The effect of hybridization on transposable element accumulation in

an undomesticated fungal species

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Abstract

Transposable elements (TEs) are mobile genetic elements that can profoundly impact the evolution of genomes and species. A long-standing hypothesis suggests that hybridization could deregulate TEs and trigger their accumulation, although it received mixed support from studies in plants and animals. Here, we tested this hypothesis in fungi using incipient species of the undomesticated yeast *Saccharomyces paradoxus*. Population genomic data revealed no signature of higher transposition in natural hybrids. As we could not rule out the elimination of past transposition increase signatures by natural selection, we performed a laboratory evolution experiment on a panel of artificial hybrids to measure TE accumulation in the near absence of selection. Changes in TE copy numbers were not predicted by the level of evolutionary divergence between the parents of a hybrid genotype. Rather, they were highly dependent on the individual hybrid genotypes, showing that strong genotype-specific deterministic factors govern TE accumulation in yeast hybrids.

Introduction

Hybridization is increasingly recognized as an important component of species evolution (Mallet, 2005; Mallet et al., 2016; Stukenbrock, 2016). Abundant empirical evidence shows that hybridization frequently leads to transgressive phenotypes in plants, animals and fungi (Langdon et al., 2019; Rieseberg et al., 1999). Mutation rates are traits for which transgressive segregation can have a major impact on the evolutionary fate of hybrids, with potential implications for the emergence of reproductive barriers and speciation. Insertions caused by transposable elements (TEs) are a class of mutations that could be particularly responsive to hybridization. TEs are diverse and ubiquitous DNA sequences that can duplicate and spread within genomes, forming families of dispersed repeated sequences (Bourque et al., 2018). They can either replicate through "copy-and-paste" (retrotransposons) or "cut-and-paste" (DNA transposons) mechanisms. TEs were first identified in cytological studies of chromosome instability in *Zea mays* (McClintock, 1950). In her seminal 1984 perspective, McClintock summarized evidence suggesting that TEs respond to various types of "genome shocks", one of which she suspected was hybridization (McClintock, 1984). Since McClintock's early insights, many studies investigated the association between TEs and hybridization across taxonomic groups and documented increased TE activity in hybrids. We refer to this increase as TE reactivation, but our use of this term implies no mechanistic basis and other terms like derepression are used in the literature. Reactivation was evidenced by increased TE abundance, novel TE insertions, TE epigenetic changes or higher TE transcript levels (Dion-Côté et al., 2014; Kidwell et al., 1977; Labrador et al., 1999; Liu and Wendel, 2000; O'Neill et al., 1998; Picard, 1976; Ungerer et al., 2006). However, hybrid reactivation is not universal, as many studies across a similar taxonomic range found no evidence for it (Coyne, 1989; Göbel et al., 2018; Hey, 1988; Kawakami et al., 2011). In fungi, it was recently shown that no TE

reactivation occurs in experimental interspecific hybrids between the yeasts *Saccharomyces*
cerevisiae and *Saccharomyces uvarum* (Smukowski Heil et al., 2020). Thus, TE reactivation
exhibits no clear taxonomic distribution, and the conditions favorable for its manifestation are
still unclear.

Studies of hybrids in controlled crosses revealed genetic bases to TE reactivation, but less
focus was put on cases of natural hybrids (but see (Kawakami et al., 2011; Ungerer et al.,
2009)). In addition to the genetic structure of populations, environmental pressures and
demographic factors can influence the action of evolutionary forces on TEs, thus impacting the
genomic landscapes of TEs in natural populations, including hybrids. Population genetic models
have investigated the action of the major evolutionary forces on mean TE copy number (CN) per
individual (Charlesworth and Charlesworth, 1983; Charlesworth and Langley, 1989). Three such
forces are deterministic (Charlesworth et al., 1994). First, TE CNs increase at a certain
transposition rate, which incorporates the various forms of transposition regulation (either by the
host or by TEs themselves). Second, TEs are removed from genomes at a certain excision rate.
Third, most TE insertions are deleterious, such that individual fitness is assumed to be a
decreasing function of CN and natural selection acts to purge TE insertions from populations.
The fourth important force is genetic drift, a stochastic force that impacts the efficiency of natural
selection by increasing the probability of fixation of deleterious mutations (Lynch, 2007). The
intensity of drift is inversely proportional to the effective population size N_e. In populations with
small N_e, drift renders natural selection inefficient, such that deleterious mutations are more
likely to fix by chance. In principle, any of those evolutionary forces could affect hybrids in a way
that differs from their parents, leading to hybrid-specific dynamics in the accumulation of TEs.

Population data in *Drosophila* indicate that deterministic forces have a predominant influence on
the allele frequency spectrum of TE insertions (Charlesworth and Langley, 1989), consistent

with a minimal contribution from drift. Among the deterministic forces, transposition rate is

arguably the most likely to be impacted by hybridization, for instance if genetic incompatibilities

impair transposition regulation mechanisms. A classic illustration of this is hybrid dysgenesis in

Drosophila, a syndrome caused by mismatches between paternal TEs and egg-loaded pools of

Piwi-interacting RNAs (piRNAs) that lead to inefficient TE repression (Brennecke et al., 2008;

Bucheton et al., 1984; Erwin et al., 2015; Rubin et al., 1982). Similarly, hybrids between

Arabidopsis thaliana and *Arabidopsis arenosa* show overexpression of a TE family from the *A.*

arenosa subgenome that is dependent on the ploidy level of the parental accessions involved,

suggesting that dosage imbalance of epigenetic regulators causes reactivation (Josefsson et al.,

2006). It can also be expected that the number of genetic incompatibilities increases with

genetic divergence, yielding a positive relationship between evolutionary divergence and TE

deregulation (Orr, 1995; Parhad and Theurkauf, 2019). Additionally, evidence from *Drosophila*

suggests that hybridization may impact excision rates for specific TE families (Coyne, 1989).

Along with these deterministic factors, reductions in natural selection efficiency should also be

considered. Evidence from *Arabidopsis* (Lockton et al., 2008; Lockton and Gaut, 2010),

Caenorhabditis (Dolgin et al., 2008) and *Daphnia* (Ye et al., 2017) suggests that long-term

differences in N_e caused by demography or mating systems are associated with variation in TE

content (Lynch and Conery, 2003). A reduction in N_e can be expected when a novel hybrid

lineage is formed. Indeed, it has been hypothesized that population bottlenecks inherent with

hybrid speciation in *Helianthus* might have played a role in the accumulation of TEs through a

decreased efficiency of purifying selection (Ungerer et al., 2009).

An emerging eukaryotic model system in ecological and evolutionary genomics is the yeast

Saccharomyces paradoxus, the undomesticated sister species of the budding yeast S.

cerevisiae. *S. paradoxus* has a complex admixture history in North America, making it an ideal

model system to test hybridization-related evolutionary hypotheses in natural populations and in

the laboratory (Figure 1) (Hénault et al., 2017). North American *S. paradoxus* lineages comprise

the Eurasian lineage *SpA*, the endemic incipient species *SpB* and *SpC* (hereafter termed pure

lineages), and the incipient hybrid species *SpC*, SpD$_1$* and *SpD$_2$* (hereafter termed hybrid

lineages; Figure 2a). *SpC** is an introgressed lineage that arose from a post-glacial secondary

contact between the endemic lineages *SpB* and *SpC* and consists of mostly *SpC*-like genomes

with ~5% introgressions from *SpB* (Leducq et al., 2016), while *SpD$_1$* and *SpD$_2$* arose more

recently from hybridization between *SpC** and *SpB* and have roughly equal genomic

contributions from both parents (Eberlein et al., 2019) (Figure 2b).

Yeasts of the *Saccharomyces* genus are valuable model species to investigate the interplay

between hybridization and transposition. *Saccharomyces* genomes contain many long terminal

repeats (LTR) retrotransposons of the Ty1/copia (Ty1, Ty2, Ty4 and Ty5) and Ty3/gypsy (Ty3)

families. Ty1/copia and Ty3/gypsy retrotransposon superfamilies (Wicker et al., 2007)

respectively correspond to the Pseudoviridae and Metaviridae virus families (Krupovic et al.,

2018) and have a broad taxonomic distribution. Ty elements comprise two overlapping open

reading frames (ORFs) within a ~5 kb internal sequence, which is flanked by two ~300 bp LTRs

in direct orientation. The two LTRs of full-length elements are prone to intra-element

recombination, leaving behind solo LTRs in the genome (Parket et al., 1995). Ty1 in the budding

yeast *S. cerevisiae* is among the best characterized eukaryotic TEs, benefiting from decades of

research on its molecular and cellular biology and, more recently, on its population genomics

(Bleykasten-Grosshans et al., 2013; Curcio et al., 2015). Although *Saccharomyces* genomes

harbor a relatively modest fraction of Ty elements (~1-3% in size), their streamlined genomes

(~12 Mbp) and short generation time (~2h) provide great power to perform TE comparative

genomics and laboratory evolutionary experiments.

Here, we use population genomic data and laboratory evolution experiments on *S. paradoxus*

and its sibling species *S. cerevisiae* to investigate the hybrid reactivation hypothesis and, more

generally, the factors governing TE accumulation in natural and experimental lineages. Our

results show that natural and experimental hybrids exhibit no reactivation of TEs. Rather, we

show that deterministic factors like population structure and the properties of individual hybrid

genotypes are major determinants of TE content evolution in hybrid genomes.

Figure 1. Population genomic data and laboratory evolution experiments allow to uncover the effects of various factors on TE accumulation in hybrids. Upon hybridization, the TE content of a hybrid is determined by equal contributions from the TE contents of its parents. Subsequent evolution can either leave the hybrid TE content unchanged, or drive it to increase or decrease. These changes can be driven by biases in transposition or excision rates or by variation in natural selection efficiency. Two complementary approaches can be used to investigate which factors drive TE dynamics in hybrids. First, genomic data from natural populations can be harnessed to understand how population structure, environmental pressures and natural selection efficiency shaped natural variation in TE content. Second, evolution experiments in laboratory controlled conditions (constant environment and relaxed natural

selection) allow to test the effect of properties of artificial hybrid genotypes on TE accumulation,

namely evolutionary divergence between parents and initial TE content.

Results

S. paradoxus lineages exhibit contrasted TE evolutionary dynamics but no reactivation in

hybrids

To investigate the TE content and dynamics in North American *S. paradoxus* pure and hybrid

lineages, we annotated six highly contiguous whole-genome assemblies previously generated

from Oxford Nanopore long reads (PRJNA514804) (Eberlein et al., 2019). We used a database

comprising reference sequences for *S. cerevisiae* Ty1, Ty2, Ty3, Ty4 and Ty5 families, along

with *S. paradoxus* Ty3-related Ty3p (Carr et al., 2012) and Ty4-related Tsu4, the latter

originating from a horizontal transfer from the *Saccharomyces eubayanus*/*Saccharomyces*

uvarum clade to *S. paradoxus* (Bergman, 2018). These annotations revealed counts of full-

length elements an order of magnitude lower than solo LTRs (Figure 2c, Figure 2—figure

supplement 1), consistent with previously reported data on *S. cerevisiae* and *S. paradoxus* (Carr

et al., 2012; Goffeau et al., 1996; Kim et al., 1998; Yue et al., 2017). To overcome the difficulty of

comparing multiple families of short and often degenerated LTR sequences within a

phylogenetic framework, we built intra-genome LTR sequence similarity networks. Sequences

within the Ty1-Ty2 and Ty3p-Ty3 LTR family pairs were indistinguishable (Figure 2—figure

supplement 2), in agreement with Ty2 and Ty3 being specific to *S. cerevisiae* (Carr et al., 2012;

Naumov et al., 1992). Consequently, Ty2 and Ty3 LTR annotations were merged with Ty1 and

Ty3p, respectively. Although they were clearly distinct, Ty4 and Tsu4 LTRs showed a close

relatedness, which justified their combination into the Ty4/Tsu4 family. No Tsu4 LTR was

annotated in the *SpA* genome, consistent with this family being specific to North American *S.*

paradoxus and absent from Eurasian *S. paradoxus* (Bergman, 2018).

Figure 2. Ty elements showed no reactivation in hybrid lineages and contrasted abundances among pure lineages. a. Bayesian phylogenetic tree of the North American *S. paradoxus* lineages based on genome-wide SNPs. **b.** Schematic admixture graph showing the evolutionary relationships between pure and hybrid lineages as inferred by (Eberlein et al., 2019; Leducq et al., 2016). Dotted lines represent admixture events. **c.** Counts of full-length elements (top) and solo LTRs (bottom) in six whole-genome assemblies based on long reads. No full-length Ty4 or Ty5 were found in any assembly, thus these families were considered extinct. **d.** CN variation in active Ty families (i.e. families which have full-length elements) measured as $\log_2$ normalized read depth (NRD) over Ty reference internal sequences for 207 diploid wild strains. Sample size for each lineage (excluding strains for which there was no coverage) is indicated between parentheses. Whiskers span 1.5 times the interquartile range.

P-values of Kruskal-Wallis tests are shown. Stars on bars show the results of Conover post-hoc

tests. *:p<0.05, **:p<0.01, ***:p<0.001.

Based on the assembly annotations, Ty1 was the most abundant family and exhibited the

largest CN variation among lineages, with a striking difference between *SpB* and *SpC* (Figure

2c). The generality of CN patterns revealed by genome assemblies was confirmed by

normalized short-read depth (NRD) over Ty reference sequences for 207 natural diploid isolates

(Figure 2d, Figure 2—figure supplement 3, Table S1; PRJNA277692, PRJNA324830,

PRJNA479851). The hybrid lineages *SpC**, *SpD$_1$* and *SpD$_2$* had intermediate full-length Ty1 and

Tsu4 CNs compared to their parental lineages, indicating no apparent persistent reactivation or

maintenance of Ty elements in these lineages. Notably, Ty3p appeared extinct in all hybrid

lineages, with no signal for internal sequences. The most outstanding profile was that of the

pure lineage *SpC*, which had significantly higher full-length Ty1 and Ty3p CNs compared to the

other lineages. For Tsu4, *SpB* was the lineage with the highest CN. Thus, lineage-wide CNs

were consistent with an absence of reactivation in hybrids, the most contrasted profiles being

found among pure lineages.

CNs alone may not reflect the actual dynamics of a TE family within a genome. For instance, a

recently expanding family may not have had the time to reach the CN of another family (or the

same family in a different lineage) that would be more abundant but at equilibrium. We thus

explored the evolutionary dynamics of Ty families within each genome. If high CNs were caused

by recent family expansions, they should be associated with a signature of low divergence

(Bergman and Bensasson, 2007). We used the sequence divergence to the most closely related

LTR for each LTR annotation to approximate evolutionary divergence since transposition. The

importance of low-divergence peaks (≤1%) for Ty1 was significantly higher in *SpC* and *SpA*

compared to *SpB* (Fisher's exact test, FDR corrected, Figure 3a; see Figure 3—figure

supplement 1 for other divergence thresholds). *SpA* also had a significant underrepresentation

of low-divergence LTRs for the composite Ty4/Tsu4 family due to the absence of Tsu4. The

height of low-divergence peaks was consistent with CNs of full-length elements (Figure 2d).

Hybrid lineages *SpC**, *SpD$_1$* and *SpD$_2$* had Ty1 peaks similar to *SpC*, although less important.

They also had no peak for Ty3p, consistent with the absence of full-length elements. Overall,

this result indicated that CN variation reflected variation in family dynamics and confirmed that

Ty elements exhibited no reactivation in hybrid lineages.

Figure 3. Evolutionary dynamics of Ty families in *S. paradoxus*. a. Distributions of minimum nucleotide divergence between LTR sequences. Heatmaps show FDR-corrected p-values for pairwise Fisher's exact tests between ratios of elements ≤1% and >1% nucleotide divergence. Color maps are centered at the significance threshold of 0.05. Blue cells depict statistically significant tests. See Figure 3—figure supplement 1 for other divergence thresholds. **b.** LTR divergence and conservation levels. Horizontal positions depict Ty orthogroups clustered from conserved (left) to private (right). Color map shows minimum nucleotide divergence. Scattered

red dots show LTRs that belong to full-length elements. FDR-corrected p-values of Fisher's

exact tests for the ratio of full-length elements in private orthogroups are shown. ns: p≥0.05,

*:p<0.05, **:p<0.01, ***:p<0.001. **c.** Lineages through time curves for Ty1 ultrametric bayesian

phylogenetic trees, using a conserved Ty4 LTR orthogroup as an outgroup. Branching time is

scaled such that values of one correspond to the present. From left to right, distributions show

all LTRs (with a close-up on recent dynamics), LTRs from private elements and LTRs from full-

length elements.

Next, we tested whether orthology between genome-wide Ty insertion loci was consistent with

transposition activity. We defined Ty orthogroups from a whole-genome multiple alignment of the

six assemblies (Figure 3b). Most orthogroups were either private to one genome or conserved

across the six (Figure 3—figure supplement 2), suggesting that a major fraction of Ty insertion

polymorphisms segregated at low frequency in North American lineages. This was consistent

with studies in *S. cerevisiae* showing that insertions are mostly fixed or at low frequency

(Bleykasten-Grosshans et al., 2013; Carr et al., 2012). Private orthogroups comprised LTRs that

had low divergence, consistent with recent transposition activity (Figure 3—figure supplement

2). Private orthogroups were significantly enriched in full-length Ty1 elements (Figure 3b),

including hybrid lineages. This indicated that all lineages had active transposition for at least

some Ty families. It also suggested that an absence of transposition in hybrid lineages could not

explain the absence of reactivation. We called lineage-wide insertion polymorphisms from

discordant mappings of the 207 short read libraries to estimate the distribution of insertion allele

frequencies. This calling method is noisy and only allows to call non-reference alleles, but

should nevertheless provide rough estimates of the insertion allele frequency spectra. Principal

component analysis (PCA) on non-reference insertions yielded the expected grouping of strains

into well-defined lineages (Figure 3—figure supplement 3). The allele frequency distributions

showed a similar excess of very low frequency alleles for the six lineages (Figure 3—figure

supplement 3), suggesting that Ty elements were subject to largely uniform evolutionary forces

across lineages.

We focused on the most abundant Ty1 to investigate the transposition dynamics in a

phylogenetic framework (Le Rouzic et al., 2013). Bayesian phylogenetic trees of Ty1 LTRs were

converted into ultrametric phylogenies to approximate the timing of branching events using a

conserved Ty4 LTR orthogroup as an outgroup. Branching times revealed that lineages had

largely similar Ty1 dynamics (Figure 3c). Due to the shortness of LTR sequences, most

branches collapsed with the outgroup, indicating an expected saturation of the molecular

substitution signal. A focus on recent dynamics revealed a clear expansion signature in *SpC* that

was shared by *SpC** and, to a lesser extent, by *SpD₁* and *SpD₂* (Figure 3c), which was reflected

by the peaks of young private and full-length elements. This suggests that any recent expansion

signature in hybrid lineages can be largely explained by inheritance from their *SpC*-related

parent. The rapid expansion in *SpC* and *SpC** can be contrasted with the more gradual

expansion in *SpA*, which harbored the second highest Ty1 CN. This contrasted with the

assumption that TE abundances can be interpreted as equilibrium copy numbers under the

transposition-selection balance hypothesis (Charlesworth and Charlesworth, 1983) and rather

suggested a recent transposition rate increase in *SpC*-related lineages.

Population structure best explains TE content variation, with minor contributions from climatic

variation

Our results showed that hybrid lineages exhibited no outstanding Ty CN profiles or sequence

dynamics. However, potential hybrid-specific differences in transposition rate might be masked

by other factors that can be expected to impact Ty dynamics. To exclude this possibility, we

investigated which factors best explain the variation in Ty CN in natural lineages. First,

population structure incorporates both host-encoded (Curcio et al., 2015) and Ty-encoded

(Czaja et al., 2020; Saha et al., 2015) determinants of Ty transposition rates. Second,

environmental factors can influence Ty transposition rate, as demonstrated by temperature

(Paquin and Williamson, 1984) and nitrogen starvation (Morillon et al., 2000) for Ty1 in *S.*

cerevisiae. If hybrid lineages experience distinct environmental conditions in nature, this may

impact their transposition rates. Finally, the efficiency at which natural selection purges

deleterious Ty insertions (which depends on N_e) could vary between lineages, ultimately

affecting the transposition-selection balance. To help discriminate global from lineage-specific

patterns arising from these factors, analyses with all lineages aggregated were complemented

by lineage-specific analyses. However, hybrid lineages were represented by relatively few

strains (≤10 in each case), which made it hard to reach meaningful conclusions for some

analyses. Thus, some of the lineage-specific analyses were focused on the more abundant *SpB*

and *SpC* pure lineages.

We first investigated the relative contribution of population structure and environmental factors.

We combined genome-wide single nucleotide variants from our population genomic data with

climatic (temperature, precipitation, solar radiation, vapor pressure, wind) and water balance

data (Abatzoglou et al., 2018) and reduced the dimensionality of both datasets by PCA, using

coordinates on the major principal components (PCs) to describe population structure (Figure 4

—figure supplement 1) and climatic variation (Figure 4—figure supplement 2, Figure 4—figure

supplement 3). We then fitted linear models to compare how genotypes-based PCs (gPCs) and

environment-based PCs (ePCs) can explain CN variation in Ty1, Ty3p and Tsu4 (Figure 4a).

Globally, gPCs reached higher levels of statistical significance as linear predictors of CN

compared to ePCs. gPC1 both explained a high proportion of variance (52%, 9% and 36% for

all lineages, *SpB* and *SpC*, respectively; Figure 4—figure supplement 1) and was a highly

significant linear predictor of Ty1 CN. Linear models that explained Ty CNs with both gPCs and

ePCs had generally lower Akaike information criterion (AIC) values than models that included

only gPCs (Table S2), suggesting that climatic variables have modest although significant

contributions in explaining CN variation. Unsupervised clustering of strains along gPCs

consistently yielded more homogeneous clusters than ePCs in terms of Ty CN (Figure 4b,

Figure 4—figure supplement 4). Overall, these results show that the genetic structure of North

American *S. paradoxus* is generally a much better predictor of Ty CNs across lineages than

climatic variables. Since locations where hybrid lineages were sampled have climatic profiles

similar to pure lineages (Figure 4—figure supplement 2), exposure to specific environmental

conditions appears unlikely to explain the absence of observable Ty reactivation in hybrid

lineages.

Figure 4. Associations between population structure, climatic variables and Ty CNs for all lineages (left), *SpB* only (middle) or *SpC* only (right). a. Linear models fitted on Ty CNs with gPCs and ePCs as independent variables. For each PC, the p-value in the linear model is plotted against the proportion of variance explained in the respective PCA. **b.** Unsupervised

clustering performed on either gPC or ePC coordinates. Cumulative distributions show the

standard deviation in Ty CN within each gPC-cluster or ePC-cluster. Distributions are also

shown as boxplots, with whiskers spanning 1.5 times the interquartile range. FDR-corrected p-

values of two-tailed Mann-Whitney U tests between gPC-cluster and ePC-cluster CN

distributions are shown. ns: p≥0.05, *:p<0.05, **:p<0.01.

Variation in natural selection efficiency cannot explain TE abundance variation in wild lineages

Most TE insertions are expected to have deleterious effects on the individual in which they occur

(Wilke and Adams, 1992), although some can be advantageous (Aminetzach et al., 2005;

Filteau et al., 2015; Gresham et al., 2008; Hof et al., 2016). Natural selection acts to purge

deleterious insertions, and its efficiency depends on the strength of genetic drift opposing it

(which is inversely proportional to N_e). Ty reactivation in hybrid lineages might be

counterbalanced by higher efficiency of natural selection in these lineages, although there is no

a priori reason to expect higher N_e in hybrids. We thus sought to investigate if CN variation

across lineages can be explained by variation in N_e. Genome-wide nucleotide diversity at four-

fold degenerate codon positions (π_S) is related to long-term N_e and was used to approximate

this parameter across diverse taxa (Lynch and Conery, 2003). Nucleotide diversity was also

used at the intraspecific level in *S. paradoxus* to estimate N_e (Tsai et al., 2008). While pure

lineages did exhibit variation in π_S (Table S3), there was no obvious negative relationship

between Ty CNs and π_S (Figure 5—figure supplement 1), as would be expected under a

transposition-selection equilibrium (assuming negligible variation in mutation rate). For instance,

nucleotide diversity was higher in *SpC* than in *SpA*, while *SpC* exhibited the highest Ty1 and

Ty3p CN. While these rough N_e estimates should be interpreted with caution, they suggested

that natural selection efficiency is likely not the main determinant in CN variation. However, our

data indicated that some Ty families experienced recent expansions and thus may not be at

equilibrium. Recent hybrid lineages also cannot be meaningfully characterized by long-term N_e,

further limiting the usefulness of π_S as a measure of the efficiency of natural selection.

We can expect that most of the fitness effects associated with TE insertions are caused by

alterations of host functions encoded in *cis.* Perhaps the TE insertions with the largest functional

alteration potential are those which interrupt ORFs of host genes. Across the six genome

assemblies, no Ty insertion was found to interrupt an ORF. Lineage-wide Ty insertion calls

revealed four Ty insertions predicted to be within host ORFs, three of which are found in *SpC*

strains and one in *SpB*. All the ORF-disrupting insertions were private alleles, and in all cases

the strain harboring them had a CN higher than the median of its lineage (Figure 5—figure

supplement 2). Examination of depth of coverage at these four loci revealed narrow peaks in the

corresponding strains that were consistent with target site duplications (TSDs) arising from Ty

integration (Figure 5—figure supplements 3-6). The absence of ORF-disrupting Ty insertions in

hybrid lineages and their scarcity in pure lineages suggests that natural selection is globally very

efficient at purging these mutations.

The number of ORF-disrupting Ty insertion alleles in *SpC* (three) compared to *SpB* (one)

suggested a slight overrepresentation in *SpC*, which could be explained by a bias towards more

deleterious insertion alleles in this lineage. To test this hypothesis, we examined the functional

annotations of the corresponding genes. There were striking patterns of functional relatedness

among genes harboring an ORF-disrupting insertion. Two *SpC* strains had independent Ty1

insertions at different loci within YDL024C (*DIA3*, which encodes a protein of unknown function

(Palecek et al., 2000)), as evidenced by the corresponding TSDs (Figure 5—figure supplement

3, Figure 5—figure supplement 4). The two other genes were involved in resistance to arsenic

compounds: 16_037 (*SpB*) had a Tsu4 insertion in YPR200C (*ARR2*, which encodes an

arsenate reductase (Mukhopadhyay and Rosen, 1998), Figure 5—figure supplement 5), while

LL2011_004 (*SpC*) had a Ty1 insertion disrupting YPR201W (*ARR3*, which encodes an

arsenite/H+ antiporter (Wysocki et al., 1997), Figure 5—figure supplement 6). This functional

relatedness could be explained by positive selection; however, this scenario is incompatible with

the extremely low frequency at which these insertions segregate. Thus, the most parsimonious

explanations for these patterns appears to be either a mutational bias favoring Ty insertions at

these loci or fitness effects too small to be visible to natural selection.

We investigated the fitness effect of the ORF-disrupting Ty1 insertion in *ARR3* by measuring the

growth of a collection of 123 *S. paradoxus* strains in culture medium supplemented with arsenite

(Figure 5a, Figure 5—figure supplement 7, Figure 5—figure supplement 8). Growth in 0.8 mM

sodium meta-arsenite ($NaAsO_2$) showed significant variation between lineages (mixed design

ANOVA, p=$9.54×10^{-39}$), and more total variance was explained by variation between lineages

(lineages: 86.58%, strains: 10.20%, residuals: 3.22%). *SpC* and *SpC** lineages exhibited poor

growth in 0.8 mM arsenite compared to *SpB* and *SpA* (Figure 5a). This phenotype was

generalized to the whole lineages, and while strain LL2011_004 was in the lower part of the

SpC distribution, it was not significantly different from many *SpC* strains not harboring an

insertion (Figure 5b, Figure 5—figure supplement 9). This indicated that the disruption of a

major gene involved in arsenite resistance had no further deleterious effect on the poor growth

ability of *SpC*-related strains in presence of arsenite. The absence of growth defect in arsenite

for LL2011_004, combined with the extremely low frequencies of all the ORF-disrupting alleles,

suggests that these insertions are effectively neutral and are not observed because of an

important reduction in purifying selection efficiency.

Figure 5. Genomic patterns of Ty insertions indicate that natural selection efficiency cannot explain CN variation in natural lineages. a. Difference between growth curves in 0.8 mM NaAsO₂ (As) and control medium (synthetic complete, SC). Curves depict the difference in culture optical density (OD) through time, averaged across replicates and bins of four timepoints

(one hour). 123 strains were assayed. *SpC* strain LL2011_004, which harbors a Ty1 insertion in

ARR3, is highlighted with a dashed line. **b.** Area under the curve (AUC) of the difference

between growth curves in As and SC. Replicates of 107 strains that passed the criteria of

number of replicates and homoscedasticity are shown. The number of strains per lineage is

shown between parentheses. Replicates of the *SpC* strain LL2011_004 are highlighted with

large empty circle symbols. Lowercase letters indicate groups that are not significantly different

following Tukey's HSD test. **c.** Counts of essential and non-essential genes in the immediate

vicinity of Ty insertions across the six whole-genome assemblies. Essential to non-essential

ratios are shown on top of the bars. **d-e.** Cumulative distributions of number of physical

interactions (d) and ω (e) for genes neighboring conserved (dashed line) or private (solid lines)

Ty insertions. Genes containing ORF-disrupting insertions are labeled with symbols. Heatmaps

show FDR-corrected p-values for pairwise Mann-Whitney U tests between distributions. Color

maps are centered at the significance threshold of 0.05. **f.** Cumulative distributions of Δ*k*

between *SpB* and *SpC*. **g.** Counts of neighboring genes with positive or negative Δk values.

To gain a more comprehensive view of the strength of selection acting on Ty insertions, we

compared the properties of genes in their neighborhood. Ty elements contain many transcription

factor binding sites and can affect the expression levels of adjacent genes (Morillon et al., 2000;

Coney and Roeder, 1988; Todeschini et al., 2005; Lesage and Todeschini, 2005). Dosage-

sensitive and functionally central genes should on average be subject to stronger selection

against mutations that affect their regulation. For instance, in *S. cerevisiae*, genes that are

essential or that have many protein-protein interactions were shown to be depleted in eQTLs,

i.e. variants that affect expression levels (Albert et al., 2018). However, the association of Ty

insertions with regulatory effects on adjacent genes is expected to be highly variable, as

transcriptional effects are known to depend on the location, orientation and nucleotide sequence

of the inserted element (Coney and Roeder, 1988; Lesage and Todeschini, 2005). Nevertheless,

if selection efficiency explains a significant part of Ty CN variation, we reason that when CNs

are high, Ty insertions should be more frequently observed near genes which properties reflect

stronger dosage and functional constraints, compared to when CNs are low. Focusing on the

Ty insertions private to each lineage, we found no significant difference in the proportion of

neighboring genes annotated as essential in *S. cerevisiae* (p>0.05, pairwise Fisher's exact

tests, FDR correction), although *SpC* did exhibit the highest essential to non-essential ratio

(Figure 5c). Distributions of physical distance to neighboring genes were similar across

lineages, as were distributions of relative orientation (Figure 5—figure supplement 10).

Additionally, genes neighboring Ty insertions private to *SpC* show no bias towards higher

number of physical interactions (Figure 5d, p>0.05, pairwise two-tailed Mann-Whitney U tests,

FDR correction). We fitted the ratio of non-synonymous to synonymous substitutions (ω) to

each neighboring gene to estimate the strength of purifying selection on protein-coding

sequences. The ω distributions for genes neighboring private Ty insertions exhibited no

significant bias (p>0.05, pairwise two-tailed Mann-Whitney U tests, FDR correction). In

particular, the distribution in *SpC* exhibited no skew towards lower values, suggesting no

association with genes under strong purifying selection (Figure 5e). Notably, genes harboring

ORF-disrupting insertions show little or no physical interactions and no signature of purifying

selection (Figure 5d-e).

We next investigated whether lineage-specific selection signatures on genes are associated

with Ty insertions. We used the RELAX branch-site codon model (Wertheim et al., 2015) to

derive estimates of selection intensity (k) along individual gene trees. Within the RELAX

framework, $k<1$ indicates relaxed selection and $k>1$ indicates strengthened (positive or

purifying) selection. We define $\Delta k = k_{SpB} - k_{SpC}$, corresponding to the difference between the

average k values for clades comprising *SpB* and *SpC* strains, respectively. Overall, the

distribution of Δk values for genes neighboring private *SpC* insertions was very similar to that for

conserved insertions (Figure 5f), while the distribution for private *SpB* insertions was slightly

biased towards genes relaxed in *SpB*. For both *SpB* and *SpC*, there were more private

insertions adjacent to genes experiencing relaxed selection in the corresponding lineage,

although comparisons with conserved insertions were not statistically significant ($p>0.05$,

Fisher's exact tests, Figure 5g). This is consistent with efficient purifying selection acting against

insertions near genes in both lineages. While genes harboring ORF-disrupting insertions have

signatures of stronger selection in *SpC* (Figure 5e, Figure 5—figure supplement 11), this

observation is mitigated by their far-right position in the ω distribution (Figure 5d). Overall, these

results show that variation in natural selection intensity is unlikely to explain a large part of the

variation in Ty CNs in natural lineages.

TE content variation in experimental evolution under relaxed selection reveal no systematic

effect of hybridization but major effects of individual genotypes

Thus far, our results showed that no reactivation is apparent in natural hybrid lineages, that

population structure over environmental variables best explains variation in Ty CNs and that

natural selection efficiency is unlikely to explain much variation in Ty genomic landscapes.

However, natural selection seems equally efficient at purging deleterious Ty insertions in all

lineages, leaving the possibility that strong purifying selection could mask any reactivation in

hybrids. Transient reactivation may also be followed by rapid re-establishment of host

repression and leave little to no signature in contemporary genomes. Moreover, natural hybrids

are somewhat limited in their ability to uncover the interplay between hybridization and

transposition, as they are relatively recent introgressed lineages that resulted from admixture

between closely related lineages. To systematically explore the effect of hybridization on Ty

transposition rates, we performed a large-scale mutation accumulation (MA) experiment on 11

artificial hybrid backgrounds, eight of which were previously published (Charron et al., 2019). A

MA experiment with yeast consists of subjecting a population to periodic single-cell bottlenecks

(typically during hundreds to thousands of mitotic generations) by streaking a randomly chosen

single colony, thus amplifying genetic drift and making natural selection inefficient (Lynch et al.,

2008) (Figure 6a). *S. paradoxus* was inferred to reproduce mitotically 99.9% of the time in the

wild (Tsai et al., 2008), making MA experiments a reasonably realistic approximation of its

natural life cycle. Pairs of haploid derivatives of natural *S. paradoxus* and *S. cerevisiae* strains

were chosen to span a range of evolutionary divergence (Figure 6b), yielding four categories of

intraspecific crosses (from the least to the most divergent): *SpC×SpC* (CC), *SpB×SpB* (BB),

SpB×SpC (BC) and *SpB×SpA* (BA); and one category of interspecific crosses: *SpB×S.*

cerevisiae (BSc; Figure 6c, Tables S4-6). The crosses used also spanned a range of variation in

parental Ty CNs (Figure 6d). For each cross, 48 to 96 MA lines derived from independent

matings were propagated for ~770 mitotic divisions.

Figure 6. Ty CN variation in MA lines reveal no effect of parental divergence and major genotype-specific effects. a. Principle of a MA experiment on a yeast population. N

corresponds to population size. **b.** Bayesian phylogenetic tree of the parental haploid strains

used to generate artificial hybrids. **c.** Design of the crosses for the MA experiment. **d.** Ty CNs in

the haploid parental strains of each cross in c. **e.** CN at the onset (T_{ini}, empty boxes) and at the

end (T_{end}, filled boxes) of the MA experiment. Whiskers span 1.5 times the interquartile range.

FDR-corrected p-values of Wilcoxon signed-rank tests and Brown–Forsythe tests for the

equality of variances are shown in black and blue, respectively. ns: $p \geq 0.05$, *:$p<0.05$, **:$p<0.01$,

***:$p<0.001$. **f.** Linear models explaining the mean variation in CN ($\Delta CN = CN_{Tend}-CN_{Tini}$) for a

given cross as a function of the joint CN and genetic divergence of its parental strains. The p-

value for each term in the models is shown. Scatter plots (CCPR plots) show the partial

residuals for each independent variable, along with the fitted regression.

We selected a random subset of 23 to 40 MA lines per cross and performed whole-genome

resequencing on stocks frozen at T_{ini} and T_{end}, totaling 372 strains sequenced at each timepoint

(PRJNA515073). All these strains were confirmed to be diploid at T_{ini} and T_{end} by quantifying

total DNA content using flow cytometry ((Charron et al., 2019), Marsit *et al.* in prep). Focusing

on full-length Ty1, Ty3p and Tsu4, no global trend emerged from the variation of NRD before

and after MA (Figure 6e, Figure 6—figure supplement 1). Mixed effects linear models fitting

cross-specific intercepts and slopes showed no statistically significant support for a global effect

of time on CNs. Nevertheless, specific combinations of crosses and Ty families showed

significant differences in either central tendency or variance. Often, crosses within a category of

parental strain divergence showed contrasting patterns. For example, although CC1, CC2 and

CC3 are genetically very similar *SpC×SpC* crosses, only CC1 exhibits a significant decrease in

Ty1 and Tsu4 CNs. There were also cases of opposite trends within the same category of cross,

for instance Ty1 CN in BC and BA crosses. This showed that individual parental genotypes,

even if they are evolutionarily closely related, can have very different dynamics, both in sign and

magnitude.

We next sought to understand how these dynamics are determined by properties of the different

hybrid genotypes tested. We fitted linear models to the variation in CN between T_{ini} and T_{end}

(ΔCN), using the sum of CNs of both parental strains of a cross and the evolutionary divergence

between them as independent variables. Although these models explain little global variance

(R^2=0.40, 0.08 and 0.06 for Ty1, Ty3p and Tsu4 respectively), we found that neither independent

variable was a significant predictor of ΔCN (Figure 6f). Only Tsu4 showed a slightly positive

regression slope with respect to evolutionary divergence, and none of them was statistically

significant, indicating that divergence between parents of a hybrid has no large influence on its

CN dynamics. On the other hand, Ty1 ΔCN had a negative relationship with parental Ty1 CN

and was marginally significant (p=0.056), suggesting that the initial CN of a hybrid genotype can

dictate the sign and magnitude of its CN change. However, this result should be interpreted with

care as some of the crosses shared common parental strains and were thus not entirely

independent. This relatedness may in fact explain part of the pattern seen for Ty1. Crosses that

shared the *SpB* parental strain UWOPS-91-202 (BB2, BC2, BA2, BSc2) seemed to have

systematically higher Ty1 ΔCN values than the corresponding crosses that shared the *SpB*

parental strain MSH-604 (BB1, BC1, BA1, BSc1). UWOPS-91-202 appeared to be a naturally

Ty-less strain, as no full-length element was found for any family in its genome (Figure 6d).

Previous reports of such Ty-less strains were shown to be permissive to the accumulation of Ty

elements when assayed in the laboratory using a reporter assay (Czaja et al., 2020; Garfinkel et

al., 2003). This observation also supports the hypothesis that genotype-specific characteristics

can be major determinants of variation in Ty CN.

Discussion

The literature on TE reactivation in hybrids demonstrates its relevance to the field of speciation (Serrato-Capuchina and Matute, 2018). Reactivation can be a source of postzygotic reproductive isolation by compromising genome stability (Dion-Côté and Barbash, 2017), which translates into a spectrum of deleterious hybrid phenotypes spanning sterility, developmental abnormalities and inviability. Many studies investigated the TE reactivation hypothesis at the interspecific level and yielded equivocal evidence. For instance, mixed evidence for TE reactivation was found within the *Drosophila* genus (Coyne, 1989; Hey, 1988; Labrador et al., 1999; Vela et al., 2014), where reactivation seems to depend on the species pair involved. On the other hand, three well-established hybrid species that emerged from parallel hybridization between the same species pair in the genus *Helianthus* all exhibit important reactivation of LTR retrotransposons, both at the genomic (Ungerer et al., 2009, 2006) and transcriptomic (Renaut et al., 2014) levels. However, a study of hybrid zones between the two parental species revealed that no reactivation is apparent in recent hybrid populations, casting doubt on the idea that hybridization alone triggered reactivation and suggesting the involvement of other factors like abiotic stress (Kawakami et al., 2011). Overall, this data nuances the hypothesis that hybridization could act as a systematic trigger of TE proliferation and reproductive isolation.

In this study, we used population genomic data on North American lineages of the yeast *S. paradoxus* to test whether TEs were reactivated in natural hybrids. It was previously shown that these lineages experienced two hybridization events that resulted in the establishment of admixed lineages that have partial postzygotic reproductive isolation with their respective parental lineages (Eberlein et al., 2019; Leducq et al., 2016). We found no evidence of TE reactivation in any of these natural hybrid lineages. Our data showed that while some hybrid lineages may have relatively high CNs and exhibit evolutionary dynamics consistent with recent

expansions, these signatures were never more pronounced than that of their parental lineages

(*SpC* in the case of *SpC** and *SpC** in the case of *SpD₁* and *SpD₂*). Moreover, our analyses

suggest that environmental and demographic factors are unlikely to be major factors affecting

TE CNs in natural lineages, which therefore likely reflect the underlying transposition rates. Our

observations would seem consistent with a role of TE reactivation in speciation, as natural

hybrids with high levels of reactivation would likely be unobserved. However, they contrast with

long-established hybrid species in *Helianthus*, showing that reactivation and long-term

maintenance can be compatible.

We note that the analysis of contemporary genomes is inherently limited in the ability to uncover

transient reactivation that would immediately follow hybridization and rapidly faint, perhaps as a

result of a re-establishment of TE repression. However, this scenario may be unlikely in yeast,

given the absence of many epigenetic and post-transcriptional repression mechanisms (see

discussion below) that can yield such a rapid re-establishment in other species. For instance,

the re-establishment of piRNA-mediated P element repression in *Drosophila* P-M dysgenic

hybrids occurs in single female individuals because paternally-inherited piRNA clusters become

expressed as they age (Khurana et al., 2011). If a hypothetical re-establishment of Ty repression

was rather driven by selection for genomic variants, this should result in a gradual improvement

of repression with time. Our data provides no support for this hypothesis, as younger hybrid

lineages *SpD₁* and *SpD₂* do not show more pronounced signatures of recent Ty expansion

compared to the older *SpC** lineage. Additionally, Ty CNs of artificial hybrids at the initial time

point of our MA experiment (i.e. early after hybridization) generally show good agreement with

the expectation from their parental haploid strains, which is incompatible with an early

reactivation of Ty elements.

Many factors are susceptible to determine whether a hybrid genotype exhibits reactivation or

not. According to the Bateson-Dobzhansky-Muller (BDM) model, genetic incompatibilities should

accumulate as populations diverge (Orr, 1995). Additionally, the evolutionary conflict between

host genomes and TEs results in an arms race that can accelerate the evolution of the involved

genes (Blumenstiel et al., 2016; Van Valen, 1973). Thus, it can be expected that

incompatibilities affecting transposition rates are more likely to be observed in hybrids between

highly diverged genotypes compared to closely related ones. Here, we report data from

mutation accumulation (MA) experiments on artificial hybrids that probe the relationship

between evolutionary divergence and transposition rates. We show that contrary to the

expectation from the BDM model, hybrids between more divergent genotypes exhibit no

deregulation of TEs. This conclusion is consistent with recent results from Smukowski Heil and

colleagues, who showed that no Ty1 reactivation occurs in experimental interspecific hybrids

between *S. cerevisiae* and *S. paradoxus* (Smukowski Heil et al., 2020). Instead of reactivation,

many hybrid genotypes exhibited marked decreases in TE CNs after evolution. This trend is

especially important for Ty3p, as eight out of ten genotypes exhibit a negative change in median

CN. Ty3p is also the least abundant Ty family in natural lineages, as it is found in rarely more

than ~5 copies and extinct in all hybrid strains sampled. Such low abundance families are likely

more prone to extinction, as stochastic fluctuation in CN would more frequently lead to the loss

of all copies. Stochastic loss of Ty copies may come from intra-element LTR recombination or

loss of heterozygosity at a locus where a Ty element is present in one parent only, although the

mechanistic causes of CN decrease remain to be investigated. This result also seems

consistent with the reported Ty CN decrease in mitotically propagated experimental populations

of *S. cerevisiae* (Bast et al., 2019), suggesting a potential intrinsic bias towards TE loss.

However, it should be noted that the latter study used haploid populations in a regime with

efficient natural selection (McDonald et al., 2016), thus mitigating comparisons with our results.

Transposition is a complex cellular process, with multiple steps regulated by both TEs and host

genomes. Genetic screens in *S. cerevisiae* revealed that genes involved in multiple cellular

functions (chromatin organization, transcription, translation, RNA and protein metabolism) affect

Ty1 transposition rate, either positively or negatively (reviewed in (Curcio et al., 2015)). Thus, it

is possible that divergent hybrids suffer from misregulation at different steps of the transposition

process that have antagonistic effects, such that they cancel out at the level of transposition

rates. It is also possible that such antagonistic interactions exist between transposition and

excision rates, yielding no net change in CN. Answering these questions will require a thorough

investigation of novel Ty insertion loci in the MA lines, which is beyond the scope of this study.

Transposition regulation being a polygenic trait with a large mutational target is also consistent

with the strong genotype specificity we observed in our artificial hybrids. Some of the contrasted

dynamics between crosses that share high genetic similarity might originate from cryptic genetic

variation in genes involved in Ty regulation, whose effect on transposition rate would be

uncovered in hybrids.

TEs are structurally very diverse and populate the genomes of a vast taxonomic range, and the

control mechanisms in response to their mobilization may exhibit a correspondingly large

diversity. In many eukaryotes, TEs are repressed epigenetically through mechanisms that

include DNA methylation, chromatin modification and mRNA degradation, processes in which

small RNAs play central roles (Lisch, 2009; Slotkin and Martienssen, 2007). In many fungi, TE

repression acts via multiple mechanisms involving RNA interference (RNAi) or cytosine

methylation, including repeat-induced point mutation (RIP) (Gladyshev, 2017). Notably,

introgressed species of the fungal genus *Coccidioides* harbor genomic signatures of efficient

RIP (Neafsey et al., 2010), suggesting that this mechanism may remain active in the face of

hybridization. Unlike other eukaryotes, *S. cerevisiae* and its closely related species lost the

genes of the RNAi pathway (Drinnenberg et al., 2009) and have no cytosine methylation (Proffitt

et al., 1984). Instead, *S. cerevisiae* Ty1 has a potent self-repression mechanism, which strength

depends on the actual Ty1 CN in the host genome (thus termed CN control, CNC) (Garfinkel et

al., 2003). CNC is a post-transcriptional mechanism that relies on p22, an N-truncated form of

the Gag (p49) Ty1-encoded capsid protein which is translated from an alternative transcript of

Ty1 (Saha et al., 2015). p22 inhibits transposition by interfering with the assembly of the

cytoplasmic virus-like particles that are essential to the replication cycle of Ty1 (Saha et al.,

2015). This unique repression mechanism may explain some of the patterns we observed in the

artificial hybrids. In particular, our data suggests a negative relationship between the initial Ty1

CN in hybrid genotypes and the change in Ty1 CN between the onset and the end of the MA

experiment. This pattern seems consistent with an efficient action of CNC, although our results

would then suggest that CNC efficiency is largely independent of the degree of evolutionary

divergence between the subgenomes of a hybrid. This contrasts with the recent finding that

interspecific sequence divergence in p22 has a dramatic impact on the action of CNC (Czaja et

al., 2020), although such impact is likely mitigated by the generally much smaller divergence

levels in our crosses. Thus, further experimental work is needed to disentangle the mechanistic

contributions of Ty1 CN and sequence divergence in the regulation of transposition rates in

Saccharomyces hybrids.

In conclusion, our work illustrates how the use of natural model systems that are amenable to

experimental investigations can bring novel insights on long-standing evolutionary hypotheses.

Our results show that natural hybrid lineages of the wild yeast *S. paradoxus* exhibit no TE

reactivation and suggest that dynamics of TE content evolution in hybrids depend on

deterministic factors that evolutionary divergence between parents can hardly predict.

 Materials and Methods

 Key Resources Table

Reagent type (species) or resource	Designation	Source or reference	Identifiers	Additional information
Sequence-based reagent	Forward primer to construct the deletion cassette for the *ADE2* gene in *SpC*	Charron, Marsit et al., 2019	CLOP97-F5	acaattaaggaatcaa gaaaccgtgataaaa aattcaagtCAGCT GAAGCTTCGTA CGC
Sequence-based reagent	Reverse primer to construct the deletion cassette for the *ADE2* gene in *SpC*	Charron, Marsit et al., 2019	CLOP97-F6	gtaattgttcgctggcc aagtatattaatacattt atataGCATAGGC CACTAGTGGAT C
Strain	Haploid parent for MA lines	*Charron et al., 2014*	LL2011_001 *MATa hoΔ::kanMX4*	
Strain	Haploid parent for MA lines	*This study*	LL2011_001 *MATa hoΔ::kanMX4 ade2Δ::hphNT1*	
Strain	Haploid parent for MA lines	*Leducq et al., 2016*	LL2011_012 *MATa hoΔ::kanMX4*	
Strain	Haploid parent for MA lines	*This study*	LL2011_012 *MATa hoΔ::kanMX4 ade2Δ::hphNT1*	
Strain	Haploid parent for MA lines	*Charron et al., 2014*	MSH-587-1 *MATa hoΔ::natMX4*	
Strain	Haploid parent for MA lines	*This study*	MSH-587-1 *MATa hoΔ::natMX4 ade2Δ::hphNT1*	
Strain	Haploid parent for MA lines	Charron, Marsit et al., 2019	LL2011_004 *MATα hoΔ::kanMX4 ade2Δ::hphNT1*	
Strain	Haploid parent for MA lines	Charron, Marsit et al., 2019	LL2011_009 *MATα*	

			hoΔ::natMX4 *ade2Δ::hphNT1*	

General tools

GNU Parallel version 20170922 (Tange, 2011) and SnakeMake version 5.5.4 (Köster and

Rahmann, 2012) were used extensively in the following analyses. Unless otherwise stated,

Python version 3.7.6 (Van Rossum and Drake, 2009) was used for the custom scripts used in

the following analyses. These scripts were run on JupyterLab version 2.0.1 (Thomas Kluyver,

Benjamin Ragan-Kelley, Fernando Pérez, Brian Granger, Matthias Bussonnier, Jonathan

Frederic, Kyle Kelley, Jessica Hamrick, Jason Grout, Sylvain Corlay, Paul Ivanov, Damián Avila,

Safia Abdalla, Carol Willing, Jupyter Development Team, 2016) and made extensive use of the

following packages: BioPython version 1.76 (Cock et al., 2009), DendroPy version 4.4.0

(Sukumaran and Holder, 2010), Matplotlib version 3.2.0 (Hunter, 2007), NetworkX version 2.4

(Hagberg et al., 2008), NumPy version 1.18.1 (van der Walt et al., 2011), pandas version 1.0.2

(McKinney, 2010), SciPy version 1.4.1 (Virtanen et al., 2020), seaborn version 0.10.0 (Waskom

et al., 2020).

Protein-coding genes annotation and orthogroup definition

Whole-genome *de novo* assemblies were previously generated for six North American *S.*

paradoxus strains using a combination of long (Oxford Nanopore) and short (Illumina) reads

(PRJNA514804). Protein-coding genes were predicted *ab initio* using AUGUSTUS version 3.3.1

with parameters species=saccharomyces genemodel=partial singlestrand=false. Predicted ORF

sequences for the six assemblies were combined with ORF annotated from the *S. cerevisiae*

strains S288c and YPS128 from (Yue et al., 2017) as references. Protein-coding genes

orthogroups were inferred from sequence similarity and synteny using the program SynChro

version Jan2015 (Drillon et al., 2014) with parameters a=0 DeltaRBH=1. Orthogroups were

defined as genes with one-to-one relationships across all pairwise comparisons between the

eight genomes. Protein-coding genes systematic names were propagated from S288c to the

corresponding orthogroups.

Annotation of Ty LTR retrotransposons

Ty LTR retrotransposons were annotated from the six whole-genome assemblies using

RepeatMasker version open-4.0.7 (Smit et al., 2015) with parameters -s -low -gccalc and the

library of reference LTR and internal sequences assembled by (Carr et al., 2012) (containing

Ty1, Ty2, Ty3, Ty4 and Ty5 from *S. cerevisiae* and Ty3p from *S. paradoxus*), additionned with

Tsu4 from *S. uvarum* (Bergman, 2018). RepeatMasker output was defragmented using

REannotate version 26.11.2007 (Pereira, 2008) with parameters -s 15000 -d 10000, yielding

defragmented elements classified as full-length, solo LTR, truncated or orphan internal

sequences. The -f parameter was used to allow defragmentation of Ty1/Ty2 hybrid elements

following (Carr et al., 2012). The defragmented annotations were manually curated to split

defragmented elements spanning entire protein-coding genes or forming concatemers.

Network-based Ty LTR sequence similarity analysis

We analyzed the sequence diversity of LTR annotations within each of our six *S. paradoxus*

assemblies using a network approach (Levy et al., 2017). The sequences of all LTR annotations

within a genome were extracted and all-against-all search was performed between LTR

sequences in a genome using BLASTN version 2.7.0 (Camacho et al., 2009) with parameters

word_size 7 dust no. Networks were built using the Python package Networkx version 2.4

(Hagberg et al., 2008) by keeping hits with a bit score higher or equal to the 10th percentile of

all bit scores for a given LTR. Visualization of the resulting networks was done using Cytoscape

version 3.7.1 (Shannon et al., 2003) using the Prefuse Force Directed layout algorithm.

Inspection of these networks justified the merging of the following LTR families: Ty1-Ty2, Ty3p-

Ty3 and Tsu4-Ty4.

Estimation of evolutionary divergence since transposition

Evolutionary divergence since transposition was estimated from inter-element LTR sequence

identity, assuming that the LTRs of a newly inserted element are identical to those of the source

element. From the sequence similarity graphs, the closest relative of each element was

retrieved by keeping the hit with the highest bit score. Hits between the two LTRs of full-length

elements were excluded, and only one LTR per full-length element was considered. For each

element, the evolutionary divergence since transposition was extracted as the percent

sequence identity to the closest relative.

Definition of Ty orthogroups from genome assemblies

Orthology of LTR retrotransposons in the six whole-genome assemblies was assessed as

follows. Whole-genome multiple sequence alignment (MSA) with rearrangements was

performed using Mugsy version 1r2.3 (Angiuoli and Salzberg, 2011) with parameters -d 20000

-nucmeropts "-b 1000 -l 15" --fullsearch --refine mugsy. MSA blocks for which at least one

genome had no corresponding sequence were trashed. The remaining alignment blocks were

scanned in 10kb sliding, non-overlapping windows in both directions and windows were trimmed

until at least 80% of sites were informative in a MSA window. Ty annotation coordinates were

transformed into an MSA coordinates system and clustered using the DBSCAN algorithm

implemented in the Python package scikit-learn version 0.22.2 with parameters min_samples=1

eps=500. This one-dimensional clustering approach allowed to define orthologous clusters

containing at least one Ty annotation in at least one genome.

39   39

We defined a set of confident orthogroups to derive an empirical distribution of physical distance

between start and end coordinates of orthologs. Clusters were selected if they contained at

most one annotation per genome and if all annotations were on the same strand and belonged

to the same family. Pairwise BLASTN searches within each cluster were used to keep elements

that produced at least one hit with ≥80% nucleotide identity and an alignment length between

90% and 110% of both query and subject lengths. From the 38 resulting confident orthogroups,

we extracted the distribution of differences between annotation start and end coordinates and

used the 95th percentile of that distribution (17 bp) as threshold for the following orthogroup

definition procedure. Within each cluster, pairs of annotations meeting the following two criteria

were considered orthologous: the pair had to include at least one best BLASTN hit, and either

start or end coordinates (or both) had to be within the defined threshold of 17 nt. This method

allowed to define 713 Ty orthogroups, 20 of which (3%) had >1 element for at least one genome

and were discarded.

Phylogenetic analysis of Ty1 evolution

For each of our six genome assemblies, we extracted the LTR sequence of all Ty1 annotations,

keeping only one LTR at random for full-length elements. As an outgroup, a conserved Ty4 LTR

orthogroup was added on the basis of minimal variance in start and end coordinates across

genomes and minimal difference between the length of the reference Ty4 sequence and the

average length of the annotations. MSAs were computed using MUSCLE version 3.8.31 (Edgar,

2004) and filtered to maximize the proportion of informative sites while keeping the outgroup,

first discarding sequences with >25% gaps and then removing sites with ≥25% gaps. Bayesian

phylogenetic trees were computed using MrBayes version 3.2.5 (Ronquist et al., 2012) with

model parameters Nst=6 Rates=invgamma. Markov chain Monte Carlo parameters were set to

10 million generations for two runs of four chains each with 25% burn-in fraction and

temperature parameter of 0.03 to improve the efficiency of Metropolis coupling. Bayesian

phylogenetic trees were converted into ultrametric phylogenetic trees using PATHd8 (Britton et

al., 2007) and rooted at the Ty4 LTR outgroup. The branching time of each internal node in the

ultrametric trees was computed as the distance from the root. Branching events were counted

as the number of children nodes for each internal node, as the bayesian phylogenetic

reconstructions yielded many polytomies at the base and tips of the trees.

Construction of a library of reference Ty sequences for S. paradoxus

We combined the Ty LTR sequences of all families and full-length sequences of Ty1, Ty3p and

Tsu4 from the defragmented annotations of our six *S. paradoxus* assemblies and aligned the

sequences of each family and type using MUSCLE. The resulting MSAs were filtered to remove

sequences and sites that had ≥70% gaps. Consensus sequences from those MSAs were

computed using EMBOSS cons version 6.6 (Rice et al., 2000) and the annotation sequence

with the lowest BLASTN E value against the resulting consensus was included in the reference

library. LTRs of the full-length reference elements were manually trimmed using MEGA version

7.0.26 (Kumar et al., 2016). The *S. paradoxus* sequences were complemented with *S.*

cerevisiae internal sequences for Ty2, Ty3, Ty4 and Ty5 (Carr et al., 2012).

Ty insertion allele calling and population structure analysis

Illumina PE 150 libraries for 207 wild *S. paradoxus* diploid strains were downloaded from NCBI

(PRJNA277692, PRJNA324830, PRJNA479851) and trimmed using Trimmomatic version 0.36

(Bolger et al., 2014) with parameters ILLUMINACLIP:combined_adapters.fa:2:30:10

TRAILING:3 SLIDINGWINDOW:4:15 MINLEN:36 and a custom library of Illumina adapter

sequences. Trimmed reads were mapped against the masked version of our *SpB* genome

assembly (strain MSH-604, from the RepeatMasker output) using bwa mem version 0.7.16a-

r1181 (Li, 2013) and sorted using samtools sort version 1.9 (Li, 2011). Duplicates were removed

using Picard RemoveDuplicates version 2.18.5-SNAPSHOT (Broad Institute, n.d.) with

parameter REMOVE_DUPLICATES=true. RetroSeq version 1.5 (Keane et al., 2013) was used

to call Ty insertion alleles from discordant pairs of mapped reads. The -discover command was

used with parameter -eref and our custom reference Ty sequences library. The -call command

was run with parameter -soft. Visual inspection of the resulting calls revealed several instances

of multiple, tightly clustered calls of the same family, likely reflecting a single true positive call. To

minimize false positives, the calls were transformed into MSA coordinates and clustered using

DBSCAN with parameters min_samples=1 eps=500. Resulting clusters were considered true

positives if they contained at least one original call of high-quality (flag 8). This strategy provided

the best balance between false positive rates and false negative rates in an analysis made with

various simulated short-read datasets (data not shown). Insertion allele frequency spectra were

computed within each lineage. Principal component analysis (PCA) was performed on Ty alleles

using the Python package scikit-learn version 0.22.2.post1.

Population structure based on genome-wide SNPs

Trimmed reads from the 207 wild strains were mapped against the unmasked version of our

SpB genome assembly (strain MSH-604) using bwa mem version 0.7.16a-r1181 and sorted

using samtools sort. Duplicates were removed using Picard RemoveDuplicates with parameter

REMOVE_DUPLICATES=true. Variants were called using freebayes version 1.3.1-17-gaa2ace8

(Garrison and Marth, 2012). Variants were filtered using bcftools view version 1.9 (Li, 2011) with

parameter -e 'QUAL<=20' -g ^het, along with the tools vcfallelicprimitives, vcfbreakmulti and

vcffilter with parameter -f 'TYPE = snp' from vcflib version 1.0.0_rc2 (Garrison, 2018). PCA on

genome-wide SNPs was performed using the Python package scikit-allel version 1.2.1 (Miles et

al., 2019), keeping only biallelic SNPs and excluding singletons. From the filtered SNPs of the

207 wild strains, 10000 SNPs were randomly selected and converted to NEXUS format using

vcf2phylip version 2.0 (Ortiz, 2019). A bayesian phylogenetic tree was computed using MrBayes

with model parameters Nst=6 Rates=invgamma. Markov chain Monte Carlo parameters were

set to 1 million generations for two runs of four chains each with 25% burn-in fraction and

temperature parameter of 0.1.

Depth of coverage of wild strains on Ty reference sequences

Trimmed reads from the 207 wild strains were mapped against our custom reference Ty

sequences library using bwa mem version 0.7.16a-r1181 and sorted using samtools sort version

1.9. Only internal sequences extracted from full-length reference sequences were used. Depth

of coverage was extracted using samtools depth. Median depth values of Ty elements were

transformed in $\log_2$ and normalized by the $\log_2$ genome-wide depth computed from the mapped

bases on the genome-wide mappings previously described. Kruskal-Wallis tests were performed

on normalized $\log_2$ depth values for families with full-length elements in natural lineages (Ty1,

Ty3p, Tsu4). Conover post-hoc tests were performed to test for pairwise differences between

lineages using the Python package scikit-posthocs version 0.6.3 (Terpilowski, 2019).

Climatic data

We used the TerraClimate climatic dataset (Abatzoglou et al., 2018) (accessed in October

2019), which incorporates high spatial and temporal resolution climatic interpolation data from

primary and derived climatic variables. For the sampling locations of our 207 wild strains, we

aggregated the 12 climatic variables over the time period from 1958 to 2015 and computed the

mean, standard deviation, minimum and maximum values over time for a given variable variable

at a given location. The resulting dataset was normalized into z-scores and summarized by PCA

using scikit-learn.

Association analysis between Ty CNs, population structure and climatic variables

SNP-based population structure PCAs and climatic variable PCAs were combined in linear

models as predictors of CNs for the three active families Ty1, Ty3p and Tsu4. To avoid spurious

effects arising from log transformation of very sparse coverage, CN values lower than -1.25

were replaced by -1.25 as for MA lines (see below). In each case, PCs explaining a cumulative

sum of 80% of the total variance in the dataset were kept and named gPCs (from PCAs on SNP

data) or ePCs (from PCAs on climatic variables). Ordinary least squares models were fitted

using the Python package statsmodels version 0.10.1 (Seabold and Perktold, 2010). Two types

of models were fitted: OLS using both gPCs and ePCs as predictors, and OLS using only gPCs

as predictors. The quality of the two model types was compared using their AIC values. Strains

were clustered according to their coordinates on gPCs using the AffinityPropagation algorithm

from scikit-learn with parameters max_iter=10000 damping=0.8, and subsequently according to

their coordinates on ePCs using the KMeans algorithm from scikit-learn with the same number

of clusters fitted for gPCs. The inconsistency of each cluster was computed as the standard

deviation of CN for each Ty family.

Nucleotide diversity at synonymous sites

Long-term effective population size for pure lineages was estimated by computing nucleotide

diversity at synonymous sites in protein-coding genes. Only SNPs at four-fold degenerate sites

in protein-coding genes (third positions of codons CTN, GTN, TCN, CCN, ACN, GCN, CGN and

GGN) were considered. Nucleotide diversity was computed using the mean_pairwise_difference

function from scikit-allel, excluding *SpD₂* strain WX21 since it is known to be heterozygous

(Eberlein et al. 2019).

Determination of the optimal NaAsO$_2$ concentration for growth assay

Three strains were chosen at random in each of the lineages *SpA*, *SpB* and *SpC** and two *SpC* strains were chosen at random, totalling 12 strains with the *SpC* strain LL2011_004. Precultures were grown in synthetic complete medium (SC; 2% glucose, 0.174% yeast nitrogen base [without amino acids without ammonium sulfate], 0.5% ammonium sulfate, 0.134% complete drop-out) until they reached a 595 nm optical density (OD$_{595}$) of ~0.5-1.0. Precultures were diluted at an OD$_{595}$ of 0.1 ml^{-1} in SC medium supplemented with 0, 0.4, 0.6, 0.8 and 1 mM NaAsO$_2$ in a single 96-well microplate, with border columns and rows filled with sterile SC medium. OD$_{595}$ was monitored during 60 hours at 15 minutes intervals at 25°C in a Tecan Spark microplate reader (Tecan Group Ltd, Männedorf, Switzerland). OD$_{595}$ values were normalized by subtracting the initial value for each well.

Growth assay in NaAsO$_2$

From the 207 wild strains used for whole genome sequence analyses, 123 strains had glycerol stocks available. Five replicates of each strain were randomly arranged into four arrays of 14x11 positions, totalling 615 replicates with one empty position, with the constraint of having at most three replicates of any given strain within an array. For each array, glycerol stocks of the 123 strains were spotted on YPD plates (1% yeast extract, 2% tryptone, 2% glucose, 2% agar) and incubated at room temperature for three days. Cells from fresh spots were diluted in 200 µl SC medium. OD$_{595}$ was monitored and 500 µl precultures were initiated at 0.08 OD$_{595}$ ml^{-1} and incubated at room temperature for 10 hours. OD$_{595}$ was monitored and precultures were diluted at 0.2 OD595 ml^{-1} in 1 ml SC medium. BRAND™ BRANDplates™ pureGrade™ S 384-well microplates (BRAND GMBH + CO KG, Wertheim, Germany) were filled with 40 µl of SC medium (even column numbers) or SC medium supplemented with 1.6 mM NaAsO$_2$ (odd column numbers). 40 µl of cultures at 0.2 OD$_{595}$ ml^{-1} were added to the plates using an

Eppendorf epMotion 5075 liquid handling platform (Eppendorf, Hamburg, Germany). Replicates

were positioned according to the predefined randomized scheme in side-by-side pairs, yielding

adjacent replicates diluted at 0.1 OD_{595} ml^{-1} in SC and SC with 0.8 mM $NaAsO_2$. Border rows

and columns of the 384-well microplates were filled with 80 µl sterile medium. Microplate lids

were treated with 0.05% Triton X-100 in 20% ethanol for 1 minute and air dried (Brewster,

2003). OD_{595} was monitored during 70 hours at 15 minutes intervals at 25°C in a Tecan Spark

microplate reader.

OD_{595} values were normalized by subtracting the initial value for each well, and values in SC

were subtracted from values in $NaAsO_2$ for each replicate. The area under the curve (AUC) of

the resulting curves was obtained by summing all timepoints. Replicates arising from

precultures that failed to grow were excluded, and strains which had less than 3 replicates left

were excluded. The standard deviation of replicates was computed for each strain, and strains

which fell outside the [-1, 1] z-score range were filtered out to meet the homoscedasticity

requirements of analysis of variance (ANOVA). To assess the significance of lineage effects, a

mixed effects linear model (with strains as random variable) was fitted in R version 3.4.3 (R

Core Team, 2017) using the lmer function from the package lme4 version 1.1-15 (Bates et al.,

2015) and an ANOVA table was produced using the package lmerTest version 3.1-2

(Kuznetsova et al., 2017). Variance components were estimated using the R package VCA

version 1.3.4 (Schützenmeister and Piepho, 2012). Post-hoc Tukey's HSD test was performed

using scikit-posthocs.

Properties of genes neighboring Ty insertions

The analysis of properties of neighboring genes considered Ty-centric immediate adjacency

relationships with protein-coding genes, meaning that a single gene could be represented more

than once if many Ty annotations are immediately adjacent to it. From the whole-genome

MSAs, protein-coding genes adjacent to Ty annotations were retrieved on the basis of synteny

with the *S. cerevisiae* S288c genome. Genes annotated as essential in *S. cerevisiae* were

extracted from YeastMine (accessed in August 2019). Physical interaction data for *S. cerevisiae*

S288c were retrieved from the BioGRID Multi-Validated Physical interaction dataset version

3.5.188 (Oughtred et al., 2019) and the interactome was built using Networkx.

The strength of purifying selection acting on individual genes was assessed at two levels: at the

species-wide level (i.e. including all North American *S. paradoxus* lineages) by fitting a single ω

ratio (non-synonymous to synonymous substitution rates) per gene; and at the lineage level, by

evaluating the relative strength of natural selection in the main pure lineages *SpB* and *SpC*. A

randomly selected subset of 34 strains from the 207 wild strains were chosen. The VCF matrix

from the SNP calling step above was used to derive the DNA sequences of protein-coding

genes in these 34 strains. Neighbor-joining gene trees were produced using the nj function from

the R package ape version 5.0 (Paradis and Schliep, 2019) and R version 3.4.3 (R Core Team,

2017). ω ratios were fitted using codeml from the PAML suite version 4.8 (Yang, 2007) with

parameters seqtype=1 model=0. Selection strength at the lineage level was evaluated using the

RELAX framework from the HyPhy software suite version develop-0d598ad (Wertheim et al.,

2015). The RELAX general descriptive model was fitted using the RELAX-scan script

(https://github.com/veg/hyphy-analyses/tree/master/RELAX-scan). From the model output, the

fitted value of the *k* parameter (describing the strength of selection) was extracted for each

branch of the gene trees. For *SpB* and *SpC*, an average *k* value was computed by selecting the

clade containing all the strains of the corresponding lineage (from the random subset) and

weighting *k* values by the length of the corresponding branch. The difference in selection

intensity between *SpB* and *SpC* was defined as $\Delta k = k_{SpB} - k_{SpC}$. For the three genes with ORF-

disrupting Ty insertions, the RELAX framework was fitted on gene trees comprising all 207 wild

strains.

Mutation accumulation

Crosses BB, BC, BA and BSc were previously published (Charron et al., 2019). CC crosses

were performed following the same methodology. The *ADE2* gene was deleted in haploid *MATa*

hoΔ::kanMX4 or *MATa hoΔ::natMX4* derivatives of the *SpC* strains LL2011_001, LL2011_012

and MSH-587-1 (Charron et al., 2014) using a *hphNT1* cassette that was amplified from plasmid

pFA6a-hphNT1 (Janke et al., 2004) using primers CLOP97-F5 and CLOP97-F6 (Tables S3 and

S4). The three strains were paired with the previously published *SpC* strains LL2011_004 *MATα*

hoΔ::kanMX4 and LL2011_009 *MATα hoΔ::natMX4* to generate three CC crosses by selection

on medium containing 100 µg ml^{-1} geneticin and 10 µg ml^{-1} nourseothricin. For each cross, 64

MA lines were initiated by selecting single colonies from independent mating cultures that were

spotted on selective medium. Diploid hybrids were propagated mitotically on Petri dishes

containing solid YPD medium (1% yeast extract, 2% tryptone, 2% glucose, 2% agar) by

streaking for single colonies. Every three days, a single colony was randomly chosen to

inoculate a new plate by picking the colony that grew the closest to a predefined mark on the

Petri plate. Colonies that had an obvious difference in red pigmentation were excluded to avoid

the propagation of mitochondrial petite mutants. MA lines were propagated for 35 passages

(approx. 770 mitotic generations), keeping glycerol stocks of all the lines every three passages.

Illumina library preparation for MA lines

For each cross, 23 to 40 MA lines were randomly chosen among the subset for which diploidy

was confirmed at T_{ini} and T_{end} (Charron et al., 2019) (Marsit *et al.* in prep.). The glycerol stocks

from passage 1 (T_{ini}) and passage 35 (T_{end}) were streaked on solid YPD medium. Single

48   48

colonies were used to inoculate liquid YPD cultures. DNA was extracted from saturated cultures

using QIAGEN Blood and Tissue kit with modifications for yeast cell wall digestion. DNA was

treated with RNase A and purified on Axygen AxyPrep Mag PCR Clean-up SPRI beads.

Genomic DNA libraries were prepared using the Illumina Nextera Tagmentase and Nextera

index kits following a modified version of the protocol described in (Baym et al., 2015). The

quality of randomly selected libraries was controlled using an Agilent BioAnalyzer 2100

electrophoresis system. Sequencing was performed at the CHU Sainte-Justine and Génome

Québec Integrated Centre for Pediatric Clinical Genomics (Montreal, Canada) on an Illumina

NovaSeq 6000 sequencer with SP flow cells.

Depth of coverage of MA lines on Ty reference sequences

Illumina PE 150 libraries for 372 MA lines and their 13 parental haploid strains (deposited at

NCBI Sequence Read Archive under PRJNA515073) were trimmed using Trimmomatic version

0.33 with parameters ILLUMINACLIP:nextera.fa:6:20:10 MINLEN:40 and a library of Illumina

Nextera adapter sequences. Trimmed reads were mapped against our *SpB* genome assembly

(strain MSH-604) using bwa mem version 0.7.17 and sorted using samtools sort version 1.8.

Duplicates were removed using Picard RemoveDuplicates version 2.18.29-SNAPSHOT with

parameter REMOVE_DUPLICATES=true. We extracted the list of *S. cerevisiae* genes with

paralogs from YeastMine (accessed in December 2018) (Balakrishnan et al., 2012) and

randomly selected 50 genes that didn't figure in this list. The protein sequences of these genes

were used as TBLASTN queries against our *SpB* genome assembly and the best hit loci were

extracted. Depth of coverage over these loci was extracted using samtools depth version 1.9

and depth values were transformed in $\log_2$. Depth values were transformed in $\log_2$ and

normalized by the $\log_2$ genome-wide depth computed from the mapped bases on the genome-

wide mappings.

The trimmed reads of the 372 MA lines and their 13 parental haploid strains were mapped

against our custom reference Ty sequences library using bwa mem version 0.7.16a-r1181 and

sorted using samtools sort version 1.9. Only internal sequences extracted from full-length

reference sequences were used. Median depth values of Ty elements were transformed in $\log_2$

and normalized by the $\log_2$ genome-wide computed from the mapped bases on the genome-

wide mappings. Normalized $\log_2$ depth values for a given cross and Ty family were discarded if

the median was lower than the 10th percentile of the normalized $\log_2$ depth values for non-

paralogous genes in the corresponding cross, thus considering the family absent from the initial

MA genotypes. Similarly, the 10th percentile of normalized $\log_2$ depth values for non-paralogous

genes in all crosses aggregated (-1.25) was used as a lower threshold for depth values of the

parental haploid strains, replacing lower values by -1.25.

Phylogenetic analysis of the MA lines haploid parents

From the mappings of the MA lines parental haploid strains, variants were called using

freebayes. Variants were filtered using bcftools view with parameter -e 'QUAL<=20 -g ^het',

along with the tools vcfallelicprimitives, vcfbreakmulti and vcffilter with parameter -f 'TYPE = snp'

from vcflib. From the filtered SNPs, 10000 SNPs were randomly selected and converted to

NEXUS format using vcf2phylip. A bayesian phylogenetic tree was computed using MrBayes

with model parameters Nst=6 Rates=invgamma Ploidy=haploid. Markov chain Monte Carlo

parameters were set to 1 million generations for two runs of four chains each with 25% burn-in

fraction and temperature parameter of 0.1. From the resulting consensus tree, the evolutionary

divergence between parental strains was measured as the sum of branch lengths connecting

each pair.

Association analysis between Ty ΔCN, evolutionary divergence between parents and initial Ty CN

For each cross, the average ΔCN was analysed in a linear model incorporating parental evolutionary divergence and initial Ty CN of the hybrid genotypes as predictors. For each cross, the Ty CNs of parental haploid strains were summed and used as a measure of the initial Ty CN in all MA lines of the cross. An OLS model was fitted using statsmodels.

Acknowledgements

We thank Hélène Martin and Anna Fijarczyk for contributing to sequencing data analyses and Philippe Després, Anne-Marie Dion-Côté, Marika Drouin, Alexandre Dubé, Clara Dubernard and Anna Fijarczyk for reading and providing useful comments on draft versions of this manuscript.

Competing interests

We declare no competing interest.

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