Intraspecific diversity loss in a predator species alters prey community structure and

ecosystem functions

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Abstract

Loss in intraspecific diversity can alter ecosystem functions, but the underlying mechanisms are still elusive, and intraspecific biodiversity-ecosystem function relationships (iBEFs) have been restrained to primary producers. Here, we manipulated genetic and functional richness of a fish consumer (*Phoxinus phoxinus*), to test whether iBEFs exist in consumer species, and whether they are more likely sustained by genetic or functional richness. We found that both genotypic and functional richness affected ecosystem functioning, either independently or in interaction. Loss in genotypic richness reduced benthic invertebrate diversity consistently across functional richness treatments, whereas it reduced zooplankton diversity only when functional richness was high. Finally, both losses in genotypic and functional richness altered essential functions (e.g. decomposition) through trophic cascades. We concluded that iBEFs lead to substantial top-down effects on entire food chains. The loss of genotypic richness was similarly prejudicial than the loss of functional richness, probably because it sustains "cryptic" functional diversity.

Introduction

Human disturbances associated with global change are increasingly altering worldwide patterns of species diversity, as well the functions and services provided by ecosystems (Parmesan 2006; Naeem *et al.* 2009; Isbell *et al.* 2017). Nonetheless changes observed at the species and ecosystem levels are always preceded by changes in phenotypic and genotypic composition within plant and animal populations (Clements & Ozgul 2016; Clements *et al.* 2017; Baruah *et al.* 2019). Accordingly, extremely rapid changes in intraspecific diversity are currently occurring worldwide (Darimont *et al.* 2009; Mimura *et al.* 2016; Miraldo *et al.* 2016; Leigh *et al.* 2019). Changes in intraspecific diversity can affect species turnover and composition (Vellend 2005; Vellend & Geber 2005), as well as ecosystem functioning (Des Roches *et al.* 2018; Raffard *et al.* 2019c). For instance, the loss of genotypes within primary producers can reduce ecosystem process rates and species diversity (Crutsinger *et al.* 2006; Johnson *et al.* 2006; Fridley & Grime 2010), suggesting the existence of biodiversity-ecosystem functioning relationships at the intraspecific level (iBEFs, Hughes *et al.* 2008; Koricheva & Hayes 2018; Raffard *et al.* 2019c), in addition to the more widely studied BEFs at the interspecific level.

The relationships between intraspecific diversity and ecosystem functioning are challenging to study and this is primarily because of the confusion between genetic and functional trait diversity. iBEF studies have initially manipulated genotypic richness within experimental populations (e.g. Crutsinger *et al.* 2006; Hughes & Stachowicz 2009; Abbott *et al.* 2017). Genetic variability is expected to encapsulate a large proportion of trait variability, and thus higher genotypic richness should maintain higher functional diversity (Hughes et al. 2008). Although this approach allows deciphering the general effects of intraspecific diversity on ecosystem functioning, it does not provide mechanisms. As a consequence, parallel studies have directly manipulated functional trait richness within populations, which enabled to

determine a direct mechanistic linkage between functional richness and community structure

and ecosystem functioning (Violle *et al.* 2007; Matthews *et al.* 2011). For instance, individual

body mass (and traits covarying with body mass) has strong ecological effects because of the

associated functional differences and resource use complementarity among individuals

(Woodward *et al.* 2005; Hildrew *et al.* 2007; Rudolf *et al.* 2014; Harmon *et al.* 2009; Rudolf

& Rasmussen 2013a). Focusing on specific traits, such as body mass, might conversely blur

the ecological effects of "cryptic" trait variation (i.e. unmeasured functional traits) that is

likely supported by genotypic richness. We therefore argue that manipulating simultaneously

genotypic richness and the diversity of key functional traits such as body mass should allow

assessing whether cryptic trait diversity is ecologically important, and a better mechanistic

understanding of iBEFs.

The ecological effects of biodiversity changes are particularly strong when the later

occur at high trophic levels such as secondary consumers or predators. Changes in the

diversity and abundance of predatory species can trigger important effects in functions

supported by lower trophic levels, especially in regulating the abundance of the lower trophic

levels and indirectly ecosystem functioning, such as biomass production (Shurin *et al.* 2002;

Barnes *et al.* 2018). High predator species richness can sometimes favour resource use

complementarity and decrease prey abundance (Cardinale *et al.* 2006; Griffin *et al.* 2013). In

some cases, however, increasing predator richness increases prey abundance through

mechanisms such as predator interference (Sih *et al.* 1998; O'Connor & Bruno 2009), and can

modify multiple ecosystem functions along food webs (Antiqueira *et al.* 2018; Barnes *et al.*

2018). These mechanisms have all been investigated at the interspecific level, but they may

also apply *within* a predatory species (intraspecific diversity), and we can expect relationships

–either positive or negative- between predator intraspecific diversity and the structure of prey

communities, that could subsequently cascade down essential ecosystem functions such as

decomposition rate or primary production (Duffy, 2002; Bruno & O'Connor, 2005).

However, it is still difficult to forecast how loss in intraspecific diversity in predatory (and

consumer) species could affect ecosystem functioning, because iBEFs studies have primarily

focused on primary producers (Koricheva & Hayes 2018; Raffard *et al.* 2019c). This is

despite the fact that human activities strongly affect predator and consumer populations, for

instance through harvest or fisheries activities, which may alter ecosystem functioning

through intraspecific changes in traits and genotypes (Hauser *et al.* 2002; Estes *et al.* 2011;

Palkovacs *et al.* 2012, 2018).

In this study, we investigated whether a loss in genotypic and functional diversity

within a consumer species at the top of a three-level trophic chain could mediate top-down

effects on key ecosystem functions. In a nine-months mesocosm experiment, we

simultaneously manipulated *genotypic richness* (number of genetic entities) and the *functional*

richness (variance in individuals body mass) of experimental populations of a freshwater fish,

the European minnow (*Phoxinus phoxinus*); a common and abundant species with important

ecosystems effects (Miró *et al.* 2018; Raffard *et al.* 2019b). We predicted that increased

functional richness should affect ecological functions; if functional richness captures the

entire functional differences among genotypes, then increasing genotypic richness should not

impact ecological functions further (Fig. 1a). At the opposite, if functional richness does not

capture all the functional differences among genotypes, then increasing genotypic richness

should increase functional diversity and affect ecological functions (Fig. 1b). The ecological

effects of genotypic and functional richness might display different shape following different

predictions based on possible "additive", "enhancing" or "compensatory" effects (Fig. 1), to

which we compared our experimental findings. Finally, we identified the mechanistic bases of

the ecological effects of genotypic and functional richness. We specifically tested whether the

loss in genotypic and functional richness affected directly populations' biomass production

and community structure, and indirectly ecosystem functioning mediated by changes in

community structure of benthic and pelagic food web.

Material and methods

Model species

The European minnow (*P. phoxinus*) is a common species occurring across Western Europe,

living in relatively cold water (summer water temperature generally lower than 24°C)

including mountains lakes, small rivers at intermediate altitude and mountain streams (Keith

et al. 2011). It is a small-bodied cyprinid fish species (<12 cm long, 4-8 cm long as an adult

in general) playing the role of secondary consumer in most ecosystems (Raffard *et al.* 2020),

with a generalist diet composed of small invertebrates, algae or zooplankton (Frost 1943;

Collin & Fumagalli 2011). Populations of European minnows differed in their genetic and

phenotypic richness (Fourtune *et al.* 2018), and previous works revealed that genetically and

phenotypically unique populations differently affect prey community abundance and

ecosystem functions (Raffard *et al.* 2019b).

We selected ten populations from a large river basin in southern France (the Garonne

catchment) based on *a priori* knowledge to maximise both genetic and functional

differentiations among populations (Fourtune *et al.* 2018; Raffard *et al.* 2019a) (Fig. S1).

Specifically, we sampled five populations mainly composed of small-bodied adults (mean

body mass ± standard error (SE) = 1.03 g ± 0.02) and five populations mainly composed of

large-bodied adults (mean body mass ± SE = 3.06 g ± 0.07) (Fig. S2a). These differences in

adult body mass are due to selective pressures from the local environment such as mean water

temperature; the higher the mean water temperature, the smallest the adult body mass due to

increased metabolic rate and an accelerated pace-of-life (Raffard *et al.* 2019a). These

populations also display unique genetic characteristic and can each be defined as unique

genetic entities (Fourtune *et al.* 2018; Prunier *et al.* 2019). Each population are geographically

distant, and they will hereafter be each considered as functionally and genetically unique

entity, and intraspecific richness will be manipulated by varying the number of populations in

independent mesocosms (see details below).

In September 2017, individuals were collected in each river by electrofishing a 200-m

section and visually selecting 30-50 individuals reflecting the size range of adults. Fish were

maintained at similar density in outdoor tanks for 3 weeks before the start of the experiment.

Fish were fed ad libitum with frozen Chironomidae during this period.

Mesocosm experiment

In October 2016, 24 outdoor mesocosms were filled with 900 L of water, and 3 cm of gravel.

Nutrients were added to the mesocosms using 10 mL of solution containing nitrogen and

phosphorus (ratio N:P:K = 3.3:1.1:5.8). Each mesocosm was then inoculated twice (October

2016 and May 2017) with 200 mL of a concentrated solution of phytoplankton and 200 mL of

concentrated solution of zooplankton collected from a unique lake nearby (Lamartine Lake,

France 43°30'21.5"N, 1°20'32.7"E). In May 2017, we introduced three adult pond-snails

(Lymnaeidae) and ten adult isopods (Asellidae) in each mesocosms. They were let uncovered

to allow natural colonization by other invertebrates and community assemblage until the start

of the experiment that occurred approximately 6 months later.

In October 2017, eight fish were weighted and introduced in each mesocosm, which

were assigned to one of four treatments according to a full-factorial design with *genotypic*

richness (two levels, *high* and *low* genotypic richness) and *functional* richness (two levels,

high and *low* functional richness) as the main factors. Genotypic richness was manipulated by

introducing individuals sourced from either two (*low genotypic richness*, four individuals

from each population) or four (*high genotypic richness*, two individuals from each

population) out of the ten populations. These two levels of genotypic richness were selected

as it has previously been shown in a meta-analysis that the effect of intraspecific richness on

ecosystem functioning increases linearly until four/six genotypes and then reaches a plateau

beyond that (Raffard *et al.* 2019c). Since we aimed at testing the effect of richness while

minimizing the effects of population identity, each replicate of each treatment of richness

contained a different (unique) assemblage of populations. Functional richness consisted in

manipulating the body mass of the individuals present in the mesocosms; hence, experimental

populations contained individuals sourced either from large or small populations (i.e., one

functional entity, *low functional richness*), or from both small and large populations (i.e., two

functional entities, *high functional richness*, see Table S1 and Fig. S2 for details on the

different experimental populations). It is noteworthy that populations containing only either

small or large individuals were actually more functionally redundant than populations

containing large and small individuals. Experimental populations hence varied simultaneously

according to their genotypic and functional richness, leading to four treatments.

The experiment lasted 30 weeks (210 days) after fish were introduced. Mesocosms

were checked daily for mortality, which was rarely observed over the course of the

experiment. At the end of the experiment, we measured several ecological parameters to

assess effects of genotypic and functional richness on population performance, community

structure and ecosystem functioning:

- *Population performance*: All tanks were emptied and we recaptured all remaining

fish to estimate mortality rate (1 - nb. survival fish/nb. introduced fish; mean per tank ±

standard error (SE) = 0.22 ± 0.01). Fish were weighted to assess fish biomass production

(biomass production = averaged final weight - averaged initial weight) of each experimental

population during the experiment. Since individuals with small initial body mass intrinsically

displayed a higher growth rate than large individuals, a standardised biomass production

index was calculated as the residuals of the relationship between biomass production and

initial fish size.

- Community structure: Zooplankton community structure was quantified by filtering

5 L of water through a 200 μm sieve. Samples were conserved in a 70% ethanol solution and

subsequently identified to the order or family levels, including Cyclopoida, Calanoida,

Daphniidae, Chydoridae and Bosminidae. The diversity of zooplankton was calculated as the

Simpson's diversity (*D-zoo*) representing the probability that two randomly chosen

individuals belong to different clades. *D-zoo* was calculated as $1 - \frac{\sum N_i * (N_i - 1)}{N_{tot} * (N_{tot} - 1)}$, where N_{tot}

was the total number of sampled individuals, and N_i the number of sampled individuals for

each group (Simpson 1949; Lande 1996). Zooplankton abundance was quantified as the total

number of individuals for all taxa pooled at the tank level.

Benthic invertebrates were collected from the mesh bags used to measure

decomposition rates (*see below*), conserved in a 70% ethanol solution, and identified as

Isopoda, Diptera, Gastropoda, Ephemeroptera, Plecoptera, Odonata, Copepoda, Cladocera,

and Ostracoda. The diversity of benthic invertebrates was calculated as the Simpson's

diversity (*D-inv*). The abundance of benthic invertebrates was quantified as the total number

of individuals for all taxa pooled at the tank level.

- Ecosystem functioning: Decomposition rate was measured by quantifying the mass

loss of black poplar (*Populus nigra*, a dominant riparian tree in southern France) abscised

leaves (Alp *et al.* 2016). On the 7th of March 2018 (19 weeks after the introduction of fish), 4

g of air-dried leaves were put in each mesocosm within a coarse mesh (1 x 1 cm) bag. At the

end of the experiment, the remaining leaf material was removed from the mesocosms, rinsed

with tap water, oven dried at 60°C for three days and weighed to the nearest 0.001 g to assess

the loss of biomass. The decomposition rate was calculated as $k = -\frac{ln\,(X)}{t}$ (Alp *et al.* 2016),

where *X* is the proportion of litter remaining and *t* is the elapsed time in days.

Pelagic algae stock was measured as the chlorophyll-a concentration ($\mu g.L^{-1}$) in the

water column using a multiparametric probe (OTT, Hydrolab DS5®). Five measurements

were taken in each mesocosm and averaged for subsequent analyses. Since phytoplankton are

the main primary producers in these mesocosms (Downing & Leibold 2002), pelagic algae

stock can be considered as a proxy for the biomass of primary producers.

Statistical analyses

Prior to analysis, the pelagic algae stock (i.e. the chlorophyll-a concentration), zooplankton

and benthic invertebrate abundances were log-transformed to reach normality of models'

residuals. After analyses of outliers, we removed one mesocosm from the final analyses that

displayed outliers for several of the measured variables. Analyses were hence run on 23

replicates.

We first assessed the ecological effects of genotypic and functional richness using

mixed effects linear models (LMMs). We ran one model for each ecological parameter as a

dependent variable, and the genotypic richness, functional richness, and the resulting two-

term interaction (that was removed when non-significant) as explanatory variables. Fish

mortality rate during the experiment was included as a fixed effect to control for a potential

effect of final density on ecological processes. To control for the disposition of the tanks

during the experiment, the position of tanks was added as a random term. To compare the

relative strength of a loss in genotypic and functional richness on ecological parameters we

calculated effect sizes. Hedges' g were computed as follow: $g = \frac{X_2 - X_1}{s_{pooled}}$, where X_1 and X_2 are

means of treatments (for each genotypic or functional richness treatment) measured for each

ecological parameters respectively, and s_{pooled} is the pooled standard deviation computed as

$s_{pooled} = \sqrt{\frac{(n_2-1)s_2^2+(n_1-1)s_1^2}{n_1+n_2-2}}$, where n is sample size, and s^2 is variance (Nakagawa &

Cuthill 2007). An effect size was hence measured for each variable response (and each

treatment), and effect sizes were averaged. The absolute mean effect size for genotypic and

functional richness (g_G and g_S, respectively) were compared visually based on 95%

confidence intervals (CI).

Secondly, we ran confirmatory path-analysis (Shipley 2009) to assess the direct and

indirect links among intraspecific diversity (genotypic and functional richness), prey

(invertebrates and zooplankton) community structure and ecosystem functioning. We

expected that intraspecific diversity in European minnows affects directly invertebrate

community (both diversity and abundance), and indirectly ecosystem functions

(decomposition rate, algae stock) through trophic cascades. Specifically, we computed two

path analysis: one for the benthic food chain (fish-invertebrates-decomposition), and one for

the pelagic food chain (fish-zooplankton-algae stock) (Fig. S3). The fits of the structural

models were assessed using the *C* statistic, that follows a χ^2 distribution and whose associated

p-value indicates whether the models adequately fit the data or not. Finally, the structural

models were compared to null models that were actually models without the effects of

genotypic and functional richness on community parameters (see Fig. S3) using Akaike

Information Criteria (AIC, Cardon *et al.* 2011; Shipley 2013). This test allows assessing

whether the indirect effects of genotypic and functional richness lead to a better explanation

of the data, and whether trophic cascades were important or not.

All statistical analysis were run using R software (R Core Team 2013), LMM were

run using the R-package lme4 (Bates *et al.* 2014), and confirmatory path-analyses were run

using the R-package ggm (Marchetti *et al.* 2020).

Results

Both genotypic and functional richness of experimental populations significantly

affected several ecological processes (Table 1). At the population level, we found that the

interaction between genotypic and functional richness tended (p = 0.057) to alter fish biomass

production of experimental populations (Table 1, Fig. 2a). Specifically, the less diversified

experimental populations (low genotypic richness and low functional richness) displayed a

lower biomass production than all other treatments (Fig. 2a). At the community level, the

diversity of benthic invertebrates was significantly higher in the high genotypic richness

treatment (mean $D\text{-}inv \pm$ SE = 0.64 ± 0.04) than in the low genotypic richness treatment (D-

$inv \pm$ SE = 0.53 ± 0.03), and this pattern was repeatable across functional richness treatments

(Fig. 2d, Table 1). Irrespective of the genotypic richness treatment, mesocosms containing

fish populations with low functional richness had higher diversity of benthic invertebrates

than mesocosms containing fish populations with high functional diversity (Fig. 2d, Table 1).

At the opposite, the diversity of zooplankton ($D\text{-}zoo$) was affected by the interaction between

genotypic and functional richness (Fig. 2e, Table 1). Zooplankton diversity was strikingly

higher in populations with both high genotypic richness and high functional richness (Fig. 2e,

Table 1), suggesting that, in this case, the effect of genotypic richness depended on the

functional richness of experimental fish populations. Benthic invertebrates and zooplankton

abundances were consistently (i.e. across genotypic richness treatment) enhanced when

increasing functional richness (Fig. 2f, g, and Table 1). Genotypic richness of populations did

not affect benthic invertebrates and zooplankton abundances (Fig. 2f, g, and Table 1). A weak

but significant negative effect of functional richness on the pelagic algae stock was detected

(Table 1, Fig. 2c) and there was no significant effects genotypic and functional richness on

decomposition rate (Table 2). Overall, genotypic and functional richness induced ecological

effects of similar intensity (mean $|g_G|$ = 0.356, CIs = 0.043-0.669, and mean $|g_S|$ = 0.564, CIs

= 0.247-0.882, see Fig. S4 for details).

Confirmatory path analyses revealed that effects of intraspecific diversity on

ecosystem functioning were primarily mediated by trophic cascades, through changes in

community structure (Fig. 3). First, genotypic and functional richness affected the diversity

and abundance of benthic invertebrates (Fig. 3), which subsequently and positively affected

the decomposition rate ($p < 0.001$, Fig. 4a, b and Table S2). Second, functional richness

positively affected abundance of zooplankton, leading to a decrease in pelagic algae stock (p

< 0.001, Fig. 4c, d, Table S2). Overall, the confirmatory path-analysis confirmed the key role

of intraspecific diversity for both food chains, and its indirect effects for ecosystem functions

(Fig. 3 and Table 2). Indeed, models including genotypic and functional richness reproduced

adequately the causal pathways (Table 2), and their AICs were better than that of the null

models for each ecosystem function (Table 2).

Discussion

The present study demonstrates that loosing intraspecific diversity in a secondary consumer

species has substantial top-down consequences for community structure and ecosystem

functioning in food webs. We showed that both losses in functional (mass) and genotypic

richness sustained these iBEFs, suggesting that differences in mass among individuals (an

important functional trait for food web dynamics, Woodward *et al.* 2005) did not capture the

entire functional space, and thus that genotypic richness encapsulates important and "cryptic"

(unmeasured) functional diversity. The loss of genotypes within consumer populations can

affect both the community structure and the abundance of lower trophic levels, as well as

ecosystem functions with a similar strength than the loss of functional entities (i.e. body

mass). Specifically, we found that diversity loss (genotypic and functional) within populations

indirectly affects primary producer biomass and organic matter recycling, two ecosystem

functions forming the bases of food chains. This suggests that intraspecific diversity is a key

but understudied facet of biodiversity as it indirectly sustains BEF relationships, even when

changes in intraspecific diversity occur at the top of the food chains.

Our study suggests that genotypic richness can support non-negligible cryptic

functional diversity. Cryptic functional traits, such as metabolic rates or behaviour, can induce

different energetic needs and ultimately affect individual diets (Brown *et al.*, 2004; Careau &

diversity, 2012; Rosenblatt & Schmitz, 2016). Hence, fish with different genotypes may share

obvious functional traits (such as body mass) but may subtly differ in other cryptic functional

traits, making them unique ecologically (Loreau 2001). Although such "ecological

dissimilarity-despite-morphological similarity" has rarely been demonstrated within species,

there is now ample evidence that cryptic species (species being morphologically and

phylogenetically similar) can actually be ecologically unique regarding their influence on

ecosystems (e.g. Fišer *et al.* 2015). Interestingly, this cryptic diversity can interact with

functional richness in various ways (i.e., additive, enhancing, or compensatory) depending on

the ecological parameter considered (i.e., benthic invertebrate diversity, zooplankton

diversity, and fish biomass production, respectively). For instance, we found that genotypic

richness can compensate for the loss of functional diversity for population performance (i.e.,

increase fish biomass production), with high genotypic richness maintaining high biomass

production even when the distribution of body mass in the population is limited. We speculate

that genotypic richness can increase niche variation among individuals, thereby limiting the

loss of biomass production when decreasing functional richness (Duffy *et al.* 2001). These

findings echo and extent studies at the interspecific level demonstrating that phylogenetic

diversity among species explains variance in ecosystem functions that is not explained either

by species or functional diversities, suggesting that phylogenetic diversity sustain unmeasured

functional differences among species (Cadotte *et al.* 2012; Mouquet *et al.* 2012; Le

Bagousse-Pinguet *et al.* 2019). We argue that iBEFs are sustained by differences in functional

traits among individuals that can be directly measured (e.g. mass) and/or indirectly captured

through quantification of the genetic pool of individuals composing populations.

Accordingly, we showed that genotypic and functional richness can affect

independently (i.e., additively) and consistently community structure, demonstrating that

multiple facets of intraspecific diversity can regulate lower trophic levels. First, genotypic

richness increased diversity benthic invertebrates. This effect was very robust, as it was

repeatable and consistent across functional richness treatments. Genotypic richness probably

enhanced resource partitioning, allowing individuals to forage on a more diverse array of

resources, regulating the abundance of each taxonomic group and leading to a higher diversity

(Duffy, 2002; Johnson *et al.*, 2006; Hughes et al. 2008). Second, functional richness led to

increased prey abundance that was repeatable across both communities of benthic

invertebrates and zooplankton. Previous studies at the inter- and intraspecific levels provide

non-consistent predictions, since an increase in consumer diversity can affect either positively

or negatively resource abundance (Sih *et al.* 1998; Griffin *et al.* 2013; Rudolf & Rasmussen

2013b; Antiqueira *et al.* 2018; Barnes *et al.* 2018). In our experiment, although functional

richness enhanced prey abundance, it also increased fish biomass production, suggesting that

functionally rich populations did not consume fewer resources than functionally poor

populations. On the opposite, flexible exploitation of resources might occur because European

minnows are omnivorous (Frost 1943; Collin & Fumagalli 2011), their diet probably included

some periphyton, decreasing the predation risk on animal resources. Because the top-down

control of intraspecific diversity on community structure is likely driven by trophic

mechanisms, quantifying individual diet and its temporal dynamic in such experiments would

allow making more accurate predictions regarding trophic niche partitioning.

Our results further revealed how changes in top consumers' genotypic and functional

richness percolate through the food web and alter ecosystem functions at the basis of the food

chains. Such trophic cascade induced by intraspecific diversity could be driven by a

"classical" (i.e. interspecific) BEF relationship between benthic invertebrate diversity and

decomposition rate (Hooper *et al.* 2005; Gessner *et al.* 2010). Specifically, fish genotypic

richness increased benthic invertebrate diversity that accelerated litter decomposition rate.

The higher decomposition rate of organic matter is likely produced by higher consumption

efficiency through trophic complementarity among clades of invertebrates in diverse

community (Gessner *et al.* 2010). Invertebrate community with a high diversity probably

harboured a high functional diversity (Cadotte *et al.* 2011), and focusing on the functional

type of invertebrates might allow a more precise understanding of this link. These results echo

those reported at the community level and those manipulating richness within primary

producer species (Downing & Leibold 2002; Crutsinger *et al.* 2006), while implying a

modification of top-down effects by intraspecific diversity in consumers. Biodiversity within

consumer species is largely impacted by anthropogenic activities (Miraldo *et al.* 2016),

involving the loss of non-measured functional diversity, which might further lead to

underestimated cascading impacts on ecosystem functioning.

In conclusion, we demonstrated that both genetic and functional richness within

consumer populations are important facets of biological diversity, inducing effects on prey

community structure and trophic cascades mediating ecosystem functions. These results are

consistent with previous synthetic works (Koricheva & Hayes 2018; Raffard *et al.* 2019c),

reinforcing the call for considering changes of intraspecific diversity in consumer species as

an important ecological predictor. Importantly, genotypic richness can sustain important

cryptic functional diversity, and future investigations should aim at developing a general framework from genes to ecosystems to better understand the global links existing among the multiple facets of biodiversity and ecosystem functioning, and ultimately ecosystem services.

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Conflict of interest disclosure

The authors of this article declare that they have no financial conflict of interest with the content of this article.

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Figure captions

Fig. 1. Predicted ecological effects of genotypic richness in relation with functional richness. Functional richness is assumed to positively affect ecological functions, whereas genotypic richness might have different effects depending on it support cryptic functional diversity. First, if functional richness does capture the essential of the functional differences among genotypes, then genotypic richness would have no ecological effects (a). Second, if genotypic richness supports cryptic functional diversity, the effects could be (b) additive, (c) enhancing, or (d) compensatory.

Fig. 2. Effects of genotypic and functional richness on populations biomass production index (a), decomposition rate (b), algae stock (c), diversity of benthic invertebrates (e), diversity of zooplankton (f), abundance of benthic invertebrates (g), and abundance of zooplankton (h). Error bars represent ± 1 SE.

Fig. 3. Causal pathways between genotypic richness, functional richness, community structure, and ecosystem functioning. Two models have been run separately on decomposition rate (a), and pelagic algae stock (b). Only significant links are drawn, blue arrows represent positive links, and yellow arrows negative links.

Fig. 4. Effects of the diversity (a) and the abundance (b) of benthic invertebrates on decomposition rate; and effects of the diversity and the abundance of zooplankton on pelagic algae stocks (c and d, respectively). Points are partial residuals extracted from models described in the statistical analysis section (see also Table S2). Straight lines represent the

615 slope predicted by the models (see statistical analyses) when significant ($\alpha < 0.05$), and grey

616 shadows represent 95% CIs.

HG = High genotypic richness
LG = Low genotypic richness

619 **Fig. 1.**

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622 **Fig. 2.**

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625 **Fig. 3**
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629 **Fig. 4.**

630 **Table 1.** Results of the mixed effect linear models quantifying the relationships between

631 genotypic richness, functional richness and ecological parameters. Significant -and near

632 significant- *p*-values are displayed in bold. Interaction terms were removed from the model

633 when not significant.

Response	Effect	*p*-value	χ^2, d.f.
Fish biomass production	Genotypic richness	0.301	(1.071, 1)
	Functional richness	0.634	(0.227, 1)
	Genotypic richness* Functional richness	**0.057**	(3.621, 1)
	Mortality	**0.023**	(5.196, 1)
Benthic invertebrates diversity	**Genotypic richness**	**0.007**	(7.152, 1)
	Functional richness	**0.002**	(9.484, 1)
	Mortality	0.212	(1.551, 1)
Zooplankton diversity	**Genotypic richness**	**0.052**	(3.756, 1)
	Functional richness	**0.004**	(8.163, 1)
	Genotypic richness* Functional richness	**0.038**	(4.287, 1)
	Mortality	0.851	(0.035, 1)
Benthic invertebrates abundance	Genotypic richness	0.300	(1.072, 1)
	Functional richness	**0.031**	(4.667, 1)
	Mortality	0.238	(1.392, 1)
Zooplankton abundance	Genotypic richness	0.885	(0.021, 1)
	Functional richness	**0.004**	(8.154, 1)
	Mortality	0.220	(1.503, 1)
Decomposition rate	Genotypic richness	0.140	(2.169, 1)
	Functional richness	0.690	(0.158, 1)
	Mortality	0.893	(0.018, 1)
Pelagic algae stock	Genotypic richness	0.834	(0.044, 1)
	Functional richness	**0.053**	(3.742, 1)
	Mortality	0.412	(0.673, 1)

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637   **Table 2.** Model fits of the two confirmatory path-analyses to explain decomposition rate and

638   algae stock. The full models included the effect of genotypic and functional richness on

639   community components while the null model did not.

Variable	C	df	p-value	AIC full model	AIC null model
Decomposition rate	8.555	6	0.200	32.555	75.043
Algae stock	6.745	10	0.749	52.745	64.679