

1 **Endochondral bone in an Early Devonian ‘placoderm’ from Mongolia**

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26 **Endochondral bone is the main internal skeletal tissue of nearly all osteichthyans^{1,2} —**
27 **the group comprising more than 60,000 living species of bony fishes and tetrapods.**
28 **Chondrichthyans (sharks and their kin) are the living sister group of osteichthyans and**
29 **have cartilaginous endoskeletons, long considered the ancestral condition for all jawed**
30 **vertebrates (gnathostomes)^{3,4}. The absence of bone in modern jawless fishes and the**
31 **absence of endochondral ossification in early fossil gnathostomes appears to lend**
32 **support to this conclusion. Here we report the discovery of extensive endochondral bone**
33 **in a new genus of ‘placoderm’-like fish from the Early Devonian (Pragian) of western**
34 **Mongolia described using x-ray computed microtomography (XR- μ CT). The fossil**
35 **consists of a partial skull roof and braincase with anatomical details providing strong**
36 **evidence of placement in the gnathostome stem group. However, its endochondral space**
37 **is filled with an extensive network of fine trabeculae resembling the endochondral bone**
38 **of osteichthyans. Phylogenetic analyses place this new taxon as a proximate sister group**
39 **of the gnathostome crown. These results provide direct support for theories of**
40 **generalised bone loss in chondrichthyans^{5,6}. Furthermore, they revive theories of a**
41 **phylogenetically deeper origin of endochondral bone and its absence in**
42 **chondrichthyans as a secondary condition^{7,8}.**

43

44 **Systematic palaeontology**

45

Placodermi M’Coy 1848

46

Minjinia turgenensis gen. et sp. nov.

47

48 **Etymology.** Generic name honours the memory of Chuluun Minjin for his extensive
49 contributions to the Palaeozoic stratigraphy of Mongolia, his enthusiastic support of this
50 work, and introducing us to the Yamaat River locality. Specific name recognises the
51 provenance of the fossil from the Turgen region, Uvs aimag of western Mongolia.

52

53 **Holotype.** Institute of Paleontology, Mongolian Academy of Sciences MPC-FH100/9.1, a
54 partial braincase and skull roof.

55

56 **Type locality.** Turgen Strictly Protected Area, Uvs province, western Mongolia; near the top
57 of the stratigraphic sequence that occurs between the Tsagaan-Salaat and Yamaat Rivers.

58

59 **Formation and age.** Upper part of Tsagaansalaat Formation, Pragian (Early Devonian)^{9,10}.

60

61 **Diagnosis.** ‘placoderm’-grade stem gnathostome with endochondral bone, deep epaxial
62 muscle cavities flanking a slender occipital ridge, and the following possible autapomorphies:
63 dermal bones covered in sparsely placed tubercles, penultimate spino-occipital nerve canal
64 substantially larger in diameter than others.

65

66 **Description**

67 MPC-FH100/9.1 consists of a partial braincase and skull roof (Fig. 1). The skull roof is
68 ornamented with sparsely distributed stellate tubercles resembling those of the Siberian
69 ‘placoderm’ *Dolganosteus*¹¹. Towards the midline of the skull roof, the tubercles are larger
70 and more pointed, and are more broadly separated from each other by unornamented fields.
71 The specimen shows signs of extensive post-mortem transport, with angles of the braincase
72 worn off and much of the skull roof and some of the braincase preserved as a mould.
73 Individual skull roof ossifications cannot be identified, although this may be due to the
74 dominantly mouldic preservation. There appears to have been a prominent nuchal plate
75 eminence comparable to acanthothoracids¹²⁻¹⁴.

76

77 **Endoskeletal tissue.** The braincase of MPC-FH100/9.1 is well ossified, comprising an
78 external bony sheath filled with an extensive matrix of spongy tissue (Fig. 2a-b; Extended
79 Data Fig. 1; Supplementary Videos 1 & 2). The trabecles forming this tissue are irregular and
80 branching, less than 1mm thick and often curved, and resemble most closely the
81 endochondral tissue of osteichthyans (Fig. 2c-d). As such, we interpret this as endochondral
82 bone. Notably, this is found in all preserved regions of the braincase, in contrast to the
83 isolated trabeculae identified as endochondral bone in *Boreaspis*¹⁵ and *Bothriolepis*¹⁶. The
84 margins of the braincase, the endocranial walls, and the boundaries of nerve and blood
85 canals, are formed from a thicker tissue which we interpret as perichondral bone. This
86 suggests that the endoskeleton of *Minjinia* comprises osteichthyan-like endochondral bone,
87 with an ossified perichondrium. To address the possible alternative explanation that it is an
88 aberrant instance of calcified cartilage, we compared the structure of this tissue with rarely-
89 preserved mineralized cartilage in the stem chondrichthyan *Diplacanthus crassismus*
90 (National Museums of Scotland specimen NMS 1891.92.334; Fig. 2e-f) observed using
91 synchrotron tomography. The cancellae within the endochondral tissue of *Minjinia* are
92 irregular, with a diameter of approximately 1-2mm. This tissue is distinctly unlike the

93 calcified cartilage of *Diplacanthus* in appearance, which consists of a densely packed matrix
94 of irregularly stacked chondrons between 20-60 μm in diameter.

95

96 **Neurocranium.** The braincase is preserved from the level of the right posterior orbital wall
97 to the posterior end of the occipital ridge. Occipital glenoid condyles are not preserved, but
98 broad, flat parachordal plates are present, separated by a midline groove that accommodated a
99 relatively narrow notochordal tunnel. A transverse fissure spans the basicranial surface at
100 about mid-length of the preserved portion. It clearly demarcates the anterior margin of the
101 parachordal plates and may correspond to the ventral cranial fissure of crown-group
102 gnathostomes. However, unlike in crown gnathostomes, it is traversed by a substantial
103 anterior extension of the cranial notochord. The courses of the lateral dorsal aortae are
104 marked by a pair of sulci on the lateral margins of the parachordal plates. A narrow, shallow
105 sulcus for the efferent hyoid artery is present on the preserved right side of the specimen,
106 immediately behind the level of the orbit (Fig. 1a).

107 The lateral surface of the braincase is preserved on the right side as a mouldic
108 impression in the matrix (Fig. 1). A sharply demarcated hyoid fossa is present on the lateral
109 wall of the otic region (Fig. 1). Posterior to this, a stout but pronounced vagal process with a
110 pair of rounded eminences likely corresponds to the branchial arch articulations. There is no
111 evidence for a pair of anterior and posterior divisions to the vagal process, which are
112 typically seen in other ‘placoderms’. A well-developed ‘placoderm’-like craniospinal
113 process is absent; its homologous position is instead covered in perichondral bone and
114 marked by a low ridge (Fig. 1).

115 In posterior view, a tall, narrow median otic ridge is evident and resembles the
116 morphology of *Romundina*¹⁷ and *Arabosteus*¹⁸. Similar to these taxa, the median otic ridge is
117 flanked by two large occipital fossae for the epaxial musculature. The notochordal tunnel is
118 approximately the same size as or smaller than the foramen magnum, as in ‘placoderms’ and
119 in contrast with crown-group gnathostomes. A metotic fissure is absent.

120

121 **Endocast.** A partial cranial endocast is preserved, consisting of the hindbrain cavity, partial
122 midbrain cavity, labyrinth cavities, and posteromedial corner of the orbital region. The two
123 primary trunk canals of the trigeminal nerve (N.V₁ and N.V_{2,3}) are preserved (Fig. 1;
124 Extended Data Fig. 2). The acoustic (N.VIII) and facial nerve (N.VII) canals share a common
125 trunk canal behind the trigeminal nerves, as in many other ‘placoderms’^{17,19-21}. The
126 supraorbital branch opens into the rear wall of the orbit and part of its supraorbital course

127 is preserved (Extended Data Figs. 2, 3). A slender branch extends below the labyrinth and
128 divides into palatine and hyomandibular branches (Extended Data Figs. 2, 3). As in other
129 ‘placoderm’-grade taxa, the vagus nerve (N. X) trunk canal is very large in diameter and exits
130 from immediately behind the labyrinth cavity (Fig. 1; Extended Data Fig. 2). The spino-
131 occipital region resembles other ‘placoderms’ in being extended. At least four spino-occipital
132 nerve canals are present in a linear series, and the penultimate canal is largest in diameter
133 (Fig. 1; Extended Data Fig. 2). Intercalating these is a network of occipital artery canals
134 branching from the dorsal aortae.

135 The skeletal labyrinth is not complete on either side of the specimen, but can mostly
136 be reconstructed according to the assumption of bilateral symmetry. The most significant
137 feature is that the labyrinth and endolymphatic cavity are joined to the main endocavity
138 chamber (Fig. 1). This is a striking contrast to other ‘placoderms’ and closely resembles
139 crown-group gnathostomes²². The endolymphatic canals are elongate and tubular, extending
140 posterolaterally to reach the skull roof, though external openings cannot be clearly identified.
141 The anterior semi-circular canal follows the saccular cavity closely as in petalichthyids²³(Fig.
142 1; Extended Data Fig. 2). However, the horizontal and posterior canals appear to extend well
143 away from the saccular chamber (Fig. 1, Extended Data Fig. 2). The dorsal junctions of the
144 anterior and posterior canals are joined in a crus commune, as in *Romundina*¹⁷ and
145 *Jagorina*¹⁹. A sinus superior is absent.

146

147 **Phylogenetic analyses**

148 We conducted phylogenetic analyses under four different protocols: equal weights
149 parsimony, implied weights parsimony, an unpartitioned Bayesian analysis, and a Bayesian
150 analysis with characters partitioned by fit determined under implied weights parsimony²⁴ (see
151 Extended Data Figs. 4-7). All phylogenetic analyses consistently place *Minjinia* as a stem-
152 group gnathostome, proximate to the gnathostome crown (Fig. 3, Extended Data Figs 4,5).
153 Equal weights parsimony recovers *Minjinia* in a position crownward of arthrodires but
154 outside of a grade consisting of *Entelognathus*, *Ramirosuarezia*, and *Janusiscus*. Under
155 implied weights, these three taxa move onto the osteichthyan stem and *Minjinia* is placed as
156 the immediate sister taxon of the gnathostome crown. Under Bayesian analyses, arthrodires
157 are resolved as more crownward than *Minjinia*. However, the latter analyses fail to recover
158 arthrodires as a clade and the node uniting them with the crown to the exclusion of *Minjinia*
159 is extremely weakly supported (posterior probability: 0.52-0.55). Under parsimony, the
160 crownward position of *Minjinia* is unambiguously supported by the skeletal labyrinth and

161 endolymphatic duct being confluent with the main cranial cavity²² (Fig. 3). In common with
162 arthrodires and the gnathostome crown, *Minjinia* possesses a division of the facial nerve deep
163 to the transverse otic process (Fig. 1; Extended Data Fig. 2). However, *Minjinia* is excluded
164 from the gnathostome crown group due to the absence of a metotic fissure and a posterior
165 dorsal fontanelle, and presence of broad, flat parachordal plates expanded behind the saccular
166 cavity (Fig. 3, Supplementary Information).

167 We undertook ancestral states reconstructions to assess the evolutionary history of
168 endochondral bone (Fig. 3, Extended Data Figs. 6 & 7). Interestingly, parsimony analysis
169 fails to recover secondary homology of this trait between *Minjinia* and osteichthyans. The
170 crownward placement of *Minjinia* is, in fact, based on independent evidence relating to
171 anatomical features of the braincase and endocast. However, the strict precision of parsimony
172 reconstructions makes it insensitive to underlying uncertainty. To explore this, we used
173 likelihood reconstructions and compared the ancestral state reconstructions under equal rates
174 (ER) and all rates different (ARD) variants of the MkV model on branch-length-rescaled
175 parsimony trees and Bayesian trees. On the parsimony trees, both models show substantial
176 non-zero probabilities (0.23 for ER; 0.39 for ARD; Extended Data Table 1) for the presence
177 of endochondral bone in the common node of *Minjinia* and Osteichthyes (Extended Data Fig.
178 6) in the parsimony trees. The ARD model shows the best likelihood score and a better AIC
179 fit for endochondral bone (Extended Data Table 1), favouring repeated losses of this tissue
180 over multiple gains (see Discussion). The values are substantially lower in the Bayesian trees
181 (Extended Data Fig. 7, Extended Data Table 1), but this results from the relative positions of
182 *Minjinia* and arthrodires, which is not well supported in those trees.

183

184 Discussion

185 *Minjinia* presents an unusual discovery of extensive endochondral bone in a ‘placoderm’-
186 grade fish, with repercussions for the phylogenetic origin of this tissue and the problem of
187 early gnathostome relationships more generally. The vertebrate skeleton is split into two
188 systems: the exoskeleton (external achondral dermal bones) and endoskeleton (internal
189 chondral bones)¹. Dermal bone evolved at least 450 million years ago in jawless stem
190 gnathostomes, but the endoskeleton in these taxa is not endochondrally ossified (but see
191 below). More crownward stem gnathostomes (osteostracans and ‘placoderms’) surround their
192 cartilaginous endoskeleton in a sheath of perichondral bone. Extant chondrichthyans lack
193 both dermal and perichondral bone, possessing a cartilaginous endoskeleton enveloped by
194 prismatic calcified cartilage. Endochondral bone, in which the cartilaginous endoskeletal

195 precursor is invaded by and eventually replaced by bone, is widely considered an
196 osteichthyan apomorphy based on clear prior polarity^{3,7,25,26}. However, recent work has cast
197 doubt on this assertion. The recognition that dermal bone is secondarily lost in
198 chondrichthyans^{27,28} is consonant with prior knowledge of the loss of perichondral bone in
199 this same lineage²⁹. Taken together, this has revived uncertainty about the true phylogenetic
200 timing of the origin of endochondral ossification⁸.

201 *Minjinia* does not represent the first report of endochondral bone outside of
202 Osteichthyes. However, it is by far the most extensive and unequivocal example, and raises
203 explicit questions in light of the proximity of *Minjinia* to the gnathostome crown. Isolated
204 examples of trabecular bone, typically restricted to a small region of the neurocranium, have
205 historically been reported in boreaspid osteostracans^{15,30}, buchanoosteid arthrodiroids³¹ and
206 petalichthyids³². However, these reports have all been dismissed as misidentifications²⁶,
207 possibly representing the retreat of perichondral bone deposited during cartilage growth³³.
208 Most recently, trabeculae in supposed endoskeletal bones of *Bothriolepis* have been termed
209 endochondral bone¹⁶, although the small scale of these is in line with ‘superficial’
210 perichondral trabeculae seen elsewhere. In line with ref. 26, we found no evidence of
211 endochondral bone in material of *Buchanosteus* held in the Natural History Museum,
212 London, or indeed in any other ‘placoderms’ we have examined. The *Epipetalichthys*
213 holotype (Museum für Naturkunde, Berlin specimen MB.f.132.1-3) shows an apparently
214 spongiose infilling in the anterior region of the braincase, but the identity of this structure, or
215 even whether it is biological, cannot be determined.

216 Does endochondral bone have a deep origin within the gnathostome stem group? This
217 would imply repeated losses of this tissue. We do find some statistical support for this
218 hypothesis (Fig. 3, Extended Data Figs. 6, 7; Extended Data Table 1), and the model is well
219 justified on prior phylogenetic and biological grounds. Endochondral bone has long been
220 known to be inconsistently developed across ‘primitive’ bony fishes: incomplete or entirely
221 absent ossification of the endoskeleton is known in both Palaeozoic actinopterygians³⁴ and
222 sarcopterygians³⁵, as well as more recent taxa³⁶. The frequent absence of endochondral bone
223 is considered secondary, and other controlling factors such as body size, maturity, mechanical
224 stress, and buoyancy can determine its degree of development¹. Our findings are also in
225 agreement with studies establishing a genetic basis for secondary loss of all bone types within
226 chondrichthyans^{5,37,38}, with the failure to produce endochondral bone likely representing
227 arrested development of chondrocytes as opposed to a primary lack of ability⁶.

228 Another confounding factor in this question is the problem of ‘placoderm’
229 relationships. Although currently resolved as a deeply pectinate grade along the gnathostome
230 stem, the backbone of this arrangement has poor statistical support and there is a lack of
231 consistency in the arrangement of plesia. *Minjinia* itself highlights this uncertainty, given its
232 highly unexpected character combinations. Notwithstanding its endochondral bone and
233 crown-gnathostome-like inner ear structure, it strongly resembles ‘acanthothoracids’—the
234 ‘placoderms’ widely considered among the most removed from the gnathostome crown (i.e.
235 most ‘primitive’). This apparent character conflict could perhaps be more easily reconciled
236 with a more coherent (though not necessarily monophyletic) ‘placoderm’ assemblage.
237 Indeed, the highly pectinate structure of the ‘placoderm’ grade seems symptomatic of an
238 overemphasis on characters and taxa resembling the crown group, thereby undersampling
239 characters that could stabilise a clear picture of ‘placoderm’ interrelationships.

240 *Minjinia* reveals new data on ‘placoderm’ endoskeleton and tissue diversity from
241 Mongolia—an otherwise extremely poorly known biogeographic realm for early
242 gnathostomes. The phylogenetic placement of this ‘acanthothoracid’-like taxon crownward of
243 all non-maxillate ‘placoderms’, in conjunction with possession of extensive endochondral
244 bone, highlights the importance of material from traditionally undersampled geographic
245 areas. The presence of endochondral bone renews the hypothesis that this tissue is
246 evolutionarily ancient and was lost secondarily in chondrichthyans^{7,8}. This view is overall
247 consistent with evidence of generalised bone loss in chondrichthyans, potentially as a result
248 of the suppression of bone-generating molecular genetic pathways^{6,38}. Continued work in
249 Mongolia and re-evaluation of phylogenetic datasets will be necessary to address this, with
250 the results likely to lead to substantial re-evaluation of gnathostome phylogeny.

251

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263 assistance of Paul Tafforeau. SG was supported by a Royal Society Dorothy Hodgkin
264 Research Fellowship. Matt Friedman is thanked for undertaking the X-ray computed
265 microtomography analysis. TNT was made available with the support of the Willi Hennig
266 Society.

267

268 **Author Contributions:** MDB conceived and designed the study. MDB, AJ, YAA, and EZ
269 participated in all field seasons. RPD and AJ undertook preliminary CT scanning and
270 segmentation that revealed the fossil was a ‘placoderm’ and had endochondral bone. RS
271 discovered the first vertebrate remains in the first field season at Yamaat Gol in 2010. SG did
272 most of the segmentation of *Minjinia* with input from MDB. AJ performed segmentation of
273 *Diplacanthus* tissue. MC provided input on occipital comparative morphology of
274 ‘placoderms’. RPD provided data and comparative analyses and data for endoskeletal tissue.
275 YAA provided background on the geology, palaeontology, and stratigraphy of the type
276 location; EZ and YAA organized field logistics and permitting. MDB, SG, MC, RPD, and AJ
277 undertook the anatomical interpretation and prepared the figures. MDB and SG conducted the
278 phylogenetic analyses. RS conducted the parsimony branch support analyses. TG wrote the
279 script for generating MrBayes partitions from TNT’s character fits table and conducted the
280 likelihood and model-fitting analyses. The manuscript was written by MDB, RPD, and SG.

281

282 **Author Information:**

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287

288 **Methods**

289

290 **X-ray computed microtomography.** We scanned MPC-FH100/9.1 using the Nikon XT
291 225s at the Museum of Paleontology, University of Michigan with the following parameters:
292 200kV, 140 μ A, over 3123 projections and a voxel size of 32.92 μ m. We conducted
293 segmentation using Mimics 19.0 (<http://biomedical.materialise.com/mimics>; Materialise,
294 Leuven, Belgium) and we imaged models for publication using Blender
295 (<https://www.blender.org>).

296 **Synchrotron light propagation phase contrast tomography.** We imaged *Diplacanthus*
297 *crassismus* specimen NMS 1891.92.334 on Beamline 19 of the European Synchrotron
298 Radiation Facility, using propagation phase-contrast synchrotron microtomography. We
299 performed a spot scan with an energy of 116keV, achieving a voxel size of 0.55 μm . We
300 processed the resulting tomograms using VG StudioMax 2.2 (Volume Graphics, Germany),
301 and prepared images in Blender.

302 **Phylogenetic analysis.** We conducted a parsimony analysis using TNT 1.5³⁹ and Bayesian
303 analysis using MrBayes v 3.2.7⁴⁰. The dataset consisted of 95 taxa and 284 discrete
304 characters based on a pre-existing dataset⁴¹. We employed Osteostraci and Galeaspida as
305 composite outgroups. We conducted parsimony analysis using both equal weights and
306 implied weights methods. Global settings were 1000 search replicates and a hold of up to 1
307 million trees. Equal weights parsimony analyses were conducted using the ratchet with
308 default settings. Implied weights parsimony used a concavity parameter of 3 and the search
309 was without the ratchet. Command lists are included in Supplementary Information. We
310 conducted Bayesian analysis using both a partitioned and unpartitioned dataset. We used the
311 Mkv model⁴² and gamma rate distribution. We ran the analyses for 5 million generations
312 with a relative burn-in fraction of 0.25. Runs were checked for convergence using Tracer⁴³.
313 We partitioned the dataset using a newly proposed method²⁴ that partitions the data according
314 to homoplasy levels. Using the results of implied weights parsimony conducted in TNT, we
315 created a text table of character fit values. We wrote an R⁴⁴ script to generate a list of
316 partition commands for MrBayes. To reduce the number of partitions with small numbers of
317 characters, we concatenated the partitions by rounding the fitness scores to 2 significant
318 figures, yielding 10 individual partitions.

319 We assessed parsimony ancestral states visually using Mesquite⁴⁵. Likelihood and
320 Bayesian ancestral states were estimated in R using the castor package⁴⁶. Prior to calculating
321 likelihood ancestral states on parsimony trees, we scaled branch lengths using PAUP*⁴⁷ and
322 calculated the likelihood scores for all of the trees under the Mkv model. The trees were then
323 exported with branch lengths. To account for overall uncertainty in tree estimates, we
324 estimated ancestral states on 100 trees randomly selected from the fundamental set of most
325 parsimonious trees and two times 50 trees selected from the 75% last trees of each posterior
326 tree distribution from the Bayesian analysis. We then run an ancestral states estimation Mk
327 model (using the castor R package) using both the Equal Rates (ER) and All Rates Different
328 (ARD) models. This resulted in 400 ancestral states estimations. For each estimation we
329 extracted the overlap log likelihood, the AIC (counting one parameter for the ER model and

330 two for the ARD model) and the scaled log likelihood (probability) for the presence and
331 absence of the endochondral bone character (character 4) for the last common node of
332 *Minjinia* and crown-group gnathostomes. We present the median value of these distributions
333 of the estimations overall log likelihoods, AICs and presence or absence of endochondral
334 bone in Extended Data Table 1.

335

336 **Data availability**

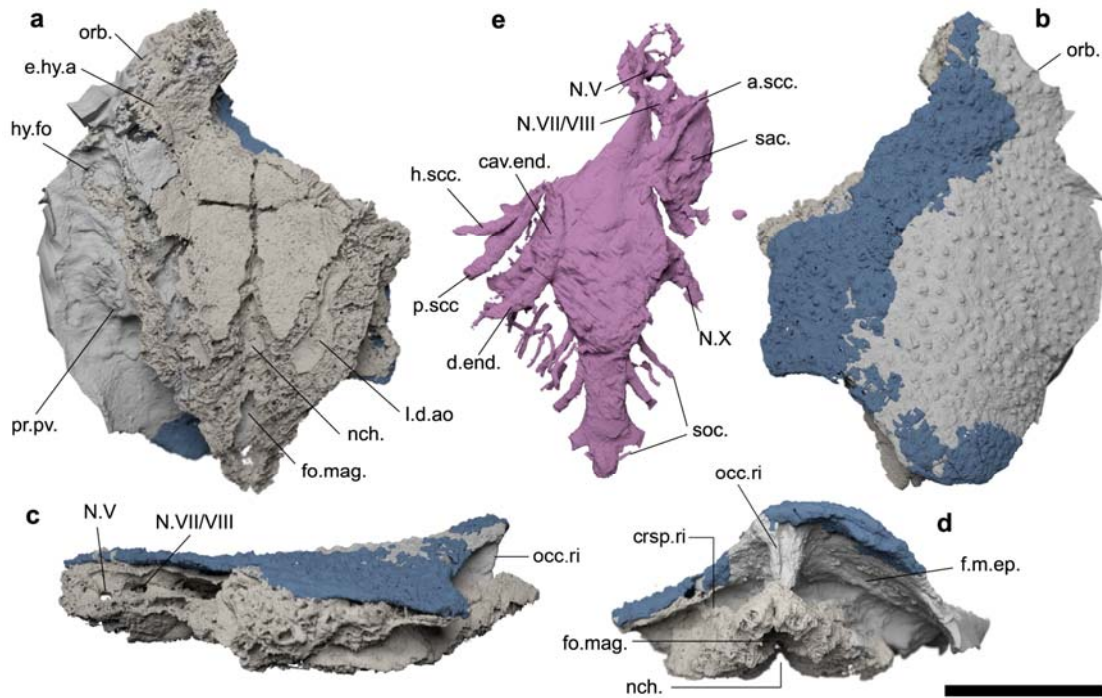
337 The holotype specimen of *Minjinia turgenensis* will be permanently deposited in the
338 collections of the Institute of Paleontology, Mongolian Academy of Sciences. Original
339 tomograms are available at (doi:10.6084/m9.figshare.12301229) and rendered models are
340 available at (doi:10.6084/m9.figshare.12301223). The phylogenetic character list and dataset
341 are available as Supplementary Information S1 and S2. The LifeScience Identifier for
342 *Minjinia turgenensis* is urn:lsid:zoobank.org:act:82A1CEEC-B990-47FF-927A-
343 D2F0B59AEA87

344

345 **Code availability**

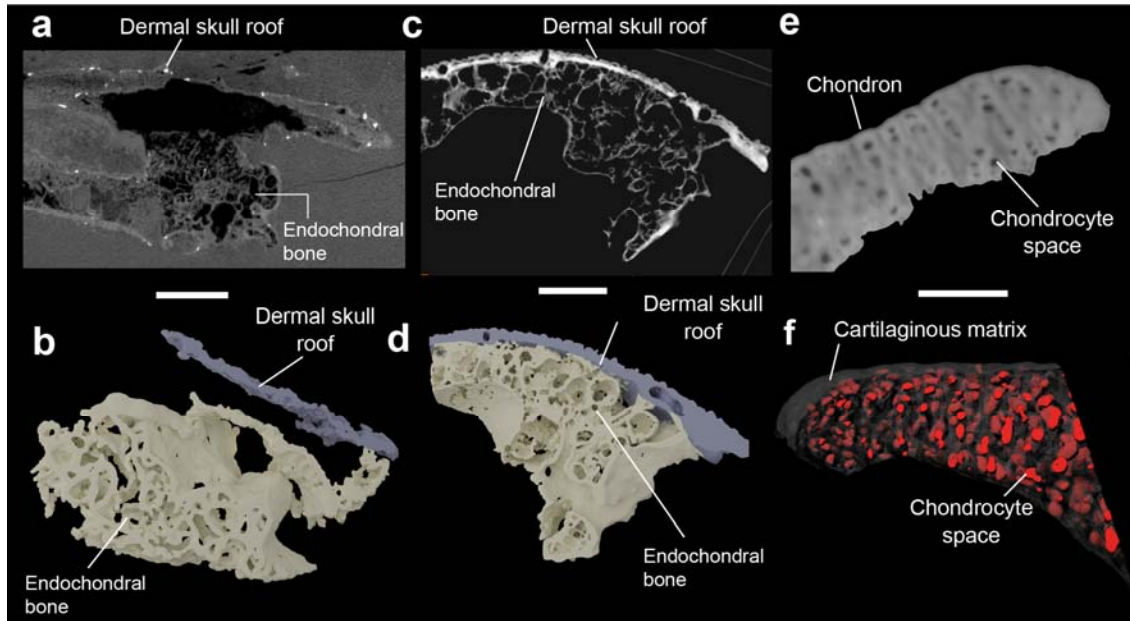
346 R code for generating partitions based on character fits and code for likelihood ancestral
347 states reconstructions and plots are available in the Supplementary Information.

348



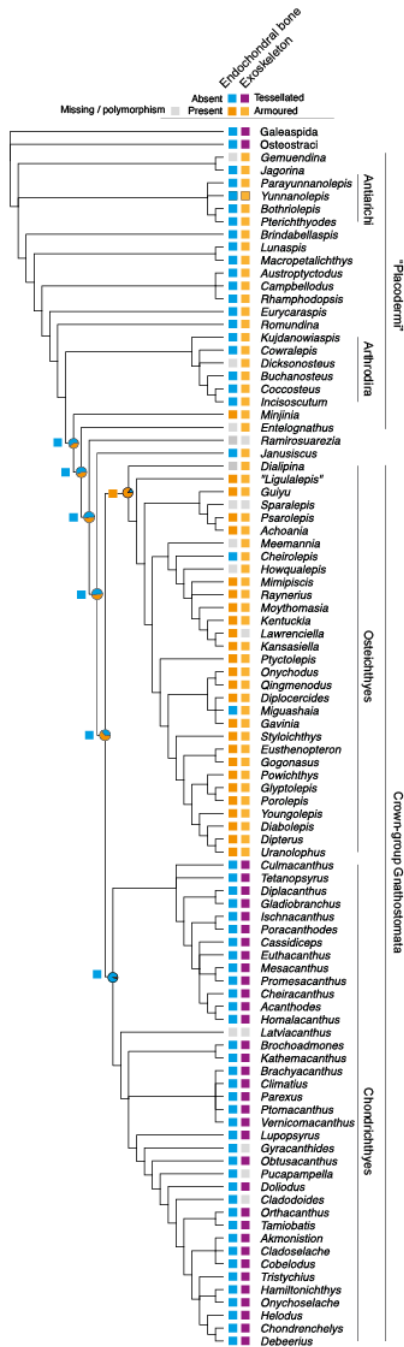
349

350 **Fig. 1 | MPC-FH100/9.1 a 'placoderm' skull roof and braincase from the Early**
351 **Devonian of Mongolia. a, Ventral view. b, Dorsal view. c, Left lateral view. d, Posterior**
352 **view. e, Braincase endocavity in dorsal view. Taupe: endoskeleton; grey: mould; pink:**
353 **endocavity; blue: exoskeleton. a.scc., anterior semicircular canal; cav.end., endolymphatic**
354 **cavity; crsp.ri, craniospinal ridge; d.end., endolymphatic duct; e.hy.a., sulcus for the efferent**
355 **hyoid artery; f.m.ep., epaxial muscle fossa; fo.mag., foramen magnum; h.scc., horizontal**
356 **semicircular canal; l.d.ao, sulcus for the lateral dorsal aorta; N.V, trigeminal nerve canal;**
357 **N.VII, facial nerve canal; N.VIII, acoustic nerve canal; N.X, vagus nerve canal; nch.,**
358 **notochordal canal; occ.ri, occipital ridge; orb., orbit; p.scc, posterior semicircular canal;**
359 **pr.pv., paravagal process; sac., saccus; soc., spino-occipital nerve canals. Scale bar, 20 mm.**



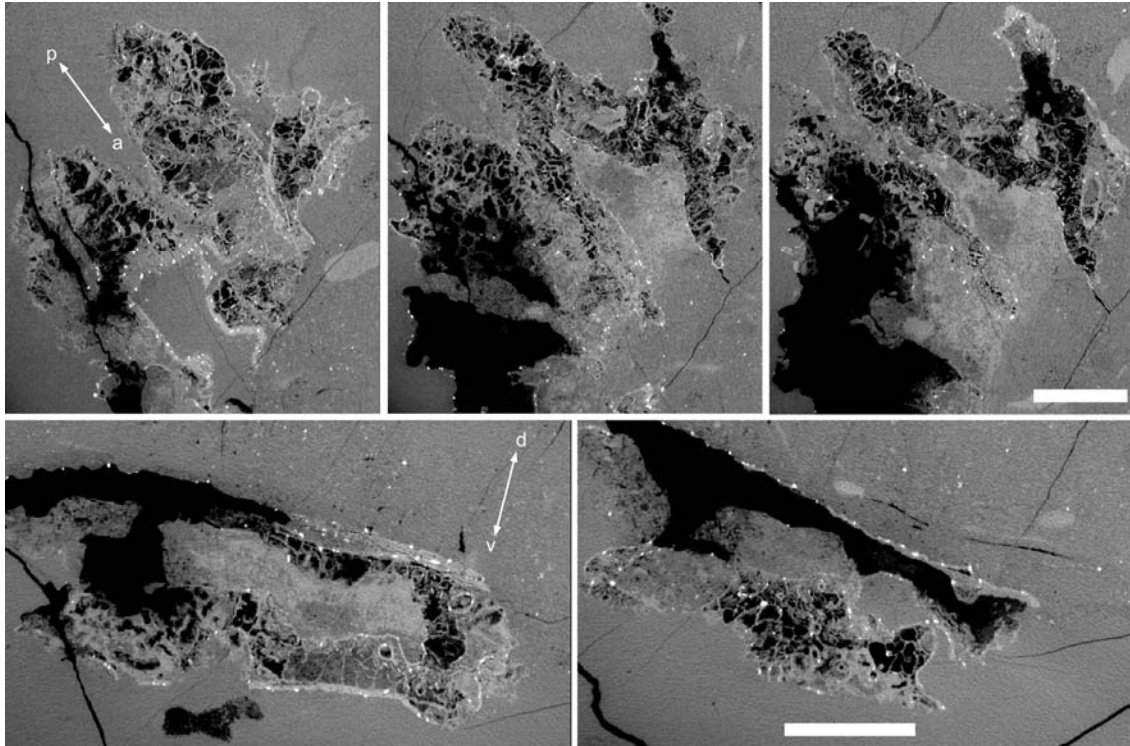
360

361 **Fig. 2 | Endoskeletal mineralisation in fossil gnathostomes.** **a**, Transverse tomographic
362 slice through MPC-FH100/9.1. **b**, Three-dimensional rendering of trabecular bone structure.
363 **c**, Transverse tomographic section through the braincase of the osteichthyan *Ligulalepis*. **d**,
364 Three-dimensional rendering of the trabecular bone in *Ligulalepis* (**c** and **d** use data from⁴¹).
365 **e**, Synchrotron tomography image of the calcified cartilage of the stem-group chondrichthyan
366 *Diplacanthus crassissimus* specimen NMS 1891.92.334. **f**, Semi-transparent three-
367 dimensional structure of calcified cartilage of NMS 1891.92.334. Scale bars, **a** and **b**, 10 mm;
368 **c** and **d**, 1 mm ; **e** and **f**, 150 µm.



369

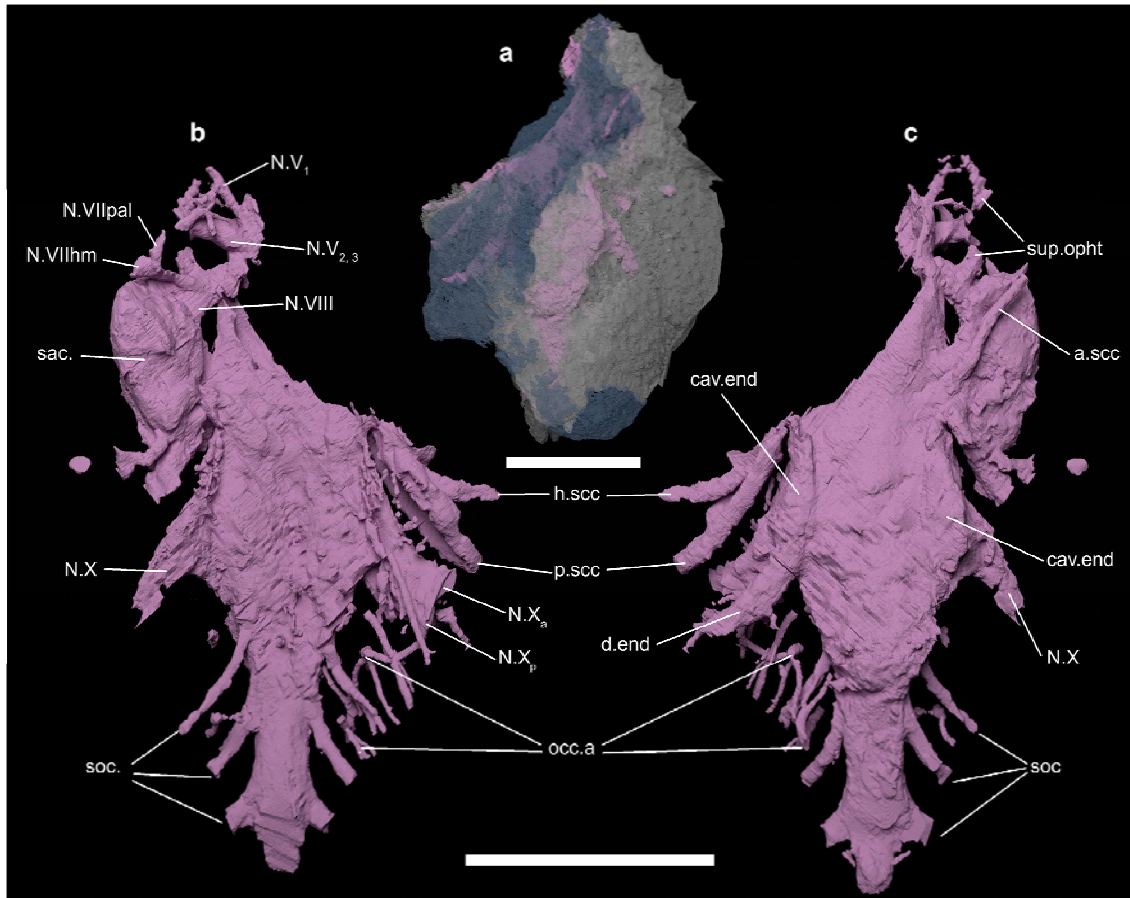
370 **Fig. 3 | Summary phylogenetic relations of early gnathostomes showing distribution of**
 371 **endochondral bone and exoskeletal armour.** Squares at nodes indicate parsimony
 372 reconstruction for endochondral bone. Pie charts at nodes show likelihood reconstructions for
 373 the same character under the all-rates-different model (see Extended Data Figs 6 & 7 for
 374 competing reconstructions). Grey box indicates uncertainty. Loss of endochondral bone maps
 375 closely with generalised loss of bone in chondrichthyes where exoskeletal armour and
 376 perichondral bone are also absent.



377

378 **Extended Data Fig. 1 | Tomograms of endoskeletal ossification in *Minjinia*.** Top row:
379 semi-coronal sections through braincase. Double-headed arrows indicate anterior-posterior
380 (a-p) dorsal-ventral (d-v) axes. Bottom row: semi-transverse sections through posterior part
381 of endocranium. Voids of black space represent mouldic preservation. Scale bars, 10 mm and
382 apply across each row of panels.

383

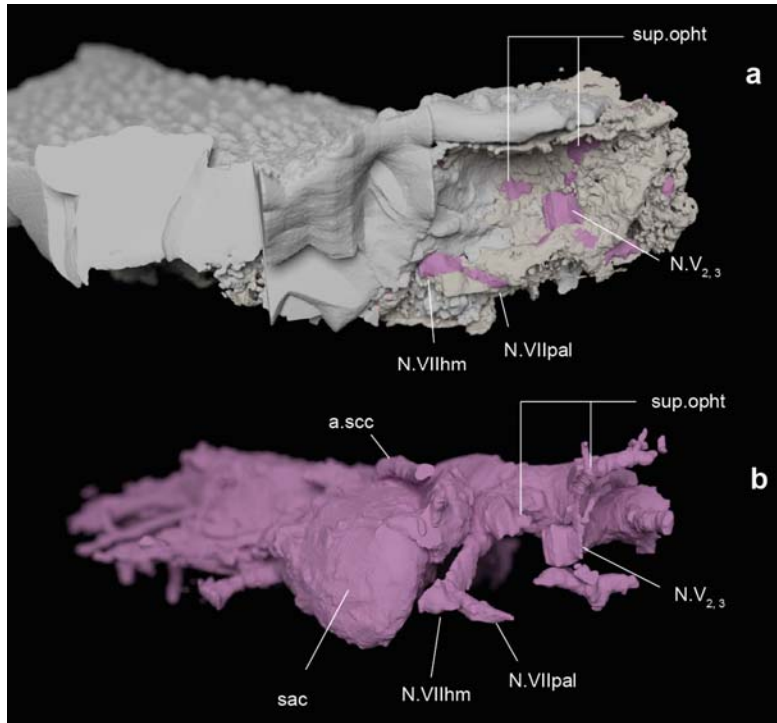


384

385 **Extended Data Fig. 2 | Braincase endocavity of *Minjinia*.** **a**, Semi-transparent rendering of
386 skull roof and braincase (grey and blue) showing extent of endocavity (pink). **b**, Ventral
387 view. **c**, Dorsal view. a.scc., anterior semicircular canal; cav.end., endolymphatic cavity;
388 d.end., endolymphatic duct; h.scc., horizontal semicircular canal; l.d.ao., sulcus for the lateral
389 dorsal aorta; N.V, trigeminal nerve canal; N.VIIhm, hyomandibular branch of facial nerve
390 canal; N.VIIpal, palatine branch of facial nerve canal; N.VIII, acoustic nerve canal; N.X,
391 vagus nerve canal, N.X_a, anterior branch of vagus nerve canal; N.X_p, posterior branch of
392 vagus nerve canal; occ.a, occipital artery canals; p.scc, posterior semicircular canal; sac.,
393 sacculus; soc., spino-occipital nerve canals; sup.opht, canal for supra-ophthalmic nerve. Scale
394 bars, 20 mm (upper scale bar associates with **a**, lower scale bar associates with **b** and **c**).

395

396



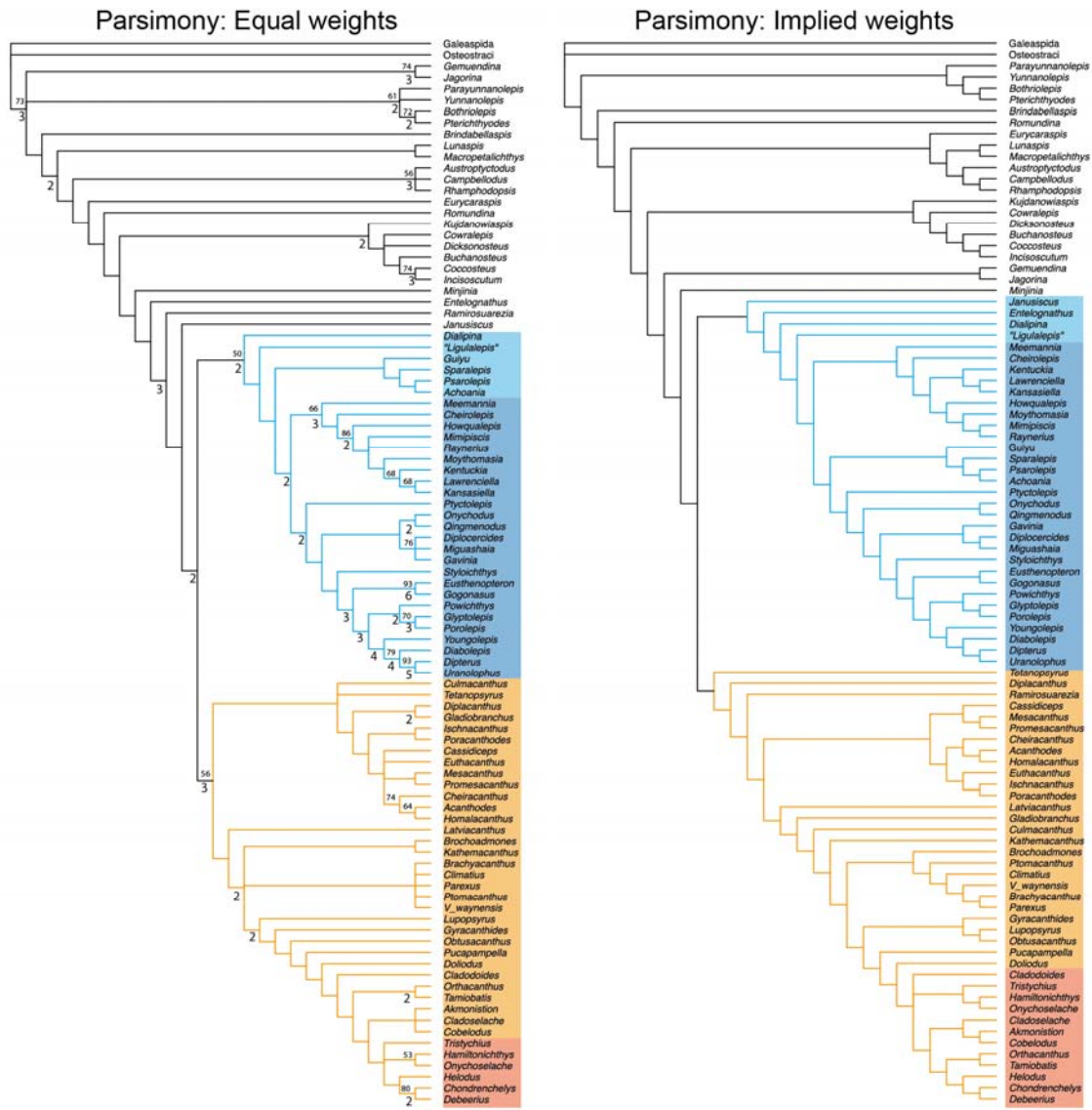
397

398 **Extended Data Fig. 3 | Right orbital wall and innervation pattern of *Minjinia*.** **a**, orbit in

399 anterolateral view showing disposition of nerve openings (pink infill). **b**, endocast in the

400 same perspective showing the relationship between never canals and endocast.

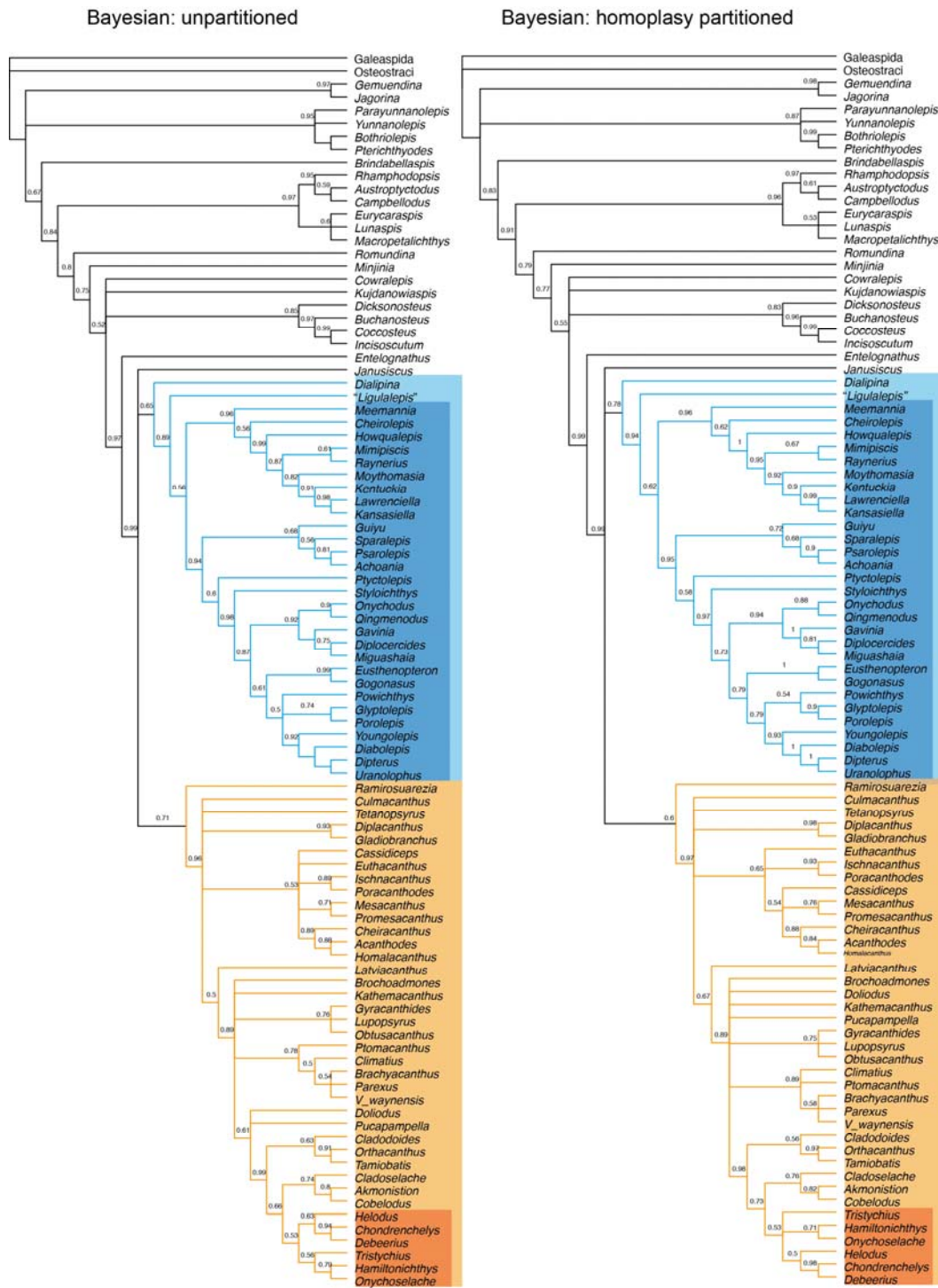
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402

403 **Extended Data Fig. 4 | Results of phylogenetic parsimony analysis.** Dataset consists of 95
 404 taxa and 284 characters. Both trees are strict consensus topologies. Equal weights parsimony
 405 analysis using the ratchet resulted in 240 trees with a length of 832 steps. Implied weights
 406 parsimony analysis using random addition sequence + branch-swapping resulted in two
 407 optimal trees with score 85.23240. Double-digit figures above internal branches are bootstrap
 408 values of 50% and over; single-digit figures below branches are Bremer decay index values.
 409 Blue shading: osteichthyan total group (dark blue: crown group); orange shading:
 410 chondrichthyan total group (dark orange: crown group).

411

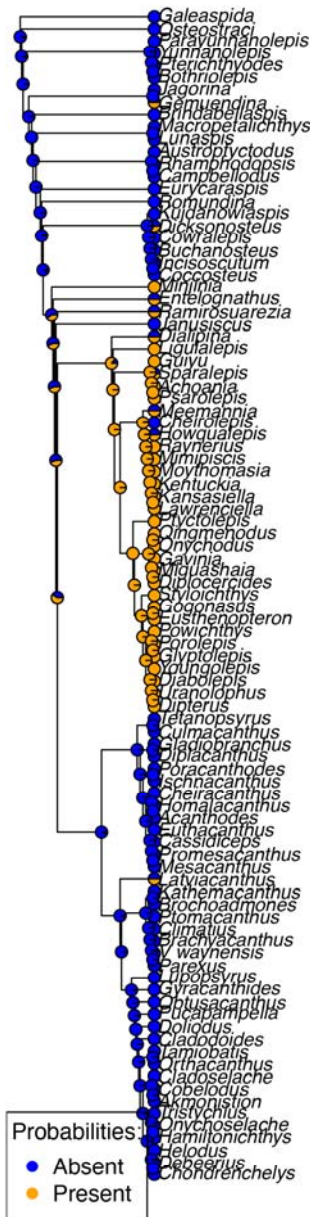


412

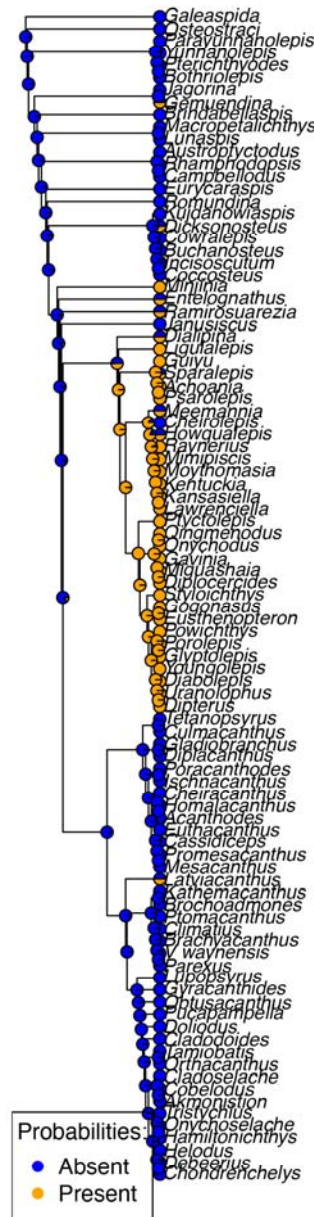
413 **Extended Data Fig. 5 | Results of Bayesian phylogenetic analysis using both partitioned**
 414 **and unpartitioned data.** Majority-rules consensus trees with posterior probabilities shown
 415 along branches. Blue shading: osteichthyan total group (dark blue: crown group); orange
 416 shading: chondrichthyan total group (dark orange: crown group).

417

Endochondral bone estimation
(parsimony – ARD)



Endochondral bone estimation
(parsimony – ER)



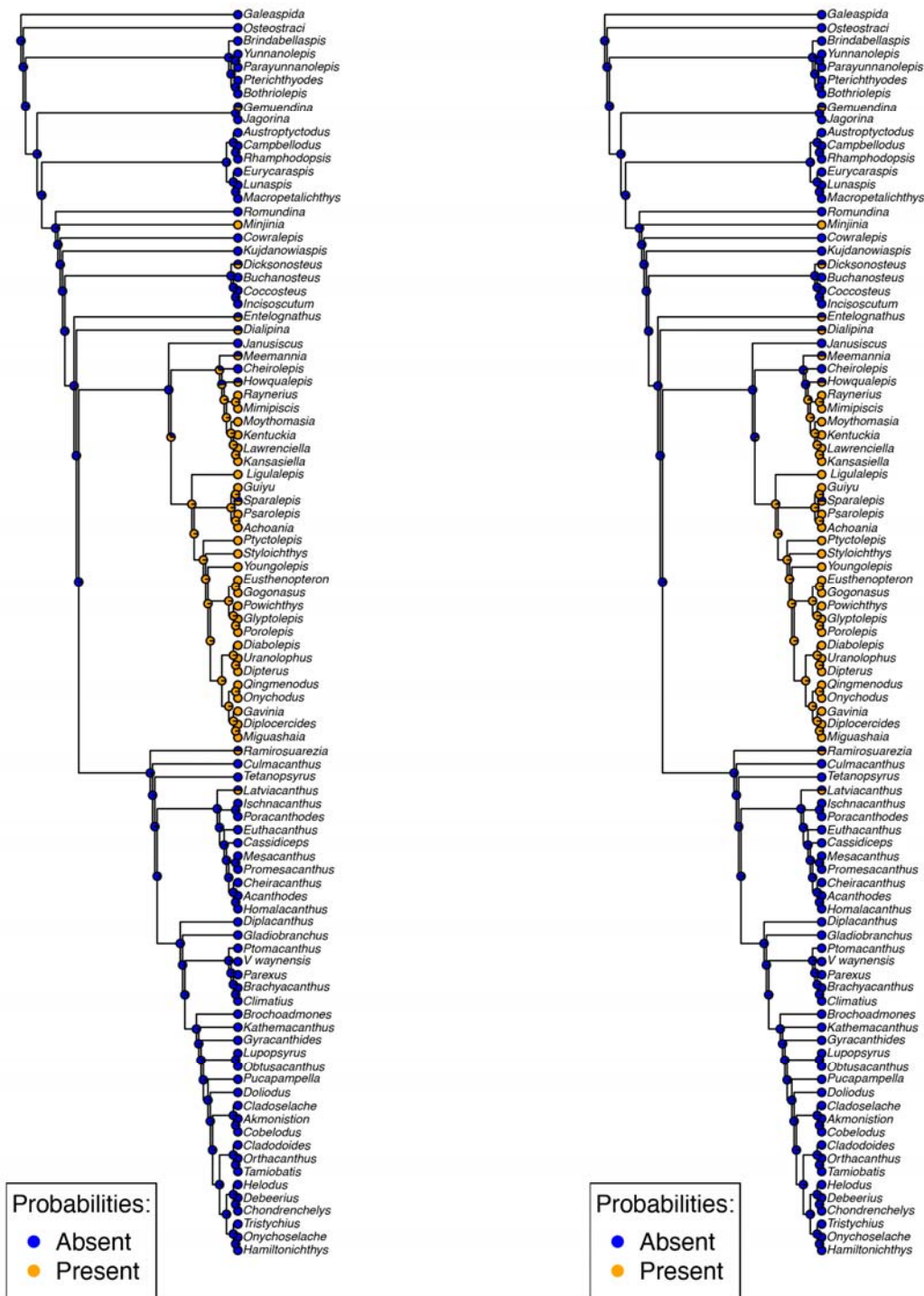
418

419 **Extended Data Fig. 6 | Likelihood ancestral state mapping of endochondral bone on**

420 **equal weights parsimony results.** ARD, all rates different model; ER, equal rates model.

Endochondral bone estimation (bayesian – ARD)

Endochondral bone estimation (bayesian – ER)



421

422

423

424

Extended Data Fig. 7 | Likelihood ancestral state mapping of endochondral bone on unpartitioned Bayesian analysis results. ARD, all rates different model; ER, equal rates model.

425 **Extended Data Table 1 | Tree distribution (100) ancestral states estimation results.** ER =
426 Equal rates model; ARD = All Rates Different model. The columns AIC and log.lik represent
427 the median AIC and log.lik across the 100 parsimony and bayesian trees (for both models).
428 The columns Absent and Present represent the median scaled likelihood for the endochondral
429 bone state.

Node	Tree	Model	log.lik	AIC	Absent	Present
<i>Minjinia</i> :crown gnathostomes	Parsimony	ER	-27.60	57.20	0.94	0.06
		ARD	-25.47	54.93	0.61	0.39
	Bayesian	ER	-29.94	61.89	0.98	0.02
		ARD	-27.69	59.38	0.82	0.18

430

431

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