

1 **Campaign-Based Citizen Science for Environmental Mycology: the “Science Solstice”**  
2 **and “Summer Soil-stice” Projects to Assess Drug Resistance in Air and Soilborne**  
3 ***Aspergillus fumigatus***

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11 **Abstract**

12 Citizen science projects are often undertaken for ecological and environmental research  
13 purposes but also have great potential for use in microbiology research to track the emergence  
14 and spread of pathogens in the environment. ‘Science Solstice’ and ‘Summer Soil-stice’ are  
15 mycology citizen science projects aimed at assessing drug resistance in *Aspergillus fumigatus*  
16 fungal spores found in air and soil, respectively, in the United Kingdom (UK). *A. fumigatus*  
17 plays an important role in the environment as a decomposer of plant material, but is also an  
18 opportunistic human lung pathogen. Infection with drug-resistant spores can lead to a worse  
19 clinical outcome for the patient.

20 On the first four solstice and equinox days between June 2018 and June 2019, volunteers were  
21 asked to collect air samples from their homes and workplaces and return them to our lab in  
22 Freepost envelopes. An additional round of samples was requested from volunteer’s gardens  
23 and/or compost on the June 2019 solstice. In total, 787 volunteers returned 2,132 air samples  
24 and 509 soil samples, which grew a total of 7,991 *A. fumigatus* colonies. The estimated total  
25 cost of the study was £2,650; the equivalent of 33 pence per *A. fumigatus* colony grown.

26 Incorporating citizen science into the environmental surveillance of drug-resistant *A. fumigatus*  
27 allowed for the simultaneous collection of hundreds of environmental samples across the entire  
28 UK on the same day. The insights generated from this study would not be practical in the  
29 absence of public participation and offers opportunities to ask scientific questions that were  
30 previously unaskable.

31

32 **Keywords:** mycology, antifungal resistance, epidemiology, citizen science, environmental  
33 sampling, pathogen monitoring

## 34 Introduction

35 Citizen science is defined as the “intentional involvement, in a non-professional capacity, of  
36 people in the scientific process, e.g. the collection... of data” (Pocock, 2015) and is becoming  
37 increasingly popular for simultaneously conducting research and engaging with the public  
38 about science. Many citizen science projects in the UK rely on volunteers to monitor population  
39 levels of native insects (Gardiner, 2012; Lye, 2012; Wilson, 2018), wildlife (Hof and Bright,  
40 2016), birds (Cannon, 2005; Sparks, 2017) and plants (Rich and Woodruff, 1990; Pescott,  
41 2015). Citizen scientists can also report environmental incidents with potentially harmful  
42 effects such as toxic algal blooms (Ransom Hardison, 2019) or river pollution (Hyder, 2017),  
43 and can aid surveillance of invasive species (Pocock and Evans, 2014), wildlife diseases  
44 (Robinson, 2010; Lawson, 2012) or plant pathogens (Brown, 2017). The majority of these  
45 projects ask participants to record their observations, either online, through an app or via post  
46 over a prolonged period of time.

47 Projects may also raise awareness of invisible health threats such as air pollution, pathogen  
48 spread and antimicrobial resistance (AMR). One example is Netherlands-based iSPEX, where  
49 participants measured atmospheric aerosols on a single day using an optical add-on for their  
50 smartphones with a corresponding app that collated data (Snik, 2014). A UK-based example is  
51 Swab & Send: an ongoing, self-funding microbiology project asking citizen scientists to take  
52 swabs of any object or environment they choose to help identify new antibiotic compounds  
53 ([www.lstmed.ac.uk/public-engagement/swab-send](http://www.lstmed.ac.uk/public-engagement/swab-send)). Public health-focused projects like these  
54 provided the inspiration for this study. We asked volunteers to collect samples from the air and  
55 soil, at home and work in the UK, for the surveillance of antifungal-resistant spores of  
56 *Aspergillus fumigatus*, a ubiquitous decomposer of dead plant matter and opportunistic human  
57 lung pathogen.

58 On average, we inhale 100s of *A. fumigatus* spores a day (Kwon-Chung and Sugui, 2013), some  
59 of which cause hypersensitisation and “fungal asthma” or aspergillosis disease ranging from  
60 chronic colonisation of the airways to invasive bloodstream infections. In the UK, as many as  
61 400,000 individuals suffer from severe asthma with fungal sensitisation (SAFS), approximately  
62 238,000 individuals with aspergillosis lung disease and an estimated 4,200 individuals with  
63 invasive aspergillosis (IA) (Pegorie, 2017). IA has a mortality rate ranging from 30-80%  
64 (Bongomin, 2017) and its prevalence is increasing in the UK due to increasing numbers of  
65 patients receiving immunosuppressive therapies for transplant, cancer or autoimmune  
66 conditions and the ageing population (Löbermann, 2012). Patients that are in critical care with  
67 severe viral infections such as influenza are at high risk of IA (Schauwvlieghe, 2018), and we  
68 are already witnessing examples of IA in patients that are ill with COVID-19. Increasingly,  
69 these infections are resistant to the medical antifungals (i.e. azole drugs) used to treat them  
70 despite no prior exposure of the patient to these drugs, suggesting environmental acquisition  
71 of resistance by the infecting spores pre-inhalation. Early diagnosis and treatment are  
72 associated with better patient outcomes, yet a survey by The Aspergillosis Trust revealed that  
73 diagnosis took between 1 to 5 years for 60% of the 128 respondents (personal comms, Sandra  
74 Hicks and Gillian Fairweather at The Aspergillosis Trust). In a survey of the scientific  
75 community on Twitter ( $n = 1,267$ ; April 2020), only 54% replied “yes” when asked if they had

76 heard of aspergillosis or knew what it was. This study aimed to raise awareness amongst  
77 participants by publishing blog posts on institute websites and including information sheets in  
78 sampling packs about *A. fumigatus*, aspergillosis and the relevance of widespread  
79 environmental sampling.

80 To date, much of the focus around environmental monitoring of airborne *A. fumigatus* spores  
81 in the UK has been on industrial composting facilities and potential risks to workers and nearby  
82 residents, with reports published by Department for Environment, Food & Rural Affairs  
83 (Defra) (Knight, 2009), Environment Agency (EA) (Environment Agency, 2018) and Health  
84 & Safety Executive (HSE) (Gilbert, 2003). Further studies have collected air and/or soil  
85 samples from areas in the UK over time to assess the prevalence of azole-resistant *A. fumigatus*:  
86 Greater Manchester from 2009-2011 (Alshareef and Robson, 2014; Bromley, 2014), Dublin  
87 from 2014-2016 (Dunne, 2017), South Wales from June to November 2015 (Tsitsopoulou,  
88 2018) and 6 sites across Southern England from May to July 2018 (Sewell, 2019). These studies  
89 give valuable insight but are limited in sample number and coverage due to sample collection  
90 being undertaken by the study authors themselves. In order to address some of these problems,  
91 this study reports the UK-wide collection of outdoor air and soil samples by citizen scientists,  
92 in a campaign-based, single timepoint manner, from which *A. fumigatus* spores were cultured  
93 and will ultimately be tested for azole antifungal-resistance.

94

## 95 Methods

96 The aims of this citizen science project were to monitor for drug-resistant *A. fumigatus* spores  
97 in outdoor air and soil across the UK at multiple timepoints. The ‘citizen science’ methodology  
98 of the study had several rationales: 1) to provide a step-change in UK spatial coverage from  
99 previous studies, 2) to raise awareness of aspergillosis diseases amongst the general public, and  
100 3) to trial the efficacy of the chosen sample collection methods on citizen scientists as a viable  
101 approach for mycological research. The outcome of the study will be to determine whether  
102 there are spatial or temporal determinants of resistance that could inform future policies to  
103 protect those at risk for aspergillosis.

104 We thereby asked individuals residing in the UK to collect spore samples from their local air  
105 on four dates (21<sup>st</sup> June 2018, 24<sup>th</sup> September 2018, Friday 21<sup>st</sup> December and 20<sup>th</sup> March 2019)  
106 and garden soil on one date (21<sup>st</sup> June 2019). These dates were chosen because they: 1) were  
107 the solstice and equinox dates making them easy to remember for participants and ‘catchy’ for  
108 the purpose of advertising; 2) were equally spaced throughout the year making it useful for  
109 examining seasonal shifts in spore recovery; and 3) allowed for sufficient time in the laboratory  
110 to process samples before the next sampling campaign.

### 111 *Recruitment for citizen science projects*

112 Participants were recruited for the projects by posts published on social media platforms  
113 Twitter and Facebook, as well as on several mycology websites and The Aspergillosis Trust  
114 website ([www.aspergillosistrust.org](http://www.aspergillosistrust.org)), containing a poster (Figure 1), a brief description of the  
115 project and a link to a Google form (Supplementary Figure 1). Printed posters were displayed  
116 outside author and co-author offices and on noticeboards around Imperial College London  
117 (ICL) and UK Centre for Ecology & Hydrology (UKCEH). The posters displayed the name of  
118 each project, all of which incorporated solstice or equinox, and an image of Stonehenge, which  
119 is iconic for such celestial events; all in an effort to make the sampling dates memorable. The  
120 posters also contained a brief description of the project that aimed to be understandable to non-  
121 scientists, links to an online blog post containing further information about the project, and the  
122 Twitter handles of the author and project to be followed for regular updates. Twitter was chosen  
123 as a way of providing project updates because Tweets are visible to the public and did not  
124 require the participants to befriend or follow the authors as on other social media platforms.  
125 Twitter updates also avoided potentially upsetting participants by sending unsolicited emails.  
126 At the bottom of each poster was a shortened URL to the Google form. Emails were sent by  
127 co-authors to ICL and UKCEH mailing lists containing a description of the project and a link  
128 to the Google form.

129 On the Google form, participants were requested to provide their name and address, for postage  
130 purposes, their email address for a reminder email sent the week before and mobile number for  
131 a reminder text to be sent the evening before. For the initial air sampling round, optional  
132 questions asked them for the research institute they are affiliated to (if any) and to say how  
133 they had heard of the project. All communications were checked for General Data Protection  
134 Regulation (GDPR)-compliance as of new rules introduced on 25<sup>th</sup> May 2018 and participants  
135 were informed *via* the Google form and *via* email about how their personal data would be used,  
136 stored and kept confidential.

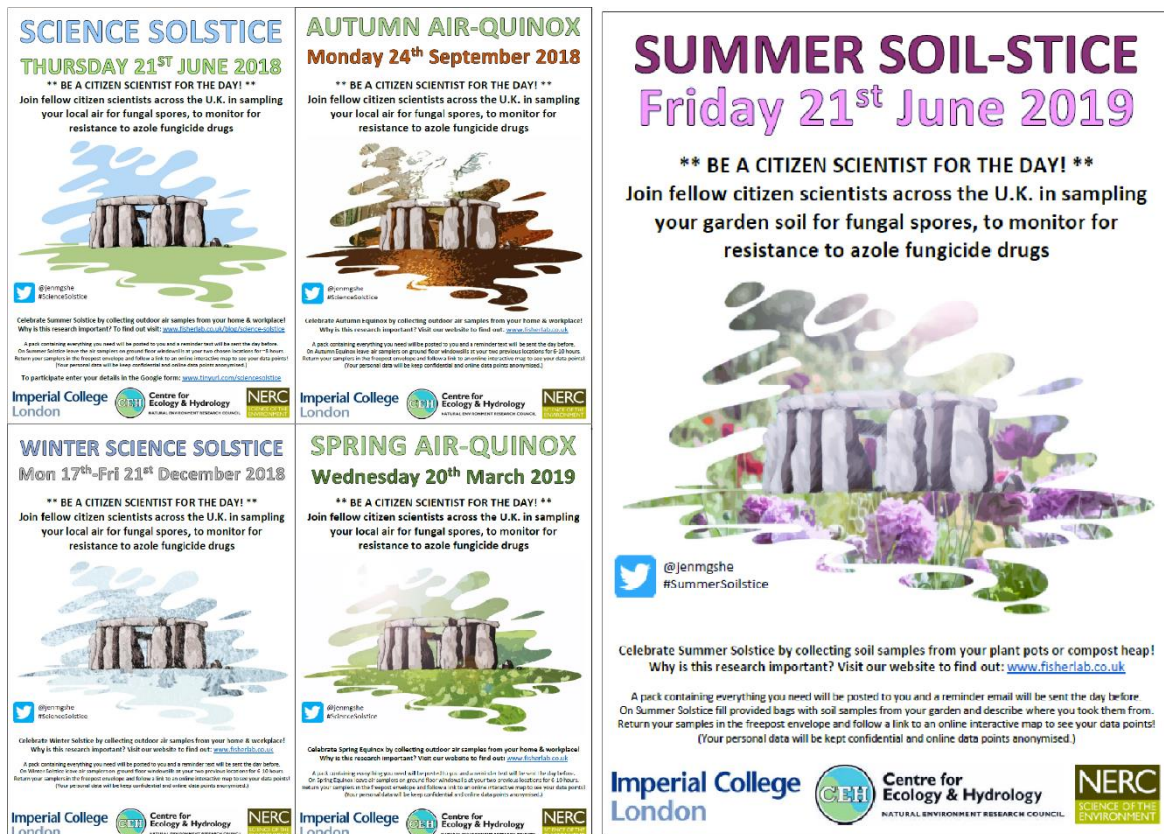


Figure 1: Posters advertising for citizen scientists to take part in UK-wide air and soil sampling projects were displayed around ICL and UKCEH and posted on social media platforms prior to sampling dates.

137

### 138 *Air sampling citizen science*

139 Participants who filled in the Google form to take part in one or more of the four air sampling  
 140 rounds were sent an air sampling pack containing a MicroAmp™ clear adhesive film (Applied  
 141 Biosystems™, UK) cut in half (to produce two “air samplers”) with adhesive putty attached  
 142 for securing in place. The pack also contained a poster (Figure 1), a questionnaire  
 143 (Supplementary Figure 2), simple instructions (Supplementary Figure 3) and a brief scientific  
 144 description of the experimental aims (Supplementary Figure 4). Participants were asked to  
 145 attach the air samplers to outdoor ground floor windowsills at their home and workplace and  
 146 expose by peeling off the backing slip for 6-10 hours on sampling day. If torrential rain was  
 147 forecast, participants were asked to sample on the soonest dry day after as rain falling on the  
 148 air samplers reduced their stickiness and therefore their ability to capture spores. They were  
 149 then asked to re-cover the air samplers and return them by post, along with the completed  
 150 questionnaire, in the Freepost envelope provided. The questionnaire asked the date and  
 151 geographical locations of sample collection, whether they were collected from outdoor ground-  
 152 floor windowsills, and for the participant to provide an email if they wished to receive updates.  
 153 Upon receipt, *A. fumigatus* colonies were cultured directly from the air samplers onto petri  
 154 dishes containing agar and stored at 4°C in a refrigerator for further analysis.

### 155 *Soil sampling citizen science*

156 Participants who filled in the Google form to take part in soil sampling, which followed after  
 157 the air sampling rounds, were sent a pack containing two plastic sachets, a wooden spatula, a

158 poster (Figure 1), simple instructions (Supplementary Figure 5) and a brief scientific  
159 description of the experimental aims (Supplementary Figure 4). They were asked to fill two  
160 plastic sachets with soil from their garden and complete a questionnaire (Supplementary Figure  
161 6) detailing the geographical location of their garden, the location of the soils within their  
162 garden (pot or planter, border, bag of compost, bag of manure, compost heap) and a brief  
163 description of the sample (e.g. plant or bulb type in pot, brand of compost or manure, contents  
164 of compost heap). They were then asked to return the sealed sachets of soil and the  
165 questionnaire in the Freepost envelope provided. Upon receipt, 2 g of each soil sample was  
166 plated onto petri dishes containing agar to culture *A. fumigatus* colonies, which were then  
167 stored at 4°C in a refrigerator for further analysis, along with the remainder of the soils.

168 For the soil sampling project, the blog post published on the UKCEH website (Supplementary  
169 Figure 7) explained that compost heaps and bags of compost might act as “hotspots” for the  
170 growth of azole-resistant *A. fumigatus*. In an effort to mitigate exposure, participants were  
171 advised to exercise caution when sampling from these locations as disturbance can lead to  
172 aerosolization of large numbers of spores. People were asked not to take part in the project if  
173 they suffer from aspergillosis, have a lung condition (chronic or acute, such as ‘flu) or are  
174 immunosuppressed, as these all put them at greater risk of contracting aspergillosis from  
175 inhaling a large number of spores. Participants were asked to sample from locations within  
176 their own garden only so they experienced equivalent or lesser exposures in taking part as from  
177 standard gardening activities such as potting, digging and compost manipulation. Participants  
178 were free to opt out at any time by emailing the primary author or by not collecting samples.

### 179 *Citizen science engagement*

180 Participants were encouraged throughout the projects to share photos of their sampling on the  
181 designated day via Twitter or email. When provided, participants were asked for their consent  
182 for this material to be used in future work and presentations by the author. Participants were  
183 also given the option to opt-out of the projects at any time by emailing the author or by not  
184 returning their samples. Participants were asked on the questionnaires to indicate whether they  
185 were happy to receive future project updates by email. Those who opted to receive updates  
186 were sent an email approximately 4-6 weeks after each round when all samples had been  
187 processed thanking them for their participation, informing them of the number of samples  
188 received, the number of *A. fumigatus* colonies grown and a link to an online Google map  
189 showing the location of each sample processed and the number of colonies grown from it.

190 On 25<sup>th</sup> June 2018, four days after the initial air sampling round, an email was sent to  
191 participants who provided an email address asking for feedback on the project via a different  
192 Google form (Supplementary Figure 8). This form asked the reason participants did not take  
193 part (if they didn’t), whether the email and text reminders were useful, whether they’d like to  
194 part again if the experiment was repeated, and had a comment box for additional feedback.

195

196

## 197 Results

### 198 *Citizen science participation in the UK*

199 Across the four air sampling projects spanning June 2018 to March 2019 a total of 485 unique  
200 individuals residing in the UK collected one or more air samples. A total of 1,293 air sampling  
201 packs were sent out and 976 were returned, equating to an overall participation rate of 75%.  
202 Participants collectively returned 1,896 air samples across the four dates, which were collected  
203 from all over England, Northern Ireland, Wales and Scotland. Screenshots of the Google maps  
204 sent out to participants after the air and soil sampling rounds is shown alongside a UK  
205 population density map in Figure 2, which shows that the majority of samples were sent in  
206 from populous areas. Concomitantly, areas with the lowest coverage of air and soil sampling  
207 are also less densely populated. Results for individual air sampling rounds and for the soil  
208 sampling round are shown in Table 1.

Type of sampling	Sampling date	Packs sent out	Packs returned	Return rate (%)	Samples collected
Air	21 <sup>st</sup> June 2018	461	365	79	712
	24 <sup>th</sup> September 2018	300	204	68	398
	21 <sup>st</sup> December 2018	231	165	71	321
	20 <sup>th</sup> March 2019	301	242	80	465
Soil	21 <sup>st</sup> June 2019	334	246	74	509

Table 1: Numbers of sampling packs sent out and number of packs and samples returned across the four air sampling dates and one soil sampling date.

209

210 Of the 365 participants in the first air sampling round, 160 (43%) also took part in the second  
211 round, 120 (32%) in the third round and 112 (30%) took part in all four air sampling rounds.  
212 Of the 246 participants in the soil sampling round, 43 (17%) had already taken part in one or  
213 more of the air sampling rounds.

### 214 *Citizen science participation globally*

215 Due to the global nature of Twitter the first air sampling round attracted 52 participants from  
216 16 countries in addition to the UK. Whilst global sampling was not the intention of this study,  
217 sampling packs were sent to these participants for comparative analysis to UK samples. For  
218 the first air sampling round global participants sent back a total of 144 samples from Australia,  
219 Belgium, Canada, Chile, China, France, Germany, Hungary, Italy, Madagascar, New Zealand,  
220 Portugal, Spain, The Gambia, The Netherlands and USA. The second air sampling round  
221 received 92 samples from 50 individuals overseas: Canada, France, Germany, New Zealand,  
222 Portugal, Spain and USA. For the third and fourth air sampling rounds it was decided not to  
223 send sampling packs abroad as the Freepost return envelopes were not valid in other countries  
224 and the authors thought it unfair for participants to pay for postage. Soil sampling was open to  
225 UK participants only due to restrictions on moving soil samples between countries.

### 226 *Isolation of A. fumigatus from samples*

227 The 1,896 air samples collected and returned from the UK across the four air sampling rounds  
228 grew a total of 2,366 of fungal colonies that were identified morphologically as *A. fumigatus*,  
229 and the 236 air samples collected globally across the first and second air sampling rounds grew

230 a total of 451 *A. fumigatus* colonies (Table 2). The 509 soil samples grew a total of 5,174  
 231 colonies. The average number of colonies per air sample ranged from 1.8 to 3.1 across the four  
 232 sampling rounds, whereas the average number per soil sample was 15.8.

Type of sampling	Sampling date	Number of samples that grew <i>A. fumigatus</i> (% of total) <sup>a</sup>	Number of <i>A. fumigatus</i> colonies cultured <sup>a</sup>	Average number of colonies grown from <i>A. fumigatus</i> -positive samples
Air	21 <sup>st</sup> June 2018			
	<i>UK</i>	408 (57)	1152	2.8
	<i>global</i>	81 (56)	280	1.9
	24 <sup>th</sup> September 2018			
	<i>UK</i>	190 (48)	429	2.3
	<i>global</i>	63 (69)	171	1.9
	21 <sup>st</sup> December 2018	152 (47)	477	3.1
	20 <sup>th</sup> March 2019	169 (36)	308	1.8
Soil	21 <sup>st</sup> June 2019	327 (64)	5174	15.8

Table 2: Numbers of samples that grew *A. fumigatus* colonies and the number of colonies cultured across the four air sampling rounds and one soil sampling round. <sup>a</sup> These numbers represent fungal isolates cultured from samples that morphologically resembled *A. fumigatus*.

233

### 234 *Recruitment method*

235 For the initial air sampling project participants were asked several optional questions on the  
 236 Google sign up form. Of the 513 individuals who completed the form, in the UK and globally,  
 237 233 (45%) belonged to a research institute and 489 (95%) indicated how they'd heard of the  
 238 project. The research institutes that recruited the most individuals were the author's institutes  
 239 UKCEH ( $n = 36$ ) and ICL ( $n = 16$ ). The ways that individuals heard of the project were on:  
 240 Facebook ( $n = 139$ ), email ( $n = 128$ ), Twitter ( $n = 103$ ), word-of-mouth ( $n = 97$ ) and other ( $n$   
 241  $= 46$ ). The Tweet recruiting individuals for the first air sampling round (#ScienceSolstice)  
 242 made 27,731 impressions and received 642 total engagements. The Tweet recruiting for  
 243 subsequent air sampling rounds (#AutumnAirquinox, #WinterScienceSolstice and  
 244 #SpringAirquinox) made 13,288 impressions and had 244 engagements, and the Tweet for soil  
 245 sampling (#SummerSoilstice) made 29,350 impressions and had 823 engagements.

### 246 *Adherence to sampling date amongst UK samples*

247 The initial air sampling round had the highest adherence to sampling date of 94%, which  
 248 dropped to ~60% for the second and fourth air sampling round (Table 3). This drop was due to  
 249 the author's communication with participants preceding the second, third and fourth air  
 250 sampling dates to collect air samples on days either side of the sampling date if they were  
 251 unable to participate on the sampling date itself. The decision was taken that sample number  
 252 was more important than sampling date, as feedback from the first air sampling round  
 253 suggested that flexibility in sampling date would increase participation. The third air sampling  
 254 round had the lowest adherence to date because it was the week before Christmas and the  
 255 weather was unpredictable so participants were encouraged to sample on any date between 17<sup>th</sup>  
 256 and 21<sup>st</sup> December 2018 with suitable weather, which 274 (85%) did. Sample date was less



257 important for the soil sampling project because it was not affected by weather conditions, so  
258 less emphasis was placed on timing for this sampling round.

259

Type of sampling	Sampling date	Samples missing date (% of total)	Number of samples collected on intended sampling date (% of total)	Range of sampling dates
Air	21 <sup>st</sup> June 2018	8 (1)	669 (94)	20 <sup>th</sup> June – 1 <sup>st</sup> July
	24 <sup>th</sup> September 2018	9 (2)	249 (62)	17 <sup>th</sup> September – 1 <sup>st</sup> October
	21 <sup>st</sup> December 2018 <sup>a</sup>	4 (1)	12 (3)	12 <sup>th</sup> December – 19 <sup>th</sup> January
	20 <sup>th</sup> March 2019	5 (1)	287 (61)	13 <sup>th</sup> March – 24 <sup>th</sup> April
Soil	21 <sup>st</sup> June 2019	2 (0)	261 (51)	16 <sup>th</sup> June – 22 <sup>nd</sup> July

Table 3: Dates that UK samples were collected for four air sampling rounds and one soil sampling round. <sup>a</sup> Due to weather conditions, and proximity to Christmas, participants for the third air sampling round were asked to collect on any suitable day between 17<sup>th</sup> – 21<sup>st</sup> December, hence low adherence to sampling date.

260

### 261 *Participant feedback*

262 During all air and soil sampling rounds, participants engaged with the author by emailing or  
263 tweeting photos of themselves or family taking part in the sampling (Figure 3). The author  
264 received many messages of support by email, on Twitter and handwritten on completed  
265 questionnaires and has been requested by several participants to talk about the citizen science  
266 projects in schools and at meetings and conferences.

267 After sending update emails for each sampling round, the primary author received email replies  
268 from participants expressing their interest in the results, pleasure in taking part and even  
269 disappointment or apologies that their sample(s) had not grown *A. fumigatus*. Leading up to,  
270 and following on from, these update emails were progress updates on Twitter that included  
271 photographs of samples being returned and the primary author in the laboratory processing  
272 samples. These Tweets were well-received and maintained contact with participants who had  
273 opted to follow the primary author and/or project on Twitter, without sending unsolicited  
274 emails to participants who had only agreed to receive the final update email. They also allowed  
275 interested individuals to follow the project even if they hadn't participated, and on several  
276 occasions these individuals got in touch asking to participate in future sampling rounds.

277 There were 118 responses to the Google feedback form emailed out after the first air sampling  
278 round and only 9 responses were from individuals who had not participated, for the following  
279 reasons: air sampler(s) blew away or was removed ( $n = 6$ ), air sampling pack did not arrive in  
280 time ( $n = 1$ ) and they hadn't made it to a post box yet but still intended to post ( $n = 2$ ). 115  
281 responders (98%) found the email reminders helpful and 2 (2%) did not receive any. 98  
282 responders (85%) found the text reminder (Supplementary Figure 9) the evening before useful,  
283 3 (3%) did not find it useful, and 14 (12%) did not receive it. 115 of the 118 responders said  
284 they would like to participate in the citizen science experiment again if it were repeated and

285 provided their email address. 79 responders left additional feedback in the comment box  
286 (Supplementary Table 1), which was overall very positive and encouraging.

287 Suggestions for improvements were made by participants in personal communications with the  
288 primary author and *via* the Google feedback form (Supplementary Table 1) and amendments  
289 were made to subsequent sampling rounds. In the first air sampling round white sticky labels  
290 that read “LOCATION 1 (or 2) air sampler: please peel this off and retain, to re-cover air  
291 sampler after ~8hrs.” were stuck to the back of each air sampler (Figure 4A), to correspond  
292 with locations 1 and 2 on the questionnaires. It quickly became apparent on 21<sup>st</sup> June 2018 that  
293 several individuals had removed this label instead of peeling off the backing of the air sampler.  
294 These individuals were contacted immediately to remedy this mistake, when possible, but 20  
295 air samplers were returned that had not been exposed correctly. It is possible to tell when an  
296 air sample has not been exposed because it appears white and debris-free whereas exposed air  
297 samples are yellow, dust and debris is visible and the sticky face and backing slip are  
298 misaligned. For subsequent air sampling rounds the sticky labels were not used and air samplers  
299 were instead labelled by hand either “LOCATION 1” or “LOCATION 2” (Figure 4B) and  
300 participants referred to the instruction page for exposing the air samplers.

301 Participants also commented during and after the first air sampling round that their air samplers  
302 had blown away because adhesive putty was insufficiently adhesive, so for the second air  
303 sampling round the author instead attached double-sided foam tape to each air sampler. After  
304 the second air sampling round participants reported that the foam tape remained stuck to their  
305 windowsills and required chemical removal, so for the third and fourth air sampling round the  
306 author provided both adhesive putty and foam tape for participants to choose between.

307 Several participants contacted the author before, during and after the first air sampling round  
308 to apologise for not taking part because they were occupied on the sampling date. As a result,  
309 the author amended correspondence for subsequent air sampling rounds asking participants to  
310 collect samples on the sampling date whenever possible, but to collect them within 3 days  
311 either side of the sampling date if more convenient. A timeline of changes made to sampling  
312 methodology as a result of participant feedback is shown in Supplementary Figure 10.

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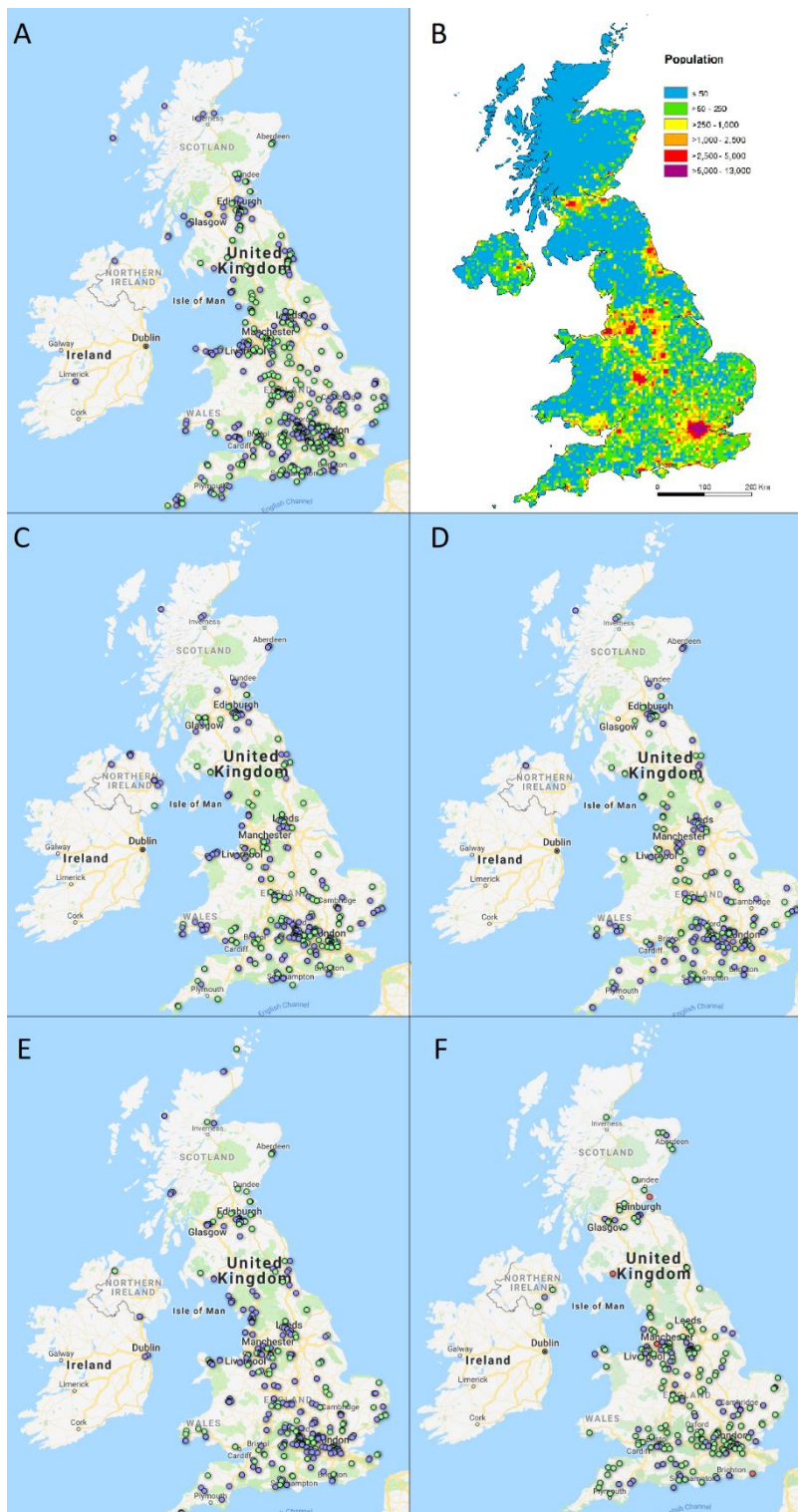


Figure 2: Google maps showing locations that participants collected air samples from on A) 21<sup>st</sup> June 2018, C) 24<sup>th</sup> September 2018, D) 21<sup>st</sup> December 2018 and E) 20<sup>th</sup> March 2019. F) shows locations that soil samples were collected from on 21<sup>st</sup> June 2019. Blue dots indicate samplers that did not grow *A. fumigatus* colonies, green dots indicate samplers that did and red dots indicate samplers that were contaminated with other fungal growth. B) is a population density map of the UK produced by Vieno et al (2015) (Heal and Williams, 2015).

315

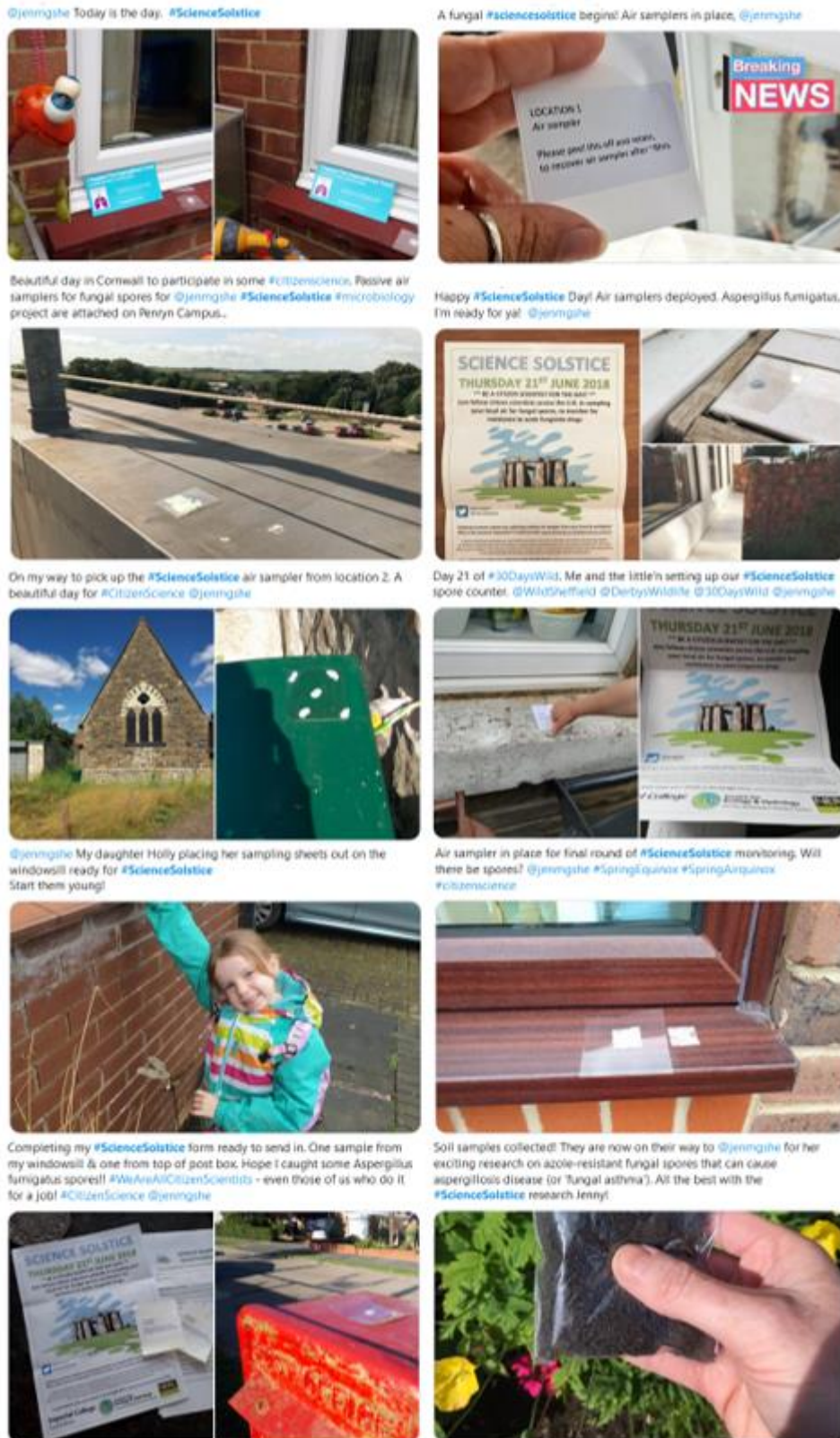


Figure 3: Several Tweets posted by participants on sampling days showing them taking part in air and soil sampling. (Permission granted by participants for these Tweets to be displayed.)

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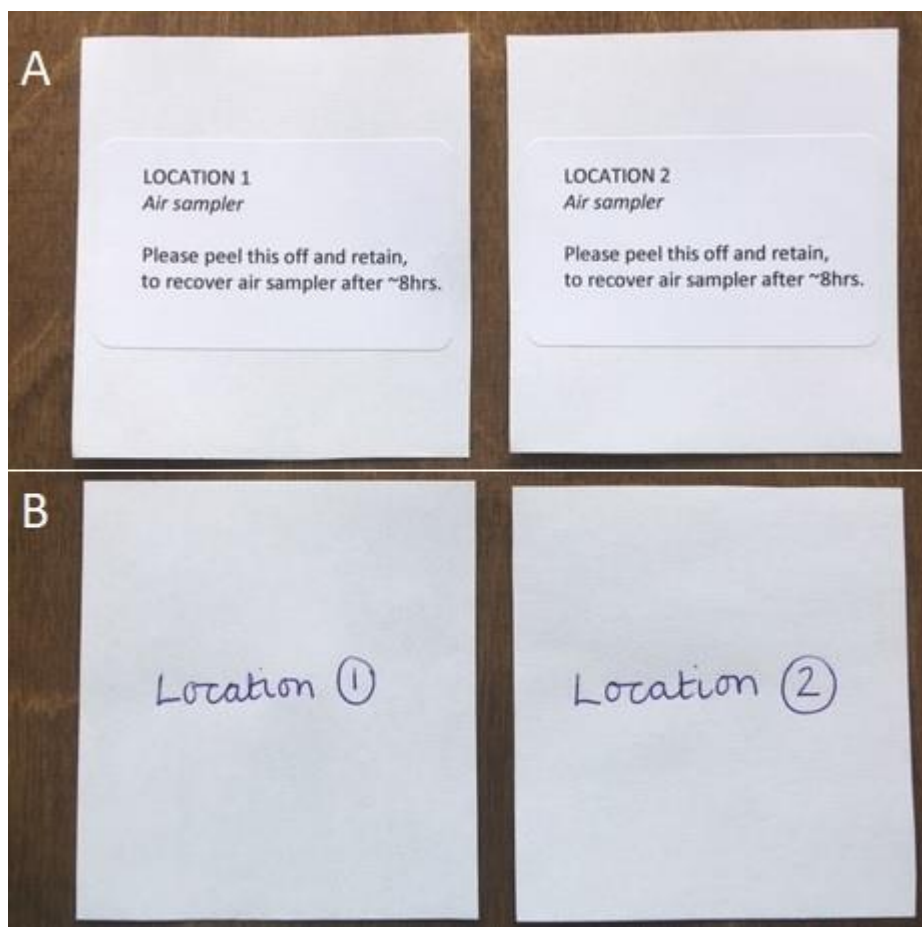


Figure 4: A) Air samplers for first air sampling round had sticky labels on the back with location number and basic instructions, but several participants peeled this off instead of the back of the air sampler. B) For subsequent air sampling rounds location number was handwritten on the back of each air samplers, to avoid confusion on how to expose the air sampler.

318

## 319 Discussion

320 This study asked citizen scientists to collect air and soil samples on five dates between June  
321 2018 and June 2019 and subsequently received a total of 2,132 air samples and 509 soil samples  
322 from 787 individuals. Advertising the first air sampling round on social media platforms  
323 Facebook and Twitter achieved a high initial enrolment due to the willingness of individuals  
324 to share posts, such that the initial Tweet containing the poster and a link to the Google sign-  
325 up form reached over 27,000 people. Individuals recruited through Facebook were mostly  
326 friends and family of the author, those recruited by email were colleagues at ICL or UKCEH  
327 and those who signed up through Twitter were colleagues, fellow mycologists, followers of  
328 The Aspergillosis Trust and the general public. Subsequent air sampling rounds benefitted from  
329 high retention rates, which participants told the author in personal communications was due to  
330 the simplicity of the sampling method, relatively small time commitment required, the  
331 enjoyment in taking part and their interest in results.

332 Organising sampling rounds to take place on single dates, which coincided with solstice and  
333 equinox days, was easier for the author and likely achieved a higher return rate than ongoing  
334 sampling. The author was able to send an email reminder several days before each sampling  
335 date and a text reminder the evening before such that, combined with television and radio  
336 broadcasts on solstice and equinox days, no participants stated they'd not taken part due to  
337 having forgotten about the project. The single sampling dates also meant that the majority of  
338 samples were returned in the following fortnight so the author was able to prepare and use lab  
339 consumables within a short timeframe, which is helpful for sterile culturing in mycology.  
340 Adherence to sampling date was exceptionally high (94%) for the first air sampling round and  
341 moderate for the second and fourth air sampling rounds (~60%) but dropped for the third (12%)  
342 due to winter weather conditions and proximity to Christmas. It is worth noting that the weather  
343 for the first, second and fourth air sampling rounds (summer, autumn and spring, respectively)  
344 was remarkably good, which likely increased participation levels and adherence to sampling  
345 dates. Importantly, the range of sampling dates did not overlap between sampling rounds so  
346 they can still be ascribed to different seasons as intended. Furthermore, every participant  
347 recorded the date of sampling on their questionnaire so, when undertaking spatial analyses in  
348 future, the authors can download meteorological data and adjust for factors such as wind speed  
349 and wind direction on the day each sample was collected.

350 The cost of sample collection by citizen scientists was considerably lower than if the author  
351 had attempted to collect this number and distribution of samples alone. Recruitment by social  
352 media platforms and email was free, stationery was provided by UKCEH and ICL and sampling  
353 packs were sent out using ICL's franking system, so major costs were the setup and renewal of  
354 Royal Mail first-class Freepost return and purchase of lab consumables. Additionally, for the  
355 first and second air sampling rounds, global participants were reimbursed for postage costs  
356 when possible but global participation was discontinued in subsequent sampling rounds. An  
357 estimated £1,800 was spent on laboratory reagents and consumables, £50 on stationery and  
358 £800 on Royal Mail postage costs bringing the total cost of this project to £2,650. This is the  
359 equivalent of £1 per sample or 33 pence per *A. fumigatus* isolate collected across all air and  
360 soil sampling rounds, both from the UK and globally.

361 The majority of participants across the five sampling rounds were resident in the UK (87%)  
362 and samples were received from England, Northern Ireland, Scotland and Wales. Without the  
363 involvement of citizen scientists, it would not have been possible for the authors to collect this  
364 number and distribution of air and soil samples at five single time points. The participation of  
365 the author's friends, family and colleagues show up as clusters of samples around  
366 Hampshire/Dorset (author's home place), Oxfordshire (UKCEH) and London (ICL), which  
367 contrasts with under-sampling in central Wales, central Scotland and Northern Ireland. The  
368 density of samples overlaps strongly with population density, so these clusters might be due to  
369 the under-sampled areas being less densely populated. The sampling discrepancies do not  
370 impact the microbiology or genetics aspect of the author's future work, but might hinder spatial  
371 analyses. Spatial coverage in low-population density regions would need to be addressed if the  
372 study was repeated.

373 The authors' reflection on this citizen science approach for sample collection is that it has  
374 exceeded our expectations in terms of participation levels, distribution of samples, timing of  
375 sample collection and numbers of *A. fumigatus* colonies grown for onward analysis. Involving  
376 citizen scientists has been an incredibly rewarding experience, because of their messages of  
377 support, Tweets and photos of them taking part in sampling, invitations to speak at schools and  
378 conferences and general enthusiasm. The authors hope this study has also raised awareness of  
379 aspergillosis diseases amongst participants both in the UK and globally. This study has resulted  
380 in a collection of 7,991 *A. fumigatus* isolates that are to be tested for susceptibility to azole  
381 drugs to determine the prevalence and distribution of azole-resistance here in the UK. The  
382 sampling techniques used were simple, inexpensive and standardized and could potentially be  
383 adapted and used to monitor environmental levels of other fungal, bacterial or viral pathogens,  
384 DNA or toxins, insects or chemicals of interest.

385

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393

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397

## 398 Competing Interests

399 The authors have no competing interests to declare.

400

401 **Author Contributions**

402 JMGS, ACS and MCF conceived the study and recruited participants. JMGS processed  
403 samples and communicated with participants. JMGS drafted the original manuscript, which  
404 ACS and MCF reviewed and provided edits.



## 405 References

- 406 Alshareef, F. and Robson, G. D. (2014) 'Prevalence , persistence , and phenotypic variation of  
407 *Aspergillus fumigatus* in the outdoor environment in Manchester , UK , over a 2-year period', *Medical*  
408 *mycology*, (April), pp. 367–375. doi: 10.1093/mmy/myu008.
- 409 Bongomin, F., Gago, S., Oladele, R. O., and Denning, D. W. (2017) 'Global and multi-national  
410 prevalence of fungal diseases—estimate precision', *Journal of Fungi*, 3(4). doi: 10.3390/jof3040057.
- 411 Bromley, M. J., Van Muijlwijk, G., Fraczek, M. G., Robson, G., Verweij, P. E., Denning, D. W., and  
412 Bowyer, P. (2014) 'Occurrence of azole-resistant species of *Aspergillus* in the UK environment',  
413 *Journal of Global Antimicrobial Resistance*. Taibah University, 2(4), pp. 276–279. doi:  
414 10.1016/j.jgar.2014.05.004.
- 415 Brown, N., Bosch, F. Van Den, Parnell, S., Denman, S., and Brown, N. (2017) 'Integrating regulatory  
416 surveys and citizen science to map outbreaks of forest diseases : acute oak decline in England and  
417 Wales', *Proceedings of the Royal Society B*.
- 418 Cannon, A. R., Chamberlain, D. A. N. E., Toms, M. P., Hatchwell, B. E. N. J., and Gaston, K. J. (2005)  
419 'Trends in the use of private gardens by wild birds in Great Britain 1995 – 2002', *Journal of Applied*  
420 *Ecology*, pp. 659–671. doi: 10.1111/j.1365-2664.2005.01050.x.
- 421 Dunne, K., Hagen, F., Pomeroy, N., Meis, J. F., and Rogers, T. R. (2017) 'Inter-country Transfer of  
422 Triazole- Resistant *Aspergillus fumigatus* on Plant Bulbs', *Clinical Infectious Diseases*, 65, pp. 147–  
423 149. doi: 10.1093/cid/cix257.
- 424 Environment Agency (2018) 'M9: Environmental monitoring of bioaerosols at regulated facilities',  
425 (July). Available at:  
426 [https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment\\_data/file/730](https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/730226/M9_Environmental_monitoring_of_bioaerosols_at_regulated_facilities.pdf)  
427 [226/M9\\_Environmental\\_monitoring\\_of\\_bioaerosols\\_at\\_regulated\\_facilities.pdf](https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/730226/M9_Environmental_monitoring_of_bioaerosols_at_regulated_facilities.pdf).
- 428 Gardiner, M. M., Allee, L. L., Brown, P. M. J., Losey, J. E., Roy, H. E., and Smyth, R. R. (2012)  
429 'Lessons from lady beetles : accuracy of monitoring data from US and UK citizen- science programs',  
430 *Frontiers in Ecology*. doi: 10.1890/110185.
- 431 Gilbert, E. J., Adrian, K., Karnon, J. D., Swan, J. R., and Crook, B. (2003) 'Occupational and  
432 environmental exposure to bioaerosols from composts and potential health effects - A critical review of  
433 published data'. Available at: <https://www.hse.gov.uk/research/rrpdf/rr130.pdf>.
- 434 Heal, M. R. and Williams, M. (2015) 'Sensitivities of UK PM2.5 concentrations to emissions  
435 reductions', *Atmospheric Chemistry and Physics*, (August 2016). doi: 10.5194/acp-16-265-2016.
- 436 Hof, A. R. and Bright, P. W. (2016) 'Quantifying the long-term decline of the West European hedgehog  
437 in England by subsampling citizen-science datasets', *European Journal of Wildlife Research*. European  
438 Journal of Wildlife Research, pp. 407–413. doi: 10.1007/s10344-016-1013-1.
- 439 Hyder, K., Wright, S., Kirby, M., and Brant, J. (2017) 'The role of citizen science in monitoring small-  
440 scale pollution events', *Marine Pollution Bulletin*. Elsevier, 120(1–2), pp. 51–57. doi:  
441 10.1016/j.marpolbul.2017.04.038.
- 442 Knight, A., Kumarwami, N., Lamarre, B., Lipscombe, R. P., Robinson, R. A., and Williams, M. (2009)  
443 'Rapid and responsive monitoring network for bioaerosol emissions: final report', (March). Available  
444 at: <http://eprintspublications.npl.co.uk/4689/>.

- 445 Kwon-Chung, K. J. and Sugui, J. A. (2013) ‘Aspergillus fumigatus-What Makes the Species a  
446 Ubiquitous Human Fungal Pathogen?’, *PLoS Pathogens*, 9(12), pp. 1–4. doi:  
447 10.1371/journal.ppat.1003743.
- 448 Lawson, B., Lachish, S., Colvile, K. M., Durrant, C., Peck, K. M., Toms, M. P., Sheldon, B. C., and  
449 Cunningham, A. A. (2012) ‘Emergence of a Novel Avian Pox Disease in British Tit Species’, *PLoS*  
450 *ONE*, 7(11). doi: 10.1371/journal.pone.0040176.
- 451 Löbermann, M., Boršo, D., Hilgendorf, I., Fritzsche, C., Zettl, U. K., and Reisinger, E. C. (2012)  
452 ‘Immunization in the adult immunocompromised host’, *Autoimmunity Reviews*, 11(3), pp. 212–218.  
453 doi: 10.1016/j.autrev.2011.05.015.
- 454 Lye, G. C., Osborne, J. L., Park, K. J., and Goulson, D. (2012) ‘Using citizen science to monitor *Bombus*  
455 populations in the UK : nesting ecology and relative abundance in the urban environment’, *Journal of*  
456 *Insect Conservation*, pp. 697–707. doi: 10.1007/s10841-011-9450-3.
- 457 Pegorie, M., Denning, D. W., and Welfare, W. (2017) ‘Estimating the burden of invasive and serious  
458 fungal disease in the United Kingdom’, *Journal of Infection*. Elsevier Ltd, 74(1), pp. 60–71. doi:  
459 10.1016/j.jinf.2016.10.005.
- 460 Pescott, O. L., Walker, K. J., Pocock, M. J. O., Jitlal, M., Outhwaite, C. L., Cheffings, C. M., Harris,  
461 F., and Roy, D. B. (2015) ‘Ecological monitoring with citizen science : the design and implementation  
462 of schemes for recording plants in Britain and Ireland’, *Biological Journal of the Linnean Society*, pp.  
463 505–521.
- 464 Pocock, M. J. O. and Evans, D. M. (2014) ‘The Success of the Horse-Chestnut Leaf-Miner , *Cameraria*  
465 *ohridella* , in the UK Revealed with Hypothesis-Led Citizen Science’, *PLoS ONE*, 9(1), pp. 1–9. doi:  
466 10.1371/journal.pone.0086226.
- 467 Pocock, M. J. O., Roy, H. E., Preston, C. D., and Roy, D. B. (2015) ‘The Biological Records Centre: A  
468 pioneer of citizen science’, *Biological Journal of the Linnean Society*, 115(3), pp. 475–493. doi:  
469 10.1111/bij.12548.
- 470 Ransom Hardison, D., Holland, W. C., Currier, R. D., Kirkpatrick, B., Stumpf, R., Fanara, T., Burris,  
471 D., Reich, A., Kirkpatrick, G. J., and Wayne Litaker, R. (2019) ‘Habscope: A tool for use by citizen  
472 scientists to facilitate early warning of respiratory irritation caused by toxic blooms of *Karenia brevis*’,  
473 *PLoS ONE*, 14(6), pp. 1–17. doi: 10.1371/journal.pone.0218489.
- 474 Rich, T. G. C. and Woodruff, E. R. (1990) ‘BSBI monitoring scheme 1987-1988’, *Chief Scientist’s*  
475 *Directorate Report*, 1265.
- 476 Robinson, R. A., Lawson, B., Toms, M. P., Peck, K. M., Kirkwood, J. K., Clatworthy, I. R., Evans, A.  
477 D., Hughes, L. A., Hutchinson, O. C., Shinto, K., Pennycott, T. W., Perkins, M. W., Rowley, P. S.,  
478 Simpson, V. R., Tyler, K. M., and Cunningham, A. A. (2010) ‘Emerging Infectious Disease Leads to  
479 Rapid Population Declines of Common British Birds’, *PLoS ONE*, 5(8). doi:  
480 10.1371/journal.pone.0012215.
- 481 Schauwvlieghe, A. F. A. D., Rijnders, B. J. A., Philips, N., Verwijs, R., Vanderbeke, L., Tienen, C.  
482 Van, Lagrou, P. K., Verweij, P. P. E., Veerdonk, F. L. Van De, Gommers, P. D., ... Wauters, P. J.  
483 (2018) ‘Invasive aspergillosis in patients admitted to the intensive care unit with severe influenza : a  
484 retrospective cohort study’, *The Lancet Respiratory*. Elsevier Ltd, 6(10), pp. 782–792. doi:  
485 10.1016/S2213-2600(18)30274-1.

- 486 Sewell, T. R., Zhang, Y., Brackin, A. P., Shelton, J. M. G., Rhodes, J., and Fisher, M. C. (2019)  
487 ‘Elevated Prevalence of Azole-Resistant *Aspergillus fumigatus* in Urban versus Rural Environments in  
488 the United Kingdom’, *Antimicrobial Agents and Chemotherapy*, (August), pp. 1–8.
- 489 Snik, F., Rietjens, J. H. H., Apituley, A., Volten, H., Mijling, B., Di Noia, A., Heikamp, S., Heinsbroek,  
490 R. C., Hasekamp, O. P., Smit, J. M., Vonk, J., Stam, D. M., Van Harten, G., De Boer, J., and Keller, C.  
491 U. (2014) ‘Mapping atmospheric aerosols with a citizen science network of smartphone  
492 spectropolarimeters’, *Geophysical Research Letters*, 41(20), pp. 7351–7358. doi:  
493 10.1002/2014GL061462.
- 494 Sparks, T. H., Atkinson, S., Lewthwaite, K., Dhap, R., and Moran, N. J. (2017) ‘Can bird abundance  
495 declines be detected by citizen science programmes? A case study using Common Cuckoo *Cuculus*  
496 *canorus*’, *Avian Biology Research*, 10(4), pp. 241–245. doi: 10.3184/175815617X15036738758862.
- 497 Tsitsopoulou, A., Posso, R., Vale, L., Bebb, S., Johnson, E., and White, P. L. (2018) ‘Determination of  
498 the prevalence of triazole resistance in environmental *Aspergillus fumigatus* strains isolated in South  
499 Wales, UK’, *Frontiers in Microbiology*, 9(JUN), pp. 1–8. doi: 10.3389/fmicb.2018.01395.
- 500 Wilson, J. F., Baker, D., Cheney, J., Cook, M., Ellis, M., Freestone, R., Gardner, D., Geen, G.,  
501 Hemming, R., Hodggers, D., Howarth, S., Jupp, A., Lowe, N., Orridge, S., Shaw, M., Smith, B., Turner,  
502 A., and Young, H. (2018) ‘A role for artificial night-time lighting in long-term changes in populations  
503 of 100 widespread macro-moths in UK and Ireland: a citizen-science study’, *Journal of Insect*  
504 *Conservation*. Springer International Publishing, 22(2), pp. 189–196. doi: 10.1007/s10841-018-0052-  
505 1.
- 506