

Comparison of manual and automated ventricle segmentation in the maternal immune stimulation rat model of schizophrenia

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1 **Abstract**

2 Maternal immune stimulation (MIS) is strongly implicated in the etiology of neuropsychiatric disorders.
3 Magnetic resonance imaging (MRI) studies provide evidence for brain structural abnormalities in
4 rodents following prenatal exposure to MIS. Reported volumetric changes in adult MIS offspring
5 comprise among others larger ventricular volumes, consistent with alterations found in patients with
6 schizophrenia. Linking rodent models of MIS with non-invasive small animal neuroimaging modalities
7 thus represents a powerful tool for the investigation of structural endophenotypes. Traditionally manual
8 segmentation of regions-of-interest, which is laborious and prone to low intra- and inter-rater reliability,
9 was employed for data analysis. Recently automated analysis platforms in rodent disease models are
10 emerging. However, none of these has been found to reliably detect ventricular volume changes in MIS
11 nor directly compared manual and automated data analysis strategies. The present study was thus
12 conducted to establish an automated, structural analysis method focused on lateral ventricle
13 segmentation. It was applied to ex-vivo rat brain MRI scans. Performance was validated for phenotype
14 induction following MIS and preventive treatment data and compared to manual segmentation. In
15 conclusion, we present an automated analysis platform to investigate ventricular volume alterations in
16 rodent models thereby encouraging their preclinical use in the search for new urgently needed
17 treatments.

18

19 **Keywords:** small animal magnetic resonance imaging, lateral ventricle, maternal immune stimulation

20

21 **Introduction**

22 Exploring new avenues in psychiatric research has the goal of combining treatment effectiveness
23 with tolerability, often with the idea of a preventive treatment strategy. One condition in order to achieve
24 this high-set goal is the availability of cross-species biomarkers. Good parameters should be easily
25 measured, reliably report disease manifestation in patients and inform preclinical treatment studies in
26 disease relevant animal models. Discovering biomarkers has proven to be very challenging for many
27 psychiatric diseases, due to the variability of symptoms and their often late onset. Most psychiatric
28 diseases, like schizophrenia, are now considered neurodevelopmental, which means that the
29 pathological alterations occur during development and are present before the symptoms develop. These
30 pathological alterations can for example be seen in morphological changes in the brain, e.g. divergent
31 volume of brain regions. One of these locations are the lateral ventricles, which are altered in a number
32 of psychiatric or neurological afflictions: for example in ischemic stroke patients (1), amyotrophic
33 lateral sclerosis (2), cystic periventricular leukomalacia (3), Alzheimer's disease (4), Parkinson's
34 disease (5) and schizophrenia (6,7). Similar changes in lateral ventricle volumes have been observed in
35 a number of established animal models with relevance to schizophrenia (8–10) as well as other
36 psychiatric or neurological diseases (11–15).

37 Brain volumes can be easily measured with magnetic resonance imaging (MRI), a non-invasive
38 imaging method available with a good soft-tissue contrast and good spatial resolution (50 - 100 μm)
39 (16,17). It utilizes longitudinal magnetization signals to create contrast images whose intensity range
40 indicate structural boundaries and tissue properties in order to further make informed decisions. To
41 quantify structural alterations due to neuropsychiatric diseases, MR images are acquired and
42 morphometry is performed. Several morphometric methods have been established, including voxel
43 based morphometry (VBM) (18) to correlate brain structure or shape differences within a population,
44 cortical thickness measurement (19,20), statistical shape analysis (21) and structural volume analysis
45 (22) with labeling of anatomical structures. For the process of morphometric analyzing the structural
46 MR images for volume, shape and/or thickness, segmentation of the volume images is typically carried
47 out. Segmentation is a process of classification of image units (pixels or voxels) into subsets based on

48 predefined criteria. The manual segmentation of the rodent's brain is often still the standard for
49 segmentation of small animal structural images (23). This method is very laborious, time consuming
50 and subjected to intra- and inter-subject variability. In the search of urgently needed new treatment
51 strategies however, there is a high demand for reliable automated processing alternatives (24,25).

52 For automated morphometric analysis of the rodent brain it is important to accurately determine
53 the anatomical structural region's parcellation. Any established method must be reliable and allow
54 repetitiveness. A method that fits this need is the atlas-based segmentation. The automated atlas-based
55 segmentation of volumetric images is based on image registrations and label propagation. It involves
56 the registration (linear and diffeomorphic) between a reference brain image to individuals of a query set
57 (or vice versa). The resulting registered brain represents the maximum similarity between the individual
58 image and the corresponding reference image. The anatomical segmentation or label of the reference
59 brain is further propagated to the individual brain sets based on the transformations and deformation
60 fields from the registration step. Registration techniques, variation between animals' anatomical
61 structures and correctness of label segmentation are critical factors for the automated segmentation of
62 the rodent brain using an atlas-based technique. More recently, some more automated workflows have
63 been implemented (26,27). These approaches are less labor-intensive and time-consuming; however,
64 they cannot be directly applied to our data depending on differences between species (mouse vs rat),
65 strains, gender or age factors. In addition, especially segmentation of lateral ventricles has been proven
66 difficult (26).

67 Thus, we aimed to establish an automated analysis platform with a focus on lateral ventricle
68 delineation. For this we chose a maternal immune stimulation (MIS) rat model for schizophrenia, which
69 is based on the finding that maternal prenatal infection increases the risk of developing schizophrenia
70 (Brown et al, 2004). Exposing pregnant rodents to the viral mimic polyriboinosinic polyribocytidilic
71 acid (poly I:C) is a commonly used neurodevelopmental approach to model schizophrenia (8–10,28,29).
72 The onset of behavioral abnormalities is seen during adolescence, similar to the clinical picture in
73 humans (10). However, the neuropathological alterations have been detected at a different time point
74 with the enlargement of lateral ventricles occurring in adulthood (30). This allows studies of preventive

75 treatment approaches. In line, we have previously reported that anodal transcranial direct current
76 stimulation (tDCS) has been shown to prevent the emergence of behavioral alterations and the
77 enlargement of lateral ventricles in the MIS animal model (8). Here, we report an analysis platform
78 construction optimized for lateral ventricle volume estimation in adult male offspring following prenatal
79 MIS induction. In addition, MR images were segmented manually and obtained results compared
80 between methods. Finally, available imaging data from our prior tDCS treatment study was employed
81 for verification of platform performance and data robustness.

82 **Materials and methods**

83 **Animals**

84 Experiments were performed according to the guidelines of the European Union Council
85 Directive 2010/63/EU for care of laboratory animals approved by the local ethic committee
86 (Landesdirektion Sachsen, Dresden, Germany). Animals were housed in a temperature and humidity-
87 controlled vivarium under a 12 h light / 12 h dark standard day cycle with food and water ad libitum.
88 Animal suffering and numbers were kept to a minimum. Female pregnant Wistar rats (Charles River
89 Laboratories, Europe) were received at gestational day (GD) 15, housed in individual cages with nesting
90 material and left to acclimatize for 2 hours. Dams were carefully anesthetized with isoflurane and the
91 viral analogue poly I:C, (4 mg/kg; Sigma, Germany) dissolved in saline, or 0.9% saline alone was
92 injected through the tail vein (volume: 100 μ l/100 gr bodyweight) (31,32). Dams were weighed daily
93 until delivery and then left undisturbed until postnatal day (PND) 21, when offspring was weaned,
94 group-housed with 2-4 animals in standard cages with bedding and shelter (dark PVC tubes). Adult
95 (>PND90) male offspring, n=21 from saline and n=20 from poly I:C injected mothers, were used for the
96 study.

97 Second, imaging data from a previous published study (8) was utilized. The study was
98 conducted to assess the preventive treatment potential of non-invasive transcranial direct current
99 stimulation (tDCS) during adolescence, prior to schizophrenia-relevant behavioral manifestation, in the
100 same MIS model of schizophrenia. The study followed a 2 \times 3 design with phenotype (saline, poly) \times

101 treatment (sham, cathodal, anodal), for details refer to Hadar et al., 2019. Groups: saline-sham (ss) n =
102 15, poly-sham (ps) n = 14, saline-cathodal (sc) n = 13, poly-cathodal (pc) n = 11, saline-anodal (sa) n =
103 11 and poly-anodal (pa) n = 12.

104 **MRI**

105 **Sample preparation**

106 For postmortem assessments the rats were deeply anaesthetized with a single i.p. injection of
107 pentobarbital (60 mg/kg) and perfused transcardially with cold 4% paraformaldehyde in 0.1 M
108 phosphate buffer, pH 7.4. Brains remained within the skull and were imaged within 1-2 weeks following
109 preparation.

110 **Acquisition**

111 MRI acquisitions were performed on a 7 Tesla rodent scanner (Bruker BioSpin MRI GmbH,
112 Ettlingen, Germany) at Charité (Berlin, Germany) or HZDR (Dresden-Rossendorf, Germany). The
113 acquisition protocol consisted of a multi slice localizer (field of view (FOV) 50 × 50 mm) and T2-
114 weighted contrast images with a rapid acquisition with relaxation enhancement (RARE) sequence
115 (imaging parameters: TR/TE = 4050/30 ms, RARE factor 8, NEX 6, FOV 30 × 30 mm, MD 256 × 256),
116 resulting in an inplane resolution of 117 μm with a slice thickness of 0.5 mm and 42 slices. Scanning
117 time was 13 minutes per brain.

118 **Manual segmentation**

119 The axial view was used for manual segmentation. Original acquired images were resliced with
120 a slice thickness of 0.12 mm. 15 slices from -0.1mm (first view of the anterior part of the anterior
121 commissure) to +0.7mm from Bregma were analyzed for each animal (33). For best image quality,
122 contrast was adjusted using MRICron. The lateral ventricles were outlined using ImageJ and special
123 attention was paid on distinguishing ventricular area from adjoining areas through identifying
124 differences in contrast by eye. Manual segmentation took up to 120 minutes per animal, as both left and
125 right lateral ventricle were separately measured through each slice and discernable contrasts were often
126 minimal, which required extensive repeated examination. Two independent raters carried out manual

127 segmentation. Volumes were obtained by multiplying the summed-up amount of pixels times pixel area
128 with slice thickness (ventricular volume = (sum (measured amount of pixels \times 0.117mm²)) \times 0.12mm \times
129 0.01). Analysis were run on means from the two independent observations per animal.

130 **Automated analysis platform**

131 The DICOM (Digital Imaging and Communications in Medicine) MR image data were
132 collected, converted into the Neuroimaging Informatics Technology Initiative (NIFTI) file format and
133 utilized for the platform's framework and evaluation. To provide computational efficiency the automated
134 analysis platform was based on the Advanced Normalization Tools (ANTs) (34), a toolkit for medical
135 image registration and segmentation and was implemented with a high-performance computing (HPC)
136 setup (see S1 Appendix for details). The structure of the automated analysis platform is shown in Fig 1.

137

138 **Fig 1. Automated analysis platform overview** (a) Flowchart with examples of b) bias correction
139 including intensity normalisation followed by field correction for inhomogeneity using the
140 N4BiasFieldCorrection (64). c) Brain extraction with registration-based skull stripping to remove non
141 brain tissues using a mask and the skullstripper tool (65). d) Atlas Creation: Iterative creation of the
142 within study template using the minimum deformation pairwise registration resulted in a study template,
143 an averaged image of selected control subjects d' with its corresponding manually delineated labels and
144 d'' regions of interest (ROI) for studies in the maternal immune activation model.

145

146 Pre-processing steps included intensity reorientation, intensity bias correction and brain
147 extraction. To achieve spatial consistency images were reoriented into standard anatomical space
148 (patient coordinate) based on the ITK-SNAP (35) employing the *orient* algorithm of the *Convert3D*
149 medical image processing tool. Hence data sets with an initial orientation LPI (Left, Posterior, Inferior),
150 common for DICOM images i.e. an image vector were converted by a linear transformation to RIP
151 (Right, Inferior, Posterior).

152 The N4BiasFieldCorrection algorithm (N4ITK), an optimized version of the N3 that
153 implements a robust B-spline approximation routine and a modified optimization scheme (36), was used

154 to correct intensity non-uniformity within images. 50 iterations over three levels with a convergence
155 threshold of 1e-6, full width at half maximum (deconvolution kernel) of 0.15mm were employed with
156 default parameters to provide optimum results for the rat subject MR images. For brain extraction the
157 python command line *SkullStrip* (37) was used for the semi-automated registration brain extraction in this
158 project. A reference stripped image with its brain mask (manually delineated) and the brain image was
159 used to generate a stripped fixed image mask that was then binarized. Image multiplication operation
160 was then performed with this mask and the original subject image (see S1 Appendix for details).

161 Post-processing steps included the segmentation of each subject, i.e. the image registration,
162 normalization, label propagation and volume extraction. For the segmentation of each subject the atlas-
163 based method was implemented using a within study created atlas from four control subjects (38). The
164 *antsMultivariateTemplateConstruction.sh* script(39) of the ANTs tool was utilized for template creation.
165 An iteration of four was employed with the greedy symmetric normalization (SyN) transformation with
166 a cross correlation similarity metric (39,40). The anatomical image of the template was manually
167 delineated into thirty-five regions using the ITK-SNAP. The standard rat brain atlas (33) was employed
168 for anatomical referencing and nomenclature. The left and right hemispheres were considered as one
169 unit. The ependymal and subependymal layers were considered the boundaries of the lateral ventricles
170 (LV). Similarly, the periventricular hypothalamic region was classified with the 3rd ventricles.

171 The *antsRegistration* command (41) of the ANTs was employed for image registration. The
172 cross-correlation (CC) similarity metric was used for diffeomorphic registration and the mutual
173 information during linear registration (26,39,42) between the fixed and moving images. The symmetric
174 normalization (SyN) (42) was the optimizer (image normalization method) algorithm used during
175 deformation with the CC similarity (39). The *antsApplyTransforms* command of the ANTs tool was
176 employed for label propagation of all subjects with a nearest neighbor interpolation scheme. From the
177 automated segmentation description using an atlas-based segmentation technique, the quality of
178 segmentation of the output is directly proportional to the performance of registration (43) and
179 interpolation scheme (see S1 Appendix for details). The volume of each structure of the label was
180 extracted using the *LabelStats* algorithm of ANTs and a MATLAB script.

181 **Statistics**

182 One way ANOVA was used to determine significant differences between phenotype (saline or
183 poly) for the lateral ventricle volumes measured through manual segmentation and the automated
184 analysis platform, respectively. To check for significant interactions between phenotype (saline or poly)
185 and treatment (saline, anodal or cathodal), two-way ANOVA was completed followed by posthoc
186 analysis with Sidak correction. Statistical significance was set at $p < 0.05$. The software SPSS was used
187 for statistical analysis of the data.

188 **Results**

189 MR images from 42 animals (n=21 saline, n=20 poly) were analyzed using the established
190 analysis platform, labels were visually quality controlled but not corrected and ventricular volumes
191 estimated. As expected, estimates were observed to be lower in saline $4.198 \pm 0.247 \text{ mm}^3$ compared to
192 poly I:C $4.434 \pm 0.279 \text{ mm}^3$ offspring, respectively (Fig. 2a,b). The difference was found to be
193 significant $T(39) = -2.858$ $p = .007$ (Fig 2c). Due to the enormous labor intensity only half of the sample
194 was also evaluated manually (n=12 saline, n=12 poly; for example of manual segmented image see S1
195 Appendix). Analysis showed similarly lower estimations of LV volumes with $3.981 \pm 0.463 \text{ mm}^3$ and
196 $4.293 \pm 0.383 \text{ mm}^3$ in saline compared to poly I:C offspring which did not reach significance $T(22) = -$
197 1.794 $p = .087$ (Fig. 2d). Reduced samples size and variance between samples in manual estimates,
198 which is two times higher compared to platform measures, can be taken in consideration, however
199 automated analysis platform data from an identically reduced sample remains significant $T(22) = -2.481$
200 $p = .023$. Thus both evaluation methods yield similar phenotype differences while the correlation
201 between measures is small $r(22) = .249$, $p = .032$.

202

203 **Fig 2. Lateral ventricle (LV) phenotype in rat MIS model.** a,b) Examples of LV delineated from a
204 saline a) and poly (b) subject by the automated analysis platform. Boxplot of the lateral ventricular
205 volume from poly I:C and saline injected offspring following c) automated and d) manual delineation
206 methods. Center lines show the medians; box limits indicate the 25th and 75th percentiles as determined

207 by R software; whiskers extend 1.5 times the interquartile range from the 25th and 75th percentiles,
208 individual data points included.

209

210 For further performance validation, imaging data from (8), which was published employing
211 manual delineation, was now re-analyzed with the newly established automated analysis platform.
212 Following visual quality control, ventricle labels were manually corrected for 15% of subjects and LV
213 volume statistics obtained (Fig. 3). A two way ANOVA yielded a significant effect for the
214 phenotype*treatment interaction ($F(5,68) = 8.626$, $p < .001$) with no main effects of phenotype or
215 treatment ($p > .05$). Posthoc analysis showed the expected, significant phenotype difference following
216 MIS between the saline control (ss) and the untreated poly I:C (ps) group $p = .002$, mean ventricular
217 volume ss: $3.97 \pm 0.25 \text{ mm}^3$ and ps: $4.28 \pm 0.21 \text{ mm}^3$. Similarly, as measured and statistically evaluated
218 in the Hadar et al., 2019 paper, treatment was found to reduce LV volume following MIS when
219 compared to ps animals for cathodal $p = .004$ and anodal $p = .006$ stimulation, respectively. Mean
220 ventricular volume for pc: $3.98 \pm 0.18 \text{ mm}^3$ and pa: $4.00 \pm 0.30 \text{ mm}^3$ groups. Finally, also differences
221 between sa and pa were replicated $p = .037$, mean ventricular volume for sa: $4.2 \pm 0.09 \text{ mm}^3$.

222

223 **Fig. 3 Performance evaluation on treatment data.** Lateral ventricle volume estimation in rat MIS
224 model from a study of preventive treatment with transcranial direct current stimulation (8) shows
225 validation of prior reported findings using automated platform method. Center lines show the medians;
226 box limits indicate the 25th and 75th percentiles as determined by R software; whiskers extend 1.5 times
227 the interquartile range from the 25th and 75th percentiles, individual data points included.

228

229 Discussion

230 The present work describes the implementation of a single atlas-based segmentation framework
231 for adult rat brain analysis, optimized for lateral ventricle segmentation. Volume estimation with this
232 automated framework was validated against the results of a standard manual method. The output of the

233 analysis platform was further used to estimate volumetric changes caused by neuropsychiatric disease
234 and the potential therapeutic treatments in the MIS rat model of schizophrenia on ex-vivo MRI data.

235 The enlargement of the lateral ventricles is consistently observed in patients with schizophrenia
236 and relevant established animal models. Such alterations are thought to originate from abnormal prenatal
237 brain development. This association has been especially well documented through the study of rodents
238 following prenatal MIS. Notably, this phenotype difference was observed in both of our independent
239 sample analysis, and with both manual and automated analysis.

240 In schizophrenia patients further structural brain abnormalities have been detected in in a
241 number of brain regions such as the striatum (44,45), the hippocampus and other medial temporal lobe
242 structures (44,46,47), as well as the cerebellum (48–50), and progressive gray matter (GM) loss is
243 present in parietal, prefrontal and superior temporal cortices (51–54). While we have not investigated
244 this further, recent semi-automated segmentation methods have already shown promising results in
245 estimating changes in striatal, hippocampal and cortical structures in rodent models (Crum et al., 2017).

246 The use of rodent models in general provides an advantage for the evaluation of temporal
247 changes in disease and repeatability at a systemic level. Animals of different strains combined with
248 several imaging techniques can provide insight regarding behavioral pharmacology, potential therapies,
249 longitudinal studies of diseases in order to measure changes, treatment efficacy and other therapeutic
250 interventions (17,55). Therefore, our results further strengthen the growing field of neuroimaging
251 techniques in preclinical animal research, for which investigations and deductions can be translated
252 towards the respective human conditions (56,57).

253 Our analysis included ex-vivo MRI data from a study of tDCS application to prevent
254 manifestation of schizophrenia associated deficits in MIA rats (8). tDCS is a brain stimulation method
255 where low electric positive (anodal) or negative (cathodal) current is applied to an area of the brain for
256 the depolarization or hyperpolarization of neurons towards facilitatory or inhibitory behavioral effects
257 respectively (58). Medial prefrontal cortex (mPFC) stimulation, based on established protocols (59) and
258 applied during adolescence, was reported to restrict the emergence of schizophrenia-related behaviors
259 as well as volumetric changes. The previously, manually segmented data sets were now reanalyzed to

260 confirm phenotype and treatment effects at adulthood. We were able to replicate the results with our
261 automated analysis platform, showing a significant difference in lateral ventricle volumes with lower
262 volumes in saline offspring compared to poly I:C offspring. In addition, the automated analysis platform
263 confirmed lower lateral ventricle sizes for poly I:C offspring treated with anodal stimulation but not for
264 poly I:C offspring treated with cathodal stimulation.

265 Manual segmentation is still frequently used to achieve volumetric measures from neuroimaging
266 techniques due to its robustness, although it has certain disadvantages. It is very time consuming, taking
267 2-3 hours per animal for a practiced expert, and therefore labor-intensive. Variance in the results is high,
268 with inter- and intra-rater biases, and the need for a better alternative is formidable. The automated
269 method as shown in this paper is accurate, allows repetitiveness and is much less time-consuming.
270 However, automated segmentation still comes with various challenges. They include often low image
271 resolution of rodent brains (smaller voxel sizes in rodents compared to humans due to their brain size),
272 physiological noise and signal-to-noise ratio (57,60). However, animal MRI scanners have higher field
273 strengths (7 - 16.4 T) than human scanners (1.5, 3T), providing better contrasts to distinguish brain
274 structures. The quality of MR image contrast influences the quality of extraction, introducing a bias by
275 raters during delineation for low quality images. It should be noted that for well contrasted images, MRI
276 acquisition requires a longer scanning time and large image outputs influence calculations during image
277 analysis. Alternatively, image contrast agents can be used during scanning to enhance the quality of
278 images while compensating for scanning time (60,61).

279 In addition, segmentation of small regions of interest can be faulted by introducing a systematic
280 bias, which increases as the region of interest gets smaller (26,62). This has been noted especially for
281 the case of lateral ventricle segmentation. It has also been noted to apply even more so when earlier
282 timepoints are investigated as the brains and corresponding ROIs are much smaller. This systematic bias
283 is inherent in automated segmentation, demanding a critical and cautious adjustment of automated
284 processes to the data at hand. Several steps have been taken in the development of this analysis platform
285 and the preparation of data to reduce a potential systematic bias. In the used registration method, a brain
286 atlas with its corresponding brain mask is used for brain extraction. Therefore, the brain extraction is

287 dependent on the availability of an atlas, its mask and the parameters of registration. The rats' size, age
288 and the similarity between the reference atlas image and the target image of the dataset must be
289 considered. To aid this, a reference subject can be chosen from the study cohort, manually segmented
290 and used for the automated registration brain extraction methods. In our approach, the *SkullStrip* (37)
291 command tool, a robust automated brain extraction tool, was used, which is dependent on the image
292 resolution. This tool was further optimized by binarizing the skull stripped output for each subject and
293 applying it to the original image to get the best "brain-only" volume image for each subject. These
294 additional steps resulted in a higher correlation in visual inspection, total brain volume and similarity
295 measure to the manual method. These efforts may be partially responsible for achieving results in line
296 with previous work on characterizing volumetric changes in lateral ventricles with the presented
297 platform contrary to the findings of other semi-automated segmentation approaches (26).

298 Limitations of the presented analysis platform include the still labor-intensive manual extracting
299 of the reference subject and its brain mask used with the optimized skull-strip method. This step, though
300 advisable to remove bias resulting from differences in strain and age when needed, is only carried out
301 once. A within-study template is highly recommended. Second the image contrast, significant for brain
302 segmentation was low in our data. There is a need for a standard segmentation protocol detailing the
303 process to delineate rodent brains for imaging modalities other than histology. Anderson et al. (63)
304 described a validation of registration parameter choices using high-performance computer clusters. They
305 indicated the computational cost-benefits and temporal performance with a voxel-based analysis. With
306 several images to be analyzed in preclinical studies and the differences observed in between registration
307 metrics there is still a strong need for a comprehensive evaluation framework to inform or guide
308 parameter choices in rodent image analysis. Finally, this automated analysis platform was optimized
309 only for the segmentation of ventricular volumes. Label propagation utilizes the transformation matrix
310 and output of the registration, so multiple labels can be propagated to measure changes in the individual
311 subject. These labels can contain different numbers of structures as well. In the future the presented
312 analysis platform should thus be extended to the measurement of other clinically relevant brain
313 alterations in rodent models.

314 **Conclusion**

315 The characterization of preclinical rodent models of neurological and psychiatric diseases with
316 several imaging modalities is challenging with an increased need for accuracy, reputability, efficiency
317 and reduced human effort except for quality control in the studies. This project thus focused on
318 implementing an automated single atlas-based segmentation framework for rat brains and validate its
319 performance against the results of the standard manual method. The analysis platform integrated several
320 pre-processing and post-processing steps including brain extraction, intensity non-uniformity bias
321 correction, linear and non-linear registrations and label propagation. The resulting analysis platform was
322 used exemplarily with ex-vivo MR images of a MIS schizophrenia rat model. It successfully detected
323 lateral ventricle volumetric changes for phenotypes in independent samples and following treatments,
324 establishing a basis for utilizing anatomical biomarkers in the rat to investigate disease progression or
325 treatment efficiency.

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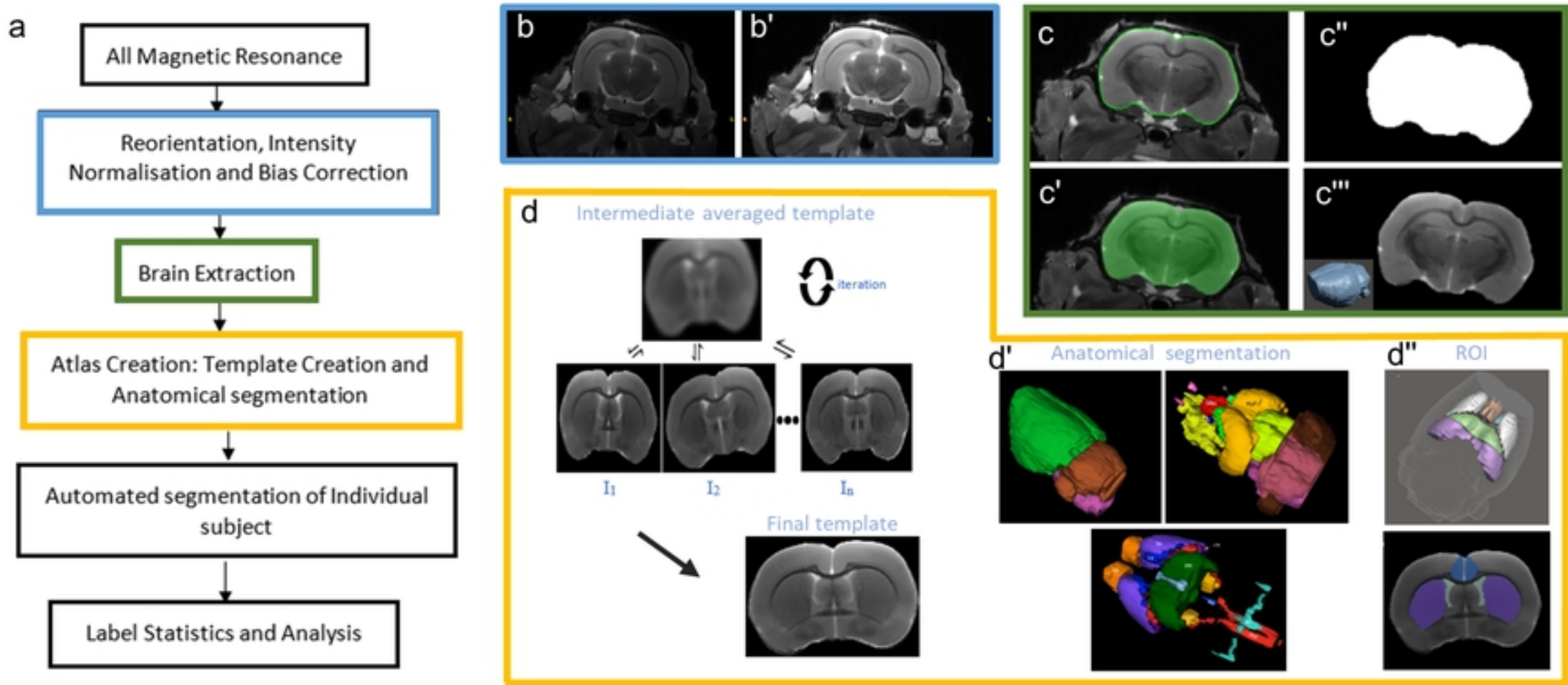
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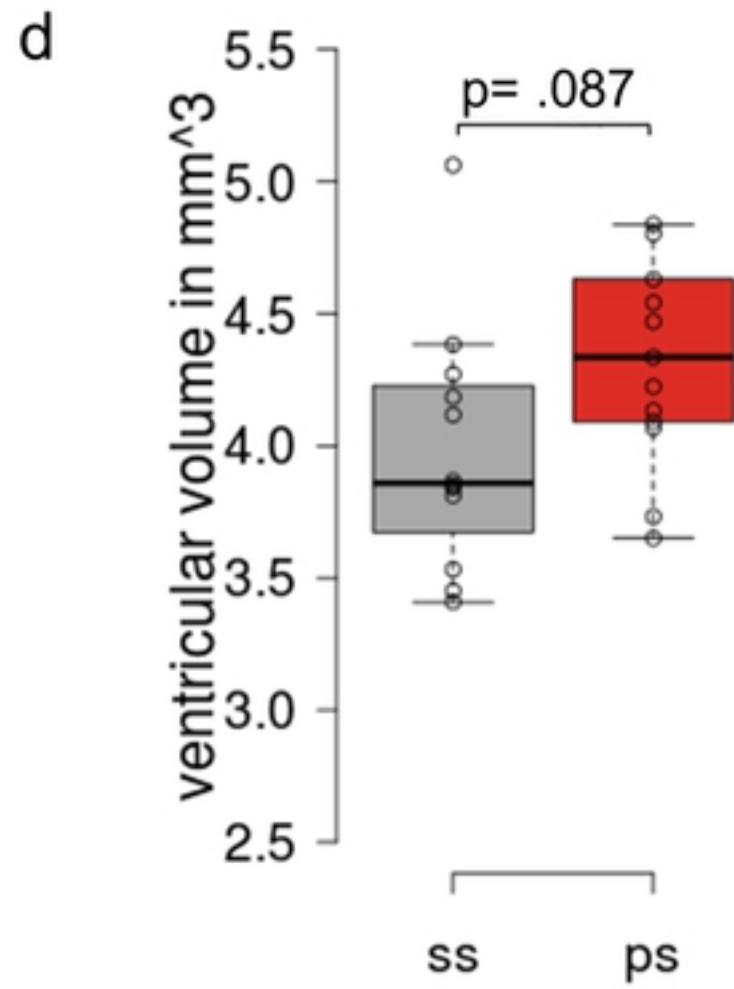
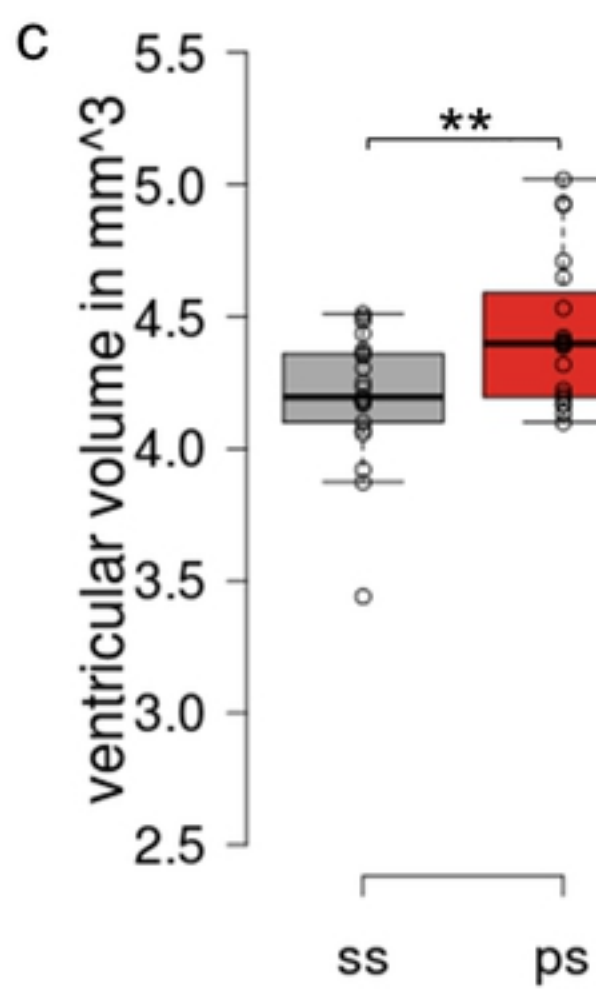
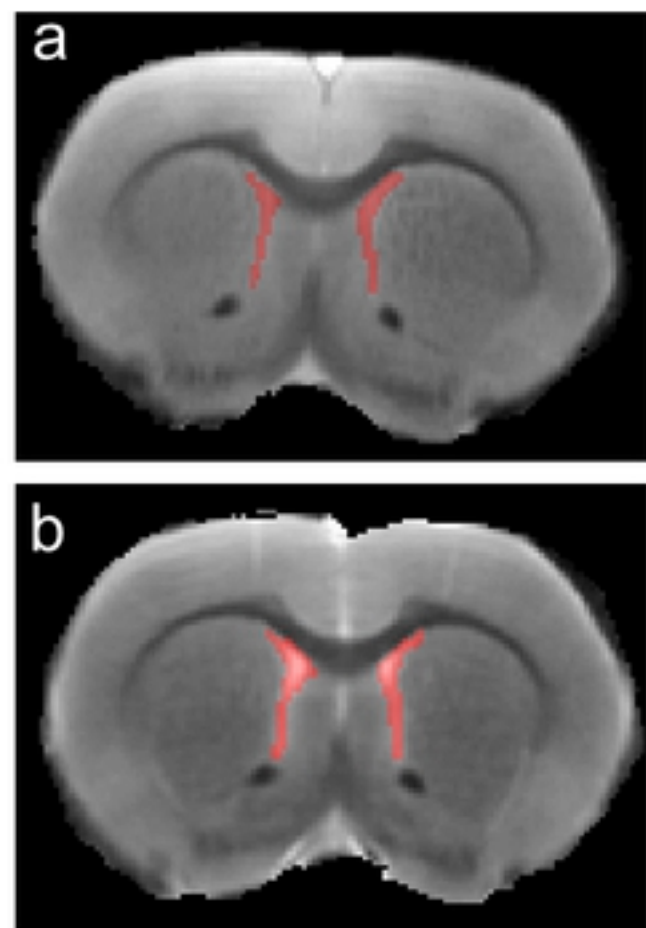
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511 **Supporting information**

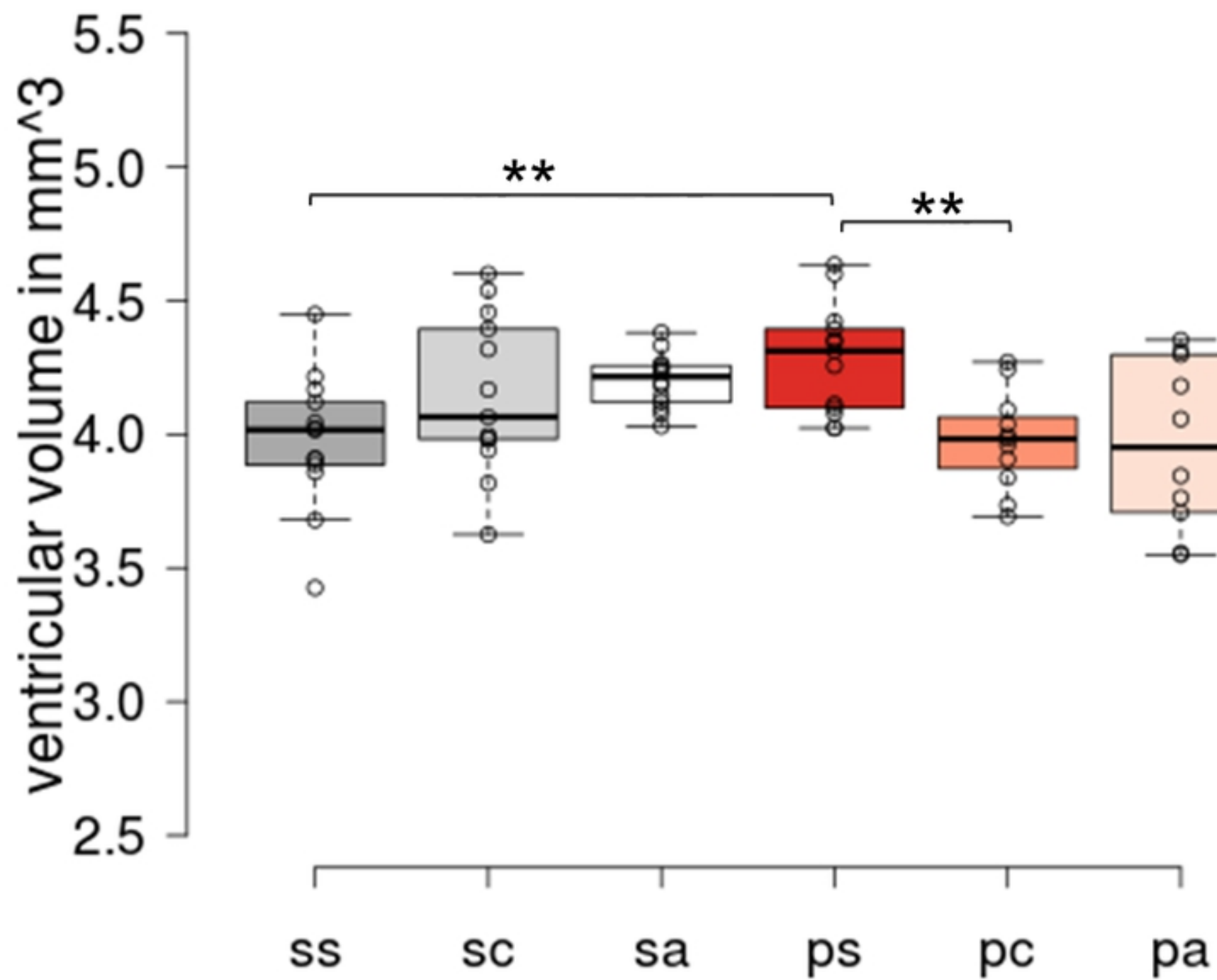
512 **S1 Appendix. Supplement**



Figure



Figure



Figure