

1 **LETTER**

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3 **Rapid inactivation of SARS-CoV-2 with Deep-UV LED irradiation**

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29

30 **Abstract**

31 The spread of novel coronavirus disease 2019 (COVID-19) infections worldwide has raised
32 concerns about the prevention and control of SARS-CoV-2. Devices that rapidly inactivate viruses
33 can reduce the chance of infection through aerosols and contact transmission. This *in vitro* study
34 demonstrated that irradiation with a deep ultraviolet light-emitting diode (DUV-LED) of 280 ± 5 nm
35 wavelength rapidly inactivates SARS-CoV-2 obtained from a COVID-19 patient. Development of
36 devices equipped with DUV-LED is expected to prevent virus invasion through the air and after
37 touching contaminated objects.

38

39 **Letter**

40 The novel coronavirus SARS-CoV-2 pandemic has spread worldwide and placed countries in
41 emerging, rapidly transforming situations. The World Health Organization (WHO) clarified that
42 more than 5.3 million cases of COVID-19 and 342,000 deaths had been reported to WHO by 25
43 May 2020 [1]. Infectious virus is detected in specimens from the respiratory tract, nasopharyngeal
44 sites, and feces in COVID-19 patients [2]. Recently, infectious SARS-CoV-2 was isolated from the
45 urine of a COVID-19 patient [3]. SARS-CoV-2 is detectable in aerosols for up to 3 h, up to 4 h on
46 copper, up to 24 h on cardboard and up to 2–3 days on plastic and stainless steel [4]. To prevent
47 exposure to contaminated material (contact infection), which is one of the major transmission routes,
48 hand hygiene with alcohol is recommended, but its effectiveness in preventing the spread of
49 SARS-CoV-2 infection may be insufficient [5, 6].

50 A deep ultraviolet light-emitting diode (DUV-LED) instrument generating around 250–300 nm
51 wavelength has been reported to effectively inactivate microorganisms, including bacteria, viruses
52 and fungi [7–10], but effects on SARS-CoV-2 have not been reported. We evaluated the antiviral
53 efficacy of irradiation by DUV-LED, generating the narrow-range wavelength (280 ± 5 nm) (Nikkiso
54 Co., Tokyo, Japan), against SARS-CoV-2.

55 A strain of SARS-CoV-2 isolated from a patient who developed COVID-19 in the cruise ship
56 *Diamond Princess* in Japan in February 2020 [11] was obtained from the Kanagawa Prefectural
57 Institute of Public Health (SARS-CoV-2/Hu/DP/Kng/19-027, LC528233). The virus was propagated
58 in Vero cells cultured in minimum essential medium (MEM) containing 2% fetal bovine serum
59 (FBS). At 48 h after infection, virus stocks were collected by centrifuging the culture supernatants of
60 infected Vero cells at 3,000 rpm for 10 min. Clarified supernatants were kept at -80 °C until use.
61 Aliquots of stock virus were diluted with phosphate-buffered saline and adjusted to 2.0×10^4
62 plaque-forming units (PFU)/ml. For the evaluation of DUV-LED inactivation, aliquots of virus stock
63 ($150 \mu\text{l}$) were placed in the center of a 60-mm Petri dish and irradiated with 3.75 mW/cm^2 at work
64 distance 20 mm for a range of times ($n=3$ each for 1, 10, 20, 30, or 60 s). Each virus stock irradiated
65 with DUV-LED was serially diluted in 10-fold steps, then inoculated onto Vero monolayers in a

66 12-well plate. After adsorption of virus for 2 h, cells were overlaid with MEM containing 1%
67 carboxymethyl cellulose and 2% FBS (final concentration). Cells were incubated for 72 h in a CO₂
68 incubator, then cytopathic effects were observed under a microscope. An unirradiated virus
69 suspension was used as a negative control. To calculate PFU, cells were fixed with 10% formalin for
70 30 min, followed by staining with 0.1% methylene blue solution. The antiviral effects of DUV-LED
71 irradiations were assessed using the logPFU ratio, calculated as $\log_{10}(\text{Nt}/\text{N0})$, where
72 Nt is the PFU count of the UV-irradiated sample, and N0 is the PFU count of the sample without UV
73 irradiation. In addition, the infectious titer reduction rate was calculated as $(1 - 1/10^{\log_{10} \text{PFU ratio}}) \times 100$
74 (%). All experiments were performed in a BSL-3 laboratory.

75 We observed a marked cytopathic effect in virus-infected cells without DUV-LED irradiation
76 (Figure 1A, see “0 s”). In contrast, virus-infected cells irradiated for 60 s showed largely comparable
77 morphology to mock cells (Figure 1A, see “60 s”). To our surprise, virus-infected cells irradiated for
78 1 s showed minimal change (Figure 1A, see “1 s”). The plaque assay (Figure 1B) revealed that short
79 time DUV-LED irradiation rapidly inactivated SARS-CoV-2 (Figure 1C and Table S1). Of note, the
80 infectious titer reduction rate of 87.4% was already recognized with irradiation of virus stock for 1 s,
81 and the rate was 99.9% with irradiation for 10 s. These results suggest that DUV-LED drastically
82 inactivated SARS-CoV-2 with irradiation for even a very short time.

83 UV-LEDs providing irradiation at various peak emission wavelengths, such as UV-A (320–400
84 nm), UV-B (280–320 nm), and UV-C (100–280 nm), have been adopted to inactivate various
85 pathogenic species, including bacteria, viruses and fungi. Devices equipped with UV-LEDs are now
86 beginning to be introduced into medical fields. UV-C is considered to be the most effective
87 germicidal region of the UV spectrum, acting through the formation of photoproducts in DNA [12].
88 These pyrimidine dimers interrupt transcription, translation and replication of DNA, eventually
89 leading to inactivation of microorganisms [13]. The efficacy of this inactivation may depend not
90 only on the wavelength, but also on factors such as the target (e.g., bacterial species), light output
91 and environmental conditions. The DUV-LED we used has the characteristics of a narrow-range
92 wavelength and high power for short exposure times and long-term use. This study demonstrated for
93 the first time the rapid inactivation of SARS-CoV-2 under DUV-LED irradiation. As shown in
94 Figure 1B, cytopathic effects were observed in control Vero cells infected with SARS-Cov-2, but not
95 in these cells with DUV-LED irradiation for only 10 s. As well as in community settings, healthcare
96 settings are also vulnerable to the invasion and spread of SARS-CoV-2, and the stability of
97 SARS-CoV-2 in aerosols and on surfaces [4] likely contributes to virus transmission in medical
98 environments. No vaccines, neutralizing antibodies, or drugs are currently available for prevention
99 and treatment of SARS-CoV-2. By revealing that SARS-Cov-2 inactivation can be achieved with
100 very short-term DUV-LED irradiation, this study provides useful baseline data toward securing a
101 safer medical environment. Development of devices equipped with DUV-LED is expected to prevent

102 the virus invasion through the air and after touching contaminated objects.

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105 **Contributors**

106 H.I. and H.S. conceived the study and wrote the manuscript. A.S. and T.O. conducted the
107 experiments dealing with viruses. S.F. contributed to the study design, study supervision and
108 manuscript revision.

109

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113

114 **Declaration of interest statement**

115 H.S. receives part of his salary from Nikkiso Co., Ltd., Tokyo, Japan. Nikkiso supplied the deep
116 ultraviolet light-emitting diode (DUV-LED) instrument for evaluation. Nikkiso had no role in study
117 design, data collection and analysis, decision to publish, or preparation of the manuscript. The other
118 authors declare no conflicts of interest.

119

120 **Figure 1.** Inhibitory effects of DUV-irradiation on SARS-CoV-2.

121 (A) Cytopathic changes in virus-infected Vero cells without DUV-LED irradiation (0 s), or with
122 DUV-LED irradiation for 1, 10, 20, 30 or 60 s.

123 (B) Plaque formation in Vero cells. Virus solutions irradiated with DUV-LED for several durations
124 were diluted (100-fold) and inoculated to Vero cells. A representative result is shown.

125 (C) Time-dependent inactivation of SARS-CoV-2 by DUV-LED irradiation. The results shown are
126 the mean and standard deviation (SD) of triplicate measurements. The dashed line indicates the limit
127 of detection.

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129 **Table S1.** Differences in infectious titer with different DUV-LED irradiation times.

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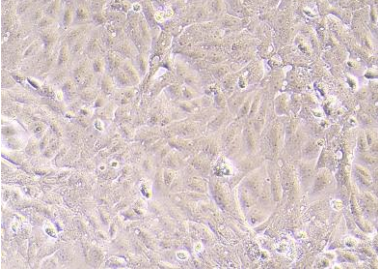
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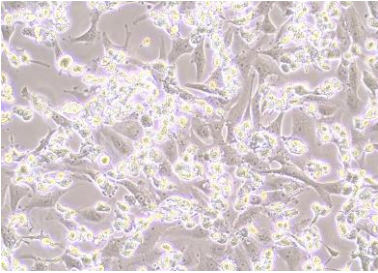
Figure 1

A

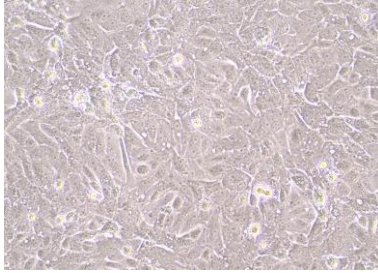
Mock



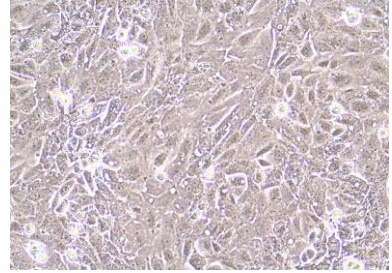
0 s



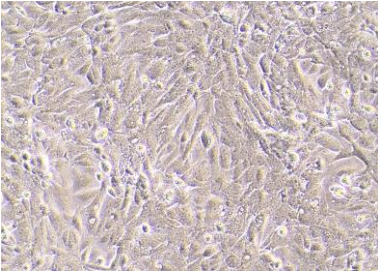
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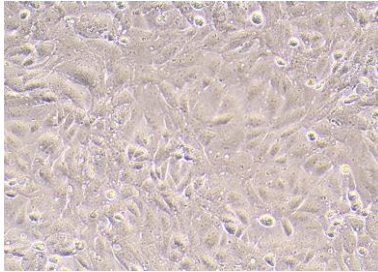
10 s



20 s



30 s



60 s

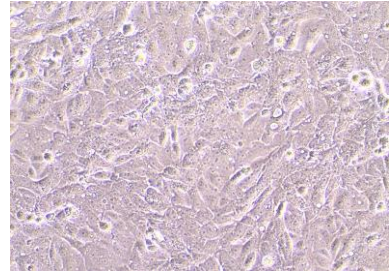
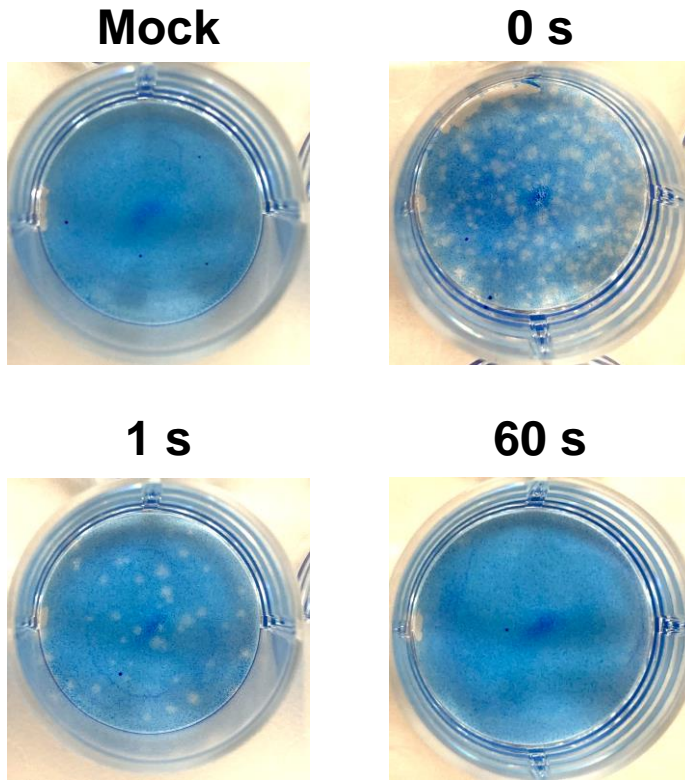
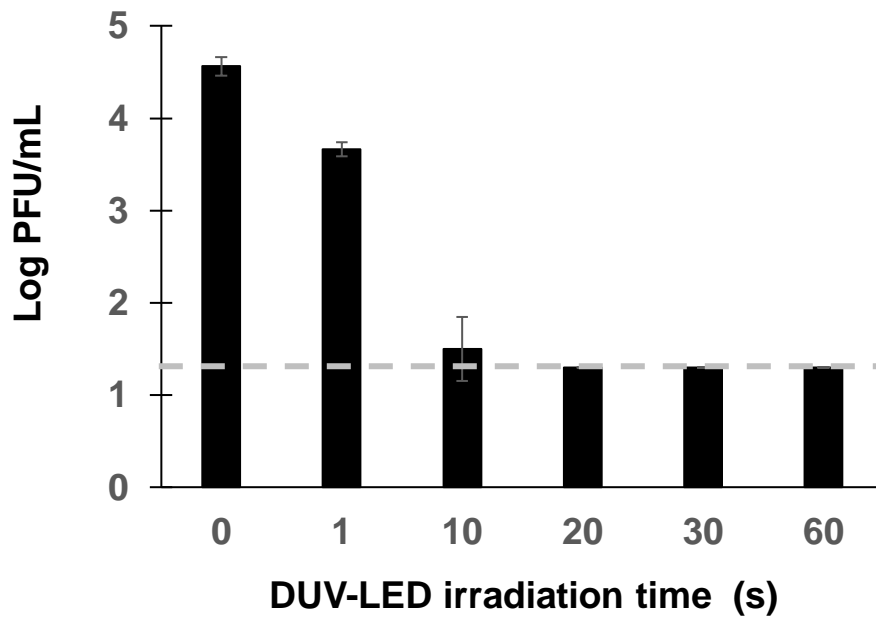


Figure 1

B



C



Supplement Table 1

Differences in infectious titer with different DUV-LED irradiation times.

| Irradiation time | control (no irradiation) | DUV-LED irradiation time | | | | |
|---|-----------------------------|--------------------------|-------------------|-------------------|--------|--------|
| | | 1 sec | 10 sec | 20 sec | 30 sec | 60 sec |
| PFU(PFU/mL) | 3.7×10^4 | 4.7×10^3 | 2.7×10^1 | 6.7×10^0 | <20 | <20 |
| Log PFU ratio ¹⁾ | — | 0.9 | 3.1 | >3.3 | >3.3 | >3.3 |
| Infection titer reduction ratio ²⁾ (%) | — | 87.4 | 99.9 | >99.9 | >99.9 | >99.9 |

¹⁾ $\log_{10} (N_t/N_0)$, where N_t is the PFU count of the UV-irradiated sample, and N_0 is the PFU count of the sample without UV irradiation. ²⁾ $(1 - 1/10^{\log \text{PFU ratio}}) \times 100$ (%).