Analysis of overlapping genetic association in type 1 and type 2 diabetes

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Abstract

Aims/hypothesis: Given the potential shared aetiology between type 1 and type 2 diabetes, we aimed to identify any genetic regions associated with both diseases. For associations where there is a shared signal and the allele that increases risk to one disease also increases risk to the other, inference about shared aetiology could be made, with the potential to develop therapeutic strategies to treat or prevent both diseases simultaneously. Alternatively, if a genetic signal colocalises with divergent effect directions, it could provide valuable biological insight into how the association affects the two diseases differently.

Methods: Using publicly available type 2 diabetes summary statistics from a genomewide association study (GWAS) meta-analysis of European ancestry individuals (74,124 cases and 824,006 controls) and type 1 diabetes GWAS summary statistics from a meta-analysis of studies on individuals from the UK and Sardinia (7,467 cases and 10,218 controls), we identified all regions of 0.5 Mb that contained variants associated with both diseases (false discovery rate<0.01). In each region, we performed forward stepwise logistic regression to identify independent association signals, then examined colocalisation of each type 1 diabetes signal with each type 2 diabetes signal using *coloc*. Any association with a colocalisation posterior probability of ≥0.9 was considered a genuine shared association with both diseases.

Results: Of the 81 association signals from 42 genetic regions that showed association with both type 1 and type 2 diabetes, four association signals colocalised between both diseases (posterior probability ≥0.9): (i) chromosome 16q23.1, near Chymotripsinogen B1 (*CTRB1*) / Breast Cancer Anti-Estrogen

Resistance Protein 1 (*BCAR1*), which has been previously identified; (ii) chromosome 11p15.5, near the Insulin (INS) gene; (iii) chromosome 4p16.3, near Transmembrane protein 129 (*TMEM129*), and (iv) chromosome 1p31.3, near Phosphoglucomutase 1 (*PGM1*). In each of these regions, the effect of genetic variants on type 1 diabetes was in the opposite direction to the effect on type 2 diabetes. Use of additional datasets also supported the previously identified colocalisation on chromosome 9p24.2, near the GLIS Family Zinc Finger Protein 3 (*GLIS3*) gene, in this case with a concordant direction of effect.

Conclusions/interpretation: That four of five association signals that colocalise between type 1 diabetes and type 2 diabetes are in opposite directions suggests a complex genetic relationship between the two diseases.

Research in Context

What is already known about this subject?

- Other than insulin, there are currently no treatments for both type 1 and type 2 diabetes.
- Findings that genetic variants near the GLIS3 gene increase risk of both type 1 and type 2 diabetes have indicated shared genetic mechanisms at the level of the pancreatic β cell.

What is the key question?

- By examining chromosome regions associated with both diseases, are there any more variants that affect risk of both diseases and could support common mechanisms and repositioning of therapeutics between the diseases?

What are the new findings?

- At current sample sizes, there is evidence that five genetic variants in different chromosome regions impact risk of developing both diseases.
- However, four of these variants have the opposite direction of effect in type 1 diabetes compared to type 2 diabetes, with only one, near *GLIS3*, having a concordant direction of effect.

How might this impact on clinical practise in the foreseeable future?

- Genetic findings have furthered research in type 1 and type 2 diabetes independently, and suggest therapeutic strategies. However, our current investigation into their shared genetics suggests that repositioning of current type 2 diabetes treatments into type 1 diabetes may not be straightforward.

86 **Introduction**

87 There is a genetic component to both type 1 and type 2 diabetes, with
88 approximately 60 chromosome regions associated with type 1 diabetes [1] and
89 over 200 associated with type 2 diabetes [2] at genomewide significance.
90 Examination of regions associated with both diseases could lead to uncovering
91 signals that simultaneously alter disease risk for both diseases, termed
92 colocalisation. Uncovering colocalising signals could provide biological insights
93 into shared disease mechanisms, and potentially reveal therapeutic targets
94 effective for both diseases. A recent analysis suggested that the same genetic
95 variant is altering risk of both type 1 and type 2 diabetes in five regions, near
96 Centromere Protein W (*CENPW*), Chymotripsinogen B1 (*CTRB1*)/Breast Cancer
97 Anti-Estrogen Resistance Protein 1 (*BCAR1*), GLIS Family Zinc Finger Protein 3
98 (*GLIS3*), B-cell Lymphoma 11A (*BCL11A*) and Thyroid Adenoma-Associated
99 Protein (*THADA*) [3].
100 Here, we identified all regions across the genome that showed evidence of
101 association to both type 1 and type 2 diseases at false discovery rate (FDR)
102 <0.01, and assessed colocalisation between the two diseases in each of these
103 regions. Furthermore, to account for the possibility of multiple causal variants
104 within an associated region, we extended the analysis to investigate
105 conditionally-independent associations within each region, to assess whether
106 any of the associations with one disease colocalised with any associations in the
107 other.

108 **Methods**

109 Type 1 diabetes meta-analysis summary statistics were generated using genome
110 wide association study (GWAS) data from 3,983 cases and 3,994 controls from
111 the UK genotyped using the Illumina Infinium 550K platform, 1,926 cases and
112 3,342 controls from the UK genotyped using the Affymetrix GeneChip 500K
113 platform and 1,558 cases and 2,882 controls from Sardinia genotyped using the
114 Affymetrix 6.0 and Illumina Omni Express platforms, totalling 7,467 cases and
115 10,218 controls (**ESM Table 1**). Genotypes were imputed using the Haplotype
116 Reference Consortium (HRC) reference panel for the UK collections [4], and a
117 custom Sardinian reference panel of 3,514 Sardinians for the Sardinian collection
118 (**ESM - Imputation)**.
119 Summary statistics for type 2 diabetes were from 74,124 cases and 824,006
120 controls of European ancestry, imputed using the HRC reference panel [2].
121 Regions associated with both diseases were identified by selecting all variants
122 with type 1 diabetes and a type 2 diabetes association with FDR<0.01 (**ESM –**
123 **Type 1 diabetes GWAS**). In each such region, windows of approximately 0.5 Mb
124 were taken to examine colocalisation (**ESM - Regions associated with both**
125 **diseases***)*. Within these regions, forward stepwise logistic regressions were
126 carried out for both diseases, and conditional summary statistics were obtained
127 so each conditionally-independent signal from both diseases could be tested
128 against each other for colocalisation (**ESM - Conditional analyses**).
129 Colocalisation of signals was assessed using *coloc* [5], a Bayesian method that
130 enumerates the posterior probability that the association signals in a region are
131 shared between traits. The prior probability of association with either disease
132 was taken to be 1×10^{-4} and the prior probability that the association signal is

133 shared across traits was taken to be 5×10^{-6}, as recommended [6]. The threshold to

134 consider signals as colocalising was conservatively chosen at a posterior

135 probability ≥0.9. Colocalisation was also examined using an alternative

136 approach, as a secondary analysis, eCAVIAR [7] (**ESM – eCAVIAR**).

137 **Results**

138 Including conditionally-independent association signals, 81 colocalisation
139 analyses were carried out across 42 chromosomal regions that showed
140 association to both diseases (**ESM Table 2**).
141 Four signals showed evidence of colocalisation using *coloc*, and they were also
142 the regions with the highest *eCAVIAR* regional colocalisation posterior
143 probabilities (**ESM Table 3**). The first was on chromosome 16q23.1, near *CTRB1*
144 and *BCAR1*, with a posterior probability of colocalisation (H4PP, hereafter) of
145 0.98 (**ESM Figure 1**). The minor A allele at the type 2 diabetes index variant,
146 rs72802342 (C>A), is protective for type 2 diabetes (OR=0.87, $p=4.00\times10^{-32}$) and
147 susceptible for type 1 diabetes (OR=1.33, $p=5.81\times10^{-10}$).
148 The second was on chromosome 11p15.5, near *INS*, where the primary type 2
149 diabetes association colocalised with the secondary type 1 diabetes association
150 (H4PP=0.95, **ESM Figure 2**). The direction of effect was opposite, with the minor
151 A allele at the type 2 diabetes index variant, rs4929965 (G>A), associated with
152 susceptibility to type 2 diabetes (OR=1.07, $p=4.80\times10^{-25}$) and protection from
153 type 1 diabetes (OR=0.87, $p=1.89\times10^{-5}$).
154 Thirdly, a region on chromosome 4p16.3 colocalised (H4PP=0.97) (**Figure 1**),
155 near Transmembrane protein 129 (*TMEM129*). The minor T allele at the type 2
156 diabetes index variant, rs56337234 (C>T), was associated with decreased risk of
157 type 2 diabetes (OR=0.94, $p=1.4\times10^{-17}$) and increased risk of type 1 diabetes
158 (OR=1.12, $p=4.07\times10^{-6}$).
159 Finally, a region on chromosome 1p31.3, near Phosphoglucomutase 1 (*PGM1*),
160 colocalised (H4PP=0.91, **ESM Figure 3**), with the minor T allele at the type 2
161 diabetes index variant rs2269247 (C>T) decreasing risk of type 2 diabetes

162 (OR=0.96, $p=4.6\times10^{-7}$) and increasing risk of type 1 diabetes (OR=1.15,
163 $p=1.9\times10^{-6}$) (**Table 1**).
164 We did not replicate the finding that the chromosome regions near *CENPW,*
165 *GLIS3*, *BCL11A or THADA* colocalised between type 1 and type 2 diabetes (H4PP
166 *CENPW*=0.12, *GLIS3*=0.29, *BCL11A*=0.28, *THADA* not examined as no type 1
167 diabetes association existed in the region (FDR=0.07)). To investigate these
168 discrepancies, we examined two other large type 2 diabetes meta-analyses: a
169 trans-ethnic study including 1,407,282 individuals [8] and a study of 433,540
170 individuals of East Asian ancestry [9]. For the *CENPW* and *BCL11A* regions, the type
171 2 diabetes signal is consistent with at least one of the other GWAS studies
172 (measured by linkage disequilibrium (LD) in Europeans to the other study index
173 variants, **ESM Table 4**); and, the type 1 diabetes index variant is not in strong LD
174 ($r^2<0.41$) with any of the index variants for type 2 diabetes across the three
175 GWAS studies. However, at *GLIS3*, there appears to be a distinct signal in the
176 European study [2] compared to the trans-ethnic and East Asian type 2 diabetes
177 studies ($r^2=0.65$), and the index variants from these two studies are in higher r^2
178 with the type 1 diabetes signal in our analysis ($r^2=0.68$), and even higher r^2 with
179 the index variant from a larger T1D genetic analysis [1] ($r^2=0.99$), indicating that
180 the signal near *GLIS3* does colocalise between type 1 and type 2 diabetes with
181 concordant direction of effect, as previously identified [10].

182 **Discussion**

183 Using genetic association summary statistics from European populations, we
184 identified 42 regions that showed association with both type 1 and type 2
185 diabetes, with 81 conditionally-independent association signals across those
186 regions. Four signals (near *CTRB1/BCAR1*, *INS*, *TMEM129* and *PGM1*) colocalised
187 between the diseases, including a signal at the complex *INS* region for the first
188 time, which was achieved by examining conditional summary statistics.
189 However, in all four cases, the allele increasing risk for one disease was
190 protective against the other. Examination of additional trans-ethnic and East
191 Asian type 2 diabetes genetic analyses, indicated that a fifth association, near
192 *GLIS3*, is likely to colocalise between diseases, with concordant direction of
193 effect.
194 Given the distinct mechanisms underlying β-cell dysfunction and cell death
195 between the two diseases [11], it is perhaps unsurprising that no additional signals
196 were detected with concordant direction of effect. However, the type 1 diabetes
197 GWAS was much smaller than the type 2 diabetes analysis, and therefore had
198 less statistical power to detect more subtle genetic effects. If a type 1 diabetes
199 GWAS were to be performed with similar power to the type 2 diabetes GWAS,
200 more regions might colocalise between the two diseases, but either the effects of
201 these additional regions on type 1 diabetes would be small compared to the
202 currently known associations, or they would be rare variants with larger effect
203 sizes.
204 That four of five colocalisation signals had opposite directions of effect implies a
205 complex genetic relationship between the two diseases. Whilst the directional
206 discordance offers little hope for effective treatments for both diseases

207 simultaneously at these particular targets, it can offer biological insight into the
208 disease pathways that these regions act upon, and even if there is directional
209 discordance, the genetics could be highlighting the same therapeutic target.
210 We did not replicate the findings that the associations near *BCL11A*, *CENPW* and
211 *THADA* colocalise between the two diseases [3], despite overlapping samples and
212 similar numbers of cases and controls in the type 1 diabetes GWAS. This is likely
213 due to three reasons: i) the previous study [3] examined colocalisation using
214 weaker association signals, for example, the colocalisation near *THADA* was
215 based on a type 1 diabetes association p-value of 0.01; ii) we used a more
216 stringent prior for colocalisation between the two diseases, as recently
217 suggested [6] (5×10^{-6} vs. 1×10^{-5}); and iii) we used a more stringent posterior
218 probability threshold to declare colocalisation (0.9 vs. 0.5). Our increased
219 stringency compared to the previous analysis [3], whilst increasing the probability
220 that any identified shared signals will be true positives, may have decreased our
221 sensitivity to detect all colocalisations. For example, by examining other large
222 type 2 diabetes GWAS analyses and a larger type 1 diabetes genetic analysis, we
223 conclude that the association near *GLIS3* likely does colocalise between the two
224 diseases, and with concordant directions of effect.
225 In conclusion, with current GWAS sample sizes, just five associations appear to
226 colocalise between type 1 diabetes and type 2 diabetes, four with opposing
227 direction of effect. Larger sample sizes would be required to identify the depth of
228 genetically identified therapeutic targets to treat or prevent both diseases
229 simultaneously.

230 ## Tables and Figures

231 **Table 1**: Regions with a colocalisation posterior probability of ≥0.9 between type 1 diabetes and type 2 diabetes. Summary statistics

232 given from the perspective of the index type 2 diabetes variant and with respect to the ALT allele. T2D= type 2 diabetes, T1D= type 1

233 diabetes. r^2 obtained from 1000 Genomes Project European population.

rsID	Proximal gene(s)	chr	pos (gr37)	REF	ALT	T2D conditional on	T2D OR (95% CI)	T2D p	r^2 to T1D index variant (T1D index variant)	T1D conditional on	T1D OR (95% CI)	T1D p
rs2269247	*PGM1*	1p31.3	64107284	C	T	-	0.96 (0.94, 0.97)	4.60×10^{-7}	0.86 (rs2269246)	-	1.14 (1.08, 1.22)	1.94×10^{-6}
rs56337234	*TMEM129*	4p16.3	1784403	C	T	-	0.94 (0.93, 0.96)	1.40×10^{-17}	0.97 (rs6829631)	-	1.12 (1.07, 1.18)	4.07×10^{-6}
rs4929965	*INS*	11p15.5	2197286	G	A	rs11042596, rs555759341, rs571342427, rs10838787	1.07 (1.06, 1.09)	4.80×10^{-25}	0.97 (rs7119275)	rs689	0.87 (0.81, 0.93)	1.89×10^{-5}
rs72802342	*CTRB1/ BCAR1*	16q23.1	75234872	C	A	rs3115960	0.87 (0.85, 0.89)	4.00×10^{-32}	0.89 (rs55993634)	-	1.33 (1.22, 1.46)	5.81×10^{-10}

234 **Figure Legends**

235 **Figure 1:** Manhattan plots showing a) gene locations and $-\log_{10}$p-value of

236 association for each variant by position along chromosome 4 (genome build 37)

237 in the *TMEM129* region for b) type 2 diabetes and c) type 1 diabetes, coloured by

238 r^2 to the type 2 diabetes index variant, rs56337234.

239

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243 **Data availability**

244 Type 1 diabetes summary statistics will be available through GWAS catalog

245 (https://www.ebi.ac.uk/gwas/). Type 2 diabetes summary statistics are already

246 publicly available.

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Authors' relationships and activities

Mark McCarthy has served on advisory panels for Pfizer, Novo Nordisk and Zoe Global, has received honoraria from Merck, Pfizer, Novo Nordisk and Eli Lilly, and research funding from Abbvie, Astra Zeneca, Boehringer Ingelheim, Eli Lilly, Janssen, Merck, Novo Nordisk, Pfizer, Roche, Sanofi Aventis, Servier, and Takeda. As of June 2019, Mark McCarthy is an employee of Genentech, and a holder of Roche stock. Anubha Mahajan is an employee of Genentech since January 2020, and a holder of Roche stock.

Jamie Inshaw is an employee of Exploristics since June 2020.

272 John Todd serves on the advisory board of GSK.

Contribution statement

JRJI carried out the type 1 diabetes meta-analysis and the colocalisation analyses, drafted the manuscript and approved the final version.

AM carried out the type 2 diabetes meta-analysis and conditional analyses, revised the article for intellectual content and approved the final version.

CS and FC were involved in data collection in the Sardinia collection and carried out the association testing in this collection, revised the article for intellectual content and approved the final version.

DC provided statistical advice and input, and made contributions to interpretation of the data, revised the article for intellectual content and approved the final version. .

MIS provided biological insight, contributed towards interpretation of the data, revised the article for intellectual content and approved the final version.

MM and JAT oversaw the research, contributed towards the conception, design and data collection, revised the article for intellectual content and approved the final version.

JAT is the guarantor of this work.

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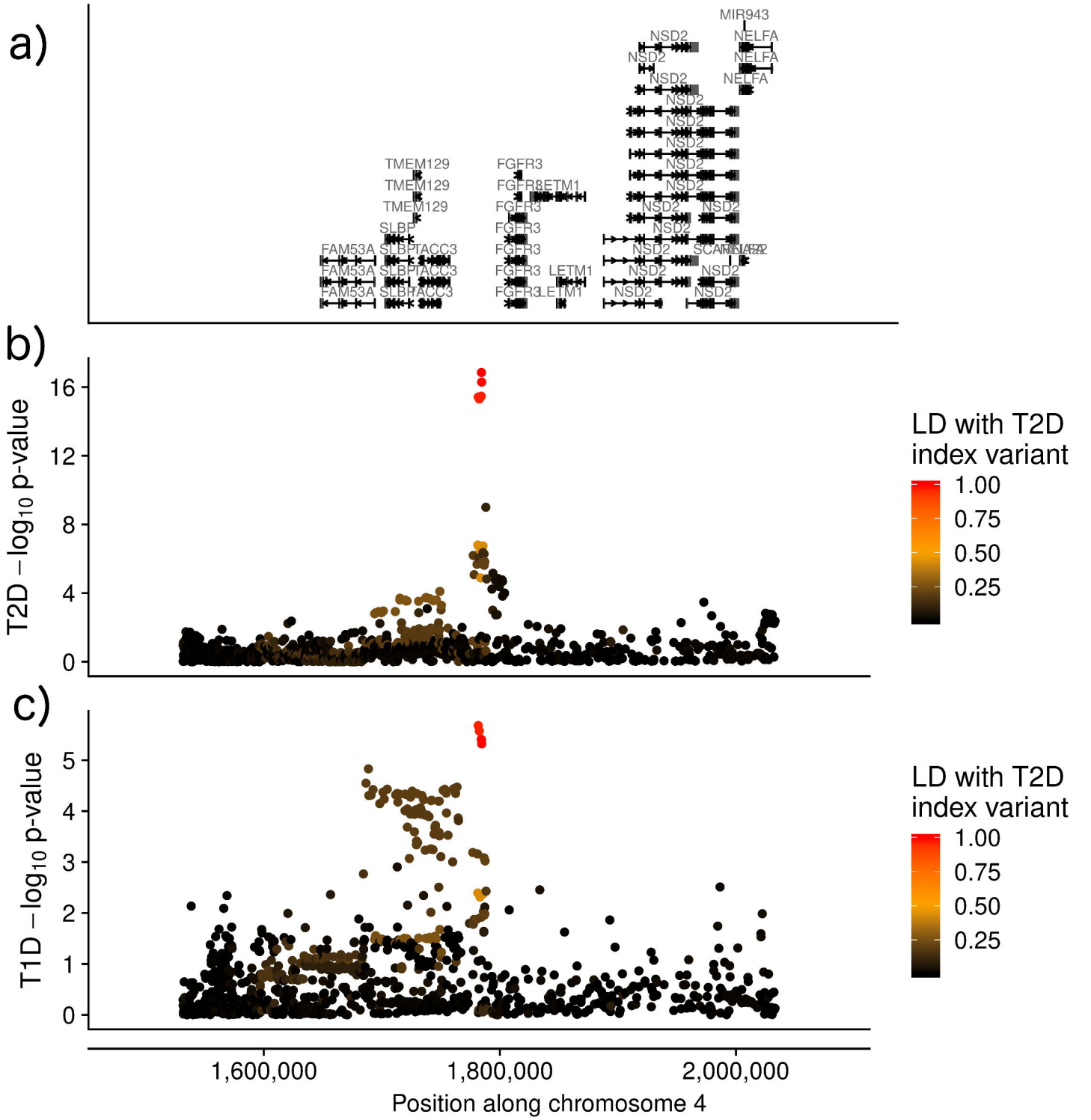
a)
MIR943
NSD2
NELFA
TMEM129
FGFR3
FGFR8
LETM1
SLBP
FAM53A
TACC3
b)
T2D −log10 p-value
16
12
8
4
0
LD with T2D index variant
1.00
0.75
0.50
0.25
c)
T1D −log10 p-value
5
4
3
2
1
0
LD with T2D index variant
1.00
0.75
0.50
0.25
1,600,000
1,800,000
2,000,000
Position along chromosome 4