

Do carbonated beverages reduce bleeding from gill injuries in angled Northern Pike?

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Alexandria Trahan^{1,*}, Auston Chhor¹, Michael J. Lawrence², Jacob W. Brownscombe¹,
Daniel Glassman¹, Connor H. Reid¹, Alice E.I. Abrams¹, Andy J. Danylchuk⁴ & Steven J.
Cooke¹

¹ Fish Ecology and Conservation Physiology Laboratory, Department of Biology and
Institute of Environmental and Interdisciplinary Science, Carleton University, Ottawa,
ON, K1S 5B6, Canada

² Department of Biological Sciences, University of Manitoba, 50 Sifton Road, Winnipeg,
MB, R3T 2N2, Canada

³ Department of Environmental Conservation, University of Massachusetts Amherst,
160 Holdsworth Way, Room 304, Amherst, MA, 01003-9485, USA

*Author for Correspondence: atrahan9@gmail.com

Abstract

The premise of catch-and-release angling is that most fish survive fisheries interactions. Therefore, it is common for anglers, management agencies, and other organizations to share information on handling practices and other strategies that are believed to improve fish welfare and survival. Recent media coverage has sensationalized the use of carbonated beverages to treat bleeding fish, an intervention that is purported to stop bleeding but has yet to be validated scientifically. We captured Northern Pike (*Esox lucius*) via hook and line, experimentally injured their gills in a standardized manner, and treated them with either Mountain Dew, Coca Cola, or carbonated lake water and observed the duration and intensity of bleeding, as well as overall blood loss (using gill colour as a proxy) while the fish was held in a lake water bath. As a control, we had a group of experimentally injured fish that did not have liquid poured over their gills before the observation period. All treatments and the control were conducted at two different water temperatures (11-18 °C and 24-27 °C) to determine if the effects of pouring carbonated beverages over injured gills is temperature dependent. When compared to the control, we found that the duration and intensity of bleeding increased regardless of the type of carbonated beverages used in this study, and there was no effect of water temperature. Use of chilled versus ambient temperature beverages similarly had no influence on outcomes. As such, there is no scientific evidence to support the use of carbonated beverages for reducing or stopping blood loss for fish that have had their gills injured during recreational angling based on the context studied here. This study reinforced the need to scientifically test angler anecdotes and theories when it comes to best practices for catch-and-release fishing.

44 **Key words:** gill injury, blood loss, fishing, mitigation, catch-and-release

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Introduction

Recreational angling is a common practice around the globe. Although some fish are harvested, a greater percentage of them are released (Cooke and Cowx 2004). Catch-and-release (C&R) occurs when recreational anglers comply with local harvest regulations or when it is adopted voluntarily based on their conservation ethic (Arlinghaus et al. 2007). Regardless of the reason, the general premise with C&R is that most released will fish survive angling-released stress and physical injuries (Wydoski 1977). Hooking injury is the most important factor influencing whether a fish survives a C&R event, with hook injury to critical areas, such as the gills or deeply in the esophagus, yielding comparatively higher mortality than when fish are hooked in areas such as the corner of the jaw (Muoneke and Childress 1994; Bartholomew and Bohnsack 2005; Arlinghaus et al., 2007; Cooke and Schramm 2007). Based on this, there has been considerable effort to develop techniques to reduce hooking injuries in critical areas, including bleeding that can occur where the hook penetrates the fish (reviewed in Brownscombe et al. 2017).

Using carbonated beverages, such as Mountain Dew and Coca Cola, poured over gills of a fish to reduce or stop bleeding caused by hooking injury is gaining in popularity within the recreational angling community as a best practice for C&R. There is even a Facebook page, “Save a Million Fish”, where recreational anglers share videos and stories of how they have saved Muskellunge (*Esox masquinongy*) with deep hooking injuries (Anderson 2018), including pouring carbonated beverages over the gills. Popular media articles in support of this practice suggest that carbon dioxide in the beverages causes vasoconstriction of the blood vessels to slow or stop bleeding (Pyzer

2015, 2019), or that phosphoric acid in carbonated beverages causes coagulation (Green 2015; Bardin 2019). However, there is no empirical research on the effectiveness of carbonated beverages for impeding bleeding in injured fish.

Gills are a multifunctional organ for fish, playing critical roles in gas exchange, ion and water balance, ammonia excretion, and acid-base balance (Evans et al. 2005). Carbonated beverages have low pH coupled with high levels of carbon dioxide (CO₂) in aqueous solution, various sugars, caffeine (if not caffeine free), phosphoric acid (H₃PO₄; Coca Cola), and citric acid (C₆H₈O₇; Mountain Dew). Several of these compounds could have an effect on gill injuries. There is a rich literature describing the effects of low pH water on ion regulation, ammonia excretion, and metabolic acid (H⁺) excretion (Wood and McDonough 1988; Evans et al. 2005; Kwong et al. 2014). The elevation of external CO₂ levels when carbonated beverages are poured over the gills may drive CO₂ into the fish's body by reversing the normal gradient for CO₂ excretion from blood to water (Gilmour, 2010, 2001; Gilmour and Perry 2009). In addition, chemoreceptors that detect changes in CO₂ are located in the gill, and may activate cardiorespiratory reflexes such as increased breathing, bradycardia, and peripheral vasoconstriction (Reid et al. 2000; Brauner et al. 2019; Tresguerres et al. 2019). Bradycardia, or slowing of heart rate, may transiently reduce bleeding (Perry and Desforges 2006). Also, caffeine is a non-specific adenosine receptor antagonist that may alter chemoreceptor signalling (Coe et al. 2017).

Regardless of the mechanism, the technique of pouring carbonated beverages over bleeding hook injuries of fish continues to be promoted, but debates as to the efficacy of its uses have also ensued (Neuharth 2019). Air exposure also occurs when

carbonated beverages are poured over injured gills, which could have negative physiological effects (Cooke and Sneddon 2007). Given the growing popularity of this technique yet uncertainty related to its effectiveness, we set out to provide the first scientific evidence whether carbonated beverages should be considered a best practice for C&R because they reduce or stop bleeding from the gills of teleost fish. For this study, we angled Northern Pike (*Esox lucius*) and simulated a hooking injury to a standardized section of gill filaments. We then poured carbonated beverages over the gills (i.e., Coca Cola, Mountain Dew, or carbonated lake water), and quantified the duration and intensity of bleeding, as well as overall blood loss, in comparison to a control. Since metabolism and related blood flow in fish are positively correlated with water temperature, we also tested the effects of carbonated beverages on bleeding in Northern Pike at two different temperature regimes. Further, at the warmer water temperature we tested whether use of chilled beverages influenced outcomes relative to beverages at ambient temperatures.

Methods

Animal Welfare

All experiments were conducted in accordance to regulations and guidelines set by the Canadian Council on Animal Care (Carleton University protocol AUP #110558), and veterinary professionals were consulted during project development. Northern Pike Fish were collected under Scientific Collection Permit #08577 from the Ontario Ministry of Natural Resources and Forestry.

Fish Capture

Northern Pike were selected for study owing to their relevance in addressing the question of interest and their popularity as a target recreational angling species (Paukert et al. 2001). The study was conducted on wild Northern Pike from Lake Opinicon, Ontario, Canada (44.5590° N, 76.3280° W), and all fish were captured from a boat using conventional medium-heavy rod and reel and a variety of crankbaits, chatter baits, and spinner baits. Once hooked, fish were retrieved, brought into the boat using a rubberized landing net, and immediately transferred to a water-filled trough for hook removal (underwater). Only fish that were hooked in the jaw were used in experiments to avoid confounding effects between lure-induced gill damage and experimental gill injury. In addition, fish that were bleeding from hooking site or the gills upon capture (<10% of fish) were not used in the study and were immediately released. Following hook removal, the total length (TL, mm) of the fish was recorded.

Experimental injury and post-injury monitoring

Gill injuries were simulated by using end-cutting pliers to remove a 0.9 cm by 0.9 cm section of gill filaments from the right middle gill arch (Fig. 1). The procedure was conducted while fish were held in the water-filled trough to eliminate the air exposure. Once gills were clipped, one of three carbonated beverage treatments or the lake water control was applied (details below), and fish transferred to a white bottomed cooler (52 cm x 26.5 cm) containing nonaerated lake water (~25 liters). When pouring carbonated beverages over the gills, a standardized volume of 150 mL was used. We also included

a reference or baseline group of fish that did not have any gill filaments clipped nor liquid poured over the gills before being transferred to the cooler.

Experiments were conducted at two different time periods. *Experiment 1* was conducted in May 2019 when water temperature ranged between 11-18 °C, whereas *Experiment 2* took place in August 2019 when water temperature ranged from 24-27 °C. *Experiment 1* had five treatment groups: 1) baseline, where the gill was not injured, and no carbonated beverages were used; 2) post injury, fish were held in lake water (pH = 6.71) without any other treatment; 3) post injury, carbonated lake water (pH = 4.37) poured over the gills; 4) post injury, Mountain Dew (pH = 3.27) poured over the gills; and 5) post injury, Coca-Cola (pH = 2.56) poured over the gills. During *Experiment 1*, beverage temperature ranged from 11-18 °C according to ambient air temperature. *Experiment 2* included baseline and control treatments (lake water pH = 6.71) as *Experiment 1*, and compared Mountain Dew (pH = 3.27) at ambient temperature (24-27 °C) with Mountain Dew (pH = 3.27) kept on ice (4-8 °C), to mimic an angler either not using or using a cooler, respectively, to hold beverages.

Following treatment fish were individually held in a cooler and visually monitored for 20 min to quantify: 1) time to bleeding cessation; 2) gill colour, which served as a proxy for blood loss; and 3) bleeding intensity. Bleeding cessation time was recorded as the time from gill injury until noticeable bleeding from the gill area stopped. Gill colour was assessed against a 20-point colour gradient, with bright red (20) at one end of the scale representing gills that were well-perfused with blood (most common), through progressively lighter shades of red to pink, to nearly white (1). Gill colour values were recorded 10 min and 20 min post-injury, as well as immediately before the gill injury

occurred, serving as a reference. Relative bleeding intensity (BIN) was based on the following scale: 0, no bleeding; 1, minor bleeding, not obvious; 2, obviously bleeding, easily observed; and 3, intense bleeding, pulsatile blood flow. For all treatment groups other than the baseline group, we recorded bleeding intensity immediately before and after liquid was poured directly onto the wound while the fish was held in a water-filled trough. Additional bleeding intensity values were recorded at 3-min and 5-min post-injury. After 20 min in the cooler, the vigour and condition of the fish were recorded using reflex action mortality predictors (RAMP), and fish that were not moribund were released. Fish that were moribund were euthanized by cerebral percussion (3 fish in total; see below).

Survival and Reflexes

The presence or absence of basic reflexes can be used to predict post-release mortality of fishes (Davis 2007; Raby et al. 2015). Our RAMP scoring system evaluated: 1) ability to maintain equilibrium; 2) reaction to grasping the tail (burst swimming); and 3) vestibular-ocular response (i.e. eye tracking) to determine whether a fish was impaired post-treatment and should be euthanized or released.

Data Analysis

R Version 1.1.447, R Studio (R Core Team 2019) was used to conduct all statistical analyses. For both *Experiment 1* and 2, bleeding time was compared among treatments using linear regression models and significant effects were assessed using Type 1 sum of squares. For bleeding intensity (BIN; ordinal scale from 0 to 3) and gill

colour (ordinal scale from 1 to 20), ordinal logistic regression was applied to each time point at which BIN or gill colour was assessed. Full models included treatment, time, and their interaction as a fixed effect and individual as a random effect to account for repeated measures. Backward model selection was used to determine final model structure using Akaike Information Criterion (AIC). When best fit models included interactions, ordinal logistic regression models were fit within each sampling time period to assess differences among treatments. Statistical significance was accepted at $\alpha = 0.05$ and, unless otherwise noted, all values are presented as means \pm SEM.

Results

For Experiment 1, 118 Northern Pike (50.9 ± 6.6 cm TL) were captured, while 38 Northern Pike (52.6 ± 6.7 cm TL, $n = 38$) were captured for *Experiment 2*. There were no significant differences in TL among the treatments for either experiment (*Experiment 1*, $F_4 = 0.641$, $p = 0.634$; *Experiment 2*, $F_3 = 0.583$, $p = 0.629$).

Time to Bleeding Cessation

Time to bleeding cessation in *Experiment 1* ranged from 0 to 690 s (mean 193 ± 95 s) and was not significantly different among treatments (Fig. 2; $F_3 = 0.83$, $p = 0.48$). For *Experiment 2*, time to bleeding cessation ranged from 0 to 193 s (mean 87 ± 40 s) and also not different among treatments (Fig. 2; $F_2 = 2.47$, $p = 0.10$).

Gill Colour Index

For *Experiment 1*, the best fitting model for gill colour index included a significant interaction between treatment and time (Table 1). Gill colour index did not differ among treatment groups prior to gill injury. Post injury, the baseline treatment (no injury) group exhibited significantly darker colour (higher score; Fig. 3) than all other treatment groups at both 10 min ($t_{126} = 3.60$, $p < 0.001$) and 20 min ($t_{127} = 5.03$, $p < 0.001$). However, no significant differences were detected among the control group (immersion in lake water) and any group treated with a carbonated beverage ($t < 5.03$, $p > 0.05$; Table 2). In *Experiment 2* there was also a significant interaction between treatment and time (Table 1). The baseline treatment (no injury) had significantly higher colour score (darker colour) than all other groups both 10 min ($t_{31} = 1.78$, $p < 0.001$) and 20 min ($t_{31} = 3.62$, $p < 0.001$) post-injury (Fig. 4). There were no significant differences were detected among the control group and use of chilled Mountain Dew ($t < -0.206$, $p > 0.05$). Fish had a significantly darker gill colour at 10 minutes when ambient temperature Mountain Dew was used in comparison to the control group ($t_{31} = -2.309$, $p = 0.021$; Table 2).

Bleeding Intensity Values

For *Experiment 1*, bleeding intensity was not different among treatments (BIN; Fig. 5A) as the best fitting model to the data included time as a fixed effect and individual as a random effect to account for repeated measures (Table 3). Similarly, treatment was not a significant contributing factor to variation in BIN in *Experiments 2* as the best fitting model to the data included time and individuals as independent predictors (Table 3).

228 *Survival and Reflexes*

229 In *Experiment 1*, no significant effect of treatment group on RAMP score was
 230 detected ($F_4 = 1.008$, $p = 0.405$). In *Experiment 2*, a significant treatment effect was
 231 detected ($F_3 = 4.002$, $p = 0.013$), with fish subjected to chilled Mountain Dew exhibiting
 232 significantly higher impairment than the baseline group ($t_{46} = -3.43$, $p = 0.001$). Only 3
 233 fish were euthanized owing to low RAMP score (2 fish in the carbonated lake water
 234 group and 1 in the Coca Cola group during *Experiment 1*).

235

236 **Discussion**

237 The main claim of those in the recreational angling community promoting the use
 238 of carbonated beverages is that this practice decreases the duration bleeding related to
 239 hooking injury, particularly of the gills (Pyzer 2019). However, our study found no
 240 differences in the time to cessation of bleeding, gill colour (which was used as an index
 241 of blood loss), or bleeding intensity among three carbonated beverages poured over
 242 bleeding gills, or between the carbonated beverages and a control group. Differences
 243 were only detected between the baseline group where no simulated hooking injury
 244 occurred, and all other groups in which injury and bleeding happened. These findings
 245 provide direct scientific evidence that the use of carbonated beverages does not curtail
 246 bleeding from gills, which is counter to anecdotal observations made by recreational
 247 anglers that use this technique. Overall, our study debunks the assertion that pouring
 248 carbonated soft drinks on the gills of injured via recreational angling is a best practice
 249 for C&R at least in the context studied here.

Although some online forums assert that exposure to CO₂ from carbonated beverages would be beneficial for reducing or even eliminating bleeding from the fills of fish, scientific studies suggest that exposure to high environmental CO₂ has negative effects on fishes (Perry and Abdallah 2012; Kaya et al. 2016; Tresguerres et al. 2019). For example, CO₂ excretion and acid-base regulation are impacted by acute exposure to elevated CO₂ and high water CO₂ triggers cardiorespiratory reflexes such as hyperventilation and bradycardia (Taylor 1988; Gilmour 2001; Perry and McKendry 2001; Claiborne et al. 2002; Perry and Reid 2002; Gilmour et al. 2005; Gilmour 2012). As such, the physiological effects of acute CO₂ exposure imply that carbonated beverages are unlikely to benefit the well-being of fishes.

Hypoxia-induced bradycardia in combination with CO₂-induced bradycardia may present a perceived positive effect when carbonated beverages are initially poured on fish gills that are bleeding. Many anglers lift fish out of the water once landed, exposing them to air, promoting hypoxia (Cooke and Sneddon 2007), which can induce bradycardia (Randall 1982; Farrell 2007). This effect of air exposure on fish has been well characterized in angled fish (Cooke et al. 2002; 2003; Furimsky et al., 2003). Therefore, when fish are removed from the water so that carbonated beverages can be used on the gills, the perception that bleeding has ceased is possible, but the effect is largely driven by hypoxia-induced decreases in cardiovascular output (Reid and Perry 2003; Perry and Desforges 2006). For our study, fish were held horizontally in a water-filled trough such that the gills were constantly submerged in well-oxygenated water. This approach should have limited hypoxia-induced bradycardia allowing effects of the carbonated beverages to be detected. Because we used white coolers, CO₂-induced

bradycardia effects were apparent. Fish would not bleed for >30 s, followed by blood spurting from the gills. If a fish were to be released by an angler or held in the water, this bleeding might not be apparent, and could account for reports of carbonated beverages stopping blood loss (for a short period).

Independent of the effects of carbonated beverages on bleeding, these acidic solutions may have damaging effects on fish gill tissues. Low pH water has been shown to cause a general inflammatory response, an increase in mucous production, and alterations in the structural morphology of the gills in teleost fish (Meyer et al. 2009). Although many of these studies use more chronic exposures (i.e. > 24 h) than used in our study, acute exposure may still have negative effects on fish (Meyer et al. 2009), particularly on ion and acid-base regulation (Wright and Wood 2009). Interestingly, Northern Pike are relatively tolerant of environmental acidification and have reported to survive in water of pH 3.5 (Zaprudnova et al. 2015), which may allow them to cope with an acute acid exposure.

Further work should address how acute instances of branchial acid exposure can affect ion and acid-base status in this context to fully appreciate the biological consequences of using carbonated beverages in an angling setting. Also, given that we limited our observations for 20 min during our study, we did not assess more chronic physiological can behavioral effects, as well as post-release mortality. Although Northern Pike are regarded as being relatively robust to hooking injury (Arlinghaus et al. 2008), and that we observed little immediate or short-term mortality in fish that were actually injured during capture for this study, the longer-term consequences of pouring chemicals associated with carbonated sodas remains unknown. Future work involving

telemetry or net pens would be useful for understanding longer-term consequences of this carbonated beverage technique. Lastly, our study used a simulated and standardized size of gill injury to a single species of fish, so further investigation is are needed to determine whether the magnitude of injury or species-specific differences would produce different results.

Overall, we provide the first evidence that counters the growing popularity of using carbonated beverages to stop bleeding in angling-caught fish. Our observations made during the 20 min holding period versus immediately release as many anglers do after they pour carbonated beverages on injured gills, sheds light on the potential perception of the curtailment of bleeding anglers witness, especially if the combination of air exposure and CO₂ cause an immediate and severe bradycardia. We found no benefit or disbenefit with pouring carbonated beverages over the gills of Northern Pike, but it is possible that there are longer term impacts. Similarly, our findings are specific to the context studied here. As such, it is possible that carbonated beverages could provide benefit or harm when used in other contexts, such as with other species (e.g., salmonids, muskellunge), using other beverages, or applying the beverages in other ways (e.g., holding them in air for longer after applying beverage). This study contributes to the growing body of literature that emphasizes the need for anglers and fisheries scientists to work collaboratively to ensure that best practices being employed truly benefit fish (Brownscombe et al. 2017).

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References

- Anderson, J. 2018. Save a million fish - home.
- <https://www.facebook.com/pages/category/Environmental-Conservation-Organization/Save-a-Million-Fish-752045115140863/> Accessed 8 March 2020.
- Arlinghaus, R., S. J. Cooke, J. Lyman, D. Policansky, A. Schwab, C. Suski, S. G. Sutton, and E. B. Thorstad. 2007. Understanding the complexity of catch-and-release in recreational fishing: an integrative synthesis of global knowledge from historical, ethical, social, and biological perspectives. *Reviews in Fisheries Science* 15(1–2):75–167. <https://doi.org/10.1080/10641260601149432>
- Arlinghaus, R., T. Klefoth, A. Kobler, and S. J. Cooke. 2008. Size selectivity, injury, handling time, and determinants of initial hooking mortality in recreational angling for Northern Pike : the influence of type and size of bait. *North American Journal of Fisheries Management* 28:123–134. <https://doi.org/10.1577/M06-263.1>
- Bardin, S. 2019. Pouring soda on fish gills: does it actually work? https://www.wired2fish.com/biology/pouring-soda-on-fish-gills-does-it-actually-work/#slide_3 Accessed 14 May 2020.
- Brauner, C. J., R. B. Shartau, C. Damsgaard, A. J. Esbaugh, R. W. Wilson, and M. Grosell. 2019. Acid-base physiology and CO₂ homeostasis: regulation and compensation in response to elevated environmental CO₂. *Fish Physiology* 37:69-132. <https://doi.org/10.1016/bs.fp.2019.08.003>
- Brownscombe, J. W., A. J. Danylchuk, J. M. Chapman, L. F. G. Gutowsky, and S. J. Cooke. 2017. Best practices for catch-and-release recreational fisheries –

- 354 angling tools and tactics. Fisheries Research 186:693–705.
- 355 <https://doi.org/10.1016/j.fishres.2016.04.018>
- 356 Claiborne, J. B., S. L. Edwards, and A. I. Morrison-Shetlar. 2002. Acid-base regulation
- 357 in fishes: cellular and molecular mechanisms. Journal of Experimental Zoology
- 358 293(3):302–319. <https://doi.org/10.1002/jez.10125>
- 359 Coe, A. J., A. J. Picard, and M. G. Jonz. 2017. Purinergic and adenosine receptors
- 360 contribute to hypoxic hyperventilation in zebrafish (*Danio rerio*). Comparative
- 361 Biochemistry and Physiology Part A: Molecular and Integrative Physiology
- 362 214:50–57. <https://doi.org/10.1016/j.cbpa.2017.09.013>
- 363 Cooke, S. J., and I. G. Cowx. 2004. The role of recreational fishing in global fish crises.
- 364 BioScience 54(9):857-859. [https://doi.org/10.1641/0006-](https://doi.org/10.1641/0006-3568(2004)054[0857:trorfi]2.0.co;2)
- 365 [3568\(2004\)054\[0857:trorfi\]2.0.co;2](https://doi.org/10.1641/0006-3568(2004)054[0857:trorfi]2.0.co;2)
- 366 Cooke, S. J., K. G. Ostrand, C. M. Bunt, J. F. Schreer, D. H. Wahl, and D. P. Philipp.
- 367 2003. Cardiovascular responses of Largemouth Bass to exhaustive exercise and
- 368 brief air exposure over a range of water temperatures. Transactions of the
- 369 American Fisheries Society 132(6):1154–1165. <https://doi.org/10.1577/t02-059>
- 370 Cooke, S. J., and H. L. Schramm. 2007. Catch-and-release science and its application
- 371 to conservation and management of recreational fisheries. Fisheries
- 372 Management and Ecology 14(2):73–79. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2400.2007.00527.x)
- 373 [2400.2007.00527.x](https://doi.org/10.1111/j.1365-2400.2007.00527.x)
- 374 Cooke, S. J., J. F. Schreer, D. H. Wahl, and D. P. Philipp. 2002. Physiological impacts
- 375 of catch-and-release angling practices on Largemouth Bass and Smallmouth
- 376 Bass. American Fisheries Society Symposium 31:489–512.

- Cooke, S. J., and L. U. Sneddon. 2007. Animal welfare perspectives on recreational angling. *Applied Animal Behaviour Science* 104(3–4):176–198.
<https://doi.org/10.1016/j.applanim.2006.09.002>
- Davis, M. W. 2007. Simulated fishing experiments for predicting delayed mortality rates using reflex impairment in restrained fish. *ICES Journal of Marine Science* 64(8):1535–1542. <https://doi.org/10.1093/icesjms/fsm087>
- Davis, M. W. 2010. Fish stress and mortality can be predicted using reflex impairment. *Fish and Fisheries* 11:1–11. <https://doi.org/10.1111/j.1467-2979.2009.00331.x>
- Evans, D. H., P. M. Piermarini, and K. P. Choe. 2005. The multifunctional fish gill: dominant site of gas exchange, osmoregulation, acid-base regulation, and excretion of nitrogenous waste. *Physiological Reviews* 85:97–177.
<https://doi.org/10.1152/physrev.00050.2003>
- Farrell, A. P. 2007. Tribute to P. L. Lutz: A message from the heart - why hypoxic bradycardia in fishes? *Journal of Experimental Biology* 210(10):1715–1725.
<https://doi.org/10.1242/jeb.02781>
- Furimsky, M., S. J. Cooke, C. D. Suski, Y. Wang, and B. L. Tufts. 2003. Respiratory and circulatory responses to hypoxia in Largemouth Bass and Smallmouth Bass: implications for “live-release” angling tournaments. *Transactions of the American Fisheries Society* 132(6):1065–1075. <https://doi.org/10.1577/t02-147>
- Gilmour, K. M. 2010. Perspectives on carbonic anhydrase. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology* 157(3):193–197.
<https://doi.org/10.1016/j.cbpa.2010.06.161>

- Gilmour, K. M., W. K. Milsom, F. T. Rantin, S. G. Reid, and S. F. Perry. 2005. Cardiorespiratory responses to hypercarbia in tambaqui *Colossoma macropomum*: chemoreceptor orientation and specificity. *Journal of Experimental Biology*, 208(6):1095–1107. <https://doi.org/10.1242/jeb.01480>
- Gilmour, K. M., and S. F. Perry. 2009. New insights into the many functions of carbonic anhydrase in fish gills. *Respiratory Physiology and Neurobiology* 184:223– 230. <https://doi.org/10.1016/j.resp.2012.06.001>
- Gilmour, K. M. 2001. The CO₂/pH ventilatory drive in fish. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology* 130:219–240. [https://doi.org/10.1016/S1095-6433\(01\)00391-9](https://doi.org/10.1016/S1095-6433(01)00391-9)
- Green, J. 2015. Can you stop a fish bleeding from the gills with coke or sprite? Nodak Angler. <https://nodakangler.com/forums/content/373-Can-You-Stop-A-Fish-Bleeding-From-The-Gills-With-Coke-Or-Sprite> Accessed 8 March 2020.
- Kaya, H., O. Hisar, S. Yilmaz, M. Gürkan, and Ş. A. Hisar. 2016. The effects of elevated carbon dioxide and temperature levels on tilapia (*Oreochromis mossambicus*): respiratory enzymes, blood pH and hematological parameters. *Environmental Toxicology and Pharmacology* 44:114–119. <https://doi.org/10.1016/j.etap.2016.05.003>
- Kwong, R. W. M., Y. Kumai, and S. F. Perry. 2014. The physiology of fish at low pH: the zebrafish as a model system. *Journal of Experimental Biology* 217:651-662. <https://doi.org/10.1242/jeb.091603>
- Meyer, E. A., R. L. Cramp, and C. E. Franklin. 2009. Damage to the gills and integument of *Litoria fallax* larvae (Amphibia: Anura) associated with

ionoregulatory disturbance at low pH. Comparative Biochemistry and Physiology

Part A: Molecular & Integrative Physiology 155(2):164-171

<https://doi.org/10.1016/j.cbpa.2009.10.032>

Muoneke, M. I., and W. M. Childress. 1994. Hooking mortality: a review for recreational fisheries. Reviews in Fisheries Science 2(2):123-156.

<https://doi.org/10.1080/10641269409388555>

Paukert, C. P., J. A. Klammer, R. B. Pierce, and T. D. Simonson. 2001. An overview of Northern Pike regulations in North America. Fisheries 26(6):6–13.

[https://doi.org/10.1577/1548-8446\(2001\)026<0006:aoonpr>2.0.co;2](https://doi.org/10.1577/1548-8446(2001)026<0006:aoonpr>2.0.co;2)

Perry, S. F., and P. R. Desforges. 2006. Does bradycardia or hypertension enhance gas transfer in rainbow trout (*Oncorhynchus mykiss*)? Comparative Biochemistry and Physiology - A Molecular and Integrative Physiology 144(2):163–172.

<https://doi.org/10.1016/j.cbpa.2006.02.026>

Perry, S. F., and J. E. McKendry. 2001. The relative roles of external and internal CO₂ versus H⁺ in eliciting the cardiorespiratory responses of *Salmo salar* and *Squalus acanthias* to hypercarbia. Journal of Experimental Biology, 204(22):3963–3971.

Perry, S. F., and Abdallah, S. (2012). Mechanisms and consequences of carbon dioxide sensing in fish. Respiratory Physiology and Neurobiology 184:309– 315.

<https://doi.org/10.1016/j.resp.2012.06.013>

Perry, S. F., and S. G. Reid. 2002. Cardiorespiratory adjustments during hypercarbia in rainbow trout *Oncorhynchus mykiss* are initiated by external CO₂ receptors on the first gill arch. Journal of Experimental Biology, 205(21):3357–3365.

- Pyzer, G. 2015. Coke: it's the real thing to stop fish bleeding. Outdoor Canada.
<https://www.outdoorcanada.ca/coke-its-the-real-thing-to-stop-fish-bleeding/>
Accessed 30 May 2019.
- Pyzer, G. 2019. Stop the bleeding: how soda water can save an injured fish.
<https://www.outdoorcanada.ca/stop-the-bleeding-how-soda-water-can-save-an-injured-fish/> Access 14 January 2020.
- Raby G. D., M. R. Donaldson, S. G Hinch, D. A. Patterson, A. G. Lotto, D. Robichaud, K. K. English, W. G. Willmore, A. P Farrell, M. W. Davis, and S. J. Cooke. 2012. Validation of reflex indicators for measuring vitality and predicting the delayed mortality of wild coho salmon bycatch released from fishing gears. Journal of Applied Ecology 49:90–98. <https://doi.org/10.1111/j.1365-2664.2011.02073.x>
- Raby, G. D., S. G. Hinch, D. A. Patterson, J. A. Hills, L. A. Thompson, and S. J. Cooke. 2015. Mechanisms to explain purse seine bycatch mortality of coho salmon. Ecological Applications, 25(7), 1757–1775. <https://doi.org/10.1890/14-0798.1>
- Randall, D. 1982. The Control of Respiration and Circulation in Fish During Exercise and Hypoxia. Journal of Experimental Biology, 100(1), 275–288.
- Reid, S. G., and S. F. Perry. 2003. Peripheral O₂ chemoreceptors mediate humoral catecholamine secretion from fish chromaffin cells. American Journal of Physiology - Regulatory Integrative and Comparative Physiology, 284(4 53-4), 990–999. <https://doi.org/10.1152/ajpregu.00412.2002>
- Reid, S. G., L. Sundin, A. L. Kalinin, F. T. Rantin, and W. K. Milsom. 2000. Cardiovascular and respiratory reflexes in the tropical fish, traíra (Hoplias

malabaricus): CO₂/pH chemoresponses. *Respiration Physiology*, 120(1):47–59.
[https://doi.org/10.1016/S0034-5687\(99\)00100-0](https://doi.org/10.1016/S0034-5687(99)00100-0)

Neuharth, S. 2019. Stop pouring soda on fish gills.
<http://themeateater.com/fish/bass/stop-pouring-soda-on-fish-gills?> Accessed 10
 June 2019.

Taylor, E. W. 1988. Acid-base regulation in animals. *FEBS Letters* 233(1):216–217.
[https://doi.org/10.1016/0014-5793\(88\)81396-6](https://doi.org/10.1016/0014-5793(88)81396-6)

Tresguerres, M., W. K. Milsom, and S. F. Perry. 2019. CO₂ and acid-base sensing. *Fish Physiology* 37:33–68. <https://doi.org/10.1016/bs.fp.2019.07.001>

Wedemeyer, G. A., and R. S. Wydoski. 2008. Physiological response of some
 economically important freshwater salmonids to catch-and-release fishing. *North
 American Journal of Fisheries Management*, 28(5):1587–1596.
<https://doi.org/10.1577/m07-186.1>

Wendelaar Bonga, S. E. 1997. The stress response in fish. *Physiological Reviews*.
 American Physiological Society. <https://doi.org/10.1152/physrev.1997.77.3.591>

Wood, C. M., and P. G. McDonough. 1988. Impact of environmental acidification on gill
 function in fish. Pages 162-182 in R. C. Ryans, editor. *Fish physiology, fish
 toxicology and fisheries management*. Environmental Protection Agency,
 International Symposium, Guangzhou.

Wright, P. A., and C. M. Wood. 2009. A new paradigm for ammonia excretion in aquatic
 animals: Role of rhesus (RH) glycoproteins. *Journal of Experimental Biology*
 212:2303-2312. <https://doi.org/10.1242/jeb.023085>

488 Zaprudnova, R. A., I. M. Kamshilov, and Y. P. Chalov. 2015. Functional properties of
 489 hemoglobin during the adaptation of fish to low environmental pH. *Inland Water*
 490 *Biology*, 8(2):188–194. <https://doi.org/10.1134/S1995082915020157>

491 Wydoski, R.S. 1977. Relation of hooking mortality and sublethal hooking stress to
 492 quality fishery management. In: Barnhart RA, Roelofs TD (eds) *Catch-and-*
 493 *release fishing as a management tool*. Humbolt State University, Arcata,
 494 California, USA, 43–87.

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Figure Captions

Figure 1. Image of the standardized 0.9 cm by 0.9 cm section of gill filaments removed from Northern Pike (*Esox lucius*) to simulate hooking injury.

Figure 2. Time to cessation of bleeding following gill injury in Northern Pike in Experiment 1 (A) and Experiment 2 (B). In A, control is compared against carbonated lake water (Carbonated LW), Mountain Dew, and Coca Cola. In B) control is compared against Mountain Dew at 4-8°C (chilled Mountain Dew) and Mountain Dew at ambient (24-27°C (Reg Mountain Dew).

Figure 3. Gill Colour Index for *Experiment 1* for 0 minutes (A), 10 minutes (B), 20 minutes (C) and the relative change from 0 to 10 minutes and 10 and 20 minutes (D). In all graphs control is compared against carbonated lake water (Carbonated LW), Mountain Dew, and Coca Cola.

Figure 4. Gill Colour Index for Experiment 2 for 0 minutes (A), 10 minutes (B), 20 minutes (C) and the relative change from 0 to 10 minutes and 10 and 20 minutes (D). In all graphs control is compared against Mountain Dew at 4-8 °C (chilled Mountain Dew) and Mountain Dew at ambient temperature, 24-27 °C (Reg Mountain Dew).

Figure 5. Bleeding intensity values following gill injury in Northern Pike in Experiment 1 (A) and Experiment 2 (B). Figures show the bleeding intensity values before the pour of

any substance, after the cut (Before Pour), after the pour of a substance (After), after 3 minutes (3min Post) and 5 minutes after the substance was poured over the gills (5min Post). In A, control is compared against carbonated lake water (Carbonated LW), Mountain Dew, and Coca Cola. In B) control is compared against Mountain Dew at 4-8°C (chilled Mountain Dew) and Mountain Dew at ambient (24-27°C (Reg Mountain Dew)).

Tables

Table 1: Model selection outputs for ordinal logistic regression models. Full models included gill color as the response, the interaction between treatment and time as fixed effects, and individual as a random effect. Backward model selection was used to determine final model structure. Significant differences in model fit are highlighted by bold italic font.

Experiment 1 (n = 118), Experiment 2 (n = 38)

Experiment	Model Specification	AIC	Residual Deviation	Loglikelihood	P value	DF
1	<i>Treatment*Time</i>	<i>2014.924</i>	<i>1954.924</i>	<i>25.575</i>	<i><0.001</i>	<i>8</i>
	Treatment	2086.274	2046.274	-29.198	1.000	2
	<i>Time</i>	<i>2053.076</i>	<i>2017.075</i>	<i>59.600</i>	<i><0.001</i>	<i>2</i>
2	<i>Treatment*Time</i>	<i>626.000</i>	<i>572.000</i>	<i>13.7312436</i>	<i>0.033</i>	<i>6</i>
	Treatment	652.3781	614.378	-0.0592199	1.000	1
	<i>Time</i>	<i>650.3189</i>	<i>614.318</i>	<i>21.8639438</i>	<i><0.001</i>	<i>2</i>

Table 2: Ordinal logistic regression model outputs for gill colour index with treatment as a predictor using control values as reference group for comparisons at individual time periods, 0, 10 and 20 minute. Significant differences are highlighted by bold italic font.

Experiment 1 (n=118, df = 439)

Time	Comparisons	Value	Standard Error	T value	P-value
0 Minutes	Carbonated Lake Water	0.14346940	0.4523524	0.31716292	0.751
	Mountain Dew	0.61190184	0.4527499	1.35152284	0.176
	Coca Cola	0.14213540	0.4630298	0.30696813	0.758
	Baseline	0.00598101	0.4948112	0.01208746	0.990
10 Minutes	Carbonated Lake Water	-0.40226948	0.4550232	-0.88406369	0.376
	Mountain Dew	-0.19233292	0.4416917	-0.43544612	0.663

	Coca Cola	-0.50898612	0.4646763	-1.09535621	0.273
	Baseline	1.78152855	0.4945991	3.60196492	<0.001
20 Minutes	Carbonated Lake Water	0.1578421	0.4628604	0.3410145	0.733
	Mountain Dew	0.6228802	0.4417568	1.4100071	0.158
	Coca Cola	-0.05273126	0.4547337	-0.1159608	0.907
	Baseline	2.500492	0.4969529	5.0316475	<0.001

Experiment 2 (n=38, df = 146)

Time	Comparisons	Value	Standard Error	T value	P-value
0 Minutes	Chilled Mountain Dew	-0.1457	0.7063	-0.2063	0.8366
	Regular Mountain Dew	-0.4886	0.6783	-0.7204	0.4713
	Baseline	0.0071	0.7534	0.0094	0.9925
10 Minutes	Chilled Mountain Dew	-0.8734	0.7416	-1.1777	0.2389
	Regular Mountain Dew	-1.6813	0.7281	-2.3093	0.0209
	Baseline	1.3084	0.7342	1.7821	0.0747
20 Minutes	Chilled Mountain Dew	-1.2230	0.7538	-1.6225	0.1047
	Regular Mountain Dew	-0.2734	0.7305	-0.3742	0.7082
	Baseline	3.3882	0.9348	3.6243	<0.001

Table 3: Model selection outputs for ordinal logistic regression models. Full models included bleeding intensity as the response, the interaction between treatment and time as fixed effects, and individual as a random effect. Backward model selection was used to determine final model structure. Significant differences in model fit are highlighted by bold italic font.

Experiment 1 (n = 118), Experiment 2 (n = 38)

Experiment	Model Specification	AIC	Residual Deviation	Loglikelihood	P value	DF
1	Treatment*Time	531.4823	495.4823	9.473	0.394	9
	Treatment	1030.8998	1018.8998	0.059	0.996	3
	Time	520.6760	508.6760	510.223	<0.001	1
2	Treatment*Time	230.3625	202.3625	1.7381	0.942	6
	Treatment	361.4302	351.4302	0.600	0.741	2
	Time	217.4912	205.4912	145.939	<0.001	1

Figure 1

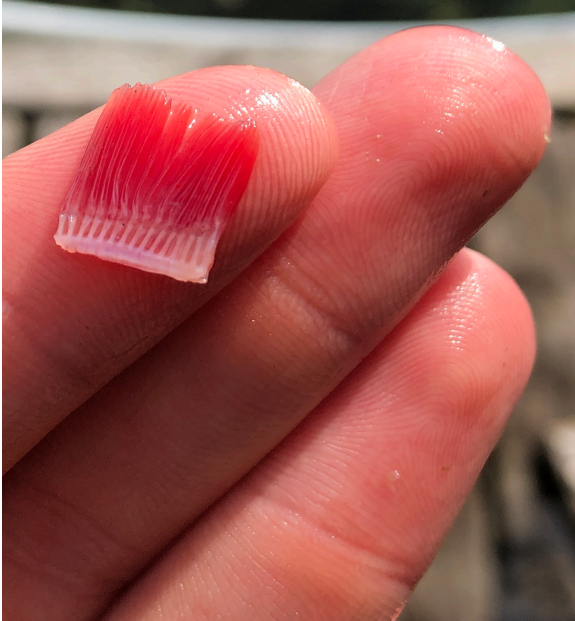


Figure 2

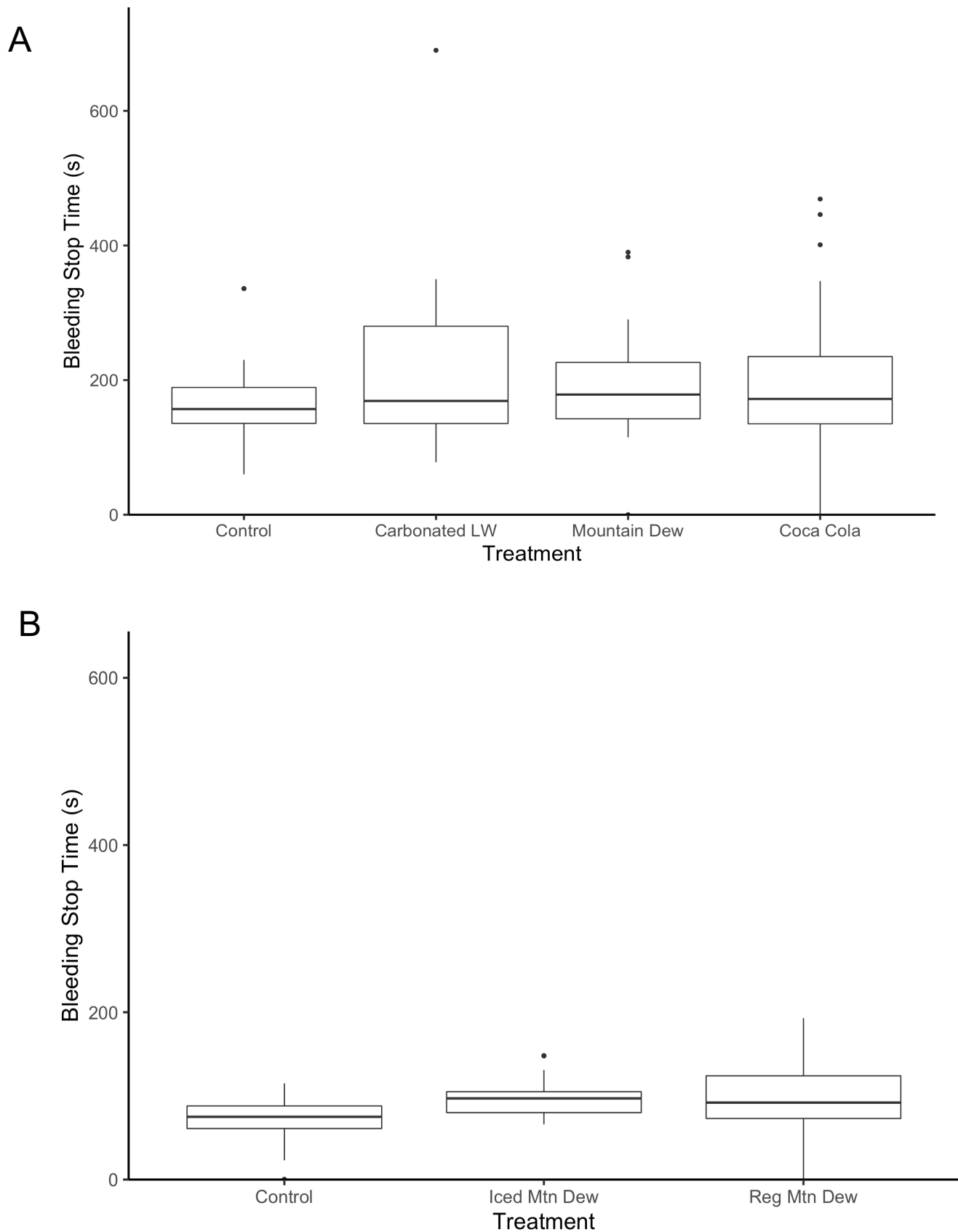


Figure 3

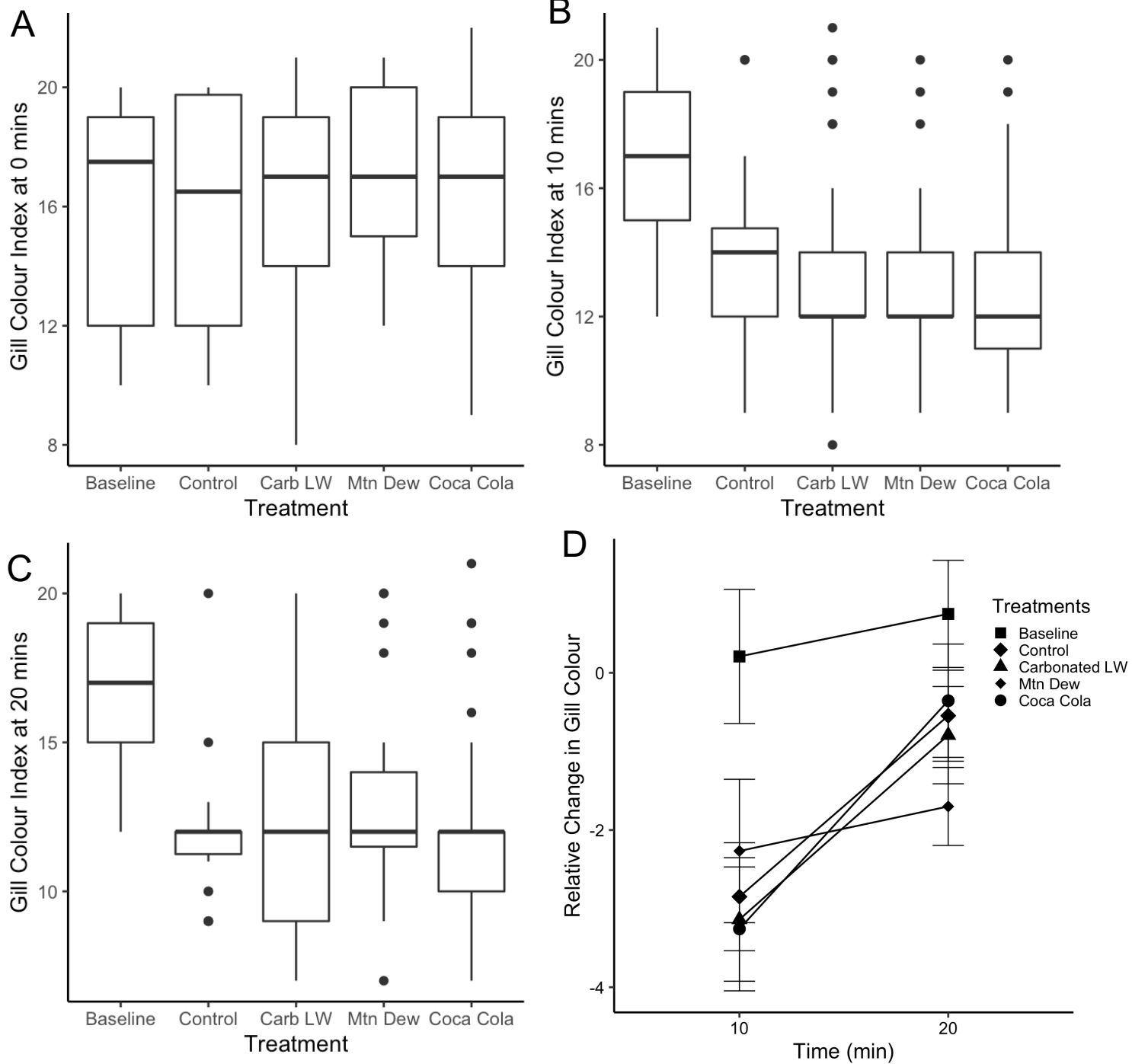


Figure 4

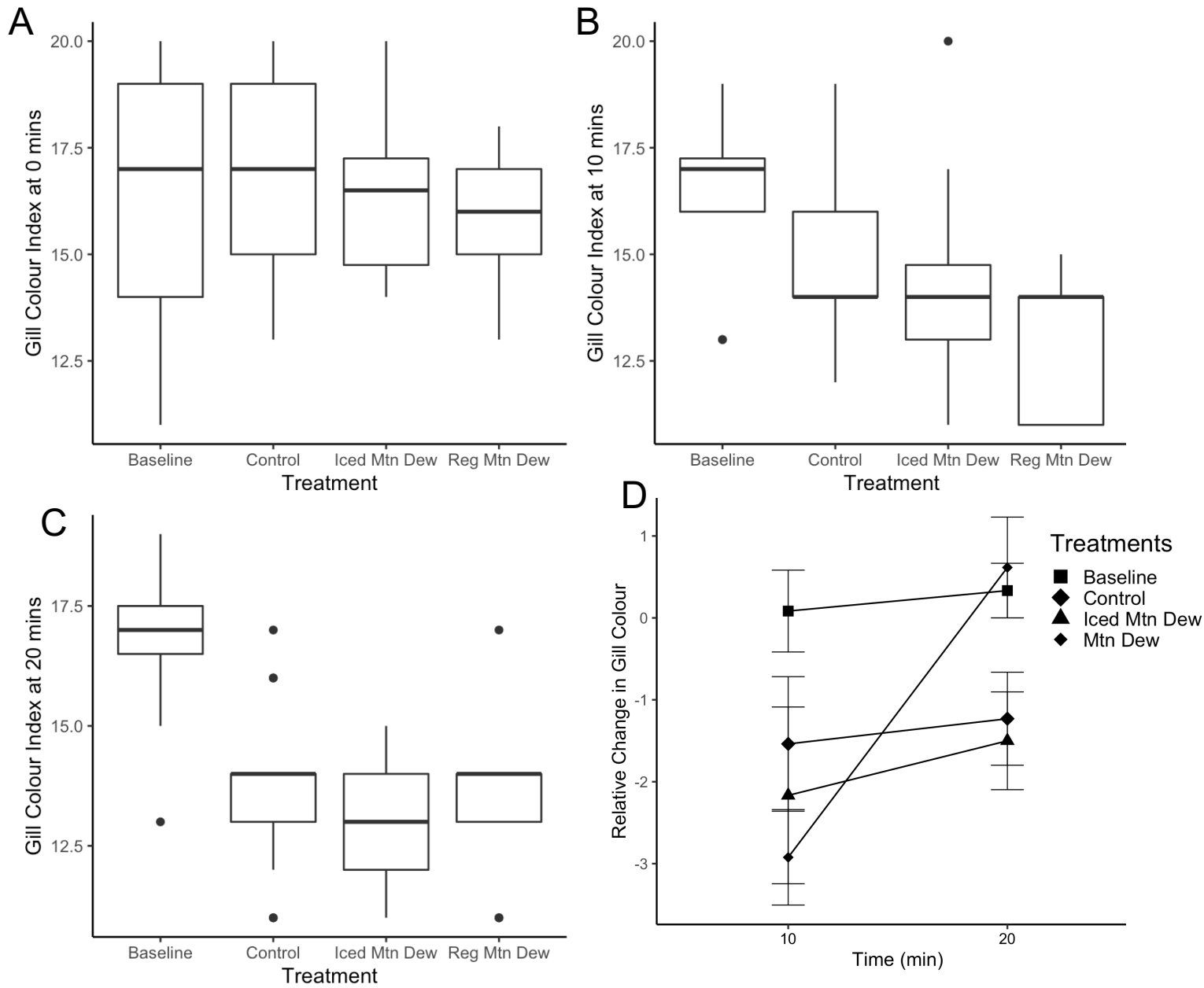


Figure 5

