Evolution of sex allocation plasticity and its predictors in a flatworm genus

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Abstract

Local sperm competition (LSC), which occurs when related sperm compete for access to a given set of eggs, can influence sex allocation (SA) in simultaneous hermaphrodites. Different factors that have been predicted to affect LSC—and hence, optimal SA—include the mating strategy (such as reciprocal mating vs. hypodermic insemination) and the ability to self-fertilize. Moreover, the level of LSC experienced could vary temporally and/or spatially, favouring the evolution of SA plasticity. However, it is unknown how the mating strategy and ability to self-fertilize can influence SA and SA plasticity. Here, we explored this by collecting estimates of SA and SA plasticity for seven species of the flatworm genus, *Macrostomum*, and performing analyses while controlling for phylogeny. While we found substantial interspecific variation in SA, neither the mating strategy nor the ability to self-fertilize had a significant effect on SA. Next, we estimated standardized effect sizes for SA plasticity in response to i) the presence of mating partners and ii) the strength of LSC for each species, and tested if the mating strategy or ability to self-fertilize predicted these effect sizes. We found considerable interspecific variation in SA plasticity, with some species being highly plastic and others showing no significant plasticity. Moreover, self-fertilization predicted SA plasticity with respect to the presence of mating partners, with SA plasticity being lower for self-fertilizing species. Finally, we showed that interspecific variation in SA is nearly three times higher than intraspecific variation, suggesting that SA estimates in field-collected specimens can, at least to some degree, be interpreted as representing interspecific rather than intraspecific variation. Our study suggests that both SA and SA plasticity can evolve rapidly, with self-fertilization being an important predictor for the latter in *Macrostomum*.

59 *Keywords*

60 phenotypic plasticity, sperm competition, self-fertilisation, reciprocal mating, hypodermic

61 insemination, local mate competition

Introduction

Sex allocation (SA) theory in simultaneous hermaphrodites predicts the optimal allocation of resources towards the male versus female function (Charnov 1982). Local sperm competition (LSC), which can be thought of as the inverse of sperm competition (Parker 1970, 1998), occurs when related sperm (usually from the same individual) compete for access to a given set of eggs, and it can influence the optimal SA in simultaneous hermaphrodites (Schärer 2009; Schärer and Pen 2013). An example of a factor that can affect LSC, and hence the optimal SA, is the mating group size (Charnov 1980, 1982), denoted as $K+1$, where K is the number of sperm donors from which a sperm recipient receives sperm at the time its eggs are fertilized. Specifically, Charnov's mating group size model predicts an influence of the mating group size on SA, such that the optimal proportion of resources allocated to the male function in an individual is predicted to increase with the number of sperm donors that are present in a local mating group.

The link between optimal SA and LSC can be visualised in terms of male fitness gain curves, which describe how much fitness is gained through the male function per unit resource investment (Charnov 1982; Schärer 2009). In small groups, e.g. under monogamy, LSC is necessarily very high, since the competing sperm are maximally related to each other, resulting in the male fitness gain curve saturating quickly. Thus, any investment into the production of more sperm than are required to fertilize the partner's eggs is a waste of resources, as the related sperm simply compete amongst themselves for fertilizations. Instead, these resources could be more profitably invested into the individual's own female function (Charnov 1982), since the female fitness gain curve is often assumed to be non-saturating and linear (Schärer 2009). Consequently, under monogamy, a simultaneous hermaphrodite is expected to have a female-biased SA (Charnov 1980, 1982). But as the mating group size increases—with more competitors contributing sperm to a given recipient—an individual's

sperm are more and more competing with unrelated sperm. So it now pays off to invest in the

male function to gain a greater share of the fertilizations, and the male fitness gain curve is

predicted to linearize, with a subsequent shift towards a more equal SA (Charnov 1982;

Schärer 2009; Schärer and Pen 2013).

An interesting question that arises then is whether and how interspecific variation in

reproductive biology, including different mating strategies or the ability to self-fertilize, could

influence the strength of LSC and hence the optimal SA. For example, hermaphrodites often

evolve different mating strategies in response to sexual conflict over mating roles, i.e. over

whether to mate as a sperm donor and/or as a sperm recipient (Charnov 1979; Michiels 1998;

Schärer et al. 2015). One such mating strategy is reciprocal mating (also called reciprocal

copulation), in which both partners mate in both mating roles, and thus donate and receive

sperm simultaneously (Michiels 1998; Schärer et al. 2015). This strategy can lead to

individuals being quite willing to engage in mating in order to get an opportunity to donate

sperm, and this general willingness likely results in reduced precopulatory sexual selection.

The ensuing higher mating rates could in turn result in increased sperm competition (and thus

a decrease in LSC) and more intense postcopulatory sexual selection. However, the presence

of certain postcopulatory processes could also increase LSC, e.g. by leading to the removal of

sperm of one or multiple donors from competition (Schärer 2009). Indeed, theoretical studies

have predicted that different postcopulatory sexual selection processes, such as cryptic female

choice, sperm digestion, and sperm displacement, can result in changes in LSC and hence the

predicted optimal SA (Charnov 1996; Pen and Weissing 1999; van Velzen et al. 2009;

Schärer and Pen 2013).

Moreover, it has been suggested that this intense postcopulatory sexual selection and sexual

conflict can lead to selection for traits and mating strategies that allow a donor to bypass these

postcopulatory processes and fertilize the eggs more directly (Charnov 1979; Michiels 1998).

One such mating strategy is forced unilateral insemination, in which one partner mates only in

the male role and donates sperm, e.g. via traumatic or hypodermic insemination (Lange et al.

2013; Reinhardt et al. 2015), while the other only mates in the female role, likely often

against its interests (Charnov 1979; Michiels 1998; Schärer et al. 2015). It is currently

difficult to predict the direction and magnitude of changes in LSC resulting from such a shift

in mating strategy—and the effects that such shifts may have on optimal SA—since we still

know relatively little about the particular postcopulatory processes involved in these mating

strategies. In species with reciprocal mating there may be some control over who or how

many partners an individual receives sperm from, and there could be strategies that lead to the

(possibly selective) removal of received sperm (van Velzen et al. 2009), possibly leading to

increased LSC. In species with hypodermic insemination such control might be more limited,

since there sperm function in a less controlled environment in which sperm from multiple

donors are more likely to mix randomly in something more closely resembling a fair-raffle-

type sperm competition, which would be expected to lower the strength of LSC (Schärer and

Janicke 2009).

Species that employ self-fertilization might differ from species that primarily outcross, since

self-fertilization will generally increase the strength of LSC and hence reduce the fitness

returns from investing in the male function, thus favouring the evolution of a more female-

biased optimal SA compared to outcrossing species (Charlesworth and Charlesworth 1981;

Charnov 1982, 1987), which is supported by empirical work in both plants and animals

(Lemen 1980; Schoen 1982; Charnov 1987; McKone 1987; Johnston et al. 1998). In general,

we would expect any process that has an effect on LSC to affect the shape of the male fitness

gain curve, and hence the optimal SA (Schärer 2009).

While most SA models investigate the effect of LSC on SA over evolutionary timescales,

LSC can also vary temporally or/and spatially within an individual's lifetime. In

hermaphrodites, altering the SA in response to the current conditions can influence the

immediate reproductive success of an individual, favouring the evolution of SA plasticity.

Indeed, plastic SA has been suggested to be one of the advantages of hermaphroditism over

separate-sex species (Charnov 1982; Michiels 1998, 1999). However, for species that do not

often experience variation in LSC during their lifetime, we may not expect high levels of

plasticity, particularly if there are costs to plasticity (DeWitt et al. 1998; Auld et al. 2010;

Siljestam and Östman 2017). Interestingly, while SA plasticity has now been documented in

many hermaphroditic species (Schärer 2009), its evolution is still comparatively poorly

understood, with few studies having investigated variation in SA plasticity across animals

species in a controlled experimental context (Schleicherová et al. 2014). Moreover, as SA can

vary both within-species (e.g. due to phenotypic plasticity) and between-species (e.g. due to

differences in mating strategy over evolutionary timescales), it is interesting to examine the

relative magnitude of the interspecific versus intraspecific variation in SA.

An excellent model system for testing the effect of the mating strategy and the ability to self-

fertilize on SA and SA plasticity is the free-living flatworm genus *Macrostomum*

(Macrostomorpha, Platyhelminthes). This genus contains many species exhibiting one if at

least two different mating strategies; one involving reciprocal mating and the other

hypodermic insemination (Brand et al., in prep.; Schärer et al. 2011). In reciprocally mating

species, a facultative postcopulatory 'suck' behaviour has been observed that has been

hypothesised to remove received ejaculate (Singh et al., in prep.; Schärer et al. 2004a, 2011;

Vizoso et al. 2010; Marie-Orleach et al. 2013). Remarkably, no such postcopulatory suck

behaviour has been documented in five hypodermically inseminating species, which

presumably exhibit forced unilateral mating, with sperm being hypodermically injected into

the partner via a needle-like male copulatory organ (Schärer et al. 2011). Interestingly,

hypodermic insemination is also associated with a suite of morphological traits that

potentially allow self-fertilisation (Ramm et al. 2012, 2015; Giannakara and Ramm 2017),

although self-fertilization has recently also been documented in at least one reciprocally

mating species, *M. mirumnovem* (Singh et al. 2020b).

Here, we collected data on SA estimates from a range of experiments performed in seven

Macrostomum species (including in a new and currently still undescribed species,

Macrostomum sp. 22). The experimental layout generally consisted of raising worms in three

different group sizes (isolated, pairs or octets) and then obtaining estimates of SA once the

worms had matured. To facilitate SA plasticity comparisons between species, we calculated

standardised effect sizes for SA plasticity in response to i) the presence of mating partners

(i.e. comparing isolated worms vs. worms with partners) and ii) the strength of LSC (i.e.

comparing paired worms vs. octet worms). Using SA estimates and SA plasticity effect sizes,

we then examined if the mating strategy and ability to self-fertilize predicted SA or SA

plasticity, while accounting for the phylogenetic interrelationships. Lastly, considering the

interspecific SA differences and the extensive SA plasticity shown by some of the species, we

examined how much SA varies within-species versus between-species by partitioning the SA

variation into its intraspecific and interspecific components. This would also allow us to more

confidently interpret SA variation observed in field-collected worms as at least partially

representing interspecific differences.

Materials and Methods

Study species

To examine how SA and SA plasticity evolves across the *Macrostomum* genus, we gathered

data on SA and SA plasticity in seven *Macrostomum* species: three that are hypodermically

inseminating, *M. pusillum* Ax 1951 (Giannakara and Ramm 2017), *M. hystrix* Ørsted 1843

sensu Luther 1905 (Winkler and Ramm 2018), and *Macrostomum* sp. 22 (reported in the

Supplementary Information S1); and four that are reciprocally mating, namely *M. janickei*

Schärer 2019, *M. cliftonensis* Schärer & Brand 2019, and *M. mirumnovem* Schärer & Brand

2019 (Schärer et al. 2020), plus multiple studies in *M. lignano* Ladurner, Schärer,

Salvenmoser & Rieger 2005 (see below). *M. mirumnovem* and all three hypodermically

inseminating species can self-fertilize (Ramm et al. 2012, 2015; Giannakara and Ramm 2017;

Singh et al. 2020; this study). We classified species as either reciprocally mating or

hypodermically inseminating using both behavioural and morphological data. Behavioural

data showed the presence of both reciprocal mating and the postcopulatory suck behaviour in

M. lignano (also see Schärer et al. 2004a)*, M. janickei*, *M. cliftonensis*, and *M. mirumnovem*

(Schärer et al. 2020), while neither of these behaviours were observed in *M. pusillum*,

M. hystrix, and *Macrostomum* sp. 22 (Singh et al., in prep.). In addition, the classification of

species using morphological traits, which is known to be correlated with the mating strategy

in *Macrostomum* (Schärer et al. 2011)*, corroborated our above classification based on

behaviour. Briefly, morphologically, species were classified as reciprocal mating or

hypodermically inseminating depending on their male and female genital morphology, the

sperm morphology, and the location of (received) allosperm (Brand et al., in prep.).

SA estimates and SA plasticity effect sizes across species

For most species the experimental procedure was similar to what we describe for

Macrostomum sp. 22 (Supplementary Information S1), i.e., worms were raised from juveniles

in three different group sizes (isolated, pairs or octets) until adulthood, when testis and ovary

size was measured in one randomly chosen worm per replicate. The SA was estimated as

testis size/(testis size+ovary size). In some experiments, the density of worms was varied

independently of group size by using two enclosure sizes (small and large) (for details and

sample sizes see Supplementary Table S2). We expect that the experiments are nevertheless

comparable, since all experiments where density was included as a factor revealed that it did

not have a significant effect on our estimate of SA.

For each experiment, we calculated the average SA across the group sizes, weighted by the

sample size for each group size. For SA plasticity estimates, we calculated standardised effect

sizes, using Cohen's d (Cohen 1988) with Hedges correction for small sample size (Hedges

1981), including confidence intervals (Howell 2011) of the effect sizes using the R package

'effsize' (Torchiano 2017), to facilitate interspecies comparisons. For each species, we

calculated an effect size for SA plasticity to assess the effect of the presence of mating

partners (i.e. comparing isolated worms vs. worms with partners) and the strength of LSC (i.e.

comparing worms in pairs vs. worms in octets). Pairs represent conditions with high LSC

whereas octets represent conditions with low LSC.

For *M. lignano*, we collected datasets from multiple published (up to 25th November 2019)

experiments, and calculated SA and SA plasticity effect sizes for each of them separately. In

total, we found two and six studies, respectively, where we could extract effect of the

presence of mating partners (Schärer and Janicke 2009; Ramm et al. 2019) and the strength of

LSC (Schärer and Ladurner 2003; Schärer et al. 2004b; Janicke and Schärer 2009; Janicke et

al. 2013; Marie-Orleach et al. 2014; Ramm et al. 2019). For Schärer and Ladurner (2003), we

calculated SA as testis size/(testis size+ovary size), since that study only gives values for

testis and ovary size, but not SA. We excluded one previous study (Janicke and Schärer 2010)

from our analysis, since the worms used in that study came from the same experiment as

Janicke and Schärer (2009), and so these were not independent datasets (we used the latter

study since it had more replicates). In addition, in Marie-Orleach et al. (2014) we combined

the data from the two different inbred lines (HUB1 and DV1), since they did not differ in any

of the traits measured. Also, note that in Schärer and Janicke (2009), there are only isolated

and paired worms and hence the estimated effect size for the presence of mating partners does

not include the effect of octets. This should not have a large effect on the calculated effect

size, since in most cases the difference between isolated and pairs is much larger than the

difference between paired and octet treatments (Figure 1).

For *M. lignano*, a weighted average across experiments was used to estimate the weighted

SA, and estimated SA plasticity effect sizes (Turner and Bernard 2006) for both the presence

of mating partners and the strength of LSC. For SA, we took the average of the weighted SA

across experiments, weighted by the total sample size of that experiment. To weight each

effect size, it was multiplied by the inverse of its variance (inverse variance weight, IVW),

which allowed us to take into consideration the differences in sample size across experiments,

namely:

$$247 \quad Hedges's\ g^* = \frac{\sum_{i=1}^{n} Hedges's\ g_i \times IVW_{Hedges's\ g_i}}{\sum_{i=1}^{n} IVW_{Hedges's\ g_i}}$$

248 And its standard error was calculated as:

$$249 \quad SE_{g^*} = \sqrt{\frac{1}{\sum_{i=1}^{n} IVW_{Hedges's\ g_i}}}$$

Evolution of SA and SA plasticity

For the phylogenetically corrected analyses of the SA and SA plasticity effect sizes, we used a

trimmed (*ape* package, Paradis et al. 2004) ultrametric phylogenetic tree of the *Macrostomum*

genus, which is based on large-scale phylogenomic analyses of *de novo* transcriptomes (C-IQ-

TREE, Brand et al., in prep.). We estimated the phylogenetic signal using Blomberg's K

(*picante* package, Blomberg et al. 2003; Kembel et al. 2010) and Pagel's λ (*phytools* package,

Pagel 1999; Revell 2012), for SA and both SA plasticity effect sizes. For both estimates of

phylogenetic signal, a value close to zero is suggestive of phylogenetic independence, while a

value close to 1 suggests that the traits evolve under Brownian motion. In our case, the

estimates of phylogenetic signal for SA showed a phylogenetic signal, while the two SA

plasticity effect sizes did not differ significantly from 0 (Supplementary Table S3). Since our

sample sizes are small, the likelihood ratio tests used to assess Pagel's λ can be unreliable

(Boettiger et al. 2012), as the asymptotic properties of maximum-likelihood estimation may

not hold for small sample sizes. Thus, we present both phylogenetically corrected analyses

and analyses without correcting for phylogeny.

For the phylogenetically corrected analyses, we compared the fit of different character

evolution models (i.e. Brownian motion, Ornstein–Uhlenbeck, and Early-burst) (*geiger*

package, Harmon et al. 2008) for SA and each effect size, and found that for each case

Brownian motion was the best character evolution model (see Supplementary Table S4).

Thus, we used Brownian motion as the preferred model for the subsequent analysis. Using

phylogenetic generalized least squares (PGLS) regressions and a Brownian motion model of

character evolution (*nlme* package, Pinheiro et al. 2014), we tested if the mating strategy

(reciprocal mating vs. hypodermic insemination) or self-fertilization (presence or absence)

predict SA, or the two SA plasticity effect sizes (presence of mating partners and strength of

LSC). PGLS allows us to account for the phylogenetic non-independence of observations

resulting from common evolutionary history of species. And for the analyses without

correcting for phylogeny, we calculated Wilcoxon rank-sum tests to assess if the mating

strategy or self-fertilization predicted the differences in SA or SA plasticity effect sizes.

Finally, to divide the observed variance in SA into a within-species and a between-species

component, we used the SA (weighted SA for *M. lignano*) for each group for each species

(thus, 3 observations per species), and fit a linear mixed model (*lme4* package, Bates et al.

2015) using species as a random effect and calculated the percentage of variance explained by

species.

All statistical analyses were carried out using R, version 3.4.1 (R Core Team 2017) unless

stated otherwise.

Results

SA and effect sizes of SA plasticity for all species

There was substantial variation in overall SA across the seven species (Figure 1,

Supplementary Table S2), both with respect to the mating strategy (solid vs. stippled lines)

and self-fertilization (circles vs. triangles). Two of the three hypodermically inseminating

species (i.e. *Macrostomum* sp. 22 and *M. pusillum)*, exhibited a low SA. Similarly, and in line

with our predictions, three of the four self-fertilizing species (*Macrostomum* sp. 22,

M. pusillum and *M. mirumnovem*), exhibited a relatively low SA, likely indicating that SA in

these species was female-biased. Interestingly, the exception to this pattern was *M. hystrix*,

which had the highest SA of all the studies species. There was also interspecific variation in

the effect size estimates of SA plasticity, even among relatively closely related species

(Figure 2, Supplementary Table S2). *M. cliftonensis* and *M. lignano* exhibited the highest SA

plasticity in response to the presence of mating partners and the strength of LSC, respectively.

In addition, *M. lignano* exhibited SA plasticity across all experiments and while the

magnitude varied somewhat, the confidence intervals overlapped, indicating that the estimates

were fairly consistent across experiments.

Evolution of SA and SA plasticity

The PGLS models (see Supplementary Table S3 and S4 for more details) showed that neither

the mating strategy ($t_{7,5}$ = -0.39, P = 0.70) nor the ability to self-fertilize ($t_{7,5}$ = -1.03, P =

0.34) significantly predicted SA. And similar to SA, the mating strategy also did not

significantly predict the SA plasticity effect size, neither due to the presence of mating

partners ($t_{7,5}$ = 0.94, P = 0.39) nor the strength of LSC ($t_{7,5}$ = 0.23, P = 0.82). In contrast, self-

fertilization significantly predicted SA plasticity with respect to the presence of mating

partners ($t_{7,5}$ = -3.07, P = 0.03), but not due to the strength of LSC ($t_{7,5}$ = -0.57, P = 0.59). SA

plasticity with respect to the presence of mating partners was lower for selfing species

(compare selfing vs. non-selfing species of green symbols in Figure 2). And essentially the

same results were observed in the analyses without correcting for phylogeny using Wilcoxon

rank-sum tests (Supplementary Table S5).

Lastly, the linear-mixed model (Intercept = 0.55±0.05, t_{value} = 10.38, 21 observations for 7

species) showed that between-species variance explained 73.6% of the total SA variance

(interspecific variance = 0.0177, and residual variance = 0.0063), suggesting that SA variance

between-species is nearly three times larger than variance within-species.

Discussion

Our study showed that there was substantial interspecific variation in both SA and SA

plasticity, also among relatively closely related species, suggesting that both SA and SA

plasticity can evolve rapidly in the *Macrostomum* genus. Moreover, although the mating

strategy did not predict SA or either of the SA plasticity effect sizes, self-fertilizing species

had a lower SA plasticity in response to presence of mating partner. In the following, we

discuss these findings in some detail.

SA and effect sizes of SA plasticity for all species

Very few studies have explored predictions of SA theory across multiple species in

hermaphroditic animals in the context of phenotypic plasticity (Hoch and Levinton 2012;

Schleicherová et al. 2014), with most studies focussing solely on intraspecific comparisons

(Schärer 2009). Hoch and Levinton (2012) tested Charnov's (1980, 1982) mating group size

model in two species of acorn barnacles, *Semibalanus balanoides* and *Balanus glandula,* by

varying both the number and density of competitors in a natural setting. They showed that

both species exhibit increased allocation in absolute, but not relative, male allocation

(estimated using the sum of the mass of testes, sperm, seminal vesicle, and penis) at higher

densities. Interestingly, the species differed in how they responded to the treatments in terms

of their female allocation (estimated using egg mass), with both the number and density of

competitors having an effect in *S. balanoides*—although in the opposite direction than

predicted by the model, while there was no effect in *B. glandula.* The authors proposed that

this might result from interspecific differences in life-history traits. In contrast to the acorn

barnacles, a study in three related polychaete worms, *Ophryotrocha diadema, O. adherens*

and *O. gracilis*, showed that while there were interspecific differences in the plasticity in

female allocation (using multiple estimates, such as resource investment in eggs, total number

of egg cocoons, and time interval between egg laying) in response to different numbers of

mating partners, there was no significant plasticity in the male function (estimated as the

number of sperm produced by an individual) across the species (Schleicherová et al. 2014).

The authors proposed that the magnitude of plasticity depended on the species-specific costs

of the sex function, and also on the mating system of the species. So, while both of these

studies document interspecific variation in SA plasticity, they lacked the statistical power to

explore whether interspecific differences in reproductive traits may affect the level of SA

plasticity.

Our results showed that both SA and SA plasticity varied considerably across the studied

Macrostomum species. Both hypodermically inseminating and self-fertilizing species tended

to show a low (i.e. more female-biased) SA, with the notable exception of *M. hystrix*.

M. hystrix represents an independent origin of hypodermic insemination in the reciprocally

mating clade (Schärer et al. 2011), and it might therefore still retain characteristics of its

reciprocally mating ancestors, which in the current sample of species appear to have relatively

high (i.e. more male-biased) SA (see Figure 1), and our analyses suggested some level of

phylogenetic signal in SA. Moreover, it has been argued that hypodermic insemination may

lead to sperm competing more in a fair-raffle-type scenario than with reciprocal mating

(Schärer and Janicke 2009), which might lead to a SA that more closely matches the group

size. In addition, *M. hystrix* is known to reach very high densities in the field (L. Schärer,

pers. obs.), which could also favour the evolution of a high SA.

M. lignano and *M. janickei,* which are sibling species capable of hybridization (Singh et al.

2020a), show clear differences in SA plasticity in response to the strength of LSC, with

M. lignano being more plastic (although the 95% confidence intervals overlap compared to

two of the *M. lignano* studies). Variation in SA plasticity in species with similar reproductive

biology could stem from different environmental conditions experienced by the species. For

example, evolution of SA plasticity might not be favoured in species that inhabit relatively

stable environments, especially if the maintenance of plasticity is costly (DeWitt et al. 1998;

Auld et al. 2010; Siljestam and Östman 2017). Even in the absence of maintenance costs,

there could still be production costs of plasticity, though if the benefits of adjusting SA

outweigh these costs, then phenotypic plasticity could still be maintained. This cost-benefit

ratio of SA plasticity can vary across species (Van Buskirk 2002; Steiner 2007) and

environments (Ratikainen and Kokko 2019), leading to the retention or loss of plasticity in a

given species. Moreover, such costs of SA plasticity might play an important role in both the

evolution and the maintenance of simultaneous hermaphroditism (St. Mary 1997), and could

also constrain changes in SA leading to a suboptimal SA. A study in *O. diadema* showed that

there were no large short-term fitness costs of sex adjustment for the species (Lorenzi et al.

2008), while in *M. lignano* production costs of SA plasticity have been documented (Sandner

2013), with an alternating group size environment leading to a lower hatchling production

compared to a stable group size environment. We currently do not have data on production

costs of plasticity in *M. janickei* when exposed to a stable vs. fluctuating environment.

Across all species, the estimates of plasticity effect sizes are larger for the presence of mating

partners than for the strength of LSC (number of mating partners), and for *M. cliftonensis* and

M. janickei, the confidence intervals of the two effect size estimates do not overlap. This

suggests that the increase in SA going from paired to octet groups is not as drastic as going

from isolated worms to worms in larger groups. A similar phenomenon has been seen in a

freshwater snail, *Lymnaea stagnalis*, where paired snails showed higher expression of six

seminal fluid protein (SFP) genes compared to isolated snails, while the SFP expression of

snails in larger (6 individuals) groups was similar to that of paired snails (Nakadera et al.

2019). In our study, this situation could potentially arise if in the presence of multiple partners

in octets, worms do not only increase their testis size, but also their rate of sperm production

per unit testis size (Schärer and Vizoso 2007), which in *M. lignano* has been shown to occur

due to an increased speed of spermatogenesis (Giannakara et al. 2016). If such effects occur

generally (but see Giannakara & Ramm 2017 for *M. pusillum*, a species that lacks SA

plasticity, see also below), we might underestimate male allocation when solely measuring

testis size. Alternatively, there could also be increased investment in other components of the

male function (Schärer and Pen 2013), e.g. seminal fluid production (Patlar and Ramm 2019;

Patlar et al. 2019; Ramm et al. 2019), or mating behaviour (Janicke and Schärer 2009). A

study in *O. diadema* showed that increased mating opportunities are accompanied by higher

investment into male function, such as aggressive behaviour (Santi et al. 2018).

Evolution of SA and SA plasticity

The SA plasticity effect sizes did not differ significantly between species exhibiting the

reciprocal and hypodermic mating strategies, while self-fertilizing species had a lower SA

plasticity (in response to presence of mating partner, but not the strength of LSC). Interesting

questions that arise then are about the causes of the variation in SA and SA plasticity between

the self-fertilizing species. While there has been both theoretical and empirical work

406 exploring the effect of self-fertilization on SA in plants (Lemen 1980; Charlesworth and

407 Charlesworth 1981; Schoen 1982; Charnov 1987; McKone 1987; Brunet 1992), there have

408 been few such studies in animals (Johnston et al. 1998). One possible explanation for

409 variation between self-fertilizing species could be if the species differ in their rate and pattern

410 of self-fertilization, which could have an effect on both optimal SA and SA plasticity. For

411 example, in obligatorily self-fertilizing species, we would expect that they do not increase

412 their SA with group size, since LSC would be expected to remain high irrespective of the

413 group size. On the other hand, for species that exhibit facultative self-fertilization, we might

414 expect SA plasticity despite a low overall SA.

415 Interestingly, our results do conform somewhat to this pattern, with *M. pusillum*—which has

416 been hypothesised to be obligatorily self-fertilizing (Giannakara and Ramm 2017)—showing

417 both the lowest overall SA and no SA plasticity. Its congener, *Macrostomum* sp. 22, showed a

418 similarly female-biased SA in isolated and paired worms, but significant SA plasticity

419 between worms in pairs and octets. If worms shifted from self-fertilization in isolation to only

420 outcrossing in pairs, then the strength of LSC would not be expected to change, while the

421 strength of LSC would clearly be expected to drop from worms in pairs to octets, a scenario

422 that would match the observed SA plasticity patterns. This could suggest that

423 *Macrostomum* sp. 22 may only self in isolation. In contrast, *M. hystrix*—thought to be a

424 preferentially outcrossing species, with studies showing delayed selfing, at least in some

425 populations, as well as costs of self-fertilization (Ramm et al. 2012; Giannakara and Ramm

426 2020)—was found to be more plastic than *M. pusillum*.

427 Finally, our results showed that there was nearly three times more variation in SA between-

428 species than within-species in our experiments. This is remarkable, since we saw extensive

429 plasticity in SA in certain species, such as across the multiple experiments in *M. lignano*.

430 Thus, this result suggests that we can interpret variation in SA among field collected worms

431   of different *Macrostomum* species as being at least partially due to interspecific variation,

432   even if they come from different social environments. This therefore facilitates the study of

433   the evolution of SA patterns across species that cannot currently be cultured in the laboratory

434   (Brand et al., in prep.). Collectively, our results suggest that SA and SA plasticity can evolve

435   rapidly in *Macrostomum*, with self-fertilization being an important predictor for the latter.

References

Auld, J. R., A. A. Agrawal, and R. A. Relyea. 2010. Re-evaluating the costs and limits of
adaptive phenotypic plasticity. Proc. R. Soc. B Biol. Sci. 277:503–511.
Ax, P. 1951. Die Turbellarien des Eulitorals der Kieler Bucht. Zool. Jahrbuecher Abteilung
fuer Syst. Oekologie und Geogr. der Tiere 80:277–378.
Bates, D., M. Mächler, B. M. Bolker, and S. C. Walker. 2015. Fitting linear mixed-effects
models using lme4. J. Stat. Softw., doi: 10.18637/jss.v067.i01.
Blomberg, S. P., T. Garland, and A. R. Ives. 2003. Testing for phylogenetic signal in
comparative data: behavioral traits are more labile. Evolution (N. Y). 57:717–745.
Boettiger, C., G. Coop, and P. Ralph. 2012. Is your phylogeny informative? Measuring the
power of comparative methods. Evolution (N. Y). 66:2240–2251.
Brunet, J. 1992. Sex allocation in hermaphroditic plants. Trends Ecol. Evol. 7:79–84.
Charlesworth, D., and B. Charlesworth. 1981. Allocation of resources to male and female
functions in hermaphrodites. Biol. J. Linn. Soc. 15:57–74.
Charnov, E. L. 1987. On sex allocation and selfing in higher plants. Evol. Ecol. 1:30–36.
Charnov, E. L. 1980. Sex allocation and local mate competition in barnacles. Mar. Biol. Lett.
1:269–272.
Charnov, E. L. 1979. Simultaneous hermaphroditism and sexual selection. Proc. Natl. Acad.
Sci. 76:2480–2484.
Charnov, E. L. 1982. The theory of sex allocation. Monogr. Popul. Biol. 18:1–355. Princeton
University Press, Cambridge.
Cohen, J. 1988. Statistical power analysis for the behavioral sciences.
DeWitt, T. J., A. Sih, and D. S. Wilson. 1998. Costs and limits of phenotypic plasticity.
Trends Ecol. Evol. 13:77–81.
Giannakara, A., and S. A. Ramm. 2020. Evidence for inter-population variation in waiting
times in a self-fertilizing flatworm. Invertebr. Reprod. Dev. 64:158–168.
Giannakara, A., and S. A. Ramm. 2017. Self-fertilization, sex allocation and spermatogenesis
kinetics in the hypodermically inseminating flatworm *Macrostomum pusillum*. J. Exp.
Biol. 220:1568–1577.
Giannakara, A., L. Schärer, and S. A. Ramm. 2016. Sperm competition-induced plasticity in
the speed of spermatogenesis. BMC Evol. Biol. 16:60.
Harmon, L. J., J. T. Weir, C. D. Brock, R. E. Glor, and W. Challenger. 2008. GEIGER:
investigating evolutionary radiations. Bioinformatics 24:129–131.
Hedges, L. V. 1981. Distribution theory for glass's estimator of effect size and related
estimators. J. Educ. Stat. 6:107–128.
Hoch, J. M., and J. S. Levinton. 2012. Experimental tests of sex allocation theory with two

species of simultaneously hermaphroditic acorn barnacles. Evolution (N. Y). 66:1332–
1343.
Howell, D. C. 2011. Confidence intervals on effect size. Univresity of Vermont, doi:
10.1134/S0362119713030110.
Janicke, T., L. Marie-Orleach, K. De Mulder, E. Berezikov, P. Ladurner, D. B. Vizoso, and L.
Schärer. 2013. Sex allocation adjustment to mating group size in a simultaneous
hermaphrodite. Evolution (N. Y). 67:3233–3242.
Janicke, T., and L. Schärer. 2009. Sex allocation predicts mating rate in a simultaneous
hermaphrodite. Proc. R. Soc. B Biol. Sci. 276:4247–4253.
Janicke, T., and L. Schärer. 2010. Sperm competition affects sex allocation but not sperm
morphology in a flatworm. Behav. Ecol. Sociobiol. 64:1367–1375.
Johnston, M. O., B. Das, and W. R. Hoeh. 1998. Negative correlation between male
allocation and rate of self-fertilization in a hermaphroditic animal. Proc. Natl. Acad. Sci.
95:617–620.
Kembel, S. W., P. D. Cowan, M. R. Helmus, W. K. Cornwell, H. Morlon, D. D. Ackerly, S.
P. Blomberg, and C. O. Webb. 2010. Picante: R tools for integrating phylogenies and
ecology. Bioinformatics 26:1463–1464.
Lange, R., K. Reinhardt, N. K. Michiels, and N. Anthes. 2013. Functions, diversity, and
evolution of traumatic mating. Biol. Rev. 88:585–601.
Lemen, C. 1980. Allocation of reproductive effort to the male and female strategies in wind-
pollinated plants. Oecologia 45:156–159.
Lorenzi, M. C., D. Schleicherova, and G. Sella. 2008. Sex adjustments are not functionally
costly in simultaneous hermaphrodites. Mar. Biol. 153:599–604.
Marie-Orleach, L., T. Janicke, and L. Schärer. 2013. Effects of mating status on copulatory
and postcopulatory behaviour in a simultaneous hermaphrodite. Anim. Behav. 85:453–
461.
Marie-Orleach, L., T. Janicke, D. B. Vizoso, M. Eichmann, and L. Schärer. 2014. Fluorescent
sperm in a transparent worm: validation of a GFP marker to study sexual selection. BMC
Evol. Biol. 14:148.
McKone, M. J. 1987. Sex allocation and outcrossing rate: a test of theoretical predictions
using Bromegrasses (Bromus). Evolution (N. Y). 41:591.
Michiels, N. K. 1998. Mating Conflicts and Sperm Competition in Simultaneous
Hermaphrodites. Pp. 219–254 in Sperm Competition and Sexual Selection. Elsevier.
Michiels, N. K. 1999. Sexual adaptations to high density in hermaphrodites. Invertebr.
Reprod. Dev. 36:35–40.
Nakadera, Y., A. Giannakara, and S. A. Ramm. 2019. Plastic expression of seminal fluid
protein genes in a simultaneously hermaphroditic snail. Behav. Ecol. 30:904–913.
Pagel, M. 1999. Inferring the historical patterns of biological evolution. Nature 401:877–884.
Paradis, E., J. Claude, and K. Strimmer. 2004. APE: Analyses of Phylogenetics and Evolution
in R language. Bioinformatics 20:289–290.
Parker, G. A. 1970. Sperm competition and its evolutionary consequences in the insects. Biol.
Rev. 45:525–567.
Parker, G. A. 1998. Sperm Competition and the Evolution of Ejaculates: Towards a Theory
Base. Pp. 3–54 in Sperm competition and Sexual selection.
Patlar, B., and S. A. Ramm. 2019. Genotype-by-environment interactions for seminal fluid
expression and sperm competitive ability. J. Evol. Biol. jeb.13568.
Patlar, B., M. Weber, and S. A. Ramm. 2019. Genetic and environmental variation in
transcriptional expression of seminal fluid proteins. Heredity (Edinb). 122:595–611.
Pen, I., and F. J. Weissing. 1999. Sperm competition and sex allocation in simultaneous
hermaphrodites: A new look at Charnov's invariance principle. Evol. Ecol. Res. 517–

525.

Pinheiro, J., D. Bates, S. DebRoy, and D. Sarkar. 2014. R Core Team (2014). nlme: linear and nonlinear mixed effects models. R package version 3.1–117. URL http//cran. r-project. org/web/packages/nlme/index. html.

Ramm, S. A., B. Lengerer, R. Arbore, R. Pjeta, J. Wunderer, A. Giannakara, E. Berezikov, P. Ladurner, and L. Schärer. 2019. Sex allocation plasticity on a transcriptome scale: Socially sensitive gene expression in a simultaneous hermaphrodite. Mol. Ecol. mec.15077.

Ramm, S. A., D. B. Vizoso, and L. Schärer. 2012. Occurrence, costs and heritability of delayed selfing in a free-living flatworm. J. Evol. Biol. 25:2559–2568.

Ramm, S. a, A. Schlatter, M. Poirier, and L. Schärer. 2015. Hypodermic self-insemination as a reproductive assurance strategy. Proc. R. Soc. B Biol. Sci. 282:20150660.

Ratikainen, I. I., and H. Kokko. 2019. The coevolution of lifespan and reversible plasticity. Nat. Commun. 10:538.

Reinhardt, K., N. Anthes, and R. Lange. 2015. Copulatory wounding and traumatic insemination. Cold Spring Harb. Perspect. Biol. 7:a017582.

Revell, L. J. 2012. phytools: an R package for phylogenetic comparative biology (and other things). Methods Ecol. Evol. 3:217–223.

Sandner, P. 2013. Impacts of sperm competition on mating behaviour and life history traits in a simultaneous hermaphrodite. University of Basel.

Santi, M., L. Picchi, and M. C. Lorenzi. 2018. Dynamic modulation of reproductive strategies in a simultaneous hermaphrodite and preference for the male role. Anim. Behav. 146:87–96.

Schärer, L. 2009. Tests of sex allocation theory in simultaneously hermaphroditic animals. Evolution (N. Y). 63:1377–1405.

Schärer, L., J. N. Brand, P. Singh, K. S. Zadesenets, C.-P. Stelzer, and G. Viktorin. 2020. A phylogenetically informed search for an alternative *Macrostomum* model species, with notes on taxonomy, mating behavior, karyology, and genome size. J. Zool. Syst. Evol. Res. 58:41–65.

Schärer, L., and T. Janicke. 2009. Sex allocation and sexual conflict in simultaneously hermaphroditic animals. Biol. Lett. 5:705–708.

Schärer, L., T. Janicke, and S. A. Ramm. 2015. Sexual conflict in hermaphrodites. Cold Spring Harb. Perspect. Biol. 7:a017673.

Schärer, L., G. Joss, and P. Sandner. 2004a. Mating behaviour of the marine turbellarian Macrostomum sp.: these worms suck. Mar. Biol. 145:373–380.

Schärer, L., and P. Ladurner. 2003. Phenotypically plastic adjustment of sex allocation in a simultaneous hermaphrodite. Proc. R. Soc. B Biol. Sci. 270:935–941.

Schärer, L., P. Ladurner, and R. M. Rieger. 2004b. Bigger testes do work more: experimental evidence that testis size reflects testicular cell proliferation activity in the marine invertebrate, the free-living flatworm Macrostomum sp. Behav. Ecol. Sociobiol. 56:420–425.

Schärer, L., D. T. J. Littlewood, A. Waeschenbach, W. Yoshida, and D. B. Vizoso. 2011. Mating behavior and the evolution of sperm design. Proc. Natl. Acad. Sci. 108:1490–1495.

Schärer, L., and I. Pen. 2013. Sex allocation and investment into pre- and post-copulatory traits in simultaneous hermaphrodites: the role of polyandry and local sperm competition. Philos. Trans. R. Soc. Lond. B. Biol. Sci. 368:20120052.

Schärer, L., and D. B. Vizoso. 2007. Phenotypic plasticity in sperm production rate: There's more to it than testis size. Evol. Ecol. 21:295–306.

Schleicherová, D., G. Sella, S. Meconcelli, R. Simonini, M. P. Martino, P. Cervella, and M.

C. Lorenzi. 2014. Does the cost of a function affect its degree of plasticity? A test on
plastic sex allocation in three closely related species of hermaphrodites. J. Exp. Mar.
Bio. Ecol. 453:148–153.

Schoen, D. J. 1982. Male reproductive effort and breeding system in an hermaphroditic plant.
Oecologia 53:255–257.

Siljestam, M., and Ö. Östman. 2017. The combined effects of temporal autocorrelation and
the costs of plasticity on the evolution of plasticity. J. Evol. Biol. 30:1361–1371.

Singh, P., D. Ballmer, M. Laubscher, and L. Schärer. 2020a. Successful mating and
hybridisation in two closely related flatworm species despite significant differences in
reproductive morphology and behaviour. bioRxiv 851972, doi:
https://doi.org/10.1101/851972.

Singh, P., J. N. Brand, and L. Schärer. n.d. Evolution of the suck behaviour, a postcopulatory
female resistance trait in a hermaphroditic flatworm genus.

Singh, P., N. Vellnow, and L. Schärer. 2020b. Variation in sex allocation plasticity in three
closely related flatworm species. Ecol. Evol. 10:26–37.

St. Mary, C. M. 1997. Sequential Patterns of Sex Allocation in Simultaneous Hermaphrodites:
Do We Need Models That Specifically Incorporate This Complexity? Am. Nat. 150:73–
97.

Steiner, U. K. 2007. Investment in defense and cost of predator-induced defense along a
resource gradient. Oecologia 152:201–210.

Torchiano, M. 2017. Package "effsize": efficient effect size computation. CRAN Packag.,
doi: 10.5281/ZENODO.196082.

Turner, H. M., and R. M. Bernard. 2006. Calculating and Synthesizing Effect Sizes. Contemp.
Issues Commun. Sci. Disord. 33:42–55.

Van Buskirk, J. 2002. A Comparative Test of the Adaptive Plasticity Hypothesis:
Relationships between Habitat and Phenotype in Anuran Larvae. Am. Nat. 160:87–102.

van Velzen, E., L. Schärer, and I. Pen. 2009. The effect of cryptic female choice on sex
allocation in simultaneous hermaphrodites. Proc. R. Soc. B Biol. Sci. 276:3123–3131.

Vizoso, D. B., G. Rieger, and L. Schärer. 2010. Goings-on inside a worm: Functional
hypotheses derived from sexual conflict thinking. Biol. J. Linn. Soc. 99:370–383.

Winkler, L., and S. A. Ramm. 2018. Experimental evidence for reduced male allocation under
selfing in a simultaneously hermaphroditic animal. Biol. Lett. 14:20180570.

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Figures

Figure 1. Effect of group size on estimates of sex allocation in seven different *Macrostomum*

species (given by different colours). The line types represent the two mating strategies and the

symbols represent the ability to self-fertilize in *Macrostomum*. The plots show means and

standard errors of the raw (untransformed) data. Note that for *M. lignano*, data from seven

independent experiments are shown.

Figure 2. Standardized SA plasticity effect sizes for the effect of the presence of mating

partners (i.e. comparing isolated worms vs. worms with partners) and the strength of LSC (i.e.

comparing worms in pairs vs. octets) in seven *Macrostomum* species, along with data on

whether the species exhibit self-fertilization or/and reciprocal mating. Note that for

M. lignano, data from multiple experiments are shown. The error bars represent 95%

confidence intervals.