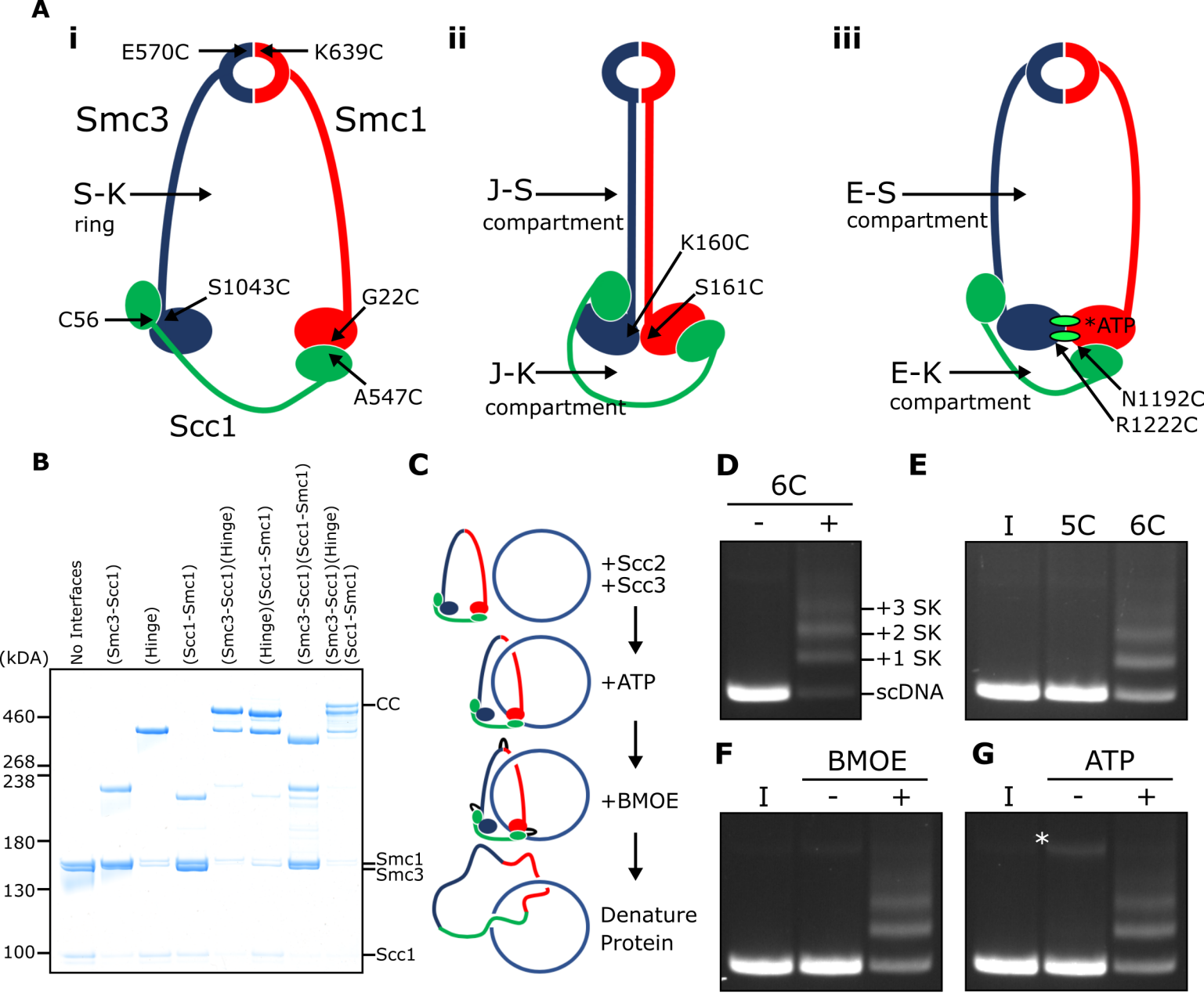
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Fig 1

Figure 1. SMC-kleisin (S-K) rings entrap circular DNA in vitro

(A) Cohesin's different compartments and the position of cysteine pairs used in our crosslinking studies. **(B)** BMOE-induced crosslinking of S-K rings with cysteine pairs in the specified interfaces. CC = circular cohesin. (**C**) The entrapment assay scheme. (**D**) Entrapment of DNA in S-K rings in the presence or absence of 6C cohesin, or (**E**) the presence 5C cohesin lacking Scc1A547C, or 6C cohesin. (**F**) For DNA in the presence of Scc2, Scc3, and 6C cohesin and the presence or absence of BMOE, or (**G**) the presence or absence of ATP. Entrapment assays incubated for 40 min (* = damaged open circular DNA) (I = input DNA).

topological entrapment, they reveal little or nothing as to its nature, namely whether DNAs are entrapped in S-K rings or other closed compartments, for example the E-S and E-K compartments between ATP-bound engaged heads and the SMC hinge and associated kleisin, respectively, or indeed other types of compartment created by multiple contacts between HAWKs and SMC proteins. For these reasons, we describe here the use of thiol-specific crosslinking to measure bona fide topological entrapment of DNAs within S-K rings in vitro. Both Scc2 and Scc3 are essential for this process, as are their abilities to bind DNA. The process is dependent on ATP binding and stimulated by its hydrolysis, a feature largely absent from previous assays (Minamino et al., 2018).

Remarkably, we find that Scc2 alone promotes the rapid entrapment of DNAs within E-S and E-K compartments in a process that is not accompanied by entrapment within S-K rings, and propose that E-S/E-K entrapment occurs simultaneously through a single mechanism. Because E-S/E-K entrapment is an order of magnitude more rapid than S-K entrapment, we suggest that creation of the former by Scc2 may be a precursor to the latter, a process contingent on the action of Scc3. Electron cryo-microscopy (cryo-EM) of complexes formed between cohesin's SMC-kleisin trimers and linear or circular DNAs in the presence of Scc2 suggests that entrapment within E-S/E-K compartments involves transport of DNA between ATPase heads prior to their engagement, whereupon DNAs are "clamped" in a sub compartment formed by Scc2's association with engaged heads in a manner similar to that recently observed in a complex between DNA and both human and *Schizosaccharomyces pombe* cohesin associated with both $Scc2^{NIPBL/Mis4}$ and $Scc3^{SA2/Psc3}$ (Higashi et al., 2020; Shi et al., 2020). Our observations reveal key insights into the biochemical activities of Scc2 and Scc3 and suggest that the recurrent clamping of DNAs by $Scc2^{NIPBL/Mis4}$ and engaged heads resulting in E-S/E-K entrapment, followed by their subsequent release, may be an integral aspect of cohesin's ability to load onto and translocate along DNA.

RESULTS

SMC-kleisin rings entrap circular DNA in vitro

We expressed cohesin trimers from *Saccharomyces cerevisiae* consisting of Smc1, Smc3, and Scc1 in insect cells using the baculovirus expression system (Fig. S1A). Scc3 was expressed separately because co-expression with trimers resulted in substoichiometric yields. We also expressed a version of Scc2 lacking its N-terminal domain (Scc2C, residues 133-1493). Though this form no longer binds Scc4, it is fully capable of activating cohesin's ATPase activity (Petela et al., 2018), and for simplicity we will refer to this as Scc2 throughout most of the text. To measure entrapment of DNAs inside S-K rings (Fig. 1A i), we introduced cysteine pairs within all three ring interfaces (Smc1K639C-Smc3E570C, Smc1G22C-Scc1A547C, and Smc3S1043C-Scc1C56) that enables them to be crosslinked using BMOE. Individual interfaces were crosslinked with efficiencies varying from 30 - 70% and by comparing the migration of proteins following crosslinking cysteine pairs at single (2C), double (4C), and triple (6C) interfaces, we identified a covalently circular species only produced when all three interfaces were crosslinked (Fig. 1B).

To measure DNA entrapment within cohesin's S-K compartment in vitro, 6C SMC-kleisin trimers were mixed with circular supercoiled DNAs, Scc2, and Scc3, and incubated for 40 min at 24˚C following addition of ATP. BMOE was then added and the mixture placed on ice for 6 min after which proteins were denatured by adding SDS to a final concentration of 1% and heating at 70˚C for 20 min. The DNA was then fractionated by agarose gel electrophoresis and visualised by ethidium bromide staining (Fig. 1C & D). Addition of 6C trimers to Scc2/Scc3/DNA mixtures greatly reduced the amount of DNA co-migrating with supercoiled monomers and produced a ladder of retarded DNA species, most likely caused by successive entrapment by one, two, three and more S-K rings (Fig. 1D).

We propose that the ladder corresponds to multiple cohesin rings entrapping individual DNAs and not entrapment of multiple DNAs by individual cohesin rings for two reasons. First, retardation caused by entrapment within E-S compartments (see below), which contain only Smc1 and Smc3, is less than that caused by entrapment within S-K or E-K compartments, which contain Scc1 as well as Smc1 and Smc3 (see Fig. 5B). Second, dimeric plasmid DNA, which is frequently present in plasmid preparations, although largely absent from these gels due to our purification protocol, runs roughly at the top of the gel with respect to our figures. Thus, if our ladders represented entrapment of multiple DNAs by

individual cohesin rings, the DNA retardation should be much greater. Those DNAs retarded by entrapment within a single ring correspond to the CMs previously observed in vivo (Gligoris et al., 2014). Ladder formation required cysteine pairs at all three interfaces. It was never observed with linear DNA (Fig. S1D) or when just a single cysteine (Fig. 1E) or BMOE (Fig. 1F) was omitted. Crucially, the ladders were strictly dependent on addition of ATP (Fig. 1G).

Entrapment of DNAs by S-K rings requires Scc3 and is stimulated by Scc2

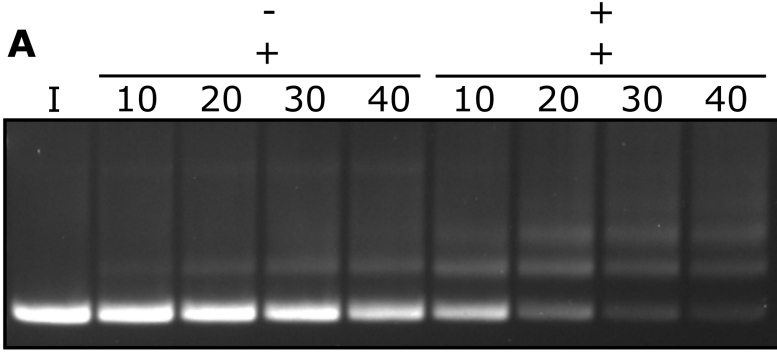
To assess the roles of Scc2 and Scc3, we measured ladder formation at four successive 10 min intervals in the presence and absence of the two proteins. Ladders indicative of entrapment increased with time (up to 40 min), suggesting that formation is a slow process, were greatly reduced by omission of Scc2 (Fig. 2A), and almost completely abolished by omission of Scc3 (Fig. 2B). In the absence of both Scc2 and Scc3, the level of entrapment was comparable to that observed in the presence of Scc2 alone (data not shown).

Entrapment of DNAs by S-K rings depends on ATP binding to Smc3 and on ATP hydrolysis

To address the role of cohesin's ATPase, we mutated Smc3's Walker A site (Smc3K38I) to abolish ATP binding to Smc3. This almost completely abolished entrapment (Fig. 2C). We did not test the effect of mutating the equivalent residue in Smc1 as this has previously been shown to abolish association of Smc1/3 heterodimers with Scc1 (Arumugam et al., 2003). We next tested the effect of mutating both Walker B sites to residues that permit ATP binding but strongly inhibit hydrolysis (Smc1E1158Q Smc3E1155Q, 'EQEQ') (Fig. S1B), which caused a more modest, albeit still significant, reduction (Fig. 2D). These data suggest that cohesin's ability to complete the ATP hydrolysis cycle stimulates entrapment but is not strictly necessary. To address whether Smc3's K112 K113 are also important we analysed the effect of substituting them by glutamine (Smc3K112Q K113Q), mutations thought to mimic the acetylated state. This also reduced S-K entrapment (Fig. S1C), an effect that parallels its abrogation of cohesin loading in vivo (Hu et al., 2015).

DNA binding to Scc3 is required for its entrapment by S-K rings

During a search for cohesin domains that bind DNA, we discovered that Scc3's association with a fragment of Scc1 containing residues 269-451 greatly stimulates its association with double stranded DNA, as measured using an electrophoretic mobility shift assay (EMSA)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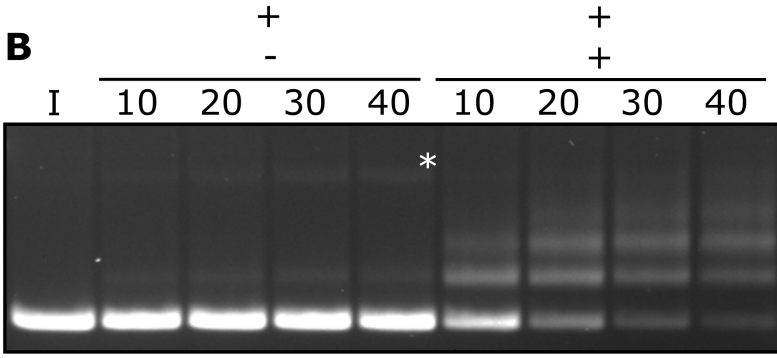
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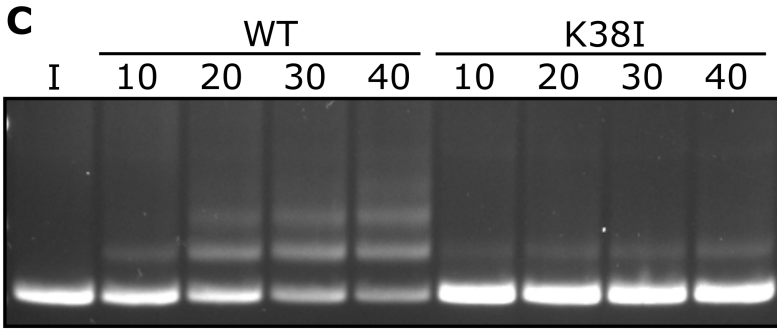
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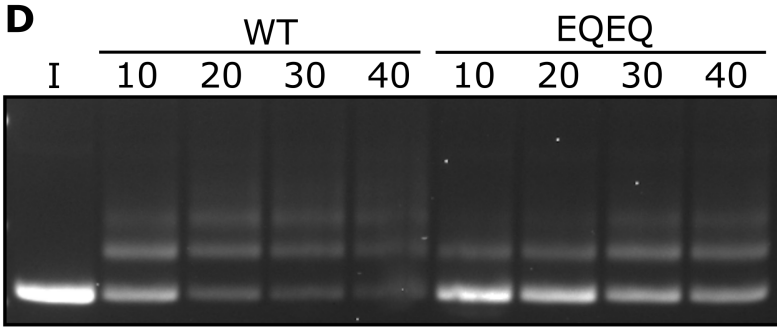
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Fig 2

Figure 2. Entrapment within S-K rings requires both Scc2 and Scc3, ATP binding to Smc3, and is stimulated by ATP hydrolysis

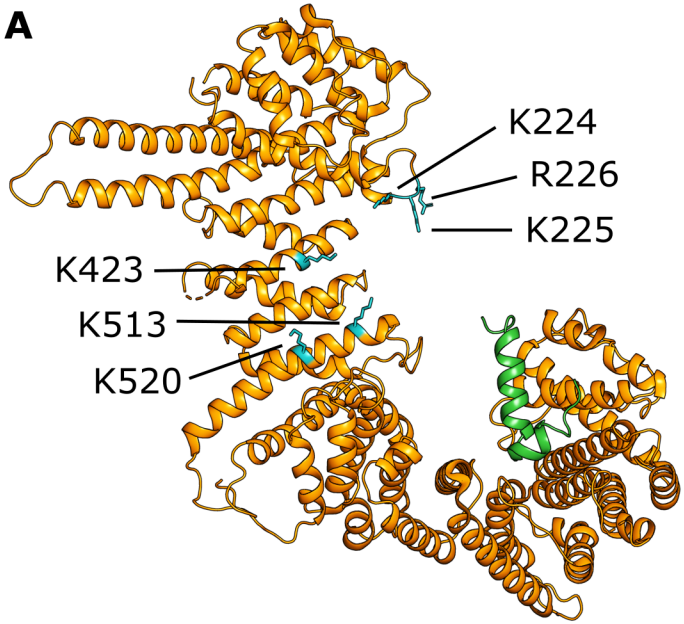
(**A**) Entrapment of DNA in S-K rings in the presence of Scc3, and the presence or absence of Scc2, or (**B**) the presence of Scc2, and the presence or absence of Scc3 (* = damaged open circular DNA). (**C**) DNA entrapment in the presence of Scc2 and Scc3, comparing WT cohesin to Smc3K38I (K38I), or (**D**) the effect of Smc1E1158Q Smc3E1155Q (EQEQ) in the presence of both Scc2 and Scc3. Entrapment assays incubated for 40 min with time points taken every 10 min (I = input DNA).

(Fig. S2A). Reasoning that Scc3/Scc1 complexes might bind DNA in a similar manner to that recently observed in a co-crystal of DNA bound to condensin's Ycg1 HAWK bound to its kleisin partner Brn1 (Kschonsak et al., 2017), we mutated two clusters of positively charged residues (Scc3K224E K225E R226E and Scc3 K423E K513E K520E) on opposite sides of the groove within Scc3 that is equivalent to Ycg1's DNA binding groove (Fig. 3A). Neither triple (3E) mutant eliminated DNA binding (Fig. S2B) nor caused lethality (Fig. S2E). Despite this, both reduced cohesin's association with all genomic sequences except point centromeres (*CEN*s) (Fig. S2C & D). In contrast, combining the two triple mutations (to create 6E) was lethal (Fig. S2E), abolished binding of Scc3 to DNA in the presence of Scc1 (Fig. 3B), and with the exception of *CEN*s eliminated cohesin's association with the genome (Fig. 3C & D).

Remarkably, cohesin containing Scc3-6E accumulated to exceptionally high levels at *CEN*s (Fig. 3C & D), which are the loading sites for most peri-centric cohesin (50 kb intervals surrounding *CEN*s). This distribution resembles that of Scc2 in wild type cells and indeed, *scc3-6E* had little or no effect on Scc2's accumulation with *CEN*s (Fig. 3E). This implies that cohesin containing Scc3-6E forms complexes with Scc2 at *CEN*s but subsequently fails to form a stable association with chromatin or translocate into neighbouring sequences. Our ability to detect such complexes at *CEN*s but not at other loading sites along chromosome arms can be attributed to the fact that Scc2's partner Scc4 binds to the kinetochore protein Ctf19 and this association transiently tethers complexes at *CEN*s while they are attempting to load (Hinshaw et al., 2017). Though accumulation of cohesin bound by Scc2 at *CEN*s does not depend on Scc3's ability to bind DNA, it does still require Scc3 (Fig. S2F). Crucially, cohesin containing Scc3-6E failed to support entrapment of DNAs inside S-K rings in vitro (Fig. 3F). During the course of our work, a crystal structure of DNA bound to a Scc3/Scc1 complex confirmed that it does indeed bind DNA (Li et al., 2018 PDB 6H8Q) in a manner resembling that of Ycg1. Moreover, K224, K225 R226, K423, K513, and K520 are all predicted to contribute to the association. These data imply that Scc3's ability to bind DNA has an important role in cohesin's ability to load onto and translocate along chromosomal DNA in vivo, as well as entrap in S-K rings in vitro.

DNA binding to Scc2 facilitates entrapment by S-K rings

The *S. pombe* Scc2/4 complex has previously been shown to bind DNA in vitro (Murayama and Uhlmann, 2014) but the physiological significance of this activity has never been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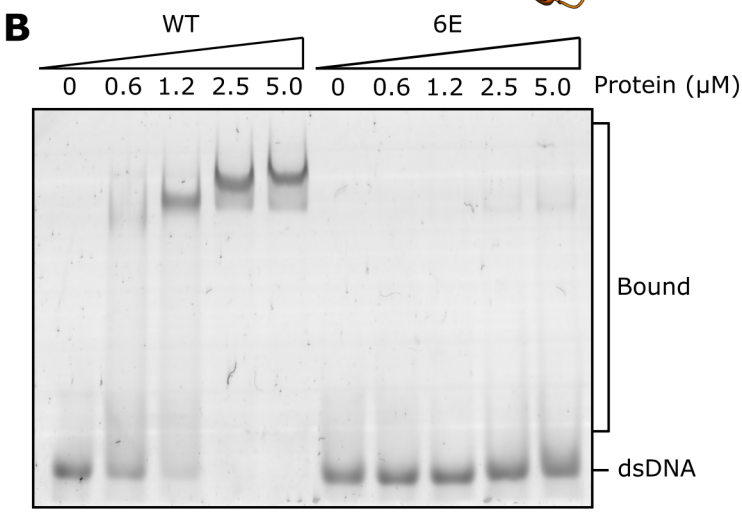
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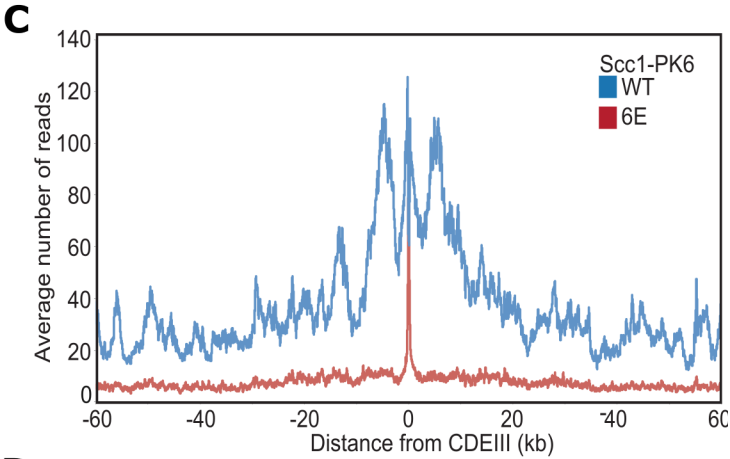
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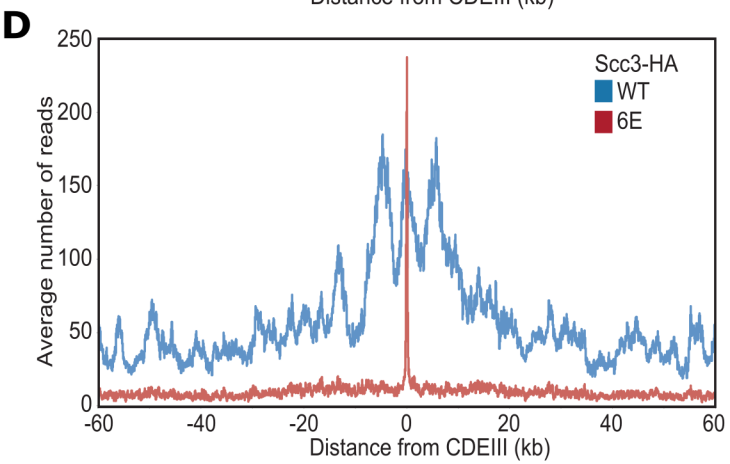
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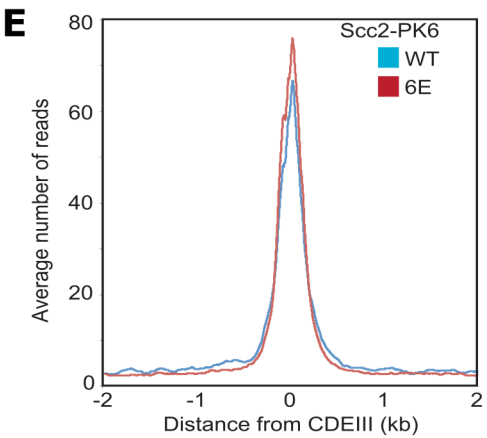
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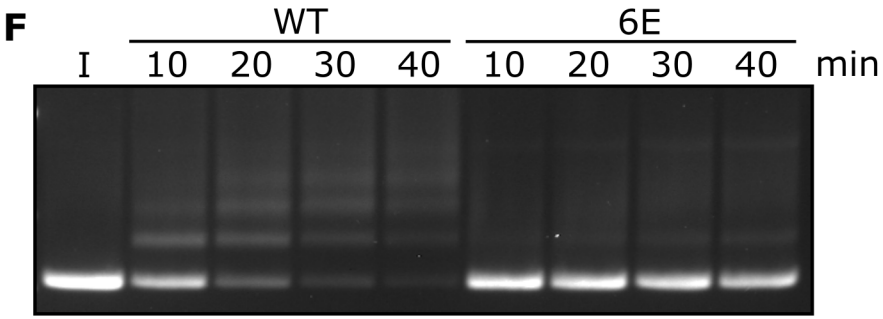
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Fig 3

Figure 3. DNA binding to Scc3 is required for its entrapment by S-K rings

(**A**) Structure of *S. cerevisiae* Scc3 (orange) protein in complex with a fragment of Scc1 (green) (PDB 6H8Q). Labelled are the six residues within the DNA binding groove of Scc3 that were mutated to glutamate (Scc3-6E). (**B**) EMSA comparing the ability of WT Scc3-Scc1$^{269-451}$ and Scc3-6E-Scc1$^{269-451}$ complexes to bind dsDNA. (**C**) Average calibrated ChIP-seq profiles of Scc1-PK6 60 kb either side of *CEN*s in the presence of ectopic WT Scc3 (KN27821) or Scc3-6E (KN27804). Cells were arrested in G1 with α-factor prior to release into auxin and nocodazole containing media at 25°C to deplete the endogenous Scc3. ChIP-seq samples were taken 60 min after release. (**D**) Average calibrated ChIP-seq profile of ectopic WT (KN27796) or mutant (KN27802) Scc3-HA performed as in C. (**E**) Average calibrated ChIP-seq profile of Scc2-PK6 in the presence of ectopic WT Scc3 (KN27821) or Scc3-6E (KN27804). Experiment was performed as in C. (**F**) Entrapment of DNA within S-K rings in the presence Scc2 and either WT Scc3 or Scc3-6E. Entrapment assay incubated for 40 min with time points taken every 10 min (I = input DNA).

investigated. EMSA revealed that *S. cerevisiae* Scc2 also binds DNA (Fig. 4A), as do Scc2/4 complexes with slightly higher affinity (Fig. S3A). Unlike Scc3, whose DNA binding was greatly enhanced by Scc1, DNA binding by Scc2 was reduced by addition of a Scc1 fragment ($Scc1^{150-298}$) that contains sequences necessary for Scc2-dependent loading in vivo (Fig. 4A) (Petela et al., 2018). Interestingly, the inhibitory effect of $Scc1^{150-298}$ was not observed in the binding of DNA to full length Scc2/4 (Fig. S3A), suggesting that DNA binding sites also exist in Scc4, or in sequences N-terminal of the deletion in our Scc2C construct.

An alignment of the crystal structure of *E. gossypii* Scc2 (Chao et al., 2017) with that of Ycg1/Brn1bound to DNA (Kschonsak et al., 2017) revealed not only a remarkable similarity in the overall shape of their hook-shaped HEAT repeats but also a set of potential DNA binding residues on the surface of the shallow concave groove corresponding to Ycg1's DNA binding pocket (Petela et al., 2018) (Fig. 4B). Four of these are particularly conserved and correspond to S717, K721, K788, and H789 in *S. cerevisiae*. Both *scc2S717L K721E* and *scc2K788E H789E* double mutants are lethal and abolish loading of cohesin throughout most of the genome (Petela et al., 2018) (Fig S3C). To test whether these residues participate in binding DNA, we used EMSAs to measure the effect on DNA binding of mutating the above residues to glutamate. Both Scc2S717E K721E and Scc2K788E H789E double mutants reduced binding (Fig. S3B) but caused only a modest reduction in S-K entrapment (data not shown). In contrast, the quadruple mutant Scc2S717E K721E K788E H789E (Scc2-4E) not only greatly reduced DNA binding (Fig. 4C) but also S-K entrapment (Fig. 4D). These results suggest that Scc2's ability to bind DNA has a crucial role in entrapping DNA within S-K rings in vitro, an activity also required for loading cohesin onto chromosomes in vivo (Petela et al., 2018) (Fig. S3C). They also demonstrate that the stimulation of DNA entrapment within S-K rings by Scc2 is not merely an adventitious property of Scc2 but an activity dependent on conserved surface residues that have unambiguous physiological functions.

DNA is never entrapped in J-S and only rarely in J-K compartments

Using the Smc1S161C-Smc3K160C cysteine pair, cohesin's ATPase heads can also be efficiently crosslinked in the J-state (Chapard et al., 2019). Moreover, this crosslinking can be combined with simultaneous crosslinking of N- and C-terminal kleisin domains to Smc3 and Smc1 ATPase heads respectively, to measure entrapment within J-K compartments

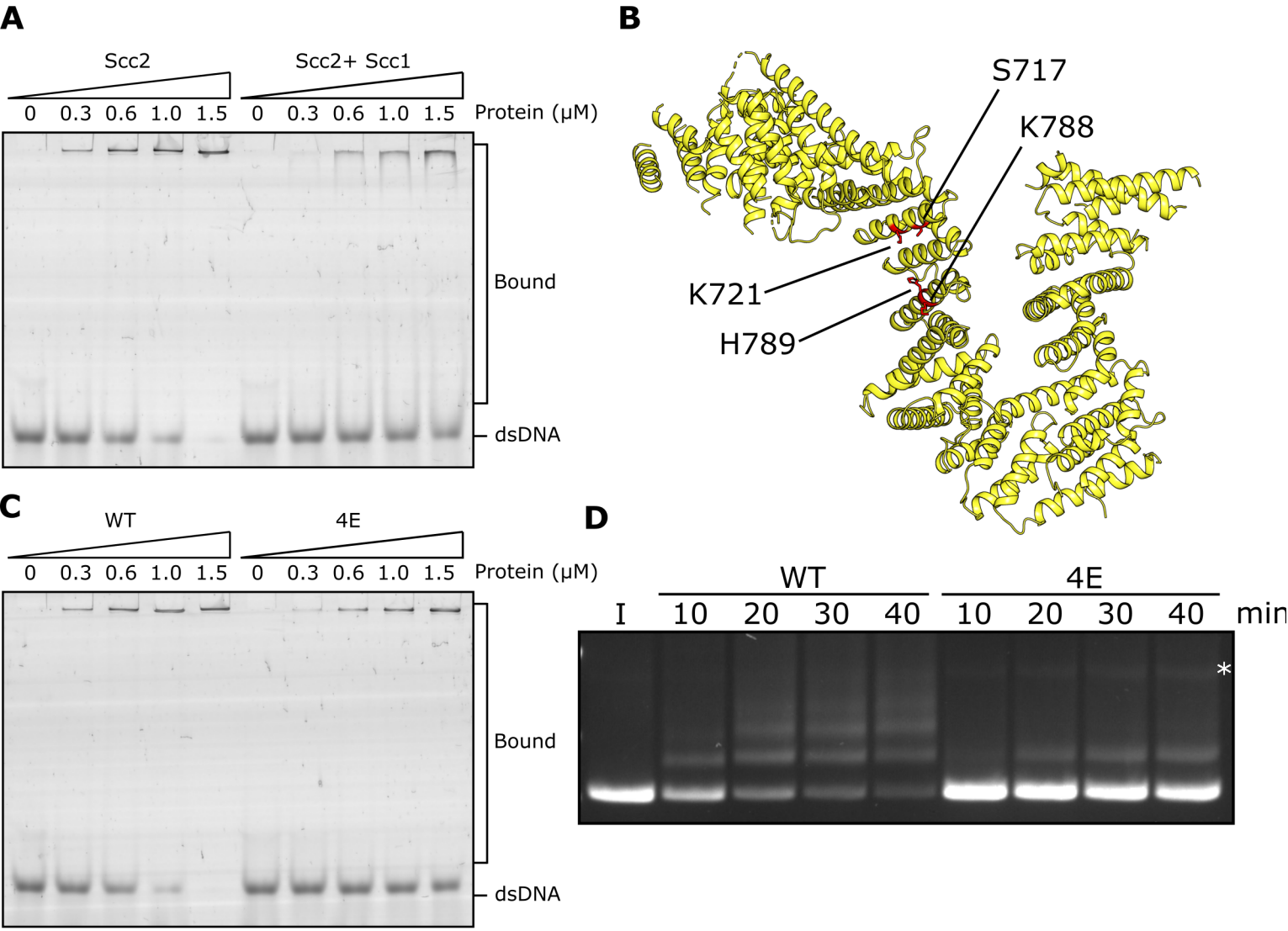

A
Scc2
Scc2+ Scc1
0 0.3 0.6 1.0 1.5 0 0.3 0.6 1.0 1.5
Protein (μM)
Bound
dsDNA
B
S717
K788
K721
H789
C
WT
4E
0 0.3 0.6 1.0 1.5 0 0.3 0.6 1.0 1.5
Protein (μM)
Bound
dsDNA
D
WT
4E
I 10 20 30 40 10 20 30 40 min
*

Fig 4

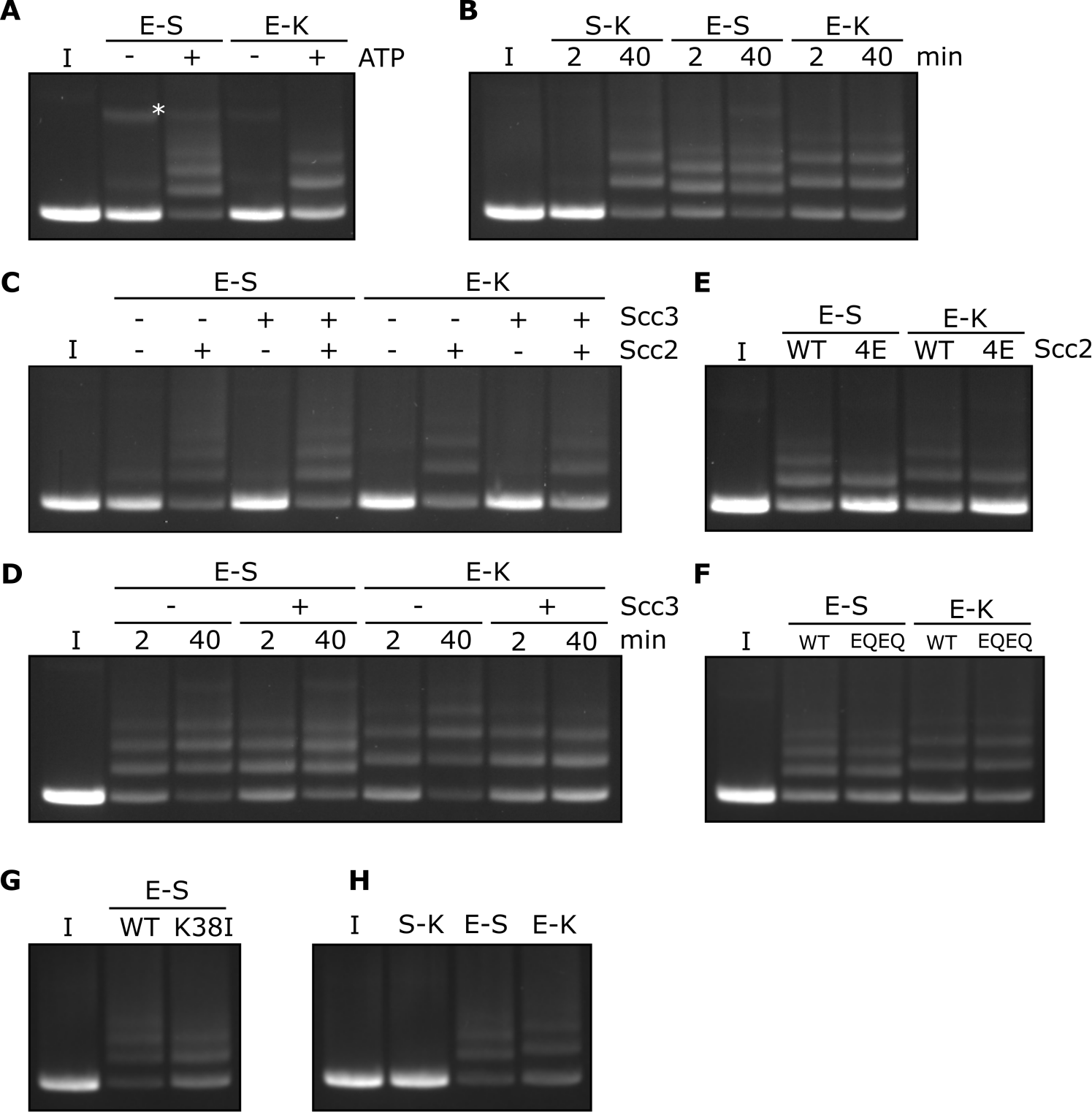
Figure 4. DNA binding to Scc2 facilitates entrapment by S-K rings

(**A**) EMSA comparing the ability of Scc2 and Scc2-Scc1$^{150-298}$ complexes to bind dsDNA. (**B**) *S. cerevisiae* Scc2 from the cryo-EM structure (Fig. 8) with the four resides within the putative DNA binding surface labelled that were mutated to glutamate (Scc2-4E). (**C**) EMSA comparing the ability of Scc2 and Scc2-4E complexes to bind dsDNA. (**D**) Entrapment of DNA in S-K rings in the presence of Scc3 and either Scc2 or Scc2-4E. Entrapment assay incubated for 40 min with time points taken every 10 min (* = damaged open circular DNA; I = input DNA).

(Fig. S4A), or with simultaneous crosslinking of the hinge (Fig. S4B), to measure entrapment of DNAs in J-S compartments (Fig. 1A ii). J crosslinking alone or in combination with hinge (J-S) or kleisin (J-K) was efficient even in the presence of ATP, DNA, Scc2, and Scc3 (Fig. S4C). In other words, both J-S and J-K circularisation occurred efficiently under conditions that promote efficient entrapment of DNAs inside S-K rings. However, DNAs were never entrapped within J-S compartments and only rarely by J-K ones (Fig. S4D). J-K entrapment was not only much less frequent than S-K entrapment but also independent of Scc2. The fact that J-K entrapment was comparable to S-K entrapment in the absence of Scc2 (compare Fig. 2A and S4D) suggests that the low-level entrapment of DNAs in S-K rings induced by Scc3 alone may in fact correspond to DNAs entrapped in J-K compartments. Though J-K circularisation by BMOE is modestly lower than that of S-K, this cannot account for its far lower DNA entrapment. We therefore suggest that most ATPase heads associated with DNA entrapped within S-K rings in vitro are not juxtaposed. They are either fully disengaged, in the E-state, or in some other conformation.

Rapid DNA entrapment in E-S and E-K compartments

We used the same approach to measure entrapment in E-S or E-K compartments (Fig. 1A iii), in this case replacing J-specific cysteines by a pair specific for the E-state (Smc1N1192C-Smc3R1222C). Unlike J crosslinking, which was readily detected in cohesin trimers, efficient E-state crosslinking was dependent on the presence of ATP (Fig. S5A). As with J-, E-state crosslinking can be combined with simultaneous crosslinking of N- and C-terminal kleisin domains to Smc3 and Smc1 ATPase heads respectively, to measure entrapment within E-K compartments (Fig. S5B), or with simultaneous crosslinking of the hinge (Fig. S5C), to measure entrapment of DNAs in E-S compartments. As previously reported (Chapard et al., 2019), Smc1/3 dimers crosslinked simultaneously at the hinge and engaged heads co-migrate with those crosslinked at the hinge alone, which hinders detection of E-S circularisation directly. Given that double crosslinking has been detected in vivo using differently tagged proteins (Chapard et al., 2019) and that DNAs are readily entrapped by Smc1/3 dimers containing hinge and E-specific cysteine pairs treated with BMOE, we can reason that efficient double crosslinking does indeed occur. DNAs were entrapped in an ATP-dependent fashion in both E-S and E-K compartments in the presence of Scc2 and Scc3 (Fig. 5A). Notably, both processes occurred much more rapidly than S-K entrapment, with significant amounts of DNA entrapped by multiple rings within 2 min (Fig. 5B). Because S-K entrapment occurs much more slowly, the efficient entrapment of DNAs


A
E-S
E-K
I
-
+
-
+
ATP
*
B
S-K
E-S
E-K
I
2
40
2
40
2
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E-S
E-K
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Scc3
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Scc2
D
E-S
E-K
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Scc3
I
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40
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40
2
40
2
40
min
F
E-S
E-K
I
WT
EQEQ
WT
EQEQ
G
E-S
I
WT
K38I
H
I
S-K
E-S
E-K

Fig 5

Figure 5. Rapid DNA entrapment in E-S and E-K compartments

(**A**) Entrapment of DNA in E-S/E-K compartments in the presence of Scc2 and Scc3, and the presence or absence of ATP, incubated for 40 min (* = damaged open circular DNA). (**B**) DNA entrapment in S-K rings, or E-S/E-K compartments in the presence of Scc2 and Scc3, incubated for either 2 or 40 min. (**C**) DNA entrapment in E-S/E-K compartments in the presence of Scc2, Scc3, Scc2 and Scc3, or absence of both, incubated for 2 min. (**D**) DNA entrapment in E-S/E-K compartments in the presence of Scc2, and either the presence or absence of Scc3, incubated for either 2 min or 40 min. (**E**) DNA entrapment in E-S/E-K compartments in the presence of either Scc2 or Scc2-4E, or (**F**) Entrapment in the presence of Scc2 alone, comparing WT and Smc1E1158Q Smc3E1155Q (EQEQ) cohesin, incubated for 2 min. (**G**) DNA entrapment in E-S compartments in the presence of Scc2 comparing WT and Smc3K38I (K38I) cohesin, or (**H**) Entrapment of DNAs in S-K rings, or E-S/E-K compartments in the presence of Scc2, incubated for 2 min (I = input DNA).

inside E-S/E-K compartments within a few minutes is presumably not accompanied by S-K entrapment. Though it occurs efficiently in vitro, entrapment of circular DNAs by cohesin in E-S compartments has not so far been detected in vivo, although *Bacillus subtilis* SMC possessing Walker B mutations have been shown to have such an activity inside cells (Vazquez Nunez et al., 2019).

Entrapment within the E-S and E-K compartments depends on Scc2 but not Scc3

In contrast to entrapment within S-K rings, which depends on Scc3, the rapid entrapment of DNA in the E-S/E-K compartments was Scc3 independent (Fig. 5C). However, it was highly dependent on Scc2, both in the presence or absence of Scc3 (Fig. 5C). Levels of E-S entrapment increased between 2 and 40 min in the presence of Scc2, as well as in the presence of Scc2 and Scc3 (Fig. 5D). However, while a similar result was seen for E-K entrapment in the presence of Scc2 alone, this increase was not observed in the presence of both Scc2 and Scc3 which instead showed no increase by the longer time point and possibly even a small reduction.

The rapid entrapment of DNAs within E-S/E-K compartments in the presence of Scc2 was reduced by Scc2-4E (Fig. 5E), suggesting that the reaction at least partly depends on Scc2's ability to bind DNA. Strikingly, both types of entrapment were unaffected by Smc1E1158Q *S*mc3E1155Q mutations (EQEQ), implying that neither form of entrapment requires ATP hydrolysis (Fig. 5F). In contrast to S-K entrapment in the presence of both Scc2 and Scc3 (Fig. 2C), E-S entrapment in the presence of Scc2 alone was only modestly reduced by Smc3K38I, implying that ATP bound merely to Smc1's ATPase head is sufficient (Fig. 5G). Indeed, Smc3K38I does not prevent E-specific crosslinking under these reaction conditions, namely in the presence of ATP, DNA, and Scc2 (Fig. S5D) and its modest reduction of E-S entrapment is in line with its effect on E-state crosslinking (Fig. 5G & S5D). Though it does not abolish head engagement, Smc3K38I clearly compromises the process. As long as ATP is present, neither DNA nor Scc2 are required for efficient E-state crosslinking of wild type complexes but both are important for Smc3K38I complexes (Fig. S5D). Smc3K38I presumably destabilises head engagement in a manner that can be overcome by the presence of Scc2 and DNA. To explore whether Scc2 and DNA also promote head engagement of otherwise wild type complexes, we tested their effect when ATP's ability to promote head engagement is compromised by omission of Mg^{2+} (Fig. S5E). Under these circumstances, addition of both Scc2 and DNA restored efficient head engagement and both

factors were required for this effect. Our finding that Scc2 and DNA collaborate to promote ATP-dependent head engagement suggests that DNA binds to a site created by head engagement as well as to Scc2.

Scc2 causes DNAs to be entrapped in E-S and E-K compartments without entering S-K rings

Because Scc3 is crucial for S-K entrapment, the rapid entrapment of DNAs within E-S and E-K compartments in the presence of Scc2 alone should be unaccompanied by S-K entrapment. This is indeed the case. In contrast with E-S or E-K entrapment, which is very efficient, few if any DNAs are entrapped in S-K compartments by 2 min (Fig. 5H). Though paradoxical, this striking observation has a very simple explanation. The similarity in kinetics suggests that E-S and E-K entrapments are created simultaneously as part of the same reaction. In other words, a single type of DNA passage followed by head engagement gives rise to both types. We envisage two types of mechanism to explain how this occurs without S-K entrapment. According to the first (and simplest), DNA moves “upwards” between disengaged ATPase heads, and is subsequently trapped in the E-S compartment following ATP-driven head engagement (Fig. 9A). An alternative is that a loop of DNA is inserted into an open S-K ring. If one of the loop’s segments were located above the ATPase domains while the other below, then subsequent head engagement would lead to simultaneous entrapment in both E-S/E-K compartments (Fig. 9B). Neither type of DNA movement involves passage through a gate created by opening the S-K ring, hence explaining the lack of S-K entrapment. Entrapment within E-S/E-K, but not S-K compartments, in the presence of Scc2 alone was also observed with relaxed (nicked) DNAs (data not shown).

Entrapment of DNA within E-S compartments disrupts coiled coil interactions proximal to the heads

To address whether the coiled coils are associated when Smc1/3 heads engage in vitro in the presence of Scc2, we combined the E-specific cysteine pair with one specific for the coiled coils (Smc1K201C-Smc3K198C), in close proximity to the joint (Fig. 6A). This revealed that double crosslinking can indeed occur in the presence of ATP (Fig. 6B). As expected, double crosslinking also occurred efficiently when the coiled coil pair was combined with one within the hinge interface (Smc1K639C Smc3E570C). These cysteine pair combinations enabled us to measure entrapment within two sub-compartments within the E-S compartment (Fig.6A iii): one created by simultaneous crosslinking of the hinge and

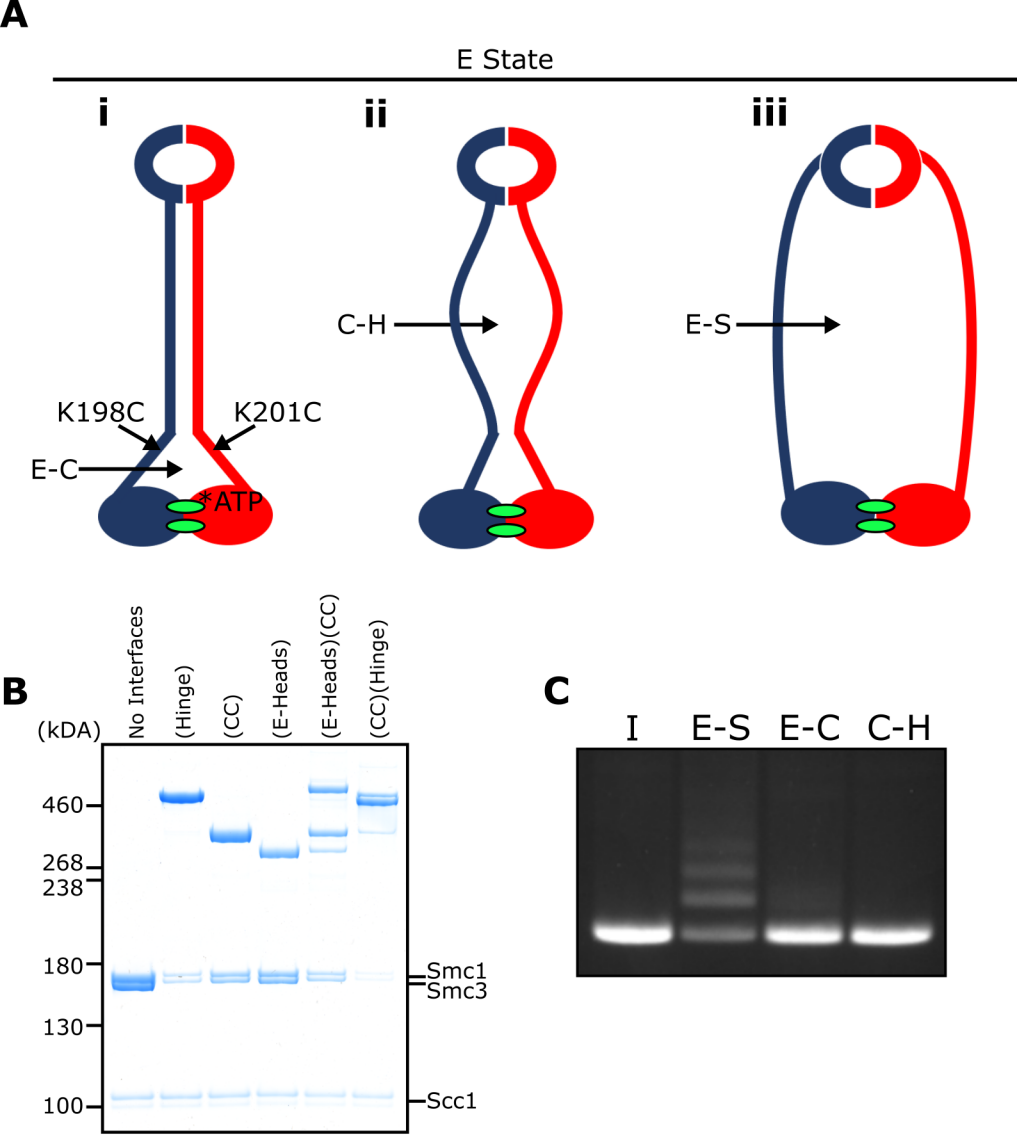

A
E State
i
ii
iii
K198C
K201C
E-C
*ATP
C-H
E-S
B
(kDA)
No Interfaces
(Hinge)
(CC)
(E-Heads)
(E-Heads)(CC)
(CC)(Hinge)
460
268
238
180
130
100
Smc1
Smc3
Scc1
C
I
E-S
E-C
C-H

Fig 6

Figure 6. Scc2 and DNA disrupt the J-state and E-S/E-K entrapment leads to dissociation of the coiled coil around the joint

(**A**) Scheme showing the location of the joint cysteine pair and how head engagement could lead to different degrees of coiled coil dissociation and sub-compartment formation. (**B**) BMOE crosslinking of cohesin containing cysteine pairs at the specified interfaces in the presence of ATP. CC = coiled coils. (**C**) DNA entrapment in E-S compartments, or either E-C or C-H sub-compartments, in the presence of Scc2, incubated for 2 min (I = input DNA).

coiled coils (C-H compartment) (Fig. 6A ii) and a complementary one made by the simultaneous crosslinking of coiled coils and engaged heads (E-C compartment) (Fig. 6A i). If DNAs entrapped in E-S compartments are in molecules whose coiled coils are associated, at least in the vicinity of their joint regions, then they must be entrapped either in the E-C or the C-H sub compartments. On the other hand, if the entrapment of DNAs within E-S compartments is accompanied by (or indeed causes), dissociation of the coiled coils in the vicinity of the Smc1K201C-Smc3K198C cysteine pair, then DNA should not be trapped in either of these sub-compartments.

Despite efficient crosslinking at both cysteine pairs (Fig. 6B), few if any DNAs were entrapped in the presence of Scc2 and ATP following BMOE treatment of cohesin trimers containing hinge and coiled coil cysteine pairs (Fig. 6C). Likewise, few if any DNAs were entrapped by cohesin trimers containing both E-state and coiled coil cysteine pairs. We deduce from this result that entrapment of DNAs within E-S compartments in the presence of Scc2 is accompanied by dissociation of their coiled coils in a manner that precludes crosslinking between Smc1K201C and Smc3K198C. It is important to point out that this feature was not apparent when analysing the crosslinking efficiency between Smc1K201C and Smc3K198C under the same conditions (data not shown), namely in the presence of Scc2, ATP and DNA. To explain this, we suggest that despite the addition of DNA, complexes exist, at least transiently, that have engaged their heads and have zipped up their coiled coils but have not in fact trapped DNA within their E-S compartments. Consistent with this notion is our finding that simultaneous crosslinking of coiled coils and engaged heads occurs efficiently even in the absence of DNA (Fig. 6B).

The DNA is 'clamped' between Scc2 and the engaged heads during entrapment within E-S/E-K compartments, as revealed by cryo-EM

The dissociation of Smc1 and Smc3 coiled coils in the vicinity of their joint regions would create space for DNA to bind to engaged heads, as observed in Mre11/Rad50 complexes (Liu et al., 2016) and more recently in both human and *S. pombe* cohesin containing $Scc2^{NIPBL/Mis4}$ and $Scc3^{SA2/Psc3}$ (Higashi et al., 2020; Shi et al., 2020). In the latter structures, the coiled coils of Smc1 and Smc3 diverge from each other at 70° as they emerge from the engaged heads, thereby creating a site for DNA to bind a surface on top of the heads. The DNA is also bound by Scc2, whose simultaneous association with Smc1's ATPase head

and the coiled coil emerging from Smc3's head creates a new type of sub compartment within which DNA bound to the engaged heads is entrapped or 'clamped'.

Our findings that Scc2 and DNA together promote head engagement (Fig. S5D & E) and that Scc2-4E reduces E-S/E-K entrapment (Fig. 5E) raise the possibility that DNAs entrapped within E-S/E-K compartments through the action of Scc2 are bound in a similar manner. However, since the human (PDB 6WG3) and *S. pombe* cryo-EM complexes (Higashi et al., 2020; Shi et al., 2020) were formed by cohesin containing Scc3$^{SA2/Psc3}$, the complexes described here, which are formed in the absence of Scc3, could in fact have a very different conformation. Furthermore, because it is not possible to trace the entire kleisin chain, and because the complexes contain only short linear DNA molecules, the topology of DNA's association with the SMC-kleisin trimers in the existing structures cannot be inferred definitively. In other words, it is not possible to determine whether the DNAs in the structures are entrapped within E-S and E-K and/or S-K compartments. As it happens, the kleisin path deduced for PDB 6WG3 (Shi et al., 2020) suggests that DNA, if it were circular, would be trapped in E-S and S-K but not E-K compartments. Thus, the presence of Scc3^{SA2} in this complex may have had an important influence on the topology of cohesin's association with DNA.

To elucidate how DNA actually associates with EQEQ cohesin trimers in the presence of ATP and Scc2, but lacking Scc3, namely under conditions in which DNAs are clearly entrapped within E-S and E-K but not S-K compartments, we used cryo-EM to solve the structure of EQEQ cohesin trimer (Smc1, Smc3 and Scc1) bound to ATP, linear DNA (40 bp), and Scc2C2 (residues 151-1493) to a resolution of 3.4 Å (Fig. 7A). Processing followed standard cryo-EM single particle procedures as implemented in RELION 3.1 pipelines (Scheres, 2012), but significant preferred orientation required the use of tilted data acquisition (Methods). With the help of previous crystal structures (PDBs 5ME3, 1W1W and 4UX3) (Chao et al., 2017; Gligoris et al., 2014; Haering et al., 2004) the electron density map enabled us to build and refine a reliable atomic model, with DNA rigidly clamped between the head domains and the HAWK subunit Scc2 (Table 1, Fig. 8A).

A key question arising from our crosslinking studies concerns the mechanism by which circular DNA is entrapped within E-S/E-K compartments without being entrapped within the S-K ring, namely whether a single segment of DNA is passed between the heads prior to their

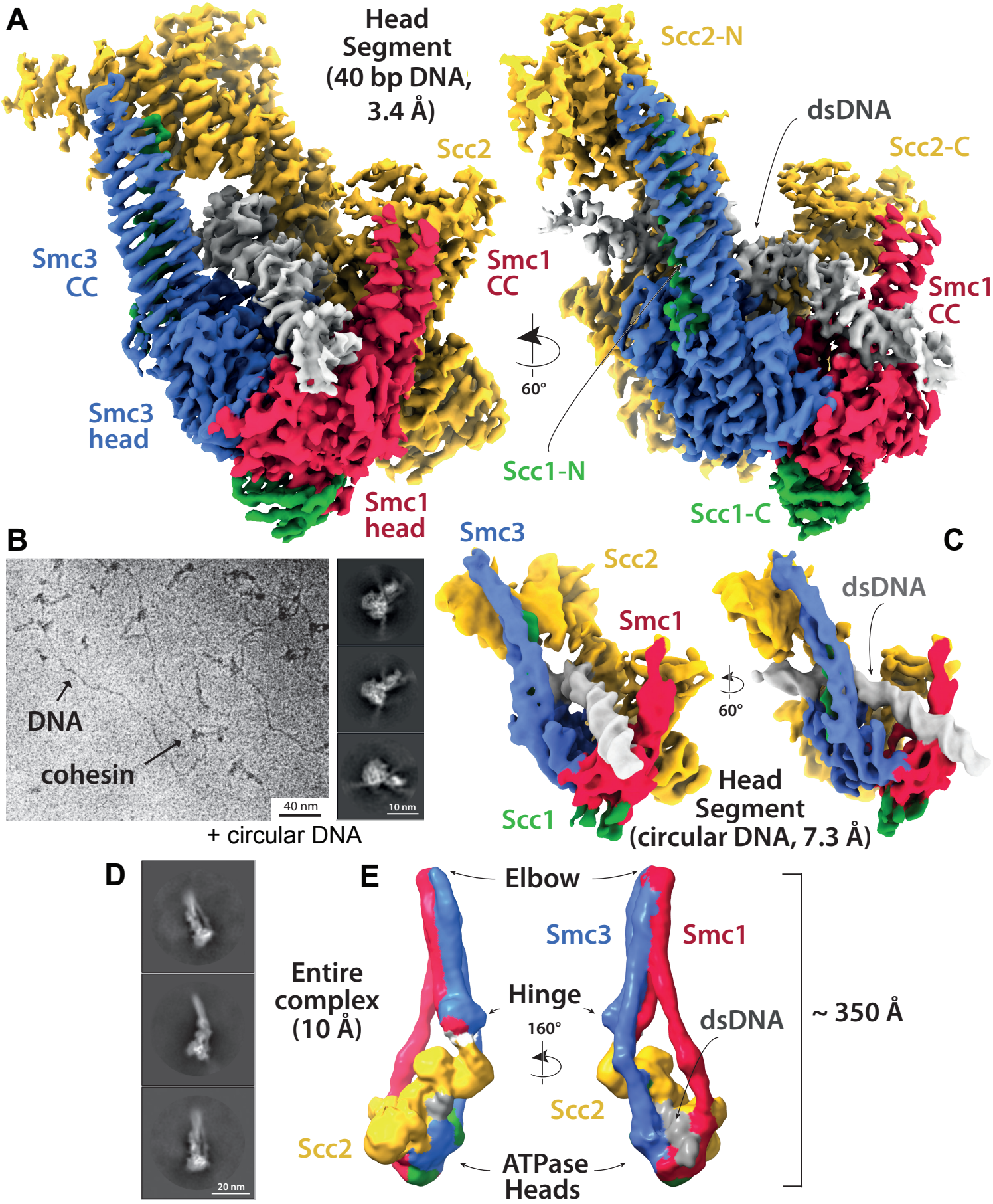

A
Head Segment (40 bp DNA, 3.4 Å)
Scc2
Smc3 CC
Smc1 CC
Smc3 head
Smc1 head
Scc2-N
dsDNA
Scc2-C
Smc1 CC
60°
Scc1-N
Scc1-C
B
DNA
cohesin
40 nm
10 nm
+ circular DNA
Smc3
Scc2
Smc1
Scc1
C
dsDNA
60°
Head Segment (circular DNA, 7.3 Å)
D
20 nm
E
Elbow
Smc3
Smc1
Entire complex (10 Å)
Hinge
160°
dsDNA
~ 350 Å
Scc2
Scc2
ATPase Heads

Fig 7

Figure 7. Cryo-EM of cohesin clamping DNA in the E-S/E-K state

(**A**) Cryo-EM map of 40 bp DNA clamped by Scc2- and ATP-bound cohesin EQEQ trimer at 3.4 Å resolution. Both front and side views are coloured by subunit. (**B**) Same complex as shown in A but bound to ~1.8 Kbp relaxed circular DNA as a cryo-EM field view (using Volta phase plate, left) and a selection of 2D class averages (right) clearly showing DNA emanating from cohesin/Scc2 complexes. (**C**) 7.3 Å resolution cryo-EM map of the complex shown in B, coloured by subunit, demonstrating that the same conformation of the complex has been obtained as with linear DNA (panel A). Same orientations and colours as in A. (**D**) 2D class averages obtained by reprocessing of the same data set as used for A with an enlarged box size show the position of the coiled coils and the hinge. (**E**) ~10 Å resolution cryo-EM map of the entire tetramer complex as shown in D. Since we used the same complex as used in the in vitro entrapment reactions, we can deduce that the DNA within the clamped structure depicted in A, C and E must be entrapped in both the E-S and E-K compartments.

engagement or whether a loop of DNA is first passed through the S-K ring before head engagement traps one segment of the loop above and another below the heads. Visualising how DNA is actually grasped by cohesin and Scc2 under these conditions should in principle be revealing. However, the linearity of the 40 bp oligonucleotide used for our high-resolution structure (Fig. 7A) precludes any conclusions as to the topology of its association with cohesin. In other words, we cannot say whether it corresponds to E-S/E-K entrapment. It also precludes any insight as to how DNA actually enters the Scc2-SMC clamp because the DNA could either have been passed through the heads prior to their engagement (or any other gate), or it could simply have been threaded through the clamp after head engagement. For this reason, we also solved to a resolution of ~7 Å the structure of the same clamped state associated with circular relaxed DNA (Table 1, Fig. 7B & C). Crucially, the structure associated with circular DNA is virtually identical to that associated with the linear oligonucleotide and since the former is known to involve E-S/E-K but not S-K entrapment, we can with some certainty infer the path of the kleisin chain with respect to the DNA (Fig. 8F). Two important conclusions can therefore be drawn. First and foremost, E-S/E-K entrapment does indeed arise from the clamping of DNA between Scc2 and engaged heads in the manner revealed by both high (Fig. 7A) and medium (Fig. 7B) resolution cryo-EM structures. Second, because the circular DNA is demonstrably not highly-bent when clamped (Fig. 7B, right), the DNA must have entered the clamped state and thereby been entrapped in E-S/E-K compartments without formation of a loop (Fig. 9B). In other words, a single segment of DNA must have been passed between the heads prior to their engagement (Fig. 9A).

Processing of the linear DNA dataset using boxes large enough to cover entire cohesin complexes (Table 1, Methods) revealed a significant subset of particles (~30%) that contained lower-resolution information about the conformation of the coiled coils and placement of the hinge (Fig. 7D & E). Despite partial head-proximal unzipping of the coiled coils in this state, the reconstructed map clearly revealed a folded conformation reminiscent of *apo* condensin, a fraction of ATP-bound condensin (Lee et al., 2020) and *apo* cohesin (Bürmann et al., 2019). A folded state was also presumed to exist in the human cohesin structure where the HAWKS and the hinge interacted directly, but the coiled coils could not be resolved (Shi et al., 2020). Our map did not have sufficient resolution to determine whether the hinge was partially open or closed (Fig. 7E).

Binding of DNA to Scc2 is consistent with Scc2-4E effects

Binding of DNA by Scc2 in *S. cerevisiae* cohesin occurs in a similar manner to that recently described for human Scc2^{NIPBL} (Shi et al., 2020) (Fig. 8A). Scc2 holds DNA by way of a curved basic and polar surface located around the transition between its neck and head regions (Fig. 8B). The surface, which causes the DNA to bend slightly (~9°), is created by the spatial arrangement into a semicircle of a series of residues (e.g. S508, N557, K714, S753, S783, and K1324/25) from the ends of six α-helices and one loop (containing K427) that together engulf the phosphate backbone. This region includes all four positions mutated in Scc2-4E (S717E K721E K788E H789E, Fig. 4), thus neatly explaining why the charge-reversing mutations lowered the binding affinity for DNA and inhibited entrapment of DNAs in vitro.

Scc2 binds both Smc1 and Smc3

Clamping involves not only the binding of DNA to Scc2 and to engaged heads (see below) but also entrapment in a novel compartment created by Scc2's association with both Smc1 and Smc3. The latter involves multiple binding sites (Fig. 8C) and is therefore much more delocalized than Scc2's DNA binding. Highly prominent is the binding of Scc2 to Smc1 through the docking of HEAT repeats 18-24 (residues 1127-1493) onto the F-loop (residues 1095-1118) on Smc1 head's C-lobe and the coiled coils that emerge above (Fig. 8C i). This mode of association is highly analogous to the binding of *S. cerevisiae* condensin Smc4 by the HAWK Ycs4 (Lee et al., 2020) (Fig. S6). Scc2 simultaneously interacts with Smc3's N-lobe - thereby providing a mechanism by which it promotes head engagement and subsequent ATP hydrolysis. One key contact in this regard involves residues in an otherwise disordered loop (1178-1203) that form a β-strand which docks onto the end of the central β-sheet of the Smc3 head (Fig. 8C iii). More conserved is a major contact mediated by salt bridges between a collection of highly conserved aspartate and glutamate residues located within two loops of Scc2 (819-EDEED-823 and 781-DD-782) and two key lysine residues (K112 K113) in Smc3 (Fig. 8C ii). Entrapment of DNA between Scc2 and engaged heads arises because in addition to the above contacts, Scc2 contacts Smc3 through HEAT repeats 1-4 (residues 151-409), which bind to the start of the joint module (coiled coil arm, residues 999-1004) in a manner that — when compared to the unbound crystal structure (Kikuchi et al., 2016) — causes a conformational rearrangement of Scc2's head segment. This movement is necessary to accommodate the simultaneous binding of DNA by both Scc2 and the heads while presumably playing a role in stabilising the unzipped conformation of the coiled coils (Fig. 8C iv). Several residues within this interface (for example, Smc3

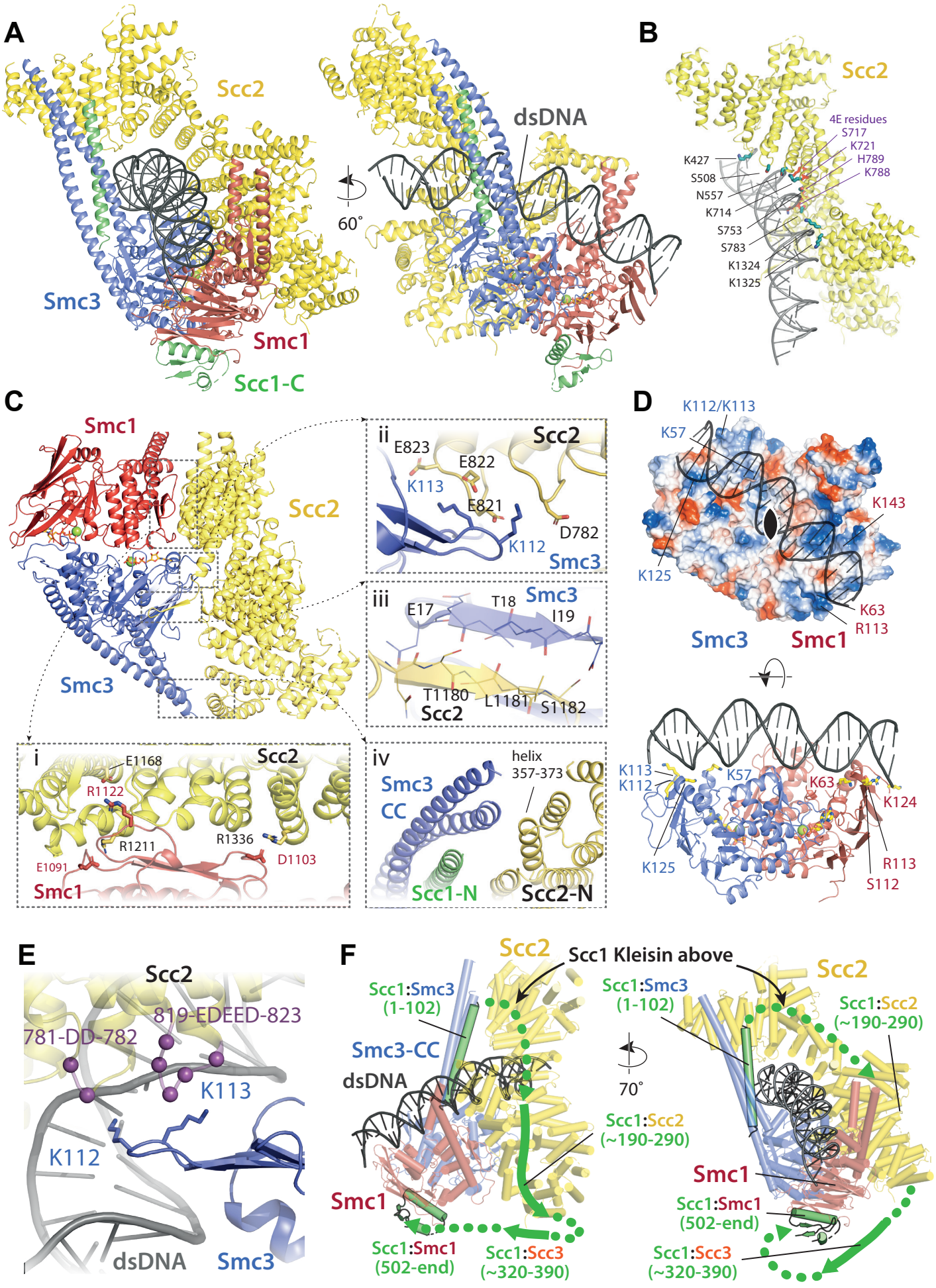

A
Scc2
dsDNA
60˚
Smc3
Smc1
Scc1-C
B
Scc2
4E residues
S717
K721
H789
K788
K427
S508
N557
K714
S753
S783
K1324
K1325
C
Smc1
Scc2
Smc3
ii
E823
E822
K113
E821
D782
K112
Smc3
iii
E17
T18
Smc3
I19
T1180
L1181
S1182
Scc2
i
E1168
Scc2
R1122
R1211
R1336
D1103
E1091
Smc1
iv
helix
357-373
Smc3
CC
Scc1-N
Scc2-N
D
K112/K113
K57
K143
K125
K63
R113
Smc3
Smc1
K113
K112
K57
K63
K124
K125
R113
S112
E
Scc2
819-EDEED-823
781-DD-782
K113
K112
dsDNA
Smc3
F
Scc2
Scc1 Kleisin above
Scc1:Smc3
(1-102)
Smc3-CC
dsDNA
Scc1:Scc2
(~190-290)
Smc1
Scc1:Smc1
(502-end)
Scc1:Scc3
(~320-390)
70˚
Scc2
Scc1:Smc3
(1-102)
Scc1:Scc2
(~190-290)
Smc1
Scc1:Smc1
(502-end)
Scc1:Scc3
(~320-390)

Fig 8

Figure 8. Molecular interactions in the E-S/E-K state

(**A**) Cartoon representation of the refined atomic model of cohesin's clamped (E-S/E-K) state based on the 3.4 Å resolution cryo-EM map (Fig. 7A, same orientation and colours, Table 1). (**B**) Basic and polar residues of Scc2 involved in the interaction with DNA. Scc2 interacts only with the backbone. Residues in its vicinity are labelled in black while those mutated in Scc2-4E (Fig. 4) in purple. (**C**) Scc2 makes extensive contacts with both Smc1 and Smc3 heads: i) Scc2 binds Smc1 through its HEAT repeats 18-24 (residues 1127-1493) that dock onto the F-loop on Smc1 (residues 1095-1118) and the emerging coiled coils above it. ii) Smc3's K112 K113, whose acetylation reduces loading efficiency, are in the vicinity of a negatively charged patch on Scc2 (819-EDEED-823 and 781-DD-782). iii) Scc2 binds to Smc3 through a β-strand (part of the otherwise disordered loop 1178-1203) that complements the central β-sheet of Smc3. iv) The N-terminal section of Scc2 contacts parts of Smc3's coiled coil arm/neck, close to where the last ordered region of Scc1's N-terminal domain is bound to the Smc3 coiled coil. (**D**) DNA binding to the SMC head domains is pseudo-symmetrical. Top: the pseudo two-fold axis of the DNA neatly aligns with that of the head domains underneath. Bottom: The head domains interact with the DNA almost exactly two full DNA turns apart, utilising pseudo symmetry-related surfaces (Smc1: K63, S112, R113, & K124; Smc3: K57, K112, K113, & K125). (**E**) The two lysines K112 K113 are in contact with a negatively charged patch on Scc2 (see panel C iii), but are also in the vicinity of the DNA backbone. (**F**) The N-and C-terminal domains of the kleisin Scc1 bind canonically to Smc3 and Smc1, linking the heads and topologically closing the tripartite Smc1/Smc3/Scc1 (S-K) cohesin ring. A tentative path of the disordered regions of Scc1, not visible in our cryo-EM map is shown to demonstrate the topology as deduced from the loading reactions and subsequent crosslinking that show that the DNA must be outside the tripartite S-K ring.

R225 K228 and Scc2 E304) are highly conserved, suggesting that it has an important function.

Pseudo-symmetric binding of DNA to the engaged Smc1/3 ATPase heads

While Scc2 holds the upper half of the DNA's backbone through a spiral of basic and polar residues (Fig. 8B), the engaged heads of Smc1 and Smc3 produce a 2-fold pseudo-symmetrical ABC ATPase heterodimer that binds DNA through two sites that are exactly two turns of the DNA apart and coincide with the major groove in the DNA (Fig. 8D). As expected for DNA-binding proteins that are not sequence specific, neither protein inserts residues into the major or minor grooves and both rely solely on interactions with the DNA backbone. A consequence of this binding mode is that the two-fold symmetry of DNA is matched almost perfectly by the Smc1/3 heterodimer (Fig. 8D top). The two pseudo-symmetrical DNA binding sites close to the major groove are formed through basic and polar amino acids in Smc1 (e.g. K63, S112, R113, & K124) and Smc3 (e.g. K57, K112, K113, & K125) (Fig. 8D bottom). The DNA is bent slightly (~9°) and it seems likely that without bending the DNA binding sites on Smc1 and Smc3 would be too close together. Overall, DNA binding is linked to head engagement and the ATPase cycle, as the complete binding path for DNA along the heads only arises when both heads come together in the E-state.

How do Smc3 K112 and K113 affect loading?

In yeast, Smc3 K112 and K113 have important roles in loading of cohesin onto chromosomes. We show here that changing KK to QQ reduces S-K entrapment in the presence of Scc2 and Scc3 in vitro (Fig. S1C), recapitulating the adverse effect on genome wide association in vivo (Hu et al., 2015). The QQ double mutation is thought to mimic acetylation of K112 K113, which takes place as cells undergo S phase and may have a role in altering how cohesin interacts with DNA, principally whether it can associate (de novo) with and translocate along chromosomes. Our high resolution cryo-EM structure (Figs. 8A and C) reveals that K112 K113 belong to the array of residues that create a basic environment for charge-mediated binding to the DNA backbone (Fig. 8E). This raises the possibility that K112 K113 participate directly in the binding of DNA to engaged heads. However, close inspection of their side chains shows that they in fact face towards the two aspartate and glutamate rich loops in Scc2 (819-EDEED-823 and 781-DD-782) (Fig. 8C ii), implying that they engage in ionic interactions between Smc3 and Scc2 as well as or instead of DNA. If so, one consequence of acetylation or replacement by QQ may be disruption of this mode

of Scc2-Smc3 binding, a notion consistent with our previous finding that QQ greatly reduces stimulation of cohesin's ATPase activity by Scc2 in the absence of DNA, at least when Scc3 is present (Petela et al., 2018).

Remarkably, the charge reversal substitution Scc2E822K was isolated as a spontaneous mutation that suppresses the lethality of *scc4* mutants whose Scc2's activity is greatly compromised (Petela et al., 2018). Because Scc2E822K would be predicted to reduce binding to Smc3 K112 K113, which might have been expected to further reduce not improve the compromised Scc2 activity in of *scc4* mutants, we suggest that E822K might loosen but not eliminate the ionic interactions between Scc2 and K112 K113, permitting the latter to make a greater contribution to DNA binding and thereby increase the affinity between engaged heads and DNA. Given the extreme conservation of residues equivalent to Smc3 K112 K113 and Scc2 E822 D823, it seems likely that the interface has a similar function in most eukaryotes and yet mutations equivalent to *smc3 K112Q K113Q* in *S. pombe* and in human tissue culture cells (where they are not lethal) (Feytout et al., 2011; Ladurner et al., 2016) do not eliminate cohesin loading in the manner observed in yeast (Davidson et al., 2016). Cohesin's ATPase is necessary for LE as well as for loading and we therefore suggest that QQ mutations may turn out to compromise LE.

Smc3 K112 K113 are required for Wapl-dependent release of cohesin from chromosomes as well as for optimal ATPase activity (Ladurner et al., 2014; Petela et al., 2018). Because, release only occurs when Scc2 is replaced by Pds5, our cryo-EM structures provide little direct insight as to their role during release. A key question is whether K112 K113 interact with Pds5 in a similar manner to Scc2 or whether their primary role during release is to bind DNA. Acetylation during S phase blocks release and helps to stabilize Pds5's association with chromosomal cohesin.

Scc1 is bound to both heads and does not engage in DNA binding

Though our map (Fig. 7A) shows little to no density for residues of Scc1 known to bind the central cleft of Scc2 (Scc1 residues ~190-290), it shows very clearly that Scc1's two structured domains,Scc1-N (residues 67-103) and Scc1-C (residues 502-555), are bound in a canonical manner to Smc3's neck and the base of Smc1's ATPase respectively (Gligoris et al., 2014; Haering et al., 2004) thereby bridging the two heads to form the S-K ring (Fig. 8F). We can therefore exclude the possibility that head engagement, at least in the presence of

Scc2 and DNA, causes the sort of rearrangement of Smc3's coiled coil thought to induce Scc1-N's release from Smc3, as suggested by recent structural studies of an ATPγS-bound cohesin trimer (Muir et al., 2020). Similarly, even though it is evident that there is a high level of conservation between the structures of human (Shi et al., 2020), *S. pombe* (Higashi et al., 2020) and *S. cerevisiae* cohesin reported here, the latter shows no sign of any structured part of Scc1 participating in the binding of DNA. Indeed, despite its conservation among most eukaryotes, the positively charged loop within Scc1-N that binds DNA in the human and *S. pombe* complexes (residues 23 - 28 in both) is not present in yeast.

Scc2 and DNA disrupt the J-state in the absence of ATP

Passage of DNA between disengaged heads prior to ATP-driven head engagement would be essential for entrapment simultaneously in E-S and E-K compartments (see Discussion). There must therefore exist a mechanism by which the heads are moved sufficiently far apart to permit DNA passage. To investigate this, we tested the effect of ATP, Scc2, and DNA on crosslinking between the J-state cysteine pair. Conditions that promote efficient E-state crosslinking, namely addition of ATP, Scc2, and DNA, caused a modest ~20% reduction in J-state crosslinking (Fig. S5F), confirming that the E-state is formed at the expense of J. As expected, other combinations of these three factors had less effect. Surprisingly, addition of Scc2 and DNA in the absence of ATP had the greatest effect, causing a ~50% reduction in J crosslinking, an effect that was highly reproducible. Such a marked reduction is presumably caused by the heads adopting a different conformation. Importantly, this does not correspond to the E-state as very little crosslinking takes place between Smc1N1192C and Smc3R1222C under these conditions (Fig. S5A). We therefore suggest that in the absence of ATP, both Scc2 and DNA reduce J-specific crosslinking by driving or indeed holding the ATPase heads apart, a process that could facilitate passage of DNA between them and thereby facilitate its entrapment in E-S compartments when heads engage in the presence of ATP.

DISCUSSION

In vitro reproduction of DNA entrapment within cohesin SMC-kleisin rings

In vivo studies have shown that cohesin entraps circular minichromosomes within its S-K ring (Gligoris et al., 2014; Srinivasan et al., 2018). We demonstrate here that purified cohesin possesses such an activity also in vitro. Unlike previous assays that have merely measured the physical association between cohesin and DNA and investigated its resistance to salt or sensitivity to kleisin cleavage (Murayama and Uhlmann, 2015, 2014), our method measures topological association directly. By covalently circularising the cohesin ring and its component compartments we can make unambiguous deductions about the topology between DNA and cohesin. The entrapment of DNAs within S-K rings measured by this method depends on Scc2, Scc3, and ATP. Importantly, it is also stimulated by ATP hydrolysis, a feature that has been lacking in previous assays but is of paramount importance for entrapment in vivo (Srinivasan et al., 2018). Four other key properties of the in vitro S-K entrapment activity reflect cohesin's behaviour in vivo, namely it depends on the ability of Scc2 and Scc3 to bind DNA, on the ability of ATP to bind Smc3 heads, and on Smc3's K112 K113 residues, whose lack of acetylation is necessary for loading in yeast (Hu et al., 2015). We therefore suggest that the in vitro DNA S-K entrapment described here involves mechanisms similar or identical to those of cohesin operating within cells. If we assume that the efficiency of BMOE induced S-K circulation is around 20%, we estimate that many DNAs are entrapped by 15 or more cohesin rings in our assay after a 40 min incubation.

Potential mechanisms for Scc2-driven E-S/E-K entrapment

An obvious question concerns the state of the Smc1 and Smc3 ATPase heads when DNA is entrapped. They could either be engaged in the presence of ATP, juxtaposed together in the absence of ATP (a state facilitated by extensive association of the Smc1 and Smc3 coiled coils), or fully disengaged. To address this, we used a Smc1-Smc3 cysteine pair specific for engaged heads (Smc1N1192C Smc3 R1222C), which revealed that DNAs are also entrapped efficiently between the hinge and engaged heads (the E-S compartment), between engaged heads and the kleisin subunit associated with them (the E-K compartment), but only rarely between juxtaposed heads and their associated kleisin (the J-K compartment). For obvious reasons, we were not able to address using cysteine-specific crosslinking whether DNAs are also entrapped within S-K rings with fully disengaged heads. However, given that cohesin's ability to hydrolyse ATP is important for S-K entrapment, it is

likely that at least some S-K rings that have entrapped DNA in vitro are in this state. Our failure to observe efficient entrapment within J-K compartments was unexpected given that this state has been documented in vivo (Chapard et al., 2019).

We were surprised to find that unlike entrapment within S-K rings, which requires Scc3, entrapment within E-S/E-K compartments was entirely Scc3 independent. In other words, entrapment within E-S/E-K compartments in the presence of Scc2 alone is not accompanied by entrapment within S-K rings. The similarity in kinetics suggests that entrapment within E-S and E-K compartments driven solely by Scc2 occurs simultaneously as part of the same reaction. The simplest explanation for this is that DNA is transported (upwards) between disengaged ATPase heads and then subsequently trapped in the E-S compartment due to ATP-driven head engagement (Fig. 9A). The simultaneous entrapment within E-K compartments arises naturally from this, as the kleisin polypeptide must be looped "upwards" to accommodate DNA entry in this manner. Crucially, this process would not require opening of the hinge or either SMC-kleisin interface, processes that would be necessary for entrapment within S-K rings. Our cryo-EM structures reveal that entrapment within E-S/E-K compartments is accompanied and probably driven by the binding of DNA to Scc2 and DNA binding sites on the upper surface of Smc1 and Smc3 ATPase heads created upon head engagement. In other words, entrapment within E-S/E-K compartments (in the absence of S-K entrapment) arises from the clamping of DNA between Scc2 and engaged heads. The remarkable similarity between this structure and one formed between DNA, $Scc2^{NIPBL}$, and tetrameric human cohesin (PDB 6WG3) (Shi et al., 2020) shows that the clamping of DNA between $Scc2^{NIPBL}$ and engaged heads not only does not involve $Scc3^{SA2}$ (whose ortholog SA2 was present in the human structure) but more importantly the same clamping happens even when Scc3 is absent (Fig. S6).

An alternative is that simultaneous E-S/E-K entrapment arises from insertion of a loop of DNA into an open S-K ring. If one segment of this loop were located above the heads while the other located below them, head engagement would also lead to simultaneous entrapment in E-S and E-K compartments (Fig. 9B). This scenario, which could also involve the clamping of DNA between Scc2 and engaged heads, is not only more complex, but it must somehow explain how DNA is bent prior to insertion, a process which would carry a clear entropic penalty and more importantly is not apparent from cryo-EM images of EQEQ cohesin associated with circular DNAs (Fig. 7B right). Importantly, the states created by the

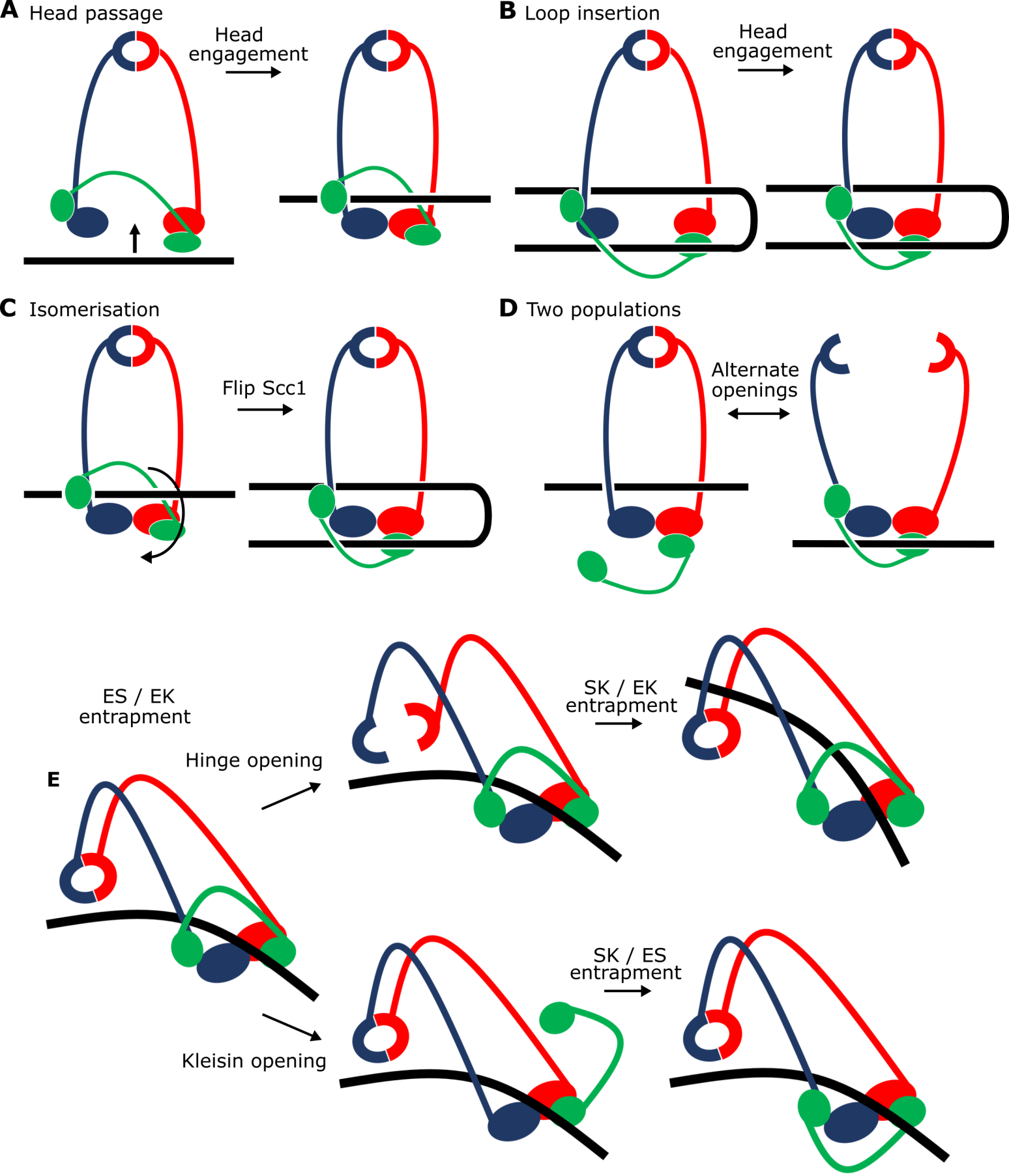

A Head passage
Head engagement
B Loop insertion
Head engagement
C Isomerisation
Flip Scc1
D Two populations
Alternate openings
E
ES / EK entrapment
Hinge opening
SK / EK entrapment
Kleisin opening
SK / ES entrapment

Fig 9

Figure 9. Potential mechanisms for Scc2-driven E-S and E-K entrapment and subsequent S-K ring entrapment

(**A**) ES/EK entrapment by DNA passing through open heads, or (**B**) through a DNA loop being inserted. (**C**) Topological isomerism between A and B. (**D**) ES/EK entrapment due to two distinct populations. (**E**) Models for converting E-S/E-K entrapment to S-K entrapment.

two mechanisms are topologically isomeric. In other words, it is possible to transform the state described in Fig. 9A to that of Fig. 9B merely by moving Scc1's central domain "downwards" to below the heads and simultaneously bending the DNA through 180°.

It is nevertheless important to point out that because our assays measuring E-S/E-K entrapment driven by Scc2 alone use complexes with different sets of cysteine pairs, they do not per se prove simultaneous entrapment of DNAs within both compartments. Thus, DNAs trapped in E-S/E-K compartments could in principle belong to separate populations (Fig. 9D). According to this scenario, and because E-S/E-K entrapment is not accompanied by S-K entrapment, DNAs entrapped solely within E-S compartments would have to be held by complexes whose kleisin subunit had dissociated from one or both ATPase heads while DNAs entrapped solely within E-K compartments would have to be trapped by complexes whose hinge had opened. There are two arguments against this interpretation. First, it is very unclear why head engagement in the presence of ATP, Scc2, and DNA should be associated with two such different events. An even more compelling argument stems from our cryo-EM structures of DNA clamped by Scc2 and engaged heads. A low resolution structure reveals coiled coils folded around their elbow and a dimerised hinge associated with Smc3's coiled coil (Fig. 7E), while a high resolution structure shows that both N- and C-terminal kleisin domains (Scc1-N and -C) are bound to Smc3's neck and the base of Smc1's ATPase respectively (Fig. 8A). In other words, association of DNA with engaged heads in the presence of Scc2 does not appear to be accompanied by opening of any of the S-K ring's three interfaces, ruling out the possibility that E-S and E-K entrapment are independent processes.

The notion that DNA can be engaged by cohesin rings in a manner that does not require opening of the hinge or either SMC-kleisin interface (Fig. 9A & B) is consistent with the recent observation that sealing all three interfaces does not adversely affect DNA loop extrusion by human cohesin complexes (Davidson et al., 2019) as well as the finding that the association between chromosomes and cohesin complexes with certain hinge mutations is not accompanied by S-K entrapment in vivo (Srinivasan et al., 2018).

Though the path of the kleisin chain connecting the ATPase heads is not discernible in any of the cryo-EM structures (this work) (Higashi et al., 2020; Shi et al., 2020), our knowledge that circular DNAs are entrapped within E-S/E-K but not S-K compartments under identical

conditions makes clear that the kleisin chain, whose Scc1-N domain is bound to Smc3's neck, must pass over the DNA bound to the engaged heads before its Scc1-C domain binds to the base of Smc1's ATPase (Fig. 9A and molecular equivalent Fig. 8F). This topology is not merely of academic interest as it provides crucial insight into the pathway by which DNAs are clamped by Scc2 and engaged ATPase heads. Because EQEQ cohesin's association with Scc2 and circular DNA does not appear to cause much DNA bending (Fig. 7B right), we favour the notion that E-S/E-K entrapment arises when DNAs pass (in an upwards direction) between heads prior to their engagement in the presence of ATP and not by insertion of a loop into the S-K ring.

Given that the ATPase heads are frequently associated either in the E- or J-state, there must exist a mechanism to create an opening between them, if only transiently, in order for DNA to pass through before being clamped by their subsequent engagement. Our observation that Scc2 and DNA disrupts the J-state, albeit only in the absence of ATP, may be relevant in this regard (Fig. S5F). This J-state disruption was not caused by adoption of the E-state (Fig. S5A), which requires ATP, and it must therefore involve transition to a state in which Smc1 and Smc3 ATPase heads adopt yet another conformation. This could be a state in which Scc2 and DNA together drive apart the ATPase head domains, thereby enabling DNA to pass between them. Subsequent ATP binding would then cause head engagement and E-state formation, trapping DNA inside both the E-S and E-K compartments. Interestingly, a conformation of this nature has recently been observed in condensin bound to Ysc4 (Lee et al., 2020), where the latter bridges the Smc2 and Smc4 heads, holding them apart by some distance. Although DNA was absent from this structure, the separation of the heads would be sufficient for DNA to pass between them and exposes the HAWK's DNA-binding surface without any impediment. If we assume that Scc2 bridges Smc1 and Smc3 heads in a similar fashion prior to their engagement, then one merely has to propose that DNA initially binds to Scc2 while in the bridged state and remains associated as the Smc3 head pivots around and the Smc3 ATPase head engages with that of Smc1 (Movie 1). We envisage that Scc2's association with Smc1 heads (which strongly resembles that between Ycs4 and Smc4, Fig. S6) remains unaltered during this transition, as it did between Smc4 and Ycs4 in condensin (Lee et al., 2020).

Our demonstration that DNAs transported into the sub-compartment created by Scc2's association with engaged ATPase heads results in entrapment in E-S/E-K but not S-K

compartments is difficult to reconcile with the proposal that DNAs must first pass through a transiently opened Scc1-Smc3 interface before they enter the clamped state created by head engagement (Higashi et al., 2020), a process that has been termed DNA “gripping”. Passage through a gate created by opening the Scc1-Smc3 interface before being clamped by engaged heads and Scc2 would be accompanied by E-K and S-K entrapment but not by E-S entrapment, which is contrary to what we observe.

It is also worth pointing out that E-K entrapment would not be possible if Scc1’s NTD were dissociated from Smc3’s neck upon head engagement, as has been suggested by a cryo-EM structure of heads engaged in the absence of both DNA and Scc2 (Muir et al., 2020). The fact that E-K entrapment accompanies E-S entrapment during our Scc2-only reaction implies that Scc1’s NTD does not in fact dissociate from Smc3’s neck upon head engagement when DNA and Scc2 are present, a feature also revealed by cryo-EM of our yeast (Fig. 8A), human (Shi et al., 2020) and *S. pombe* (Higashi et al., 2020) structures of cohesin heads in complex with DNA and $Scc2^{NIPBL/Mis4}$. Because Scc2 is necessary to prevent cohesin’s release from chromosomes during G1, and because release is accompanied by disengagement of Scc1’s NTD from Smc3’s neck (Srinivasan et al., 2019), we suggest that head engagement may indeed promote Scc1’s dissociation from Smc3 but that this process is actively inhibited by Scc2. One of the functions of Pds5 and Wapl in mediating release during G1 when Smc3 is not acetylated may be to replace Scc2 and thereby abrogate this protection mechanism. In this regard, it is interesting that Scc2 contacts the joint region within Smc3’s coiled coil adjacent to where Scc1’s NTD binds to Smc3's neck (Fig. 8C iv), an interaction also observed in the human structure (Shi et al., 2020) and could have a role in hindering Scc1’s dissociation from Smc3 upon head engagement.

The remarkable similarity in the structures by which yeast and human cohesin clamp DNA between $Scc2^{NIPBL}$ and engaged ATPase heads (Fig. S6) suggests that this highly conserved conformation must have crucial physiological functions. We propose two possibilities. The first is as follows. Because Scc2 is required for S-K entrapment in the presence of Scc3, as well as for E-S/E-K entrapment in its absence, we suggest that entrapment driven by Scc2 and the binding of DNA to engaged heads is necessary for subsequent S-K entrapment. However, this does not exclude the possibility that Scc2 has roles in S-K entrapment additional to formation of an E-S/E-K intermediate. In other words,

the clamping of DNAs between Scc2^{NIPBL} and engaged heads may be a key intermediate during the process of S-K entrapment and hence crucial for the establishment of sister chromatid cohesion.

The observation that S-K entrapment is clearly not necessary for DNA translocation or loop extrusion and may in fact be a rare event in the life of chromosomal cohesin suggests another possibility. The clamping of DNA between Scc2^{NIPBL} and engaged heads and subsequent release upon ATP hydrolysis, all in the absence of S-K entrapment, may be the driving force for cohesin's translocation along DNA, a notion fully consistent with Scc2's key role in stimulating DNA-dependent ATP hydrolysis (Petela et al., 2018) and loop extrusion (Davidson et al., 2019). If so, a crucial question for the future is how transport of DNA into the sub-compartment created by Scc2 and engaged heads is harnessed to mediate translocation along DNA. We presume that DNA translocation is accompanied (and indeed driven) by recurrent cycles of DNA uptake into the clamped state, with each cycle involving segments of DNA further along the chromosome fibre. However, functional translocation would not be possible without a second (reciprocal) mechanism by which DNAs are recurrently bound and released. Scc3's ability to bind DNA may be crucial in this regard. Another idea is that cohesin's hinge provides the second site and that the clamp/release transport cycle is accompanied by changes in the folding of Smc1/3 coiled coils around their elbow region, which could be the key to walking along the DNA. However, this notion is difficult to reconcile with the observation that cohesin's coiled coils can be folded whether its ATPase heads are engaged (Fig. 7E) or disengaged (Bürmann et al., 2019).

Scc3 catalyses entry of DNA inside the SMC-kleisin ring

Our finding that Scc3 is essential for S-K but not E-S/E-K entrapment reveals that Scc3 has a unique role in promoting entry of DNA inside the SMC-kleisin ring as well as being necessary for loop extrusion (Davidson et al., 2019). In principle, Scc3 could catalyse DNA entry either via a gate created by transient hinge opening or through one produced by transient dissociation of one or both SMC-kleisin interfaces (Fig. 9E). Crucially, S-K entrapment in vivo is not abolished by fusing Scc1's NTD to Smc3 or by fusing its CTD to Smc1, implying that DNA must enter either through the hinge or through dissociation of either one of the two SMC-kleisin interfaces (possibly through simultaneous dissociation of both) (Srinivasan et al., 2018). There is little or no direct evidence regarding which mechanism is correct. SMC-kleisin dissociation has been strongly implicated in release and

is therefore also a plausible mechanism for entry (Beckouët et al., 2016). Nevertheless, hinge opening is equally plausible, especially in the light of recent findings that folding of cohesin's coiled coils around an elbow brings its hinge domain into close proximity to DNA bound to the heads, and that Scc3^{SA2} interacts with a half opened hinge when DNA is bound to human cohesin-Scc2^{NIPBL} complexes (Bürmann et al., 2019; Shi et al., 2020). Ascertaining which mechanism is at play will require a method to measure the effect on S-K entrapment of chemically linking interfaces together in a manner that is orthogonal to the BMOE-induced crosslinking. For example, prior crosslinking of both SMC-kleisin interfaces would abolish entrapment via a kleisin gate (Fig. 9E bottom pathway) but not via a hinge gate (Fig. 9E top pathway).

This feature of Scc3's activity depends on its ability to bind DNA in a manner similar to that employed by Scc2 (this work) (Li et al., 2018, Shi et al., 2020) and condensin's Ycg1 HAWK (Kschonsak et al., 2017). Two sets of residues are implicated in DNA binding (K224 K225 R226 and K423 K513 K520). Mutation of one or other set does not abrogate DNA binding or cause lethality but does reduce cohesin's association with chromosomes while mutation of both sets (Scc3-6E) abolishes not only DNA binding and S-K entrapment in vitro but is lethal and abolishes all loading throughout the genome in vivo. Though it abrogates entrapment of DNA within S-K compartments (Fig. 3F), Scc3-6E has no effect on Scc2 driven E-S/E-K entrapment (data not shown). Thus, if S-K entrapment in vivo involved prior formation of an E-S/E-K intermediate, which is consistent with the latter's more rapid kinetics in vitro, then cohesin containing Scc3-6E should form this intermediate and accumulate in this state, possibly at loading sites. Our observation that, despite failing to associate with the vast majority of the genome, Scc3-6E cohesin accumulates at especially high levels at *CEN* sequences, which are highly efficient loading sites, suggests that this may indeed be the case. Unlike Scc3-6E, complete depletion of Scc3 abrogates cohesin's association at *CEN*s as well as along chromosomes arms, which implies that Scc3 has additional functions that do not involve or require its ability to bind DNA.

The notion that entrapment of DNA within S-K rings is preceded by its prior entrapment within E-K/E-S compartments to Scc2 and SMC heads in a manner observed in our cryo-EM structure raises the interesting possibility that DNA is eventually entrapped within the S-K ring, not by passing from outside to inside, but instead by being allowed to exit from either the E-S or the E-K compartment by transiently opening one of the S-K ring's three interfaces.

Transient hinge opening would permit DNA's escape from the E-S compartment (Fig. 9E top pathway) while transient dissociation of one or another, or indeed both, kleisin-head interfaces would permit escape from the E-K compartment (Fig. 9E bottom pathway). In both cases, the subsequent closing of these exit gates would lead to entrapment of DNA within the S-K ring. According to these scenarios, exit via the hinge or via a SMC-kleisin interface without head disengagement would lead, at least initially, to the selective loss of E-S and E-K entrapment respectively. If true, clamping of the DNA would provide the opportunity to open gates without losing grip of the DNA while doing so. It is interesting in this regard that whereas E-K entrapment does not increase between 2 and 40 min when both Scc2 and Scc3 are present, E-S entrapment continues to increase in parallel with the rise in S-K entrapment. Whether this asymmetry is a hint that Scc3 promotes entrapment within S-K rings by opening an SMC-kleisin interface will require far more rigorous types of experiments. Though EQEQ mutants reduce S-K entrapment, they do not eliminate it, suggesting that DNA entry can in principle occur without head disengagement, as depicted in Fig. 9E.

The notion that a key function of Scc3, dependent on its ability to bind DNA, is to facilitate entrapment of DNA within S-K rings has an important corollary. S-K entrapment is thought to be a crucial feature of sister chromatid cohesion. Hitherto, direct evidence for this mechanism has been confined to the observation of small circular minichromosomes entrapped within S-K rings in vivo. We show here that a function of Scc3, not shared by Scc2, is to facilitate entrapment within S-K rings. If this is also an essential function of Scc3 in vivo, it follows that S-K entrapment must also be an essential cohesin function and one that applies to proper chromosomes as well as small circular ones.

We have known for two decades that Scc2 and Scc3 have different roles in promoting cohesin's association with chromosomes (Ciosk et al., 2000; Tóth et al., 1999). The various topological assays described in this paper have finally revealed some of these. Scc2 promotes entrapment of DNA in E-S/E-K compartments by promoting its binding to engaged SMC ATPase heads, while Scc3 promotes entrapment inside S-K rings. These findings are supported by our cryo-EM structure that reveals the molecular identify of the clamped E-S/E-K state and also by recent cryo-EM structures containing cohesin, DNA, and $Scc2^{NIPBL/Mis4}$ as well as $Scc3^{SA2/Psc3}$ (Higashi et al., 2020; Shi et al., 2020). Crucially, our assays reveal the topology of DNA's association with cohesin and the path of the kleisin for

the clamped E-S/E-K state, at least when formed by Scc2 alone and head engagement.

Entrapment of DNAs within E-S compartments has not hitherto been detected in vivo, emphasizing the value of in vitro systems in revealing reactions that are otherwise difficult to detect. Future work will be required to address whether E-S/E-K entrapment also occurs inside cells, to elucidate the mechanism of S-K entrapment, and to reveal conditions that promote J-K entrapment, a form that has been detected in vivo but not yet efficiently in vitro.

ACKNOWLEDGEMENTS

We thank all members of the Nasmyth and Löwe groups for valuable discussions and many unseen contributions over the course of this work. We would like to thank Nicolas Jean (MRC-LMB, Cambridge, UK) for a sample of the relaxed plasmid DNA, Yang Lee (MRC-LMB) for help with tilted data processing, and Dima Chirgadze and Steven W. Hardwick for help with cryo-EM data collection (Biochemistry Department, Cambridge University, UK). This work was partially funded by the Medical Research Council (U105184326 to J.L), Wellcome (202754/Z/16/Z to J.L and 107935/Z/15/Z to K.A.N), and Cancer Research UK (26747 to K.A.N).

AUTHOR CONTRIBUTIONS

J.E.C and K.A.N. conceived the entrapment study. J.E.C. conducted most the in vitro experiments and analysed the data, with contributions from M.B.R., S.Y., and M.V. In vivo experiments were carried out by M.B.R, N.J.P, and J.M. Cohesin purification protocol was developed by B.-G.L. and J.L. Cryo-EM, processing and analysis was performed by B.-G.L and to a minor extent by A.G.L. and J.L. The manuscript was written by J.E.C., A.G.L., B.-G.L, J.L. and K.A.N.

DECLARATION OF INTERESTS

The authors declare no competing interests.

MATERIALS AND METHODS

Recombinant yeast cohesin complex cloning

The *S. cerevisiae* genes *SMC1, SMC3, SCC3, SCC2, SCC1, and SCC4* were codon optimised for expression in *Spodoptera frugiperda* cells and synthesised using the Genescript Thermo Fisher service. These were then cloned into MultiBac vectors. Tag introduction and mutagenesis was achieved through Gibson assembly (New England Biolabs) to generate *SMC1*-His, 2xStrepII-*SCC3*, *SCC1*-2xStrepII, *SCC2*-2xStrepII, *SCC2C*^{133}*SCC2*$^{133-1493}$-2xStrepII, and His-*SCC4*. *SMC3 SMC1*-His, 2xStrepII-*SCC3, SCC2*-2xStrepII, and *SCC2*$^{133-1493}$-2xStrepII were cloned into pACEbac1 vectors, and *SCC1*-2xStrepII and His-*SCC4* cloned into pIDC vectors. *SMC1*-His and *SMC3* were then combined into the same vector via cloning to create a pACEbac1 *SMC1*-His *SMC3*. Vectors containing cohesin trimers were generated by combining pACEbac1 *SMC1*-His *SMC3* with pIDC *SCC1*-2xStrepII by a Cre recombinase reaction (New England Biolabs). The vector for the Scc2/4 expression was also created by combining the pACEbac1 *SCC2*-2xStrepII with pIDC His-*SCC4* using Cre recombinase.

Virus generation and protein expression

DNAs were first transformed into DH10Bac (Thermo Fisher) cells and bacmids containing the expression vector screened for by blue-white selection. DNA was then extracted and 2 µg of bacmid DNA was transfected into 2 ml *S. frugiperda* Sf9 cells (Thermo Fisher) at a cell density of 1x10^{6} cells ml^{-1} using FuGENE HD reagent (Promega), grown in Sf900 II SFM media (Thermo Fisher). These were then incubated at 27°C for 5 days to create P1 virus. P2 virus was then amplified by infecting 50 ml Sf9 cells at a density of 2x10^{6} cells ml^{-1} with 500µl P1 virus and incubating in the dark at 27°C for 3 days with shaking at 100 rpm. P2 virus was then harvested by pelleting cells by centrifugation at 4000g and decanting into 5% FBS (Sigma), and then stored in the dark at 4°C. Typically, proteins were then expressed by adding 5 ml P2 virus to 500 ml Sf900 cells at a density of 2x10^{6} cells ml^{-1} and incubating in the dark at 27ºC for 2 days with shaking at 100 rpm. Cells were then harvested by centrifugation at 1000g, washed with PBS, and then frozen in liquid nitrogen and stored at -80ºC.

Protein purification

Cells were thawed in Buffer A (50 mM HEPES pH 7.5, 150 mM NaCl, 1 mM TCEP (Thermo Fisher), 5% glycerol) supplemented with 1 Complete Protease Inhibitor (EDTA-free) tablet

(Roche), 70 µg RNAse A (Roche), and 100 U ml^{-1} Supernuclease (Sino Biological) and then lysed by sonication. Following sonication, cell lysate was supplemented with 1 mM PMSF (Sigma). Proteins were then purified via a three strep purification protocol. First, proteins were purified via affinity pulldown of their StrepII tags using a StrepTrap HP column (Fisher Scientific) and eluted into Buffer A supplemented with 2.5mM desthiobiotin (Fisher Scientific). Scc2 constructs were eluted into 50 mM Tris pH 8.0 rather than 50 mM HEPES pH 7.5. Proteins were then further purified by anion exchange chromatography using a 5 ml HiTrap Q HP column (GE Healthcare) across a gradient of 100 mM to 1M NaCl. Scc2 constructs were eluted across a gradient of 0 mM to 1 M NaCl. Finally, proteins were purified via size exclusion chromatography using a Superose 6 increase 10/300 GL column (VWR) for cohesin trimers and a HiLoad 16/600 Superdex 200 column (GE Healthcare) for Scc3 and the Scc2 constructs.

Protein gel electrophoresis and western blotting

Samples were mixed with 4xLDS sample buffer (Thermo Fisher), loaded onto a 3-8% Tris-acetate gel (Thermo Fisher) and separated at 150 V for 50 min. Gels were then either stained with Quick Coomassie stain (Generon) or transferred onto a 0.2 µm nitrocellulose membrane using a Trans-blot Turbo transfer pack (Bio-Rad). The antibodies used for western blotting were anti-His (Sigma), anti-Strep HRP conjugated (iba) and anti-Smc3 (Bethyl Laboratories). Primary antibodies were probed with anti-mouse HRP conjugated antibodies (Thermo Fisher).

Purification of pUC19 plasmid DNA

pUC19 plasmid was transformed into TOP10 (Thermo Fisher) cells and grown overnight at 37°C. The next day a single colony was inoculated into 250 ml SOB++ media and grown at 37°C overnight for 16 hours. DNA was then purified via MaxiPrep (Qiagen) using precooled reagents and equipment and eluted into 50 mM HEPES pH 7.5. DNA was then further purified by $CsCl_2$ density gradient centrifugation in the presence of EtBr (Thermo Fisher). The DNA was then extracted, and the EtBr removed by washing several times with butanol saturated with 50 mM HEPES pH 7.5 and then the butanol phase discarded. The $CsCl_2$ was then removed by dialysis against 2 L 50 mM HEPES pH 7.5 buffer over 24 hours at 4°C, with two buffer changes. The DNA was then collected and stored at -20°C.

Protein crosslinking assay

1002 For protein crosslinking assays, 10 µl reactions were prepared containing 570 nM protein
1003 (buffered with 50 mM HEPES pH 7.5, 50 mM NaCl, 5 mM $MgCl_2$, 1 mM TCEP, 5% glycerol).
1004 If added, ATP (Sigma) and pUC19 were added to a concentration of 5 mM and 60 nM
1005 respectively. Reactions mixes were first incubated on ice for 5 min before adding 1 µl BMOE
1006 (Thermo Fisher) to a final concentration of 0.64 mM. Reactions were then incubated on ice
1007 for 6 min. Samples were denatured by adding 4xLDS buffer and heating at 70°C for 10 min
1008 before being separated in 3-8% Tris-acetate gels (Thermo Fisher) run at 150 V for 3 hours.
1009 Gels were stained with Quick Coomassie stain (Generon).
1010
1011 **DNA entrapment assay**
1012 For DNA entrapment assays, 13 µl reactions were prepared containing 165 nM protein and
1013 9.3 nM supercoiled pUC19 (buffered with 50 mM HEPES pH 7.5, 20 mM NaCl, 1 mM $MgCl_2$,
1014 5% glycerol). Scc2C was added to a concentration of 40 nM. These were incubated on ice
1015 for 5 min before reactions were initiated by addition ATP (Sigma) to a final concentration of
1016 5 mM (1 µl ATP added to 12 µl protein DNA mix). Typically, reactions were then incubated
1017 at 24°C for either 40 min or 2 min depending on the compartment being assessed. To these,
1018 1.5 µl BMOE (Thermo Fisher) was added to a final concentration of 0.64 mM and samples
1019 were incubated on ice for 6 min. Samples were then denatured by addition of 1.5 µl 10%
1020 SDS and heated at 70°C for 20 min. DNA loading dye was added and mixtures separated
1021 in a 0.8% agarose gel, run at 50 V for 17 hours at 4°C. The gel was stained with EtBr (Thermo
1022 Fisher) and visualised by UV light. Images shown are representative of 2 independent
1023 experiments.
1024
1025 **ATPase assay**
1026 ATPase experiments were carried out as described in (Petela et al., 2018).
1027
1028 **Calibrated ChIP-sequencing**
1029 ChIP sequencing experiments were carried out as described in (Petela et al., 2018).
1030
1031 **Electromobility shift assay (EMSA)**
1032 A FAM-labelled 39 base pair HPLC purified DNA oligo (Invitrogen,
1033 GAATTCGGTGCGCATAATGTATATATTATGTTAAATAAGCTT) was annealed to a
1034 complementary DNA oligo to form dsDNA by heating to 95°C for 5 min and then decreasing
1035 the temperature to 4°C in 0.1°C increments (buffered in 100 mM potassium acetate, 50 mM

HEPES pH 7.5) to a final concentration of 45 μM. The reactions were then prepared by adding 0.3 μM FAM-dsDNA to increasing concentrations of protein (buffered in 50 mM HEPES pH 7.5, 75 mM NaCl, 1 mM TCEP, 10% glycerol) in a final volume of 10 μL. The reaction mix was then incubated on ice for 30 min in the dark. These were then separated in 5% acrylamide gels prepared with 0.5% TAE (40 mM Tris, 20 mM acetic acid, 1 mM EDTA) run at 100 V for 1 hour at 4ºC in the dark. FAM-labelled DNA was visualised directly on a Fujifilm FLA7000 scanner with the LD473/Y[520] filter.

Cell viability of Scc3 mutants

Mutant *scc3* alleles (under their native promoter) were incorporated at the *leu2* locus in heterozygous *SCC3/Δscc3* cells. Diploids were sporulated and tetrads dissected onto YPD plates. The genotype of the resulting haploids was determined by replica plating. All mutations were confirmed by DNA sequencing.

Cryo-EM sample preparation

1 μM purified *S. cerevisiae* cohesin EQ/EQ trimer (Smc1E1158Q, Smc3E1155Q, Scc1-2xStrepII) was incubated for 30 min at 4˚C with 1 μM Scc2C2(151-1493), forming cohesin tetramer, 5 mM ATP, and 1.3 μM of a 40 bp dsDNA (5'-GAATTCGGTGCG-CATAATGTATATTATGTTAAATAAGCTT-3', 5'-AAGCTTATTTAACATAA-TATACATTATGCGCACCGAATTC-3') or relaxed plasmid DNA. The plasmid was 1789 bp, derived from pUC19, containing a single site for Nt.BspQI nicking endonuclease. Nicking was performed according to the manufacturer's instructions for 60 min at 50 °C (NEB) and the product was purified using a PCR purification kit (Qiagen) and eluted in water. For vitrification, 3 μl of sample were applied to glow-discharged Quantifoil Au 2/2 holey carbon 200 mesh grids or Ultrafoil Au 2/2 holey gold 200 mesh grids (Quantifoil), and flash frozen in liquid ethane using an FEI Vitrobot Mark IV (Thermo Fisher Scientific) and a liquid-ethane cryostat set to −180 °C (Russo et al., 2016).

Cryo-EM data collection

All cryo-EM images were collected on a Titan Krios electron microscope operated at 300 kV (Thermo Fisher Scientific). Images for the cohesin tetramer:40 bp dsDNA complex were collected with a Quantum energy filter (GIF) in front of a K3 Summit direct electron camera in super-resolution mode (both Gatan). The nominal defocus range was set to 1.5 ~ 3.3 μm. Each image was dose-fractionated over 55 frames with a dose rate of 1 electrons per Å per

image. A total 11,944 micrographs were collected in three separate sessions using Krios III at LMB (calibrated pixel size: 1.069 Å / pixel at nominal magnification of 81,000) and the Krios microscope at the Biochemistry Department at the University of Cambridge (pixel size: 1.07 Å / pixel at nominal magnification of 81,000). Because the images showed strong orientation bias, 6,580 micrographs of the datasets were collected at tilts of 25° or 30°. For the tetramer:plasmid DNA complex, 535 images were collected using the Volta phase plate (VPP) (Danev and Baumeister, 2016) and a Gatan K2 Summit direct electron camera in counting mode on Krios II at LMB (calibrated pixel size: 1.00 Å / pixel at magnification of 105,000) with total doses of 45 electrons per $Å^2$, dose fractionated into 40 movie frames using a nominal defocus range of 0.6 ~ 1.0 µm.

Cryo-EM data processing and reconstruction

RELION 3.1 was used for all data processing unless otherwise specified (Scheres, 2012). The resolution was determined based on gold standard Fourier shell correlation (FSC) using the 0.143 criterion (Rosenthal and Henderson, 2003). Using RELION's own motion correction implementation, movie frames were aligned and combined with dose weighting using 7 × 5 patches or 5 x 5 patches for K3 and K2 datasets, respectively. CTF parameters were estimated with CtfFind4 (Rohou and Grigorieff, 2015) for un-tilted images. Focus-gradient patch CTF estimation was performed for tilted images using the programme Warp (Tegunov and Cramer, 2019). For particle picking, RELION and crYOLO were used (Wagner et al., 2019).

For the cohesin tetramer:40 bp DNA complex, initially particles were picked from a subset of ~1,000 images with a Laplacian-of-Gaussian blob as a template using RELION, followed by particle extraction and reference-free 2D classification. An initial 3D model was obtained using particles from selected 2D class images showing different orientations. Then, particle coordinates from images that formed good 2D classes were used to train a model in crYOLO, and particle picking was performed with the trained model. Picked particles were extracted using a box size of 320^2 pixels followed by 3D classification. After several rounds of 3D classification, 588,164 particles with clear density for the Smc1/3 heads and Scc2 were selected and 3D refined to 3.8 Å resolution. Further CTF refinement and Bayesian polishing were performed in RELION followed by another round of 3D auto-refinement, which resulted in a final 3.4 Å map of the cohesin Smc1EQ, Smc3EQ, Scc1, Scc2, ATP, 40 bp DNA complex showing the head part only.

For processing of the tetramer:plasmid DNA complex, picked particles from crYOLO were binned and extracted in a box of 160^2 pixels (2 Å per pixel), followed by several rounds of 2D/3D classifications with the map of the tetramer:40 bp DNA complex as initial 3D model. 23,704 particles showing the head part clearly were selected and re-extracted using a box size of 320^2 pixels (1.07 Å/pix) and were 3D refined to 7.3 Å resolution.

In order to obtain a map of the entire complex, particles from the first 3D classification during the processing of the tetramer:40 bp DNA complex (above) containing clear DNA density were re-centred on the joint region of the complex and re-extracted in a box size of 320^2 pixels (2 Å per pixel). After initial 2D classification, an initial model was generated with a few class averages in different orientations, followed by 3D classification. After several rounds of 3D classification, 21,343 particles containing well-ordered coiled coil and hinge density were 3D refined to 10 Å resolution.

Model building

A homology model of yeast Scc2 was obtained from SWISS-MODEL (Waterhouse et al., 2018) using a crystal structure of Scc2 from *E. gossypii* (PDB 5ME3) as the template (Chao et al., 2017). Crystal structures of yeast Smc1 head (PDB 1W1W; (Haering et al., 2004)) and Smc3 head (PDB:4UX3; (Gligoris et al., 2014)) and the Scc2 homology model were docked into the tetramer:40 bp DNA cryo-EM density map using UCSF Chimera X (Goddard et al., 2018). MAIN (Turk, 2013) and COOT (Emsley et al., 2010) were used for manual rebuilding, followed by refinement using Phenix.real_space_refinement (Afonine et al., 2018). Manual-rebuilding and refinement were repeated for several cycles. The cryo-EM map and the atomic model of the cohesin head segment at 3.4 Å resolution (Table 1) were deposited in the EM Data Bank (EMDB) and Protein Data Bank (PDB) with accession numbers EMD-11585 and PDB 6ZZ6.

REFERENCES

Afonine P V., Poon BK, Read RJ, Sobolev O V., Terwilliger TC, Urzhumtsev A, Adams PD. 2018. Real-space refinement in PHENIX for cryo-EM and crystallography. *Acta Crystallogr Sect D Struct Biol* **74**:531–544. doi:10.1107/S2059798318006551

Arumugam P, Gruber S, Tanaka K, Haering CH, Mechtler K, Nasmyth K. 2003. ATP Hydrolysis Is Required for Cohesin's Association with Chromosomes. *Curr Biol* **13**:1941–1953. doi:10.1016/j.cub.2003.10.036

Beckouët F, Srinivasan M, Roig MB, Chan KL, Scheinost JC, Batty P, Hu B, Petela N, Gligoris T, Smith AC, Strmecki L, Rowland BD, Nasmyth K. 2016. Releasing Activity Disengages Cohesin's Smc3/Scc1 Interface in a Process Blocked by Acetylation. *Mol Cell* **61**:563–574. doi:10.1016/j.molcel.2016.01.026

Bürmann F, Lee BG, Than T, Sinn L, O'Reilly FJ, Yatskevich S, Rappsilber J, Hu B, Nasmyth K, Löwe J. 2019. A folded conformation of MukBEF and cohesin. *Nat Struct Mol Biol* **26**:227–236. doi:10.1038/s41594-019-0196-z

Chan KL, Gligoris T, Upcher W, Kato Y, Shirahige K, Nasmyth K, Beckouët F. 2013. Pds5 promotes and protects cohesin acetylation. *Proc Natl Acad Sci U S A* **110**:13020–13025. doi:10.1073/pnas.1306900110

Chan KL, Roig MB, Hu B, Beckouët F, Metson J, Nasmyth K. 2012. Cohesin's DNA exit gate is distinct from its entrance gate and is regulated by acetylation. *Cell* **150**:961–974. doi:10.1016/j.cell.2012.07.028

Chao WCH, Murayama Y, Muñoz S, Jones AW, Wade BO, Purkiss AG, Hu X-W, Borg A, Snijders AP, Uhlmann F, Singleton MR. 2017. Structure of the cohesin loader Scc2. *Nat Commun* **8**:13952. doi:10.1038/ncomms13952

Chapard C, Jones R, van Oepen T, Scheinost JC, Nasmyth K. 2019. Sister DNA Entrapment between Juxtaposed Smc Heads and Kleisin of the Cohesin Complex. *Mol Cell* **75**:224-237.e5. doi:10.1016/j.molcel.2019.05.023

Ciosk R, Shirayama M, Shevchenko Anna, Tanaka T, Toth A, Shevchenko Andrej, Nasmyth K. 2000. Cohesin's Binding to Chromosomes Depends on a Separate Complex Consisting of Scc2 and Scc4 Proteins. *Mol Cell* **5**:243–254. doi:10.1016/S1097-2765(00)80420-7

Danev R, Baumeister W. 2016. Cryo-EM single particle analysis with the volta phase plate. *Elife*. doi:10.7554/eLife.13046

Davidson IF, Bauer B, Goetz D, Tang W, Wutz G, Peters J-M. 2019. DNA loop extrusion by human cohesin. *Science (80-)* **366**:1338–1345. doi:10.1126/science.aaz3418

Davidson IF, Goetz D, Zaczek MP, Molodtsov MI, Huis in ’t Veld PJ, Weissmann F, Litos G, Cisneros DA, Ocampo-Hafalla M, Ladurner R, Uhlmann F, Vaziri A, Peters J. 2016. Rapid movement and transcriptional re-localization of human cohesin on DNA. *EMBO J* **35**:2671–2685. doi:10.15252/embj.201695402

Diebold-Durand ML, Lee H, Ruiz Avila LB, Noh H, Shin HC, Im H, Bock FP, Bürmann F, Durand A, Basfeld A, Ham S, Basquin J, Oh BH, Gruber S. 2017. Structure of Full-Length SMC and Rearrangements Required for Chromosome Organization. *Mol Cell* **67**:334-347.e5. doi:10.1016/j.molcel.2017.06.010

Emsley P, Lohkamp B, Scott WG, Cowtan K. 2010. Features and development of Coot. *Acta Crystallogr Sect D Biol Crystallogr* **66**:486–501. doi:10.1107/S0907444910007493

Feytout A, Vaur S, Genier S, Vazquez S, Javerzat J-P. 2011. Psm3 Acetylation on Conserved Lysine Residues Is Dispensable for Viability in Fission Yeast but Contributes to Eso1-Mediated Sister Chromatid Cohesion by Antagonizing Wpl1. *Mol Cell Biol* **31**:1771–1786. doi:10.1128/mcb.01284-10

Gligoris TG, Scheinost JC, Bürmann F, Petela N, Chan KL, Uluocak P, Beckouët F, Gruber S, Nasmyth K, Löwe J. 2014. Closing the cohesin ring: Structure and function of its Smc3-kleisin interface. *Science (80-)* **346**:963–967. doi:10.1126/science.1256917

Goddard TD, Huang CC, Meng EC, Pettersen EF, Couch GS, Morris JH, Ferrin TE. 2018. UCSF ChimeraX: Meeting modern challenges in visualization and analysis. *Protein Sci* **27**:14–25. doi:10.1002/pro.3235

Golfier S, Quail T, Kimura H, Brugués J. 2020. Cohesin and condensin extrude DNA loops in a cell-cycle dependent manner. *Elife* **9**. doi:10.7554/eLife.53885

Gruber S, Haering CH, Nasmyth K. 2003. Chromosomal cohesin forms a ring. *Cell* **112**:765–777. doi:10.1016/S0092-8674(03)00162-4

Haering CH, Farcas AM, Arumugam P, Metson J, Nasmyth K. 2008. The cohesin ring concatenates sister DNA molecules. *Nature* **454**:297–301. doi:10.1038/nature07098

Haering CH, Löwe J, Hochwagen A, Nasmyth K. 2002. Molecular architecture of SMC proteins and the yeast cohesin complex. *Mol Cell* **9**:773–788. doi:10.1016/S1097-2765(02)00515-4

Haering CH, Schoffnegger D, Nishino T, Helmhart W, Nasmyth K, Löwe J. 2004. Structure and stability of cohesin’s Smc1-kleisin interaction. *Mol Cell* **15**:951–964. doi:10.1016/j.molcel.2004.08.030

Higashi TL, Eickhoff P, Simoes JS, Locke J, Flynn HR, Snijders AP, Papageorgiou G,

Reilly O, Chen ZA, Reilly FJO, Rappsilber J, Uhlmann F. 2020. A Structure-Based Mechanism for DNA Entry into the Cohesin Ring. doi:10.1101/2020.04.21.052944

Hinshaw SM, Makrantoni V, Harrison SC, Marston AL. 2017. The Kinetochore Receptor for the Cohesin Loading Complex. *Cell* **171**:72-84.e13. doi:10.1016/j.cell.2017.08.017

Hu B, Petela N, Kurze A, Chan KL, Chapard C, Nasmyth K. 2015. Biological chromodynamics: A general method for measuring protein occupancy across the genome by calibrating ChIP-seq. *Nucleic Acids Res* **43**. doi:10.1093/nar/gkv670

Kikuchi S, Borek DM, Otwinowski Z, Tomchick DR, Yu H. 2016. Crystal structure of the cohesin loader Scc2 and insight into cohesinopathy. *Proc Natl Acad Sci U S A* **113**:12444–12449. doi:10.1073/pnas.1611333113

Kim Y, Shi Z, Zhang H, Finkelstein IJ, Yu H. 2019. Human cohesin compacts DNA by loop extrusion. *Science (80-)* **366**:1345–1349. doi:10.1126/science.aaz4475

Kschonsak M, Merkel F, Bisht S, Metz J, Rybin V, Hassler M, Haering CH. 2017. Structural Basis for a Safety-Belt Mechanism That Anchors Condensin to Chromosomes. *Cell* **171**:588-600.e24. doi:10.1016/j.cell.2017.09.008

Ladurner R, Bhaskara V, Huis In ’T Veld PJ, Davidson IF, Kreidl E, Petzold G, Peters JM. 2014. Cohesin’s ATPase activity couples cohesin loading onto DNA with Smc3 acetylation. *Curr Biol* **24**:2228–2237. doi:10.1016/j.cub.2014.08.011

Ladurner R, Kreidl E, Ivanov MP, Ekker H, Idarraga-Amado MH, Busslinger GA, Wutz G, Cisneros DA, Peters J. 2016. Sororin actively maintains sister chromatid cohesion. *EMBO J* **35**:635–653. doi:10.15252/embj.201592532

Lammens A, Schele A, Hopfner K-P. 2004. Structural Biochemistry of ATP-Driven Dimerization and DNA-Stimulated Activation of SMC ATPases. *Curr Biol* **14**:1778–1782. doi:10.1016/j.cub.2004.09.044

Lee B-GB, Merkel F, Allegretti M, Hassler M, Cawood C, Lecomte L, O’Reilly FJ, Sinn LRL, Gutierrez-Escribano P, Kschonsak M, Bravo S, Nakane T, Rappsilber J, Aragon L, Beck M, Löwe J, Haering CH, Reilly FJO, Sinn LRL, Gutierrez-Escribano P, Bravo S, Nakane T, Rappsilber J, Aragon L. 2020. Cryo-EM structures of holo condensin reveal a subunit flip-flop mechanism. *Nat Struct Mol Biol*. doi:10.1038/s41594-020-0457-x

Li Y, Muir KW, Bowler MW, Metz J, Haering CH, Panne D. 2018. Structural basis for scc3-dependent cohesin recruitment to chromatin. *Elife* **7**. doi:10.7554/eLife.38356

Liu Y, Sung S, Kim Y, Li F, Gwon G, Jo A, Kim A, Kim T, Song O, Lee SE, Cho Y. 2016. ATP -dependent DNA binding, unwinding, and resection by the Mre11/Rad50 complex . *EMBO J* **35**:743–758. doi:10.15252/embj.201592462

Marcos-Alcalde Í, Mendieta-Moreno JI, Puisac B, Gil-Rodríguez MC, Hernández-Marcos M, Soler-Polo Di, Ramos FJ, Ortega J, Pié J, Mendieta J, Gómez-Puertas P. 2017. Two-step ATP-driven opening of cohesin head. *Sci Rep* **7**:1–14. doi:10.1038/s41598-017-03118-9

Minamino M, Higashi TL, Bouchoux C, Uhlmann F. 2018. Topological in vitro loading of the budding yeast cohesin ring onto DNA. *Life Sci Alliance* **1**. doi:10.26508/lsa.201800143

Muir KW, Li Y, Weis F, Panne D. 2020. The structure of the cohesin ATPase elucidates the mechanism of SMC–kleisin ring opening. *Nat Struct Mol Biol* **27**:233–239. doi:10.1038/s41594-020-0379-7

Murayama Y, Uhlmann F. 2015. DNA Entry into and Exit out of the Cohesin Ring by an Interlocking Gate Mechanism. *Cell* **163**:1628–1640. doi:10.1016/j.cell.2015.11.030

Murayama Y, Uhlmann F. 2014. Biochemical reconstitution of topological DNA binding by the cohesin ring. *Nature* **505**:367–371. doi:10.1038/nature12867

Nasmyth K. 2001. Disseminating the Genome: Joining, Resolving, and Separating Sister Chromatids During Mitosis and Meiosis. *Annu Rev Genet* **35**:673–745. doi:10.1146/annurev.genet.35.102401.091334

Oliveira RA, Hamilton RS, Pauli A, Davis I, Nasmyth K. 2010. Cohesin cleavage and Cdk inhibition trigger formation of daughter nuclei. *Nat Cell Biol* **12**:185–192. doi:10.1038/ncb2018

Petela NJ, Gligoris TG, Metson J, Lee BG, Voulgaris M, Hu B, Kikuchi S, Chapard C, Chen W, Rajendra E, Srinivisan M, Yu H, Löwe J, Nasmyth KA. 2018. Scc2 Is a Potent Activator of Cohesin's ATPase that Promotes Loading by Binding Scc1 without Pds5. *Mol Cell* **70**:1134-1148.e7. doi:10.1016/j.molcel.2018.05.022

Rohou A, Grigorieff N. 2015. CTFFIND4: Fast and accurate defocus estimation from electron micrographs. *J Struct Biol* **192**:216–21. doi:10.1016/j.jsb.2015.08.008

Rosenthal PB, Henderson R. 2003. Optimal determination of particle orientation, absolute hand, and contrast loss in single-particle electron cryomicroscopy. *J Mol Biol* **333**:721–745. doi:10.1016/j.jmb.2003.07.013

Russo CJ, Scotcher S, Kyte M. 2016. A precision cryostat design for manual and semi-automated cryo-plunge instruments. *Rev Sci Instrum*. doi:10.1063/1.4967864

Scheres SHW. 2012. RELION: Implementation of a Bayesian approach to cryo-EM structure determination. *J Struct Biol*. doi:10.1016/j.jsb.2012.09.006

Shi Z, Gao H, Bai X, Yu H. 2020. Cryo-EM structure of the human cohesin-NIPBL-DNA complex. *Science (80-)* eabb0981. doi:10.1126/science.abb0981

Soh YM, Bürmann F, Shin HC, Oda T, Jin KS, Toseland CP, Kim C, Lee H, Kim SJ, Kong MS, Durand-Diebold ML, Kim YG, Kim HM, Lee NK, Sato M, Oh BH, Gruber S. 2015. Molecular basis for SMC rod formation and its dissolution upon DNA binding. *Mol Cell* **57**:290–303. doi:10.1016/j.molcel.2014.11.023

Srinivasan M, Petela NJ, Scheinost JC, Collier J, Voulgaris M, Roig MB, Beckouët F, Hu B, Nasmyth KA. 2019. Scc2 counteracts a wapl-independent mechanism that releases cohesin from chromosomes during G1. *Elife* **8**:1–34. doi:10.7554/eLife.44736.001

Srinivasan M, Scheinost JC, Petela NJ, Gligoris TG, Wissler M, Ogushi S, Collier JE, Voulgaris M, Kurze A, Chan KL, Hu B, Costanzo V, Nasmyth KA. 2018. The Cohesin Ring Uses Its Hinge to Organize DNA Using Non-topological as well as Topological Mechanisms. *Cell* **173**:1508-1519.e18. doi:10.1016/j.cell.2018.04.015

Tegunov D, Cramer P. 2019. Real-time cryo-electron microscopy data preprocessing with Warp. *Nat Methods* **16**:1146–1152. doi:10.1038/s41592-019-0580-y

Tóth A, Ciosk R, Uhlmann F, Galova M, Schleiffer A, Nasmyth K. 1999. Yeast cohesin complex requires a conserved protein, Eco1p(Ctf7), to establish cohesion between sister chromatids during DNA replication. *Genes Dev* **13**:320–333. doi:10.1101/gad.13.3.320

Turk D. 2013. MAIN software for density averaging, model building, structure refinement and validation. *Acta Crystallogr Sect D Biol Crystallogr* **69**:1342–1357. doi:10.1107/S0907444913008408

Uhlmann F, Lottspelch F, Nasmyth K. 1999. Sister-chromatid separation at anaphase onset is promoted by cleavage of the cohesin subunit Scc1. *Nature* **400**:37–42. doi:10.1038/21831

Uhlmann F, Wernic D, Poupart M-A, Koonin E V, Nasmyth K. 2000. Cleavage of Cohesin by the CD Clan Protease Separin Triggers Anaphase in Yeast. *Cell* **103**:375–386. doi:10.1016/S0092-8674(00)00130-6

Vazquez Nunez R, Ruiz Avila LB, Gruber S. 2019. Transient DNA Occupancy of the SMC Interarm Space in Prokaryotic Condensin. *Mol Cell* **75**:209-223.e6. doi:10.1016/j.molcel.2019.05.001

Wagner T, Merino F, Stabrin M, Moriya T, Antoni C, Apelbaum A, Hagel P, Sitsel O, Raisch T, Prumbaum D, Quentin D, Roderer D, Tacke S, Siebolds B, Schubert E, Shaikh TR, Lill P, Gatsogiannis C, Raunser S. 2019. SPHIRE-crYOLO is a fast and accurate fully automated particle picker for cryo-EM. *Commun Biol* **2**:218. doi:10.1038/s42003-019-0437-z

Waterhouse A, Bertoni M, Bienert S, Studer G, Tauriello G, Gumienny R, Heer FT, De Beer TAP, Rempfer C, Bordoli L, Lepore R, Schwede T. 2018. SWISS-MODEL: Homology modelling of protein structures and complexes. *Nucleic Acids Res* **46**:W296–W303. doi:10.1093/nar/gky427

Wells JN, Gligoris TG, Nasmyth KA, Marsh JA. 2017. Evolution of condensin and cohesin complexes driven by replacement of Kite by Hawk proteins. *Curr Biol*. doi:10.1016/j.cub.2016.11.050

Yatskevich S, Rhodes J, Nasmyth K. 2019. Organization of Chromosomal DNA by SMC Complexes. *Annu Rev Genet* **53**:445–482. doi:10.1146/annurev-genet-112618-043633

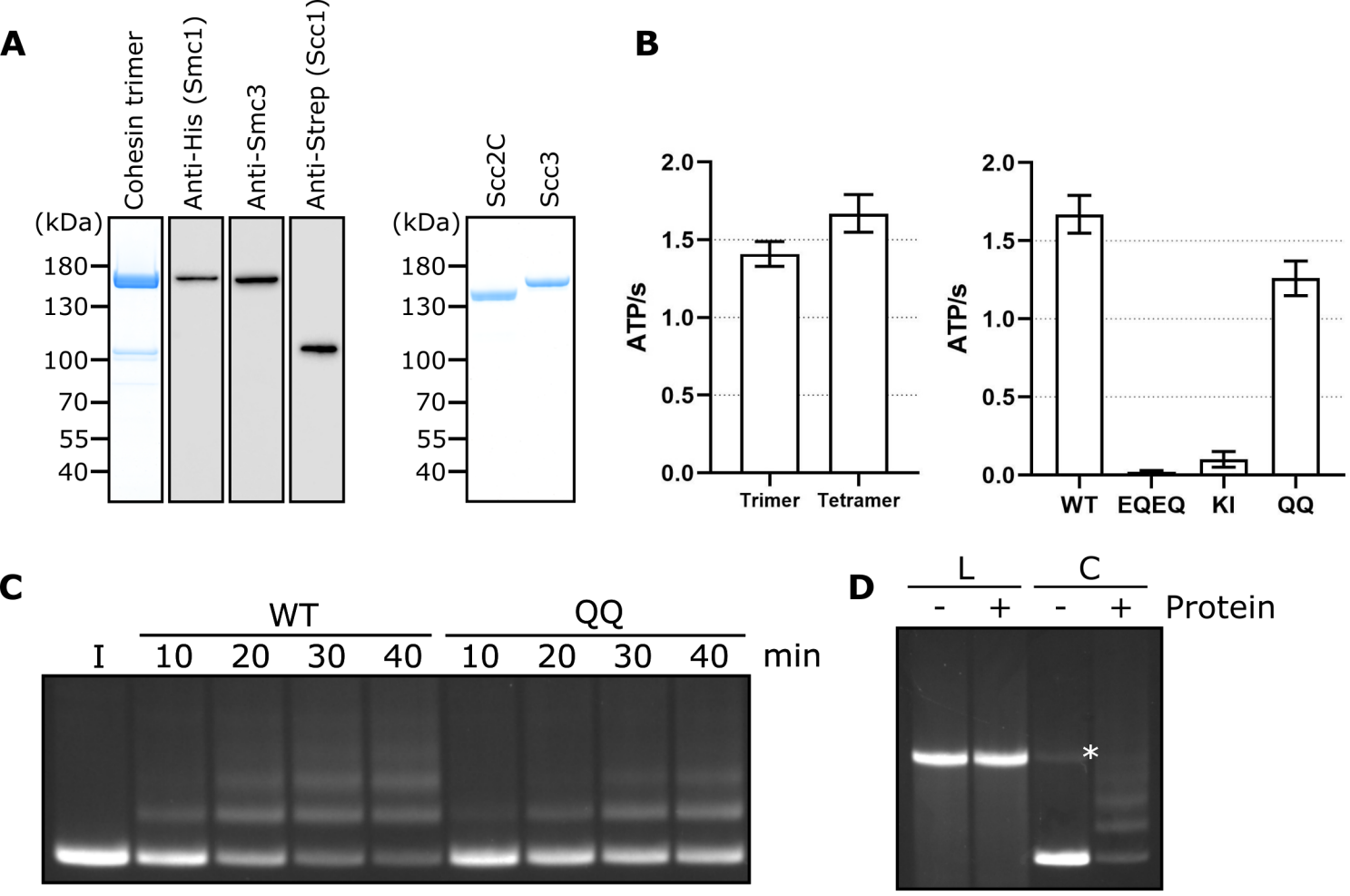

A
Cohesin trimer
Anti-His (Smc1)
Anti-Smc3
Anti-Strep (Scc1)
(kDa)
180
130
100
70
55
40
Scc2C
Scc3
B
ATP/s
2.0
1.5
1.0
0.5
0.0
Trimer
Tetramer
WT
EQEQ
KI
QQ
C
WT
QQ
I
10
20
30
40
min
D
L
C
-
+
Protein
*

Fig S1

Figure S1. Related to Figures 1 and 2

(**A**) Left hand panel - Coomassie strain of cohesin trimers. Individual subunits were then probed for by western blotting. Right hand panel – Coomassie stain of Scc2C and Scc3. (**B**) Left hand panel - ATPase activity for purified cohesin trimers incubated with DNA, Scc2, and ATP in the presence of Scc3 (tetramer) or absence (trimer). Right hand panel – ATPase activity for either WT cohesin tetramer, Smc1E1158Q Smc3E1155Q (EQEQ), Smc3K38I (KI) or Smc3K112Q K113Q (QQ). (**C**) Entrapment of DNA in S-K rings in the presence of Scc2 and Scc3 comparing WT cohesin to Smc3K112Q K113Q (QQ) (I = input DNA), or (**D**) entrapment of linear DNA (L) or circular supercoiled DNA (C) in the presence or absence of protein (Trimer, Scc2 and Scc3) (* = damaged open circular DNA).

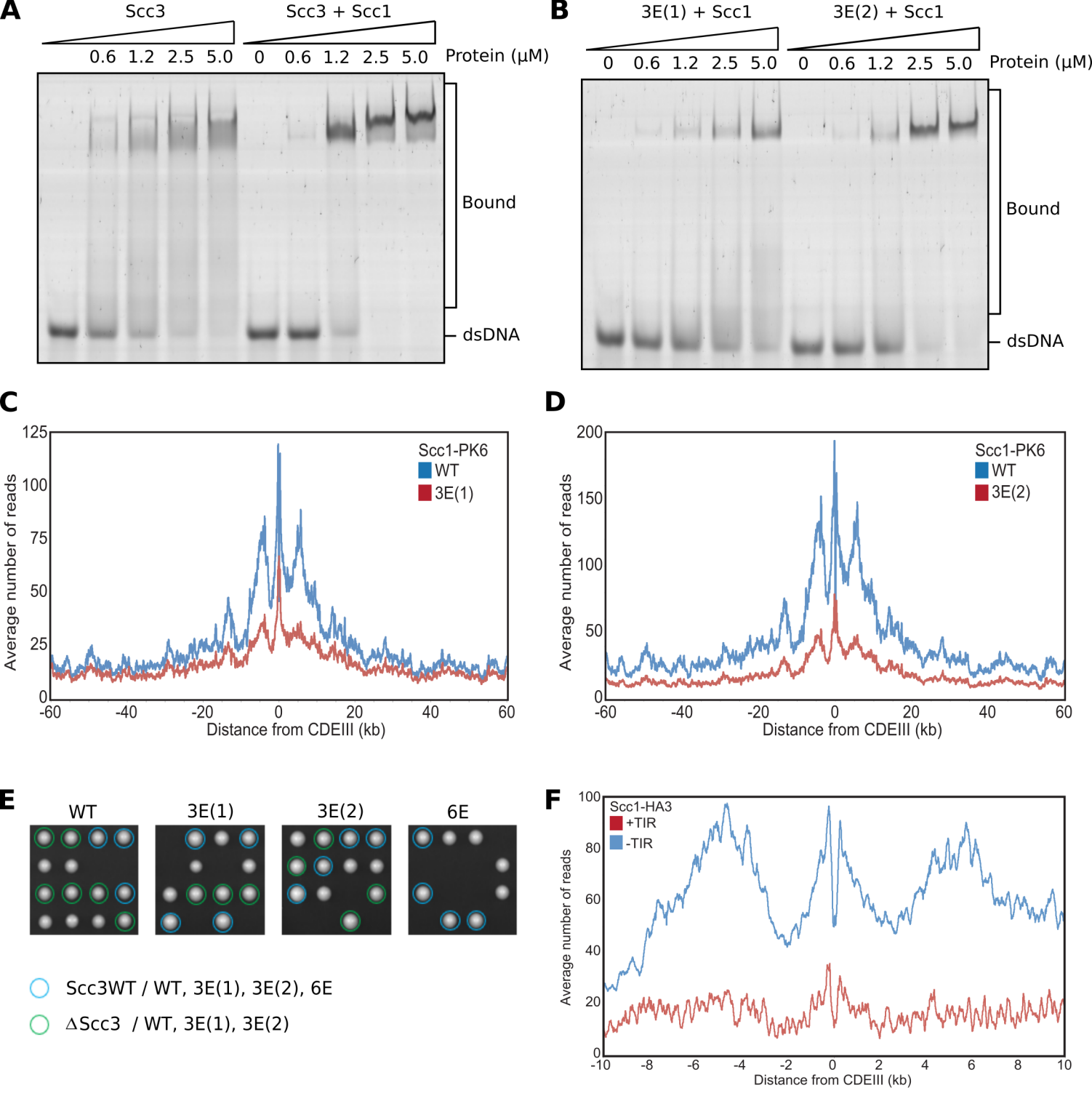
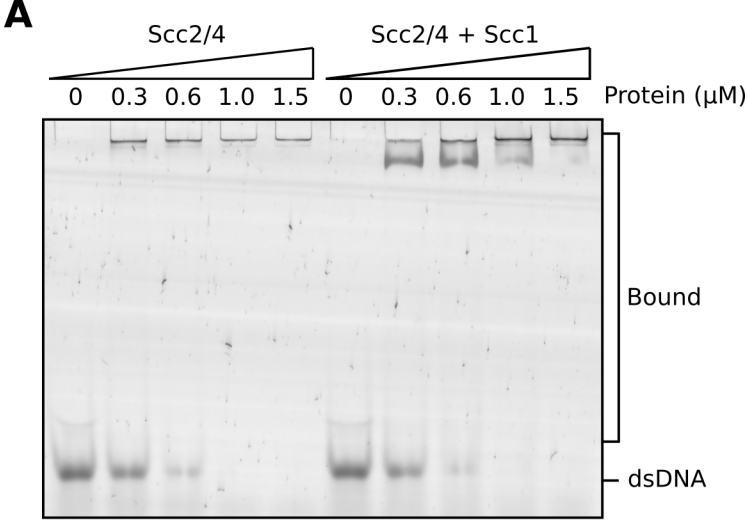
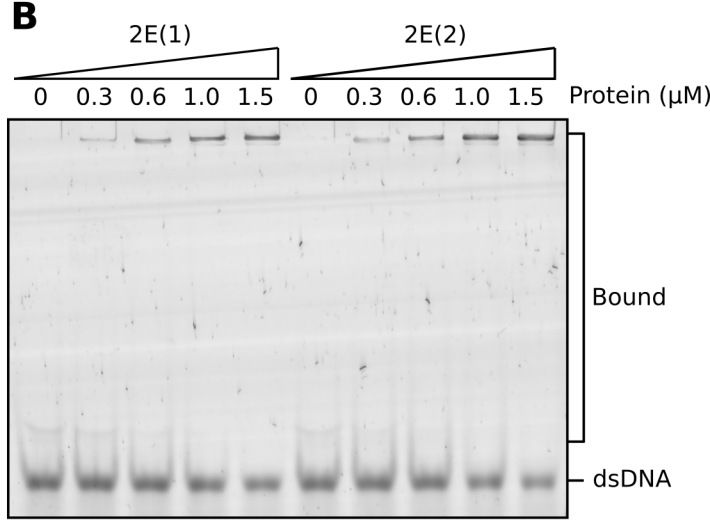

A
Scc3
Scc3 + Scc1
0.6 1.2 2.5 5.0
0 0.6 1.2 2.5 5.0
Protein (μM)
Bound
dsDNA
B
3E(1) + Scc1
3E(2) + Scc1
0 0.6 1.2 2.5 5.0
0 0.6 1.2 2.5 5.0
Protein (μM)
Bound
dsDNA
C
Scc1-PK6
WT
3E(1)
Average number of reads
Distance from CDEIII (kb)
D
Scc1-PK6
WT
3E(2)
Average number of reads
Distance from CDEIII (kb)
E
WT
3E(1)
3E(2)
6E
Scc3WT / WT, 3E(1), 3E(2), 6E
ΔScc3 / WT, 3E(1), 3E(2)
F
Scc1-HA3
+TIR
-TIR
Average number of reads
Distance from CDEIII (kb)

Fig S2

Figure S2. Related to Figure 3

(**A**) EMSA comparing the ability of Scc3 to bind dsDNA against Scc3-Scc1$^{269-451}$ complexes, or (**B**) of Scc3-3E(1)-Scc1$^{269-451}$ (Scc3K224E K225E R226E) against Scc3-3E(2)-Scc1$^{269-451}$ (Scc3K423E K513E K520E) complexes. (**C**) Average calibrated ChIP-seq profiles 60 kb either side of *CEN*s of Scc1-PK6 in the presence of WT Scc3 (KN27542) or Scc3-3E(1) (KN27547), taken in cycling cells at 25°C. (**D**) Average calibrated ChIP-seq profiles of Scc1-PK6 in the presence of WT Scc3 (KN27542) or Scc3-3E(2) (KN27697), taken in cycling cells at 25°C. (**E**) Vertical tetrad dissection of spores from diploid cells heterozygous for deletion of the endogenous *SCC3* gene, expressing a single copy of either WT Scc3 (KN21273), Scc3-3E(1) (KN27539), Scc3-3E(2) (KN27696), or Scc3-6E (KN27763) from an ectopic locus. WT Scc3, Scc3-3E(1), Scc3-3E(2) are able to support viability but Scc3-6E is not. Further marker selection, protein expression and genotype sequencing confirmed that Scc3-6E causes lethality. (**F**) Average calibrated ChIP-seq profiles 10 kb either side of *CEN*s of Scc1-HA3 either expressing TIR necessary for auxin inducible degradation (KN20783) or not (KN20785). Samples were arrested in G1 with α-factor at 25°C prior to release into auxin containing media to deplete endogenous Scc3. ChIP-seq samples were taken 60 min after release.


A
Scc2/4
Scc2/4 + Scc1
0 0.3 0.6 1.0 1.5 0 0.3 0.6 1.0 1.5
Protein (μM)
Bound
dsDNA


B
2E(1)
2E(2)
0 0.3 0.6 1.0 1.5 0 0.3 0.6 1.0 1.5
Protein (μM)
Bound
dsDNA

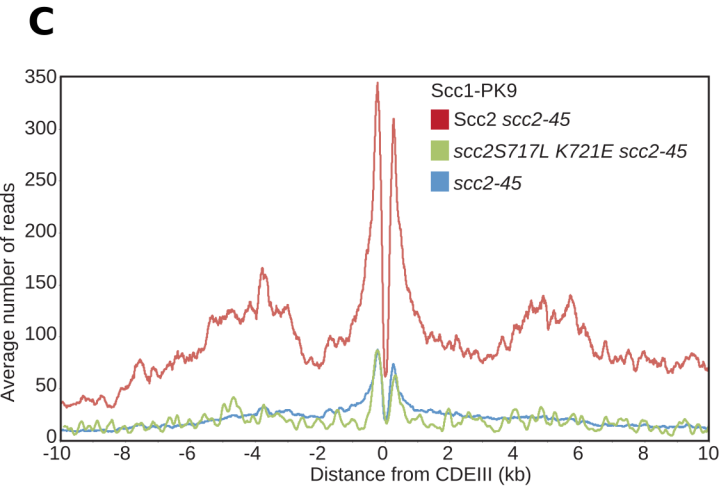

C
Scc1-PK9
Scc2 scc2-45
scc2S717L K721E scc2-45
scc2-45
Average number of reads
350
300
250
200
150
100
50
0
-10 -8 -6 -4 -2 0 2 4 6 8 10
Distance from CDEIII (kb)

Fig S3

Figure S3. Related to Figure 4

(**A**) EMSA comparing the ability of full length Scc2/4 and Scc2/4-Scc1$^{150-298}$ complexes to bind dsDNA or (**B**) Comparing Scc2-2E(1) (Scc2S717E K721E) and Scc2-2E(2) (Scc2K788E H789E). (**C**) Average calibrated ChIP-seq profiles 10 kb either side of *CEN*s of cells expressing WT (KN24185), *S717L K721E* double mutant (KN27010) or no ectopic copy of *SCC2* (KN22390), over endogenous *scc2-45*. Cells were arrested in G1 with α-factor at 25°C before release into medium containing nocodazole at 37°C. Samples were taken 75 min after release. *scc2-45* is a temperature sensitive allele.

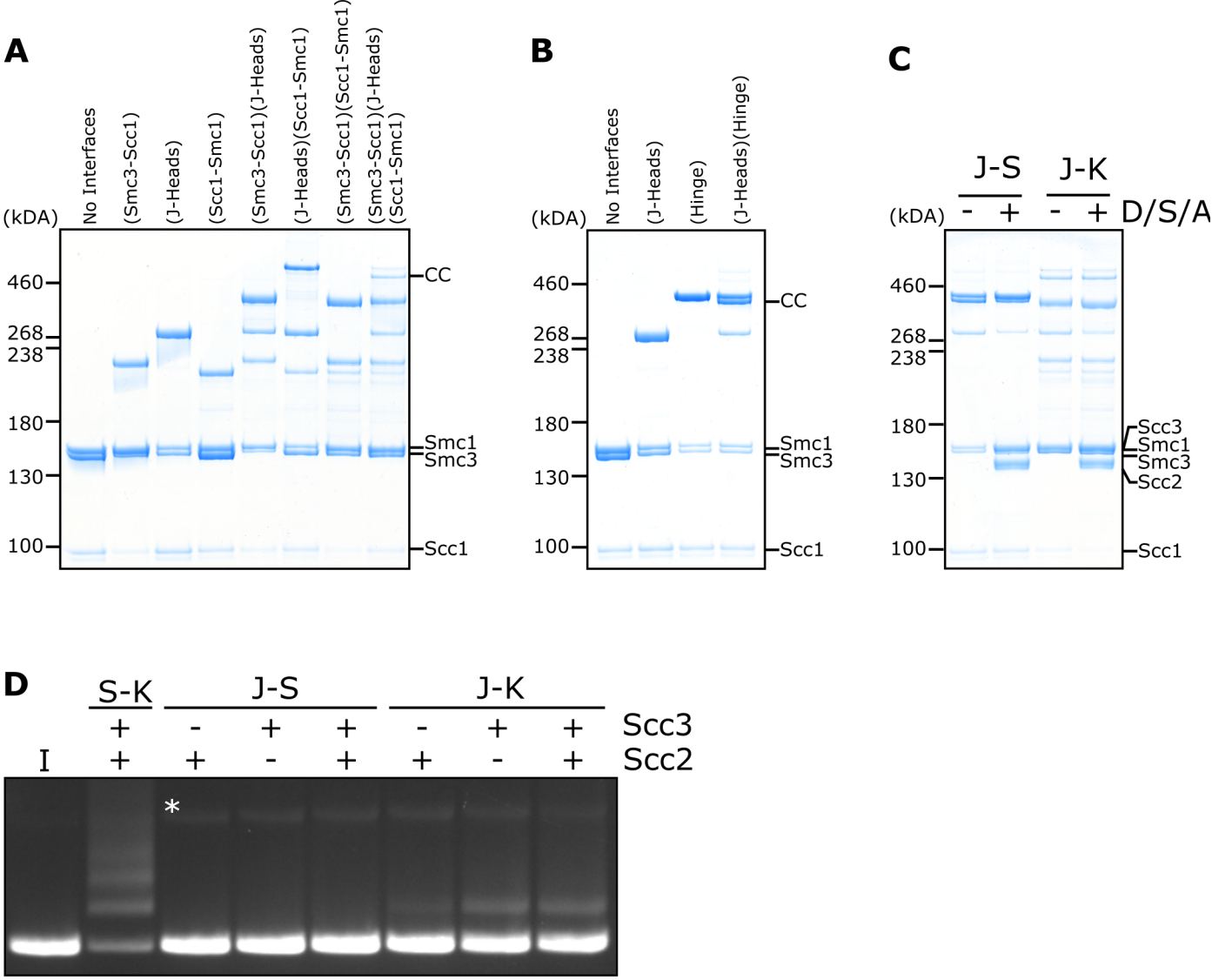

A
No Interfaces
(Smc3-Scc1)
(J-Heads)
(Scc1-Smc1)
(Smc3-Scc1)(J-Heads)
(J-Heads)(Scc1-Smc1)
(Smc3-Scc1)(Scc1-Smc1)
(Smc3-Scc1)(J-Heads)
(Scc1-Smc1)
(kDA)
460
268
238
180
130
100
CC
Smc1
Smc3
Scc1
B
No Interfaces
(J-Heads)
(Hinge)
(J-Heads)(Hinge)
(kDA)
460
268
238
180
130
100
CC
Smc1
Smc3
Scc1
C
J-S
J-K
- + - + D/S/A
(kDA)
460
268
238
180
130
100
Scc3
Smc1
Smc3
Scc2
Scc1
D
S-K
J-S
J-K
Scc3
Scc2
I
*

Fig S4

Figure S4. DNA is never entrapped in J-S and only rarely in J-K compartments

(**A**) BMOE crosslinking of J-K compartments, or (**B**) of J-S compartments, with cysteine pairs in the designated interfaces. CC = circular cohesin. (**C**) Crosslinking of J-S or J-K compartments in the presence of DNA, Scc2, Scc3 and ATP (D/S/A) or absence of all four. (**D**) Entrapment of DNA in either S-K rings, or J-S/J-K compartments in the presence of Scc2, Scc3, or both Scc2 and Scc3. Incubated for 40 min (* = damaged open circular DNA; I = input DNA).

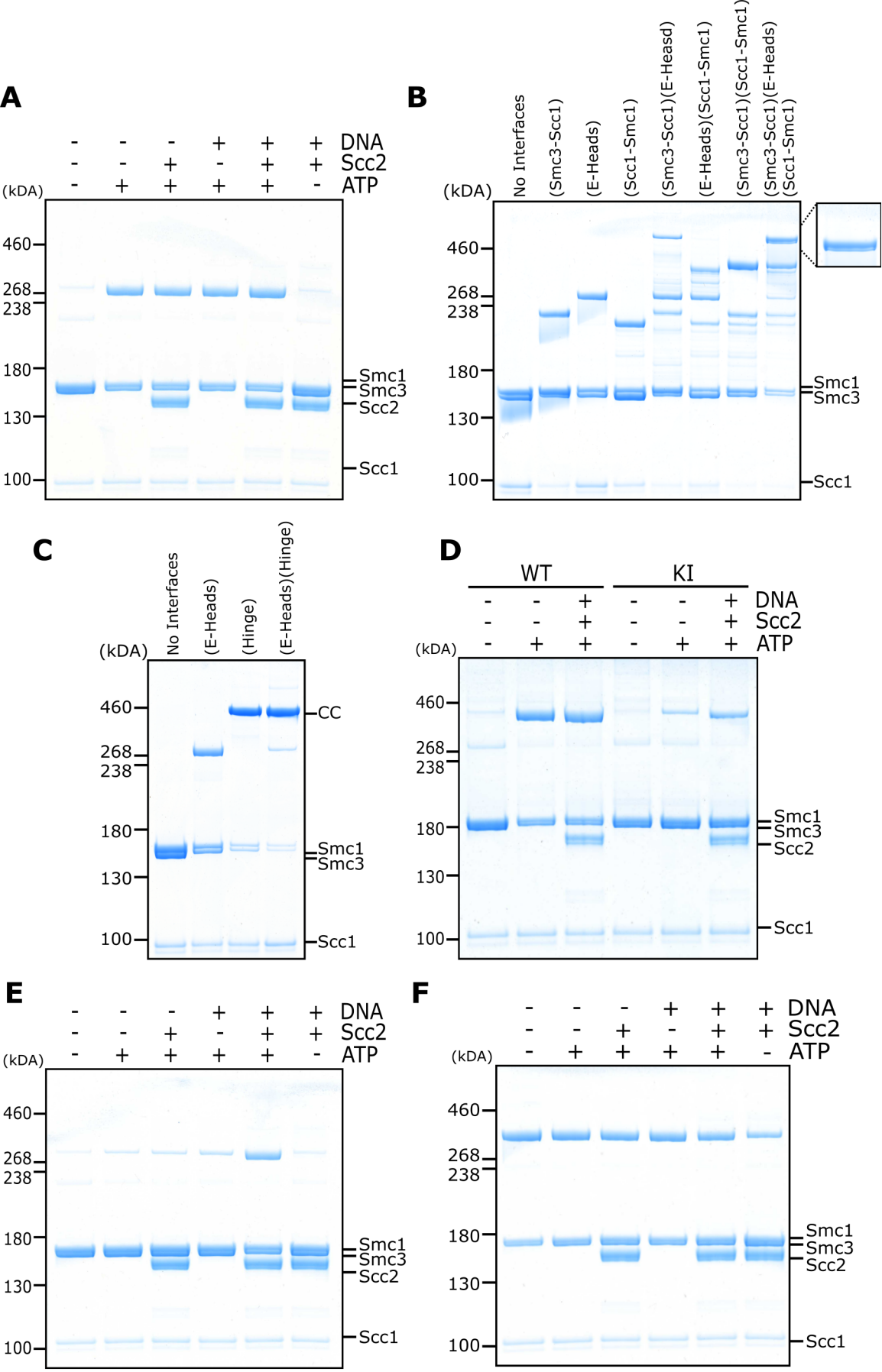

A
DNA
Scc2
ATP
(kDA)
460
268
238
180
130
100
Smc1
Smc3
Scc2
Scc1
B
No Interfaces
(Smc3-Scc1)
(E-Heads)
(Scc1-Smc1)
(Smc3-Scc1)(E-Heasd)
(E-Heads)(Scc1-Smc1)
(Smc3-Scc1)(Scc1-Smc1)
(Smc3-Scc1)(E-Heads)
(Scc1-Smc1)
C
No Interfaces
(E-Heads)
(Hinge)
(E-Heads)(Hinge)
CC
D
WT
KI
E
F

Fig S5

Figure S5. Related to Figure 5

(**A**) BMOE crosslinking of cohesin containing a cysteine pair for E head association, in the presence of ATP, DNA, or Scc2 in various combinations and in the presence of 1mM $MgCl_2$. (**B**) Crosslinking of E-K compartments, or (**C**) of E-S compartments, with cysteine pairs in the designated interfaces. CC = circular cohesin. (**D**) Crosslinking of cohesin containing a cysteine pair for E head association, comparing WT cohesin against Smc3K38I (KI) in the presence of ATP, DNA, or Scc2 in various combinations. (**E**) Crosslinking of cohesin containing a cysteine pair specific for engaged heads, in the presence of ATP, DNA, or Scc2 in various combinations and in the absence of 1mM $MgCl_2$. (**F**) BMOE crosslinking of cohesin with a cysteine pair specific for J head association, in the presence of ATP, DNA, or Scc2 in various combinations.

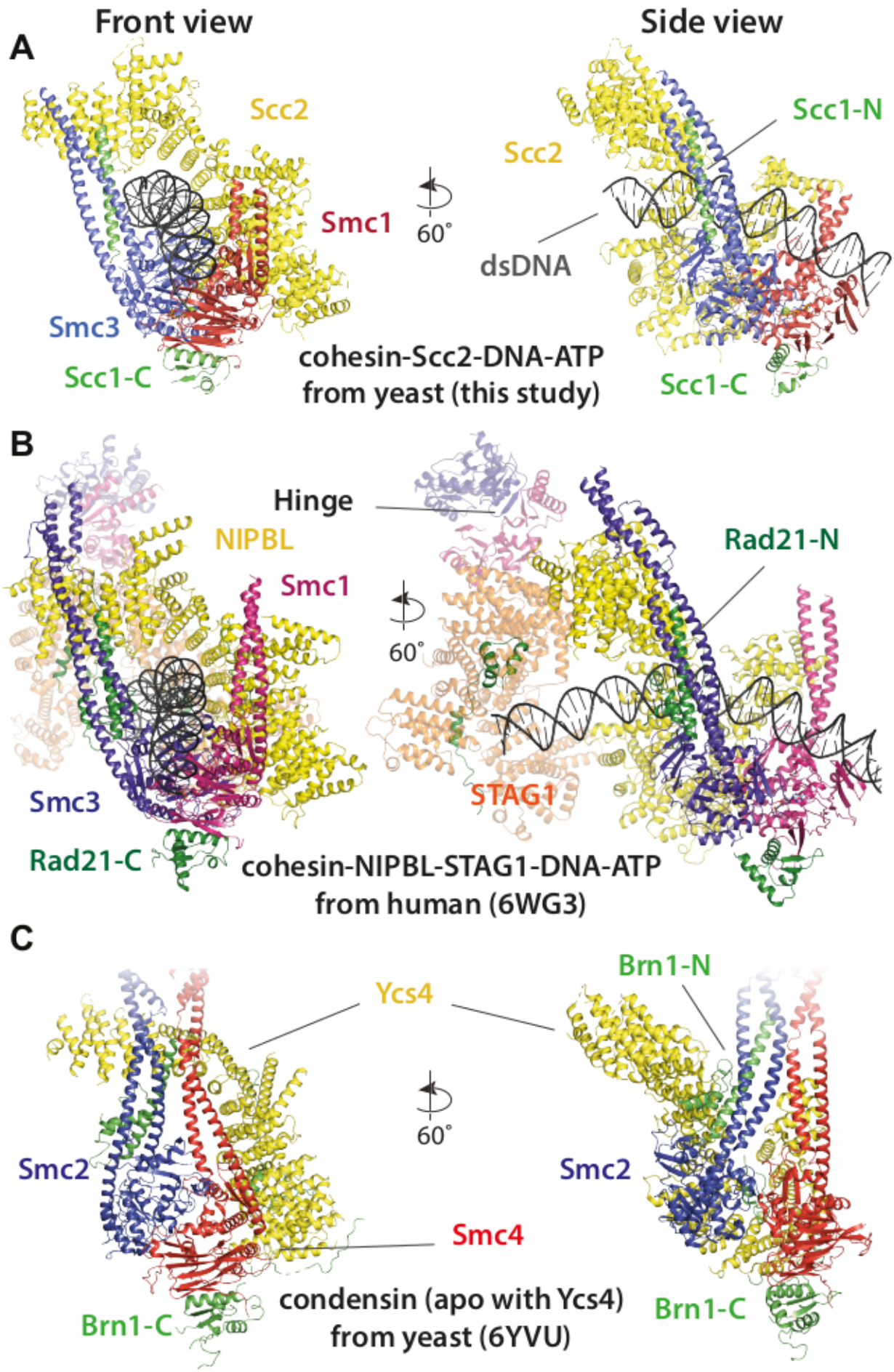

A
Front view
Side view
Scc2
Scc1-N
Scc2
Smc1
60°
dsDNA
Smc3
Scc1-C
cohesin-Scc2-DNA-ATP
from yeast (this study)
Scc1-C
B
Hinge
NIPBL
Rad21-N
Smc1
60°
Smc3
STAG1
Rad21-C
cohesin-NIPBL-STAG1-DNA-ATP
from human (6WG3)
C
Brn1-N
Ycs4
60°
Smc2
Smc2
Smc4
condensin (apo with Ycs4)
from yeast (6YVU)
Brn1-C
Brn1-C

Figure S6. Comparison of cryo-EM structures

(**A**) Cohesin tetramer (yeast, this study), (**B**) pentamer (human, 6WG3) (Shi et al., 2020) and (**C**) condensin (yeast, 6YVU) (Lee et al., 2020) are shown in two orientations (same or similar to Fig. 7A and 8A).

Movie 1. A model for the formation of the clamped E-S/E-K state of cohesin. According to this model cohesin transitions from a putative 'bridged state' (modelled on the same state of yeast apo condensin as observed by cryo-EM) (Lee et al., 2020) in which Scc2, analogous to Ycs4 in condensin, bridges the Smc1/3 heads. In the bridged state Scc2's DNA binding surface becomes accessible for DNA to attach without impediment and positions it for the next step, namely clamping. ATP-binding driven head engagement, achieved through a rotation of the Smc3 head relative to the rest of the complex, and with Scc2's and Smc1's relative orientations staying the same, results in entrapment in the E-S compartment. Because the disordered kleisin chain has to be pushed upwards during the clamping, the DNA is also in the E-K compartment. The initial binding of DNA to the Scc2 DNA binding site guides the DNA through the large opening of the heads generated by the bridged state and leads to the final clamped state that has been described in this study. The movie is a simple morph between a putative bridged state of cohesin modelled on the same state in condensin and the high-resolution cryo-EM structure of the clamped state determined in this study, with a few clashes removed manually because cohesin and condensin subunits, in particular Scc2 and Ycs4 are not completely homologous structurally.

Table 1.

S. cerevisiae cohesin	+ 40 bp DNA		+ circular DNA
	Tetramer Smc1E1158Q, Smc3E1155Q, Scc1, Scc2C2, ATP	**Tetramer head segment** Smc1E1158Q, Smc3E1155Q, Scc1, Scc2C2, ATP (EMD-11585, PDB 6ZZ6)	**T. head segment** Smc1E1158Q, Smc3E1155Q, Scc1, Scc2C2, ATP
Data collection and processing			
Magnification	81,000	81,000	105,000
Voltage (kV)	300	300	300
Electron exposure (e–/Å^2)	55	55	40
Defocus range (μm)	1.5 ~ 3.3	1.5 ~ 3.3	0.6 ~ 1.0
Pixel size (Å)	1.07	1.07	1.0
Symmetry imposed	*C1*	*C1*	*C1*
Initial particle images (no.)	1,516,413	2,314,881	65,442
Final particle images (no.)	21,343	588,164	23,728
Map resolution (Å)	10	3.35	7.3
FSC threshold	0.143	0.143	0.143
Map resolution range (Å)	10 – 50	3.2 – 50	7.3 – 50
Refinement			
Initial model used (PDB)		1W1W, 4UX3, 5ME3	
Model resolution (Å)		3.35	
FSC threshold		0.143	
Model resolution range (Å)		3.2 – 4.5	
Map sharpening *B* factor (Å^2)		-88.25	
Model composition			
Non-hydrogen atoms		16556	
Protein residues		1909	
		Smc1(2-71, 87-195, 1044-1224); Smc3(1-224, 997-1071, 1104-1222); Scc1(67-103, 502-510, 519-555); Scc2(221-236, 250-262, 278-291, 307-322, 336-373, 386-589, 597-634, 647-662, 678-617, 927-1049, 1059-1078, 1090-1184, 1203-1342, 1355-1398, 1413-1434, 1448-1456, 1466-1475)	
Nucleotide residues		68 (poly-A)	
Ligands		2 ATP, 2 Mg	
B factors (Å^2)			
Protein		11.21	
Nucleotide		68.00	
Ligand		19.62	
R.m.s. deviations			
Bond lengths (Å)		0.002	
Bond angles (°)		0.556	
Validation			
MolProbity score		1.80 (85th percentile)	
Clashscore		8.36	
Poor rotamers (%)		0.06	
Ramachandran plot			
Favored (%)		94.98	
Allowed (%)		5.02	
Disallowed (%)		0.00	

REAGENTS	SOURCE	IDENTIFIER
Antibodies		
Anti-His (mouse)	Sigma	Cat# SAB1305538-400UL
Anti-mouse HRP	ThermoFisher	Cat# 62-6520
Anti-Smc3 (mouse)	Bethyl Laboratories	Cat# A300-060A
Anti-Strep HRP	iba	Cat# 2-1502-001
Bacterial Strains		
DH10Bac	ThermoFisher	Cat# 10361012
PIR1	ThermoFisher	Cat# C101010
TOP10	ThermoFisher	Cat# C404010
Cell Lines		
Sf9	ThermoFisher	Cat# 11496015
Chemicals and Recombinant Proteins		
ATP Lithium Salt	Sigma	Cat# 11140965001
Bismaleimidoethane (BMOE)	ThermoFisher	Cat# 22323
Complete EDTA free protease inhibitor cocktail	Roche	Cat# 4693132001
Cre Recombinase	New England Biolabs	Cat# M0298S
Desthiobiotin	Fisher Scientific	Cat# 12753064
EtBr	ThermoFisher	Cat# 15585011
Fetal Bovine Serum	Sigma	Cat# 12303C
FuGENE HD Transfection reagent	Promega	Cat# E2311
Gibson Assembly Mix	New England Biolabs	Cat# E2611L
HiLoad 16/60 Superdex 200	GE Healthcare	Cat# GE28-9893-35
HiSpeed Plasmid Maxi Kit	Qiagen	Cat# 12663
HiTrap Q HP	GE Healthcare	Cat# GE29-0513-25
Immobilon Western ECL	Millipore	Cat# WBLKS0500
NuPAGE 3-8% Tris-Acetate Protein Gels	ThermoFisher	Cat# EA0378BOX
PMSF	Sigma	Cat# 329-98-6
Quick Coomassie Stain	Generon	Cat# GEN-QC-STAIN-1L
RNase A	Roche	Cat# 10109169001
Sf900 II SFM	ThermoFisher	Cat# 10902104
StrepTrap HP	Fisher Scientific	Cat# 11540654
Supernuclease	SinoBiological	Cat# SSNP01
Superose 6 Increase 10/300 GL	VWR	Cat# 29-0915-96
TCEP	ThermoFisher	Cat# 20490
4xLDS	ThermoFisher	Cat# NP0007
Critical Commercial Assays		
EnzChek phosphate assay kit	Invitrogen	Cat# E6646

Recombinant DNA
pACEbac1 *SMC1-His*
pACEbac1 *SMC3*
pACEbac1 *SMC1-His SMC3*
pACEbac1 *SCC2*$^{133-1493}$*-2xStrepII*
pACEbac1 *SCC2-2xStrepII*
pACEbac1 *2xStrepII-Scc2*$^{151-1493}$
pACEbac1 *2xStrepII-SCC3*
pIDC *SCC1-2xStrepII*
pIDC *Scc1*$^{269-451}$*-2xStrepII*
pIDC *Scc1*$^{150-298}$*-2xStrepII*
pIDC *His-SCC4*

***S. cerevisiae* strains**	
MATa ura::ADH1 promoter-OsTIR1-9myc::URA3 Scc3-PK3-aid::KanMX4 SCC1-HA3::HIS3	KN20783
MATa Scc3-PK3-aid::KanMX4 SCC1-HA3::HIS3	KN20785
MATa/alpha scc3::NatMX4/WT	KN21079
MATa/alpha scc3::NatMX4/WT, leu::Scc3-HA3::LEU	KN21273
MATa Scc1-PK9::KanMX scc2-45::natMX (L545P D575G)	KN22390
C. glabrata Mata, SCC1-PK9::NATMX4	KN23308
MATa Scc1-PK9::KanMX scc2-45::natMX (L545P D575G) lys2::Scc2-HyGMX	KN24185
C. glabrata Mata, SCC1-HA3::NATMX4	KN25532
MATa Scc1-PK9::KanMX scc2-45::natMX (L545P D575G) LYS2::Scc2(S717L,K721E)-HygMX	KN27010
MATa/alpha scc3::NatMX4/WT, leu::Scc3 (K224E, K225E, R226E)-HA3::LEU	KN27539
MATa scc3::NatMX4, Scc1-PK6::TRP1, leu::Scc3-HA3::LEU	KN27542
MAT alpha scc3::NatMX4, Scc1-PK6::TRP1, leu::Scc3 (K224E, K225E, R226E)-HA3::LEU	KN27547
MATa/alpha scc3::NatMX4/WT, leu::Scc3 (K423E, K513E, K520E)-HA3::LEU	KN27696
MATa scc3::NatMX4, Scc1-PK6::TRP1, leu::Scc3 (K423E, K513E, K520E)-HA3::LEU	KN27697
MATa/alpha scc3::NatMX4/WT, leu::Scc3 (K224E, K225E, R226E, K423E, K513E, K520E)-HA3::LEU	KN27763
MATa ura::ADH1promoter-OsTIR1-9myc::URA3, Scc3-PK3-aid::KanMX4, leu::Scc3-HA3::LEU	KN27796
MATa ura::ADH1 promoter-OsTIR1-9myc::URA3, Scc3-PK3-aid::KanMX4 leu::Scc3 (K224E, K225E, R226E, K423E, K513E, K520E)-HA3::LEU	KN27802
MATa ura::ADH1promoter-OsTIR1-9myc::URA3, Scc3-HA3-aid::KanMX4, Scc1-PK6::TRP1, leu::Scc3 (K224E, K225E, R226E, K423E, K513E, K520E)-HA3::LEU	KN27804
MATa ura::ADH1promoter-OsTIR1-9myc::URA3, Scc3-HA3-aid::KanMX4, Scc1-PK6::TRP1, leu::Scc3-HA3::LEU	KN27821
MATa ura::ADH1promoter-OsTIR1-9myc::URA3, Scc3-HA3-aid::KanMX4, leu::Scc3-HA3::LEU, Scc2-PK9::NatMX	KN28075
MATa ura::ADH1 promoter-OsTIR1-9myc::URA3, Scc3-HA3-aid::KanMX4, Scc2-PK9::NatMX, leu::Scc3 (K224E, K225E, R226E, K423E, K513E, K520E)-HA3::LEU	KN28287

software, algorithm	
RELION 3.1	doi:10.1016/j.jsb.2012.09.006
CtfFind4	doi:10.1016/j.jsb.2015.08.008
Warp	doi:10.1038/s41592-019-0580-y
CrYOLO 1.5	doi:10.1038/s42003-019-0437-z
Chimera	https://www.cgl.ucsf.edu/chimera/
ChimeraX 1.0	https://www.cgl.ucsf.edu/chimerax/
COOT	doi:10.1107/S0907444910007493
MAIN	doi:10.1107/S0907444913008408
Phenix.real_space_refinement	doi:10.1107/S2059798318006551
PYMOL 2	https://pymol.org/2/
SWISS-MODEL	https://swissmodel.expasy.org

Other	
Quantifoil Au 2/2 holely carbon 200 mesh cryoEM grids	Quantifoil GmbH
Ultrafoil 2/2 holely gold 200 mesh cryoEM grids	Quantifoil GmbH