Effect of Diverse Temporal Recoding of Granule Cells on Pavlovian Eyeblink Conditioning in The Cerebellum

Sang-Yoon Kim* and Woochang Lim†

Institute for Computational Neuroscience and Department of Science Education,
Daegu National University of Education, Daegu 42411, Korea

We consider the Pavlovian eyeblink conditioning (EBC) via repeated presentation of paired conditioned stimulus (tone) and unconditioned stimulus (airpuff), and investigate the effect of diverse temporal recoding of granule (GR) cells on the EBC in a cerebellar network by varying the connection probability p_c from Golgi to GR cells. For an optimal value of p_c^* (= 0.029), individual GR cells exhibit diverse spiking patterns which are well- or ill-matched with the unconditioned stimulus. Then, these diversely-recoded signals via parallel-fibers (PFs) from the GR cells are effectively depressed by the error-teaching signals via climbing fibers (CFs) from the inferior olive. Synaptic weights at well-matched PF-Purkinje cell (PC) synapses of active GR cells are strongly depressed via strong long-term depression (LTD), while practically no LTD occurs at ill-matched PF-PC synapses. This kind of "effective" depression at PF-PC synapses coordinates firings of PCs effectively, which then exert effective inhibitory coordination on cerebellar nucleus neuron [which evokes conditioned response (CR; eyeblink)]. When the learning trial passes a threshold, acquisition of CR begins. In this case, the timing degree $\mathcal{T}_d$ of CR becomes good due to presence of ill-matched spiking group which plays a role of protection barrier for the timing. With further increase in the trial, strength $\mathcal{S}$ of CR (corresponding to the amplitude of eyelid closure) increases due to strong LTD in the well-matched spiking group, while its timing degree $\mathcal{T}_d$ decreases. In this way, the well- and the ill-matched spiking groups play their own roles for the strength and the timing of CR, respectively. Thus, the (overall) learning efficiency degree $\mathcal{L}_e$ (taking into consideration both timing and strength of CR) for the CR increases with learning trial, and eventually it becomes saturated. By changing p_c from p_c^*, we also investigate the effect of diverse spiking patterns of GR cells on the EBC. Plots of both the diversity degree $\mathcal{D}$ of spiking patterns of GR cells and the saturated learning efficiency degree $\mathcal{L}_e^*$ of CR versus p_c are found to form bell-shaped curves with peaks at p_c^*, and they also have a strong correlation. Consequently, the more diverse in temporal recoding of GR cells, the more effective in motor learning for the Pavlovian EBC.

PACS numbers: 87.19.lw, 87.19.lu, 87.19.lv
Keywords: Eyeblink conditioning, Effective motor learning, Diverse recoding, Granule cells, Synaptic plasticity, Cerebellar ring network

I. INTRODUCTION

The cerebellum plays an essential role in precise temporal and spatial motor control for coordinating voluntary movements such as posture, balance, and locomotion, leading to smooth and balanced body movement [1–3]. In addition, it is also involved in higher cognitive functions such as attention, language, and speech [2, 3]. The temporal information of movements (e.g., initiation or termination) is called "timing," while the spatial information of movements (e.g., amplitude or velocity) is called "gain" [4]. The goal of cerebellar motor learning is to perform precise timing and gain control for movements. The cerebellar mechanisms for timing and gain control for eye movements have been studied in the two types of experimental paradigms; (1) timing control for the Pavlovian eyeblink conditioning (EBC) [5–11] and (2) gain control for the vestibulo-ocular reflex and the optokinetic response [1, 12].

Here, we are interested in the Pavlovian EBC [see Fig. 1(a)] which is a representative example for the classical conditioning [13]. An animal (e.g., rabbit, rat, or mouse) receives repeated presentations of paired conditioned stimulus (CS; tone) and (eyeblink-eliciting) unconditioned stimulus (US; airpuff). When the training trial passes a threshold, the animal acquires the ability to elicit eyelid conditioned response (CR; acquisition of learned eyeblink) via learning representation of the time passage between the onsets of CS and US (i.e., the animal becomes conditioned to closes its eye in response to the tone CS with a time delay equal to the inter-stimulus interval (ISI) between the CS and the US onsets). The CRs exhibit two distinct features: (1) gradual acquisition of CR (i.e., CRs are acquired gradually over many training trials of repeated CS-US pairings) [14–18] and (2) adaptive timing of CR (i.e., CRs are well timed such that the time of peak eyelid closure matches well the ISI between the onsets of CS and US) [19–23]. Experimental works on EBC have been done in several animal species such as humans [24], monkeys [15], dogs [14], ferrets [18], rabbits [16], rats [17], and mice [25]. Particularly, since a series of groundbreaking experiments in rabbits [26, 27], EBC in restrained rabbits has served as a good model for

———————
*Electronic address: sykim@icn.re.kr
†Electronic address: wclim@icn.re.kr

motor learning.

Marr [28] and, later, Albus [29] formulated their seminal theory for cerebellar motor learning on the basis of its structure. Particularly, they paid attention to the recurrent network between the granule (GR) and the Golgi (GO) cells as a device of representation of spatial information (i.e., spatial coding). The input spatial patterns, conveyed via the mossy fibers (MFs), become more sparse and less similar to each other via recoding process in the granular layer composed of GR and GO cells. Then, the recoded inputs are fed into the Purkinje cells (PCs) via the parallel fibers (PFs) [i.e., the axons of GR cells]. In addition to the PF recoded signals, the PCs also receive the error-teaching signals through the climbing-fiber (CF) from the inferior olive (IO). The PF-PC synapses are assumed to be the only synapses at which motor learning occurs. Thus, synaptic plasticity may occur at PF-PC synapses (i.e., their synaptic strengths may be potentiated or depressed). Marr in [28] assumes that a Hebbian type of long-term potentiation (LTP) (i.e., increase in synaptic strengths) occurs at the PF-PC synapses when both the PF and the CF signals are conjunctively excited [30, 31]. In contrast to Marr's learning via LTP, Albus in [29] assumes that synaptic strengths at PF-PC synapses are depressed [i.e., an anti-Hebbian type of long-term depression (LTD)] via conjunctive excitation of both the PF and the CF signals. In the case of Albus' learning via LTD, firing activities of PCs get reduced [i.e., they learn when to stop their inhibition (i.e. when to disinhibit) rather than when to fire]. Several later experimental works have provided the support for the Albus' learning via LTD [32–34]. Thus, LTD was established as a unique type of synaptic plasticity for cerebellar motor learning [35–38].

A number of computational works for the EBC have been done. Several artificial models have been proposed for the time-passage representation (i.e., time coding) in the cerebellum [39–44]. However, these artificial models lacked biological plausibility. A realistic cerebellar model, based on many biological properties, has been built by focusing on the recurrent loop between the GR and the GO cells in the granular layer as a time-coding device [45]. Then, the realistic model generated a temporal code based on the population of active GR cells, and also, it was extended to reproduce the experimental results of the Pavlovian EBC [46, 47]. However, the computational mechanism to generate such a temporal code was unclear mainly due to complexity of the realistic model. To understand the computational mechanism for the time coding, a rate-coding model was developed in a simple recurrent inhibitory network, and its dynamics was analyzed in both the numerical and analytical way [48]. This rate-coding model generated a non-recurrent sequence of active neurons, corresponding to representation of a time-passage. Due to randomness in the recurrent connections, individual neurons exhibited random repetition of transitions between the active (bursting) and the inactive (silent) states which were

persistent long-lasting ones. However, this rate-coding model is free of actual time scales. A spiking neural network model (with actual time scales) was built to examine representation of time passage in the cerebellum [49], and a large-scale computational simulation was also performed to reproduce some features of the EBC in the experiments.

However, the effect of diverse temporal recoding of GR cells on the Pavlovian EBC remains to be necessary for more clarification in several dynamical aspects, although much understanding on the representation of time passage in the cerebellum was achieved in previous computational works. As in the case of spatial coding for the optokinetic response [50], we first make dynamical classification of diversely-recoded PF signals completely to clearly understand their conjunction with the error-teaching CF signals. Then, based on such dynamical classification of diverse spiking patterns of GR cells, we may understand clearly synaptic plasticity at PF-PC synapses and the subsequent learning progress. Consequently, understanding on the rate of acquisition and the timing and strength of CR is expected to be so much improved.

To this purpose, we consider a cerebellar spiking ring network for the Pavlovian EBC, and first make a dynamical classification of diverse spiking patterns of GR cells by varying the connection probability p_c from GO to GR cells in the granular layer. The whole population of GR cells is divided into GR clusters. These GR clusters show diverse spiking patterns which are well- or ill-matched relative to the US (i.e., airpuff unconditioned stimulus) signal. Each spiking pattern is characterized in terms of the "conjunction" index, representing the "matching" degree between the spiking pattern and the US. To quantify the degree of diverse recoding of GR cells, we introduce the diversity degree $\mathcal{D}$, given by the relative standard deviation in the distribution of conjunction indices of all the spiking patterns in the GR clusters. We mainly focus on an optimal case of $p_c^* = 0.029$ where the spiking patterns of GR clusters are the most diverse. In this optimal case, $\mathcal{D}^* \simeq 1.842$ which is a quantitative measure for diverse temporal recoding of GR cells in the granular layer. Dynamical origin of these diverse spiking patterns of GR cells is also investigated. Diverse total synaptic inputs (including both the excitatory inputs via MFs and the inhibitory inputs from the pre-synaptic GO cells) into the GR clusters lead to generation of diverse spiking patterns (i.e. outputs) in the GR clusters.

Based on dynamical classification of diverse spiking patterns in the GR clusters, we next make an intensive investigation on the effect of diverse temporal recoding of GR cells on synaptic plasticity at PF-PC synapses and subsequent learning process by employing a refined rule for synaptic plasticity (constructed from the experimental result in [51]). PCs (corresponding to the cerebellar output) receive both the diversely-recoded PF signals from the GR cells and the error-teaching CF signals from the IO neuron; the CF signals are well-matched with the US signal. In this case, CF signals can be regarded as "in-

structors," while PF signals may be considered as "students" [50]. Then, well-matched PF student signals are strongly depressed (i.e., their synaptic weights at PF-PC synapse are greatly decreased) via strong LTD by the CF instructor signals. On the other hand, practically no LTD occurs for the ill-matched PF student signals because most of them have no associations with the US signals which are strongly localized in the middle of each trial. In this way, at the PF-PC synapses, the student PF signals are effectively depressed by the instructor CF signals.

During learning trials with repeated presentation of CS-US pairs, the "effective" depression at PF-PC synapses coordinates activations of PCs effectively, which then exert effective inhibitory coordination on the cerebellar nucleus (CN) neuron [which evokes CR (i.e., learned eyeblink)]. In the optimal case of $p_c^* = 0.029$, when passing the 141th threshold trial, acquisition of CR starts. In this optimal case, the timing degree $\mathcal{T}_d$ for the CR becomes good because of presence of ill-matched spiking group which plays a role of protection barrier for the timing. With further increase in the trial, strength $\mathcal{S}$ of CR [denoting the amplitude of eyelid closure (measured by the electromyography (EMG))] increases due to strong LTD in the well-matched spiking group, while its timing degree $\mathcal{T}_d$ decreases. In this way, the well- and the ill-matched spiking groups play their own roles for the strength and the timing of CR, respectively. Thus, the (overall) learning efficiency degree $\mathcal{L}_e$ (considering both timing and strength of CR) for the CR increases with the training trial, and eventually it gets saturated.

Saturation in learning progress may be clearly shown in the inferior olive (IO) system. During the learning trials, the IO neuron receives both the excitatory US signal and the inhibitory signal from the CN neuron. We introduce the learning progress degree $\mathcal{L}_p$, given by the ratio of the time-averaged inhibitory input from the CN neuron to the time-averaged excitatory input of the US signal. With increasing trial from the threshold, the time-averaged inhibitory input (from the CN neuron) increases (i.e., $\mathcal{L}_p$ increases), and converges to the constant time-averaged excitatory input (through the US signal). Thus, at about the 250th trial, the learning progress degree becomes saturated at $\mathcal{L}_p = 1$. At this saturated stage, the time-averaged excitatory and inhibitory inputs to the IO neuron get balanced, and we obtain the saturated learning efficiency degree $\mathcal{L}_e^*$ ($\simeq 11.19$) of the CR in the CN.

Finally, we vary p_c from p_c^*, and investigate the effect of diverse temporal recoding of GR cells on the Pavlovian EBC. The distribution of both the diversity degree $\mathcal{D}$ of spiking patterns of GR cells and the saturated learning efficiency degree $\mathcal{L}_e^*$ of CR versus p_c are found to form bell-shaped curves with peaks at p_c^*. Moreover, both $\mathcal{D}$ and $\mathcal{L}_e^*$ have a strong correlation. As a result, the more diverse in temporal recoding of GR cells, the more effective in motor learning for the Pavlovian EBC.

This paper is organized as follows. In Sec. II, we describe the cerebellar ring network for the Pavlovian EBC.

FIG. 1: Pavlovian eyeblink conditioning (EBC). (a) Eyelid conditioned response (CR) (i.e., learned eyeblink) via repeated presentation of paired CS (conditioned stimulus) and US (unconditioned stimulus). Firing rates of (b1) transient conditioned stimulus (TCS) for resetting and (b2) sustained conditioned stimulus (SCS) (EBC signal). (c) Firing rate of transient unconditioned stimulus (US) for timing (eliciting unconditioned response).

The governing equations for the population dynamics in the ring network are also presented, together with a refined rule for the synaptic plasticity at the PF-PC synapses. Then, in the main Sec. III, we investigate the effect of diverse temporal recoding of GR cells on motor learning for the Pavlovian EBC by changing p_c. Finally, we give summary and discussion in Sec. IV.

II. CEREBELLAR RING NETWORK FOR THE PAVLOVIAN EYEBLINK CONDITIONING

In this section, we describe our cerebellar ring network for the Pavlovian EBC. Figure 1(a) shows the Pavlovian EBC. During the training trials, repeated presentations of paired tone CS and delayed airpuff US are made to an animal (e.g., rabbit, rat, or mouse). As the training trial passes a threshold, the animal acquires the ability to elicit eyelid CR (i.e., acquisition of learned eyeblink) through learning representation of the time passage between the CS and the US onsets. Accordingly, the animal gets conditioned to closes its eye in response to the tone CS with a time delay equal to the ISI between the onsets of CS and US.

FIG. 2: Cerebellar ring network for the EBC. (a) Box diagram for the cerebellar network. Lines with triangles and circles represent excitatory and inhibitory synapses, respectively. GR (granule cell), GO (Golgi cell), and PF (parallel fiber) in the granular layer, PC (Purkinje cell) and BC (basket cell) in the Purkinje-molecular layer, and other parts for CN (cerebellar nuclei), IO(inferior olive), MF (mossy fiber), and CF (climbing fiber). (b) Schematic diagram for granular-layer ring network with concentric inner GR and outer GO rings. Numbers represent granular layer zones (bounded by dotted lines) for $N_C = 32$. In each Ith zone ($I = 1, \cdots, N_C$), there exists the Ith GR cluster on the inner GR ring. Each GR cluster consists of GR cells (solid circles), and it is bounded by 4 glomeruli (stars). On the outer GO ring in the Ith zone, there exists the Ith GO cell (diamonds). (c) Schematic diagram for Purkinje-molecular-layer ring network with concentric inner PC and outer BC rings. Numbers represent the Purkinje-molecular-layer zones (bounded by dotted lines) for $N_{\rm PC} = 16$. In each Jth zone, there exist the Jth PC (solid circle) on the inner PC ring and the Jth BC (solid triangle) on the outer BC ring.

A. Conditioned Stimulus and Unconditioned Stimulus

We first consider the CS for the EBC signal. When the CS is a tone, the pontine nucleus in the pons receives the auditory information, and then it sends the "context" signal for the EBC via MFs to both the GR cells and the CN neuron. There are a transient CS for resetting and a sustained CS (representing the tone) [49]. Each step ($0 < t < 2000$ msec) for EBC learning consists of the trial stage ($0 < t < 1000$ msec) and the break stage ($1000 < t < 2000$ msec); t denotes the time. In the trial stage, the transient CS is modeled in terms of strong and brief Poisson spike trains of 200 Hz for $0 < t < 5$ msec and the subsequent background Poisson spike trains of 5 Hz for $5 < t < 1000$ msec. On the other hand, the sustained CS is modeled in terms of Poisson spike trains of 30 Hz for $0 < t < 1000$ msec. In the following break stage, the CS is modeled in terms of the background Poisson spike trains of 5 Hz for $1000 < t < 2000$ msec. The firing rates $f_{\rm TCS}(t)$ and $f_{\rm SCS}(t)$ of the transient CS and the sustained CS are shown in Figs. 1(b1) and 1(b2), respectively. These figures also show the preparatory step for $-500 < t < 0$ msec where the CS is modeled in terms of the background Poisson spike trains of 5 Hz; this preparatory step precedes just the 1st step for the EBC learning.

Next, we consider the US for the desired timing signal. When an airpuff US is delivered to the cornea of an eye, sensory information is carried to the sensory trigeminal nucleus (which extends through the whole of midbrain, pons, and medulla and into the high cervical spinal cord). Then, the trigeminal nucleus also sends the desired timing signal to the IO. In the trial stage ($0 < t < 1000$ msec), the US (eliciting unconditioned response) is modeled in terms of the strong and brief Poisson spike trains of 25 Hz for a short interval in the middle of the trial stage, $t^* - \Delta t < t < t^* + \Delta t$ ($t^* = 500$ msec and $\Delta t = 5$ msec) [49]. The firing rate $f_{\rm US}(t)$ of the US is shown in Fig. 1(c).

B. Framework for Cerebellar Ring Network

As in [50], we consider the box diagram for the cerebellar network and our cerebellar network for the EBC. As in the famous small-world ring network [52, 53], we employ a one-dimensional ring network with a simple archi-

tecture, in contrast to the two-dimensional square-lattice network [4, 49]. Our ring network has advantage for computational and analytical efficiency, and its visual representation may also be easily made.

Figure 2(a) shows the box diagram. The granular layer, corresponding to the input layer of the cerebellar cortex, is composed of the excitatory GR cells and the inhibitory GO cells. On the other hand, the Purkinje-molecular layer, corresponding to the output layer of the cerebellar cortex, consist of the inhibitory PCs and the inhibitory BCs (basket cells). The MF context signals for the EBC are fed from the pontine nucleus in the pons to the GR cells; each GR cell receives two transient and two sustained CS signals via four MFs (i.e., two pairs of transient and sustained CS signals are fed into each GR cell). Diverse temporal recoding is made in the granular layer via inhibitory coordination of GO cells on GR cells. Then, these diversely-recoded outputs are fed via PFs to the PCs and the BCs in the Purkinje-molecular layer.

The PCs receive another excitatory error-teaching CF signals from the IO, along with the inhibitory inputs from the BCs. Then, depending on the type of PF signals (i.e., well- or ill-matched PF signals), diverse PF (student) signals are effectively depressed by the error-teaching (instructor) CF signals. Such "effective" depression at the PF-PC synapses coordinates firings of PCs effectively, which then exert effective inhibitory coordination on the CN neuron. The CN neuron also receives two excitatory signals; one transient and one sustained CS signals via MFs. In the earlier trials, the CN neuron can not fire, due to strong inhibition from the PCs. As the learning trial passes a threshold, the CN neuron starts firing, and then it exerts excitatory projections onto the eyeblink pre-motoneurons in the midbrain which then supply motor commands to eyeblink motoneurons. Thus, acquisition of CR begins (i.e., acquisition of learned eyeblink starts). This CN neuron also provides inhibitory inputs to the IO neuron which also receives the excitatory signals for the desired timing from the trigeminal nucleus. Then, the IO neuron supplies excitatory error-teaching CF signals to the PCs.

Figure 2(b) shows a schematic diagram for the granular-layer ring network with concentric inner GR and outer GO rings. Numbers represent granular-layer zones (bounded by dotted lines); the numbers 1, 2, $\cdots$, and N_C represent the 1st, the 2nd, $\cdots$, and the N_Cth granular-layer zones, respectively. Thus, the total number of granular-layer zones is N_C; Fig. 2(b) shows an example for $N_C = 32$. In each Ith zone ($I = 1, \cdots, N_C$), there exists the Ith GR cluster on the inner GR ring. Each GR cluster consists of N_{GR} excitatory GR cells (solid circles). Then, location of each GR cell may be denoted by the two indices (I, i) which represent the ith GR cell in the Ith GR cluster, where $i = 1, \cdots, N_{\mathrm{GR}}$. Here, we consider the case of $N_C = 2^{10}$ and $N_{\mathrm{GR}} = 50$, and thus the total number of GR cells is 51,200. In this case, the Ith zone covers the angular range of $(I - 1) \theta^*_{\mathrm{GR}} < \theta < I \theta^*_{\mathrm{GR}}$ ($\theta^*_{\mathrm{GR}} = 0.35°$). On the outer GO ring in each Ith zone,

there exists the Ith inhibitory GO cell (diamond), and thus the total number of GO cells is N_C.

We note that each GR cluster is bounded by 4 glomeruli (corresponding to the axon terminals of the MFs) (stars) at both boundaries of the GR cluster; at each boundary, a pair of glomeruli (upper and lower ones) exist. GR cells within each GR cluster share the same inhibitory and excitatory synaptic inputs through their dendrites which contact the four glomeruli at both ends of the GR cluster. Each glomerulus receives inhibitory inputs from nearby 81 (clockwise side: 41 and counter-clockwise side: 40) GO cells with a random connection probability p_c (= 0.029). Hence, on average, about 2 GO cell axons innervate each glomerulus. Thus, each GR cell receives about 9 inhibitory inputs through 4 dendrites which synaptically contact the glomeruli at both boundaries. In this way, each GR cell in the GR cluster shares the same inhibitory synaptic inputs from nearby GO cells through the intermediate glomeruli at both ends.

In addition, each GR cell shares the same four excitatory inputs via the four glomeruli at both boundaries, because a glomerulus receives an excitatory MF input. We note that transient CS signals are supplied via the two upper glomeruli, while sustained CS signals are fed through the two lower glomeruli. Here, we take into consideration stochastic variability of synaptic transmission from a glomerulus to GR cells, and supply independent Poisson spike trains with the same firing rate to each GR cell for the excitatory MF signals. In this GR-GO feedback system, each GO cell receives excitatory synaptic inputs through PFs from GR cells in the nearby 49 (central side: 1, clockwise side: 24 and counter-clockwise side: 24) GR clusters with a random connection probability 0.1. Hence, 245 PFs (i.e. GR cell axons) innervate a GO cell.

Figure 2(c) shows a schematic diagram for the Purkinje-molecular-layer ring network with concentric inner PC and outer BC rings. Numbers represent the Purkinje-molecular-layer zones (bounded by dotted lines). In each Jth zone ($J = 1, \cdots, N_{\mathrm{PC}}$), there exist the Jth PC (solid circles) on the inner PC ring and the Jth BC (solid triangles) on the outer BC ring. Here, we consider the case of $N_{\mathrm{PC}} = 16$, and thus the total numbers of PC and BC are 16, respectively. In this case, each Jth ($J = 1, \cdots, N_{\mathrm{PC}}$) zone covers the angular range of $(J-1) \theta^*_{\mathrm{PC}} < \theta < J \theta^*_{\mathrm{PC}}$, where $\theta^*_{\mathrm{PC}} \simeq 22.5°$ (corresponding to about 64 zones in the granular-layer ring network). We note that diversely-recoded PFs innervate PCs and BCs. Each PC (BC) in the Jth Purkinje-molecular-layer zone receives excitatory synaptic inputs via PFs from all the GR cells in the 288 GR clusters (clockwise side: 144 and counter-clockwise side: 144 when starting from the angle $\theta = (J-1) \theta^*_{\mathrm{PC}}$ in the granular-layer ring network). Thus, each PC (BC) is synaptically connected via PFs to the 14,400 GR cells (which corresponds to about 28 % of the total GR cells). In addition to the PF signals, each PC also receives inhibitory inputs from nearby 3 BCs (central side: 1, clockwise side: 1 and counter-clockwise

side: 1) and excitatory error-teaching CF signal from the IO.

Here, for simplicity, we consider just one CN neuron and one IO neuron. Both excitatory inputs (corresponding to one transient and one sustained CS signals) via 2 MFs and inhibitory inputs from all the 16 PCs are fed into the CN neuron. Then, the CN neuron provides excitatory input to the eyeblink pre-motoneurons in the midbrain and also supplies inhibitory input to the IO neuron. One additional excitatory desired timing signal from the trigeminal nucleus is also fed into the IO neuron. Then, through integration of both excitatory and inhibitory inputs, the IO neuron provides excitatory error-teaching CF signals to the PCs.

C. Elements of The Cerebellar Ring Network

We choose leaky integrate-and-fire (LIF) neuron models as elements of the cerebellar ring network [54]. Here, the LIF neuron models incorporate additional afterhyperpolarization (AHP) currents that determine refractory periods. This LIF neuron model is one of the simplest spiking neuron models. Due to its simplicity, it can be easily analyzed and simulated. Hence, it has been very popularly used as a neuron model.

Dynamics of states of individual neurons in the X population are governed by the following equations:

$$C_X \frac{dv_i^{(X)}}{dt} = -I_{L,i}^{(X)} - I_{AHP,i}^{(X)} + I_{ext}^{(X)} - I_{syn,i}^{(X)}, \quad i = 1, \cdots, N_X, \quad (1)$$

where N_X is the total number of neurons in the X population, X = GR and GO in the granular layer, X = PC and BC in the Purkinje-molecular layer, and in the other parts X = CN and IO. The state of the ith neuron in the X population at a time t (msec) is characterized by its membrane potential $v_i^{(X)}$ (mV), and C_X (pF) denotes the membrane capacitance of the cells in the X population. The time-evolution of $v_i^{(X)}(t)$ is governed by 4 types of currents (pA) into the ith neuron in the X population; the leakage current $I_{L,i}^{(X)}$, the AHP current $I_{AHP,i}^{(X)}$, the external constant current $I_{ext}^{(X)}$ (independent of i), and the synaptic current $I_{syn,i}^{(X)}$.

We consider a single LIF neuron model [without the AHP current and the synaptic current in Eq. (1)] which describes a simple parallel resistor-capacitor circuit. Here, the leakage term is due to the resistor and the integration of the external current is due to the capacitor which is in parallel to the resistor. Thus, in Eq. (1), the 1st type of leakage current $I_{L,i}^{(X)}$ for the ith neuron in the X population is given by:

$$I_{L,i}^{(X)} = g_L^{(X)}(v_i^{(X)} - V_L^{(X)}), \quad (2)$$

where $g_L^{(X)}$ and $V_L^{(X)}$ are conductance (nS) and reversal potential for the leakage current, respectively.

The ith neuron fires a spike when its membrane potential $v_i^{(X)}$ reaches a threshold $v_{th}^{(X)}$ at a time $t_{f,i}^{(X)}$. Then, the 2nd type of AHP current $I_{AHP,i}^{(X)}$ follows after firing (i.e., $t \geq t_{f,i}^{(X)}$):

$$I_{AHP,i}^{(X)} = g_{AHP}^{(X)}(t) \, (v_i^{(X)} - V_{AHP}^{(X)}) \quad \text{for } t \geq t_{f,i}^{(X)}, \quad (3)$$

where $V_{AHP}^{(X)}$ is the reversal potential for the AHP current. The conductance $g_{AHP}^{(X)}(t)$ is given by an exponential-decay function:

$$g_{AHP}^{(X)}(t) = \bar{g}_{AHP}^{(X)} \, e^{-(t - t_{f,i}^{(X)})/\tau_{AHP}^{(X)}}, \quad (4)$$

where $\bar{g}_{AHP}^{(X)}$ and $\tau_{AHP}^{(X)}$ are the maximum conductance and the decay time constant for the AHP current. As $\tau_{AHP}^{(X)}$ increases, the refractory period becomes longer.

The 3rd type of external constant current $I_{ext}^{(X)}$ for spontaneous firing is provided to only PCs because of their high spontaneous firing rate [55, 56]. In Appendix, Table I shows the parameter values for the capacitance C_X, the leakage current $I_L^{(X)}$, the AHP current $I_{AHP}^{(X)}$, and the external constant current $I_{ext}^{(X)}$. These values are adopted from physiological data [49].

D. Three Kinds of Synaptic Currents

Here, we are concerned about the 4th type of synaptic current $I_{syn,i}^{(X)}$ into the ith neuron in the X population in Eq. (1). It is composed of the following 3 kinds of synaptic currents:

$$I_{syn,i}^{(X)} = I_{\text{AMPA},i}^{(X,Y)} + I_{\text{NMDA},i}^{(X,Y)} + I_{\text{GABA},i}^{(X,Z)}. \quad (5)$$

Here, $I_{\text{AMPA},i}^{(X,Y)}$ and $I_{\text{NMDA},i}^{(X,Y)}$ are the excitatory AMPA (α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid) receptor-mediated and NMDA (N-methyl-D-aspartate) receptor-mediated currents from the pre-synaptic source Y population to the post-synaptic ith neuron in the target X population. In contrast, $I_{\text{GABA},i}^{(X,Z)}$ is the inhibitory GABA$_A$ (γ-aminobutyric acid type A) receptor-mediated current from the pre-synaptic source Z population to the post-synaptic ith neuron in the target X population.

As in the case of the AHP current, the R (= AMPA, NMDA, or GABA) receptor-mediated synaptic current $I_{R,i}^{(T,S)}$ from the pre-synaptic source S population to the ith post-synaptic neuron in the target T population is given by:

$$I_{R,i}^{(T,S)} = g_{R,i}^{(T,S)}(t) \, (v_i^{(T)} - V_R^{(S)}), \quad (6)$$

where $g_{(R,i)}^{(T,S)}(t)$ and $V_R^{(S)}$ are synaptic conductance and synaptic reversal potential (determined by the type of

the pre-synaptic source S population), respectively. We obtain the synaptic conductance $g_{R,i}^{(T,S)}(t)$ from:

$$g_{R,i}^{(T,S)}(t) = \bar{g}_R^{(T)} \sum_{j=1}^{N_S} J_{ij}^{(T,S)} \, w_{ij}^{(T,S)} \, s_j^{(T,S)}(t), \qquad (7)$$

where $\bar{g}_R^{(T)}$ and $J_{ij}^{(T,S)}$ are the maximum conductance and the synaptic weight of the synapse from the jth pre-synaptic neuron in the source S population to the ith post-synaptic neuron in the target T population, respectively. The inter-population synaptic connection from the source S population (with N_s neurons) to the target T population is given in terms of the connection weight matrix $W^{(T,S)}$ ($= \{w_{ij}^{(T,S)}\}$) where $w_{ij}^{(T,S)} = 1$ if the jth neuron in the source S population is pre-synaptic to the ith neuron in the target T population; otherwise $w_{ij}^{(T,S)} = 0$.

The post-synaptic ion channels are opened because of the binding of neurotransmitters (emitted from the source S population) to receptors in the target T population. The fraction of open ion channels at time t is represented by $s^{(T,S)}$. The time course of $s_j^{(T,S)}(t)$ of the jth neuron in the source S population is given by a sum of exponential-decay functions $E_R^{(T,S)}(t - t_f^{(j)})$:

$$s_j^{(T,S)}(t) = \sum_{f=1}^{F_j^{(S)}} E_R^{(T,S)}(t - t_f^{(j)}), \qquad (8)$$

where $t_f^{(j)}$ and $F_j^{(S)}$ are the fth spike time and the total number of spikes of the jth neuron in the source S population, respectively. The exponential-decay function $E_R^{(T,S)}(t)$ (which corresponds to contribution of a pre-synaptic spike occurring at $t = 0$ in the absence of synaptic delay) is given by:

$$E_R^{(T,S)}(t) = e^{-t/\tau_R^{(T)}} \Theta(t) \quad \text{or} \qquad (9a)$$

$$= (A_1 e^{-t/\tau_{R,1}^{(T)}} + A_2 e^{-t/\tau_{R,2}^{(T)}})\Theta(t), \quad (9b)$$

where $\Theta(t)$ is the Heaviside step function: $\Theta(t) = 1$ for $t \geq 0$ and 0 for $t < 0$. Depending on the source and the target populations, $E_R^{(T,S)}(t)$ may be a type-1 single exponential-decay function of Eq. (9a) or a type-2 dual exponential-decay function of Eq. (9b). In the type-1 case, there exists one synaptic decay time constant $\tau_R^{(T)}$

(determined by the receptor on the post-synaptic target T population), while in the type-2 case, two synaptic decay time constants, $\tau_{R,1}^{(T)}$ and $\tau_{R,2}^{(T)}$ appear. In most cases, the type-1 single exponential-decay function of Eq. (9a) appears, except for the two synaptic currents $I_{\text{GABA}}^{(\text{GR,GO})}$ and $I_{\text{NMDA}}^{(\text{GO,GR})}$.

In Appendix, Table II shows the parameter values for the maximum conductance $\bar{g}_R^{(T)}$, the synaptic weight $J_{ij}^{(T,S)}$, the synaptic reversal potential $V_R^{(S)}$, the synaptic decay time constant $\tau_R^{(T)}$, and the amplitudes A_1 and A_2 for the type-2 exponential-decay function in the granular layer, the Purkinje-molecular layer, and the other parts for the CN and IO, respectively. These values are adopted from physiological data [49].

E. Refined Rule for Synaptic Plasticity

As in [50], we employ a rule for synaptic plasticity, based on the experimental result in [51]. This rule is a refined one for the LTD in comparison with the rule used in [4, 49], the details of which will be explained below.

The coupling strength of the synapse from the pre-synaptic neuron j in the source S population to the post-synaptic neuron i in the target T population is $J_{ij}^{(T,S)}$. Initial synaptic strengths for $J_{ij}^{(T,S)}$ are given in Table II. In this work, we assume that learning occurs only at the PF-PC synapses. Hence, only the synaptic strengths $J_{ij}^{(\text{PC,PF})}$ of PF-PC synapses may be modifiable (i.e., they are depressed or potentiated), while synaptic strengths of all the other synapses are static. [Here, the index j for the PFs corresponds to the two indices (M,m) for GR cells representing the mth ($1 \leq m \leq 50$) cell in the Mth ($1 \leq M \leq 2^{10}$) GR cluster.] Synaptic plasticity at PF-PC synapses have been so much studied in many experimental [32–35, 51, 57–65] and computational [4, 29, 49, 66–72] works.

As the time t is increased, synaptic strength $J_{ij}^{(\text{PC,PF})}(t)$ for each PF-PC synapse is updated with the following multiplicative rule (depending on states) [50, 51]:

$$J_{ij}^{(\text{PC,PF})}(t) \rightarrow J_{ij}^{(\text{PC,PF})}(t) + \Delta J_{ij}^{(\text{PC,PF})}(t), \qquad (10)$$

where

$$\Delta J_{ij}^{(\text{PC,PF})}(t) = \Delta\text{LTD}_{ij}^{(1)} + \Delta\text{LTD}_{ij}^{(2)} + \Delta\text{LTP}_{ij}, \qquad (11)$$

$$\Delta\text{LTD}_{ij}^{(1)} = -\delta_{LTD} \cdot J_{ij}^{(\text{PC,PF})}(t) \cdot CF_i(t) \cdot \sum_{\Delta t=0}^{\Delta t_r^*} \Delta J_{LTD}(\Delta t), \qquad (12)$$

$$\Delta\text{LTD}_{ij}^{(2)} = -\delta_{LTD} \cdot J_{ij}^{(\text{PC,PF})}(t) \cdot [1 - CF_i(t)] \cdot PF_{ij}(t) \cdot D_i(t) \cdot \sum_{\Delta t=0}^{\Delta t_l^*} \Delta J_{LTD}(\Delta t), \qquad (13)$$

$$\Delta\text{LTP}_{ij} = \delta_{LTP} \cdot [J_0^{(\text{PC,PF})} - J_{ij}^{(\text{PC,PF})}(t)] \cdot [1 - CF_i(t)] \cdot PF_{ij}(t) \cdot [1 - D_i(t)]. \qquad (14)$$

Here, $J_0^{(\text{PC,PF})}$ is the initial value (=0.006) for the synaptic strength of PF-PC synapses. Synaptic modification (LTD or LTP) occurs, depending on the relative time difference Δt [= t_{CF} (CF activation time) - t_{PF} (PF activation time)] between the spiking times of the error-teaching instructor CF and the diversely-recoded student PF. In Eqs. (12)-(14), $CF_i(t)$ denotes a spike train of the CF signal coming into the ith PC. When $CF_i(t)$ activates at a time t, $CF_i(t) = 1$; otherwise, $CF_i(t) = 0$. This instructor CF activation gives rise to LTD at PF-PC synapses in conjunction with earlier ($\Delta t > 0$) student PF activations in the range of $t_{\text{CF}} - \Delta t_r^* < t_{\text{PF}} < t_{\text{CF}}$ ($\Delta t_r^* \simeq 277.5$ msec), which corresponds to the major LTD in Eq. (12).

We next consider the case of $CF_i(t) = 0$, which corresponds to Eqs. (13) and (14). Here, $PF_{ij}(t)$ denotes a spike train of the PF signal from the jth pre-synaptic GR cell to the ith post-synaptic PC. When $PF_{ij}(t)$ activates at time t, $PF_{ij}(t) = 1$; otherwise, $PF_{ij}(t) = 0$. In the case of $PF_{ij}(t) = 1$, PF firing may cause LTD or LTP, depending on the presence of earlier CF firings in an effective range. If CF firings exist in the range of $t_{\text{PF}} + \Delta t_l^* < t_{\text{CF}} < t_{\text{PF}}$ ($\Delta t_l^* \simeq -117.5$ msec), $D_i(t) = 1$; otherwise $D_i(t) = 0$. When both $PF_{ij}(t) = 1$ and $D_i(t) = 1$, the PF activation causes another LTD at PF-PC synapses in conjunction with earlier ($\Delta t < 0$) CF activations [see Eq. (13)]. The probability for occurrence of earlier CF firings within the effective range is very low because mean firing rates of the CF signals (corresponding to output firings of individual IO neurons) are ~ 1.5 Hz [73, 74]. Hence, this 2nd type of LTD is a minor one. In contrast, in the case of $D_i(t) = 0$ (i.e., absence of earlier associated CF firings), LTP occurs because of the PF firing alone [see Eq. (14)]. The update rate δ_{LTD} for LTD in Eqs. (12) and (13) is 0.005, while the update rate δ_{LTP} for LTP in Eqs. (14) is 0.0005 (=$\delta_{LTD}/10$) [4].

In the case of LTD in Eqs. (12) and (13), the synaptic modification $\Delta J_{LTD}(\Delta t)$ changes depending on the relative time difference Δt (= $t_{\text{CF}} - t_{\text{PF}}$). We use the following time window for the synaptic modification $\Delta J_{LTD}(\Delta t)$ [50, 51]:

$$\Delta J_{LTD}(\Delta t) = A + B \cdot e^{-(\Delta t - t_0)^2/\sigma^2}, \qquad (15)$$

where $A = -0.12$, $B = 0.4$, $t_0 = 80$, and $\sigma = 180$. Figure 3 shows the time window for $\Delta J_{LTD}(\Delta t)$. As shown well in Fig. 3, LTD occurs in an effective range of $\Delta t_l^* < \Delta t < \Delta t_r^*$. We note that a peak appears at $t_0 = 80$ msec, and hence peak LTD takes place when PF firing precedes CF firing by 80 msec. A CF firing gives rise to LTD in association with earlier PF firings in the region hatched with horizontal lines ($0 < \Delta t < \Delta t_r^*$), and it also causes to another LTD in conjunction with later PF firings in the region hatched with vertical lines ($\Delta t_l^* < \Delta t < 0$). The effect of CF firing on earlier PF firings is much larger than that on later PF firings. However, outside the effective range (i.e., $\Delta t > \Delta t_r^*$ or $< \Delta t_l^*$), PF firings alone results in occurrence of LTP, because of absence of effectively associated CF firings.

Our refined rule for synaptic plasticity has the following advantages for the ΔLTD in comparison with that in [4, 49]. Our rule is based on the experimental result in [51]. In the presence of a CF firing, a major LTD ($\Delta\text{LTD}^{(1)}$) occurs in conjunction with earlier PF firings in the range of $t_{\text{CF}} - \Delta t_r^* < t_{\text{PF}} < t_{\text{CF}}$ ($\Delta t_r^* \simeq 277.5$ msec), while a minor LTD ($\Delta\text{LTD}^{(2)}$) takes place in conjunction with later PF firings in the range of $t_{\text{CF}} < t_{\text{PF}} < t_{\text{CF}} - \Delta t_l^*$ ($\Delta t_l^* \simeq -117.5$ msec). The magnitude of LTD varies depending on Δt (= t_{CF} - t_{PF}); a peak LTD occurs when $\Delta t = 80$ msec. In contrast, the rule in [4, 49] considers only the major LTD in association with earlier PF firings in the range of $t_{\text{CF}} - 50 < t_{\text{PF}} < t_{\text{CF}}$, the magnitude of major LTD is equal, independently of Δt, and minor LTD in conjunction with later PF firings is not considered. Outside the effective range of LTD, PF firings alone lead to LTP in both rules.

F. Numerical Method

Numerical integration of the governing Eq. (1) for the time-evolution of states of individual neurons, together with the update rule for synaptic plasticity of Eq. (10), is made by using the 2nd-order Runge-Kutta method with the time step 1 msec. In each realization, we choose random initial points $v_i^{(X)}(0)$ for the ith neuron in the X population with uniform probability in the range of

FIG. 3: Time window for the LTD at the PF-PC synapse. Plot of synaptic modification $\Delta J_{LTD}(\Delta t)$ for LTD versus Δt.

$v_i^{(X)}(0) \in (V_L^{(X)} - 5.0, V_L^{(X)} + 5.0)$; the values of $V_L^{(X)}$ are given in Table I.

III. EFFECT OF DIVERSE TEMPORAL RECODING IN GR CLUSTERS ON MOTOR LEARNING FOR THE PAVLOVIAN EYEBLINK CONDITIONING

In this section, we investigate the effect of diverse temporal recoding of GR cells on motor learning for the EBC by changing the connection probability p_c from the GO to the GR cells. We mainly focus on an optimal case of $p_c^* = 0.029$ where the spiking patterns of GR clusters are the most diverse. In this case, we first make dynamical classification of diverse spiking patterns of the GR clusters. Next, we study the effect of diverse spiking patterns on the synaptic plasticity at the PF-PC synapses and the subsequent learning process in the PC-CN-IO system. Finally, we change p_c from the optimal value p_c^*, and investigate dependence of the diversity degree $\mathcal{D}$ of spiking patterns and the saturated learning efficiency degree $\mathcal{L}_e^*$ for the CR on p_c. Both $\mathcal{D}$ and $\mathcal{L}_e^*$ are found to form bell-shaped curves with peaks at p_c^*, and they have strong correlation with the Pearson's coefficient $r \simeq 0.9982$ [75]. Consequently, the more diverse in temporal recoding in the GR clusters, the more effective in the motor learning for the EBC.

A. Collective Firing Activity in The Whole Population of GR Cells

Temporal recoding process is performed in the granular layer (corresponding to the input layer of the cerebellar cortex), composed of GR and GO cells (see Fig. 2). GR cells (principal output cells in the granular layer) receive excitatory context signals for the EBC via the MFs [see Figs. 1(b1) and 1(b2)] and make diverse recoding of context signals through receiving effective inhibitory coordination of GO cells. Thus, diversely recoded signals are fed into the PCs (principal output cells in the cerebellar cortex) via PFs.

We first consider the firing activity in the whole population of GR cells for $p_c^* = 0.029$. Collective firing activity may be well visualized in the raster plot of spikes which is a collection of spike trains of individual neurons. As a collective quantity showing whole-population firing behaviors, we use an instantaneous whole-population spike rate $R_{\mathrm{GR}}(t)$ which may be got from the raster plots of spikes [76–83]. To get a smooth instantaneous whole-population spike rate, we employ the kernel density estimation (kernel smoother) [84]. Each spike in the raster plot is convoluted (or blurred) with a kernel function $K_h(t)$, and then a smooth estimate of instantaneous whole-population spike rate $R_{\mathrm{GR}}(t)$ is got by averaging the convoluted kernel function over all spikes of GR cells

FIG. 4: Firing activity of GR cells in an optimal case of p_c (connection probability from GO to GR cells) = 0.029. (a) Raster plots of spikes of 10^3 randomly chosen GR cells and (b) instantaneous whole-population spike rate $R_{\mathrm{GR}}(t)$ in the whole population of GR cells for the 1st step in the learning process for the EBC. Plots of the activation degrees (c1) $A(t)$ in the whole population of GR cells and (c2) $A^{(G)}(t)$ in the G spiking group [$G (= w)$: well-matched (solid curve) and $G (= i)$: ill- matched (dotted curve)]. Plots of (d1) instantaneous individual firing rate $f_{\mathrm{GR}}^{(i)}(t)$ for the active GR cells and (d2) instantaneous population spike rate $f_{\mathrm{GR}}^{(p)}(t)$ in the whole population of GR cells

in the whole population:

$$R_{\mathrm{GR}}(t) = \frac{1}{N} \sum_{i=1}^{N} \sum_{s=1}^{n_i} K_h(t - t_s^{(i)}), \qquad (16)$$

where $t_s^{(i)}$ is the sth spiking time of the ith GR cell, n_i is the total number of spikes for the ith GR cell, and N is the total number of GR cells (i.e., $N = N_c \cdot N_{\mathrm{GR}} = 51,200$). As a kernel function $K_h(t)$, we use a Gaussian kernel function of band width h:

$$K_h(t) = \frac{1}{\sqrt{2\pi}h} e^{-t^2/2h^2}, \qquad -\infty < t < \infty. \qquad (17)$$

Throughout the paper, the band width h of $K_h(t)$ is 10 msec.

Figure 4(a) shows a raster plot of spikes of 10^3 randomly chosen GR cells. At the beginning of trial stage ($0 < t < 7$ msec), all GR cells fire spikes due to the effect of strong transient CS signal of 200 Hz. In the remaining part of the trial stage ($7 < t < 1000$ msec), GR cells make random repetition of transitions between active and inactive states because of sustained CS signal of 30 Hz, and thus they seem to exhibit diverse spiking trains. Time passage from the CS onsets may be represented by the diverse spiking patterns of GR cells, which will be explained in details in Figs. 5 and 6. In the break stage ($1000 < t < 2000$ msec), GR cells fire very sparse spikes. For simplicity, only the raster plot in the range of $1000 < t < 1100$ msec is shown; the raster plot in a part of the preparatory stage (essentially the same as the break stage) ($-100 < t < 0$ msec), just before the 1st

trial stage, is also shown. Figure 4(b) shows the instantaneous whole-population spike rate $R_{GR}(t)$ in the whole population of GR cells. $R_{GR}(t)$ is basically in proportion to the transient and sustained CS inputs via MFs [see Figs. 1(b1)-1(b2)]. Thus, $R_{GR}(t)$ is sharply peaked in the beginning of the trial stage due to the strong transient CS, and then it becomes nearly flat in the remaining part of the trial stage where the sustained CS is present. However, due to the inhibitory effect of GO cells, the overall firing rates are uniformly lowered as follows. The time-averaged whole-population spike rates $\overline{R_{GR}(t)}$ in the time intervals of $0 < t < 5$ msec, $5 < t < 1000$ msec, and $1000 < t < 2000$ msec are 155.4 Hz, 32.5 Hz, and 3.4 Hz, respectively.

We next consider the activation degree of GR cells. To examine it, we divide the whole learning step into bins. In the beginning of the trial stage ($0 < t < 10$ msec), we divide the time interval into small bins (bin size: 1 msec) to properly take into consideration the effect of strong transient CS; the effect of transient CS seems to persist until the 7th bin. Then, in the remaining trial stage ($10 < t < 1000$ msec), to use wide bins (bin size: 10 msec) seems to be sufficient for considering the effect of sustained CS. Thus, we obtain the activation degree A_i for the active GR cells in the ith bin:

$$A_i = \frac{N_{a,i}}{N}. \tag{18}$$

Here, $N_{a,i}$ and N ($= 51,200$) are the number of active GR cells in the ith bin and the total number of GR cells, respectively. Figure 4(c1) shows a plot of the activation degree $A(t)$ in the whole population of GR cells. In the initial 7 bins ($0 < t < 7$ msec), $A = 1$ due to the effect of strong transient CS. In the presence of sustained CS for $7 < t < 1000$ msec, the activation degree A decreases monotonically from 0.189 to 0.131. In this case, the time-averaged activation degree $\overline{A(t)}$ is 0.161. Hence, representation of time passage from the onsets of CS can be made in a sparse recoding scheme (i.e., MF inputs become more sparse via recoding in the granular layer). In the break stage ($1000 < t < 2000$ msec), the time-averaged activation degree $\overline{A(t)}$ is 0.011 and small variations occur, which may be regarded as nearly "silent" stage, in comparison with the trial stage.

The whole population of GR cells may be decomposed into two types of well-matched and ill-matched spiking groups; details will be given in Figs. 5 and 6. Spiking patterns in the well-matched (ill-matched) spiking group are well (ill) matched with the airpuff US signal. In this case, the activation degree $A_i^{(G)}$ of active GR cells in the ith bin in the G spiking group is given by:

$$A_i^{(G)} = \frac{N_{a,i}^{(G)}}{N^{(G)}}. \tag{19}$$

Here, $N_{a,i}^{(G)}$ and $N^{(G)}$ are the number of active GR cells in the ith bin and the total number of GR cells in the G spiking group, respectively ($G = w$ and i for the well-matched

and the ill-matched spiking groups, respectively). The number of clusters, belonging to the well- and the ill-matched spiking groups are 841 and 183, respectively, and hence $N^{(w)} = 42,050$ and $N^{(i)} = 9,150$ because $N_{GR} = 50$ (number of GR cells in each cluster).

Figure 4(c2) shows plots of activation degree $A^{(G)}(t)$ in the well-matched (solid line) and the ill-matched (dotted curve) spiking groups. In the beginning of the trial stage [i.e., in the initial 7 bins ($0 < t < 7$ msec)], $A^{(G)} = 1$, independently of the spiking groups, due to the effect of strong transient CS. On the other hand, in the remaining trial stage ($7 < t < 1000$ msec) where the sustained CS is present, $A^{(G)}(t)$ varies, strongly depending on the type of spiking groups. In the case of well-matched spiking group, $A^{(w)}(t)$ decreases monotonically from 0.2 to 0.133, which is a little higher than $A(t)$ in the whole population. In contrast, in the case of ill-matched spiking group, $A^{(i)}(t)$ forms a well-shaped curve with a central "zero-bottom" with the time-averaged activation degree $\overline{A^{(i)}(t)} = 5.32 \cdot 10^{-4}$ for $330 < t < 580$ msec. Due to appearance of the central zero-bottom, contribution of the ill-matched spiking group to $A(t)$ in the whole population may be negligible in the range of $330 < t < 580$ msec. In the break stage ($1000 < t < 2000$ msec), the time-averaged activation degree $\overline{A^{(G)}(t)} = 0.011$ ($G = w$ or i) with small variations, independently of the spiking groups, which is the same as $\overline{A(t)}$ in the whole population.

In each ith bin, the contribution $C_i^{(G)}$ of each spiking group to the activation degree A_i in the whole population is given by the product of the fraction $F^{(G)}$ and the activation degree $A_i^{(G)}$ of the spiking group:

$$C_i^{(G)} = F^{(G)} \cdot A_i^{(G)} = \frac{N_{a,i}^{(G)}}{N}, \tag{20}$$

where $F^{(G)} = N^{(G)}/N$; $F^{(w)} = 0.821$ (82.1%) and $F^{(i)} = 0.179$ (17.9%) [see Fig. 6(b)]. Hence, we can easily get the contribution $C_i^{(G)}$ of each spiking group by just multiplying $A_i^{(G)}$ in Fig. 4(c2) with the fraction $F^{(G)}$. The sum of $C_i^{(G)}(t)$ over the well- and the ill-matched spiking groups is just the activation degree $A_i(t)$ in the whole population (i.e., $A_i = C_i^{(w)} + C_i^{(i)}$). In this case, contribution $C_i^{(i)}$ of the ill-matched spiking group becomes small due to both low activation degree $A_i^{(i)}$ and small fraction $F^{(i)}$. Particularly, because of existence of the central zero-bottom, contribution $C_i^{(i)}$ is negligibly small in the middle ($330 < t < 580$ msec) of the trial stage.

In the whole population, the activation degree $A(t)$ showing decreasing tendency is in contrast to the instantaneous whole-population spike rate $R_{GR}(t)$ which is flat in the trial stage. To understand this discrepancy, we consider the bin-averaged instantaneous individual firing

rate $f_{GR}^{(i)}$ of active GR cells:

$$f_{GR}^{(i)} = \frac{N_{s,i}}{N_{a,i}\,\Delta t}. \qquad (21)$$

Here, $N_{s,i}$ is the number of spikes of GR cells in the ith bin, $N_{a,i}$ is the number of active GR cells in the ith bin, and Δt is the bin size. Figure 4(d1) shows a plot of $f_{GR}^{(i)}(t)$ for the active GR cells. In the initial 7 bins $(0 < t < 7$ msec) of the trial stage where $A(t) = 1$, $f_{GR}^{(i)}(t)$ decreases very slowly from 155.6 to 155.3 Hz with the time t (i.e., the values of $f_{GR}^{(i)}(t)$ are nearly the same). In the remaining part $(7 < t < 1000$ msec) of the trial stage, $f_{GR}^{(i)}(t)$ increases monotonically from 173 to 258 Hz. In this case, the bin-averaged instantaneous population spike rate $f_{GR}^{(p)}$ is given by the product of the activation degree A_i of Eq. (18) and the instantaneous individual firing rate $f_{GR}^{(i)}$ of Eq. (21):

$$f_{GR}^{(p)} = A_i\,f_{GR}^{(i)} = \frac{N_{s,i}}{N\,\Delta t}. \qquad (22)$$

Figure 4(d2) shows a plot of the instantaneous population spike rate $f_{GR}^{(p)}(t)$. It is flat except for the sharp peak in the beginning of the trial stage, as in the case of $R_{GR}(t)$. We note that both $f_{GR}^{(p)}(t)$ and $R_{GR}(t)$ correspond to bin-based estimate and kernel-based smooth estimate for the instantaneous whole-population spike rate of the GR cells, respectively [76]. Although the activation degree $A(t)$ of GR cells decreases with t, their population spike rate keeps the flatness (i.e., $f_{GR}^{(p)}(t)$ becomes flat), because of the increase in the individual firing rate $f_{GR}^{(i)}(t)$. As a result, the bin-averaged instantaneous population spike rate $f_{GR}^{(p)}(t)$ in Fig. 4(d2) becomes essentially equal to the instantaneous whole-population spike rate $R_{GR}(t)$ in Fig. 4(b).

B. Dynamical Classification and Dynamical Origin of Diverse Spiking Patterns in The GR Clusters

There exist N_C $(= 2^{10})$ GR clusters in the whole population. N_{GR} $(= 50)$ GR cells in each GR cluster share the same inhibitory and excitatory inputs via their dendrites which synaptically contact the four glomeruli (i.e., terminals of MFs) at both ends of the GR cluster [see Fig. 2(b)]. Nearby inhibitory GO cell axons innervate the four glomeruli. Due to the shared inputs, GR cells in each GR cluster exhibit similar firing behaviors.

As in the case of $R_{GR}(t)$ in Eq. (16), the firing activity of the Ith GR cluster is described in terms of its instantaneous cluster spike rate $R_{GR}^{(I)}(t)$ $(I = 1, \cdots, N_C)$:

$$R_{GR}^{(I)}(t) = \frac{1}{N_{GR}} \sum_{i=1}^{N_{GR}} \sum_{s=1}^{n_i^{(I)}} K_h(t - t_s^{(I,i)}), \qquad (23)$$

FIG. 5: Diverse spiking patterns in the GR clusters in an optimal case of $p_c^* = 0.029$. Raster plots of spikes and instantaneous cluster spike rates $R_{GR}^{(I)}(t)$ for diverse spiking patterns. Six well-matched spiking patterns in the Ith GR clusters; $I =$ (a1) 245, (a2) 174, (a3) 505, (a4) 722, (a5) 154, and (a6) 458. Four ill-matched spiking patterns in the Ith GR cluster; $I =$ (b1) 288, (b2) 654, (b3) 411, and (b4) 1001. $\mathcal{C}^{(I)}$ represents the conjunction index of the spiking pattern in the Ith GR cluster. Distribution of the reproducibility degree $\mathcal{R}^{(I)}$ in the (c1) whole population and the (c2) well- and (c3) ill-matched spiking groups. Bin size for the histograms in (c1)-(c3) is 0.01.

where $t_s^{(I,i)}$ is the sth spiking time of the ith GR cell in the Ith GR cluster and $n_i^{(I)}$ is the total number of spikes for the ith GR cell in the Ith GR cluster.

We introduce the conjunction index $\mathcal{C}^{(I)}$ of each GR cluster, denoting the matching degree of the spiking behavior $[R_{GR}^{(I)}(t)]$ of each Ith GR cluster with the airpuff US signal $f_{US}(t)$ for the desired timing [see Fig.1(c)]. The conjunction index $\mathcal{C}^{(I)}$ is given by the cross-correlation at the zero-time lag [i.e., $Corr_{GR}^{(I)}(0)$] between $R_{GR}^{(I)}(t)$ and $f_{US}(t)$:

$$Corr_{GR}^{(I)}(\tau) = \frac{\overline{\Delta f_{US}(t + \tau)\Delta R_{GR}^{(I)}(t)}}{\sqrt{\overline{\Delta f_{US}^2(t)}}\sqrt{\overline{\Delta R_{GR}^{(I)}}^2(t)}}, \qquad (24)$$

where $\Delta f_{US}(t) = f_{US}(t) - \overline{f_{US}(t)}$, $\Delta R_{GR}^{(I)}(t) = R_{GR}^{(I)}(t) - \overline{R_{GR}^{(I)}(t)}$, and the overline denotes the time average. We note that $\mathcal{C}^{(I)}$ represents well the phase difference between the spiking patterns $[R_{GR}^{(I)}(t)]$ of GR clusters and the US signal $[f_{US}(t)]$.

Figure 5 shows diverse spiking patterns of GR clusters. This type of diversity results from inhibitory coordination of GO cells on the firing activity of GR cells in the GR-GO feedback loop in the granular layer. Time passage from the CS onsets may be well represented by the diverse spiking patterns of GR clusters because MF inputs become less similar (i.e., more orthogonal) to each other through recoding in the granular layer.

Six examples for the well-matched spiking patterns in the Ith ($I = 245, 174, 505, 722, 154,$ and 458) GR clusters are given in Figs. 5(a1)-5(a6), respectively. Raster plot of spikes of N_{GR} ($= 50$) GR cells and the corresponding instantaneous cluster spike rate $R_{\mathrm{GR}}^{(I)}(t)$ are shown, along with the value of the conjunction index $\mathcal{C}^{(I)}$ in each case of the Ith GR cluster. In all these cases, the instantaneous cluster spike rates $R_{\mathrm{GR}}^{(I)}(t)$ are well-matched with the US signal $f_{\mathrm{US}}(t)$, and hence these well-matched GR clusters have positive conjunction indices (i.e., $\mathcal{C}^{(I)} > 0$).

In the 1st case of $I = 245$ with the highest $\mathcal{C}^{(I)}$ ($= 0.79$), $R_{\mathrm{GR}}^{(245)}(t)$ is strongly localized around the middle of the trial stage (i.e. a central band of spikes is formed around $t = 500$ msec), and hence it is the most well-matched with the US signal $f_{\mathrm{US}}(t)$. In the 2nd case of $I = 174$ with $\mathcal{C}^{(I)} = 0.73$, $R_{\mathrm{GR}}^{(174)}(t)$ is also localized around $t = 500$ msec, but its central spiking band spreads a little more to the left side, in comparison with the case of $I = 245$. Hence, its conjunction index relative to $f_{\mathrm{US}}(t)$ is a little decreased.

We note that LTD at the PF-PC synapses occurs within an effective range of Δt_l^* ($\simeq -117.5$) $< \Delta t <$ Δt_r^* ($\simeq 277.5$) (see Fig. 3). Here, Δt [$= t_{\mathrm{CF}}$ (CF activation time) - t_{PF} (PF activation time)] is the relative time difference between the spiking times of the error-teaching instructor CF and the diversely-recoded student PF. The CF activation occurs approximately at $t_{\mathrm{CF}} = 500$ msec due to the strong brief US (strongly localized at $t = 500$ msec). Then, LTD may occur when the PF activation time t_{PF} lies in an effective LTD range of 222.5 msec $< t_{\mathrm{PF}} < 617.5$ msec. In the above two cases of GR clusters ($I = 245$ and 174) with higher $\mathcal{C}^{(I)}$, their PF signals (corresponding to axons of the GR cells) are strongly depressed by the US instructor signal because most parts of their spiking bands are well localized in the effective LTD range.

We next consider the 3rd and the 4th cases of the Ith GR cluster ($I = 505$ and 722) with intermediate $\mathcal{C}^{(I)}$. In the cases of $I = 505$ (722), the spiking band in the raster plot extends to the left (right) until $t \simeq 0$ (1000) msec. Thus, big left- and right-extended spiking bands appear for $I = 505$ and 722, respectively. Some part of this big spiking band lies inside the effective LTD range where LTD occurs in conjunction with the CF firing. On the other hand, its remaining part lies outside the effective LTD range, and hence LTP occurs for the PF firings alone without association with the CF signal.

We also consider the case of lower $\mathcal{C}^{(I)}$ for $I = 154$ and 458 (i.e., the 5th and the 6th cases). In both cases, they have tendency to fill the raster plots with more spikes via appearance of two or more spiking bands. Thus, some central part of these bands lies inside the effective LTD range where LTD occurs. In contrast, LTP occurs in the other left and right parts of the spiking bands because they lie outside the effective LTD range; in comparison with the case of intermediate $\mathcal{C}^{(I)}$, LTP region is extended. In this way, as $\mathcal{C}^{(I)}$ is decreased toward the zero, the raster plot tends to be filled with more spikes (constituting spiking bands), and hence the region where LTP occurs is extended.

In addition to the well-matched spiking patterns, ill-matched spiking patterns with negative conjunction indices (i.e., $\mathcal{C}^{(I)} < 0$) also appear. Four examples for the ill-matched spiking patterns in the Ith ($I = 288, 654, 411,$ and 1001) GR clusters are given in Figs. 5(b1)-5(b4), respectively. We first consider the case of $I = 288$ with the lowest $\mathcal{C}^{(I)}$ ($= -0.49$) (i.e., its magnitude $|\mathcal{C}^{(I)}|$: largest). This lowest case corresponds to the "opposite" case of the highest one for $I = 245$ with $\mathcal{C}^{(I)} = 0.79$ in the following sense. A central gap with negligibly small number of spikes (i.e., practically no spikes) appears around $t = 500$ msec, in contrast to the highest case where a central spiking band exists. Hence, in this lowest case, occurrence of LTD in the central gap may be practically negligible. On the other hand, mainly LTP occurs in the left and right spiking bands, most of which lie outside the effective LTD range. The right spiking band lies completely outside the effective LTD range, and hence no LTD occurs. The width of the central gap is larger than the width of the effective LTD range. However, since the gap is shifted a little to the right, a small part near the right boundary ($t \simeq 261$ msec) of the left spiking band overlaps with a small region near the left boundary ($t \simeq 222.5$ msec) of the effective LTD region. In this small overlapped region of $239 < \Delta t < 277.5$ msec, the values of the synaptic modification ΔJ_{LTD} (i.e., the average synaptic modification $\langle \Delta J_{\mathrm{LTD}} \rangle \simeq 0.031$) are very small, and hence very weak LTD may occur.

As the magnitude $|\mathcal{C}^{(I)}|$ is decreased, the central gaps becomes widened, and the widths of the left and the right spiking bands also get decreased, as shown in the cases of $I = 654, 411,$ and 1001. In these cases, the two left and right spiking bands lie completely outside the effective LTD range, and hence only LTP occurs for the PF signals alone without conjunction with the CF signal. In this way, as $|\mathcal{C}^{(I)}|$ approaches the zero from the negative side, spikes become more and more sparse, which is in contrast to the well-matched case where more and more spikes fill the raster plot as $\mathcal{C}^{(I)}$ goes to the zero from the positive side.

The above spiking patterns are shown in the 1st learning step [consisting of the 1st trial stage ($0 < t < 1000$ msec) and the 1st break stage ($1000 < t < 2000$ msec)]. For simplicity, they are shown in the range of $1000 < t < 1100$ msec in the break stage, and a part of the preliminary stage ($-100 < t < 0$ msec), preceding the 1st learning step, is also shown. We examine the reproducibility of the spiking patterns across the learning steps. To this end, we consider the cross-correlation between the instantaneous cluster spike rates $R_{\mathrm{GR}}^{(I,l)}(t)$ in the Ith GR cluster for the kth ($l = k$) and the ($k+1$)th ($l = k+1$) learning

steps;

$$Corr_{\mathrm{GR}}^{(I,k)}(\tau) = \frac{\overline{\Delta R_{\mathrm{GR}}^{(I,k)}(t+\tau)\Delta R_{\mathrm{GR}}^{(I,k+1)}(t)}}{\sqrt{\overline{\Delta R_{\mathrm{GR}}^{(I,k)2}(t)}}\sqrt{\overline{\Delta R_{\mathrm{GR}}^{(I,k+1)2}(t)}}}, \quad (25)$$

where $\Delta R_{\mathrm{GR}}^{(I,l)}(t) = R_{\mathrm{GR}}^{(I,l)}(t) - \overline{R_{\mathrm{GR}}^{(I,l)}(t)}$ ($l = k$ and $k + 1$) and the overline represents the time average. Then, the reproducibility degree $\mathcal{R}^{(I)}$ of the Ith GR cluster is given by the average value of the cross-correlations at the zero-time lag between the instantaneous cluster spike rates $R_{\mathrm{GR}}^{(I,l)}(t)$ for the successive learning steps:

$$\mathcal{R}^{(I)} = \frac{1}{N_{\mathrm{step}}-1}\sum_{k=1}^{N_{\mathrm{step}}-1} Corr_{\mathrm{GR}}^{(I,k)}(0), \quad (26)$$

where N_{step} is the total number of learning steps. Here, we consider the case of $N_{\mathrm{step}} = 100$.

Figure 5(c1) shows the distribution of the reproducibility degrees $\mathcal{R}^{(I)}$ for the whole GR clusters. Double peaks appear; large broad peak and small sharp peaks at $\mathcal{R}^{(I)} = 0.925$ and 0.815, respectively. The range of $\mathcal{R}^{(I)}$ is $(0.812, 0.997)$. Hence, the spiking patterns are highly reproducible across the learning steps. Figures 5(c2) and 5(c3) also show the distributions of $\mathcal{R}^{(I)}$ of the GR clusters in the well- and the ill-matched spiking groups, respectively. In the case of well-matched spiking group, the distribution of $\mathcal{R}^{(I)}$ has a broad peak and its range is $(0.842, 0.997)$. On the other hand, in the ill-matched case, the distribution decreases from its maximum at $\mathcal{R}^{(I)} = 0.815$, and its range is $(0.812, 0.879)$. We note that the average values of $\{\mathcal{R}^{(I)}\}$ in the well- and the ill-matched spiking group are 0.927 and 0.828, respectively. Hence, on average, the spiking patterns of the GR clusters in the well-matched spiking group may be more reproducible than those in the ill-matched spiking group, because the average individual firing rate in the well-matched spiking group is higher than that in the ill-matched spiking group.

Results on characterization of the diverse well- and ill-matched spiking patterns are given in Fig. 6. Figure 6(a) shows the plot of the fraction of conjunction indices $\mathcal{C}^{(I)}$ in the whole GR clusters. $\mathcal{C}^{(I)}$ increases slowly from the negative value to the peak at 0.35, and then it decreases rapidly. For this distribution of $\{\mathcal{C}^{(I)}\}$, the range is $(-0.49, 0.79)$, the mean is 0.3331, and the standard deviation is 0.6135. Then, we obtain the diversity degree $\mathcal{D}$ for the spiking patterns $[R_{\mathrm{GR}}^{(I)}(t)]$ of all the GR clusters:

$$\mathcal{D} = \text{Relative Standard Deviation}$$
$$\text{for the Distribution of } \{\mathcal{C}^{(I)}\}, \quad (27)$$

where the relative standard deviation is just the standard deviation divided by the mean. In the optimal case of $p_c^* = 0.029$, $\mathcal{D}^* \simeq 1.842$, which is just a quantitative measure for the diverse recoding made through feedback cooperation between the GR and the GO cells in the

FIG. 6: Characterization of diverse spiking patterns in the GR clusters in an optimal case of $p_c^* = 0.029$. (a) Distribution of conjunction indices $\{\mathcal{C}^{(I)}\}$ in the whole population. (b) Fraction of well-matched and ill-matched spiking groups. Distribution of conjunction indices $\{\mathcal{C}^{(I)}\}$ for the (c1) well- and (c2) ill-matched spiking groups. Bin size for the histograms in (a) and in (c1)-(c3) is 0.1. (d) Plots of reproducibility degree $\mathcal{R}^{(I)}$ versus $\mathcal{C}^{(I)}$ in the well-matched ($\mathcal{C}^{(I)} > 0$) and the ill-matched ($\mathcal{C}^{(I)} < 0$) spiking groups.

granular layer. It will be seen that $\mathcal{D}^*$ is just the maximum in Fig. 16(b) for the plot of $\mathcal{D}$ versus p_c. Hence, spiking patterns of the GR clusters at p_c^* is the most diverse.

We decompose the whole GR clusters into the well-matched ($\{\mathcal{C}^{(I)}\} > 0$) and the ill-matched ($\{\mathcal{C}^{(I)}\} < 0$) spiking groups. Figure 6(b) shows the fraction of spiking groups. The well-matched spiking group is a major one with fraction 82.1%, while the ill-matched spiking group is a minor one with fraction 17.9%. In this case, the spiking-group ratio $\mathcal{R}_{\mathrm{sp}}$, given by the ratio of the fraction of the well-matched spiking group to that of the ill-matched spiking group is 4.59. For this spiking-group ratio, spiking patterns of the GR clusters are the most diverse.

Figures 6(c1)-6(c2) also show the plots of conjunction indices $\mathcal{C}^{(I)}$ of the GR clusters in the well- and the ill-matched spiking groups, respectively. In the case of well-matched spiking group, the distribution of $\mathcal{C}^{(I)}$ with a peak at 0.35 has only positive values in the range of $(0.0, 0.79)$, and its mean and standard deviations are 0.428 and 0.372, respectively. On the other hand, in the case of the ill-matched spiking group, the distribution of $\mathcal{C}^{(I)}$ with a maximum at -0.05 has only negative values in the range of $(-0.49, 0.0)$, and its mean and standard deviations are -0.104 and 0.129, respectively. In this case, $\mathcal{C}^{(I)}$ increases slowly to the maximum. As will be seen in the next subsection, these well- and the ill-matched spiking groups play their own roles in the synaptic plasticity at PF-PC synapses and the subsequent learning process

FIG. 7: Dynamical origin of diverse spiking patterns in the GR clusters in an optimal case of $p_c^* = 0.029$. Well-matched spiking patterns for $I=$ (a1) 245 and (a2) 722 and ill-matched spiking patterns for $I=$ (a3) 288 and (a4) 654. In (a1)-(a4), top panel: raster plots of spikes in the sub-population of pre-synaptic GO cells innervating the Ith GR cluster, 2nd panel: plots of $f_X^{(I)}(t)$: bin-averaged instantaneous spike rates of the MF signals ($X =$ MF) into the Ith GR cluster (gray line) and bin-averaged instantaneous sub-population of pre-synaptic GO cells ($X =$ GO) innervating the Ith GR cluster (black line); $\langle \cdots \rangle_r$ represents the realization average (number of realizations is 100), 3rd panel: time course of $\langle g_{tot}^{(I)}(t) \rangle_r$: conductance of total synaptic inputs (including both the excitatory and inhibitory inputs) into the Ith GR cluster, and bottom panel: plots of $R_{\mathrm{GR}}^{(I)}(t)$: instantaneous cluster spike rate in the Ith GR cluster. (b) Distribution of conjunction indices $\{\mathcal{C}_{\mathrm{in}}^{(I)}\}$ for the conductances of total synaptic inputs into the GR clusters.

for the EBC, respectively.

We also examine the correlation between the conjunction index $\mathcal{C}^{(I)}$ and the reproducibility degree $\mathcal{R}^{(I)}$. Figure 6(d) show the plots of $\mathcal{C}^{(I)}$ versus $\mathcal{R}^{(I)}$ in the well- and the ill-matched spiking groups. In both cases, there appear strong negative correlations between $\mathcal{C}^{(I)}$ and $\mathcal{R}^{(I)}$; for the well-matched (ill-matched) spiking group, the Pearson's correlation coefficient is $r = -0.9999$ (-0.9978). When left-right reflections are made on Figs. 6(c1)-6(c2), shapes of the reflected ones are similar to the shapes of Figs. 5(c2)-5(c3), respectively, which implies the negative correlation between $\mathcal{C}^{(I)}$ and $\mathcal{R}^{(I)}$ in each spiking group. As shown in Figs. 5(a1)-5(a6), as $\mathcal{C}^{(I)}$ decreases to the zero from the positive side, the raster plot tends to be filled with more spikes due to increased individual firing rates, which leads to increase in $\mathcal{R}^{(I)}$. On the other hand, as $\mathcal{C}^{(I)}$ increases to the zero from the negative side, the raster plot of spikes tends to be more sparse because of decreased individual firing rates [see Figs. 5(b1)-5(b4)], which results in decrease in $\mathcal{R}^{(I)}$. Consequently, there appears a gap at the limit $\mathcal{C}^{(I)} = 0$.

Finally, we study the dynamical origin of diverse spiking patterns in the Ith GR clusters. As examples, we consider two well-matched spiking patterns for $I = 245$

and 722 [see the spiking patterns in Figs. 5(a1) and 5(a4)] and two ill-matched spiking patterns for $I = 288$ and 654 [see the spiking patterns Figs. 5(b1) and 5(b2)]. In Fig. 7, (a1)-(a4) correspond to the cases of $I =$245, 722, 288, and 654, respectively.

Diverse recodings for the MF signals are made in the GR layer, consisting of excitatory GR and inhibitory GO cells (i.e., in the GR-GO cell feedback loop). Thus, firing activities of GR cells are determined by two types of synaptic input currents (i.e., excitatory synaptic inputs via MF signals and inhibitory synaptic inputs from randomly connected GO cells). Then, investigations on the dynamical origin of diverse spiking patterns of the GR clusters are made via analysis of total synaptic inputs into the GR clusters. As in Eq. (6), synaptic current is given by the product of synaptic conductance g and potential difference. In this case, synaptic conductance determines the time-course of synaptic current. Hence, it is sufficient to consider the time-course of synaptic conductance. The synaptic conductance g is given by the product of synaptic strength per synapse, the number of synapses M_{syn}, and the fraction s of open (post-synaptic) ion channels [see Eq. (7)]. Here, the synaptic strength per synapse is given by the product of maximum synaptic conductance $\bar{g}$ and synaptic weight J, and the time-course of $s(t)$ is given by a summation for exponential-decay functions over pre-synaptic spikes, as shown in Eqs. (7) and (8).

Here, we make an approximation of the fraction $s(t)$ of open ion channels (i.e., contributions of summed effects of pre-synaptic spikes) by the bin-averaged spike rate $f_X^{(I)}(t)$ of pre-synaptic neurons ($X =$ MF and GO); $f_{\mathrm{MF}}^{(I)}(t)$ is the bin-averaged spike rate of the MF signals into the Ith GR cluster and $f_{\mathrm{GO}}^{(I)}(t)$ is the bin-averaged spike rate of the pre-synaptic GO cells innervating the Ith GR cluster. In the case of MF signal, we get:

$$f_{\mathrm{MF}}^{(I)}(t) = f_{\mathrm{TCS}}^{(I)}(t) + f_{\mathrm{SCS}}^{(I)}(t), \tag{28}$$

where $f_{\mathrm{TCS}}^{(I)}(t)$ and $f_{\mathrm{SCS}}^{(I)}(t)$ are the bin-averaged spike rates of the transient and the sustained CS signals, respectively.

Then, the conductance $g_X^{(I)}(t)$ of synaptic input from X (=MF or GO) into the Ith GR cluster ($I = 1, \cdots, N_C$) is given by:

$$g_X^{(I)}(t) \simeq \mathrm{M}_f^{(R)} \cdot f_X^{(I)}(t). \tag{29}$$

Here, the multiplication factor $\mathrm{M}_f^{(R)}$ [= maximum synaptic conductance $\bar{g}_R$ × synaptic weight $J^{(\mathrm{GR},X)}$ × number of synapses $M_{\mathrm{syn}}^{(\mathrm{GR},X)}$] varies depending on X and the receptor R on the post-synaptic GR cells. In the case of excitatory synaptic currents into the Ith GR cluster with AMPA receptors via TCS or SCS MF signal, $\mathrm{M}_f^{(\mathrm{AMPA})} = 2.88$; $\bar{g}_{\mathrm{AMPA}} = 0.18$, $J^{(\mathrm{GR,MF})} = 8.0$, and $M_{\mathrm{syn}}^{(\mathrm{GR},X)} = 2$ ($X=$TCS, SCS). On the other hand, in the case of the Ith GR cluster with NMDA receptors,

$\bar{g}_{\mathrm{NMDA}} = 0.025$, and hence $\mathrm{M}_f^{(\mathrm{NMDA})} = 0.4$, which is much less than $\mathrm{M}_f^{(\mathrm{AMPA})}$. For the inhibitory synaptic current from pre-synaptic GO cells to the Ith GR cluster with GABA receptors, $\mathrm{M}_f^{(\mathrm{GABA})} = 2.63$; $\bar{g}_{\mathrm{GABA}} = 0.028$, $J^{(\mathrm{GR,GO})} = 10$, and $M_{\mathrm{syn}}^{(\mathrm{GR,GO})} = 9.4$. Then, the conductance $g_{tot}^{(I)}$ of total synaptic inputs (including both the excitatory and the inhibitory inputs) into the Ith GR cluster is given by:

$$
\begin{aligned}
g_{tot}^{(I)}(t) &= g_{\mathrm{MF}}^{(I)} - g_{\mathrm{GO}}^{(I)} = g_{\mathrm{AMPA}}^{(I)} + g_{\mathrm{NMDA}}^{(I)} - g_{\mathrm{GO}}^{(I)} \\
&= 3.28\, f_{\mathrm{MF}}^{(I)}(t) - 2.63\, f_{\mathrm{GO}}^{(I)}(t). \quad (30)
\end{aligned}
$$

In Figs. 7(a1)-7(a4), the top panels show the raster plots of spikes in the sub-populations of pre-synaptic GO cells innervating the Ith GR clusters. From these raster plots, bin-averaged (sub-population) spike rates $f_{\mathrm{GO}}^{(I)}(t)$ may be obtained. The bin-averaged spike rate of pre-synaptic GO cells in the ith bin is given by $\frac{n_i^{(s)}}{N_{pre}\,\Delta t}$, where $n_i^{(s)}$ is the number of spikes in the ith bin, Δt is the bin size, and N_{pre} (=10) is the number of pre-synaptic GO cells. As in Fig. 4, in the beginning of the trial stage ($0 < t < 10$ msec), we employ the small bin-size of $\Delta t = 1$ msec to properly take into consideration the effect of strong transient CS, and in the remaining trial stage ($10 < t < 1000$ msec), we use the wide bin-size of $\Delta t = 10$ msec for considering the effect of sustained CS. Through an average over 100 realizations, we obtain the realization-averaged (bin-averaged) spike rate of pre-synaptic GO cells $\langle f_{\mathrm{GO}}^{(I)}(t) \rangle_r$ because N_{pre} (= 10) is small; $\langle \cdots \rangle_r$ represent a realization-average. The 2nd panels show $\langle f_{\mathrm{GO}}^{(I)}(t) \rangle_r$ (black line) and $\langle f_{\mathrm{MF}}^{(I)}(t) \rangle_r$ (gray line). We note $\langle f_{\mathrm{GO}}^{(I)}(t) \rangle$ varies depending on I, while $\langle f_{\mathrm{MF}}^{(I)}(t) \rangle$ is independent of I. Then, we obtain the realization-averaged conductance $\langle g_{tot}^{(I)}(t) \rangle_r$ of total synaptic inputs in Eq. (30), which is shown in the 3rd panels.

We note that the shapes of $\langle g_{tot}^{(I)}(t) \rangle_r$ (corresponding to the total input into the Ith GR cluster) in the 3rd panels are nearly the same as those of $R_{\mathrm{GR}}^{(I)}(t)$ (corresponding to the output of the Ith GR cluster) in the bottom panels. It is thus expected that well-matched (ill-matched) inputs into the GR clusters may lead to generation of well-matched (ill-matched) outputs (i.e., responses) in the GR clusters. To confirm this point clearly, as in case of the spiking patterns $[R_{\mathrm{GR}}^{(I)}(t)]$ in the GR clusters, we introduce the conjunction index for the total synaptic input of the Ith GR cluster, representing the matching degree between $\langle g_{tot}^{(I)}(t) \rangle_r$ (conductance of total synaptic input into the Ith GR cluster) and the (airpuff) US signal $f_{\mathrm{US}}(t)$ for the desired timing. Similar to the conjunction index $\mathcal{C}^{(I)}$ for the spiking patterns (i.e. outputs) in the Ith GR cluster [see Eq. (24)], the conjunction index $\mathcal{C}_{\mathrm{in}}^{(I)}$ for the total synaptic input is given by the cross-correlation at the zero-time lag (i.e., $Corr_{\mathrm{in}}^{(I)}(0)$) between $\langle g_{tot}^{(I)}(t) \rangle_r$ and $f_{\mathrm{US}}(t)$:

$$
Corr_{\mathrm{in}}^{(I)}(\tau) = \frac{\overline{\Delta f_{\mathrm{US}}(t+\tau)\Delta\langle g_{tot}^{(I)}(t)\rangle_r}}{\sqrt{\overline{\Delta f_{\mathrm{US}}^2(t)}}\sqrt{\overline{\Delta\langle g_{tot}^{(I)}(t)\rangle_r}^2}}, \quad (31)
$$

where $\Delta f_{\mathrm{US}}(t) = f_{\mathrm{US}}(t) - \overline{f_{\mathrm{US}}(t)}$, $\Delta\langle g_{tot}^{(I)}(t)\rangle_r = \langle g_{tot}^{(I)}(t)\rangle_r - \overline{\langle g_{tot}^{(I)}(t)\rangle_r}$, and the overline represents the time average. Thus, we have two types of conjunction indices, $\mathcal{C}^{(I)}$ [output conjunction index: given by $Corr_{\mathrm{GR}}^{(I)}(0)$] and $\mathcal{C}_{\mathrm{in}}^{(I)}$ [input conjunction index: given by $Corr_{\mathrm{in}}^{(I)}(0)$] for the output and the input in the Ith GR cluster, respectively.

Figure 7(b) shows the plot of fraction of input conjunction indices $\{\mathcal{C}_{\mathrm{in}}^{(I)}\}$ in the whole GR clusters. We note that the distribution of input conjunction indices in Fig. 7(b) is nearly the same as that of output conjunction indices in Fig. 6(a). $\mathcal{C}_{\mathrm{in}}^{(I)}$ increases slowly from the negative value to the peak at 0.35, and then it decreases rapidly. In this distribution of $\{\mathcal{C}_{\mathrm{in}}^{(I)}\}$, the range is (-0.49, 0.79), the mean is 0.3332, and the standard deviation is 0.6137. Then, we get the diversity degree $\mathcal{D}_{\mathrm{in}}$ for the total synaptic inputs $\{\langle g_{tot}^{(I)}(t)\rangle_r\}$ of all the GR clusters:

$$
\begin{aligned}
\mathcal{D} =\ & \text{Relative Standard Deviation} \\
& \text{for the Distribution of } \{\mathcal{C}_{\mathrm{in}}^{(I)}\}. \quad (32)
\end{aligned}
$$

Hence, $\mathcal{D}_{\mathrm{in}} \simeq 1.842$ for the synaptic inputs, which is nearly the same as $\mathcal{D}^*$ ($\simeq 1.842$) for the spiking patterns of GR cells. Consequently, diverse synaptic inputs into the GR clusters results in generation of diverse outputs (i.e., spiking patterns) of the GR cells.

C. Effect of Diverse Temporal Recoding on Synaptic Plasticity at PF-PC Synapses

Based on dynamical classification of spiking patterns of GR clusters, we study the effect of diverse temporal recoding in the GR clusters on synaptic plasticity at PF-PC synapses. As shown in the preceding subsection, MF context input signals for the EBC are diversely recoded in the granular layer (corresponding to the input layer of the cerebellar cortex). The diversely-recoded well- and ill-matched PF (student) signals (coming from the GR cells) are fed into the PCs (i.e., principal cells of the cerebellar cortex) and the BCs in the Purkinje-molecular layer. The PCs also receive well-matched error-teaching CF (instructor) signals from the IO, together with the inhibitory inputs from the BCs. Then, the synaptic weights at the PF-PC synapses vary depending on the matching degree between the PF and the CF signals.

We first consider the change in normalized synaptic weights $\tilde{J}$ of active PF-PC synapses during the learning trials in the optimal case of $p_c^* = 0.029$;

$$
\tilde{J}_{ij}(t) = \frac{J_{ij}^{(\mathrm{PC,PF})}(t)}{J_0^{(\mathrm{PC,PF})}}. \quad (33)
$$

FIG. 8: Change in synaptic weights of active PF-PC synapses during learning trials in the optimal case of $p_c^* = 0.029$. (a1)-(a9) Trial-evolution of distribution of normalized synaptic weights $\tilde{J}(t)$ of active PF signals. (b1)-(b9) Trial-evolution of bin-averaged (normalized) synaptic weights $\langle \tilde{J}(t) \rangle$ of active PF signals. Bin size: $\Delta t = 50$ msec.

FIG. 9: Plots of (a) trial-averaged mean $\overline{\langle \tilde{J}(t) \rangle}$ and (b) modulation $\mathcal{M}_J$ for the bin-averaged (normalized) synaptic weights $\langle \tilde{J}(t) \rangle$ in Figs. 8(b1)-8(b9) versus trial. Dynamical balance between ΔLTD and ΔLTP during learning trials. Plots of (c1) trial-summed ΔLTD [$\langle \Delta$LTD$\rangle_s$ (solid circles)] and ΔLTP [$\langle \Delta$LTP$\rangle_s$ (open circles)] of active PF signals and (c2) ratio of $|\langle \Delta$LTD$\rangle_s|$ to $\langle \Delta$LTP$\rangle_s$ versus trial.

Here, the initial synaptic strength ($J_0^{(PC,PF)} = 0.006$) is the same for all PF-PC synapses. Figures 8(a1)-8(a9) show trial-evolution of distribution of $\tilde{J}$ of active PF-PC synapses. As the learning trial is increased, normalized synaptic weights $\tilde{J}$ change due to synaptic plasticity at PF-PC synapses. We note that the distribution of $\tilde{J}$ in each trial is composed of two markedly separated structures (i.e., a combination of separate top horizontal line with a central gap and lower band). Here, the top horizontal line with a central gap has no essential change with the trials, while the lower bands come down with the trials and their vertical widths increase. This kind of distribution of $\tilde{J}$ becomes saturated at about the 250th trial.

The top horizontal line with a central gap arises from the ill-matched spiking group (with negative conjunction indices). In the case of ill-matched PF signals, practically no LTD occurs because most of them have no associations with the error-teaching CF signals which are strongly localized in the middle of trial (i.e., near $t = 500$ msec). As shown in Fig. 4(c2), the activation degree $A^{(i)}(t)$ (denoted by the dotted line) of GR cells in the ill-matched spiking group has a central "zero-bottom" where $A^{(i)}(t) \simeq 0$ (i.e., negligibly small number of spikes in the middle part of trial). In the initial and the final parts of the trial (outside the middle part), practically no LTD takes place due to no practical conjunctions with the strongly-localized CF signals. Thus, the normalized synaptic weights $\tilde{J}$ of the active GR cells in the ill-matched spiking group forms the top horizontal line with a central gap which is nearly invariant with the

trial.

On the other hand, lower bands arise from the well-matched spiking group (with positive conjunction indices). In the case of well-matched PF signals, they are strongly depressed by the error-teaching CF signals (i.e., strong LTD occurs) in each trial due to good association between the well-matched PF and CF signals. As a result, a lower band is formed, it comes down with the trial, and eventually becomes saturated.

To more clearly examine the above trial evolutions, we obtain the bin-averaged (normalized) synaptic weight in each ith bin (bin size: $\Delta t = 50$ msec):

$$\langle \tilde{J}(t) \rangle_i = \frac{1}{N_{s,i}} \sum_{f=1}^{N_{s,i}} \tilde{J}_{i,f}(t), \qquad (34)$$

where $\tilde{J}_{i,f}$ is the normalized synaptic weight of the fth active PF signal in the ith bin, and $N_{s,i}$ is the total number of active PF signals in the ith bin. Figures 8(b1)-8(b9) show trial-evolution of bin-averaged (normalized) synaptic weights $\langle \tilde{J}(t) \rangle$ of active PF signals. In each trial, $\langle \tilde{J}(t) \rangle$ forms a step-well-shaped curve. As the trial is increased, the step-well curve comes down, its width and depth increase, and saturation seems to occur at about the 250th cycle.

We also obtain the trial-averaged mean $\overline{\langle \tilde{J}(t) \rangle}$ via time average of $\langle \tilde{J}(t) \rangle$ over a trial:

$$\overline{\langle \tilde{J} \rangle} = \frac{1}{N_b} \sum_{i=1}^{N_b} \langle \tilde{J}(t) \rangle_i. \qquad (35)$$

Here, N_b is the number of bins for trial averaging, and the overbar represents the time average. Figures 9(a) and 9(b) show plots of the trial-averaged mean $\overline{\langle \tilde{J}(t) \rangle}$ and the modulation [=(maximum - minimum)/2] $\mathcal{M}_J$ for $\langle \tilde{J}(t) \rangle$

versus trial. The trial-averaged mean $\overline{\langle \tilde{J}(t) \rangle}$ decreases monotonically from 1.0 due to LTD at PF-PC synapses, and it becomes saturated at 0.367 at about the 250th trial.

However, strength of the LTD varies depending on the parts of the trial. In the middle part without practical contribution of ill-matched spiking group, strong LTD occurs, due to contribution of only well-matched active PF signals. On the other hand, at the initial and the final parts, somewhat less LTD takes place, because both the ill-matched spiking group (with practically no LTD) and the well-matched spiking group make contributions together. Consequently, with increasing trial, the middle-stage part comes down more rapidly than the initial and final parts. Hence, the modulation $\mathcal{M}_J$ increases monotonically from 0, and it gets saturated at 0.0867 at about the 250th trial.

We investigate the dynamical origin for the saturation of trial-averaged mean $\overline{\langle \tilde{J}(t) \rangle}$ in Fig. 9(a). As explained in the Subsec. II E, a major LTD occurs when a CF firing makes association with earlier PF signals [see the $\Delta \mathrm{LTD}_{ij}^{(1)}$ in Eq. (12)], while a minor LTD takes place when the CF firing makes conjunction with later CF signals [see the $\Delta \mathrm{LTD}_{ij}^{(2)}$ in Eq. (13)]. We consider the trial-summed $\Delta \mathrm{LTD}$ ($\langle \Delta \mathrm{LTD} \rangle_s$) in each trial, given by the sum of $\Delta \mathrm{LTD}_{ij}$ occurring at all active PF-PC synapses during the whole trial ($0 < t < 1000$ msec), where $\Delta \mathrm{LTD}_{ij} = \Delta \mathrm{LTD}_{ij}^{(1)} + \Delta \mathrm{LTD}_{ij}^{(2)}$. Similarly, we also consider the trial-summed $\Delta \mathrm{LTP}$ ($\langle \Delta \mathrm{LTP} \rangle_s$) in each trial, given by the sum of $\Delta \mathrm{LTP}_{ij}$ occurring at all active PF-PC synapses during the whole trial. Here, LTP occurs for the PF signals alone without association with CF signals; $\Delta \mathrm{LTP}_{ij}$ is given in Eq. (14).

Figures 9(c1) shows trial-evolution of trial-summed $\Delta \mathrm{LTD}$ [$\langle \Delta \mathrm{LTD} \rangle_s$ (solid circles)] and $\Delta \mathrm{LTP}$ [$\langle \Delta \mathrm{LTP} \rangle_s$ (open circles)] occurring at all active PF-PC synapses. Here, $\langle \Delta \mathrm{LTP} \rangle_s$ ($\simeq 0.00386$), independently of the trial. In the beginning trials, $|\langle \Delta \mathrm{LTD} \rangle_s|$ (i.e., magnitude of $\langle \Delta \mathrm{LTD} \rangle_s$) is larger than $\langle \Delta \mathrm{LTP} \rangle_s$, and hence $\Delta \mathrm{LTD}$ is dominant. However, with increasing the trial, $|\langle \Delta \mathrm{LTD} \rangle_s|$ decreases monotonically, and becomes saturated at 0.00386 at about the 250th trial. Thus, the saturated value of $|\langle \Delta \mathrm{LTD} \rangle_s|$ becomes the same as that of $\langle \Delta \mathrm{LTP} \rangle_s$. This process may be well seen in Fig. 9(c2). With increasing the trial, the ratio of $|\langle \Delta \mathrm{LTD} \rangle_s|$ to $\langle \Delta \mathrm{LTP} \rangle_s$ decreases from 4.71, and it converges to 1 at about the 250th trial. Thus, trial-level balance between $\Delta \mathrm{LTD}$ and $\Delta \mathrm{LTP}$ occurs [i.e., $|\langle \Delta \mathrm{LTD} \rangle_s| = \langle \Delta \mathrm{LTP} \rangle_s$ ($\simeq 0.00386$)] at about the 250th trial, which results in the saturation of trial-averaged mean $\overline{\langle \tilde{J}(t) \rangle}$.

FIG. 10: Change in firing activity of PCs during learning trial in the optimal case of $p_c^* = 0.029$. (a1)-(a9) Raster plots of spikes of PCs and (b1)-(b9) instantaneous population spike rates $R_{\mathrm{PC}}(t)$. (c1)-(c9) Realization-averaged instantaneous population spike rates $\langle R_{\mathrm{PC}}(t) \rangle_r$; number of realizations is 100. Plots of (d1) trial-averaged mean $\overline{\langle R_{\mathrm{PC}}(t) \rangle_r}$ and (d2) modulations $\mathcal{M}_{\mathrm{PC}}$ for $\langle R_{\mathrm{PC}}(t) \rangle_r$ versus trial.

D. Effect of PF-PC Synaptic Plasticity on Subsequent Learning Process in The PC-CN-IO System

As a result of diverse recoding in the GR clusters, well- and ill-matched spiking groups appear. In the case of well-matched PF signals, they are strongly depressed by the CF signals due to good association between the PF and CF signals. On the other hand, in the case of ill-matched PF signals, practically no LTD occurs because most of them have no conjunctions with the error-teaching CF signals. In this subsection, we investigate the effect of this kind of effective PF-PC synaptic plasticity on the subsequent learning process in the PC-CN-IO system.

Figure 10 shows change in firing activity of PCs during learning trial in the optimal case of $p_c^* = 0.029$. Trial-evolutions of raster plots of spikes of 16 PCs and the corresponding instantaneous population spike rates $R_{\mathrm{PC}}(t)$ are shown in Figs. 10(a1)-10(a9) and Figs. 10(b1)-10(b9), respectively. Realization-averaged smooth instantaneous population spike rates $\langle R_{\mathrm{PC}}(t) \rangle_r$ are also shown in Figs. 10(c1)-10(c9). Here, $\langle \cdots \rangle_r$ denotes realization average and the number of realizations is 100.

$\langle R_{\text{PC}}(t)\rangle_r$ seems to be saturated at about the 250th cycle.

As shown in Figs. 8(b1)-8(b9), bin-averaged normalized synaptic weights $\langle \tilde{J}(t)\rangle$ form a step-well-shaped curve. In the middle part of each trial, strong LTD occurs due to contribution of only well-matched spiking group. On the other hand, at the initial and the final parts, somewhat less LTD takes place because both the ill-matched spiking group (with practically no LTD) and the well-matched spiking group make contributions together. As a result of this kind of effective depression at the (excitatory) PF-PC synapses, with increasing the learning trial, raster plots of spikes of all the 16 PCs become more and more sparse in the middle part of each trial (i.e, near $t = 500$ msec), which may be clearly seen in the instantaneous population spike rate $\langle R_{\text{PC}}(t)\rangle_r$. $\langle R_{\text{PC}}(t)\rangle_r$ becomes lower in the middle part than at the initial and the final parts. Thus, $\langle R_{\text{PC}}(t)\rangle_r$ also forms a step-well-shaped curve with a minimum in the middle part.

As the trial is increased, such step-well-shaped curve for $\langle R_{\text{PC}}(t)\rangle_r$ comes down and the (top) width and the depth of the well increase. Eventually, at the 141th trial, a "zero-bottom" is formed in the step-well in the middle part of the trial (i.e., $\langle R_{\text{PC}}(t)\rangle_r \simeq 0$ for $468 < t < 532$ msec). Appearance of the zero-bottom in the step-well is the prerequisite condition for acquisition of CR. At the zero-bottom of the step-well, PCs stop inhibition completely. This process may be seen well in Figs. 10(c1)-10(c5). Thus, from the 141th threshold trial, the CN neuron may fire spikes which evoke CR, which will be seen in Fig. 11. With increasing the trial from the 141th trial, both the (top) width of the step-well and the zero-bottom width are increased, although the depth of the well remains unchanged [see Figs. 10(c6)-10(c9)]. As a result, the strength $\mathcal{S}$ of CR increases, while its timing degree $\mathcal{T}_d$ is decreased; the details will be given in Fig. 11. The (overall) learning efficiency degree $\mathcal{L}_e$, taking into consideration both $\mathcal{T}_d$ and $\mathcal{S}$, increases with the trial, and becomes saturated at about the 250th trial.

Figures 10(d1) and 10(d2) show plots of trial-averaged mean $\overline{\langle R_{\text{PC}}(t)\rangle_r}$ (i.e., time average of $\langle R_{\text{PC}}(t)\rangle_r$ over the trial) and modulation $\mathcal{M}_{\text{PC}}$ of $\langle R_{\text{PC}}(t)\rangle_r$ versus trial, respectively. Due to effective LTD at the PF-PC synapses, the trial-averaged mean $\overline{\langle R_{\text{PC}}(t)\rangle_r}$ decreases monotonically from 92.47 Hz, and it gets saturated at 19.91 Hz at about the 250th cycle. On the other hand, the modulation $\mathcal{M}_{\text{PC}}$ increases monotonically from 0.352 Hz, and it becomes saturated at 16.11 Hz at about the 141th cycle. After the 141th threshold trial, $\mathcal{M}_{\text{PC}}$ remains unchanged due to no change in the depth of the step-well, unlike the case of $\overline{\langle R_{\text{PC}}(t)\rangle_r}$. These PCs (principal cells of the cerebellar cortex) exert effective inhibitory coordination on the CN neuron which evokes the CR (i.e., learned eyeblink).

Figure 11 shows change in firing activity of the CN neuron which produces the final output of the cerebellum during learning trial in the optimal case of $p_c^* = 0.029$. Trial-evolutions of raster plots of spikes of the CN neu-

FIG. 11: Change in firing activity of the CN neuron during learning trial in the optimal case of $p_c^* = 0.029$. (a1)-(a6) Raster plots of spikes of the CN neuron (i.e., collection of spike trains for all the realizations; number of realizations is 100) and (b1)-(b6) bin-averaged instantaneous individual firing rate $f_{\text{CN}}(t)$; the bin size is $\Delta t = 50$ msec. Plots of (c1) timing degree $\mathcal{T}_d$ and (c2) strength $\mathcal{S}$ of CR versus trial. (d) Plot of (overall) learning efficiency degree $\mathcal{L}_e$ for the CR versus trial.

ron (i.e., collection of spike trains for all the realizations; number of realizations is 100) and the bin-averaged instantaneous individual firing rates $f_{\text{CN}}(t)$ (i.e., the number of spikes of the CN neuron in a bin with the bin width $\Delta t = 50$ msec) are shown in Figs. 11(a1)-11(a6) and Figs. 11(b1)-11(b6), respectively. At the 140th trial, the CN neuron does not fire due to strong inhibition from the PCs, and thus it is silent (i.e., it lies in the silent period) during the whole trial stage ($0 < t < 1000$ msec). However, as a result of appearance of the zero-bottom in the step-well for $\langle R_{\text{PC}}(t)\rangle_r$ at the 141th threshold trial, the CN neuron begins to fire spikes in the middle part of the trial. In this case, as the time is increased from $t = 0$, the CN neuron first lies in the silent period, then a transition to the firing state occurs in the middle part, and finally another transition to the silent state also takes place. With increasing the trial, raster plots of spikes of the CN neuron become more and more dense in the middle part of each trial, in contrast to the case of PCs.

This process may be clearly seen in the instantaneous individual firing rates $f_{\text{CN}}(t)$. Due to the effective inhibitory coordinations of PCs on the CN neuron, $f_{\text{CN}}(t)$ begins to increase from 0 in the middle part of the trial, it reaches a peak, and then it decreases to 0 relatively slowly. The peak of $f_{\text{CN}}(t)$ appears a little earlier than the US presentation ($t = 500$ msec) which may denote the anticipatory CR [10, 49]. Thus, $f_{\text{CN}}(t)$ forms a bell-shaped curve. As the trial is increased, the bottom-base width and the peak height of the bell are increased, and $f_{\text{CN}}(t)$ seems to be saturated at about the 250th trial.

Figures 11(c1) and 11(c2) show plots of the timing degree $\mathcal{T}_d$ and the strength $\mathcal{S}$ of the CR versus trial, respectively. The timing degree $\mathcal{T}_d$, representing the match-

ing degree between the firing activity of the CN neuron [$f_{CN}(t)$] and the US signal $f_{US}(t)$, is given by the cross-correlation $Corr_T(0)$ at the zero lag between $f_{CN}(t)$ and $f_{US}(t)$:

$$Corr_T(\tau) = \frac{\overline{\Delta f_{US}(t+\tau)\Delta f_{CN}(t)}}{\sqrt{\overline{\Delta f_{US}^2(t)}}\sqrt{\overline{\Delta f_{CN}^2(t)}}}, \qquad (36)$$

where $\Delta f_{US}(t) = f_{US}(t) - \overline{f_{US}(t)}$, $\Delta f_{CN}(t) = f_{CN}(t) - \overline{f_{CN}(t)}$, and the overline denotes the time average. Practically, $\mathcal{T}_d$ reflects the width of the bottom base of the bell curve. With increasing the trial, the width of the bottom base increases, due to increase in the (top) width of the step-well curve for the PCs. As a result, $\mathcal{T}_d$ decreases monotonically from 0.912 at the 141th trial, and it becomes saturated at 0.346 at about the 250th trial. On the other hand, as the trial is increased, the peak height of the bell increases. Thus, the strength $\mathcal{S}$ of CR (representing the amplitude of eyelid closure), given by the modulation [(maximum - minimum)/2] of $f_{CN}(t)$, increases monotonically from 1.803 at the 141th trial, and it gets saturated at 32.38 at about the 250th trial.

Then, the (overall) learning efficiency degree $\mathcal{L}_e$ for the CR, taking into consideration both the timing degree $\mathcal{T}_d$ and the strength $\mathcal{S}$ of CR, is given by their product:

$$\mathcal{L}_e = \mathcal{T}_d \cdot \mathcal{S}. \qquad (37)$$

Figure 11(d) shows a plot of $\mathcal{L}_e$ versus trial. $\mathcal{L}_e$ increases monotonically from 1.645 at the 141th trial, and it becomes saturated at about the 250th cycle. Thus, we get the saturated learning efficiency degree $\mathcal{L}_e^*$ ($\simeq 11.19$). As will be seen in the next subsection, $\mathcal{L}_e^*$ is the largest one among the available ones. Hence, in the optimal case of $p_c^* = 0.029$ where spiking patterns of GR clusters with the diversity degree $\mathcal{D}^*$ ($\simeq 1.842$) are the most diverse, motor learning for the EBC with the saturated learning efficiency degree $\mathcal{L}_e^*$ is the most effective.

Learning progress can be clearly seen in the IO system. During the learning trial, the IO neuron receives both the excitatory (airpuff) US signal for the desired timing and the inhibitory signal from the CN neuron (representing a realized eye-movement). Then, the learning progress degree $\mathcal{L}_p$ is given by the ratio of the time-averaged inhibitory input from the CN neuron to the magnitude of the time-averaged excitatory input of the desired US timing signal:

$$\mathcal{L}_p = \frac{\overline{I_{GABA}^{(IO,CN)}}}{|\overline{I_{AMPA}^{(IO,US)}}|}. \qquad (38)$$

Here, $\overline{I_{GABA}^{(IO,CN)}}$ is the trial-averaged inhibitory GABA receptor-mediated current from the CN to the IO, and $\overline{I_{AMPA}^{(IO,US)}}$ is the trial-averaged excitatory AMPA receptor-mediated current into the IO via the desired US timing signal; no (excitatory) NMDA receptors exist on the IO.

FIG. 12: Change in firing activity of the IO neuron during learning trial in the optimal case of $p_c^* = 0.029$. Plots of (a1) realization-average for the time-averaged inhibitory synaptic current from the CN neuron ($\langle \overline{I_{GABA}^{(IO,CN)}} \rangle_r$) (open circles) and realization-average for the time-averaged excitatory synaptic current via the (airpuff) US signal ($\langle |\overline{I_{AMPA}^{(IO,US)}}| \rangle_r$) versus trial; number of realizations $\langle \cdots \rangle_r$ is 100. (a2) Plot of learning progress degree $\mathcal{L}_p$ versus trial. (b1)-(b5) Raster plots of spikes of the IO neuron (i.e., collection of spike trains for all the realizations; number of realizations is 100) and (c1)-(c5) bin-averaged instantaneous individual firing rate $f_{IO}(t)$; the bin size is $\Delta t = 40$ msec. (d) Plot of trial-averaged individual firing rate $\overline{f_{IO}(t)}$ versus trial.

[Note that the 4th term in Eq. (1) is given by $-I_{syn,i}^{(X)}(t)$, because $I_{GABA}^{(IO,CN)} > 0$ and $I_{AMPA}^{(IO,US)} < 0$.]

Figure 12(a1) shows plots of $\langle \overline{I_{GABA}^{(IO,CN)}} \rangle_r$ (open circles) and $\langle |\overline{I_{AMPA}^{(IO,US)}}| \rangle_r$ (crosses) versus trial in the optimal case of $p_c^* = 0.029$; $\langle \cdots \rangle_r$ represents an average over 100 realizations. At the 141th threshold trial, acquisition of CR starts (i.e., the CN neuron begins to fire spikes). Hence, before the threshold the trial-averaged inhibitory input from the CN neuron is zero, it begins to increase from the threshold and converges to the constant trial-averaged excitatory input through the US signal for the desired timing. Thus, with increasing the trial, $\mathcal{L}_p$ is zero before the threshold, it begins to increase from the threshold, and becomes saturated at $\mathcal{L}_p = 1$, as shown well in Fig. 12(a2). In this saturated case, the trial-averaged excitatory and inhibitory inputs to the IO are balanced.

We also investigate the firing activity of IO neuron during learning process. Figures 12(b1)-12(b6) and Figures 12(c1)-12(c6) show trial-evolutions of raster plots of spikes of the IO neuron (i.e., collection of spike trains for all the realizations; number of realizations is 100) and the bin-averaged instantaneous individual firing rates f_{IO} (i.e., the number of spikes of the IO neuron in a bin with the bin width $\Delta t = 40$ msec), respectively. Before the 141th threshold trial, relatively dense spikes appear in the middle part of the trial in the raster plot of spikes,

FIG. 13: Highly-connected ($p_c = 0.3$) and lowly-connected ($p_c = 0.003$) cases. Raster plots of spikes of 10^3 randomly chosen GR cells for (a1) $p_c = 0.3$ and (a2) $p_c = 0.003$. Instantaneous whole-population spike rates $R_{GR}(t)$ in the whole population of GR cells for (b1) $p_c = 0.3$ and (b2) $p_c = 0.003$. Band width for $R_{GR}(t)$: $h = 10$ msec. Distributions of conjunction indices $\{\mathcal{C}^{(I)}\}$ of the spiking patterns in the GR clusters in the whole population for (c1) $p_c = 0.3$ and (c2) $p_c = 0.003$. Bin size for the histograms in (c1) and (c2) is 0.1. Fractions of well-matched and ill-matched spiking groups for (d1) $p_c = 0.3$ and (d2) $p_c = 0.003$.

due to the effect of excitatory US signal. However, with increasing the trial from the threshold, spikes in the middle part become sparse, because of increased inhibitory input from the CN neuron. In this case, the bin-averaged instantaneous individual firing rate $f_{IO}(t)$ of the IO neuron forms a bell-shaped curve due to the US signal into the IO neuron. With increasing the trial from the 141th threshold, the amplitude of $f_{IO}(t)$ begins to decrease due to the increased inhibitory input from the CN neuron, and it becomes saturated at about the 250th trial. Thus, the trial-averaged individual firing rate $\overline{f_{IO}(t)}$ is constant (=1.5 Hz) before the threshold without the inhibitory input from the CN neuron. However, with increasing the trial from the threshold, it is decreased from 1.326 Hz due to increase in the inhibitory input from the CN neuron, and gets saturated at 0.0902 Hz at about the 250th trial, as shown in Fig. 12(d).

The firing output of the IO neuron is fed into the PCs via the CFs. Hence, with increasing the trial from the threshold, the error-teaching CF instructor signals become weaker and saturated at about the 250th cycle. While the saturated CF signals are fed into the PCs, trial-level balance between ΔLTD and ΔLTP occurs at

the PF-PC synapses, as shown in Figs. 9(c1) and 9(c2). As a result, saturation for the trial-averaged bin-averaged synaptic weights $\langle \overline{\tilde{J}(t)} \rangle$ appears [see Fig. 8(c)]. Then, the subsequent learning process in the PC-CN system also gets saturated, and we obtain the saturated learning efficiency degree $\mathcal{L}_e^*$ ($\simeq 11.19$), which is shown in Fig. 11(d).

E. Variation of The Connection Probability p_c and Strong Correlation between Diversity Degree $\mathcal{D}$ and Efficiency Degree $\mathcal{L}_e$ of CR

In the above subsections, we consider only the optimal case of $p_c^* = 0.029$ (i.e., 2.9%) where the spiking patterns of the GR clusters are the most diverse. From now on, we change the connection probability p_c from GO to GR cells, and study dependence of the diversity degree $\mathcal{D}$ for the spiking patterns in the GR clusters and the learning efficiency degree $\mathcal{L}_e$ of CR on p_c.

We first consider both the highly-connected case of $p_c = 0.3$ (i.e., 30%) and the lowly-connected case of $p_c = 0.003$ (i.e., 0.3%). Figures 13(a1) and 13(a2) show the raster plots of spikes of 10^3 randomly chosen GR cells for $p_c = 0.3$ and 0.003, respectively. The population-averaged firing activities in the whole population of GR cells may be well seen in the instantaneous whole-population spike rates $R_{GR}(t)$ in Figs. 13(b1) and 13(b2) for $p_c = 0.3$ and 0.003, respectively.

As shown in Fig. 2(b), each GR cluster is bounded by four glomeruli (corresponding to the terminals of the MFs) at both ends. Each glomerulus receives inhibitory inputs from nearby 81 GO cells with the connection probability p_c. In the highly-connected case of $p_c = 0.3$, on average, about 24 GO cell axons innervate each glomerulus. Then, each GR cell in a GR cluster receives about 97 inhibitory inputs via four dendrites which contact the four glomeruli at both ends. In this highly-connected case, inhibitory inputs from the pre-synaptic GO cells are increased, in comparison with the optimal case of $p_c^* = 0.029$. As a result, spike density in the raster plot of spikes is decreased (i.e., spikes become sparse) due to decreased individual firing rates, and hence the flat top part of $R_{GR}(t)$ becomes lowered, in comparison to the optimal case in Fig. 4(b).

In the highly-connected case of $p_c = 0.3$, differences between total inhibitory synaptic inputs from pre-synaptic GO cells to each GR cells are decreased due to increase in the number of pre-synaptic GO cells. In addition, the excitatory inputs into each GR cells via MFs are Poisson spike trains with the same firing rates, and hence they are essentially the same. Hence, differences between the total synaptic inputs (including both the inhibitory and the excitatory inputs) into each GR cells become reduced. These less different inputs into GR cells produce less different outputs (i.e. spiking activities) of GR cells, which become more similar to the population-averaged firing activity $R_{GR}(t)$ with a flat top in Fig. 13(b1). Thus, GR cells tend to exhibit relatively regular firings during the

whole trial stage ($0 < t < 1000$ msec), in comparison with optimal case of $p_c^* = 0.029$. Consequently, the raster plot of sparse spikes for $p_c = 0.3$ becomes relatively uniform [compare Fig. 13(a1) with Fig. 4(a)].

On the other hand, in the lowly-connected case of $p_c = 0.003$, the inhibitory inputs from GO cells into GR cells are so much reduced, and the excitatory MF signals into the GR cells become dominant inputs. Hence, spike density in the raster plot of spikes is increased (i.e., spikes become dense), because of increased individual firing rate, and the flat top part of $R_{GR}(t)$ becomes raised, in comparison to the optimal case in Fig. 4(b). Furthermore, differences between the total synaptic inputs into each GR cells become reduced, because the dominant excitatory MF signals, generated by the Poisson spike trains with the same firing rates, are essentially the same, and thus firing activities of GR cells become more similar to $R_{GR}(t)$ with a flat top in Fig. 13(b2). Hence, as in the highly-connected case, GR cells tend to show relatively regular spikings during the whole trial stage. As a result, the raster plot of dense spikes for $p_c = 0.003$ also becomes relatively uniform, as shown in Fig. 13(a2), in comparison with the optimal case in Fig. 4(a).

Figures 13(c1) and 13(c2) show the distributions of conjunction indices $\{\mathcal{C}^{(I)}\}$ for $p_c = 0.3$ and 0.003, respectively. The ranges in the distributions of $\{\mathcal{C}^{(I)}\}$ for $p_c = 0.3$ and 0.003 are (-0.21, 0.44) and (-0.18, 0.48), respectively. In both cases, their ranges are narrowed from both the positive and the negative sides, in comparison with the range (-0.49, 0.79) in the optimal case of $p_c^* = 0.029$. As explained above, in both the highly- and the lowly-connected cases of $p_c = 0.3$ and 0.003, GR cells tend to exhibit relatively regular spikings in the whole trial stage, due to decrease in the differences in the total synaptic inputs from GO cells into each GR cells, which is in contrast to the optimal case of $p_c^* = 0.029$ where random repetitions of transitions between bursting and silent states (both of which are persistent long-lasting ones) occur. Then, in both the highly- and the lowly-connected cases, highly well-matched spiking patterns with higher $\mathcal{C}^{(I)}$ and highly ill-matched spiking patterns with higher magnitude $|\mathcal{C}^{(I)}|$ disappear, which leads to reduction in the ranges of the distributions of $\{\mathcal{C}^{(I)}\}$ arise.

Due to the narrowed distribution of $\{\mathcal{C}^{(I)}\}$, both the mean ($\simeq 0.239$) and the standard deviation ($\simeq 0.360$) in the highly-connected case of $p_c = 0.3$ are decreased, in comparison to the optimal case where the mean and the standard deviation are 0.333 and 0.614, respectively. Then, the diversity degree $\mathcal{D}$ for the spiking patterns $[R_{GR}^{(I)}(t)]$ in the GR clusters, denoting a quantitative measure for diverse recoding in the granular layer, is given by the relative standard deviation for the distribution of $\{\mathcal{C}^{(I)}\}$ [see Eq. (27)]. For $p_c = 0.3$, its diversity degree is $\mathcal{D} \simeq 1.506$ which is smaller than $\mathcal{D}^* (\simeq 1.842)$ in the optimal case of $p_c^* = 0.029$. Similar to the highly-connected case, for $p_c = 0.003$ both the mean ($\simeq 0.272$) and the standard deviation ($\simeq 0.315$) for the distribution of $\{\mathcal{C}^{(I)}\}$ are also decreased. In this lowly-connected case,

FIG. 14: Change in firing activity of the CN neuron in the highly-connected ($p_c = 0.3$) and the lowly-connected ($p_c = 0.003$) cases. Case of $p_c = 0.3$: (a1)-(a6) raster plots of spikes of the CN neuron (i.e., collection of spike trains for all the realizations; number of realizations is 100) and (b1)-(b6) bin-averaged instantaneous individual firing rate $f_{CN}(t)$; the bin size is $\Delta t = 50$ msec. Case of $p_c = 0.003$: (c1)-(c6) raster plots of spikes of the CN neuron and (d1)-(d6) bin-averaged instantaneous individual firing rate $f_{CN}(t)$.

the diversity degree $\mathcal{D}$ for the spiking patterns $[R_{GR}^{(I)}(t)]$ in the GR clusters is $\mathcal{D} \simeq 1.157$ which is much smaller than $\mathcal{D}^* (\simeq 1.842)$ in the optimal case. We also note that the diversity degree $\mathcal{D}$ for $p_c = 0.003$ is smaller than that for $p_c = 0.3$; $\mathcal{D}^* = 1.842$ ($p_c^* = 0.029$) $> \mathcal{D} = 1.506$ ($p_c = 0.3$) $> \mathcal{D} = 1.157$ ($p_c = 0.003$).

Figures 13(d1) and 13(d2) show fractions of well-matched ($\{\mathcal{C}^{(I)} > 0\}$) and ill-matched ($\{\mathcal{C}^{(I)} < 0\}$) spiking groups for $p_c = 0.3$ and 0.003, respectively. In the highly-connected case of $p_c = 0.3$, the well-matched spiking group is a major one with fraction 88.2%, while the ill-matched spiking group is a minor one with fraction 11.8%. In comparison with the optimal case where the fraction of well-matched spiking group is 82.1%, the fraction of well-matched spiking group for $p_c = 0.3$ is increased. In this highly-connected case, the spiking-group ratio, given by the ratio of the fraction of the well-matched spiking group to that of the ill-matched spiking group, is $\mathcal{R}_{sp} \simeq 7.48$ which is larger than that ($\mathcal{R}_{sp}^* \simeq 4.59$) in the optimal case. In the lowly-connected case of $p_c = 0.003$, the fraction of well-matched spiking group is more increased to 93.9%. Hence, the spiking-group ratio is $\mathcal{R}_{sp} \simeq 13.8$ which is much larger than that in the optimal case.

Due to decrease in differences between the total synaptic inputs into each GR cells, firing activities of GR cells for $p_c = 0.3$ and 0.003 become more similar to the population-averaged firing activity $R_{GR}(t)$, in comparison with the optimal case of $p_c^* = 0.029$. We note that $R_{GR}(t)$ is well-matched with the US signal $f_{US}(t)$ [i.e, $R_{GR}(t)$ has a positive conjunction index with respect to $f_{US}(t)$], which results in increase in the fraction of well-matched spiking group for $p_c = 0.3$ and 0.003. In contrast to the case of $p_c = 0.3$, in the lowly-connected case

of $p_c = 0.003$, inhibitory inputs into each GR cells are so much reduced, and hence the dominant inputs are just the excitatory MF signals which are well-matched with the US signal $f_{US}(t)$. Thus, the fraction of well-matched spiking group for $p_c = 0.003$ becomes larger than that for $p_c = 0.3$.

These changes in the diversity degree $\mathcal{D}$ of the spiking patterns in the GR clusters have direct effect on the synaptic plasticity at the PF-PC synapses and the subsequent learning process in the PC-CN system. As shown in the optimal case of $p_c^* = 0.029$, the ill- and the well-matched spiking groups play their own roles for the CR. The ill-matched spiking group plays a role of protection barrier for the timing of CR, while the strength of CR is determined by strong LTD in the well-matched spiking group. Due to break-up of highly well-matched and highly ill-matched spiking patterns, the distributions of $\{\mathcal{C}^{(I)}\}$ for the highly- and the lowly-connected cases of $p_c = 0.3$ and 0.003 are narrowed. Hence, both the timing degree $\mathcal{T}_d$ and the strength $\mathcal{S}$ of CR are decreased for both $p_c = 0.3$ and 0.003, which are well shown in Figs. 14 and 15.

Figure 14 shows change in the firing activity of the CN neuron which generates the final output of the cerebellum during learning trial in the highly- and the lowly-connected cases of $p_c = 0.3$ and 0.003. For $p_c = 0.3$, trial-evolutions of the raster plots of spikes of the CN neuron (i.e., collection of spike trains for all the realizations; number of realizations is 100) and the bin-averaged instantaneous individual firing rates $f_{CN}(t)$ (i.e., the number of spikes of the CN neuron in a bin with the bin width $\Delta t = 50$ msec) are shown in Figs. 14(a1)-14(a6) and Figs. 14(b1)-14(b6), respectively. In this highly-connected case, at the 142th threshold trial, the CN neuron begins to fire in the middle part of the trial. Thus, acquisition of CR occurs a little later, in comparison with the optimal case of $p_c^* = 0.029$ with the 141th threshold. In this case, $f_{CN}(t)$ forms a bell-shaped curve. With increasing the trial from the threshold, raster plots of spikes of the CN neuron become more and more dense in the middle part of each trial, and bottom-base width and peak height of the bell curve for $f_{CN}(t)$ increase. At about the 250th trial, $f_{CN}(t)$ seems to become saturated.

For $p_c = 0.3$, the bottom-base width of the bell curve is wider and its peak height is shorter, in comparison with the optimal case (see Fig. 11). Due to break-up of highly ill-matched spiking patterns (which play the role of protection barrier for timing of CR), bottom-base width (associated with the reciprocal of timing degree of CR) of the bell increases. Also, peak height of the bell (related to the strength of CR) decreases because of break-up of highly well-matched spiking patterns (which induce strong LTD and determine the strength of CR). Consequently, as the diversity degree $\mathcal{D}$ of the spiking patterns in the GR clusters is deceased from 1.842 ($p_c^* = 0.029$) to 1.507 ($p_c = 0.3$), the bottom-base width of the bell curve is increased, and the peak height is decreased.

For $p_c = 0.003$, trial-evolutions of the raster plots

FIG. 15: Timing degree, strength, and learning efficiency degree of the CR in the highly-connected ($p_c = 0.3$) and the lowly-connected ($p_c = 0.003$) cases. (a) Plots of timing degree $\mathcal{T}_d$ of the CR versus trial. (b) Plots of strengths $\mathcal{S}$ of the CR versus trial. (c) Plots of learning efficiency degree $\mathcal{L}_e$ for the CR versus trial. Solid circles, open circles, and crosses represent data in the cases of $p_c = 0.3$, 0.003, and 0.029, respectively.

of spikes of the CN neuron and the bin-averaged instantaneous individual firing rates $f_{CN}(t)$ are shown in Figs. 14(c1)-14(c6) and Figs. 14(d1)-14(d6), respectively. In this lowly-connected case, acquisition of CR occurs at the 143th trial which is a little later in comparison with the 142th threshold for $p_c = 0.3$. Similar to the highly-connected case, with increasing the trial from the threshold, raster plots of spikes of the CN neuron become more and more dense in the middle part of each trial, and the bottom-base width and the peak height of the bell curve for $f_{CN}(t)$ increase. Eventually, saturation occurs at about the 250th trial. In comparison to the highly-connected case, the bottom-base width of the bell curve is wider and its peak height is shorter, because of more break-up of highly ill-matched spiking patterns and highly well-matched spiking patterns (which results in more decrease in the diversity degree of spiking patterns). As a result, with decreasing the diversity degree $\mathcal{D}$ from 1.507 ($p_c = 0.3$) to 1.157 ($p_c = 0.003$), the bottom-base width of the bell curve increases, and the peak height decreases (i.e., less diversity in the spiking patterns results in decrease in the timing degree and the strength of CR).

Figures 15(a) and 15(b) show plots of the timing degree $\mathcal{T}_d$ and the strength $\mathcal{S}$ of CR versus trial for $p_c = 0.3$ (solid circles), 0.003 (open circles), and 0.029 (crosses), respectively. The timing degree $\mathcal{T}_d$, denoting the matching degree between the firing activity of the CN neuron [$f_{CN}(t)$] and the US signal $f_{US}(t)$ for a timing, is given by the cross-correlation $Corr_T(0)$ at the zero lag between $f_{CN}(t)$ and $f_{US}(t)$ in Eq. (36). This timing degree $\mathcal{T}_d$ reflects the width of the bottom base of the bell curve. With increasing the trial, the width of the bottom base increases, as shown in Fig. 14, and hence $\mathcal{T}_d$ decreases monotonically, and it becomes saturated at about the 250th trial. We note that, as the diversity degree $\mathcal{D}$ of the spiking patterns is decreased

FIG. 16: Strong correlation between the diversity degree $\mathcal{D}$ and the saturated learning efficiency degree $\mathcal{L}_e^*$. (a) Fractions of well-matched (solid circles) and ill-matched (open circles) spiking groups versus the connection probability p_c. (b) Plot of diversity degree $\mathcal{D}$ of the spiking patterns in the GR clusters versus p_c. Plots of saturated (c1) timing degree $\mathcal{T}_d^*$, (c2) strengths $\mathcal{S}^*$, and (c3) learning efficiency degree $\mathcal{L}_e^*$ of the CR versus p_c. (d) Plot of $\mathcal{L}_e^*$ versus $\mathcal{D}$.

($\mathcal{D} = 1.507$, 1.157, and 1.842 for $p_c = 0.3$, 0.003, and 0.029, respectively), $\mathcal{T}_d^*$ (saturated value of $\mathcal{T}_d$) decreases; $\mathcal{T}_d^* = 0.346$ ($p_c^* = 0.029$) $> \mathcal{T}_d^* = 0.266$ ($p_c = 0.3$) $> \mathcal{T}_d^* = 0.187$ ($p_c = 0.003$).

On the other hand, as the trial is increased, the peak height of the bell increases. Hence, the strength $\mathcal{S}$ of CR, given by the modulation of $f_{\mathrm{CN}}(t)$, increases monotonically, and it becomes saturated at about the 250th trial. In this case, $\mathcal{S}^*$ (saturated value of $\mathcal{S}$) also is decreased with decrease in the diversity degree $\mathcal{D}$; $\mathcal{S}^* = 32.382$ ($p_c^* = 0.029$) $> \mathcal{S}^* = 27.099$ ($p_c = 0.3$) $> \mathcal{S}^* = 21.656$ ($p_c = 0.003$).

We then consider the learning efficiency degree $\mathcal{L}_e$ for the CR, given by product of the timing degree $\mathcal{T}_d$ and the strength $\mathcal{S}$ in Eq. (37). Figure 15(c) shows a plot of $\mathcal{L}_e$ versus trial. $\mathcal{L}_e$ increases monotonically from the threshold trial, and it becomes saturated at about the 250th cycle. Thus, we obtain the saturated learning efficiency degree $\mathcal{L}_e^*$, the values of which are 7.216, 4.054, and 11.19 for $p_c = 0.3$, 0.003, and 0.029, respectively. Among the three cases, $\mathcal{L}_e^*$ ($= 11.19$) in the optimal case is the largest one, and $\mathcal{L}_e^*$ is decreased with decrease in the diversity degree $\mathcal{D}$; $\mathcal{L}_e^* = 11.19$ ($p_c^* = 0.029$) $> \mathcal{L}_e^* = 7.216$ ($p_c = 0.3$) $> \mathcal{L}_e^* = 4.054$ ($p_c = 0.003$).

Finally, based on the above two examples for the highly- and the lowly-connected cases, we investigate dependence of the diversity degree $\mathcal{D}$ for the spiking patterns of the GR clusters and the saturated learning efficiency degree $\mathcal{L}_e^*$ on p_c by varying it from the optimal value ($p_c^* = 0.029$). Figure 16(a) shows plots of fractions of the well- and the ill-matched spiking groups versus p_c. The fraction of the well-matched spiking group (solid cir-

cles) forms a well-shaped curve with a minimum in the optimal case of p_c^* ($= 0.029$), while the fraction of the ill-matched spiking group (open circles) forms a bell-shaped curve with a maximum in the optimal case. In the optimal case of $p_c^* = 0.029$, the spiking-group ratio (i.e., ratio of fraction of the well-matched spiking group to fraction of the ill-matched spiking group) is $\mathcal{R}_{\mathrm{sp}}^* = 4.59$. As p_c is changed (i.e., increased or decreased) from p_c^*, the fraction of the in-phase spiking group increases, and then the spiking-group ratio $\mathcal{R}_{\mathrm{sp}}$ increases from $\mathcal{R}_{sp}^*$ in the optimal case.

Figure 16(b) show a plot of the diversity degree $\mathcal{D}$ for the spiking patterns in the GR clusters. The diversity degrees $\mathcal{D}$ forms a bell-shaped curve with a maximum $\mathcal{D}^*$ ($\simeq 1.842$) in the optimal case of $p_c^* = 0.029$. With changing p_c from p_c^*, $\mathcal{D}$ decreases from $\mathcal{D}^*$. Hence, in the optimal case, temporal recoding of GR cells is the most diverse. Figures 16(c1)-16(c3) show plots of the saturated timing degree $\mathcal{T}_d^*$, the saturated strength $\mathcal{S}^*$, and the saturated learning efficiency degree $\mathcal{L}_e^*$ of CR, respectively. All of them form bell-shaped curves with maxima $\mathcal{T}_d^*$ ($\simeq 0.346$), $\mathcal{S}^*$ ($\simeq 32.38$), and $\mathcal{L}_e^*$ ($\simeq 11.19$) in the optimal case of $p_c^* = 0.029$. As p_c is changed from p_c^*, $\mathcal{T}_d^*$, $\mathcal{S}^*$, and $\mathcal{L}_e^*$ are decreased. Figure 16(d) shows a plot of $\mathcal{L}_e^*$ versus $\mathcal{D}$. As shown clearly in Fig. 16(d), both $\mathcal{L}_e^*$ and $\mathcal{D}$ have a strong correlation with the Pearson's correlation coefficient $r \simeq 0.9982$. Consequently, the more diverse in temporal recoding of the GR cells, the more effective in motor learning for the Pavlovian EBC.

IV. SUMMARY AND DISCUSSION

We are concerned about the Pavlovian EBC. Diverse experimental works on the EBC have been done in many mammalian species such as humans, monkeys, dogs, ferrets, rabbits, rats, and mice [14–18, 24–27]. Also, computational works reproduced some features (e.g., representation of time passage) of the EBC in artificial models [39–44], a realistic biological model [45–47], a rate-coding model [48], and a spiking neural network model [49]. However, effects of diverse temporal recoding in GR clusters on the EBC in previous computational works are necessary to be more clarified in several dynamical aspects. Particularly, the previous computational works lacked complete dynamical classification of diverse spiking patterns in the GR clusters and their conjunction with the error-teaching CF signals.

To the best of our knowledge, for the first time, we made complete quantitative classification of diverse spiking patterns in the GR clusters in terms of the newly-introduced conjunction index $\{\mathcal{C}^{(I)}\}$ and diversity degree $\mathcal{D}$ in the case of Pavlovian EBC. Each spiking pattern is characterized in terms of its conjunction index $\mathcal{C}^{(I)}$, representing the matching degree between the spiking pattern and the US signal for the desired timing. Then, the whole spiking patterns are clearly decomposed into the well-matched ($\mathcal{C}^{(I)} > 0$) and the ill-matched ($\mathcal{C}^{(I)} < 0$)

spiking groups. Furthermore, the degree of diverse re-coding of the GR cells may be quantified in terms of the diversity degree $\mathcal{D}$, given by the relative standard deviation in the distribution of $\{\mathcal{C}^{(I)}\}$. Thus, $\mathcal{D}$ provides a quantitative measure for diverse temporal recoding of GR cells. It is also shown that diverse total synaptic inputs (including both the excitatory inputs via MFs and the inhibitory inputs from the pre-synaptic GO cells) into the GR clusters result in generation of diverse spiking patterns (i.e. outputs) in the GR clusters.

Based on the above dynamical classification of diverse spiking patterns in the GR clusters, we made clear investigations on the effect of diverse recoding of GR cells on the Pavlovian EBC (i.e., their effect on the synaptic plasticity at the PF-PC synapses and the subsequent learning process in the PC-CN-IO system). To the best of our knowledge, this kind of approach, based on the well- and the ill-matched spiking groups, is unique in studying the Pavlovian EBC. The well-matched spiking patterns are strongly depressed by the error-teaching CF signals, and they make dominant contributions in the middle part of each trial. In contrast, for the ill-matched spiking patterns with central gaps in the middle part, practically no LTD occurs due to no conjunction with the CF signals. Thus, in the middle part of each trial, strong LTD occurs via dominant contributions of well-matched spiking group. On the other hand, at the initial and the final parts of each trial, less LTD takes place because both the ill-matched spiking group with practically no LTD and the well-matched spiking group make contributions together. As a result, a big modulation in bin-averaged synaptic weight $\langle \tilde{J} \rangle$ arises via interplay of the well- and ill-matched spiking groups.

Due to this type of effective synaptic plasticity at the PF-PC synapses, the (realization-averaged) population spike rate $\langle R_{\text{PC}}(t) \rangle$ of PCs forms a step-well-shaped curve with a minimum in the middle part of each trial. When passing a threshold trial, a "zero-bottom" (where $\langle R_{\text{PC}}(t) \rangle \simeq 0$) appears in the central well. At this threshold trial, the CN neuron begins to fire in the middle part of trial. Hence, appearance of the zero-bottom in the step-well for $\langle R_{\text{PC}}(t) \rangle$ is a prerequisite condition for acquisition of CR. In the subsequent trials, the individual firing rate $f_{\text{CN}}(t)$ of the CN neuron forms a bell-shaped curve with a maximum in the middle part (which is up-down reversed with respect to $\langle R_{\text{PC}}(t) \rangle$). Outside the bottom of the bell, the CN neuron cannot fire, due to inhibition of ill-matched spiking group (with practically no LTD). Hence, the ill-matched spiking group plays a role of protection barrier for timing, and the timing degree of CR is reciprocally associated with the bottom width of the bell. In this case, the peak of the bell in the middle part is formed due to strong LTD in the well-matched spiking group, and its height is directly related to the strength of CR (corresponding to the amplitude of eyelid closure). In this way, both the well- and the ill-matched spiking groups play their own roles for the timing and the strength of CR, respectively.

By changing p_c, we investigated dependence of the diversity degree $\mathcal{D}$ of the spiking patterns and the saturated learning efficiency degree $\mathcal{L}_e^*$ for the CR (given by the product of the timing degree and the strength of CR) on p_c. Both $\mathcal{D}$ and $\mathcal{L}_e$ have been found to form bell-shaped curves with peaks at the same optimal value of $p_c^* = 0.029$, and they also have a strong correlation. Consequently, the more diverse in the spiking patterns of GR cells, the more efficient in motor learning for the Pavlovian EBC. This main result in our work seems to be examined in an experiment suggested in the below.

From now on, we summarize our results in more details. In this work, we mainly focused on an optimal case of p_c^* ($= 0.029$) where the spiking patterns of GR clusters are the most diverse. The instantaneous whole-population spike rate $R_{\text{GR}}(t)$ describes well collective firing activity in the whole population of GR cells (receiving tone CS inputs via MFs). It is basically in proportion to the CS signal, but its amplitude is uniformly lowered due to the effect of inhibitory GO cells. This whole population of GR cells is divided into 2^{10} GR clusters. Due to the shared excitatory and inhibitory inputs, GR cells in each Ith GR cluster ($I = 1, \cdots, 2^{10}$) show similar spiking behaviors, and their averaged firing activity is described by the instantaneous cluster spike rate $R_{\text{GR}}^{(I)}(t)$ in the Ith GR cluster. Then, GR clusters exhibit diverse spiking patterns for $R_{\text{GR}}^{(I)}(t)$ which are well- and ill-matched ones with the airpuff US signal.

The diverse spiking patterns are grouped into the well-matched and the ill-matched spiking groups. The fraction of well-matched (ill-matched) spiking group is 82.1 (17.9) %, and hence the spiking-group ratio (given by the ratio of the fraction of well-matched spiking group to that of the ill-matched spiking group) is $\mathcal{R}_{\text{sp}}^* = 4.59$. In this case, the spiking pattern of each Ith GR cluster has been quantitatively characterized in terms of its conjunction index $\mathcal{C}^{(I)}$, given by the cross-correlation at the zero-time lag between the instantaneous cluster spike rate $R_{\text{GR}}^{(I)}(t)$ and the firing rate $f_{\text{US}}(t)$ of the airpuff US signal, representing the matching degree between the spiking behavior $[R_{\text{GR}}^{(I)}(t)]$ of each Ith GR cluster and the US signal $f_{\text{US}}(t)$. The distribution for $\{\mathcal{C}^{(I)}\}$ forms a bell-shaped curve. Here, spiking patterns in the well-matched spiking group have positive values of $\mathcal{C}^{(I)}$, while those in the ill-matched spiking group have negative values of $\mathcal{C}^{(I)}$. Then, the diversity of spiking patterns of all the GR clusters is quantitatively characterized in terms of the diversity degree $\mathcal{D}$, given by the relative standard deviation of the distribution $\{\mathcal{C}^{(I)}\}$. In the optimal case of $p_c^* = 0.029$, $\mathcal{D}^* \simeq 1.842$, which is a quantitative measure for the diversity of spiking patterns in the GR clusters. We also made investigation on dynamical origin of diverse spiking patterns in the GR clusters. It is thus found that diverse total synaptic inputs (including both the excitatory inputs via MFs and the inhibitory inputs from the pre-synaptic GO cells) into the GR clusters lead to production of diverse spiking patterns (i.e. outputs) in the

GR clusters.

Next, based on the dynamical classification of diverse spiking patterns in the GR clusters in the optimal case of $p_c^* = 0.029$, we made an intensive investigation on their effects on synaptic plasticity at PF-PC synapses and subsequent learning process in the PC-CN-IO system. During the whole training trials, well-matched and ill-matched spiking groups have been found to play their own roles, respectively. Both the diversely-recoded PF (student) signals from GR cells and the error-teaching CF (instructor) signals from the IO are fed into the PCs. We note that the CF signals are also well-matched with the US signal $f_{US}(t)$.

For the ill-matched PF signals (with negative conjunction indices), practically no LTD occurs because most of them have no conjunctions with the error-teaching CF signals which are strongly localized in the middle of each trial (i.e., near $t = 500$ msec). Hence, synaptic weights at the PF-PC synapses form a top horizontal line with a central gap in each trial. On the other hand, well-matched PF signals (with positive conjunction indices) have been found to be strongly depressed by the CF signals. Thus, synaptic weights at the PF-PC synapses form a lower band; with increasing the trial, this band becomes lowered due to strong LTD. In this way, the distribution of synaptic weights at the PF-PC synapses consists of two distinctly separated structures (i.e., a combination of separated top horizontal line with a central gap and lower band). Consequently, the bin-averaged synaptic weights of active PF signals have been found to form a step-well-shaped curve (i.e., appearance of a minimum in the middle part). In this way, at the PF-PC synapses, diversely-recoded PF student signals are effectively depressed by the error-teaching CF instructor signals.

Due to this kind of effective depression at PF-PC synapses, the (realization-averaged) instantaneous population spike rate $\langle R_{PC}(t) \rangle_r$ of PCs (corresponding to the principal outputs of the cerebellar cortex) has also been found to form a step-well-shaped curve with a minimum in the middle part of each trial. With increasing the learning trial, the whole step-well curve becomes lowered (i.e., PCs learn when to stop inhibition), and the top width and the depth of the well show increasing tendency (i.e., the cental well becomes wider and deeper). Eventually, at the 141th threshold trial, $\langle R_{PC}(t) \rangle_r$ at the bottom of the well becomes zero (i.e., a "zero-bottom" of the step-well appears). We note that these PCs exert effective inhibitory coordination on the CN neuron [which evokes the eyelid CR (i.e., learned eyeblink)]. Before the threshold trial, the CN neurons can not fire spikes, due to strong inhibition from the PCs. At the threshold trial, the CN neuron begins to fire in the middle part of trial, due to the presence of zero-bottom of the step-well for $\langle R_{PC}(t) \rangle_r$ where PCs supply no inhibition to the CN neuron. Hence, the appearance of zero-bottom of the step-well is the prerequisite condition for the acquisition of CR. In the subsequent learning trials following the threshold, the (realization-averaged) instantaneous indi-

vidual firing rate f_{CN} of the CN neuron has been found to form a bell-shaped curve with a maximum in the middle part; with increasing the trials, the bottom width and the peak-height of the bell tend to increase.

At the threshold trial, the timing degree $\mathcal{T}_d$ of CR, given by the cross-correlation at the zero-time lag between f_{CN} and $f_{US}(t)$, becomes good due to presence of ill-matched spiking group which plays a role of protection barrier for the timing. With further increasing the trial, strength $\mathcal{S}$ of CR [representing the amplitude of eyelid closure (measured by the EMG)], given by the modulation of f_{CN}, has been found to increase due to strong LTD in the well-matched spiking group, while its timing degree $\mathcal{T}_d$ has been found to decrease. In this way, the ill- and the well-matched spiking groups have been found to play their own roles for the timing and the strength of CR, respectively. Thus, the (overall) learning efficiency degree $\mathcal{L}_e$ for the CR, given by product of $\mathcal{T}_d$ and $\mathcal{S}$, has been found to increase with learning trial and to be saturated at about the 250th trial. The saturated learning efficiency degree in the optimal case of $p_c^* = 0.029$ was $\mathcal{L}_e^* (\simeq 11.19)$.

During the learning trial, the IO neuron receives the inhibitory input from the CN neuron and the excitatory US signal. In this case, learning progress is characterized in terms of the learning progress degree $\mathcal{L}_p$, given by the ratio of the time-averaged inhibitory input from the CN neuron to the magnitude of the time-averaged excitatory input of the US signal. As the learning trial is increased, the time-averaged inhibitory input increases, and converges to the constant time-averaged excitatory input. Thus, with increasing the learning trial, the learning progress degree $\mathcal{L}_p$ has been found to increase and eventually become saturated at $\mathcal{L}_p = 1$ at about the 250th cycle. In this saturated case, the time-averaged excitatory and inhibitory inputs to the IO neuron become balanced.

One of main purposes for our study is to investigate the effect of the connection probability p_c (from GO to GR cells) on diverse recoding of GR cells. By changing (i.e., increasing or decreasing) p_c from its optimal value $p_c^* (= 0.029)$, the fraction of well-matched spiking group has been found to increase. Thus, the spiking-group ratio $\mathcal{R}_{sp}$ (i.e., the ratio of the fraction of well-matched spiking group to that of the ill-matched spiking group) has a minimum value of $\mathcal{R}_{sp}^* (= 4.59)$ for $p_c^* = 0.029$. In this optimal case, each GR cell receives inhibitory inputs from about 9 nearby GO cells, and spiking patterns in the GR clusters are the most diverse. In Refs. [4, 49] where the parameter values were taken, based on physiological data, the average number of nearby GO cell axons innervating each GR cell is about 8, which is very close to that in the optimal case. Thus, we hypothesize that the granular layer in the cerebellar cortex has evolved toward the goal of the most diverse recoding.

As the spiking-group ratio $\mathcal{R}_{sp}$ is increased from the minimum $\mathcal{R}_{sp}^*$, the diversity degree $\mathcal{D}$ for the spiking patterns in the GR clusters decreases because of increase in

the fraction of well-matched spiking group. Thus, the curve of $\mathcal{D}$ has been found to form a bell-shaped curve with a maximum $\mathcal{D}^*(\simeq 1.842)$ at $p_c = p_c^*$. Consequently, in the optimal case ($p_c^* = 0.029$), spiking patterns in the GR clusters are the most diverse. In this case, a plot of saturated learning efficiency degree $\mathcal{L}_e^*$ for the CR versus p_c has also been found to form a bell-shaped curve with a peak $\mathcal{L}_e^*(\simeq 11.19)$ for $p_c = p_c^*$. Moreover, both the saturated learning efficiency degree $\mathcal{L}_e^*$ and the diversity degree $\mathcal{D}$ have a strong correlation with the Pearson's correlation coefficient $r \simeq 0.9982$. As a result, the more diverse in temporal recoding of GR cells, the more effective in the motor learning for the Pavlovian EBC, which is our main result.

To examine the main result in our work, we also suggest a real experiment for the EBC. To control p_c in a given species of animals (e.g., a species of rabbit, rat, or mouse) in an experiment seems to be practically difficult, in contrast to the case of computational neuroscience where p_c may be easily varied. Instead, we may consider an experiment for several species of animals (e.g., 3 species of rabbit, rat, and mouse). In each species, a large number of randomly chosen GR cells ($i = 1, \cdots, M$) are considered. Then, through many CS-US trials, one may get the peristimulus time histogram (PSTH) for each ith GR cell [i.e., (bin-averaged) instantaneous individual firing rate $f_{\text{GR}}^{(i)}(t)$ of the ith GR cell]. GR cells are expected to exhibit diverse PSTHs. Then, in the case of each ith GR cell, one can get its conjunction index $\mathcal{C}_i$, representing the similarity degree between its PSTH $f_{\text{GR}}^{(i)}(t)$ and the CF signal for the desired timing [i.e., the PSTH of the IO neuron $f_{\text{IO}}(t)$]. In this case, the conjunction index $\mathcal{C}_i$ is given by the cross-correlation at the zero-time lag between $f_{\text{GR}}^{(i)}(t)$ and $f_{\text{IO}}(t)$. Thus, one may get the diversity degree $\mathcal{D}$ of PSTHs of GR cells, given by the relative standard deviation in the distribution of $\{\mathcal{C}_i\}$, for the species.

In addition to the PSTHs of GR cells, under the many CS-US trials, one can also obtain a bell-shaped PSTH of a CN neuron [(bin-averaged) instantaneous individual firing rate $f_{\text{CN}}(t)$ of the CN neuron]. The reciprocal of bottom-base width and the peak height of the bell curve correspond to timing degree $\mathcal{T}_d$ and strength $\mathcal{S}$ for the EBC, respectively. In this case, the (overall) learning efficiency degree $\mathcal{L}_e$ for the EBC is given by the product of $\mathcal{T}_d$ and $\mathcal{S}$. In this way, a set of $(\mathcal{D}, \mathcal{L}_e)$ may be experimentally obtained for each species, and depending on the species, the set of $(\mathcal{D}, \mathcal{L}_e)$ may change. Then, for example in the case of 3 species of rabbit, rat, and mouse, with the three different data sets for $(\mathcal{D}, \mathcal{L}_e)$, one can examine our main result (i.e., whether more diversity in PSTHs of GR cells leads to more efficient motor learning for the EBC).

Finally, we discuss limitations of our present work and future works. In the present work, we investigated the effect of diverse temporal recoding of GR cells on the Pavlovian EBC. In the present work, we set the ISI between the onsets of CS and US at 500 msec. The acquisi-

TABLE I: Parameter values for LIF neuron models with AHP currents for the granule (GR) cell and the Golgi (GO) cell in the granular layer, the Purkinje cell (PC) and the basket cell (BC) in the Purkinje-molecular layer, and the cerebellar nucleus (CN) and the inferior olive (IO) neurons.

X-population		Granular Layer		Purkinje-Molecular Layer		CN neuron	IO neuron
		GR cell	GO cell	PC	BC		
C_X		3.1	28.0	107.0	107.0	122.3	10.0
$I_L^{(X)}$	$g_L^{(X)}$	0.43	2.3	2.32	2.32	1.63	0.67
	$V_L^{(X)}$	-58.0	-55.0	-68.0	-68.0	-56.0	-60.0
$I_{AHP}^{(X)}$	$\bar{g}_{AHP}^{(X)}$	1.0	20.0	100.0	100.0	50.0	1.0
	$\tau_{AHP}^{(X)}$	5.0	5.0	5.0	2.5	2.5	10.0
	$V_{AHP}^{(X)}$	-82.0	-72.7	-70.0	-70.0	-70.0	-75.0
	$v_{th}^{(X)}$	-35.0	-52.0	-55.0	-55.0	-38.8	-50.0
$I_{ext}^{(X)}$		0.0	0.0	250.0	0.0	0.0	0.0

tion rate and the timing degree and strength of CR have been known to depend on the ISI [49, 85]. Hence, in a future work, it would be interesting to study dependence of EBC on the ISI. Based on the results of our work, it would also be interesting to study extinction of CR, as a future work. After acquisition of CR, we turn off the air-puff US. Then, the CR is expected to become gradually extinct via LTP at the PF-PC synapses [7]. In this work, we considered only the synaptic plasticity at the PF-PC synapses. In the cerebellum, diverse synaptic plasticity occurs at other synapses [86, 87] (e.g., MF-CN and PC-CN synapses [88], PF-BC and BC-PC synapses [89], and MF-GR cells synapses [90]). Hence, as a future work, it would be interesting to study the effect of diverse synaptic plasticity at other synapses on the cerebellar motor learning for the Pavlovian EBC; particularly, we are interested in the effect of the synaptic plasticity at the MF-CN synapse on the EBC [7]. In addition to change in p_c (i.e., connection probability from GO to GR cells), another possibility to vary synaptic inputs into the GR cells is to change NMDA receptor-mediated maximum conductances $\bar{g}_{\text{NMDA}}^{(\text{GR})}$ and $\bar{g}_{\text{NMDA}}^{(\text{GO})}$, related to persistent long-lasting firing activities [49]. It would also be interesting to study the effect of NMDA receptor-mediated synaptic inputs on diverse recoding of GR cells and motor learning for the EBC by varying $\bar{g}_{\text{NMDA}}^{(\text{GR})}$ and $\bar{g}_{\text{NMDA}}^{(\text{GO})}$. This work is beyond the scope of the present work, and hence it is left as a future work. In the present work, we considered just inter-population synaptic connections, and hence no motor rhythms appear due to absence of intra-population synaptic connections. When we add intra-population couplings, diverse motor rhythms appear; a granular motor rhythm of $7 \sim 25$ Hz in the granular layer [91], ultrafast motor rhythm of ~ 200 Hz in the Purkinje (molecular) layer [92], and ~ 10 Hz motor rhythm in the system of IO neurons [93]. Hence, in a future work, it

TABLE II: Parameter values for synaptic currents $I_R^{(T,S)}(t)$ into the granule (GR) and the Golgi (GO) cells in the granular layer, the Purkinje cells (PCs) and the basket cells (BCs) in the Purkinje-molecular layer, and the cerebellar nucleus (CN) and the inferior olive (IO) neurons in the other parts. In the granular layer, the GR cells receive excitatory inputs via mossy fibers (MFs) and inhibitory inputs from GO cells, and the GO cells receive excitatory inputs via parallel fibers (PFs) from GR cells. In the Purkinje-molecular layer, the PCs receive two types of excitatory inputs via PFs from GR cells and through climbing fibers (CFs) from the IO and one type of inhibitory inputs from the BCs. The BCs receive excitatory inputs via PFs from GR cells. In the other parts, the CN neuron receives excitatory inputs via MFs and inhibitory inputs from PCs, and the IO neuron receives excitatory input via the US signal and inhibitory input from the CN neuron.

Target Cells (T)	Granular Layer					Purkinje-Molecular Layer				Other Parts				
	GR			GO		PC			BC	CN			IO	
Source Cells (S)	MF	MF	GO	PF	PF	PF	CF	BC	PF	MF	MF	PC	US	CN
Receptor (R)	AMPA	NMDA	GABA	AMPA	NMDA	AMPA	AMPA	GABA	AMPA	AMPA	NMDA	GABA	AMPA	GABA
$\bar{g}_R^{(T)}$	0.18	0.025	0.028	45.5	30.0	0.7	0.7	1.0	0.7	50.0	25.8	30.0	1.0	0.18
$J_{ij}^{(T,S)}$	8.0	8.0	10.0	0.00004	0.00004	0.006	1.0	5.3	0.006	0.002	0.002	0.008	1.0	5.0
$V_R^{(S)}$	0.0	0.0	-82.0	0.0	0.0	0.0	0.0	-75.0	0.0	0.0	0.0	-88.0	0.0	-75.0
$\tau_R^{(T)}$	1.2	52.0	7.0, 59.0	1.5	31.0, 170.0	8.3	8.3	10.0	8.3	9.9	30.6	42.3	10.0	10.0
A_1, A_2			0.43, 0.57		0.33, 0.67									

would be interesting to study the effect of motor rhythms on diverse recoding of GR cells and motor learning for the EBC by adding intra-population couplings.

Acknowledgments

This research was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education (Grant No. 20162007688).

Appendix: Parameter Values for The LIF Neuron Models and The Synaptic Currents

In this appendix, we list four tables which show parameter values for the LIF neuron models in Subsec. II C and the synaptic currents in Subsec. II D. These values are adopted from physiological data [4, 49].

For the LIF neuron models, the parameter values for the capacitance C_X, the leakage current $I_L^{(X)}$, the AHP current $I_{AHP}^{(X)}$, and the external constant current $I_{ext}^{(X)}$ are shown in Table I.

For the synaptic currents, the parameter values for the maximum conductance $\bar{g}_R^{(T)}$, the synaptic weight $J_{ij}^{(T,S)}$, the synaptic reversal potential $V_R^{(S)}$, the synaptic decay time constant $\tau_R^{(T)}$, and the amplitudes A_1 and A_2 for the type-2 exponential-decay function in the granular layer, the Purkinje-molecular layer, and the other parts for the CN and IO are shown in Table II.

[1] M. Ito, *The Cerebellum and Neural Control* (Raven Press, New York, 1984).

[2] M. Ito, Ann. N. Y. Acad. Sci. **978**, 273 (2002).

[3] M. Ito, *The Cerebellum: Brain for an Implicit Self* (Pearson Education Inc., New Jersey, 2012).

[4] T. Yamazaki and S. Nagao, PLoS One **7**, e33319 (2012).

[5] I. Gormezano, E. J. Kehoe, and B. S. Marshall, Prog. Psychobio. Physiol. Psychol. **10**, 197 (1983).

[6] R. B. Ivry, Curr. Opin. Neurobiol. **6**, 851 (1996).

[7] M. D. Mauk and N. H. Donegan, Learn. Mem. **3**, 130 (1997).

[8] K. M. Christian and R. F. Thompson, Learn. Mem. **11**, 427 (2003).

[9] R. B. Ivry and R. M. Spencer, Curr. Opin. Neurobiol. **14**, 225 (2004).

[10] J. F. Medina, W. L. Nores, T. Ohyama, and M. D. Mauk, Curr. Opin. Neurobiol. **10**, 717 (2000).

[11] T. Ohyama, W. L. Nores, M. Murphy, and M. D. Mauk, trends Neurosci. **26**, 222 (2003).

[12] M. Ito, Trends Cogn. Sci. **2**, 313 (1998).

[13] A. R. Wagner and S. E. Brandon, in *Contemporary Learning Theories: Pavlovian Conditioning and The Status of Traditional Learning Theories*, edited by S. B. Klein and R. R. Mowrer, (Erlbaum, Hillsdale, NJ, 1989), pp. 149-189.

[14] E. R. Hilgard and D. G. Marquis, J. Comp. Psychol. **19**,

29 (1935).

[15] E. R. Hilgard and D. G. Marquis, Psychol. Monogr. **47**, 186 (1936).

[16] N. Schneiderman, I. Fuentes, and I. Gormezano, Science **136**, 650 (1962).

[17] R. W. Skelton, Behav. Neurosci. **102**, 586 (1988).

[18] M. Ivarsson and P. Svesson, J. Neurophysiol. **83**, 796 (2000).

[19] C. A. Boneau, J. Exp. Psychol. **56**, 471 (1958).

[20] M. D. Mauk and B. P. Ruiz, Behav. Neurosci. **106**, 666 (1992).

[21] J. A. Domingo, A. Gruart, and J. M. Delagado-Garcia, J. Neurophysiol. **78**, 2518 (1997).

[22] J. H. Jr. Freeman, D. A. Nicholson, A. S. Mukler, C. A. Rabinak, and N. T. DiPietro, Behav. Neurosci. **117**, 283 (2003).

[23] S. K. E. Koekoek, H. C. Hulscher, B. R. Dortland, R. A. Hensbroek, Y. Elgersma, T. J. H. Ruigrok, and C. I. De Zeeuw, Science **301**, 1736 (2003).

[24] E. R. Hilgard and A. A. Campbell, J. Exp. Psychol. **19**, 227 (1936).

[25] S. A. Heiney, M. P. Wohl, S. N. Chettih, L. I. Rufflo, and J. F. Medina, J. Neurosci. **34**, 14845 (2014).

[26] D. A. McCormick, G. A. Clark, D. G. Lavond, and R. F. Thompson, Proc. Natl. Acad. Sci. U. S. A. **79**, 2731 (1982).

[27] D. A. McCormick and R. F. Thomson, Science **223**, 296 (1984).

[28] D. Marr, J. Physiol. **202**, 437 (1969).

[29] J. S. Albus, Math. Biosci. **10**, 25 (1971).

[30] D. O. Hebb, *The Organization of Behavior; A Neuropsychological Theory* (Wiley & Sons, New York, 1949).

[31] G. S. Brindley, IBRO Bull. **3**, 80 (1964).

[32] M. Ito, M. Sakurai, and P. Tongroach, J. Physiol. **324**, 113 (1982).

[33] M. Ito and M. Kano, Neurosci. Lett. **33**, 253 (1982).

[34] M. Sakurai, J. Physiol. **394**, 463 (1987).

[35] M. Ito, Ann. Rev. Neurosci. **12**, 85 (1989).

[36] M. Ito, Brain Res. **886**, 237 (2000).

[37] M. Ito, Physiol. Rev. **81**, 1143 (2001).

[38] M. Ito, Nat. Rev. Neurosci. **3**, 896 (2002).

[39] J. Desmond and J. Moore, Biol. Cybern. **58**, 405 (1988).

[40] J. W. Moore, J. E. Desmond, and N. E. Berthier, Giol. Cybern. **62**, 17 (1989).

[41] M. A. Gluck, E. S. Reifsneider, and R. F. Thompson, in *Neuroscience and Connectionest Theory*, edited by M. A. Gluck and D. E. Rumelhart, (Erlbaum, Hillsdale, New Jersy, 1990), pp. 131-186.

[42] F. Chapeau-Blondeau and G. Chauvet, Biol. Cybern. **65**, 267 (1991).

[43] D. Bullock, J. C. Fiala, and S. Grossberg, Neural Netw. **7**, 1101 (1994).

[44] J. C. Fiala, S. Grossberg, and D. Bullock, J. Neurosci. **16**, 3760 (1996).

[45] D. V. Buonomano and M. D. Mauk, Neural Comput. **6**, 38 (1994).

[46] J. F. Medina, K. S. Garcia, W. L. Nores, N. M. Taylor, and M. D. Mauk, J. Neurosci. **20**, 5516 (2000).

[47] J. F. Medina and M. D. Mauk, Nature Neurosci. **3**, 1205 (2000).

[48] T. Yamazaki and S. Tanaka, Neural Comput. **17**, 1032 (2005).

[49] T. Yamazaki and S. Tanaka, Eur. J. Neurosci. **26**, 2279 (2007).

[50] S.-Y. Kim and W. Lim. 2020. *Effect of Diverse Recoding of Granule Cells on Optokinetic Response in A Cerebellar Ring Network with Synaptic Plasticity.* bioRxiv doi: https://doi.org/10.1101/2020.03.25.007245.

[51] P. Safo and W. G. Regehr, Neuropharmacology **54**, 213 (2008).

[52] D. J. Watts and S. H. Strogatz, Nature **393**, 440 (1998).

[53] S. H. Strogatz, Nature **410**, 268 (2001).

[54] W. Gerstner and W. Kistler, *Spiking Neuron Models* (Cambridge University Press, New York, 2002).

[55] W. T. Thach, J. Neurophysiol. **31**, 785 (1968).

[56] M. Häusser and B. A. Clark, Neuron **19**, 665 (1997).

[57] E. De Schutter, Trends Neurosci. **18**, 291 (1995).

[58] C. Chen and R. F. Thompson, Learn. Mem. **2**, 185 (1995).

[59] S.-H. Wang, W. Denk, and M. Häusser, Nat. Neurosci. **3**, 1266 (2000).

[60] M. Coesmans, J. T. Weber, C. I. De Zeeuw, and C. Hansel, Neuron **44**, 691 (2004).

[61] V. Steuber, W. Mittmann, F. E. Hoebeek, R. A. Silver, C. I. De Zeeuw, M. Häusser, and E. De Schutter, Neuron **54**, 121 (2007).

[62] V. Lev-Ram, S. B. Mehta, D. Kleinfeld, and R. Y. Tsien, Proc. Natl. Acad. Sci. USA **100**, 15989 (2003).

[63] E. Molnár, J. Neurochem. **131**, 1 (2014).

[64] Y. Yang and S. G. Lisberger, Nature **510**, 529 (2014).

[65] A. R. Gallimore, T. Kim, K. Tanaka-Yamamoto, and E. De Schutter, Cell Rep. **22**, 722 (2018).

[66] W. Gerstner and J. L. van Hemmen, Network **3**, 139 (1992).

[67] D. V. Buonomano and M. D. Mauk, Neural Comput. **6**, 38 (1994).

[68] G. T. Kenyon, J. F. Medina, and M. D. Mauk, J. Comput. Neurosci. **5**, 17 (1998).

[69] J. F. Medina, K. S. Garcia, W. L. Nores, N. M. Taylor, and M. D. Mauk, J. Neurosci. **20**, 5516 (2000).

[70] P. D. Roberts, J. Comput. Neurosci. **22**, 283 (2007).

[71] P. Achard and E. De Schutter, Front. Comput. Neurosci. **2**, 8 (2008).

[72] G. Bouvier, J. Aljadeff, C. Clopath, C. Bimbard, J. Ranft, A. Blot, J.-P. Nadal, N. Brunel, V Hakim, and B. Barbour, eLife **7**, e31599 (2018).

[73] A. Mathy, S. S. N. Ho, J. T. Davie, I. C. Duguid, B. A. Clark, and M. Häusser, Neuron **62**, 388 (2009).

[74] R. R. Llinás, Front. Neural Circuit. **7**, 96 (2014).

[75] K. Pearson, Proc. Royal Soc. Lond. **58**, 240 (1895).

[76] S.-Y. Kim and W. Lim, J. Neurosci. Meth. **226**, 161 (2014).

[77] X.-J. Wang, Physiol. Rev. **90**, 1195 (2010).

[78] N. Brunel, J. Comput. Neurosci. **8**, 183 (2000).

[79] N. Brunel and V. Hakim, Neural Comput. **11**, 1621 (1999).

[80] N. Brunel and X.-J. Wang, J. Neurophysiol. **90**, 415 (2003).

[81] C. Geisler, N. Brunel, and X.-J. Wang, J. Neurophysiol. **94**, 4344 (2005).

[82] N. Brunel and D. Hansel, Neural Comput. **18**, 1066 (2006).

[83] N. Brunel and V. Hakim, Chaos **18**, 015113 (2008).

[84] H. Shimazaki and S. Shinomoto, J. Comput. Neurosci. **29**, 171 (2010).

[85] M. D. Mauk and B. P. Ruiz, Behav. Neurosci. **106**, 666 (1992).

[86] C. Hansel, D. J. Linden, and E. D'Angelo, Nat. Neurosci.

4, 467 (2001).

[87] Z. Gao, B. J. van Beugen, and C. I. De Zeeuw, Nat. Rev. Neurosci. **13**, 619 (2012).

[88] N. Zheng and I. M. Raman, Cerebellum **9**, 56 (2010).

[89] W. Lennon, T. Yamazaki, and R. Hecht-Nielsen, Front. Comput. Neurosci. **9**, 150 (2015).

[90] E. D'Angelo and C. I. De Zeeuw, Trends Neurosci. **32**, 30 (2008).

[91] E. D'Angelo, S. K. E. Koekkoek, P. Lombardo, S. Solinas, E. Ros, J. Garrido, M. Schonewille, and C. I. De Zeeuw, Neuroscience **162**, 805 (2009).

[92] C. de Solages, G. Szapiro, N. Brunel, V. Hakim, P. Isope, P. Buisseret, C. Rousseau, B. Barbour, and C. Léna, Neuron **58**, 775 (2008).

[93] R. R. Llinás, J. Physiol. **589**, 3423 (2011).