Multiple spatial behaviours govern

² social network positions in a wild

³ ungulate

- 4 Gregory F Albery^{1,2*}, Alison Morris², Sean Morris², Josephine M
- 5 Pemberton², Tim H. Clutton-Brock^{2,3}, Daniel H Nussey², Josh A Firth^{4,5}
- 6 1. Department of Biology, Georgetown University, Washington, DC
 - 2. Institute of Evolutionary Biology, University of Edinburgh, Edinburgh, UK
- 8 3. Department of Zoology, University of Cambridge, Cambridge, UK
- 9 4. Department of Zoology, University of Oxford, Oxford, UK
- 10 5. Merton College, University of Oxford, Oxford, UK
- 11 *gfalbery@gmail.com

12 Abstract

7

- 13 The structure of wild animal social systems depends on a complex combination of intrinsic
- 14 and extrinsic drivers. Population structuring and spatial behaviour are key determinants of
- 15 individuals' observed social behaviour, but quantifying these spatial components alongside
- 16 multiple other drivers remains difficult due to data scarcity and analytical complexity. We
- 17 used a 43-year dataset detailing a wild red deer population to investigate how individuals'
- 18 spatial behaviours drive social network positioning, while simultaneously assessing other
- 19 potential contributing factors. Using Integrated Nested Laplace Approximation (INLA) multi-
- 20 matrix animal models, we demonstrate that social network positions are shaped by two-
- 21 dimensional landscape locations, pairwise space sharing, individual range size, and spatial
- 22 and temporal variation in population density, alongside smaller but detectable impacts of a
- 23 selection of individual-level phenotypic traits. These results indicate strong, multifaceted
- 24 spatiotemporal structuring in this society, emphasising the importance of considering
- 25 multiple spatial components when investigating the causes and consequences of sociality.
- 26 Word Count: Abstract: 143 words; Main text: 4720 words
- 27 Number of figures: 4; Number of tables: 0; Number of references: 53

28 Authorship Statement

- 29 GFA conceived the study, analysed the data, and wrote the manuscript, advised by JAF. AM
- 30 and SM collected the data. JAF, JMP, THCB, and DN commented on the manuscript.

31 Data Accessibility Statement

- 32 The code used here is available at <u>https://github.com/gfalbery/Spocial_Deer</u>. On
- acceptance, the data will be uploaded to the same repo, which will be archived on Zenodo.

34 Introduction

35 Social behaviour is an integral component of an animal's phenotype, driving processes 36 including disease transmission, mating, learning, and selection (Croft et al. 2008; 37 VanderWaal et al. 2014; Krause et al. 2015; Firth et al. 2018; Sah et al. 2018; Silk et al. 38 2019; Firth 2020). Contemporary studies of animal behaviour often use social networks to 39 derive individual-level social network positions, under the notion that between-individual 40 variation in network positioning is indicative of between-individual variation in social 41 behaviour (Franks et al. 2010; Krause et al. 2015; Sosa et al. 2020). However, an animal's 42 position in its social network is also dependent on its own spatial behaviour (Webber & 43 Vander Wal 2018; Albery et al. 2020a), and on a range of extrinsic factors: demography 44 determines local population density and structuring (Shizuka & Johnson 2019), while the 45 environment shapes resource distributions, movement corridors, and emergent patterns of 46 space use, all of which will influence the architecture of the social system (Firth & Sheldon 47 2016; Webber & Vander Wal 2018; Farine & Sheldon 2019; He et al. 2019). As such, it is 48 important to consider spatial behaviour and environmental context when assessing the 49 causes and consequences of individual-level social network positioning (Webber & Vander 50 Wal 2018; He et al. 2019; Albery et al. 2020a), yet doing so remains difficult in most systems 51 due to the complexity of spatial-social analyses that incorporate these processes.

52 The spatial drivers of social network structure are poorly understood because they are highly 53 multivariate and (therefore) difficult to analyse. On the one hand, there is strong support for 54 simpler "first-order" associations between spatial and social behaviour. For example, spatial 55 proximity and social connections are often correlated, because individuals that share more 56 space are more likely to associate or interact. This finding holds for diverse taxa including elk 57 (Vander Wal et al. 2014), raccoons (Robert et al. 2012), birds (Firth & Sheldon 2016), and 58 myriad other systems. Similarly, spatial and social network centrality are occasionally found 59 to correlate (Mourier et al. 2019), as are temporal variation in population density and social 60 contact rates (Sanchez & Hudgens 2015), and the social environment can drive spatial 61 behaviour (Firth & Sheldon 2016; Spiegel et al. 2016). Spatial behaviours can be 62 summarised using a wide range of metrics, including individuals' spatial activity levels (e.g. 63 home range area), pairwise space sharing (e.g. distances or home range overlaps), 64 demographic structure (e.g. temporal population size or local conspecific density), and point 65 location on the two-dimensional landscape. For example, are more social individuals simply 66 wider-ranging, leading them to make more contacts? Do they most often inhabit areas of 67 high population density or well-used movement corridors? These variable spatial 68 components take a combination of different data structures, and are therefore difficult to 69 include in the same models, particularly in large numbers and alongside a range of other

individual-level phenotypes. It is therefore unclear to what extent individuals' social network
positions emerge from 1) their own social behaviour; 2) their own spatial behaviour; 3) their
situation within the population and broader social network; 4) other aspects of their biotic and
abiotic environment such as landscape structure; and 5) intrinsic phenotypic traits that

74 researchers are commonly interested in investigating.

75 Several frameworks have been proposed to facilitate the untangling of spatial and social 76 processes in wild animals (Jacoby & Freeman 2016; Silk et al. 2018, 2019; Webber & 77 Vander Wal 2018; Mourier et al. 2019; Albery et al. 2020a). To date, statistical methodology 78 focusses on incorporating spatial behaviours into the node-and-edge structure of network 79 data, using e.g. null network permutations (Firth & Sheldon 2016), spatially embedded 80 networks (Daraganova et al. 2012), and nested "networks of networks" composed of 81 movement trajectories (Mourier et al. 2019). Many such analyses involve reducing 82 movement patterns into some form of spatial network based on home range overlap or 83 spatial proximity between dyads (Mourier et al. 2019). For example, statistical models 84 named "animal models" can examine spatial variation by fitting such matrices as variance 85 components, potentially alongside other dyadic similarity matrices (i.e., pairwise measures of 86 similarity), to quantify genetic and non-genetic contributions to individuals' phenotypes 87 (Kruuk 2004; Stopher et al. 2012b; Regan et al. 2016; Thomson et al. 2018; Webber & 88 Vander Wal 2018). As yet, the focus on controlling for spatial autocorrelation using space 89 sharing and network permutations has contributed to a lack of clarity concerning the role that 90 spatial behaviour and environmental context play in driving social network positioning 91 (Albery et al. 2020a).

92 Studies across ecological disciplines increasingly use Integrated Nested Laplace

93 Approximation (INLA) models to control for spatial autocorrelation in a multitude of contexts

94 (Lindgren et al. 2011; Lindgren & Rue 2015; Zuur et al. 2017). As well as including fixed and

95 random effects to quantify individual-level drivers, these models can incorporate dyadic

96 space sharing components (Holand *et al.* 2013) and stochastic partial differentiation

97 equation (SPDE) effects to model 2-dimensional spatial patterns in the response variable,

98 thereby controlling for and estimating spatiotemporal variation associated with fine-scale

99 positioning within the landscape (Albery et al. 2019). As such, these models offer an exciting

100 opportunity to test and compare the roles of a range of spatial behaviours and

101 autocorrelation structures, alongside phenotypic drivers, in determining social network

102 positioning.

103 We address this question using the long-term study in the Isle of Rum red deer (Cervus

104 *elaphus*). These study animals comprise an unmanaged wild population with a contiguous

105 fission-fusion social system (Clutton-Brock et al. 1982). They experience strong 106 environmental gradients and exhibit spatial autocorrelation in a number of important 107 phenotypes: individuals with greater home range overlap have more similar behavioural and 108 life history traits (Stopher et al. 2012b), and those in closer proximity have more similar 109 parasite burdens (Albery et al. 2019); further, as with other matrilineal mammalian systems, 110 closely related individuals frequently associate (Clutton-Brock et al. 1982) and live closer 111 together (Stopher et al. 2012b). Individuals have highly repeatable home ranges (Stopher et 112 al. 2012b) that decline in size over their lifetimes, predicting declining survival probability 113 (Froy et al. 2018). The study area has a strong spatial gradient in resource availability, with 114 high-quality grazing heavily concentrated in the far north of the system, and with most 115 individuals aggregating around this area, such that population density decreases outwards 116 towards the edge of the study population (Clutton-Brock et al. 1982). As such, the deer 117 comprise an ideal system for assessing spatial-social relationships in the wild. 118 To assess how individuals' spatial behaviours translate to social network positions, we 119 constructed fine-scale social networks from 43 years of censuses of the study population. 120 We derived 8 different individual-level network positioning measures of varying complexity 121 that are important to different social processes (Krause et al. 2015; Sosa et al. 2020). Using 122 multi-matrix animal models in INLA, we examined whether spatial locations, space sharing, 123 home range area, and local population density explained variation in network position 124 metrics, alongside a range of individual-, temporal-, and population-level factors. Specifically, 125 we aimed to test two hypotheses: that the structure of the social network would be highly 126 dependent on the distribution of population density in space; and that individuals' social 127 network centrality would be largely explained by their ranging behaviour, where wide-ranging 128 individuals were more likely to be socially well-connected. We further expected that space 129 sharing and point locations would uncover substantial spatial autocorrelation in social 130 network positioning, and that different social network metrics would exhibit different spatial

- 131 patterns and vary drivers. This not only comprises a large-scale empirical examination of the
- 132 factors shaping social network positions in this extensively monitored wild mammal, but also
- 133 provides a methodological advancement in developing powerful, flexible new methods
- (INLA-based multi-matrix animal models) with broad potential for examining spatial-socialprocesses in this and other systems.

136 Methods

137 Study system and censusing

The study was carried out on an unpredated long-term study population of red deer on the
Isle of Rum, Scotland (57°N,6°20'W). The natural history of this matrilineal mammalian

140 system has been studied extensively (Clutton-Brock et al. 1982), and we focussed on 141 females aged 3+ years, as these individuals have the most complete associated census 142 data, and few males live in the study area except during the mating period. Individuals are 143 monitored from birth, providing substantial life history and behavioural data, and >90% of 144 calves are caught and tagged, with tissue samples taken (Clutton-Brock et al. 1982). The 145 population thus has comprehensive genomic data, allowing high-powered quantitative 146 genetic analyses: most individuals born since 1982 have been genotyped at >37,000 SNPs, 147 distributed throughout the genome (e.g. Huisman, Kruuk, Ellis, Clutton-Brock, & Pemberton, 148 2016). Census data were collected for the years 1974-2017, totalling 423,070 census 149 observations. Deer were censused by field workers five times a month, for eight months of 150 the year, along one of two alternating routes (Clutton-Brock et al. 1982). Individuals' 151 identities, locations (to the nearest 100M), and group membership were recorded. Grouping 152 events were estimated by seasoned field workers according to a variant of the "chain rule" 153 (e.g. Castles et al., 2014), where individuals grazing in a contiguous group within close 154 proximity of each other (under ~10 metres) were deemed to be associating, with mean 130.4 155 groups observed per individual across their lifetime (range 6-943). The mortality period falls 156 between Jan-March, when there is least available food, and minimal mortality occurs outside 157 this period. We only used census records in each May-December period, from which we 158 derived annual social network position measures as response variables (Figure 1-2). We 159 elected to investigate this seasonal period because it stretches from the spring calving 160 period until the beginning of the mortality period, simplifying network construction and 161 avoiding complications arising from mortality events. Our dataset totalled 3356 annual 162 observations among 532 grown females (Figure 1).

163 In this system, female reproduction imposes substantial costs for immunity and parasitism

- 164 (Albery *et al.* 2020c), and for subsequent survival and reproduction (Clutton-Brock, Albon, &
- 165 Guinness, 1989; Froy, Walling, Pemberton, Clutton-Brock, & Kruuk, 2016). If a female
- reproduces, she produces 1 calf per year in the spring, generally beginning in May; the "deer
- 167 year" begins on May 1 for this reason. Here, reproductive status was classified into the
- 168 following four categories using behavioural observations: True Yeld (did not give birth);
- 169 Summer Yeld (the female's calf died in the summer, before 1st October); Winter Yeld (the
- 170 female's calf died in the winter, after 1st October); and Milk (calf survived to 1st May the
- 171 following calendar year).
- 172 Generating spatial and social matrices
- 173 All code is available online at https://github.com/gfalbery/Spocial_Deer. We constructed the
- 174 home range overlap (HRO) matrix using the R package AdeHabitatHR (Calenge 2011),
- 175 following previous methodology (Stopher *et al.* 2012b; Regan *et al.* 2016; Froy *et al.* 2018).

First, using a kernel density estimation method, we derived lifetime home ranges for each
individual with more than five census observations. Previous analysis has shown that this
system is robust to the number of observations used to generate home ranges (Froy *et al.*2018). We used lifetime home ranges to fit one value per individual in the animal models;
individual ranges (and range sizes) correlate strongly from year to year (Stopher *et al.*2012b; Froy *et al.* 2018). We derived proportional HRO of each dyad using Bhattacharya

Affinity (following Stopher *et al.* 2012b), producing values between 0-1 (i.e. no overlap tocomplete overlap).

184 To control for individuals' two-dimensional point locations, we used a Stochastic Partial

185 Differentiation Equation (SPDE) effect in INLA. This effect models the distance between

points to calculate spatial autocorrelation, using Matern covariance (Lindgren *et al.* 2011).

187 This random effect used individuals' annual centroids (mean easting and northing in a given

188 year) or lifetime centroids (mean easting and northing across all observations) as point

189 locations to approximate spatial variation in the response variable (Lindgren *et al.* 2011;

190 Albery *et al.* 2019).

191 We used a genomic relatedness matrix (GRM) using homozygosity at 37,000 Single

192 Nucleotide Polymorphisms, scaled at the population level (Yang *et al.* 2011; for a population-

193 specific summary, see Huisman *et al.* 2016). This matrix is well-correlated with pedigree-

194 derived relatedness metrics (Huisman et al. 2016). HRO was well-correlated with distance

195 between lifetime centroids (i.e., closer individuals tended to share more range), and both

196 were weakly but significantly correlated with genetic relatedness (Supplementary Figure 1).

197 To test whether social network positions could be explained by population density, we 198 derived the local density of individuals again using AdeHabitatHR (Calenge 2011). We 199 generated density kernels of observations, and then assigned individual deer their local 200 population density based on their location on this kernel, following previous methodology 201 developed in badgers (Albery et al. 2020b). This local density value was then fitted as a fixed 202 explanatory variable. We used four different density metrics, each examining the density of a 203 different observation type: lifetime centroids ("lifetime density"); annual centroids ("annual 204 density"); all observations across the study period ("sighting density"); and all observations in 205 the focal year ("annual sighting density"). Only one such density metric was fitted at once. 206 We also calculated annual home range areas (HRA) by taking the 70% isopleth of each 207 individual's annual space use distribution, following previous methodology (Froy et al. 2018). 208 This HRA variable was fitted as a fixed effect in the same way as local density.

209 We constructed a series of 43 annual social networks using "gambit of the group," where 210 individuals in the same grouping event (as described above) were taken to be associating

- 211 (Franks et al. 2010). Dyadic associations were calculated using the 'simple ratio index'
- 212 (Cairns & Schwager 1987) derived as a proportion of total sightings (grouping events) in
- 213 which the focal individuals were seen together: Sightings_{A,B}/(Sightings_A+Sightings_B-
- 214 Sightings_{A,B}), or Intersect_{A,B}/Union_{A,B}. In this dyadic matrix, 0=never seen together and
- 215 1=never seen apart.

216 Statistical Analysis

217 Metrics. Using the annual social networks, we derived eight individual-level network metrics 218 which are commonly used across animal social networks and have been considered in 219 detail(Whitehead 2008; Brent 2015; Krause et al. 2015; Firth et al. 2017). We set each of 220 these network metrics for use as response variables in separate INLA Generalised Linear 221 Mixed Models (GLMMs) with a Gaussian family specification. In increasing order of 222 complexity, our measures included four direct sociality metrics, which only take into account 223 an individual's connections with other individuals: 1) Group Size – the average number of 224 individuals a deer associated with per sighting; 2) Degree – the number of unique individuals 225 she was observed with; 3) Strength – sum of all their weighted social associations to others; 226 4) Mean Strength – the average association strength to each of the unique individuals she 227 was observed with (equivalent to strength divided by degree). We also included four more 228 complex "indirect" metrics (all using algorithms as specified from (Csardi & Nepusz 2006)), 229 which also take into account an individual's connections' connections: 5) Eigenvector 230 centrality – which considers the sum of their own connections and the sum of their 231 associates' connections; 6) Weighted Eigenvector – which is akin to eigenvector centrality 232 but also accounts for the weights of theirs, and their associates, connections; 7) 233 Betweenness – the number of shortest paths that pass through the focal individual to 234 traverse the whole network; 8) Clustering (local) – the tendency for an individual's contacts 235 to be connected to one another, forming triads. The raw, untransformed correlations were 236 assessed for all metrics, and R lay between -0.5 and 0.879 across metrics (Supplementary 237 Figure 2). When modelling them as response variables, to approximate normality, all social 238 metrics were square root-transformed apart from eigenvector centralities (which were left 239 untransformed), group size (which was cube root-transformed), and betweenness (which 240 was log(X+1)-transformed). Each social network metric was fitted as a response variable in 241 a separate model set (as outlined conceptually in Figure 1).

242 **Base model structure.** We ensured that all models followed the same base structure.

243 Random effects included individual identity and year (categorical random intercepts), as well

- as the genetic relatedness matrix. Fixed effects included Age (continuous, in years),
- 245 Reproductive Status (four categories: True Yeld; Summer Yeld; Winter Yeld; and Milk), and
- 246 Number of observations (continuous, log-transformed to approximate normality), as well as

247 year-level continuous factors including Year (continuous) and that year's study Population

248 Size (log-transformed). All continuous response and explanatory variables were

standardised to have a mean of zero and a standard deviation of 1. Fixed effect estimates

- were provided by the mean and 95% credibility intervals of the posterior estimate
- 251 distribution.

252 Adding spatial components. To investigate the divergent value of different spatial

- 253 behaviours, we iteratively added spatial effects to the base model, investigating which
- 254 behaviours best fit the data. These spatial behaviours corresponded to four broad
- components in Figure 1: space sharing (HRO matrix); home range area (HRA); point
- 256 locations (SPDE effect); and local population density (density fixed effect). For space
- sharing, we only used one metric: lifetime HRO (see above). For point locations, we selected
- between 1) lifetime centroids; 2) annual centroids; and 3) annual centroids with a
- 259 spatiotemporally varying annual field. For density, we used the four metrics outlined above
- 260 ("lifetime", "annual", "sighting", and "annual sighting" density). To distinguish between
- 261 competitive models we used Deviance Information Criterion (DIC). In each round, we added
- each spatial behaviour individually and then kept the best-fitting one, until all four had been
- added or their addition did not improve the model, using a cutoff of 2 DIC.
- 264 Comparing all spatial and non-spatial drivers. To compare the relative importance of all 265 fixed and random effects, we examined the model's predicted values and their correlations 266 with the observed values, representing the proportion of the variance that was explained by the model (i.e., R²). We used the model to predict each social behaviour metric, and 267 268 iteratively held each explanatory variable's predictions at the mean, one at a time. We then 269 assessed the squared correlations of these values with the observed values, relative to 270 those of the full model. Variables with greater effects in the model produced less accurate 271 predicted values when held constant.

272 **Results**

- 273 Spatial behaviours were important in determining all eight individual-level social network
- 274 position variables. The non-spatial model was far the poorest-fitting for all eight metrics, and
- the DIC changes associated with adding spatial components were substantial (Figure 3A).
- 276 Generally, wide-ranging individuals and those living in areas of greater population density
- tended to be more central, and space sharing and point location effects both revealed
- 278 substantial spatial autocorrelation (Figure 3).
- As expected, home range area and population density had substantial effects on social
 network centrality (Figure 3). Population density was positively associated with all centrality

measures except betweenness and clustering (Figure 3A), and the best-fitting density metric
was annual density. Individuals with larger home ranges likewise tended to be more social,
except in the case of clustering (no effect) and mean strength, which were negatively
associated with HRA (Figure 3A).

285 Notably, point location-based SPDE effects tended to improve model fit over these fixed 286 effects, and had a greater effect than on model fit space sharing HRO effects, even when 287 conceptualised at the same timescale (i.e., across the individual's lifetime). Investigating the 288 R^2 components of the models containing only HRO (i.e., without SPDE effects) revealed that 289 in general spatial overlap accounted for more variation than the genetic matrix 290 (Supplementary Figure 3), but comparing these with the other models revealed that the point 291 location effects contributed more than either of these matrices (Figure 3B). Annually varying 292 centroids further improved model fit, and allowing the spatial field to vary between years in 293 our spatiotemporal models improved models even more (Figure 3A). Although the space 294 sharing and genomic relatedness matrices had similar sized impacts on the full models 295 (Figure 3B), removing the SPDE effect resulted in a substantial increase in the HRO effect, 296 but with very little impact on the GRM's R^2 (Supplementary Figure 3). These findings were 297 relatively consistent across all metrics (Figure 3A-B), although the SPDE effect was notably 298 smaller for betweenness (Figure 3B). Taken together, these results reveal that lifetime space 299 sharing was good at accounting for variation in social behaviour, but that its effect was 300 surpassed by increasingly complex temporal formulations of point location effects. 301 We compared the importance of all fixed and random effects by predicting selectively from

302 the model, revealing overwhelmingly strong effects of spatiotemporal drivers (Figure 3B).

303 Our models fit well and explained a substantial amount of variation in social network

304 centrality (>70%); the majority of the fit was lent by a combination of the INLA SPDE effect,

305 fixed effects of local population density, and random effects of year (Figure 3B). Space

306 sharing (HRO) and home range area (HRA) had comparatively small effects.

307 Observations also had a notable impact for Degree, Betweenness, and Clustering (Figure 308 3B). Fixed effects for year and observation numbers were generally strong and significantly 309 positive across metrics, except in the case of clustering, for which observation number's 310 effect was significantly negative (Figure 3B). There were also small positive effects of 311 population size on betweenness and degree centrality (Figure 3B).

312 Although individual-level drivers (reproduction, age, and individual identity) had a negligible

impact on all variables' R² (Figure 3B), many had a significant effect (i.e., their 95%)

314 credibility intervals did not overlap with zero; Figure 3C). Individuals whose calves lived to

315 the winter and then either died before the 1st May ("Winter Yeld") or survived ("Milk") were

316 generally less central than those that did not give birth ("True Yeld") or whose calf died

317 before 1st October ("Summer Yeld"). Similarly, there were minor age-related decreases in

318 network centrality for the direct metrics (Group Size, Degree, and Strength; Figure 3C).

319 To investigate spatial patterns of sociality when accounting for our fixed and random effects, 320 we projected the annual SPDE random effect in two-dimensional space (Figure 4; 321 Supplementary Figures 5-12). The spatial distributions of network centrality metrics were 322 highly variable, but direct metrics generally peaked in the centre of the study system and 323 decreased outwards (Figure 4). Mean Strength was an exception, being lowest in the centre 324 and increasing outward (Figure 4D); Clustering was patchily distributed, such that no clear 325 pattern was evident (Figure 4H); and Betweenness was slightly offset, being highest in the 326 north-northeast of the study area rather than in the central north (Figure 4G). The range of 327 autocorrelation also varied among metrics; Betweenness and Clustering had notably shorter 328 ranges than the other metrics (Supplementary Figure 4). We also plotted the spatial fields 329 through time, revealing substantial variation in the spatial fields across the study period 330 (Supplementary Figures 5-12).

331 Discussion

332 The role of spatial behaviour in driving social network structure

333 The position individuals occupy within their social networks can affect many aspects of their 334 ecology and evolution (Krause et al. 2015; Firth et al. 2018; Sah et al. 2018), and our results 335 confirm the powerful role of fine-scale spatial context in shaping such traits (e.g. Farine & 336 Sheldon, 2019; Mourier et al., 2019; Webber & Vander Wal, 2018). Capitalising on our 337 models' ability to compare the influence of a wide range of spatial and non-spatial 338 components, we found that spatial behaviour and environmental context were the most 339 important determinants of social network centrality -- more so than a suite of individual-level 340 phenotypes and demographic factors. Individuals with larger ranges and inhabiting higher-341 density areas were more central in the social network, revealing the important role of 342 individual spatial activity levels and location within the broader population structure. As 343 expected, models were further improved when we incorporated pairwise space sharing and 344 two-dimensional point locations, demonstrating that an individual's social network position is 345 not determined simply by the density of nearby individuals and by its own spatial activity, but 346 by other aspects of the fine-scale surrounding environment such as microclimate, resource 347 distribution, and landscape architecture (Spiegel et al. 2018; Webber & Vander Wal 2018; 348 He et al. 2019). Reciprocally, individuals may be altering their spatial behaviour, e.g. opting 349 to share more space or live closer together if they are more socially connected (Firth & 350 Sheldon 2016; Spiegel et al. 2016). As such, we propose that social network studies should

351 more regularly incorporate both space sharing and (temporally varying) point locations in 352 their statistical approaches to anticipate these effects, alongside specific spatial behaviours 353 thought to drive social network position. This practice will help to buffer for the fact that the 354 spatial environment not only correlates with social proximity, but can alter the fabric of the 355 network itself.

356 The landscape of sociality

357 One of the foremost advantages of our approach is the ability to flexibly investigate two-358 dimensional spatial patterns of social network centrality. This allowed us to qualitatively 359 assess the spatial structure of the social network, while providing clues towards the causal 360 factors. Most notably, betweenness peaked in the north-northeast of the system, likely 361 because the far northeastern community is relatively isolated from the rest of the population 362 due to the landscape structure (Figure 2), so that many 'social paths' that traverse the 363 population (the criteria for betweenness centrality) go through individuals in this intermediate 364 (north-northeast) area. That is, individuals living in this area are more likely to be connected 365 to both the far eastern communities and the central and western ones.

366 As expected, direct centrality metrics (group size, degree, and strength) were affected by 367 local population density, which peaks in the central north study area due to the concentration 368 of high quality grazing (Clutton-Brock et al. 1982). Individuals' resource selection behaviours 369 increase local density in this area (Clutton-Brock et al. 1982), and will increase social 370 connectivity as a result (Ostfeld et al. 1986; Sanchez & Hudgens 2015; Webber & Vander 371 Wal 2018). This comprises strong evidence for density-related increases in social contact 372 frequency, and accentuates the vital importance of considering resource distribution, habitat 373 selection, and population structure when examining social network correlates (Spiegel et al. 374 2016; Webber & Vander Wal 2018; Farine & Sheldon 2019; He et al. 2019). However, 375 because density was accounted for as a fixed effect in the models, the spatial patterns of 376 location effects for the direct metrics did not strictly follow the spatial pattern of density. 377 Instead, these metrics peaked in the centre of the study population, demonstrating that 378 individuals living in this central region are more well-connected when accounting for 379 population density. Combining these spatial components allowed us to effectively 380 differentiate what we do know (that greater population density drives increased social 381 connectedness) from what we do not (the drivers of greater sociality for individuals in the 382 central area). Without using the SPDE effect (i.e., relying only on generalised pairwise space 383 sharing rather than accounting for specific two-dimensional spatial patterns), these insights 384 into these patterns may have been harder to detect. An alternative method could involve 385 splitting the population into subpopulations and analysing them separately or comparing 386 them, but this method has been shown to be less powerful in this population (Albery et al.

2019), and is ultimately based on arbitrary choices if a population is mixed. The causes of the spatial distribution of clustering remain unresolved, but the pattern highlights areas where individuals are connected together in triads or tight cliques, and appears to be negatively correlated with betweenness (Figure 4). For traits such as this, it is unlikely that a simpler explanatory variable could be formulated to quantify the spatial-social processes at play.

393 Regardless of the causes of the spatial patterns, such fine-scale variation across the 394 landscape holds important ecological consequences, particularly for the more complex 395 network metrics. For instance, the areas of high clustering may act as 'incubator' areas 396 where cliques can develop new socially influenced behaviours (Centola 2018; Guilbeault et 397 al. 2018; Firth 2020) such as cooperative behaviours (Rand et al. 2011). The high contact 398 rates in the northern central areas might sustain high local burdens of directly transmitted 399 diseases (Cote & Poulin 1995), while individuals inhabiting the high-betweenness 400 intermediate areas may be important for transmitting novel diseases across the population 401 as a whole (VanderWaal et al. 2014).

402 Analytical benefits of INLA animal models

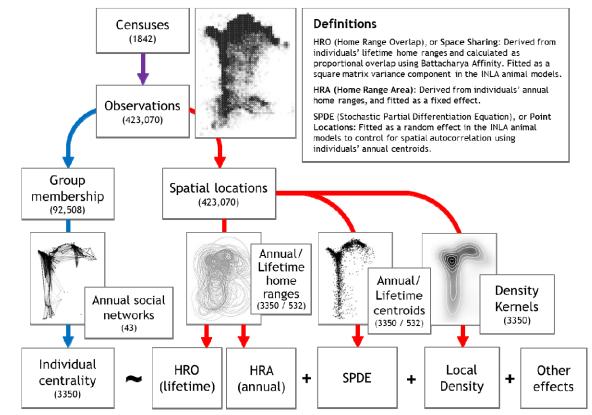
Analyses using multiple layers of different behaviours are well-suited to extricating space 403 404 and sociality in wild animal systems (Silk et al. 2018; Webber & Vander Wal 2018; Finn et al. 405 2019), and there is increasing conceptual and analytical overlap with the related field of 406 movement ecology (Jacoby & Freeman 2016; Mourier et al. 2019; Pasquaretta et al. 2020). 407 Notably, many spatial-social studies suffer from the necessity to reduce complex movement 408 patterns into simpler metrics, which risks losing important information in the process. As 409 such, recent studies have pushed for researchers to incorporate movement trajectories 410 themselves into complex network data structures (Mourier et al. 2019). Our approach allows 411 incorporation of multiple dyadic and non-dyadic behavioural measures, and with several 412 analytical timescales, offering an alternative workaround to this problem. Although other 413 methods can control for point locations (e.g. using autoregressive processes and 414 row/column effects; Stopher et al. 2012b), INLA models allow greater precision, fit quickly, 415 and allow incorporation of spatiotemporal structuring. Furthermore, plotting the SPDE effect 416 in two dimensions, as in Figure 4, gives an easily interpretable and intuitive portrayal of 417 network traits in space that can be hard to visualise using other methods. For these reasons, 418 we highly recommend further exploration of INLA animal models as a flexible method with 419 which to extricate individual, demographic, spatial, and temporal contributors to sociality 420 where sample sizes are sufficient (Thomson et al. 2018; Webber & Vander Wal 2018). In 421 addition to carrying out network-level manipulations (Daraganova et al. 2012; Davis et al. 422 2015; Firth & Sheldon 2016; Farine 2017), researchers concerned about spatial confounding

423 could implement relatively familiar linear models of social behaviour, but with additional

424 spatial components such as SPDE random effects and similarity matrix variance

425 components, with trustworthy and interpretable results (Albery et al. 2020a).

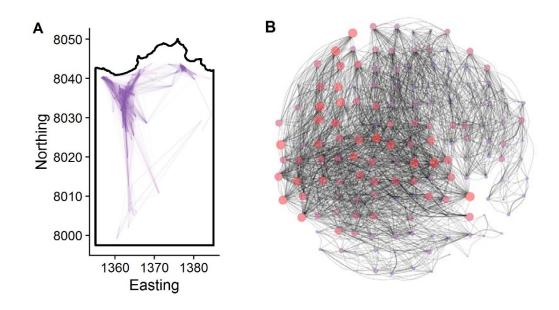
426 Although space accounted for an overwhelming amount of variation, many non-spatial 427 factors had substantial effects. The categorical random effect for interannual variation was 428 substantial, and there were detectable linear annual effects and population size effects, as 429 expected given the important roles of demography in structuring social networks (Shizuka & 430 Johnson 2019). Interestingly, there was a substantial positive association with study year 431 that was not attributable to the growth in population size over the same period. It is possible 432 that this represents a change in the deer's social phenotypes over time, although the 433 potential specific mechanisms now would benefit from further examination. Individual-level 434 factors had weaker contributions to model fit and smaller effect sizes: most notably, genetic 435 and individual random effects were negligible when spatial autocorrelation was accounted 436 for, confirming the importance of considering space when assessing heritability 437 independently of space in this population (Stopher et al. 2012a). Nevertheless, individual-438 level effects were encouragingly still detectable and significant, particularly for simpler 439 "direct" metrics. It is possible that more complex social network positions are less 440 determined by individual social behaviours, particularly for animals with fission-fusion 441 societies such as the deer; this hypothesis could be tested using similar spatial-social 442 analyses in a number of other systems. This finding demonstrates that even when spatial 443 structuring plays a vital role in determining social network structure, controlling for this 444 structuring analytically can reveal important, conservative individual-level effects. Future 445 analyses within this population, and potentially other long-term studies, could take 446 advantage of this framework by including environmental drivers such as food availability and 447 climatic factors to explain patterns of social connectivity, while further unpicking the causes 448 of the individual-level trends that we observed.



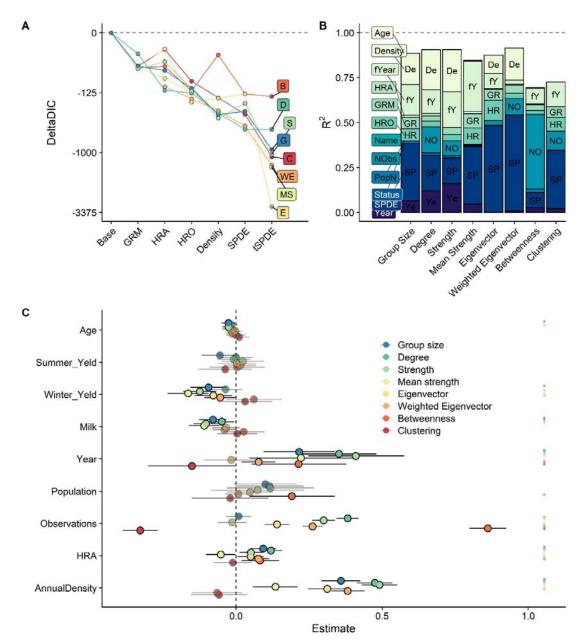
450

451

Figure 1: Data processing and analysis pipeline, demonstrating how behavioural census
data were collected, used to derive social and spatial behavioural traits, and fitted in INLA
animal model GLMMs. Numbers in brackets represent sample sizes, and only include
females aged 3+ years. Blue arrows represent social behaviour; red arrows represent spatial
behaviours. See methods for the fixed and random effects. The text box displays the
definitions for the different spatial effects.



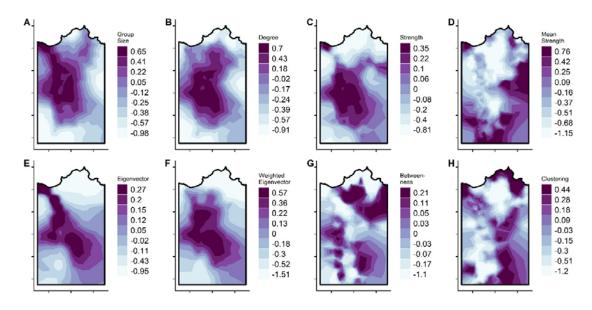
460 461	Figure 2: Spatial structuring of the 2016 social association network as a representative example. A: the spatial locations (centroids) of individual deer, connected by their social
401	
462	associations. Line opacity and width are weighted by connection strength. Ten axis units =
463	1KM. B: the same social network with the nodes positioned in a network spring-layout
464	(Csardi & Nepusz 2006) and then expanded into an even, circular grid according to their
465	nearest spatial positions in A. The points' (i.e. nodes') sizes and colours show individuals'
466	strength centrality (large and red=high strength; small and blue=low strength). Thickness of
467	the lines (i.e. edges) connecting them shows dyadic association strength between
468	individuals.
469	



470

471 Figure 3: Model outputs demonstrating strong effects of spatial and non-spatial drivers on 472 social network positions. A: DIC changes associated with addition of different spatial 473 components, for all eight social network centrality measures. Variables are arranged in order 474 of mean contribution to model fit, which varied little among response variables. Different 475 colours correspond to different network centrality response variables, with the same colour 476 key as panel C. GRM = Genomic Relatedness Matrix. HRA = Home Range Area. HRO = 477 home range overlap. The SPDE models are differentiated into those using annual centroids 478 ("SPDE") and the version with spatiotemporally varying annual spatial fields ("tSPDE"). B: 479 Variance accounted for by each variable for all eight network position measures, expressed 480 as contribution to R² in the annual model (squared correlation between observed and 481 predicted values). Different shades correspond to different variables. fYear = year as a 482 categorical random effect. HRA = Home Range Area. GRM = Genomic Relatedness Matrix. 483 HRO = home range overlap. Name = individual identity. NObs = number of observations 484 (i.e., sampling bias). PopN = population size. Status = reproductive status. SPDE = point 485 location effects estimated using the Stochastic Partial Differentiation Equation effect in the

486 INLA models. For all response variables, individual level effects (Age, Reproductive Status, 487 Name) had a negligible effect. C: Fixed effect estimates for the models. Fixed effects are 488 grouped into individual factors (age and three reproductive status effects), annual factors 489 (continuous time in years since study began, and annual population size), and sampling 490 factors (observation number). Reproductive status effects are separated into four levels: did 491 not reproduce (the intercept); calf died in the first few months of life ("Summer Yeld"); calf 492 died during the winter ("Winter Yeld"); and calf survived to May the following year ("Milk"). 493 Different colours correspond to different network centrality response variables. Points 494 represent the posterior mean; error bars denote the 95% credibility intervals for the effects. 495 Asterisks denote significant variables (i.e., those whose estimates did not overlap with zero). 496 Significant variables are fully opaque, while non-significant ones are transparent. 497



498

Figure 4: Spatial fields for the SPDE random effect for each response variable, taken from the INLA animal models and based on annual centroid point locations. Metrics can be conceptualised as simpler "direct" metrics (top row) and more complex "indirect" metrics (bottom row). Darker colours correspond to greater values. Each axis tick corresponds to 1km; for the values associated with the Easting and Northings, see Figure 1.

504 Acknowledgements

505 We thank Scottish Natural Heritage and its predecessors for permission to work on the Isle 506 of Rum NNR. The field project has been supported by grants mainly from the UK NERC with 507 some additional funding from BBSRC, the Royal Society and ERC. We thank all who have 508 contributed to the maintenance of the project over time, especially Loeske Kruuk. We thank 509 multiple dedicated field workers who have contributed to field data collection, especially 510 Fiona Guinness who collected the first 20 years of census data. GFA was funded by NSF 511 grant number 1414296, and by a Bruce McEwen Career Development Fellowship the Animal 512 Models for the Social Dimensions of Health and Aging Research Network (NIH/NIH R24 513 AG065172). JAF was supported by a fellowship from Merton College and BBSRC

514 (BB/S009752/1) and funding from NERC (NE/S010335/1). We thank Amy Sweeny and

515 Quinn Webber for comments on the manuscript, as well as Matt Silk, Orr Spiegel, and one 516 anonymous reviewer.

517 References

- 518 Albery, G.F., Becker, D.J., Kenyon, F., Nussey, D.H. & Pemberton, J.M. (2019). The fine-
- scale landscape of immunity and parasitism in a wild ungulate population. *Integr. Comp. Biol.*, icz016, 1–11.
- Albery, G.F., Kirkpatrick, L., Firth, J.A. & Bansal, S. (2020a). Unifying spatial and social
 network analysis in disease ecology. *J. Anim. Ecol.*, 1–17.
- Albery, G.F., Newman, C., Ross, J.G.B., Macdonald, D.W., Bansal, S. & Buesching, C.D.
 (2020b). Negative density-dependent parasitism in a group-living carnivore.
- 525 Albery, G.F., Watt, K.A., Keith, R., Morris, S., Morris, A., Kenyon, F., et al. (2020c).
- 526 Reproduction has different costs for immunity and parasitism in a wild mammal. *Funct.*527 *Ecol.*, 34, 229–239.
- 528 Brent, L.J.N. (2015). Friends of friends: Are indirect connections in social networks important 529 to animal behaviour? *Anim. Behav.*, 103, 211–222.
- 530 Cairns, S.J. & Schwager, S.J. (1987). A comparison of association indices. *Anim. Behav.*,
 531 35, 1454–1469.
- 532 Calenge, C. (2011). *Home range estimation in R: the adehabitatHR package*. Available at:
 533 https://cran.r-project.org/web/packages/adehabitatHR/index.html. Last accessed 10
 534 March 2020.
- 535 Castles, M., Heinsohn, R., Marshall, H.H., Lee, A.E.G., Cowlishaw, G. & Carter, A.J. (2014).
 536 Social networks created with different techniques are not comparable. *Anim. Behav.*,
 537 96, 59–67.
- 538 Centola, D. (2018). *How Behavior Spreads: The Science of Complex Contagions*. Princeton
 539 Analytical Sociology Series.
- 540 Clutton-Brock, T.H., Albon, S.D. & Guinness, F.E. (1989). Fitness costs of gestation and
 541 lactation in wild mammals. *Nature*, 337, 260–262.
- 542 Clutton-Brock, T.H., Guinness, F.E. & Albon, S.D. (1982). *Red Deer: Behavior and Ecology*543 *of Two Sexes*. University of Chicago Press, Chicago, IL.
- 544 Cote, I.M. & Poulin, R. (1995). Parasitism and group size in social animals: A meta-analysis.
 545 *Behav. Ecol.*, 6, 159–165.

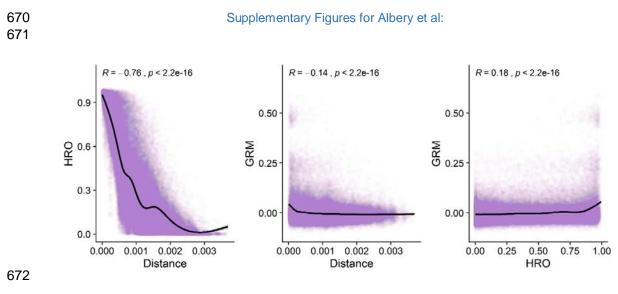
- 546 Croft, D.P., James, R. & Krause, J. (2008). *Exploring animal social networks*. Princeton
 547 University Press.
- 548 Csardi, G. & Nepusz, T. (2006). The igraph software package for complex network research.
 549 *InterJournal Complex Syst.*, Complex Sy, 1695.
- 550 Daraganova, G., Pattison, P., Koskinen, J., Mitchell, B., Bill, A., Watts, M., et al. (2012).
- 551 Networks and geography: Modelling community network structures as the outcome of 552 both spatial and network processes. *Soc. Networks*, 34, 6–17.
- 553 Davis, S., Abbasi, B., Shah, S., Telfer, S. & Begon, M. (2015). Spatial analyses of wildlife 554 contact networks. *J. R. Soc. Interface*, 12.
- Farine, D.R. (2017). A guide to null models for animal social network analysis. *Methods Ecol. Evol.*, 8, 1309–1320.
- Farine, D.R. & Sheldon, B.C. (2019). Stable multi-level social structure is maintained by
 habitat geometry in a wild bird population. *bioRxiv*, 1–30.
- Finn, K.R., Silk, M.J., Porter, M.A. & Pinter-Wollman, N. (2019). The use of multilayer
 network analysis across social scales in animal behaviour. *Anim. Behav.*, 149, 7–22.
- 561 Firth, J.A. (2020). Considering complexity: animal social networks and behavioural 562 contagions. *Trends Ecol. Evol.*, 35, 100–104.
- 563 Firth, J.A., Cole, E.F., Ioannou, C.C., Quinn, J.L., Aplin, L.M., Culina, A., et al. (2018).
- 564 Personality shapes pair bonding in a wild bird social system. *Nat. Ecol. Evol.*, 2, 1696–
 565 1699.
- Firth, J.A. & Sheldon, B.C. (2016). Social carry-over effects underpin trans-seasonally linked
 structure in a wild bird population. *Ecol. Lett.*, 19, 1324–1332.
- Firth, J.A., Voelkl, B., Crates, R.A., Aplin, L.M., Biro, D., Croft, D.P., *et al.* (2017). Wild birds
 respond to flockmate loss by increasing their social network associations to others. *Proc. R. Soc. B Biol. Sci.*, 284.
- 571 Franks, D.W., Ruxton, G.D. & James, R. (2010). Sampling animal association networks with 572 the gambit of the group. *Behav. Ecol. Sociobiol.*, 64, 493–503.
- 573 Froy, H., Börger, L., Regan, C.E., Morris, A., Morris, S., Pilkington, J.G., et al. (2018).
- 574 Declining home range area predicts reduced late-life survival in two wild ungulate 575 populations. *Ecol. Lett.*, 21, 1001–1009.
- 576 Froy, H., Walling, C.A., Pemberton, J.M., Clutton-brock, T.H. & Kruuk, L.E.B. (2016).

- 577 Relative costs of offspring sex and offspring survival in a polygynous mammal. *Biol.*578 *Lett.*, 12, 20160417.
- 579 Guilbeault, D., Becker, J. & Centola, D. (2018). Complex Contagions: A Decade in Review.
 580 pp. 3–25.
- He, P., Maldonado-chaparro, A.A. & Farine, D.R. (2019). The role of habitat configuration in
 shaping social structure: a gap in studies of animal social complexity. *Behav. Ecol. Sociobiol.*, 73.
- Holand, A.M., Steinsland, I., Martino, S. & Jensen, H. (2013). Animal Models and Integrated
 Nested Laplace Approximations. *Genes/Genomes/Genetics*, 3, 1241–1251.
- 586 Huisman, J., Kruuk, L.E.B., Ellis, P.A., Clutton-Brock, T.H. & Pemberton, J.M. (2016).
- Inbreeding depression across the lifespan in a wild mammal population. *Proc. Natl. Acad. Sci.*, 113, 201518046.
- Jacoby, D.M.P. & Freeman, R. (2016). Emerging Network-Based Tools in Movement
 Ecology. *Trends Ecol. Evol.*, 31, 301–314.
- Krause, J., James, R., Franks, D.W. & Croft, D.P. (2015). *Animal social networks*. Oxford
 University Press, Oxford, UK.
- Kruuk, L.E.B. (2004). Estimating genetic parameters in natural populations using the "animal
 model." *Philos. Trans. R. Soc. B Biol. Sci.*, 359, 873–890.
- Lindgren, F. & Rue, H. (2015). Bayesian Spatial Modelling with R-INLA. J. Stat. Softw., 63,
 1–25.
- Lindgren, F., Rue, H. & Lindstrom, J. (2011). An explicit link between Gaussian fields and
 Gaussian Markov random fields: the stochastic partial differential equation approach. *J. R. Stat. Soc. B*, 73, 423–498.
- Mourier, J., Lédée, E.J.I. & Jacoby, D.M.P. (2019). A multilayer perspective for inferring
 spatial and social functioning in animal movement networks. *bioRxiv*.
- Ostfeld, R.S., Lidicker, W.Z., Heske, E.J., Ostfeld, R.S., Lidicker, W.Z. & Heske, E.J. (1986).
 The Relationship between Habitat Heterogeneity, Space Use, and Demography in a
 Population of California Voles. *J. Mammal.*, 45, 433–442.
- Pasquaretta, C., Dubois, T., Gomez-Moracho, T., Perilhon Delepoulle, V., Le Loc'h, G.,
 Heeb, P., *et al.* (2020). Analysis of temporal patterns in animal movement networks. *Methods Ecol. Evol.*, 0–1.

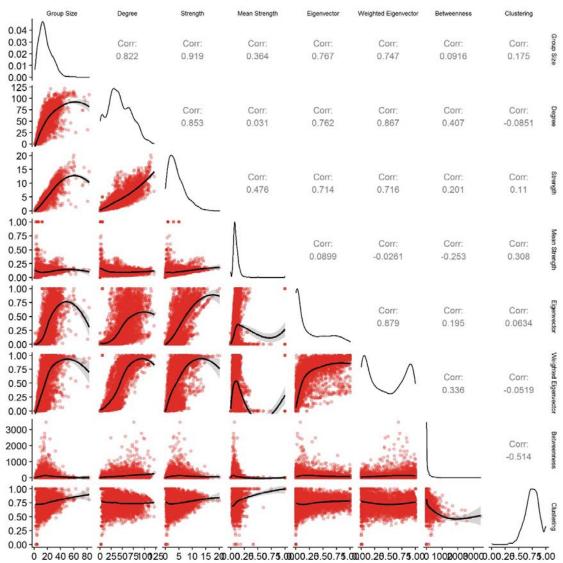
- Rand, D.G., Arbesman, S. & Christakis, N.A. (2011). Dynamic social networks promote
- cooperation in experiments with humans. *Proc. Natl. Acad. Sci. U. S. A.*, 108, 19193–
 19198.
- 611 Regan, C.E., Pilkington, J.G., Berenos, C., Pemberton, J.M., Smiseth, P.T. & Wilson, A.J.
- 612 (2016). Accounting for female space sharing in St. Kilda Soay sheep (Ovis aries)
- 613 results in little change in heritability estimates. *J. Evol. Biol.*, 30, 96–111.
- Robert, K., Garant, D. & Pelletier, F. (2012). Keep in touch: Does spatial overlap correlate
- 615 with contact rate frequency? J. Wildl. Manage., 76, 1670–1675.
- Sah, P., Mann, J. & Bansal, S. (2018). Disease implications of animal social network
 structure: A synthesis across social systems. *J. Anim. Ecol.*, 87, 546–558.
- Sanchez, J.N. & Hudgens, B.R. (2015). Interactions between density, home range
 behaviors, and contact rates in the Channel Island fox (*Urocyon littoralis*). *Ecol. Evol.*,
 5, 2466–2477.
- Shizuka, D. & Johnson, A.E. (2019). How demographic processes shape animal social
 networks. *Behav. Ecol.*, 1–11.
- Silk, M.J., Finn, K.R., Porter, M.A. & Pinter-Wollman, N. (2018). Can Multilayer Networks
 Advance Animal Behavior Research? *Trends Ecol. Evol.*, 33, 376–378.
- 625 Silk, M.J., Hodgson, D., Rozins, C., Croft, D., Delahay, R., Boots, M., et al. (2019).
- Integrating social behaviour, demography and disease dynamics in network models:
 applications to disease management in declining wildlife populations. *Philos. Trans. R. Soc. B*, 374, 20180211.
- Sosa, S., Sueur, C. & Puga-Gonzalez, I. (2020). Network measures in animal social network
 analysis: their strengths, limits, interpretations and uses. *Methods Ecol. Evol.*
- Spiegel, O., Leu, S.T., Sih, A. & Bull, C.M. (2016). Socially interacting or indifferent
 neighbours

 Randomization of movement paths to tease apart social preference and
 spatial constraints. *Methods Ecol. Evol.*, 971–979.
- Spiegel, O., Sih, A., Leu, S.T. & Bull, C.M. (2018). Where should we meet? Mapping social
 network interactions of sleepy lizards shows sex-dependent social network structure. *Anim. Behav.*, 136, 207–215.
- Stopher, K. V., Nussey, D.H., Guinness, F., Morris, A., Pemberton, J.M., Clutton-Brock, T.H., *et al.* (2012a). Re-mating across years and intralineage polygyny are associated with
 greater than expected levels of inbreeding in wild red deer. *J. Evol. Biol.*, 25, 2457–

- 641 Stopher, K. V, Walling, C. a, Morris, A., Guinness, F.E., Clutton-brock, T.H., Pemberton,
- 542 J.M., et al. (2012b). Shared spatial effects on quantitative genetic parameters:
- accounting for spatial autocorrelation and home range overlap reduces estimates of
 heritability in wild red deer. *Evolution*, 66, 2411–26.
- Thomson, C.E., Winney, I.S., Salles, O.C. & Pujol, B. (2018). A guide to using a MultipleMatrix animal model to disentangle genetic and nongenetic causes of phenotypic
 variance. *PLoS One*, 13, e0197720.
- VanderWaal, K.L., Atwill, E.R., Isbell, L.A. & McCowan, B. (2014). Linking social and
 pathogen transmission networks using microbial genetics in giraffe (Giraffa
 camelopardalis). *J. Anim. Ecol.*, 83, 406–414.
- Vander Wal, E., Laforge, M.P. & McLoughlin, P.D. (2014). Density dependence in social
 behaviour: Home range overlap and density interacts to affect conspecific encounter
 rates in a gregarious ungulate. *Behav. Ecol. Sociobiol.*, 68, 383–390.
- Webber, Q.M.R. & Vander Wal, E. (2018). An evolutionary framework outlining the
 integration of individual social and spatial ecology. *J. Anim. Ecol.*, 87, 113–127.
- Whitehead, H. (2008). Analyzing animal societies : quantitative methods for vertebrate
 social analysis. University of Chicago Press.
- Yang, J., Lee, S.H., Goddard, M.E. & Visscher, P.M. (2011). GCTA: A Tool for Genome-wide
 Complex Trait Analysis. *Am. J. Hum. Genet.*, 88, 76.
- Zuur, A.F., leno, E.N. & Saveliev, A.A. (2017). Beginner's guide to spatial, temporal, and
 spatial-temporal ecological data analysis with R-INLA. Highstat Ltd.
- 662
- 663
- 664
- 665
- 666
- 667
- 668
- 669

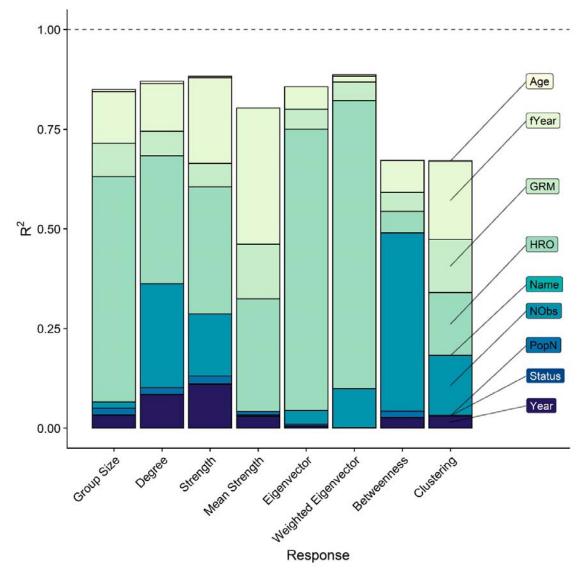


Supplementary Figure 1: Correlation among pairwise values in lifetime point location distances, home
 range overlap matrix, and genomic relatedness matrix. The data have been fitted with a Generalised
 Additive Model (GAM) smooth.



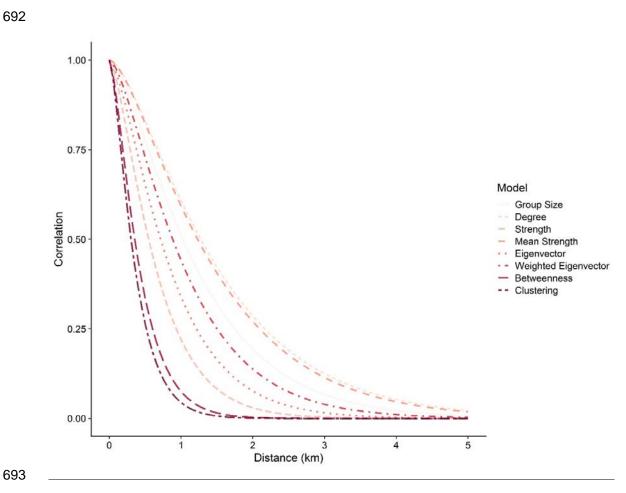
678

679 Supplementary Figure 2: Pairwise correlations among network position response variables. Values 680 were transformed, scaled to have a mean of 0 and a standard deviation of 1, with outliers removed, 681 before analysis. The data have been fitted with a Generalised Additive Model (GAM) smooth.

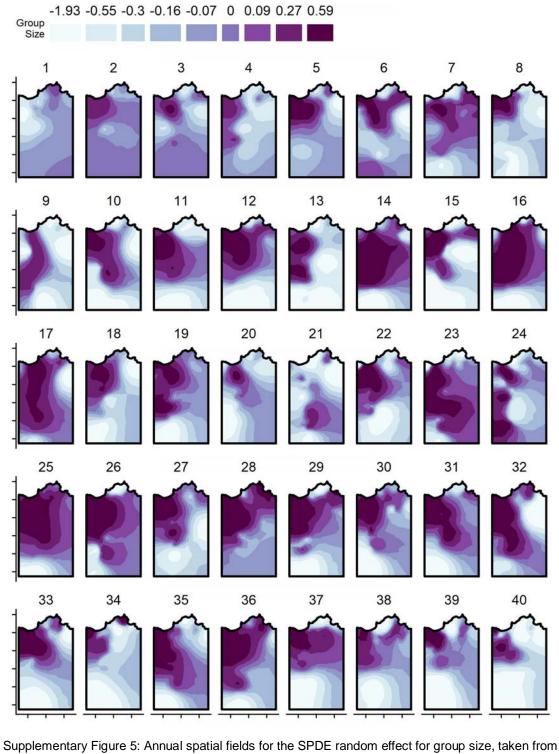


682

683 Supplementary Figure 3: Variance accounted for by each variable for all eight network position 684 measures, expressed as contribution to R² in the models with home range overlap. This output differs 685 from Figure 3B in the main text because the model did not include the SPDE random effect. Different 686 shades correspond to different variables. fYear = year as a categorical random effect. GRM = 687 Genomic Relatedness Matrix. HRO = home range overlap. Name = individual identity. NObs = 688 number of observations (i.e., sampling bias). PopN = population size. Status = reproductive status. 689 For all response variables, individual level variables (Age, Reproductive Status, Identity) had a 690 negligible effect.

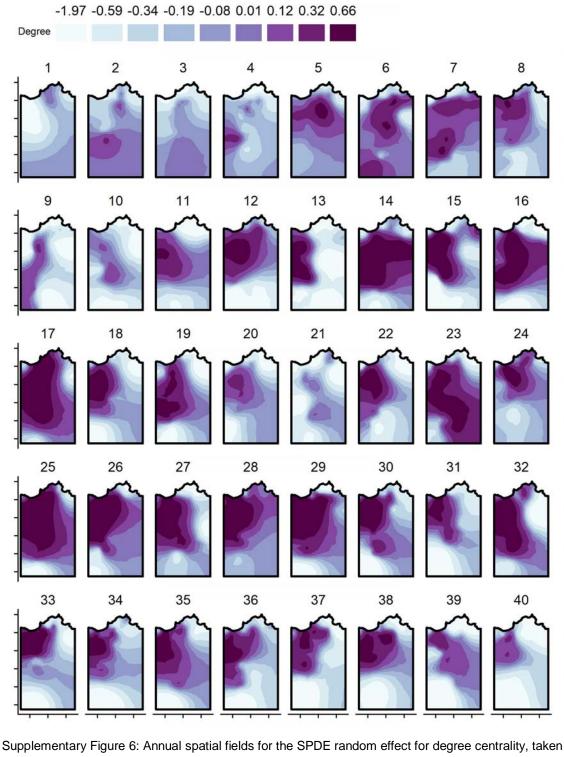


694 Supplementary Figure 4: The INLA SPDE autocorrelation ranges for each response variable. Different 695 shades and line types correspond to different response variables. The X axis is in kilometres.

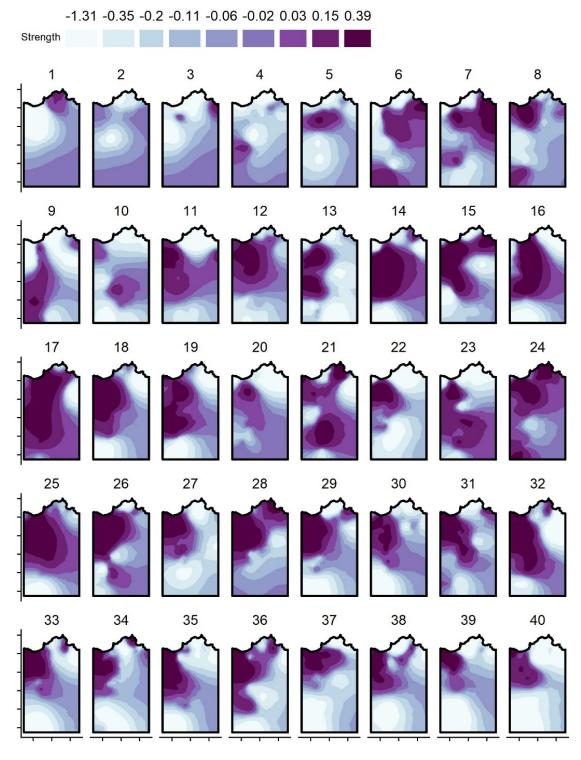


697

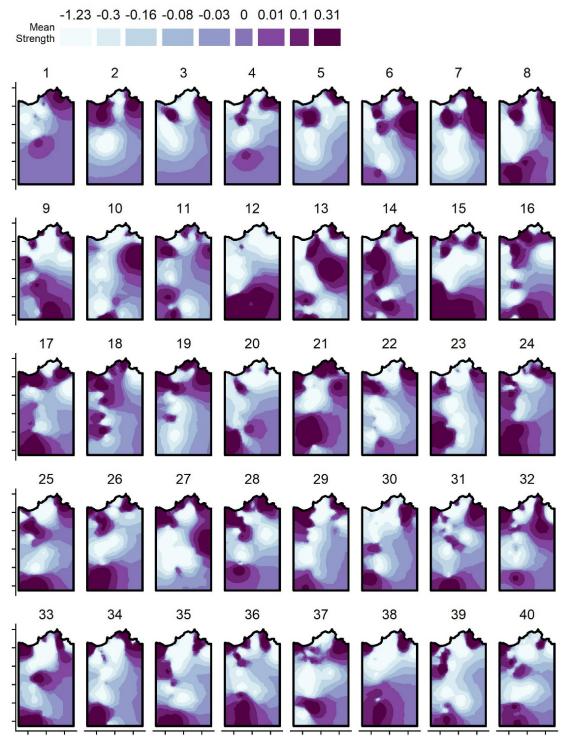
Supplementary Figure 5: Annual spatial fields for the SPDE random effect for group size, taken from
 the INLA animal models and based on annual centroid point locations. Darker colours correspond to
 greater values. Each axis tick corresponds to 1km distance; for the values associated with the Easting
 and Northings, see Figure 1 in the main text.



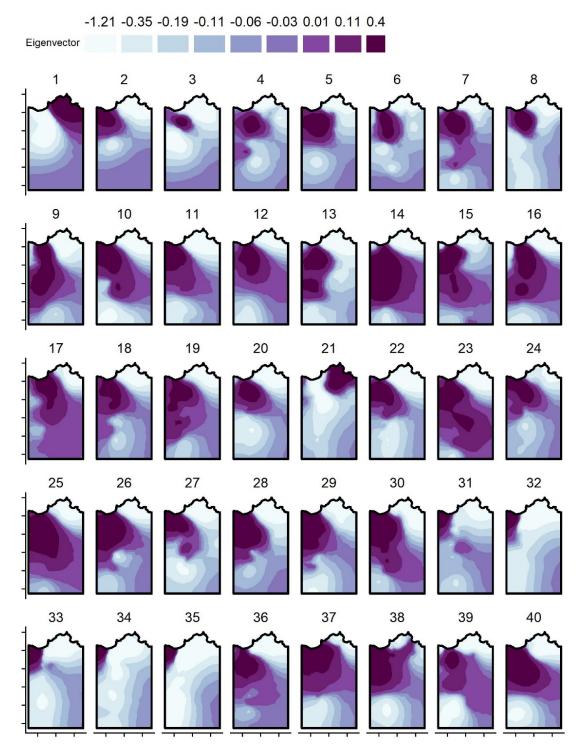
Supplementary Figure 6: Annual spatial fields for the SPDE random effect for degree centrality, taken
 from the INLA animal models and based on annual centroid point locations. Darker colours
 correspond to greater values. Each axis tick corresponds to 1km distance; for the values associated
 with the Easting and Northings, see Figure 1 in the main text.



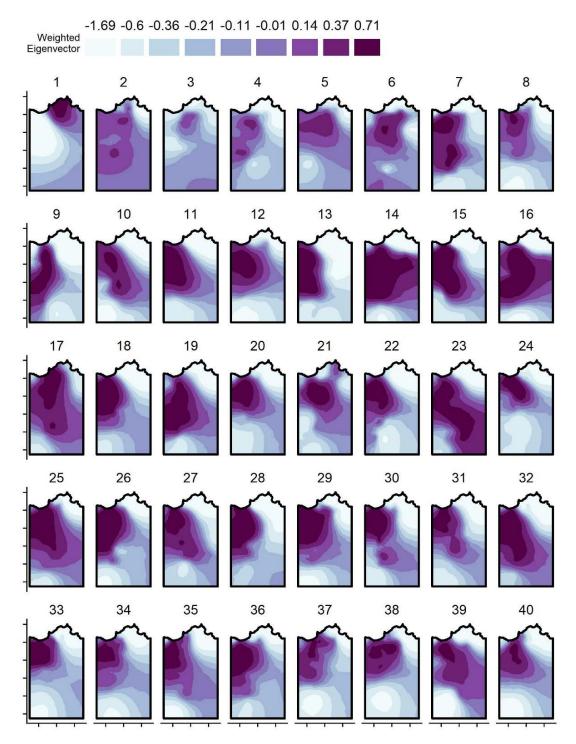
Supplementary Figure 7: Annual spatial fields for the SPDE random effect for strength centrality,
taken from the INLA animal models and based on annual centroid point locations. Darker colours
correspond to greater values. Each axis tick corresponds to 1km distance; for the values associated
with the Easting and Northings, see Figure 1 in the main text.



Supplementary Figure 8: Annual spatial fields for the SPDE random effect for mean strength
centrality, taken from the INLA animal models and based on annual centroid point locations. Darker
colours correspond to greater values. Each axis tick corresponds to 1km distance; for the values
associated with the Easting and Northings, see Figure 1 in the main text.

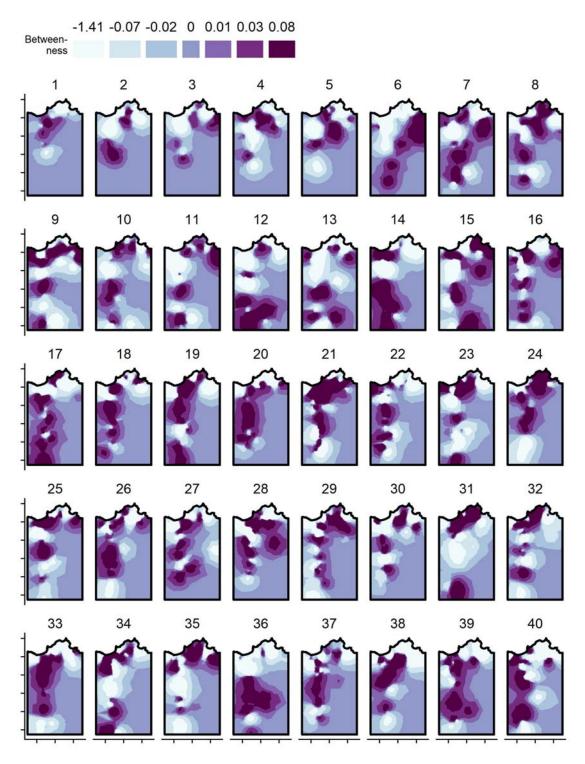


Supplementary Figure 9: Annual spatial fields for the SPDE random effect for Eigenvector centrality,
 taken from the INLA animal models and based on annual centroid point locations. Darker colours
 correspond to greater values. Each axis tick corresponds to 1km distance; for the values associated
 with the Easting and Northings, see Figure 1 in the main text.

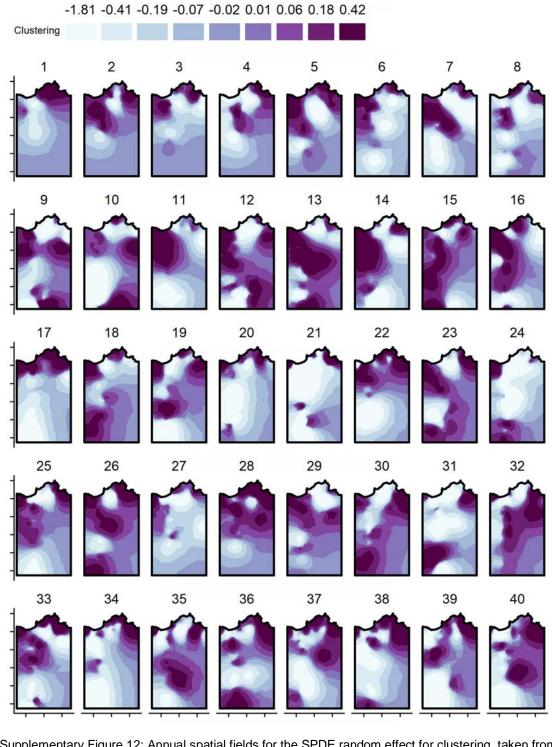


723

Supplementary Figure 10: Annual spatial fields for the SPDE random effect for weighted Eigenvector
 centrality, taken from the INLA animal models and based on annual centroid point locations. Darker
 colours correspond to greater values. Each axis tick corresponds to 1km distance; for the values
 associated with the Easting and Northings, see Figure 1 in the main text.



Supplementary Figure 11: Annual spatial fields for the SPDE random effect for betweenness centrality, taken from the INLA animal models and based on annual centroid point locations. Darker colours correspond to greater values. Each axis tick corresponds to 1km distance; for the values associated with the Easting and Northings, see Figure 1 in the main text.



733

Supplementary Figure 12: Annual spatial fields for the SPDE random effect for clustering, taken from
the INLA animal models and based on annual centroid point locations. Darker colours correspond to
greater values. Each axis tick corresponds to 1km distance; for the values associated with the Easting
and Northings, see Figure 1 in the main text.