Coronavirus testing indicates transmission risk increases along

wildlife supply chains for human consumption in Viet Nam,

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Abstract

Outbreaks of emerging coronaviruses in the past two decades and the current pandemic of a novel coronavirus (SARS-CoV-2) that emerged in China highlight the importance of this viral family as a zoonotic public health threat. To gain a better understanding of coronavirus presence and diversity in wildlife at wildlife-human interfaces in three southern provinces in Viet Nam 2013-2014, we used consensus Polymerase Chain Reactions to detect coronavirus sequences. In comparison to previous studies, we observed high proportions of positive samples among field rats (34.0%, 239/702) destined for human consumption and insectivorous bats in guano farms (74.8%, 234/313) adjacent to human dwellings. Most notably among field rats, the odds of coronavirus RNA detection significantly increased along the supply chain from field rats sold by traders (reference group; 20.7% positivity, 39/188) by a factor of 2.2 for field rats sold in large markets (32.0%, 116/363) and 10.0 for field rats sold and served in restaurants (55.6%, 84/151). Coronaviruses were detected in the majority of wildlife farms (60.7%, 17/28) and in the Malayan porcupines (6.0%, 20/331) and bamboo rats (6.3%, 6/96) that are farmed. We identified six known coronaviruses in bats and rodents, clustered in three *Coronaviridae* genera, including the *Alpha-, Beta-*, and *Gammacoronaviruses*. Our analysis also suggested either mixing of animal excreta in the environment or interspecies transmission of coronaviruses, as both bat and avian coronaviruses were detected in rodent feces in the trade. The mixing of multiple coronaviruses, and their apparent amplification along the wildlife supply chain into restaurants, suggests maximal risk for end consumers and likely underpins the mechanisms of zoonotic spillover to people.

74 **Key words:** coronavirus, One Health, rodents, bats, wildlife trade, wildlife farm, Viet Nam,

75 surveillance, guano farming, amplification

Introduction

Human-wildlife contact with a bat or an intermediate host species in China likely

triggered a coronavirus spillover event that may have involved wildlife markets and led to the

pandemic spread of SARS-CoV-2 [1,2]. The pandemic risk of commercial trade in live wildlife

was first recognized during the 2002-2003 Severe Acute Respiratory Syndrome (SARS)

outbreak due to SARS-CoV [3]. This virus spread to more than 29 countries in Asia, Europe, and

the Americas with 8,096 people infected and 774 deaths, costing the global economy about $US

40 billion in response and control measures [4,5]. Unfortunately, the global impact of COVID-

19, the disease caused by SARS-CoV-2 has reached nearly every country and greatly surpassed

those numbers by many orders of magnitude [6]. While bats are thought to be the ancestral hosts

for all groups of coronaviruses, including those that were previously thought to be in the rodent

and avian clades [7], for both SARS-CoV and SARS-CoV-2 wildlife trade supply chains are

suspected to have contributed the additional conditions necessary for the emergence, spillover,

and amplification of these viruses in humans [8,9]. To better understand the presence and

diversity of coronaviruses in wildlife we conducted coronavirus surveillance at high-risk

interfaces in Viet Nam from 2009 to 2014 [10]. We sampled in live field rat trade (*Rattus* sp. and

Bandicota sp.) and wildlife farm interfaces to assess risk from different wildlife supply chains

destined for human consumption, and sampled bat guano farms to assess the potential

occupational risk of this practice given that bat guano farm artificial roost structures are often

erected near human dwellings.

In the early 2000s, the Vietnamese field rat trade was estimated to process 3,300-3,600

tons of live rats annually for consumption, a market valued at US$2 million [11]. Although rats

are still commonly traded in wet markets and sold live for food consumption along the Mekong

Delta in southern Viet Nam, no recent published data on the scale and scope of the trade is

available [12]. This human-wildlife interface involves the capture of wild caught field rats,

subsequent trade, and consumption along a supply chain involving the entire Mekong Delta

region, particularly Cambodia and Viet Nam [13]. Driving this trade are consumers in Viet Nam

and Cambodia, who report eating rats at least once per week because of their good flavor, low

cost, and perception of rats as 'healthy, nutritious, natural, or disease free' [13]. Rodent parts

(heads, tails, and internal organs discarded at slaughter) are also often fed to domestic livestock

or herptiles raised in captivity including frogs, snakes, and crocodiles [12]. Records of this local

trade in field rats include official rat hunts, instituted by French administrators, that killed

upwards of 10,000 rats a day prior to the arrival of bubonic plague in Ha Noi in 1903 [14].

Over the past three decades, commercial wildlife farming has developed in many

countries in Southeast Asia, including Viet Nam. Although there are historic references to the

occurrence of wildlife farms in Viet Nam dating back to the late 1800s, the rapid expansion in

terms of farm numbers, species diversity, and scale of operations has occurred in recent decades

in response to growing domestic and international demand for wildlife [15]. A 2014 survey

across 12 provinces in southern Viet Nam identified 6,006 registered wildlife farms of which

4,099 had active operations. The surveyed farms were stocked with approximately one million

wild animals including, rodents, primates, civets, wild boar, Oriental rat-snakes, deer, crocodiles,

and softshell turtles. Ninety-five percent of the farms held 1-2 species of wildlife, and 70% of the

farms also raised domestic animals on the same premises [16]. A key component of the wildlife

farm industry in Viet Nam is the raising of wild species for meat for human consumption [16].

These farms sell to urban wild meat restaurants serving increasingly affluent populations

throughout the country and also supply international markets with wild meat [17]. Commercial

wildlife farming in Viet Nam is part of the expanded international trade of wildlife that has been

hypothesized to contribute to the cause of global epidemics, such as SARS [18] and now

COVID-19.

Emerging evidence suggests zoonotic virus spillover risk is a concern at bat-human

interfaces in Asia. Guano harvested from a cave in Thailand were positive for a group C

betacoronavirus, which includes MERS-CoV, and 2.7% of 218 people living in close proximity

to bats known to carry viruses related to SARS-CoV tested positive for SARS-related antibodies

in China [19,20]. The traditional practice of guano farming in parts of Cambodia and Viet Nam

involves the construction of artificial bat roosts in gardens or backyard farms, under which

domestic animals and crops are raised, and children often play [21,22]. Cambodian development

programs promoted the practice in 2004 to enhance soil fertility, reduce reliance on chemical

fertilizers, generate income ($US 0.50/kg), control insect pests, and protect the lesser Asiatic

yellow bats (*Scotophilus kuhlii*) that were being hunted [21–23]. No personal protection

measures are taken when harvesting the guano, which is used as fertilizer and is reported to

improve the growth rate in five economically important plant species [24].

In this study we investigated the presence and diversity of coronavirus sequences in the

field rat trade distribution chain, wildlife farms specializing in rodents for human consumption,

and bat guano "farms" and roosts near human dwellings to better understand the natural hosts of

coronaviruses and the risk for these interfaces to facilitate spillover into humans.

Materials and Methods

Sampling Locations

145  Sampling was performed at multiple sites representing several high-risk interfaces for

contacts among people, rodents, and bats. Rodent sampling focused on the live rodent trade

supply chain and rodent farms. Along the supply chain, we targeted eight sites involved in the

private sale and processing of live rodents for consumption, defined as 'traders' for the purpose

of this study in Dong Thap and Soc Trang provinces, 14 large markets sites in Dong Thap and

Soc Trang provinces (>20 vendors), and two restaurant sites in Soc Trang province (Fig 1). The

28 rodent farm sites we targeted in Dong Nai province produced Malayan porcupines (*Hystrix*

brachyura) and bamboo rats (*Rhizomys* sp.) for human consumption (Fig 2). Other species

observed or raised at the wildlife farm sites included dogs, cattle, pigs, chickens, ducks, pigeons,

geese, common pheasant, monitor lizards, wild boar, fish, python, crocodiles, deer, civets, non-

human primates as pets or part of private collections, free-flying wild birds, and free-ranging

peri-domestic rodents.

Fig 1. Slaughtering rodents (left) and rodent market (right) in Dong Thap province,
October 2013.

Fig 2. Malayan porcupines (*Hystrix brachyura*) farm in Dong Nai province, November
2013.

Bat sampling occurred at bat guano "farms" and a natural bat roost located at a religious

site. Bat guano farms consisted of artificial roosts constructed with a concrete base and pillars

topped with fronds of coconut palm or Asian Palmyra Palm (*Borassus flabellifer*) (Fig 3).

Seventeen bat guano farms were sampled in the two provinces of Dong Thap and Soc Trang. The

natural bat roost was located at a religious site in Soc Trang province known as the "bat pagoda",

where *Pteropus* sp. have historically roosted in trees protected from hunting, and light and noise

pollution [25].

Fig 3. Bat guano farms in Soc Trang Province, October 2013.

All study sampling occurred from January 2013 to March 2014 at 41 sites in the wet

(south Viet Nam: May 1st - November 30th) and 30 in the dry (south Viet Nam: December 1st -

April 30th) seasons. Given the distances between sites, all sites were sampled once except the bat

pagoda natural roost in Soc Trang province, which was visited three times and sampled in both

seasons.

Animal sampling

Samples were humanely collected using standard and previously published protocols

[26]. Feces, swabs of the pen floors, and urine/urogenital swabs were collected from rodents at

wildlife farms. Samples were classified as 'fecal sample' when collected from animals housed

individually, and as 'environmental sample' when collected below cages housing multiple

individuals. Samples from rodents in the trade included primarily oral swabs in addition to

tissues (i.e. brain, kidney, lung, and small intestine), rectal swabs, and urine/urogenital swabs.

These samples were collected from individual carcasses after the rodents were slaughtered by a

market vendor, trader, or restaurant kitchen staff. However, the rodents were usually butchered at

a common site for each observed time period that was only cleaned intermittently following the

trader's, vendor's, or restaurant's regular practices. Oral swabs were taken from the severed

heads, and additional tissue samples were collected from the internal organs and the

gastrointestinal tracts which were removed during the butchering process.

Fecal samples and a small number of urine samples from bats in guano farms and the

natural roost site were collected on clean plastic cover sheets within 1-2 hours after placement

under bat roosts, and thus each sample may represent one or multiple bats. Oral and rectal swabs

were also collected from live-captured bats at the natural pagoda roost site.

Animals were identified in the field to the lowest taxonomic level possible based on

morphological characteristics, and species was identified in a subset of animals through genetic

barcoding [15]. Due to difficulty of morphologic identification in the field, unless barcoded,

rodents (*Rattus argentiventer*, *R. tanezumi*, *R. norvegicus*, *R. exulans*, *R. losea,* and *Bandicota*

indica; [12,27]) were categorized as "field rats". Bats were classified as "*Microchiroptera*"

following the traditional taxonomic classification (new classification of two new suborders

Yangochiroptera and *Yinpterochiroptera*, was only published near the end of the study, so for

consistency we used the historical classification [28]).

All samples were collected in cryotubes containing RNAlater (RNA stabilization reagent,

Qiagen), and stored in liquid nitrogen in the field before being transported to the laboratory for

storage at -80 ˚C. Samples were tested by the Regional Animal Health Office No. 6 (RAHO6)

laboratory in Ho Chi Minh City. The study was approved by the Department of Animal Health of

the Ministry of Agriculture and Rural Development and protocols were reviewed by the

Institutional Animal Care and Use Committee at the University of California at Davis (protocol

number 16048).

Sample Testing

RNA was extracted (RNA MiniPrep Kit, Sigma-Aldrich) and cDNA transcribed

(SuperScript III First Strand cDNA Synthesis System, Invitrogen). Coronavirus RNA was

detected using two broadly reactive consensus nested-PCR assays targeting the *RNA dependent*

RNA polymerase (RdRp) gene [29,30]. The positive control was a synthetic plasmid containing

the primer-binding sites for both assays. Distilled water was used as a negative control and

included in each test batch. PCR products were visualized using 1.5% agarose gels, and bands of

the correct size were excised, cloned, and sequenced by Sanger dideoxy sequencing using the

same primers as for amplification.

Phylogenetic analysis

For sequence analysis and classification operating taxonomic units were defined with a

cut off of 90% identity, i.e. virus sequences that shared less than 90% identity to a known

sequence were labelled sequentially as PREDICT_CoV-1, -2, -3, etc. and groups sharing $\geq$ 90%

identity to a sequence already in GenBank were given the same name as the matching sequence

[7]. A phylogenetic tree was constructed for sequences amplified using the Watanabe protocol,

as this PCR protocol yielded longer sequences and more positive results than the Quan protocol.

Several representative sequences for each viral species found in our study were included for

analysis and are available in GenBank (Table S3). Alignments were performed using MUSCLE,

and trees were constructed using Maximum likelihood and the Tamura 3-parameter model in

MEGA7 [31]. The best-fit model of DNA substitution was selected in MEGA7 using BIC scores

(Bayesian Information Criterion) and Maximum Likelihood values (*lnL*). Bootstrap values were

calculated after 1000 replicates. In addition, a median-joining network was constructed using

Network 5.0.0.3 [32] to explore phylogenetic relationships among bat coronavirus 512/2005

sequences at the intraspecies level, as haplotype networks may better represent the relationships

among viral sequences with low sequence diversity compared with phylogenetic trees [33].

Statistical analyses

Visualization of sampling locations in provinces in Viet Nam, along with the distribution

by species and interface was constructed with the ggmap, ggplot2, and sp packages [34]. All

analyses were done using R version 3.5.0 or higher (R Development Core Team, Vienna,

Austria). Data (S1 Data) and code (S1 R Code) are available in the supplementary materials. The

effect of risk factors (season, sub-interface type) was examined and limited to interfaces for

which the distribution of samples across factors could support the analysis. These included

season for *Pteropus* bat samples collected in the bat pagoda natural roost and the effect of season

and sub-interface for samples collected in the rodent trade in southern Viet Nam. Given the low

sample size, the effect of season for *Pteropus* bats samples positive for coronaviruses was

assessed using a Fisher exact test. The effect of season (dry, wet, with dry season as reference

category) and sub-interface type (trader, large markets, restaurants, with trader as reference

category) in traded rodent samples positive for coronaviruses was assessed with a mixed effect

multivariable logistic regression, with sites as random effect (i.e. grouping variable) using the

lme4 R package [35]. A p-value of less than 0.05 was considered statistically significant. The

95% binomial confidence intervals for proportions were calculated using binom.test in R.

The comparison of the proportion of coronavirus positives in different sample types was

performed on positive individuals sampled in the rodent trade with multiple sample types

collected per individual. We then calculated the proportion of individuals positive for each

sample type, as a proxy for the probability of detection by each sample type.

Results

Detection of coronavirus by animal taxa and interface

A total of 2,164 samples collected between January 2013 and March 2014 from rodents

and bats were tested for coronaviruses (Table 1, S1 Table). Assuming that non-invasive samples

from bats and farmed rodents represented unique distinct individuals, these samples came from

1,506 individuals, including 1,131 rodents and 375 bats from 70 sites sampled in Dong Thap,

Soc Trang, and Dong Nai provinces in the southern region near the Mekong River Delta (Fig 4).

Fig 4. Map of sampling sites by province and multi-panel plots showing individual counts

of animals sampled by province, taxa, and interface. The color of each bar represents the

animal taxonomic group sampled in Dong Nai, Dong Thap, and Soc Trang provinces. *Sciuridae*

and *Rattus argentiventer* were only sampled one time apiece from wildlife farms.

Out of 70 sites, coronavirus positives were detected at 58 including 100% (24/24) of live

rodent trade sites, 60.7% (17/28) of rodent wildlife farm sites, 94.1% (16/17) of bat guano farm

sites, and at the one natural pteropid bat roost. Wildlife farms were only sampled in Dong Nai

province and the live rodent trade and bat interfaces were sampled in Dong Thap and Soc Trang

provinces (Fig 4).

Table 1: Summary of coronavirus positives by taxa and interface. Co-infection is defined as the detection of two different coronavirus taxonomic units in an individual animal.

Taxa group	Interface	Sub-interface	Taxa group	% site positive	% individual positive	Viral species	# of co-infected animals
Rodents	Rodent trade	Trader	Field rat[a]	100% (8/8)	20.7% (39/188)	Murine coronavirus (n=36), Longquan aa coronavirus (n=5)	2
		Large market	Field rat[a]	100% (14/14)	32.0% (116/363)	Murine coronavirus (n=103), Longquan aa coronavirus (n=31)	18
		Restaurant	Field rat[a]	100% (2/2)	55.6% (84/151)	Murine coronavirus (n=70), Longquan aa coronavirus (n=20)	6
	Wildlife farm		*Hystrix* sp.	47.8% (11/23)	6.0% (20/331)	Bat coronavirus 512/2005 (n=19), Infectious bronchitis virus (IBV) (n=1)	0
			Rhizomys sp.	45.5% (5/11)	6.3% (6/96)	Bat coronavirus 512/2005 (n=5), Infectious bronchitis virus (IBV) (n=1)	0
			Rattus sp.[b]	100% (1/1)	100% (1/1)	Bat coronavirus 512/2005 (n=1)	0
			Sciuridae sp.	0% (0/1)	0% (0/1)		
Bats	Human dwelling	Natural bat roost					
			Pteropus sp.	100% (1/1)	6.7% (4/60)	PREDICT_CoV-17 (n=3), PREDICT_CoV-35 (n=1)	0
			Cynopterus horsfieldii	0% (0/1)	0% (0/2)		
		Bat guano farm	*Microchiroptera*[c]	94.1% (16/17)	74.8% (234/313)	PREDICT_CoV-17 (n=1), PREDICT_CoV-35 (n=38), Bat coronavirus 512/2005 (n=216)	21[d]
				82.9% (58/70)	**33.5% (504/1506)**		**47**

[a] Field rat here refers to a mix of *Rattus* sp. and *Bandicota* sp.

[b] This environmental sample collected from a porcupine cage on a porcupine farm was barcoded as *Rattus* sp., suggesting this species
was free-ranging at the site (Fig 2). The detection of a bat virus from this sample is suggestive of either environmental mixing or viral
sharing.
[c] Suborder
[d] Co-infections included PREDICT_CoV-17 with Bat coronavirus 512/2005 (n=1) and PREDICT_CoV-35 with Bat coronavirus
512/2005 (n=20).

Coronaviruses were detected in the field rat trade (a mix of *Rattus* and *Bandicota* genera)

at all sites in Dong Thap (n=16) and Soc Trang (n=8) provinces, with 34.6% (95% CI 29.8 –

39.7%, 129/373) and 33.4% (95% CI 28.4 – 38.9%, 110/329) positives respectively. The overall

proportion of positives in field rats was 34.0% (95% CI 30.6 – 37.7%, 239/702), ranging from

3.2% to 74.4% across sites. Field rats sampled in the rodent trade had an increasing proportion of

positives along the distribution chain. Starting with traders, the proportion positive was 20.7%

(95% CI 15.3 – 27.4%, 39/188), 32.0% (95% CI 27.2 – 37.1%, 116/363) in large markets, and

55.6% (95% CI 47.3 – 63.6%, 84/151) at restaurants (Fig 5). The proportion of positives was

higher in the wet season (36.7%, 95% CI 32.8 – 40.8%, 210/572) than the dry season (22.3%,

95% CI 15.7 – 30.6%, 29/130). In a multivariate model with site as random effect, both season

and interface type were significantly associated with the risk of rodent infection, with higher risk

of infection in the wet season (OR=4.9, 95% CI 1.4 – 18.0), and increasing risk along the supply

chain from traders (baseline) to large markets (OR=2.2, 95% CI 1.05 – 4.7), to restaurants

(OR=10.0, 95% CI 2.7 – 39.5) (S2 Table). It should be noted, however, that since sites were only

visited during one season, both independent variables were defined at the site level and

confounding effects with other site-level characteristics cannot be excluded.

Fig 5. Plot of the proportion of coronavirus positives in field rats by interface. Bars show

95% confidence intervals.

Among the positive field rats with more than one sample tested (n=220), the proportion

positive by sample type was 79.9% (95% CI 73.9 – 84.9%, 175/219) in oral swabs, 52.9% (95%

CI 38.6 – 66.8%, 27/51) in lung, 51.6% (95% CI 43.5 – 59.7%, 80/155) in small intestine, 31.2%

(95% CI 12.1 – 58.5%, 5/16) in brain, 23.1% (95% CI 6.2 – 54.0%, 3/13) in kidney, 50.0% in

feces (1/2), 100% in spleen (1/1), and 0% in urine/urogenital swabs (0/1).

At the rodent farm interface, 6.0% (95% CI 3.8 – 9.3%, 20/331) of *Hystrix brachyura* and

6.3% (95% CI 2.6 – 13.6%, 6/96) of *Rhizomys* sp. were positive. The overall proportion of

positives was 6.3% (95% CI 4.3 – 9.1%, 27/429) (Table 1 and Fig 4). There was no difference

among species or season and proportion positive in rodent farms, and low sample size and

unequal sampling limited analysis.

The proportion of coronavirus positives at the two bat interfaces differed by an order of

magnitude as 74.8% (95% CI 69.5 – 79.4%) of the non-invasive samples collected from

Microchiroptera bats at bat guano farms were positive, and 6.7% (95% CI 2.2 – 17.0%) of the

Pteropus genus samples at the natural roost in Soc Trang province (Fig 4) were positive (Table

1). Pteropid bats sampled at the natural roost had higher proportions of positives in the wet

season (27.3%, 95% CI 7.3 – 60.7%, 3/11) compared with the dry season (2.0%, 95% CI 0.1 –

12.2%, 1/50; Fisher exact test p=0.02, OR=16.6 [1.2 – 956.8]), although low sample size and

single sampling per season warrants cautious interpretation.

Phylogenetic analysis

Six distinct taxonomic units of coronaviruses corresponding to bat coronavirus 512/2005,

Longquan aa coronavirus, avian infectious bronchitis virus (IBV), murine coronavirus,

PREDICT_CoV-17, and PREDICT_CoV-35 were detected. All these viruses were detected

using both the Watanabe and Quan assays, except IBV sequences that were detected only using

the Quan protocol. Of the 504 positive animals, 433 were positive by the Watanabe assay, 410

were positive by the Quan assay, and 339 were positive by both. Phylogenetic analysis showed

that among the six coronaviruses detected, PREDICT_CoV-35 and bat CoV 512/2005 clustered

within the *Alphacoronaviruses*, while PREDICT_CoV-17, Longquan aa CoV and murine CoV

clustered within the *Betacoronaviruses*. The virus identified within the *Gammacoronavirus*

genus was avian IBV.

PREDICT_CoV-17 and PREDICT_CoV-35 were first reported by Anthony et al. [17].

We found PREDICT_CoV-17 in *Pteropus* bats and in *Microchiroptera* (Table 1). The

PREDICT_CoV-17 sequences from *Pteropus* detected in this study clustered closely with

PREDICT_CoV-17 sequences from *Pteropus giganteus* bats in Nepal and *Pteropus lylei* bats in

Thailand [36] (Fig 6, S3 Table). PREDICT_CoV-35 was found in *Microchiroptera* in bat guano

farms and in a pteropid bat (Table 1). PREDICT_CoV-35 sequences from Viet Nam clustered

with other PREDICT_CoV-35 sequences found previously in samples from hunted *Scotophilus*

kuhlii bats in Cambodia (S3 Table; Dr. Lucy Keatts personal communication), and with

sequences found in bats from an earlier study in the Mekong Delta region in Viet Nam (Fig 6).

Bat coronavirus 512/2005 was detected in *Microchiroptera* bat guano; and in *H.*

brachyura (feces and environmental samples), *R. pruinosus* (feces barcoded), and *R.*

argentiventer (barcoded environmental sample) in wildlife farms (Table 1 and S1 Table). In

Microchiroptera, Bat coronavirus 512/2005 was frequently found in co-infection with

PREDICT_CoV-35 (Table 1, S1 Table). Network analysis showed the relationships among the

bat coronavirus 512/2005 sequences from the three provinces in south Viet Nam (Fig 7). We

observed two main clusters and a shallow geographic structure of genetic diversity, perhaps

illustrative of sampling effort but also of localized transmission and circulation of bat

coronavirus 512/2005 strains in these provinces. One cluster was exclusively detected in

Microchiroptera and mostly restricted to Dong Thap province and another cluster included

sequences shared among all hosts and distributed in the three provinces (Fig 7). Parts of the

network showed a star-like topology (Fig 7), typical of populations in expansion that have

recently increased size. There were three sequence types that were shared among

Microchiroptera and rodents.

358   Murine coronavirus and Longquan aa coronavirus were detected in 209 and 56 field rat

samples, respectively, and 26 were coinfected with both (Table 1). Two sequences of IBV were

detected in rodent feces collected on two wildlife farms, one in a bamboo rat and another in a

Malayan porcupine. The rodent interfaces where bat and avian coronaviruses were detected in

feces were not full containment facilities and possibly had bats and birds flying and roosting

overhead (Fig 2). The IBV positives were detected in fecal samples from wildlife farms that had

chickens, pigs, and dogs on site.

Fig 6. Phylogenetic tree of bat and rodent coronavirus sequences detected in Viet Nam. The

analysis is based on 387 bp fragment of the *RdRp* gene using maximum likelihood with the

Tamura 3-parameter model, Gamma distributed with Invariant sites (G+I), and 1000 bootstrap

replicates via MEGA7. The analysis included 17 sequences from this study (red from bat hosts,

blue from rodent hosts), six sequences (in gray) from a previous study in Viet Nam [27], and 25

reference sequences (in black) available in the GenBank database (S3 Table). The tree was

rooted by a strain of Night-heron coronavirus HKU19 (GenBank accession No. NC_016994).

Fig 7. Median-joining networks of bat coronavirus 512/2005 *RdRp* sequences color-coded

according to (A) host and (B) sampling location. Each circle represents a sequence, and circle

size is proportional to the number of animals sharing a sequence. Numbers on branches indicate

the number of mutations between sequences (if >1). Circles are colored-coded by animal host:

bat (*Microchiroptera*), rodent (*Rattus* & *Bandicota*, *Rhizomys*, and *Hystrix*) and sampling

location (Dong Thap (blue), Dong Nai (yellow) and Soc Trang (green)). Small black circles

represent median vectors (ancestral or unsampled intermediate sequence types).

Discussion

High prevalence and amplification along the supply chain for human consumption

Significant findings of this study are the high proportion of coronavirus positive animals and the increasing proportion of positives found along the rodent trade supply chain from the capture site to restaurants. The transit of multiple animal species through the supply chain offers opportunities for inter- and intra-species mixing. Overcrowding and close confinement of live animals in cages results in increased animal contact, likely leading to stress. While methodologically similar to rodent surveys in Zhejiang province, China (2%), Dong Thap province, Viet Nam (4.4%), and globally (0.32%), our overall proportion of coronavirus positives was much higher among field rats (34.5%) and somewhat higher among farmed rodents (6.3%) [7,27,37]. Stress and poor nutrition likely contributes to shedding by reducing animal condition and altering immune functions [38]. Together, these factors may result in increased shedding and amplification of coronaviruses along the supply chain for human consumption.

The amplification of coronavirus along the supply chain may be associated with season as field rats were significantly more positive in the wet season. *Rattus argventiventer* generally reproduce year-round in Viet Nam, but are particularly abundant in the wet season (May through October) following the rice harvest when an abundance of food supports the population increase [39]. If these seasonal population increases affect density dependent contact, there could be increased coronavirus prevalence and shedding in wild field rats during certain times of the year, which could then be further amplified along the trade.

Our survey was not a comprehensive multi-year evaluation of the field rat supply chain

and it was restricted to two provinces with this interface. These limitations mean we are not able

to make inferences about larger spatial patterns or the inter-annual variability of coronavirus

prevalence in wildlife populations found in this interface, which spans into neighboring

Cambodia.

However, from a mechanistic perspective as animals progress along the wildlife supply

chain, opportunity for human contact increases, including close direct contact with traders,

butchers, cooks, and consumers [40]. The combination of increased coronavirus prevalence in

traded wildlife and greater opportunity for human-wildlife contact as well as intra- and inter-

species contact in trade systems is likely to increase the risk of zoonotic transmission of

coronaviruses in wildlife markets, restaurants, and other trade interfaces.

414   Viral sharing or environmental mixing

We detected avian and bat coronaviruses in wildlife farm rodents, including Malayan

porcupines and bamboo rats, but we did not detect rodent-associated coronaviruses. The only

previously published coronavirus testing of Malayan porcupine samples carried out in China

were negative [41]. It is unclear if the Malayan porcupine samples from animals screened in this

study were infected with the avian or bat viruses or if environmental contamination or mixing

occurred with avian and bat guano. Chickens were present at the two sites where the IBV-

positive rodents were detected, and bats fly and potentially roost overhead at most farms.

'Artificial market' studies of influenza A viruses have found cage-stacking of species on top of

other species and shared water sources facilitate viral transmission [42,43]. Nevertheless, viral

sharing between species and environmental contamination or mixing (i.e. bat/bird guano landing

on rat feces) are two equally likely explanations for the presence of bat and avian coronaviruses

detected in rodent fecal and environmental samples.

The field rats were co-infected with the Longquan aa coronavirus and the murine

coronaviruses, both of which are from the Lineage A (*Embecovirus*) *Betacoronavirus* genus. Co-

infections with multiple coronaviruses deserve particular attention as this co-occurrence may

facilitate viral recombination leading to the emergence of new viruses [44,45].

At the very least, we conclude that rodents in the field and farmed rodent supply chains

are being exposed to coronaviruses from rodents, bats, and birds and perhaps creating

opportunities for coronavirus recombination events, which may lead to viruses that could spill

over into humans [46]. Repeated and more direct individual sampling of these species at these

interfaces would be useful to determine if viral sharing was occurring versus environmental

contamination of samples.

Bat guano farms

The high proportion of positive bat feces at bat guano farms indicates the potential risk of

bat guano farmers, their families, and their animals being exposed to bat coronaviruses. The

overall proportion of positives (74.8%) was higher than previous studies using similar testing

methods targeting bats in Viet Nam (22%), Thailand (7.6%), Lao PDR (6.5%), and Cambodia

(4.85%) [27,47,48]. In this region of Viet Nam, artificial roosts are typically erected in backyard

family owned plots that incorporate a mosaic of duck, goat, or pig production and crops such as

guava tress or other fruit trees and large scale kitchen gardens.

Bats have been shown to be an important evolutionary hosts of coronaviruses, including

those infecting humans [7,49–52]. Both PREDICT_CoV-17 and PREDICT_CoV-35 have been

detected previously in the *Pteropus* and *Microchiroptera* bats in Viet Nam, Cambodia, and

Nepal, which confirms that coronaviruses are capable of infecting distantly related hosts [7]. The

finding of the same virus in different bat species raises the question of whether they co-roost

and/or share viruses through contact during other activities. Utilizing shared resources such as

water or feeding on and around crops and fruit could lead to contact and facilitate a host jump.

The presence of the same virus in bat species in multiple neighboring countries supports the

suggestion by others that virus distribution coincides with their bat host distribution [7,53,54].

While there has been no testing of the pathogenicity of these bat coronaviruses in humans or

animals, they are found at close contact bat-human interfaces and further characterization is

needed to understand their host range and potential for spillover. Any general persecution of bats

because of zoonotic viruses they may carry can actually increase the number of susceptible bats

and increase transmission risk to people [56], and would interfere with the important ecosystem

services that bats provide, such as controlling insect pests of rice fields [55], plant pollination,

and seed dispersal.

462 Capacity building and outreach

Beyond the viral findings, this work represented an important opportunity for capacity

development in field, laboratory, and scientific disciplines, as well as opportunities for social

engagement and education of high-risk communities on zoonotic disease threats. The consensus

PCR approach for viral detection provides a cost-effective tool to detect emerging viruses in low-

resource settings. Our work adds to the growing body of research demonstrating the utility of this

approach to detect both known and novel viruses and co-infections in a variety of taxa, sample

types, and interfaces. In Viet Nam, the direct result is an enhanced One Health surveillance

capacity to detect important emerging or unknown viruses in humans, wildlife, and livestock. In

the communities with which we partnered, strong engagement enabled teams to sample a wide

diversity of wild animals at high-risk interfaces. Importantly, we have returned to these same

communities to share the viral findings and to educate participants with an outreach program on

how to live safely with bats [57].

Conclusions

Large percentages of coronaviruses were detected at high risk interfaces in bats and

rodents, which is of concern when assessing the potential for human exposure and spillover. The

observed viral amplification along the wildlife trade supply chain for human consumption likely

resulted from the mixing and close confinement of stressed live animals, such as field rats, and

sheds light on the potential for coronavirus shedding in other wildlife supply chains (e.g., civets,

pangolins) where similarly large numbers of animals are collected, transported, and confined.

Livestock and people living in close contact with rodents, bats, and birds shedding coronaviruses

provides opportunities for intra- and inter-species transmission and potential recombination of

coronaviruses.

Human behavior is facilitating the spillover of viruses, such as coronavirus, from animals

to people. The wildlife trade supply chain from the field to the restaurant provides multiple

opportunities for such spillover events to occur [1]. To minimize the public health risks of viral

disease emergence from wildlife and to safeguard livestock-based production systems, we

recommend precautionary measures that restrict the killing, commercial breeding, transport,

buying, selling, storage, processing and consuming of wild animals. The emergence of SARS-

CoV, MERS-CoV, and now SARS-CoV-2 highlight the importance of the coronavirus viral

family to affect global public health. The world must increase vigilance through building and

improving detection capacity; actively conducting surveillance to detect and characterize

coronaviruses in humans, wildlife, and livestock; and to inform human behaviors in order to

reduce zoonotic viral transmission to humans.

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Supporting information

S1 Table. Summary of all testing results by genus, interface, sub-interface, sample types, sites, percentage of samples testing positive, and viral species.

S2 Table: Multivariate mixed effect logistic regression showing the association between season and interface with coronavirus positives in field rats in the rodent trade.

S3 Table: GenBank accession numbers for coronavirus sequences detected in this study and for reference sequences

S1 Data. Data required for all analysis and metadata for each parameter is available at (pending DOI processing): https://doi.org/10.5061/dryad.7h44j0zrj OR https://datadryad.org/stash/share/pk3wVUxFNzTuCYZ9t8haKRPmx7V8YhTDBuHpG8JJ 9kU

S1 R Code. Code used to conduct the analysis described.

Proportion of coronavirus-positive field rats

A

Hystrix

Microchiroptera

Rattus and *Bandicota*

Rhizomys

B

Dong Nai

Dong Thap

Soc Trang