Overlapping roles of JIP3 and JIP4 in promoting axonal transport of lysosomes in human iPSC-derived neurons

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Running Title: JIP3/4-dependent regulation of axonal lysosomes in human neurons

Abstract:

The dependence of neurons on microtubule-based motors for the movement of lysosomes over long distances raises questions about adaptations that allow neurons to meet these demands. Recently, JIP3/MAPK8IP3, a neuronally enriched putative adaptor between lysosomes and motors, was identified as a critical regulator of axonal lysosome abundance. In this study, we establish a human induced pluripotent stem cell (iPSC)-derived neuron model for the investigation of axonal lysosome transport and maturation and show that loss of JIP3 results in the accumulation of axonal lysosomes and the Alzheimer's disease-related amyloid precursor protein (APP)-derived Aβ42 peptide. We furthermore reveal an overlapping role of the homologous JIP4 gene in lysosome axonal transport. These results establish a cellular model for investigating the relationship between lysosome axonal transport and amyloidogenic APP processing and more broadly demonstrate the utility of human iPSC-derived neurons for the investigation of neuronal cell biology and pathology.

Introduction

The endo-lysosomal system plays key house-keeping roles in all cells, but neurons are particularly dependent on efficient lysosome function owing to their extreme size, polarity and post-mitotic state (Ferguson, 2019). Indeed, rare loss-of-function mutations in multiple genes encoding lysosome proteins result in lysosome storage diseases, which frequently manifest with severe neurological and neurodegenerative pathologies (Sun, 2018; Marques and Saftig, 2019; Ballabio and Bonifacino, 2020). More recently, advances in understanding the genetics of neurodegenerative diseases associated with aging such as Alzheimer's disease, Parkinson's disease, amyotrophic lateral sclerosis and frontotemporal dementia, have identified a number of genes encoding endo-lysosomal proteins as modulators of disease risk (Lambert *et al.*, 2013; Guerreiro and Hardy, 2014; Chang *et al.*, 2017; Klein and Mazzulli, 2018; Wallings *et al.*, 2019). Human genetics thus points to the importance of optimal lysosome function for the maintenance of neuron health and raises fundamental questions about how the endo-lysosomal system is adapted to the unique demands of neurons.

Primary cultures of embryonic or neonatal rodent neurons are a well-established model for the investigation of neuronal cell biology (Barnes and Polleux, 2009; Banker, 2018). In particular, primary cultures of cortical and hippocampal neurons from transgenic and knockout mice have been widely used to investigate the functions of specific genes in neurons. Although this has been a tremendously valuable model system, it has several significant limitations. These include practical issues such as the time required for establishing mouse colonies for each new mutation and the ongoing mouse breeding and genotyping required to provide embryos/pups for each new primary culture. Furthermore, combining multiple mutations into one mouse at minimum requires lengthy breeding schemes and often has negative impacts on mouse viability. Excitingly, the

development of human induced pluripotent stem cells (iPSCs), combined with highly efficient protocols for their differentiation into defined neuronal cell types along with CRISPR-Cas9-dependent genome editing tools has opened up new opportunities for investigating the impact of targeted genetic perturbations on neuronal cell biology (Dolmetsch and Geschwind, 2011; Zhang *et al.*, 2013; Paquet *et al.*, 2016; Wang *et al.*, 2017; Fernandopulle *et al.*, 2018; Tian *et al.*, 2019). In particular, the development of robust neuronal differentiation protocols based on transcription factor over-expression now allows for the generation of human neuronal cultures with unprecedented speed and consistency (Zhang *et al.*, 2013; Fernandopulle *et al.*, 2018).

In this study, we used of human iPSC-derived neurons to investigate the relationships between lysosome axonal transport and Alzheimer's disease pathology. This research direction builds on longstanding reports of lysosome-filled axonal dilations around Aβ deposits at amyloid plaques (Terry *et al.*, 1964; Cataldo and Nixon, 1990; Nixon *et al.*, 2005; Condello *et al.*, 2011; Dikranian *et al.*, 2012; Gowrishankar *et al.*, 2015) Although this relationship between axonal lysosome accumulations and amyloid plaques has been known for many years, it has been challenging to define the relevance of these lysosomes to the broader disease pathology due to the lack of tools to selectively alter the abundance of axonal lysosomes. Recent progress on this topic came from observations that axonal lysosomes accumulate in JIP3 mutant mice with a major impact on amyloid plaques (Gowrishankar *et al.*, 2017). JIP3 is thought to act as an adaptor between lysosomes and the dynein motor; a function supported by axonal accumulation of lysosomes and/or hybrid organelles with properties of lysosomes, late endosomes and autophagosomes in response to JIP3 loss-of-function mutations in multiple model organisms (Drerup and Nechiporuk, 2013; Edwards *et al.*, 2013; Gowrishankar *et al.*, 2017; Hill and Colon-Ramos, 2019).

Our new results define baseline lysosome properties in human iPSC-derived neurons and show that KO of the human *JIP3* gene results in the accumulation of protease-deficient lysosomes in swollen axons and increased Aβ42 production. Furthermore, we found that this axonal lysosome accumulation is enhanced by the additional KO of *JIP4*, revealing that JIP4, previously shown to regulate lysosome movement in non-neuronal cells (Willett *et al.*, 2017), has an overlapping function with JIP3 in regulating axonal lysosome transport in neurons. Our results, which parallel and extend recent observations from rodent models, emphasize the power of human iPSC-derived neurons as a valuable tool for investigating human neuronal cell biology and neurodegenerative disease mechanisms. These new observations are also relevant for the development of models to study recently discovered neurodevelopmental defects arising from mutations in the human *JIP3* (also known as *MAPK8IP3*) gene (Iwasawa *et al.*, 2019; Platzer *et al.*, 2019).

Materials and Methods:

CRISPR-Cas9 mediated knock out of JIP3

We used an established WTC-11 human iPSC line (provided by Michael Ward, NINDS) that was engineered to harbor a doxycycline-inducible NGN2 transgene expressed from the AAVS1 safe harbor locus in order to support an efficient protocol for their differentiation into neurons (referred to as i^3Neurons) with properties of layer 2/3 cortical glutamatergic pyramidal (Wang *et al.*, 2017; Fernandopulle *et al.*, 2018). These iPSCs were grown on Matrigel coated dishes in E8 media (Gibco). For CRISPR-Cas9-mediated gene editing, cells were harvested using accutase and 1.5 million cells were resuspended in Mirus nucleofector solution and electroporated with 5 ug of px458 plasmid (Addgene plasmid #48138) containing a small guide RNA (see Table S2 for

oligonucleotide information) targeted against the JIP3 (also known as MAPK8IP3) gene using an Amaxa 2D nucleofector. Electroporated cells were then plated into one well of a 24 well plate. GFP-positive cells were then selected by FACS after 4 days. Cells were once again plated communally post-sorting and serially diluted 4 days later to yield clonal populations for screening. 10 colonies were selected per sgRNA and screened for mutations using PCR amplification of genomic DNA flanking the sgRNA target site followed by sequencing of the amplicons (see Supplemental Table S2 for primer details).

Generation of JIP3/4 Double knock out cells

The JIP3 KO IPSC line was subjected to a second round of CRISPR-based editing by the method described above to disrupt the JIP4 gene in these cells (see Table S2 for oligonucleotide information). PCR amplification of genomic DNA flanking the sgRNA target site followed by sequencing of the amplicons (see Supplemental Table S2 for primer details), was carried out to confirm the double knock out.

Generation of stable IPSC line expressing LAMP1-GFP

LAMP1-GFP was amplified using PCR (using primers listed in Table S2) from Addgene plasmid #34831 and then cloned into lentiviral vector FUGW (Addgene plasmid #14883) from which the GFP fragment had been excised, by digesting the vector with BamHI and EcoRI. FUGW was a gift from David Baltimore (Addgene plasmid # 14883; http://n2t.net/addgene: 14883; RRID:Addgene_14883). LAMP1-mGFP was a gift from Esteban Dell'Angelica (Addgene plasmid # 34831; http://n2t.net/addgene: 34831; RRID:Addgene_34831).

HEK 293 FT cells were transfected with the newly made LAMP1-GFP lentiviral vector, containing along with the psPAX2 (Addgene #12260) and PCMV (Addgene #8454)

packaging plasmids. pCMV-VSV-G was a gift from Bob Weinberg (Addgene plasmid # 8454; http://n2t.net/addgene:8454; RRID:Addgene_8454). psPAX2 was a gift from Didier Trono (Addgene plasmid # 12260; http://n2t.net/addgene:12260; RRID:Addgene_12260) The supernatant from these cells (containing the virus) was collected, concentrated and then diluted into E8 medium containing the Y-27632 ROCK inhibitor (Tocris). This was added to freshly split IPSCs (1.5 million cells in one well of a 6-well plate). The media on the cells was changed after 48 hours. These cells were then expanded and LAMP1-GFP expressing cells were selected by FACS.

ELISA-based measurements of Aβ42 in i^3Neurons

I^3 Neurons were differentiated from WT and JIP3 KO IPSCs as described previously (Fernandopulle *et al.*, 2018). i^3 Neurons were grown on Poly-Ornithine(PO)-coated 6 well plates for 21 days before Aβ42 ELISA measurements were performed as per a previously validated ELISA assay for the measurement of human Aβ42 (Teich *et al.*, 2013). In brief, the cells were lysed into Tris-EDTA and then processed for ELISA based measurements of Aβ42 using a high-sensitivity ELISA kit (catalog no. 292–64501; Wako). Standard curves were generated using Aβ1-42 peptide standards for each set of experiments. Such measurements were made from four independent WT and KO cultures.

Immunofluorescence analysis of i^3 Neurons

I^3Neurons differentiated for one to six weeks on 35 mm Mattek glass bottom dishes were processed for immunostaining as described previously (Gowrishankar *et al.*, 2017). See Table S1 for antibody information.

Immunoblotting experiments

I^3Neurons (control and hJIP3 KO) were grown on PO-coated six-well plates (500,000 cells/well). After 14 or 21 days of differentiation, i^3Neurons were washed with ice-cold PBS and then lysed in lysis buffer [Tris-buffered saline (TBS) with 1% Triton, protease inhibitor cocktail and phosphatase inhibitor] and spun at 13,000 *g* for 5 min. The supernatant was collected and incubated at 95°C for 5 min in SDS sample buffer before SDS-PAGE, transfer to nitrocellulose membranes, and immunoblotting. For immunoblotting experiments comparing iPSCs with i^3Neurons, lysates of the parent IPSC line that is 70% confluent were compared to the lysates from i^3Neurons differentiated from same parental split). See Table S1 for antibody information.

Microscopy

Images were acquired using a Zeiss 880 Airyscan confocal microscope via a 100X plan-Apochromatic objective (1.46 NA) with 2X optical zoom. Live imaging of lysosome dynamics in i^3Neurons was carried out using Airy Scan imaging mode using 100X objective with 1.5 or 2x optical zoom to achieve scan speeds of 1-2 frames per second. Airyscan processing of images was done using Zen software. Further analysis of the videos was performed using ImageJ software.

Analysis of lysosome properties

To measure the mobility of LAMP1-positve and Lysotracker-positive organelles, i^3Neurons stably expressing LAMP1-GFP were incubated with 50nM Lysotracker Deep Red for 3 minutes and then and then rinsed gently twice with imaging medium warmed to 37 degrees (136 mM NaCl, 2.5 mM KCl, 2 mM $CaCl_2$, 1.3 mM $MgCl_2$, and 10 mM Hepes, pH 7.4, supplemented with BSA and glucose) and imaged at 37 degrees in the same buffer for 10 min in Airyscan mode as described above. Kymographs were generated from the videos using the “reslice” function on “straightened” axons

(Gowrishankar *et al.*, 2017) (ImageJ), and the percentage of Lysotracker-positive LAMP1 organelles, per axon, was computed. The dynamics and directionality of LAMP1 organelles with respect to Lysotracker-positive vesicles was measured using the kymographs. Organelles were considered stationary if they moved <2 μm in 2 min. A total of eleven neurons from three independent experiments were analyzed.

Statistical analysis

Data are represented as mean ± SEM unless otherwise specified. Statistical analysis was performed using Prism 7 software. Detailed statistical information (statistical test performed, number of independent experiments, and p-values) is described in the respective Figure legends.

Results

To evaluate the suitability of the i^3 Neuron model (Wang *et al.*, 2017; Fernandopulle *et al.*, 2018); (Figure 1A) for the investigation of neuronal lysosome biology, we began with a characterization of key properties of these organelles based on recent findings from studies of lysosome distribution and maturation in mouse neurons and brain tissue (Gowrishankar *et al.*, 2015; Farias *et al.*, 2017; Cheng *et al.*, 2018; Yap *et al.*, 2018). As we had previously observed in mouse neurons both in vitro and in vivo (Gowrishankar *et al.*, 2015), LAMP1-positive organelles in the i^3neurons were more abundant in the soma, as compared to dendrites and axons (Figure.1B, C) and the LAMP1-positive organelles localized in the soma of i^3neurons were more enriched in multiple cathepsins (luminal proteases) compared to those localized in axons and dendrites (Figure 1D-E). This relationship between the abundance of luminal hydrolases that correlates with lysosome subcellular positioning in neurons likely reflects a spectrum of maturation between late endosomes, autophagosomes and lysosomes (Gowrishankar *et al.*, 2015; Farias *et al.*,

2017; Cheng *et al.*, 2018; Yap *et al.*, 2018) and matches previous in vivo observations (Gowrishankar et al, 2015). For simplicity, and reflecting the eventual degradative function mediated by this pathway (Ferguson, 2018), we will subsequently refer to this collection of closely related organelles as lysosomes.

In order to visualize lysosomes in live cell imaging studies, we generated iPSCs stably expressing LAMP1-GFP, an abundant integral membrane protein of lysosomes (Eskelinen, 2006). This transgene yielded expression levels that were comparable to the levels of endogenous LAMP1 (Figure 2A). LAMP1-GFP-positive organelles were less abundant in axons when compared to the soma or dendrites (Figure 2B), consistent with what is observed with endogenous LAMP1 staining in the i^3Neurons (Figure 1). They also exhibited bi-directional movement (Video 1). Since LAMP1 localizes to a spectrum of endo-lysosomal organelles (non-degradative organelles, degradative lysosomes as well as biosynthetic intermediates(Swetha *et al.*, 2011; Pols *et al.*, 2013; Cheng *et al.*, 2018), we examined the dynamics of LAMP1-GFP-positive organelles in axons as well as their labeling with Lysotracker, a dye that is commonly used to identify lysosomes based on its pH-dependent accumulation within their acidic lumen. These experiments revealed distinct populations of LAMP1-positive organelles in axons of WT iNeurons based on differences in their motility and acidification. Firstly, only ~20% of the LAMP1 organelles in the axon were Lysotracker-positive (Figure 2C, D). Interestingly, these LAMP1 and lysotracker double positive organelles moved predominantly in the retrograde direction (Figure. 2C; Videos 2-4). In contrast, when the directionality of all LAMP1-positive organelles was examined, we observed that a slight majority of LAMP1 vesicles moved in the anterograde direction (Figure. 3E; Video 2-4). These anterogradely-moving organelles that do not label with lysotracker (Videos 2 and 3) may represent Golgi-derived biosynthetic, LAMP1-containing vesicles that deliver newly

made lysosomal proteins for the maturation of endosomes and autophagosomes into lysosomes within the axon (Swetha *et al.*, 2011; Pols *et al.*, 2013).

Having established that the i^3Neurons exhibit similar baseline axonal lysosome properties to those previously observed in rodent neuron primary culture models, we next tested whether they also responded similarly to a JIP3 loss-of-function mutation, a genetic perturbation which promotes axonal lysosome accumulation in other more established neuronal model systems (Drerup and Nechiporuk, 2013; Edwards *et al.*, 2013; Gowrishankar *et al.*, 2017). Consistent with past reports of its neuronal enrichment (Gowrishankar *et al.*, 2017), the JIP3 protein was undetectable in iPSCs but highly abundant in the i^3Neurons, a similar pattern was observed for VAMP2, a synaptic vesicle protein (Figure 3A). Meanwhile, LAMP1 was present at comparable levels in both iPSCs and i^3Neurons (although it exhibited a difference in mobility between iPSCs and i^3Neurons (Figure.3B). This mobility shift could reflect differences in extent of glycosylation of LAMP1 in neuronal cells as compared to non-neuronal cells and might have an impact on the ability of LAMP1 to serve as a transporter for the efflux of cholesterol from neuronal lysosomes (Li and Pfeffer, 2016).

Having established that JIP3 is highly expressed in the i^3Neuron model, we next used CRISPR-Cas9 mediated gene editing to generate a JIP3 KO iPSC line and obtained cells with a homozygous 1 bp insertion in exon 1 (Figure 3C). This results in a frameshift and premature stop codon that will prevent JIP3 protein translation and is furthermore predicted to trigger nonsense mediated decay of the mutant transcript. Immunoblotting confirmed the loss of JIP3 protein in i^3Nneurons derived from these JIP3 mutant iPSCs (Figure 3D). Consistent with studies of mouse JIP3 KO cortical neuron primary cultures, the human JIP3 KO i^3Neurons developed lysosome-filled axonal swellings as revealed

by lysotracker labeling (Figure 3E, F) and LAMP1 immunofluorescence (Figure 3 G, H). These swellings were observed by 10 days of differentiation (Figure 3F) and became more penetrant with age (Figure 3H). In contrast to lysosomes, mitochondria did not prominently accumulate in these swellings (Figure 3I).

Having established that loss of JIP3 results in axonal lysosome-filled swellings, a phenotype seen in mouse JIP3 KO neurons as well as at Alzheimer's disease amyloid plaques, we next interrogated the role of human JIP3 on amyloid precursor protein (APP) processing into Aβ peptides in this cellular model. Our previous studies had demonstrated that accumulation of axonal lysosomes in JIP3 KO mouse neurons was accompanied by increased levels of BACE1 (β-site APP cleaving enzyme), the protease that initiates amyloidogenic processing of APP. We found a similar build-up of BACE1 protein in the human JIP3 KO i^3Neurons (Figure 4 A, B). This was accompanied by a substantial increase in levels of intracellular Aβ42 peptide in the JIP3 KO cultures (Figure 4C).

Although the JIP3 KO phenotypes are striking, the impact on axonal lysosome accumulation is age dependent and lysosomes still move in both anterograde and retrograde directions in JIP3 KO axons. This indicates the existence of redundancy in mechanisms for lysosome axonal transport. Human JIP4 was previously proposed to play a role in the dynein-dependent movement of lysosomes from the periphery to perinuclear region, in non-neuronal cells (Willett *et al.*, 2017). Studies in cultured neurons from JIP3+JIP4 double KO mice indicated a functional redundancy in these molecules for kinesin-dependent axonal transport (Sato *et al.*, 2015). However, the impact on lysosomes was not investigated in these double KO neurons. Due to sequence conservation between JIP3 and JIP4 and the ability of JIP4 to promote

lysosome movement in other cell types (Willett *et al.*, 2017), we next examined the effect of knocking out both JIP3 and JIP4 in our human iPSC model system. Cas9-based editing of JIP4 in the JIP3 mutant IPSC line resulted in a 4 bp deletion in exon 9 of JIP4 (Figure 5A). iPSCs lacking both JIP3 and JIP4 were viable and could still be efficiently differentiated into i^3Neurons. Immunoblotting confirmed the loss of both proteins in i^3Neurons differentiated from this JIP3+4 double knock out (DKO) iPSC line (Figure 5B). When compared to even the JIP3 KO line, the JIP3+JIP4 double KO cells exhibited a dramatic axonal lysosome accumulation phenotype (Figure.5 C-E).

Discussion

In this study, we have used human i^3Neurons as a model for cell biological and genetic studies of axonal lysosome traffic with potential human disease relevance. Our results from JIP3 KO i^3Neurons corroborate past observations from JIP3 KO mouse neuron primary cultures, revealing potential Alzheimer's disease relevant consequences of lysosome axonal transport defects. This strengthens confidence that this human iPSC-derived model yields neurons that are sufficiently differentiated to serve as a valuable alternative to traditional rodent neuron primary culture models for the investigation of neuronal cell biology, including the transport of endo-lysosomal and autophagic organelles, as also suggested by other recent studies (Liao *et al.*, 2019; Boecker *et al.*, 2020). We further generated JIP4 KO and JIP3 +JIP4 double KO lines and established that JIP3 and JIP4 proteins have over-lapping functions in promoting the axonal transport of lysosomes.

Our experiments additionally yielded several interesting observations relating to heterogeneity of the axonal LAMP1-positive organelles. First, only a small fraction of them (~20%) were labeled with the lysotracker dye, indicating an acidic lumen. Second,

these lysotracker-positive LAMP1 vesicles were less mobile than the overall pool of LAMP1-positive organelles and their movement was most frequently in the retrograde direction (Videos 2-4; Figure 2C-E). Third, non-acidified LAMP1 organelles were biased toward movement in the anterograde direction and were generally smaller in appearance. Collectively, these observations are consistent with a model wherein the majority of the anterograde LAMP1-positive organelles represent precursors, which deliver LAMP1 and other cargos into the axon. Distinct identities for anterograde versus retrograde axonal LAMP1-positive organelles is consistent with electron microscopy analysis of axonal organelle morphology proximal and distal to a localized blockade of transport where organelles with late endosome/lysosome morphology preferentially accumulated distal to the block (Tsukita and Ishikawa, 1980). More recently, LAMP1-positive anterogradely moving organelles that do not have degradative properties but which also deliver synaptic vesicle proteins, were reported (Vukoja *et al.*, 2018). Our results define LAMP1 vesicle sub-populations that can be discriminated based on their apparent size, movement direction and lysotracker labeling affinity. These observations are consistent with previous studies, which documented a gradient of lysosome acidification along the axon with a higher frequency of acidified lysosomes closer to the cell body(Overly *et al.*, 1995; Overly and Hollenbeck, 1996). However, differences in probe pH sensitivity could explain why other investigators using pHluorin (pKa~7) fused to LAMP1 concluded that essentially all LAMP1 organelles are acidified (Farias PNAS 2017). The growing appreciation for heterogeneity in axonal endo-lysosome sub-populations raises a host of questions about regulatory mechanisms and physiological functions (Farias *et al.*, 2017; Cheng *et al.*, 2018; Ferguson, 2018). The experimental tractability of the i^3Neuron systems for imaging and genetic manipulations makes it an ideal tool for answering these questions.

The increased abundance of BACE1 and Aβ42 peptide that accompanies lysosome transport defects in the JIP3 KO i^3Neurons parallels past observations in mouse JIP3 KO neuronal primary cultures, transgenic Alzheimer's disease mouse models as well as human Alzheimer's disease brains (Fukumoto *et al.*, 2002; Holsinger *et al.*, 2002; Zhao *et al.*, 2007; Zhang *et al.*, 2009; Gowrishankar *et al.*, 2017). These observations establish the utility of the i^3Neurons model for investigating the mechanistic basis of the relationships between lysosome axonal transport and APP cleavage as well as for the development of new strategies to modulate these processes for therapeutic purposes.

In addition to serving as a model for investigating Alzheimer's disease, the JIP3 KO i^3Neuron model may support the investigation of other human neurological diseases. Of most obvious relevance, de novo heterozygous loss-of-function variants in the human *JIP3/MAPK8IP3* gene were recently identified as a cause of neurodevelopmental defects resulting in intellectual disability (Iwasawa *et al.*, 2019; Platzer *et al.*, 2019). The emergence of a severe human neurodevelopmental disease from what is predicted to be just a partial reduction JIP3 expression suggests an exquisite sensitivity to gene dosage at critical developmental stages. Consistent with this interpretation, large scale exome sequencing revealed strong selective pressure against predicted deleterious variants in the human *MAPK8IP3* gene (Lek *et al.*, 2016). Thus, in addition to our original focus on the role of JIP3 in lysosome transport and APP processing mechanisms, there are now broader questions about how JIP3 contributes to human neuronal development that can potentially be answered in the i^3Neuron model system.

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Figure Legends

Figure 1: Cathepsin-rich lysosomes are concentrated in the soma of i^3Neurons.

Phase contrast images of an IPSC colony (left) and i^3Neurons differentiated for two weeks (right). B. Stitched confocal image of i^3Neurons (3 weeks of differentiation) co-stained for LAMP1 (cyan; endo-lysosomal compartments) and MAP2 (magenta; dendrites). Scale Bar, 10μm. C. Confocal immunofluorescence image of an i^3Neuron stained for LAMP1 (cyan) and MAP2 (magenta), showing concentration of LAMP1-positive organelles in the soma (asterisk) when compared to the neurites (yellow arrows). Scale Bar, 10μm. D. Immunofluorescence staining of i^3Neurons for cathepsin L (grey), LAMP1 (cyan) and Tau (magenta), showing that LAMP1-postive organelles in neurites are relatively deficient of cathepsin L compared to those in the neuronal cell body. Inset scale bar,1 μm. E. Immunofluorescence staining of i^3Neurons for cathepsin D (grey), LAMP1 (cyan) and Tau (magenta), showing that LAMP1-postive organelles in neurites are relatively deficient in cathepsin D compared to those in neuronal cell bodies. Scale Bar, 10μm. Inset scale bar, 1μm.

Figure 2: Acidification and movement of LAMP1-positive organelles in i^3Neurons.

A. Immunoblot showing levels of LAMP1-GFP and endogenous LAMP1 in lysates from an i^3Neuron culture that was differentiated from an iPSC line that stably expresses a LAMP1-GFP transgene. B. LAMP1-GFP subcellular distribution aligns well with endogenous LAMP1 in i^3Neurons (10 days of differentiation). Scale Bar, 10μm. C. Fluorescence image of a portion of an axon, 'straightened' using ImageJ, from an

i^3Neuron expressing LAMP1-GFP (cyan) and stained with Lysotracker-Deep Red (magenta). Scale Bar, 5μm. D. Kymograph of lysotracker (magenta) and LAMP1-GFP (cyan) dynamics in an axon, from a video taken over 2.5 minutes at 1 frame per second. Scale Bar, 5μm. White arrow: Lysotracker-positive+LAMP1 double positive vesicle. Yellow arrow: small, anterogradely moving LAMP1 vesicle. E. Quantification of the fraction of axonal lysotracker and LAMP1-positive organelles in individual neurons. Data reflects 157 LAMP1 organelles analyzed from a total 12 i^3 Neurons analyzed from three independent experiments F. Plot summarizing the direction of movement of LAMP1-positive organelles. Each point on the graph represents the fraction of LAMP1 organelles moving in that direction from a single neuron. Data reflects LAMP1 organelles from 11 i^3Neurons from three independent experiments. ($p<0.01$, unpaired t test).

Figure 3: Lysosomes accumulate within axonal swellings in JIP3 KO i^3Neurons.

A. Immunoblots of samples from the parental iPSC line and differentiated i^3Neurons (21 days of differentiation). B. Immunoblot for LAMP1 in the parental iPSC line and differentiated i^3Neurons. C. Alignment of a portion of the exon 1 sequence for human JIP3 that shows the location of a single base-pair insertion in the KO cells. The sgRNA target sequence is highlighted in blue. D. JIP3 immunoblot reveals the complete loss of the protein in CRISPR-edited JIP3 KO i^3Neurons (21 days of differentiation), while it is abundant in the i^3Neurons differentiated from control IPSCs. E and F. Fluorescence images of Lysotracker Red DND-99 stained Control and JIP3 KO i^3Neurons grown at low density (12 days of differentiation). Yellow arrows highlight axonal swellings filled with lysosomes in the JIP3 KO neurons. G and H. Stitched immunofluorescence image of control (G) and JIP3 KO (H) i^3Neurons (21 days of differentiation) showing large LAMP1-positive accumulations in the KO culture while relatively fewer vesicles seen in neurites

of control cultures. Scale bar, 50 µm. I. Confocal images of JIP3 KO i^3Neurons (12 days of differentiation) stained with Lysotracker Deep Red (cyan) and Mitotracker Green (magenta). Although some mitochondria are present in the LAMP1-positive axonal swellings, they are not enriched as compared to the LAMP1 signal. Scale bar, 10 µm.

Figure 4: JIP3 KO i^3Neurons exhibit increased BACE1 and Aβ42 levels.

A. Immunoblot reveals increased levels of BACE1 protein in JIP3 KO i^3Neurons compared to Control (VAPB-loading control) B. Quantification of BACE1 levels in JIP3 KO i^3Neurons compared to Control (n=3; mean ± SEM; $p<0.05$; unpaired t-test). C. Quantification of endogenous human Aβ42 levels based on ELISA measurements from n=4 independent Control and JIP3 KO i^3Neuron cultures differentiated for 21 days. (mean ± SEM; $p<0.01$, unpaired t test).

Figure 5: JIP3/4 DKO i^3Neurons exhibit exacerbation of lysosomal phenotype compared to JIP3 single KO neurons.

A. Alignment of a portion of the exon 9 sequence for human JIP4 that shows the location of a four base-pair deletion in the DKO cells. The sgRNA target sequence is highlighted in blue. (B) Immunoblot showing levels of JIP3 and JIP4 in control, JIP3 KO and JIP3+JIP4 DKO i^3Neurons differentiated for 13 days; ribosomal protein S6 (rS6) was used as a loading control. (C-E) Confocal image of control i^3Neurons, JIP3 KO i^3Neurons and JIP3+JIP4 i^3Neurons (DIV 13), stably expressing LAMP1-GFP. Scale bar, 10 µm. Insets highlight lysosome accumulations in the KO neurons.

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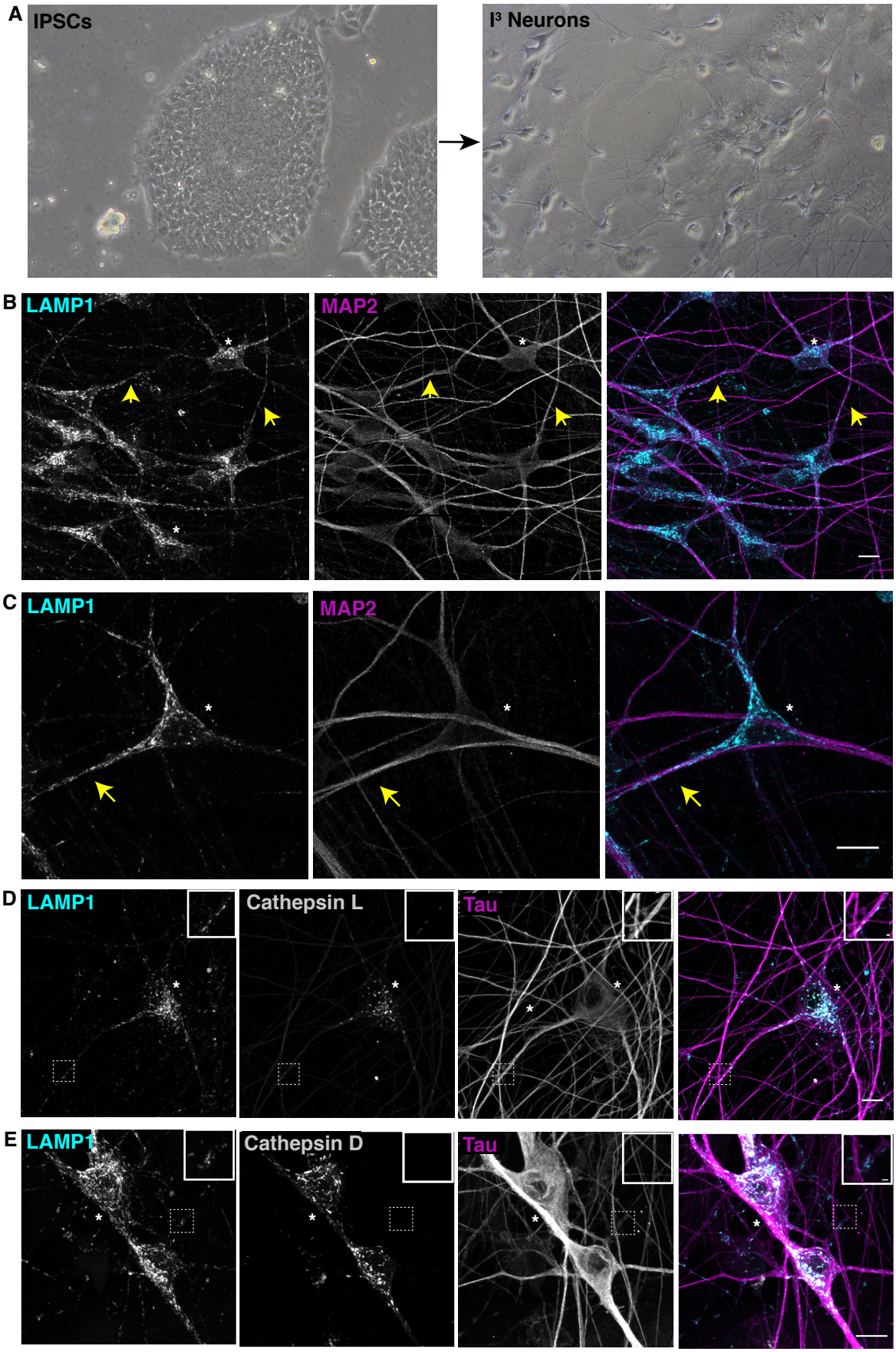
A
IPSCs
I³ Neurons
B
LAMP1
MAP2
C
LAMP1
MAP2
D
LAMP1
Cathepsin L
Tau
E
LAMP1
Cathepsin D
Tau

Figure 1

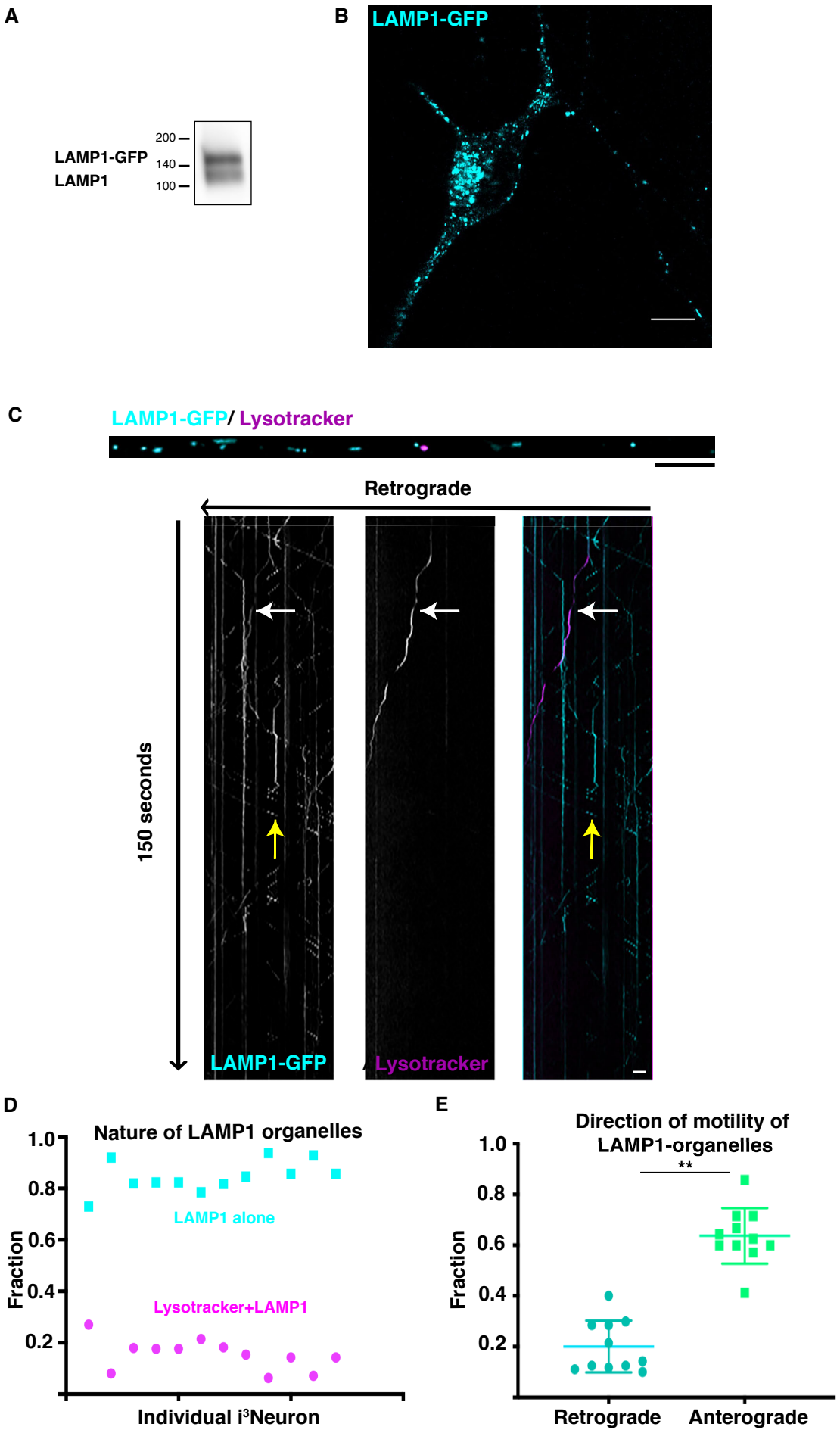
A
LAMP1-GFP
LAMP1
200
140
100
B
LAMP1-GFP
C
LAMP1-GFP/ Lysotracker
Retrograde
150 seconds
LAMP1-GFP
Lysotracker
D
Nature of LAMP1 organelles
1.0
0.8
0.6
0.4
0.2
Fraction
LAMP1 alone
Lysotracker+LAMP1
Individual i³Neuron
E
Direction of motility of
LAMP1-organelles
**
1.0
0.8
0.6
0.4
0.2
Fraction
Retrograde
Anterograde

Figure 2

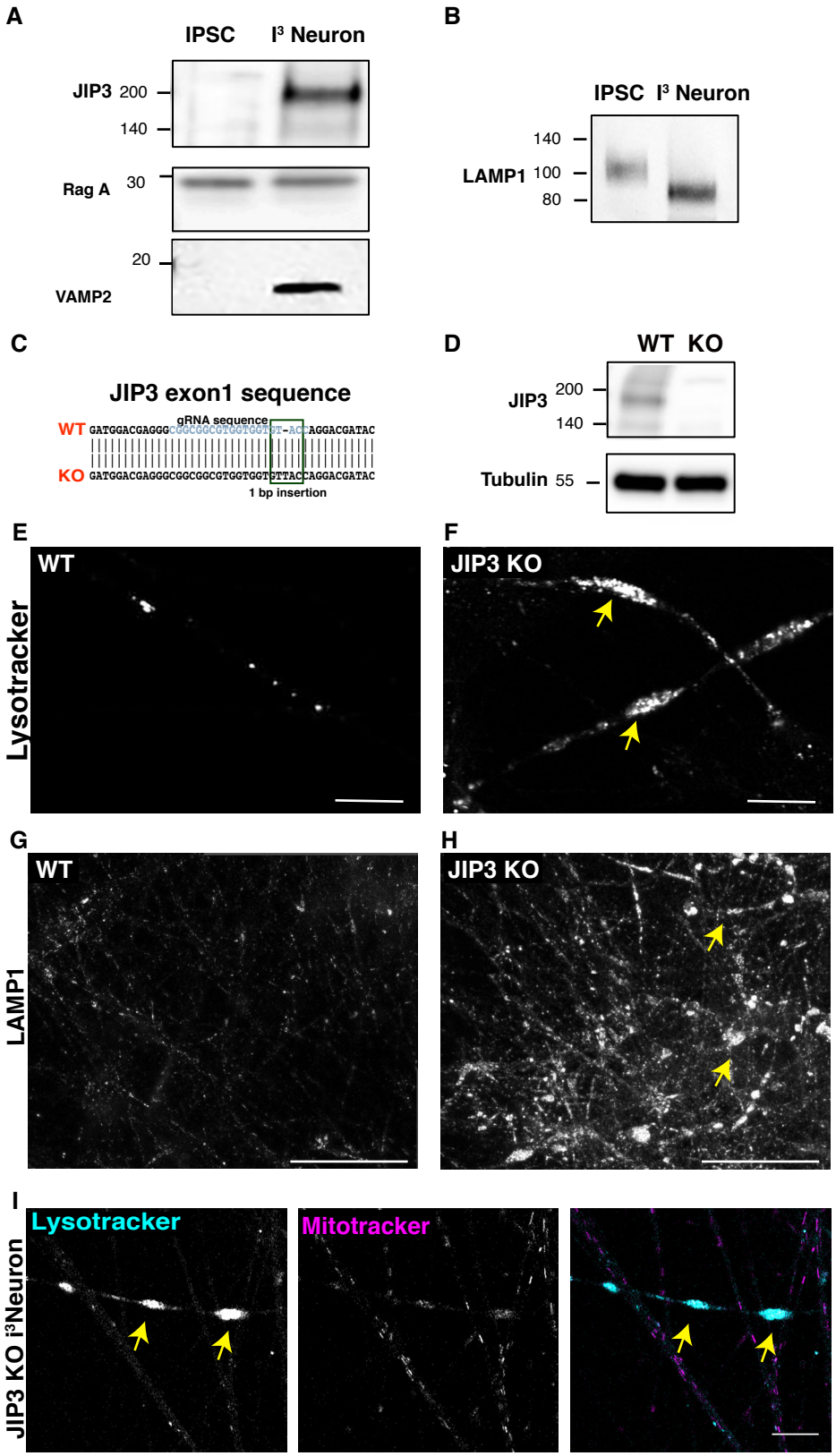
A
IPSC
I³ Neuron
JIP3 200
140
Rag A 30
20
VAMP2
B
IPSC I³ Neuron
140
LAMP1 100
80
C
JIP3 exon1 sequence
gRNA sequence
WT GATGGACGAGGCGGCGGCGTGGTGGTGTACAGGACGATAC
KO GATGGACGAGGGCGGCGGCGTGGTGGTTTACCAGGACGATAC
1 bp insertion
D
WT KO
JIP3 200
140
Tubulin 55
E
WT
Lysotracker
F
JIP3 KO
G
WT
LAMP1
H
JIP3 KO
I
Lysotracker
Mitotracker
JIP3 KO i³Neuron

Figure 3

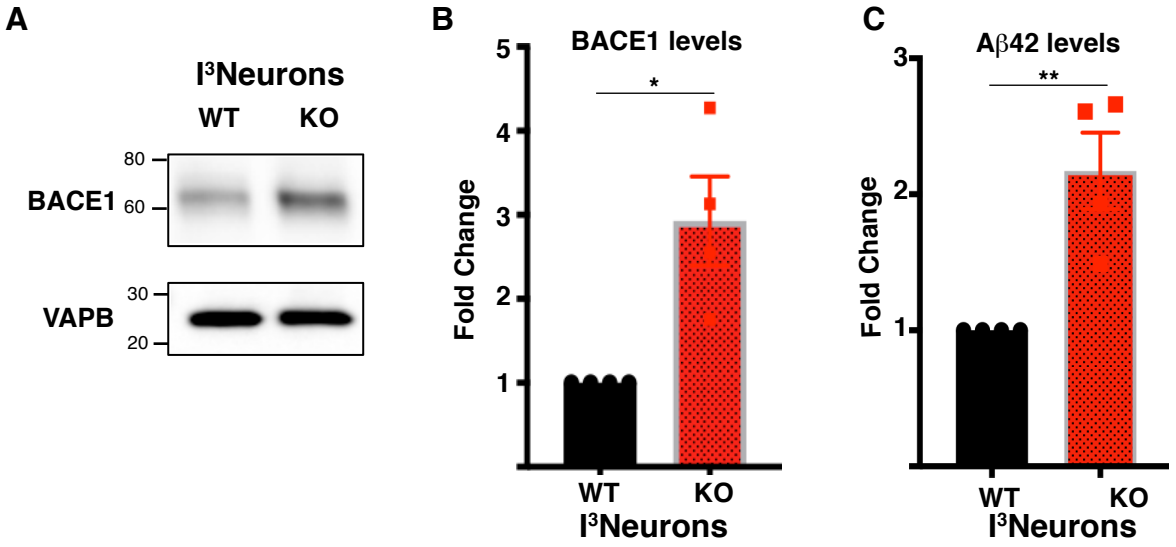
A
I³Neurons
WT
KO
80
BACE1
60
30
VAPB
20
B
BACE1 levels
*
5
4
3
2
1
Fold Change
WT
KO
I³Neurons
C
Aβ42 levels
**
3
2
1
Fold Change
WT
KO
I³Neurons

Figure 4

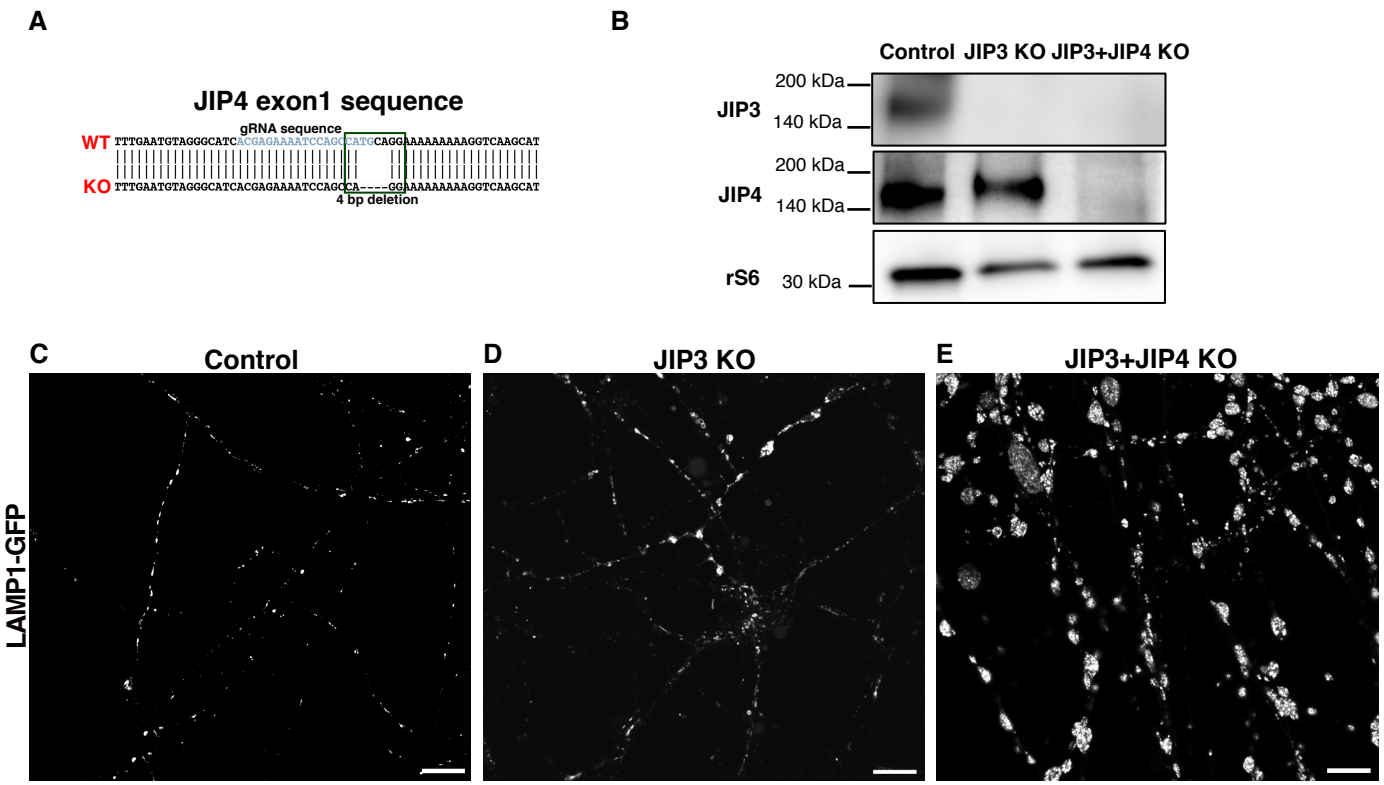
A
JIP4 exon1 sequence
gRNA sequence
WT TTTGAATGTAGGGCATCACGAGAAAATCCAGCCATGCAGGAAAAAAAAAGGTCAAGCAT
KO TTTGAATGTAGGGCATCACGAGAAAATCCAGCCA----GGAAAAAAAAAGGTCAAGCAT
4 bp deletion
B
Control JIP3 KO JIP3+JIP4 KO
JIP3
200 kDa
140 kDa
JIP4
200 kDa
140 kDa
rS6
30 kDa
C
Control
D
JIP3 KO
E
JIP3+JIP4 KO
LAMP1-GFP

Figure 5

Supplemental Material for:

Overlapping roles of JIP3 and JIP4 in promoting axonal transport of lysosomes in human iPSC-derived neurons

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Supplemental Video Legends

LAMP1 and Lysotracker dynamics in i^3Neurons

Video 1: Video of LAMP1-positive vesicles in i^3Neuron that stably express LAMP1-GFP reveals that LAMP1-positive organelles in neurites are highly dynamic and move bi-directionally. Images were acquired at 2 frames/s, playback rate 21 frames/s. Scale bar, 10 µm.

Videos 2-4: Videos show dynamics of Lysotracker-positive (acidic; Video 3,4) and Lysotracker-negative LAMP1 vesicles (Video 2,4) in axon of an i^3Neuron stably expressing LAMP1-GFP and differentiated for a week. Images were acquired at 1 frame/s, playback rate 7 frames/s. Scale bar, 5 µm.

Table S1: Antibody Summary

Antibody	Source	Catalog Number
BACE1	Cell Signaling Technology	5606
Cathepsin D	R&D Systems	AF1014
Cathepsin L	R&D Systems	AF 952
JIP3	Sigma	HPA069311
JIP4	Cell Signaling Technology	5519
LAMP1	University of Iowa DSHB	H4A3
LAMP1	Cell Signaling Technology	9091
MAP2B	Millipore	AB5622
MAP2	BD Biosciences	610410
Neurofilament	Biolegend	SM312-R
Rag A	Cell Signaling Technology	4357
S6 Ribosomal protein	Cell Signaling Technology	2217

Tau	Cell Signaling Technology	4019S
Tubulin	Sigma	T5168
VAMP2	Synaptic systems	104211
VAPB	Sigma Aldrich	HPA013144

Table S2: Summary of oligonucleotide primers used in the study

Gene Target	Use	Sense sequence (5' 3')	Antisense sequence
JIP3	CRISPR KO	CACCGCGGCGGCGTGGTGGTGTACC	AAACGGTACACCACCACGCCGCCGC
JIP4	CRISPR KO	CCGACGAGAAAATCCAGCCATGC	AAACGTGAGAAAGATGTGCTGCAA
JIP3	Genomic sequencing	ACGCTCGTACTGGGTGA	GCGATGATGGAGATCCAGATG
LAMP1	PCR for cloning of LAMP1-GFP into FUGW lentiviral vector	GGCTGCAGGTCGACTCTAGAGGATCCATGGCGGCCCCC GGCAGCGC	TTATCGATAAGCTTGATATCGAATTCTT ACTTGTACAGCTCGTCCA