

1 **Spatial context of trait variation: morphology of spotted salamanders (*Ambystoma***
2 ***maculatum*) varies more within ponds than between ponds**

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4 Elizabeth T. Green^{1,2}, Anthony I. Dell^{1,3}, John A. Crawford¹, Elizabeth G. Biro^{3,4}, David R.
5 Daversa^{1,3,5*}

6

7 ¹ National Great Rivers Research and Education Center (NGRREC), East Alton, IL 62024,
8 USA

9 ² Department of Biology, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599,
10 USA

11 ³ Department of Biology, Washington University in St. Louis, St. Louis, MO 63130, USA

12 ⁴ Tyson Research Center, Washington University in St. Louis, St. Louis, MO 63130, USA

13 ⁵ La Kretz Center for California Conservation Science, University of California, Los Angeles,
14 Los Angeles, CA, 90095, USA

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17 *Corresponding Author: Dave Daversa, ddaversa@gmail.com, ORCID: 0000-0002-8984-
18 8897

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22

23 **Abstract**

24 The influence of intraspecific trait variation on species interactions makes trait-based
25 approaches critical to understanding eco-evolutionary processes. Because species occupy
26 habitats that are patchily distributed in space, advancement in trait-based ecology hinges
27 on understanding how trait variation is distributed within and between habitat patches.
28 We sampled larval spotted salamanders (*Ambystoma maculatum*) across spatially discrete
29 ponds to quantify within- and between-pond variation in mass, length, and their allometric
30 relationship. Between-pond variation explained 7–35% of total observed variation in the
31 length and shape of salamander larvae, depending on the body segment measured (i.e.,
32 head, body, or tail). Salamander tail morphology was more variable and exhibited more
33 between-pond variation than head or body morphology. Salamander mass was highly
34 variable and strongly correlated with total length. Allometric analysis revealed that the
35 slopes of mass-length relationships were similar across ponds, but that intercepts differed
36 across ponds. Preliminary evidence hinted that newly constructed ponds were a driver of
37 the observed differences in mass-length relationships. Pond construction may therefore
38 bolster trait diversity across the broader landscape, and in so doing instil more adaptive
39 potential of salamander populations under current and future environmental change.

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41 **Key Words:** Allometry; Amphibian; Conservation; Ecology; Eco-morphology; Intraspecific
42 Trait variation; Morphometrics; Phenotypic plasticity

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45 Morphological traits underpin many eco-evolutionary processes. The range of
46 morphology exhibited by individuals of a species, a form of intraspecific trait variation
47 (Bolnick et al., 2011), shapes the niche breadth of populations, which in turn affects their
48 resiliency to environmental disturbances and biological invasions (Tack et al., 2014).
49 Because of this, models of population and community dynamics that incorporate
50 intraspecific trait variation have become central to ecology and evolution (Bolnick et al.,
51 2011; Moran et al., 2016). A key outstanding challenge for this ‘trait-based’ paradigm is to
52 understand trait variation in a spatially explicit context (Violle et al., 2012; Moran et al.,
53 2016).

54 Species tend to occur in landscapes comprised of spatially discrete habitat patches,
55 and traits may vary both among individuals within habitat patches and among groups
56 across habitat patches. Theory predicts that the influence of trait variation on population
57 and community dynamics depends on how variation is partitioned within versus between
58 habitat patches (Moran et al., 2016; Banitz, 2019). Between-patch variation potentially
59 allows for a broader range of adaptive responses to environmental disturbances (Moran et
60 al., 2016) and antagonistic interactions (Tack et al., 2014) than does within-patch variation.
61 Alternatively, between-patch trait variation may heighten extinction risk by geographically
62 isolating trait diversity to specific habitat patches. Population and community dynamics are
63 influenced not just by the degree of intraspecific trait variation but also the relative
64 proportion of trait variation that occurs within- versus between-patches (Violle et al., 2012;
65 Moran et al., 2016).

66 Trait co-variation describes relationships between multiple individual traits.
67 Between-patch variation in how traits co-vary may also be influential in eco-evolutionary

68 processes (but see Evangelista et al., 2019). Measures of trait co-variation are useful for
69 describing species growth patterns (Hirst et al., 2014), their adaptive constraints (Voje et
70 al., 2014), and complex morphological characteristics such as body shape (Laughlin and
71 Messier, 2015). Practically, they provide a tool for filling data gaps in ecological models
72 (Madin et al., 2016). Allometric scaling of mass with body length is a basic form of trait co-
73 variation that is widely used for this purpose (Madin et al., 2016). Additionally,
74 morphometrics integrates co-variation between body length, depth, and sometimes width,
75 to characterize body shapes of individuals. Body shape, being a strong proxy of
76 performance and fitness, is arguably a better predictor of species adaptive responses to
77 environmental change than linear body measurements (Laughlin and Messier, 2015). As
78 such, shape and allometry can be powerful predictors of ecology and evolution in patchy
79 landscapes.

80 In this study, we assessed within- and between-patch trait variation and co-
81 variation in larval spotted salamanders (*Ambystoma maculatum*). We sought to quantify
82 the extent of between-patch differences in salamander mass and length, as well as
83 allometric and morphometric relationships among multiple traits. We sampled larval
84 spotted salamanders among a network of spatially discrete wetlands (i.e., ponds) and
85 measured body length and mass of 519 individuals. We used these data to quantify
86 individual variation in mass and length within and among ponds. We then examined the
87 contribution of between-patch (i.e., between-pond) differences to the total observed
88 variation and co-variation of those traits. As an initial exploration into potential drivers of
89 between-pond differences in salamander morphology, we also performed a preliminary

90 examination of whether salamander mass, length, mass-length allometry, and shape were
91 influenced by the age and predator density of ponds.

92

93 **Materials and Methods**

94 *Study species.* — Spotted salamanders are broadly distributed throughout the Northeastern
95 and Midwestern United States (Petranka, 1998). Spotted salamanders are semi-terrestrial
96 pond breeders, annually migrating from terrestrial hibernacula to reproduce in fishless
97 wetlands. Breeding in our study area occurs between March–April (Sexton et al., 1990).
98 After hatching from eggs, aquatic larvae develop and metamorphose in 6–10 weeks
99 (Petranka, 1998). Larvae feed on invertebrates and anuran tadpoles, and are themselves
100 prey for adult salamanders and larger aquatic invertebrates such as odonate larvae and
101 beetles (Urban, 2010). Spotted salamander populations are useful systems to study
102 morphological variation in a spatial context because: i) individuals occupy and move
103 among spatially discrete ponds that comprise functional metapopulations with patchy
104 habitat structure (Patrick et al., 2008); and ii) larval stages exhibit substantial
105 morphological plasticity in response to heterogeneity in biotic and abiotic conditions
106 (Scott, 1990; Urban, 2010; Shaffery and Relyea, 2015), which permits a range of trait
107 expressions across individuals and ponds.

108

109 *Field sampling and husbandry.* — We sampled multiple ponds in east-central Missouri,
110 distributed across three distinct conservation properties – Tyson Research Center (800
111 ha), Forest 44 Conservation Area (400 ha) and Shaw Nature Reserve (700 ha) (Fig. 1). We
112 focused on six ponds in which pilot surveys confirmed Spotted salamander larvae were

113 present. Three ponds (Mincke Pond, Arthur Christ Pond, Beth's Pond) were constructed in
114 2008 for research purposes and had similar sizes and dimensions (Burgett 2015). As part
115 of a separate experiment, Rotenone was applied to Beth's Pond in 2008. While this initially
116 reduced microbial biodiversity (Woods et al., 2016) the micro-organismal community
117 structure had returned to pre-treatment conditions well before sampling for this study
118 (Woods et al., 2016). The other three focal ponds (Salamander Pond, Forest 44 Pond, Shaw
119 Pond) were older and variable in size (Table S1). Salamander Pond was created in 1965,
120 and Forest 44 Pond and Shaw Pond between 1990 and 1996 (data extracted from Google
121 Earth Historical Imagery). All ponds contained multiple predators of larval spotted
122 salamanders including: dragonfly (*Anax* sp.) and damselfly (*Lestes* sp.) nymphs, diving
123 beetles (*Dytiscidae* sp.), hydrophilid beetles (*Tropisternus* sp), and adult newts
124 (*Notophthalmus viridescens*) (Tables S1 and S4; E.G. Biro unpublished data). All ponds were
125 located within forested habitats typical of temperate deciduous ecosystems found in the
126 Midwestern US. A major highway bisected the study area, separating Forest 44 and Shaw
127 ponds from the other four ponds.

128 We intensively sampled one pond per week (30 June–08 August 2016) by dip-
129 netting near the perimeter of the pond. This sampling design confounded between-pond
130 variation in spotted salamander morphology with possible temporal morphological
131 variation due to growth and development (Landberg and Azizi, 2010; Musseau et al., 2020).
132 To minimize the influence of temporal factors in salamander morphological variation, we
133 focused sampling on the latter stage of the developmental period of salamanders (Harrison
134 stage 45–46; Harrison, 1969), when growth and development had slowed (Landberg and
135 Azizi, 2010). At each sampling event, we dip-netted until 10 minutes passed without a

136 capture to maximize coverage of morphological variation within ponds. We retained all
137 larvae that did not show overt signs of injury or illness (e.g., damaged tails or legs,
138 tumorous growth, etc.) and immediately transported them to the National Great Rivers
139 Research and Education Center (NGRREC) – less than 1 hr drive – where they were housed
140 for seven days before being returned to their original ponds.

141 We housed larvae individually in circular plastic arenas (28 cm diameter) filled with
142 500 mL dechlorinated water (approximately 2.5 cm depth). Larvae were maintained at
143 18 °C with a 14:10 h light:dark cycle, consistent with ambient conditions at the surveyed
144 ponds. Salamanders were fed a single gray tree frog (*Hyla chrysoscelis-versicolor*) tadpole
145 on the fifth day as part of a separate experiment. Observing that not all salamanders ate the
146 tree frogs fed to them the prior day, we tested whether feeding influenced measures of
147 mass for a random subset of 237 individuals for which feeding data were available (see
148 Supplementary Material for detailed methods).

149 As an initial exploration into potential drivers of between-pond variation in
150 salamander morphology, we assessed whether salamander mass, length, mass-length co-
151 variation, and shape were influenced by the age and predator density of ponds. We
152 consider these assessments preliminary because of the low replication of ponds in our
153 sample (N = 6). We used historical information described above to classify pond age as
154 ‘new’ (N = 3; Mincke Pond, Arthur Christ Pond, Beth’s Pond) or ‘old’ (N = 3; Salamander
155 Pond, Forest 44 Pond, Shaw Pond) (Table S1).

156 To estimate predator density, we systematically dip-netted the focal ponds and
157 recorded the abundance and composition of two broad types of predators of spotted
158 salamander larvae: macro-invertebrates and adult amphibians (Table S5, below); none of

159 the focal ponds contained fish. We were unable to sample Beth's Pond for predators, and
160 instead used historical data collected by E.G.B. in 2013. We checked predator density
161 counts in 2013 against our 2016 sampling using Mincke Pond, the pond for which we had
162 data from both years. Predator densities in Mincke Pond in 2013 were similar to those that
163 we observed in 2016, so we considered our predator density estimates for Beth's Pond to
164 be representative for our sampling period.

165

166 *Trait measurements.* — On the sixth day after capture, we measured the length and mass of
167 salamanders, distinguishing between head length, body length, and tail length. We
168 photographed lateral and dorsal images of each larvae placed into clear tanks that
169 minimized movement (Fig. S1). We blot-dried individuals on paper towels before weighing.
170 We measured the length of salamander heads, bodies, and tails from images using ImageJ
171 (Fig. S1) (Rasband, 1997).

172 To obtain measurements of the shape of larvae we digitized landmarks on lateral
173 images using the software *tpsDig2* (Rohlf, 2006). Following Van Buskirk & Schmidt, (2000)
174 we tagged twenty landmarks that outlined larval shape (Fig. S1). Landmarks 1–3 described
175 the shape of the head of the larvae, landmarks 4–11 described body shape, and landmarks
176 9–20 outlined tail shape. Landmarks were rotated, scaled by size, and aligned to a
177 coordinated system using the Procrustes least-squares superimposition available in the
178 *geomorph* package for R statistical software (Adams and Otárola-Castillo 2013). We
179 conducted four principal component analyses to explore the scaled two-dimensional shape
180 variation, again distinguishing head shape, body shape, and tail shape. The first principal
181 component (PC) score accounted for most of the variation in head shape (37%), body

182 shape (33%), and tail shape (33%). We therefore used PC1 as the shape metric in our
183 analyses.

184

185 *Data Analyses.* — We calculated the coefficient of variation – the ratio of the standard
186 deviation to the mean – as a standardized measure of individual morphological variation
187 within ponds. We partitioned observed variation in salamander length and mass to within-
188 and between-pond trait differences with generalized linear mixed models (GLMM), using
189 the *lme4* package in R (R Core Team, 2019). Specifically, we calculated the intra-class
190 correlation coefficient (ICC) from GLMMs that included ‘pond’ as a random intercept term
191 (Nakagawa and Schielzeth, 2010). The ICC, also called the variance partitioning coefficient
192 (Messier et al., 2010), is the proportion of total variation in response variables that is
193 attributable to group-level (between-pond in our case) differences. In our analysis, the ICC
194 indicated the proportion of total observed variation in salamander length and mass that
195 came from between-pond differences in those traits. We ran separate GLMMs for mass and
196 the four length measurements – head length, body length, tail length, and total length. All
197 models used a Gaussian error structure, as all morphological data were normally
198 distributed (Fig. 3).

199 We then assessed within- and between-pond variation in mass-length allometry, a
200 form of trait co-variation. Specifically, we ran GLMMs to test whether the slopes and
201 intercepts of mass-length regressions differed across the six focal ponds, again
202 distinguishing head length, body length, and tail length. GLMMs included mass as the
203 response, length as a fixed effect, pond as a random intercept term, and length as a random
204 slope term. We log-transformed both length and mass and used a Gaussian error structure

205 for the normalized data in all models. To enable convergence of these more complex
206 models, we multiplied (log-transformed) mass by a factor of 10 to standardize the units
207 with length measurements. To test for differences in regression intercepts and slopes
208 across focal ponds, we used likelihood ratio tests comparing the fit of models that included
209 both terms with models omitting the random intercept or random slope term. We also
210 calculated the marginal and conditional R^2 of the models using the *MUMIn* package in R.
211 Marginal R^2 is a measure of the amount of variation in mass that was explained by the fixed
212 effect of length, while conditional R^2 considers variation explained by both fixed and
213 random effect terms (Johnson, 2014).

214 To further assess differences in the co-variation of morphological traits among focal
215 ponds, we determined the extent to which between-pond variation in salamander head,
216 body, tail, and overall (all segments combined) shape contributed to total observed
217 variation in these multidimensional morphological traits. Again, we calculated the ICC from
218 GLMMs, including 'pond' as a random intercept term. PC1 scores were used as response
219 variables. The shape data were also normally distributed (Fig. 3), so we used a Gaussian
220 error structure for all models.

221 To perform our preliminary test of the influence of pond age and predator density
222 on salamander mass, length, and shape, we ran GLMMs that included pond age (new vs.
223 old) and predator density as fixed effects, and the pond name as a random effect. To test
224 how the age and predator density of ponds influenced mass-length co-variation, we ran
225 GLMMs with log-transformed mass as the response and log-transformed length, the focal
226 factor (i.e., pond age or predator density) and their interaction as fixed effects. We also
227 included salamander length (again, log-transformed) as a random slope term, and pond

228 name as a random intercept term. We ran separate GLMMs for the two interactions to
229 prevent model overfitting. We also ran separate GLMMs for our different length measures:
230 head length, body length, tail length, and total length. We compared the fit of models
231 including the interaction terms with models omitting the interaction terms, using
232 likelihood ratio tests, to test the influence of pond age and predator density on the
233 relationship between mass and length.

234

235 **Results**

236 We measured a total of 519 spotted salamander larvae (Forest 44: N = 101, Shaw: N = 88,
237 Salamander pond: N = 65, Arthur Christ: N = 116, Beth's: N = 30, Mincke: N = 119) in this
238 study. Salamander mass was not influenced by their feeding on the previous day in the
239 subset of 227 individuals tested ($F_{(1,82)} = 3.43, p = 0.068$). Within all ponds, salamander
240 mass varied more among individuals than any of the length measurements (Fig. 2a).
241 Morphological variation was consistently lower in Beth's Pond (Fig. 2b), although this was
242 likely due to the lower sample size. Otherwise, there was no indication that specific ponds
243 had more or less morphological variation (Figs. 2b and 3).

244 Between-pond differences in average trait values accounted for 7 to 35% of the total
245 observed variation in salamander mass and length, depending on the specific body section
246 measured (Table 1). Specifically, between-pond differences accounted for proportionally
247 more of the observed variation in salamander mass (35%), total length (35%), and tail
248 length (27%) than in head (11%) and body length (17%) (Table 1).

249 Salamander mass strongly co-varied with total length, which makes sense because
250 we took single measures of mass that incorporated all body segments. Mass also strongly

251 co-varied with body length and tail length, but was less correlated with head length, likely
252 because heads are the smallest body segment of salamanders (Fig. S1). There were
253 detectable differences in the intercepts of mass-length relationships across ponds
254 (including pond as a random intercept term improved model fit; mass-head length: $X^2_1 =$
255 111.84, $p < 0.001$; mass-body length: $X^2_1 = 141.09$, $p < 0.001$; mass-tail length: $X^2_2 = 39.089$,
256 $p < 0.001$; total length: $X^2_1 = 81.136$, $p < 0.001$; Fig. 4). For a given length, individuals from
257 Salamander Pond tended to be heavier than individuals from other ponds (Fig. 4) whereas
258 individuals from Beth's Pond were generally lighter in mass per unit length (Fig. 4). The
259 slopes of mass-length relationships were less influenced by pond of capture (Table 2). Only
260 the slopes for mass-head length and mass-tail length relationships were influenced by the
261 pond of capture (mass-head length: $X^2_2 = 8.68$, $p = 0.013$; mass-tail length: $X^2_2 = 11.25$, $p =$
262 0.004; Fig. 4); the slopes of mass-body length and mass-total length relationships were
263 consistent across sampled ponds (mass-body length: $X^2_2 = 4.24$, $p = 0.120$; mass-total length:
264 $X^2_2 = 5.05$, $p = 0.080$; Fig. 3). The slope exponents were always < 3 (Table S3), indicating
265 that larger salamander larvae generally had more elongate heads, bodies, and tails than
266 smaller larvae.

267 Similar to length measures, the shape of salamander tails exhibited more between-
268 pond variation than did the heads or bodies. Between-pond trait differences contributed
269 25% of the total observed variation in tail shape, compared with 11%, 9%, and 7% of head,
270 body, and overall shape, respectively (Table 1). Furthermore, there was little evidence that
271 PC scores were clustered by pond (Fig. 5), indicating a weak signal of between-pond
272 variation in salamander shape.

273 Neither pond age nor predator density influenced salamander mass or any
274 measures of length (Table S4). However, these pond attributes did influence certain mass-
275 length relationships and body shapes (Fig. S4, Table S5). Pond age influenced the scaling of
276 mass with head and tail length (Fig. S4, Table S5). In contrast, predator density influenced
277 mass-body length relationships (Fig S2, Table S2). Both pond age and predator density
278 influenced salamander head shape, but neither influenced body or tail shape. Pond age also
279 influenced the overall shape of salamanders, but predator density did not (Table S4).

280

281 **Discussion**

282 Measuring the lengths and masses of salamander larvae in a network of spatially discrete
283 ponds showed that most morphological variation and co-variation occurred within ponds.
284 Between-pond morphological differences were not negligible however, particularly for
285 salamander tails. Salamander tails exhibited more between-pond variation than heads or
286 bodies. Between-pond differences in trait co-variation were also evident in salamander
287 tails. Scaling of mass with salamander tail lengths differed across ponds both in terms of
288 the intercepts and the slopes of the relationships, and between-pond differences
289 contributed more to total observed variation in tail shape than for head or body shape. The
290 spatial discreteness of pond habitats therefore seems to act strongly on salamander tail
291 morphology (see below for further discussion).

292 The substantial within-pond variation in salamander morphology suggests that local
293 factors, such as microhabitat heterogeneity, influence salamander morphology. Given that
294 many ponds were spaced within the documented dispersal ranges of salamanders
295 (Zamudio and Wiczorek, 2006; Patrick et al., 2008), movement between ponds could also

296 have reduced the contribution of between-pond differences to morphological variation by
297 sustaining mixing of genotypes and phenotypes. At the metapopulation scale, local and
298 spatial factors likely interact to shape varying degrees of between-pond morphological
299 variation similar to what we observed in salamanders.

300 The stronger contribution of between-pond differences to salamander tail variation
301 may be because tails play an important role in locomotive (i.e., swimming) performance.
302 Being meso-predators, swimming performance for salamanders is critical to both capturing
303 prey and evading predators (Van Buskirk and Schmidt, 2000; Urban, 2010; Landberg and
304 Azizi, 2010). Tails may therefore be more closely linked to fitness, hence under stronger
305 selection, than heads and bodies, at least in habitats where predation is a significant threat
306 (Landberg and Azizi, 2010). Multiple predators of larval salamanders were found in our
307 focal ponds, so predation risk is likely to be a strong selective force in the salamander
308 metapopulation studied here. In a preliminary analysis of the data, we did not detect an
309 influence of predator density on tail length of salamanders (see Supplementary Material),
310 but this analysis was based on our limited sample of ponds and warrants further
311 investigation. Regardless of the factors driving salamander tail variation, our findings
312 suggest that salamander responses to habitat alterations, biological invasions, and other
313 pond-level disturbances may manifest as changes in tail morphology as opposed to changes
314 in head and body morphology. If this prediction holds, the more common body size
315 measure for amphibians, snout-to-vent length, which only takes head and body length into
316 account, would be insufficient for predicting the eco-evolutionary responses of this species
317 to landscape-scale environmental changes in aquatic habitats.

318 Between-pond variation in the allometric relationship between salamander mass

319 and total length arose specifically from differences in intercepts; slopes of the relationship
320 were highly consistent. Pond-level effects appear to act on the relationship of salamander
321 mass to total body length, but they do not appear to alter how mass scales with total body
322 length. This spatially robust scaling of mass and total body length may explain why the two
323 traits separately exhibited nearly identical degrees of between-pond variation. More
324 broadly, this pattern of allometric scaling, in which intercepts but not slopes of trait
325 relationships differ, is consistent with allometric relationships documented across many
326 other taxa (Voje et al., 2014), suggesting a general constraint to the plasticity and evolution
327 of the slopes of trait relationships.

328 One important caveat to the above findings is that our sampling could not
329 distinguish between-pond variation in salamander morphology from possible temporal
330 variation in salamander morphology. Spotted salamander larvae exhibit growth and
331 developmental changes within summer months that could have contributed to observed
332 morphological variation. We expect the contributions of temporal changes to salamander
333 morphological variation to have been minor because we sampled salamander larvae during
334 latter developmental stages, evidenced by all salamanders being in the final Harrison
335 stages (45-46) (Harrison, 1969) after most growth and development had occurred
336 (Landberg and Azizi, 2010). In support of this expectation, body lengths and mass did not
337 increase monotonically throughout the sampling period, which should be the case when
338 growth and development drive morphological variation. Nevertheless, we cannot rule out
339 the possible influence of temporal morphological variation and advocate for further work
340 that corroborates our findings through longitudinal pond sampling, which would account
341 for growth and ontogeny.

342 The inclusion of morphological diversity data in biodiversity conservation stems
343 from the idea that different populations of the same species are not equal in terms of eco-
344 evolutionary history. As such, exploring various approaches to the conservation of
345 morphological diversity is important to developing strategies for reducing biodiversity
346 losses under global change (Des Roches et al., 2018). The mix of within- and between-pond
347 morphological variation in salamanders provides promise that pond construction can
348 utilize local and spatial processes to bolster morphological diversity. Capitalizing on the
349 presence of new constructed ponds in our study area, we made a preliminary comparison
350 of salamander morphology and allometry between new and old ponds (see Supplementary
351 Material for detailed methods and results). Although our analysis did not detect differences
352 in mass or length of salamanders between new and old ponds, we did find differences in
353 mass-length relationships and body shape (Supplementary Material Table S4). Habitat
354 restoration through pond construction may therefore bolster diversity in trait co-variation,
355 and in so doing may instill more adaptive potential under environmental change (Laughlin
356 and Messier, 2015). Although there are several studies for various taxa that quantify
357 functional connectivity between habitat patches (and local populations) using genetic
358 techniques, we encourage additional studies on morphological parameters and patterns to
359 better understand the mechanisms that promote long-term population persistence in
360 fragmented landscapes.

361

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370

371 **Conflicts of Interest**

372 We declare no conflicts of interest

373

374 **Author Contributions**

375 DRD and ETG designed the study and executed field sampling and trait measurements, with
376 supervisory support from AID and JAC. ETG completed the morphometrics and principal
377 components analyses. DRD performed the rest of the statistical analyses. All authors
378 contributed to the writing and revision of the manuscript.

379

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- 465

466 **Tables and Figures**

body segment	between-pond variance	SD	residual variance	SD	ICC (%)	95% CI - lower	95% CI - upper
mass							
	0.013	0.112	0.023	0.152	35.3	8.62	58.71
length							
head	0.15	0.39	1.27	1.13	10.8	0.73	26.11
body	0.92	0.96	4.36	2.09	17.4	8.72	53.62
tail	4.27	2.07	11.29	3.36	27.5	4.08	52.07
combined	18.23	4.27	32.91	5.74	35.6	6.25	57.80
shape							
	between-pond variance	SD	residual variance	SD	ICC	95% CI - lower	95% CI - upper
head	0.0004	0.019	0.003	0.053	11.0	0.7	26.5
body	0.0002	0.016	0.002	0.049	9.4	0.3	23.6
tail	0.001	0.026	0.002	0.049	22.8	1.2	35.0
combined	0.000	0.009	0.001	0.033	7.60	0.2	17.7

467

468 **Table 1. Proportion of mass, length, and shape variation attributable to between-**
 469 **pond differences.** Displayed are the variance components of generalized linear mixed
 470 models used for our analyses of within- and between-pond variation in salamander mass,
 471 length, and shape. The intra-class correlation coefficient (ICC), or the variance partitioning
 472 component, is the proportion of between-pond variation explaining total observed trait
 473 variation (between-pond variance + residual variance). Higher ICC values denote higher
 474 degree of between-pond variation in salamander traits. CI = confidence intervals of the ICC
 475 derived from bootstrapping over 500 resampling events.

476

mass-length co-variation								
body segment	intercept variance	SD	slope variance	SD	residual variance	SD	marginal R ²	conditional R ²
head	0.11	0.33	0.09	0.29	0.02	0.13	0.26	0.49
body	0.04	0.20	0.02	0.13	0.01	0.09	0.65	0.77
tail	0.08	0.28	0.04	0.19	0.01	0.08	0.73	0.79
combined	0.07	0.26	0.00	0.07	0.00	0.07	0.81	0.87

477

478 **Table 2. Variation in intercepts and slopes of mass-length relationships.** Displayed

479 are the variance components of generalized linear mixed models used for our analyses of

480 between-pond variation in the scaling of salamander mass with different length

481 measurements. Models included length (log-transformed) as a fixed effect and a random

482 slope term, and pond as a random intercept term. Marginal R² denotes the amount of

483 variation in mass explained by the length measurement alone, whereas conditional R²

484 considers variation explained by the random intercept and slope terms.

485

486

487 **Fig. 1. Map of study area.** The six focal ponds were located in Eastern Missouri (US across
488 three conservation areas. Mincke Pond, Arthur Christ Pond, and Beth's Pond are located in
489 Tyson Research Center. Shaw Pond is located in the Shaw Nature Reserve. Forest 44 Pond
490 is located in Forest 44 Conservation Area. All ponds occurred in Oak-Hickory forests typical
491 of the region.

492

493 **Fig. 2. Within-pond variation in salamander morphology.** The coefficient of variation,
494 or the extent of variance in relation to mean trait values, is shown for **(a)** mass and length
495 measures and **(b)** the six ponds where we sampled salamanders, ordered from left to right
496 in the chronological order in which they were sampled. Colors distinguish pond of capture
497 in (a) and the focal trait in (b). Note that the same data are reported in (a) and (b).

498

499 **Fig. 3. Salamander mass and length variation within and between ponds.** Panels on
500 left **(a,c,e,g,i)** are frequency distributions of mass and length measurements across all focal
501 ponds. Violin plots on the right-side panels **(b,d,f,h,j)** distinguish individual-level and
502 between-pond morphological variation to show how the traits were spatially structured.
503 Box plots within the violin plots denote the mean, standard error, and 95% confidence
504 intervals of trait measures. Ponds in the right-side panels are ordered from left to right in
505 the chronological order in which they were sampled.

506

507

508 **Fig. 4. Between-pond variation in mass-length allometry in salamanders.** Regression
509 lines of salamander mass with (a) head length, (b) body length, (c) tail length, and (d) are
510 shown for the six focal ponds, distinguished by line colors. Shaded areas show the 95%
511 confidence intervals of the regression lines. Mass and length are plotted on a \log_{10} scale in
512 all cases.

513

514 **Fig. 5. Salamander shape variation within and among ponds.** The shape values are
515 based on sets of landmarks at different points along the lateral surface of salamander
516 bodies (a). The overall (b), head (c), body (d), and tail (e) shape of salamanders collected
517 from the six focal ponds. The shape values are based on sets of landmarks at different
518 points along the lateral surface of salamander bodies. PC1 and PC2 values increase with
519 elongation of shape and increasing length:height ratio.

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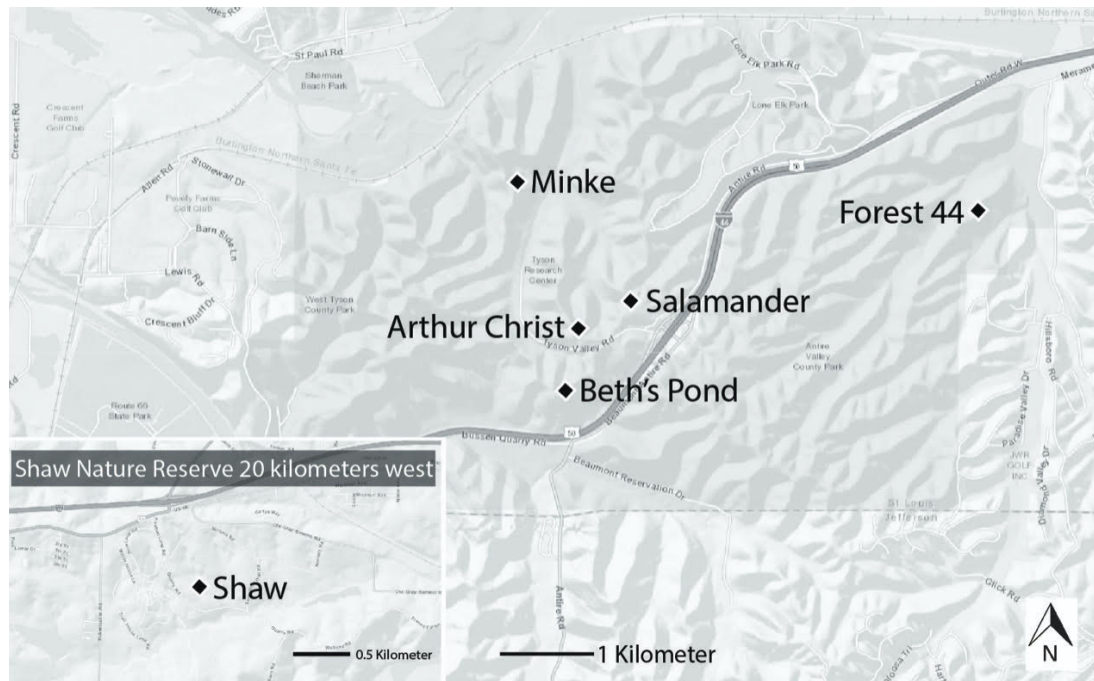
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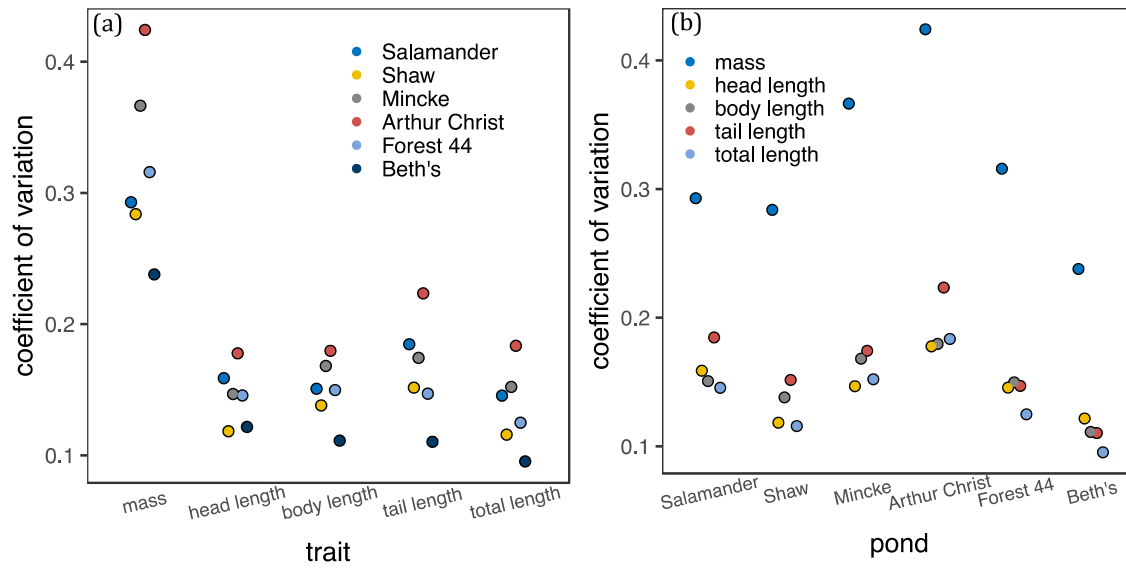
530 Fig. 1



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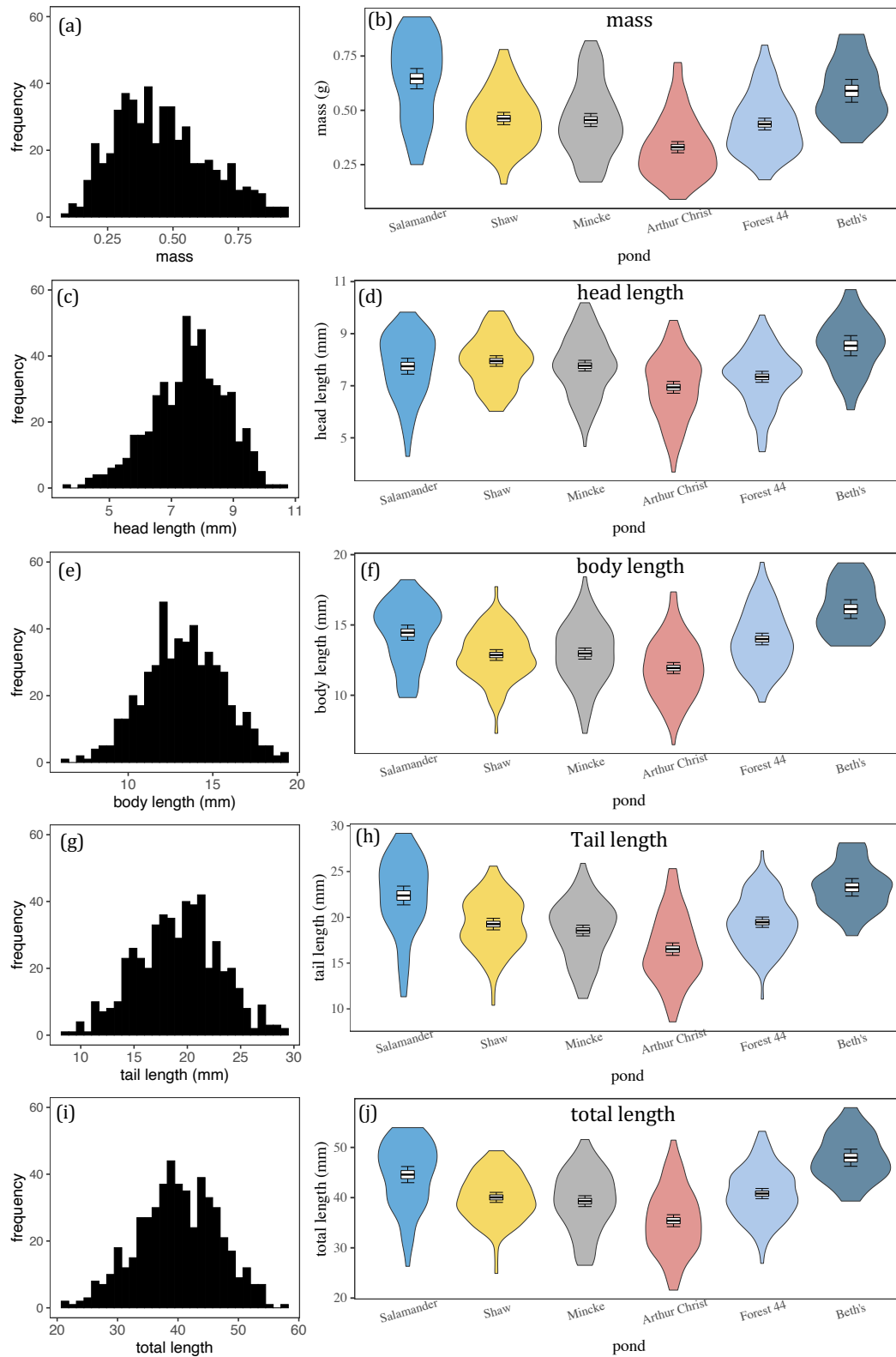
533 Fig. 2



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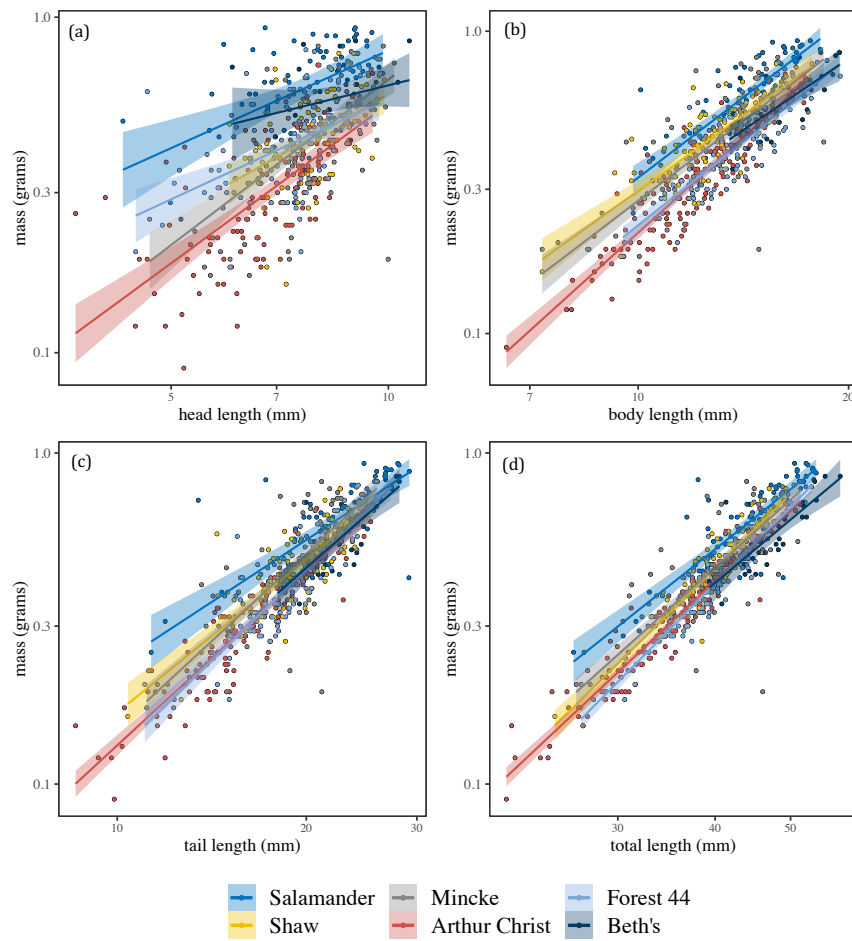
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536 Fig. 3

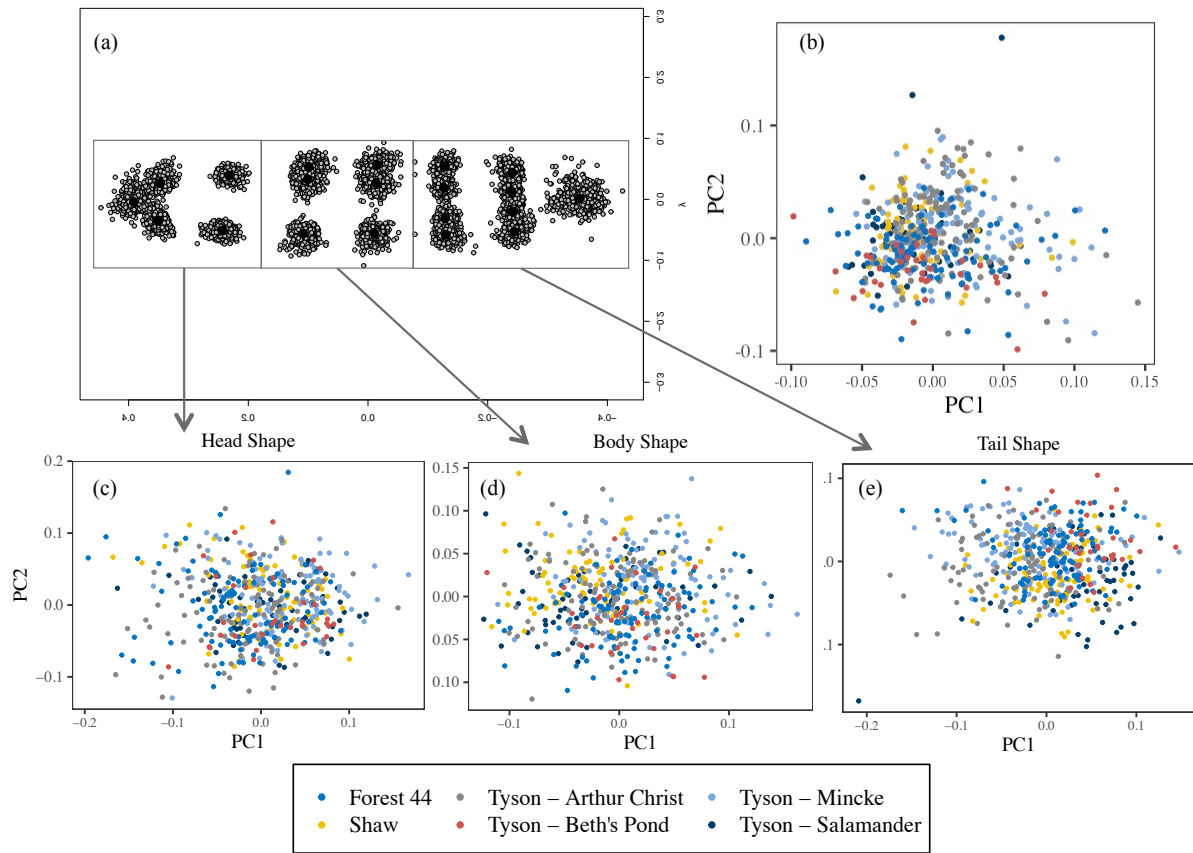


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538 Fig. 4



549 Fig. 5



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