Deep ancestral introgression shapes evolutionary history of dragonflies and

damselflies

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SUMMARY

Introgression is arguably one of the most important biological processes in the evolution of groups of related species, affecting at least 10% of the extant species in the animal kingdom. Introgression reduces genetic divergence between species, and in some cases can be highly beneficial, facilitating rapid adaptation to ever-changing environmental pressures. Introgression also significantly impacts inference of phylogenetic species relationships where a strictly binary tree model cannot adequately explain reticulate net-like species relationships. Here we use phylogenomic approaches to understand patterns of introgression along the evolutionary history of a unique, non-model, relic insect system: dragonflies and damselflies (Odonata). We demonstrate that introgression is a pervasive evolutionary force across various taxonomic levels within Odonata. In particular, we show that the morphologically "intermediate" species of Anisozygoptera (one of the three primary suborders within Odonata besides Zygoptera and Anisoptera), which retain phenotypic characteristics of the other two suborders, experienced high levels of introgression probably coming from zygopteran genomes. Additionally, we found evidence for multiple cases of deep inter-familial ancestral introgression. However, weaker evidence of introgression was found within more recently diverged focal clades: Aeshnidae, Gomphidae+Petaluridae and Libellulidae.

INTRODUCTION

In recent years there has been a large body of evidence amassed demonstrating that multiple parts of the Tree of Life did not evolve according to a strictly bifurcating phylogeny [1, 2]. Instead, many organisms experience reticulate network-like evolution that is caused by an exchange of inter-specific genetic information via various biological processes. In particular, lateral gene transfer, incomplete lineage sorting (ILS) and introgression can result in gene trees that are discordant with the species tree [3, 4]. Lateral transfer and introgression both involve gene flow following speciation, thereby producing "reticulate" phylogenies. Incomplete lineage sorting (ILS), on the other hand, occurs when three or more lineages coalesce in their ancestral population. This often arises in an order that conflicts with that of the true species tree, and thus involves no post-speciation gene flow and therefore does not contribute to reticulate evolution.

Thus, phylogenetic species-gene tree incongruence observed in empirical data can provide

insight into underlying biological factors that shape the evolutionary trajectories of a set of taxa.

The major source of reticulate evolution for eukaryotes is introgression where it affects

approximately 25% of flowering plant and 10% of animal species [1, 5]. Introgressed alleles can

be fitness-neutral, deleterious [6] or adaptive. For example, adaptive introgression has been

shown to provide an evolutionary rescue from polluted habitats in gulf killifish (*Fundulus*

grandis) [7], yielded mimicry adaptations among *Heliconius* butterflies [8] and archaic

introgression has facilitated adaptive evolution of altitude tolerance [9], immunity and

metabolism in modern humans [10]. Additionally, hybridization and introgression are important

and often overlooked mechanisms of invasive species establishment and spread [11].

Odonata, the insect order that contains dragonflies and damselflies, lacks a strongly

supported backbone tree to clearly resolve higher-level phylogenetic relationships [12, 13].

Current evidence places odonates together with Ephemeroptera (mayflies) as the living

representatives of the most ancient insect lineages to have evolved wings and active flight [14].

Odonates possess unique anatomical and morphological features such as a specialized body

form, specialized wing venation, a distinctive form of muscle attachment to the wing base [15]

allowing for direct flight and accessory (secondary) male genitalia that support certain unique

behaviors (e.g., sperm competition). They are among the most adept flyers of all animals and are

exclusively carnivorous insects relying primarily on vision [16, 17] to capture prey. They spend

much of their adult life in flight [18]. Biogeographically, odonates exhibit worldwide dispersal

[19] and play crucial ecological roles in local freshwater communities, being a top invertebrate

predator [20]. Due to this combination of characteristics, odonates are quickly becoming model

organisms to study specific questions in ecology, physiology and evolution [21, 22]. However,

the extent of introgression at the genomic scale within Odonata remains primarily unknown.

Two early attempts to tackle introgression/hybridization patterns within Odonata were

undertaken in [23, 24]. The studies showed that two closely related species of damselflies,

Ischnura graellsii and *I. elegans,* can hybridize under laboratory conditions and that genital

morphology of male hybrids shares features with putative hybrids from *I. graellsii–I. elegans*

natural allopatric populations [23]. The existence of abundant hybridization and introgression in

natural populations of *I. graellsii* and *I. elegans* has received further support from an analysis of

microsatellite data [25]. Putative hybridization events have also been identified in a pair of

calopterygoid damselfly species, *Mnais costalis* and *M. pruinosa* based on the analyses of two

molecular loci (mtDNA and nucDNA) [26], and between *Calopteryx virgo* and *C. splendens*

using 16S ribosomal DNA and 40 random amplified polymorphic DNA (RAPD) markers [27]. A

more recent study identified an inter-specific hybridization between two cordulegasterid

dragonfly species, *Cordulegaster boltonii* and *C. trinacriae* using two molecular markers

(mtDNA and nucDNA) and geometric morphometrics [28].

Here we present a comprehensive analysis of transcriptomic data from 83 odonate

species. First, we reconstruct a robust phylogenetic backbone using up to 4341 genetic loci for

the order and discuss its evolutionary history spanning from the Carboniferous period (~360

mya) to present day. Second, we infer signatures of introgression with varying degrees of

phylogenetic depth using genomic data across the tree primarily focusing on four focal clades,

namely Anisozygoptera ("deep"), Aeshnidae ("shallow"), Gomphidae+Petaluridae

("intermediate") and Libellulidae ("shallow"). We find that introgression is pervasive in Odonata

throughout its entire evolutionary history. In particular, we identify a strong signal of deep

introgression in the Anisozygoptera suborder, species of which possess traits of both main

suborders, Anisoptera and Zygoptera.

RESULTS

Overview of Odonata Phylogeny
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We compiled transcriptomic data for 83 odonate species including 49 new transcriptomes

sequenced for this study (Data S1). To assess effects of various steps of our phylogenetic

pipeline on species tree inference, we examined different methods of sequence homology

detection, multiple sequence alignment strategies, postprocessing filtering procedures and tree

estimation methods. Specifically, three types of homologous loci (gene clusters) were used to

develop our supermatrices, namely 1603 conserved single-copy orthologs (CO), 1643 all single-

copy orthologs (AO) and 4341 paralogy-parsed orthologs (PO) with ≥ 42 (~50%) species present

(for more details, see Method Details). To date, our data represents the most comprehensive

resource available for Odonata in terms of gene sampling. Each gene cluster was aligned,

trimmed and concatenated resulting in five main supermatrices, CO (DNA/AA), AO (AA) and

PO (DNA/AA), which included 2,167,861 DNA (682,327 amino acid (AA) sites), 882,417 AA

sites, 6,202,646 DNA (1,605,370 AA) sites, respectively. Thus, the largest alignment that we

used to infer the odonate phylogeny consists of 4341 loci concatenated into a supermatrix with

>6 million nucleotide sites. All supermatrices are summarized in Data S2; the inferred odonate

relationships are shown in Figure S1A whereas topologies of all inferred phylogenies are plotted

in Figure S1B and topologies of 1603 CO gene trees are shown in Figure S1C. Additionally, we

performed nodal dating of the inferred phylogeny using 20 fossil calibration points (Data S3).

The inferred ML phylogenetic tree of Odonata using DNA supermatrix of 1603 BUSCO

loci (Figure 1) was used as a primary phylogenetic hypothesis throughout this study as it agrees

with the majority of relationships inferred by other methods (Figures S1A, S1B). Divergence of

Zygoptera and Epiprocta (Anisozygoptera+Anisoptera) from the Most Recent Common Ancestor

(MRCA) occurred in the Middle Triassic ~242 mya (Figure 1), which is in line with recent

estimates [14, 29]. Comprehensive phylogenetic coestimation of subordinal relationships within

Odonata showed that the suborders were well supported (Figure S1A), as they were consistently

recovered as monophyletic clades in all analyses. In several previous studies, paraphyletic

relationships of Zygoptera had been proposed based on wing vein characters derived from fossil

odonatoids and extant Odonata [30], analysis of 12S [31], analysis of 18S, 28S, Histone 3 (H3)

and morphological data [32] and analysis of 16S and 28S data [33]. In most of these studies,

Lestidae was inferred to be sister to Anisoptera. Functional morphology comparisons of flight

systems, secondary male genitalia and ovipositors also supported a non-monophyletic Zygoptera

with uncertain phylogenetic placement of multiple groups [34]. Nevertheless, the relationships

inferred from these previous datasets seem to be highly unlikely due to apparent morphological

differentiation (e.g., eye spacing, body robustness, wing shape) between the suborders and

support for monophyletic Anisoptera and Zygoptera from more recent morphological [35],

molecular [14, 17, 36, 37] and combined studies using both data types [38]. Our analyses recover

Zygoptera as monophyletic consistently (Figure S1A).

Divergence time estimates suggest a TMRCA of Anisoptera and Anisozygoptera in the

Late Triassic (~205 mya; Figure 1). Epiprocta as well as Anisoptera were consistent with more

recent studies and recovered as monophyletic with very high support. We also note here that our

divergence time estimates of Anisoptera tend to be younger than those found in ref. [39]. The

fossil-calibration approach based on penalized likelihood has been shown to overestimate true

nodal age [40] preventing direct comparison between our dates and those estimated in ref. [39].

The phylogenetic position of Gomphidae and Petaluridae, both with respect to each other

and the remaining anisopteran families, has long been difficult to resolve. Several phylogenetic

hypothesis have been proposed in the literature based on molecular and morphological data

regarding the placement of Gomphidae as sister to the remaining Anisoptera [41] or to

Libelluloidea [42]. Petaluridae has exhibited stochastic relationships with different members of

Anisoptera, including sister to Gomphidae [42], sister to Libelluloidea [37], sister to

Chlorogomphidae+Cordulegasteridae [38] and sister to all other Anisoptera [30, 43]. The most

recent analyses of the major anisopteran lineages using several molecular markers [13] suggest

Gomphidae and Petaluridae as a monophyletic group, but without strong support. Here the

majority of our supermatrix analyses (Figure S1A) strongly support a sister relationship between

the two families, and in our phylogeny (Figure 1) they split from the MRCA ~165 mya in the

Middle Jurassic (Figure 1); however, almost all the coalescent-based species trees reject such a

relationship with high confidence (Figure S1A). In the presence of incomplete lineage sorting,

concatenation methods can be statistically inconsistent [44] leading to an erroneous species tree

topology with unreasonably high support [45]. Thus, inconsistency in the recovery of a sister

group relationship between Gomphidae and Petaluridae can be explained by elevated levels of

incomplete lineage sorting between the families and/or possible introgression events [46] (see

below).

New zygopteran lineages originated in the Early Jurassic ~191 mya with the early split of

Lestoidea and the remaining Zygoptera (Figure 1). Subsequent occurrence of two large

zygopteran groups, Calopterygoidea and Coenagrionoidea, was estimated within the Cretaceous

(~145-66 mya) and culminated with the rapid radiation of the majority of extant lineages in the

Paleogene (~66-23 mya). Our calibrated divergences generally agree with estimates in [14].

However, any further comparisons are precluded by the lack of comprehensive divergence time

estimation for Odonata in the literature. The backbone of the crown group Calopterygoidea that

branched off from Coenagrionoidea ~129 mya in the Early Cretaceous was well supported as

monophyletic in most of our inferred phylogenies (Figures 1, S1A). Previous analyses struggled

to provide convincing support for the monophyly of the superfamily [12, 37, 38], whereas only

11 out of 48 phylogenetic reconstructions rejected Calopterygoidea (Figure S1A).

We used quartet sampling [47] to provide additional information about nodal support and

investigate biological explanations for alternative evolutionary histories that received some

support. We found that for most odonate key radiations, the majority of quartets (i.e. Frequency
> 0.5) support the proposed phylogenetic hypothesis with Quartet Concordance (QC) scores > 0
(Figure S2) across all estimated putative species trees (Figure S1B). The few exceptions consist
of the Gomphidae+Petaluridae split and the A+B split, where we have Frequency < 0.5 and QC
< 0, which suggests that alternative relationships are possible. Quartet Differential (QD),
inspired by f and D statistics for introgression, provides an indicator of how much the
proportions of discordant quartets are skewed (i.e. whether one discordant is more common than
the other), suggestive of introgression and/or introgression or substitution rate heterogeneity
[47]. Interestingly, we identified skewness (i.e. QD < 0.5) for almost every major radiation
(Figure S2), which suggests that alternative relationships can be a result of underlying biological
processes (e.g. introgression) rather than ILS alone [47]; however, this score may not be highly
informative if the majority of quartets agree with the focal topology (i.e Frequency > 0.5 and QC
> 0). However, for both the Gomphidae+Petaluridae and the A+B splits, we have Frequency >
0.5, QC < 0 and QD < 0.5, implying that alternative phylogenetic relationships are plausibly not
only due to ILS but also possible ancestral introgression.
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199   **Major Trends in Evolutionary History of Odonata**
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Investigation of diversification rates in Odonata highlighted two major trends correlated with two
mass extinction events in the Permian-Triassic (P-Tr) ~252 mya and Cretaceous-Paleogene (K-
Pg) ~66 mya. First, it appears likely that the ancient diversity of Odonata was largely eliminated
after the P-Tr mass extinction event (see the temporal distribution of fossil samples in Figure 1)
as was also the case for multiple insect lineages [48]. According to the fossil record at least two
major odonatoid lineages went extinct (Protodonata and Protanisoptera [49]) and likely many
genera from other lineages as well (e.g., Kargalotypidae from Triadophlebimorpha [50]). The
establishment of major odonate lineages was observed during the Cretaceous starting ~135 mya
(Figure 1, red dot). This coincided with the radiation of angiosperm plants that, in turn, triggered
the formation of herbivorous insect lineages [29]. Odonates are exclusively carnivorous insects
and their diversification was likely driven by the aforementioned sequence of events.
Interestingly, molecular adaptations in the odonate visual systems are coupled with their
diversification during the Cretaceous as well [17].

Figure 1. Evolutionary History of Odonata

Fossil calibrated ML phylogenetic tree of Odonata using a DNA supermatrix consisting of 1603 BUSCO genes with a total of 2,167,861 aligned sites. The blue densities at each node represent posterior distributions of ages estimated in MCMCTree using 20 fossil calibration points. Lineage Through Time (LTT, pink solid line) trajectory shows accumulation of odonate lineages through time. The red dot on the LTT line indicates the beginning of establishment for major odonate lineages originating in and spanning the Cretaceous. The histogram (blue bars) represents temporal distribution of fossil Odonatoptera samples. The dashed vertical lines mark major extinction events, namely Permian-Triassic (P-Tr, ~ 251 mya), Triassic-Jurassic (Tr-J, ~ 201.3 mya) and Cretaceous-Paleogene (K-Pg, ~66 mya)

Figure 2. Detection of Introgression/Hybridization Trajectories by Different Methods in Anisozygoptera.

Three site pattern-based (D statistic, HyDe and D_{FOIL}), gene tree count/branch length-based (χ^2 count test/QuIBL) and network ML inference (PhyloNet) methods were used to test for introgression/hybridization. Arrows denote introgression. The figure panel represents larval and adult stages for three Odonata suborders. Species from top to bottom: *Lestes australis, Epiophlebia superstes* and *Anax junius.* Image credit: *Epiophlebia superstes* adult by Christian Dutto Engelmann; *Lestes australis* and *Anax junius* adults by John Abbott.

Signatures of Introgression within Odonata

We tested introgression/hybridization scenarios within Odonata on different taxonomic

ranks, such as order, suborder, superfamily (except Aeshnoidea, since it has been recovered as

paraphyletic, and Platystictoidea, which contains only one species in our phylogeny; Figure 1)

and certain specific focal clades. Specifically, we used five different methods to test for

introgression within Odonata, as exemplified for the Anisozygoptera suborder in Figure 2. As

our test cases for introgression, we *a priori* selected Aeshnidae and Libellulidae families as

possible examples of "shallow" introgression, Gomphidae+Petaluridae as a possible example of

"intermediate" introgression and Anisozygoptera as a possible case of "deep" introgression.

Since the introgression signatures within Odonata are unknown, we chose these test clades based

on possession of characteristic traits of both suborders for Anisozygoptera [35, 51-53], the

unstable phylogenetic placement of Gomphidae relative to Petaluridae [41, 42] and the presence

of closely related species within Libellulidae and Aeshnidae. For Aeshnidae, Libellulidae and

Gomphidae+Petaluridae, we tested all possible introgression scenarios between species within

each of these clades using methods outlined in Figure 2. For Anisozygoptera we tested whether

the lineage leading to *E. superstes* experienced ancestral introgression with Anisoptera and/or

Zygoptera (see Figure 2). Additionally, we searched for introgression within other taxonomic

groups, including the remaining suborders (Zygoptera and Anisoptera) and five superfamilies

within Zygoptera and Anisoptera (Lestoidea, Calopterygoidea, Coenagrionoidea,

Cordulegastroidea and Libelluloidea; Figure 1). The introgression results for the entire Odonata

order either comprise of all tests performed within the entire phylogeny (HyDe) or of a union of

tests performed within Anisoptera, Zygoptera and Anisozygoptera (D_{FOIL} and χ^2 count

test/QuIBL).

Figure 3. Distributions of the Patterson's *D* Statistic, HyDe γ and Their Relations Across Odonate Taxonomic Levels.
(A) Estimated distributions of significant (Bonferroni corrected *P* < 0.05) *D* statistic from ABBA-BABA site pattern counts for each quartet using HyDe output. Black dots mark medians of violin plots.
(B) Distribution of significant (Bonferroni corrected $P < 10^{-6}$) γ values for each quartet estimated by HyDe. In general, γ values that are not significantly different from 0 (or 1) denote no relation of a putative hybrid species to either of the parental species P_1 (1-γ) or P_2 (γ) in a quartet.
(C) Proportions of quartets that support or reject introgression based on simultaneous significance of *D* statistics and γ.
(D) Non-linear relationships between absolute values of significant *D* statistics and γ. The black line denotes a GAM fit. The color legend shows density of quartets across γ-*D* plane.
(E)-(F) tSNE projections of the 15 site pattern counts derived for each of the 51367 significant quartets from HyDe output (each dot on a tSNE map represents a quartet). Color schemes reflect significant values of *D* statistic (E) and significant values of of γ (F).

First, we compared the distributions of *D*-statistic values within different taxa computed

from ABBA-BABA [54] site patterns (Figure 3A). The multimodal shape of the D distribution

for Odonata can be explained by the presence of quartets of older as well as more recently

diverged taxa. Although quartets with older divergence times tend to have only slightly

decreased values of D [55], this effect can be more pronounced for extremely old divergences

leading to significantly smaller D. Indeed, quartets that were formed within more recently

diverged clades of Coenagrionoidea, Aeshnidae (both in Early Cretaceous, ~ 140 mya) and

Libellulidae (Oligocene ~ 33 mya) have significantly greater D values (Wilcoxon rank-sum test

[WRST], $P = 8.366 \times 10^{-10}$) if compared with quartets that were formed from distantly related

species of Zygoptera and Anisoptera (Late Triassic ~237 mya). Comparisons of D distributions

derived from the entire order with those from other odonate monophyletic groups showed that

only Coenagrionoidea, Gomphidae+Petaluridae and Libelluloidea exhibit significantly higher D

statistic values (WRST, all $P < 10^{-4}$), whereas Zygoptera, Anisozygoptera and Cordulegastroidea

showed reduced average D values (WRST, all $P < 10^{-4}$). Second, we estimated the γ parameter

of HyDe's hybrid speciation model [56], where γ and (1-γ) corresponds to the parental fractions

in a putatively hybrid genome, with values of 1 or 0 indicating the absence of hybridization.

Comparisons of γ distributions (Figure 3B) show similar trends except that here Anisozygoptera,

Anisoptera and Libellulidae along with Coenagrionoidea, Gomphidae+Petaluridae and

Libelluloidea exhibit elevated average γ values (WRST, all $P < 0.00064$) compared to the total

distribution of γ. Additionally, we detected an overall excess of quartets with both significant γ

and D for Anisozygoptera (Fisher exact test [FET], $P = 1.865584 \times 10^{-10}$) and Calopterygoidea

(FET, $P = 0.016$) which further support signatures of introgression within these clades (Figure

3C). Overall D and γ are significantly correlated (Spearman's rank correlation test, $\rho = 0.78$, $P =$

0) with a nonlinear relationship, and the majority of quartets with low D also have low γ values

(brighter region, Figure 3D).

There are a variety of other test statistics that have been developed to detect introgression

(e.g., [55, 57-59]). Because, like D and γ, many of these statistics are computed from different

invariants, we attempted to visualize the relationships between all 15 site patterns computed by

HyDe using t-distributed stochastic neighbor embedding (tSNE; [60]) for dimensionality

reduction, along with the corresponding values of D and γ (Figures 3 E and F, respectively).

Specifically, we showed that quartets with similar signatures of introgression tend to cluster

together. Moreover, tSNE maps further demonstrate the correlation between D and γ. The

clustering of quartets with significant introgression according to a two-dimensional

representation of their site patterns may suggest the presence of additional site-pattern signatures

of introgression.

Further, we tested introgression using an alternative invariant-based method D_{FOIL} (Figure

4), which is capable of detecting ancestral as well as inter-group introgression and inferring its

polarization (see Method Details). Overall, the distributions of D_{FOIL} statistics (Figure 4A) except

for Lestoidea and Aeshnidae were significantly different from the D_{FOIL} distribution within the

entire order across all taxonomic levels (WRST, all $P < 3.4 \times 10^{-4}$). This observation is indicative

of differences in the amount of ancestral introgression, which is determined significance of D_{FOIL}

statistics. Further we note that average D_{FOIL} (Figure 4A) was significantly different from 0 (one

sample t-test [OSTT], all $P < 0.0223$) for all taxa except Lestoidea, Aeshnidae,

Gomphidae+Petaluridae and Libellulidae. Together these significant deviations from 0 of D_{FOIL}

statistics support the hypothesis of introgression for the tested taxonomic levels [61]. Tests of

individual quintets as implemented in D_{FOIL} identified significant cases of introgression within

all clades except Lestoidea (Figure 4B). Considering polarization of introgression for

Anisozygoptera, almost ubiquitously (1157 out of 1158) cases with positive ancestral

introgression quintets suggest that Anisozygoptera is a recipient taxon whereas Zygoptera is a

donor taxon (Figure 2). Also, 4473 evaluated quintets with significant inter-group introgression

are indicative of one-way introgression from Zygoptera to Anisozygoptera as well. As did D and

γ, D_{FOIL} detected an excess of cases that support introgression for Anisozygoptera (FET, $P =$

0.04165). The other focal clade, Aeshnidae showed two significant quintets with only inter-

group type introgression (Figure 4B) where *Aeshna palmata* putatively introgressed into

Austroaeschna subapicalis. For Gomphidae+Petaluridae group D_{FOIL} identified a single case of

ancestral introgression where *Phenes raptor* introgressing into ancestral Gomphidae lineage. For

Libellulidae, two ancestral and five inter-group introgression scenarios were significant.

Specifically, for inter-group introgression we found that *Acisoma variegatum* putatively

Figure 4. Distributions of the D_{FOIL} Statistics and Their Relations Across Odonate Taxonomic Levels.

(A) Estimated distributions of D_{FOIL} statistics from different site pattern counts for each symmetric quintet. D_{FOIL} allows to determine introgression and its polarization between a donor and recipient taxa by comparison of a sign (+/-/0) for all D_{FOIL} statistics. Black dots mark medians of violin plots. Lestoidea had no significant cases.

(B) Counts of quintets that support ancestral, inter-group or no introgression scenarios based on significance of D_{FOIL} statistics ($P < 0.01$) and their directionality.

introgressed into *Rhyothemis variegate*, *Libellula saturata* into *Ladona fulva*, *Ladona fulva* into

Erythrodiplax connata and two cases of *Pantala flavescens* into *Sympetrum frequens*. Despite

the notion that D_{FOIL} approach exhibits low false positive rate, it requires tree symmetry (Figure

2) [61], thus not all the quintet combinations of taxa can be tested for introgression with this

method. For this reason we were unable to form more total quintets for Anisozygoptera, i.e.

symmetric trees (((Zygoptera, Zygoptera),(Anisozygoptera, Anisoptera)),Outgroup).

307   Next, we used an approach based on gene tree topology counts to identify introgression

308   within Odonata (Figure 5). Briefly, for a triplet of species, under ILS one would expect equal

309   proportions of gene tree topologies supporting the two topologies disagreeing with the species

310   tree, and any imbalance may suggest introgression. Thus, deviation from equal frequencies of

Figure 5. Overview of χ^2 Count Test for Odonate Taxonomic Levels Classification of triplets based on χ^2 test results. Counts of triplets that are concordant with the species tree (FDR < 0.05 and maximum number of gene trees agreeing with a triplet), extreme ILS (FDR > 0.05 for all gene tree counts), ILS (FDR > 0.05 for the discordant gene tree counts) and introgression+ILS (FDR < 0.05 for the discordant gene tree counts)

gene tree counts among discordant gene trees can be assessed using a χ^2 test, similar to the

method proposed in [62]. With this method, we identified a clear excess of significant triplets

that support introgression for Anisozygoptera (FET, $P = 0$), in fact they account for almost 1/3 of

all significant cases within Odonata. In agreement with D_{FOIL}, D and γ, we found introgression

patterns between Anisozygoptera and Zygoptera. Out of 1048 significant triplets 1021 suggest

introgression between Anisozygoptera and Zygoptera. An alternative gene tree branch length-

based approach, QuIBL [63], also showed an excess of significant triplets (FET, $P =$

9.556092×10^{-6}; Figure S3) compared to the entire order. Strikingly, out of 1042 significant

triplets, 1015 suggest introgression between Anisozygoptera and Zygoptera as well. Among

other focal clades only Libellulidae has a single triplet (Figure 5) with significant imbalance of

discordant gene tree topology counts, which supports introgression of *Ladona fulva* and *Libellula*

forensis.

Finally, we performed phylogenetic network inference in PhyloNet [64, 65] for our four

focal clades (Figure 6) specifying a single reticulation scenario. For the Anisozygoptera,

Figure 6. Phylogenetic Networks for Focal Clades Inferred in PhyloNet.

(A) Phylogenetic networks estimated from a set of ML gene trees using maximum pseudolikelihood approach. *Epiophlebia superstes* was specified as a putative hybrid for Anisozygoptera clade. Blue lines indicate a reticulation event with the value of PhyloNet's estimated γ. Numbers above the network indicates the log-likelihood score.

(B) Phylogenetic networks estimated from a set of ML gene trees using maximum likelihood approach. *Epiophlebia superstes* was specified as a putative hybrid for Anisozygoptera clade. Blue lines indicate a reticulation event with the value of PhyloNet's estimated γ. Number above the network indicates the log-likelihood score.

Aeshnidae and Gomphidae+Petaludaridae clades maximum pseudolikelihood (Figure 6A) and

full maximum likelihood (Figure 6B) approaches recovered topologically similar networks. In

Anisozygoptera pseudolikelihood analysis suggests a reticulation event between Aeshnidae

(Anisoptera) and Zygoptera, whereas full likelihood infers reticulation between Anisoptera and

Zygoptera suborders. For the Aeshnidae and Gomphidae+Petaludaridae clades both

pseudolikelihood and full likelihood approaches both recover the same reticulation. For the

Libellulidae clade pseudolikelihood and full likelihood methods did not converge to the same

results, however we note significantly better log-likelihood score for the full likelihood network.

Overall, we note that the full likelihood approach returned more optimal networks based on the

log-likelihood scores. This observation is most likely due to the fact that we performed full

likelihood analysis in an exhaustive manner (Data S4) testing every possible reticulation event

Figure 7. Overlap between Putatively Introgressed Species Pairs Inferred by Hyde/D, D_{FOIL} and χ^2.

The numbers within sets represent the number of unique introgressing species pairs identified by a corresponding method.

within a focal clade topology congruent with the species tree (Figure 1), whereas

pseudolikelihood analysis used the hill-climbing algorithm to search a network space, which is

not guaranteed to retrieve the most optimal solution.

We examined the agreement across site- and gene tree-based methods for detecting

introgression (i.e. Hyde, D, and D_{FOIL} and the topology count test) and summarize the results in

Figure 7. Specifically, we compared sets of unique introgressing species pairs that were

identified with significant pattern of introgression. Overall, all methods were able to identify

abundant introgression within the entire Odonata order and the three suborders with strong

agreement. However, there was little intersection among the predictions from these methods

within our focal groups (except for Anisozygoptera). Notably, our topology count test produced

very few predictions that were not in agreement with Hyde, D and/or D_{FOIL}. Further, the analyses

of phylogenetic networks suggest that we were not able to identify consistent patterns of

introgression for Aeshnidae, Gomphidae+Petaluridae and Libellulidae.

DISCUSSION

Identification of reticulations within the Tree of Life has become possible for non-model

organisms due to cost-effective approaches for obtaining genomic data as well as rapid

development of phylogenetic tools for testing models of introgression.

Introgression among Odonata has been previously thought to be uncommon [66, 67] as a

result of probable reproductive isolation mechanisms such as ethological barriers, phenotypic

divergence (e.g., variable morphology of genitalia [68, 69]) and habitat and temporal isolation.

Contrary to this statement, we found abundant patterns of ancestral introgression (Figure 7)

among distantly related odonate taxa. This observation may be explained by reduced sexual

selection pressures in the early stages of odonate evolution that may have inhibited rapid genital

divergence [70], which is probably a primary source of reproductive isolation in Odonata [71].

Most of the introgression and hybridization research in Odonata has been done within the

zygopteran Coenagrionidae family and especially between populations of genus *Ishnura*

focusing on mechanisms of reproductive isolation (e.g. [25, 72]). These studies established

"hybridization thresholds" for these closely related species and showed positive correlation

between isolation and genetic divergence. We found multiple patterns of introgression within

Coenagrionoidea superfamily (Figures 3-6, S3) which is perhaps attributed to the signal of

introgression within several *Ishnura* species included in our taxon sampling.

In particular we found compelling evidence for deep ancestral introgression in

Anisozygoptera suborder, that based on the fossil record, became genetically isolated after the

Lower Jurassic [73] and species of this suborder exhibit anatomical characteristics of both

Anisoptera and Zygoptera suborders. Some general features of Anisozygoptera that relate them

to Zygoptera include dorsal folding of wings during perching in adults, characteristic anatomy of

proventriculus (a part of digestive system that is responsible for grinding of food particles) and

absence of specific muscle groups in the larval rectal gills; whereas abdominal tergite shape, rear

wing geometry and larval structures are similar to Anisoptera [51]. More recent studies also

revealed that Anisozygoptera ovipositor morphology shares similarity with Zygoptera [52];

muscle composition of the head resemble characteristics of both Anisoptera and Zygoptera [53];

thoracic musculature of Anisozygoptera larva exhibit similarity between Anisoptera and

Zygoptera [35]. Thus Anisozygoptera represent a morphological and behavioral "intermediate"

[74], which is supported by our findings where we consistently recovered introgression between

Zygoptera and Anisozygoptera (Figures 2-7, S3). Moreover, with the D_{FOIL} method the

Anisozygoptera was inferred as a recipient taxon from a zygopteran donor. Strikingly, the

average HyDe probability parameter $\gamma \sim 0.7$ (Figure 3) inferred from multiple quartets is very

similar to PhyloNet's inheritance probability $\gamma \sim 0.62$. This may suggest that between 30–40% of

the Anisozygoptera genome descends from zygopteran lineages.

Interestingly, we were not able to detect a robust introgression signal within our other three focal

clades, namely Aeshnidae, Gomphidae+Petaluridae and Libellulidae, where we previously

suspected introgression due to the reasons outlined in the Results section. Phylogenetic

instability of Gomphidae+Petaluridae is most likely explained by ILS rather than introgression.

Indeed, monophyly of Gomphidae+Petaluridae has been strongly rejected by the ILS-aware

phylogenetic reconstructions in ASTRAL (Figure S1A). Overall, due to limited taxon sampling

within Aeshnidae and Libellulidae our methods did not infer a clear introgression signal. We

note that most of the PhyloNet's $\gamma's$ for these clades are ≥ 0.92 (Figure 6) which suggests that the

majority of loci have not experienced introgression. Further the phylogenetic networks

reconstructed by PhyloNet suggest that these clades may contain species that experienced

introgression, but for whom the donor taxa were not sampled. In particular, the topology of a

reticulation edge that branches off from the base of the tree for Gomphidae+Petaluridae,

Aeshnidae and Libellulidae may be indicative of this scenario. Additionally, the absence of the

clear introgression signal within these focal clades can be explained by the fact that a potential

donor can exist outside of the focal clades or went extinct.

402   Our findings provide the first insight into global patterns of introgression that were

pervasive throughout the evolutionary history of Odonata. Further work based on full genome

sequencing data will be necessary to accurately estimate the fraction of the introgressed genome

in various species. We also stress the importance of further method development to allow for

more accurate and scalable inference of phylogenetic networks and enable researchers to pin-

point instances of inter-taxonomic introgression on the phylogeny.

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AUTHOR CONTRIBUTIONS

AS, CS and SMB conceived the research. AS, CS, MSF and PB analyzed the data. MC, KAC,

MFW and DRS supervised the project. AS, CS, DRS and SMB wrote the manuscript.

DECLARATION OF INTERESTS

The authors declare no competing interests.

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METHOD DETAILS

Taxon sampling and RNA-seq

In this study, we used distinct 85 species (83 ingroup and 2 outgroup taxa). 35 RNA-seq libraries were obtained from NCBI (see Data S1). The remaining 58 libraries were sequenced in Bybee Lab (some species have several RNA-seq libraries; Data S1). Total RNA was extracted for each taxon from eye tissue using NucleoSpin columns (Clontech) and reverse-transcribed into cDNA libraries using the Illumina TruSeq RNA v2 sample preparation kit that both generates and amplifies full-length cDNAs. Prepped mRNA libraries with insert size of ~200bp were multiplexed and sequenced on an Illumina HiSeq 2000 producing 101-bp paired-end reads by the Microarray and Genomic Analysis Core Facility at the Huntsman Cancer Institute at the University of Utah, Salt Lake City, UT, USA. Quality scores, tissue type and other information about RNA-seq libraries are summarized in Data S1.

Transcriptome assembly and CDS prediction

RNA-seq libraries were trimmed and de novo assembled using Trinity [75, 76] with default parameters. Then only the longest isoform was selected from each gene for downstream analyses using Trinity utility script. In order to identify potentially coding regions within the transcriptomes, we used TransDecoder with default parameters specifying to predict only the single best ORF. Each predicted proteome was screened for contamination using DIMOND BLASTP [77] with an E-value cutoff of 10^{-10} against custom protein database. Non-arthropod hits were discarded from proteomes (amino acid, AA sequences) and corresponding CDSs. To mitigate redundancy in proteomes and CDSs, we used CD-HIT [78] with the identity threshold of 0.99. Such a conservative threshold was used to prevent exclusion of true paralogous sequences; thus, reducing possible false positive detection of 1:1 orthologs during homology searches.

Homology assessment

In the present study three types of homologous loci (gene clusters) namely conserved single-copy orthologs (CO), all single-copy orthologs (AO) and paralogy-parsed orthologs (PO) identified by BUSCO v1.22 [79], OrthoMCL [80] and Yang's [81] pipelines respectively were used in phylogenetic inference.

BUSCO arthropod Hidden Markov Model Profiles of 2675 single-copy orthologs were

used to find significant COs matches within CDS datasets by HMMER's *hmmersearch* v3 [82]

with group-specific expected bit-score cutoffs. BUSCO classifies loci into complete [duplicated]

and fragmented. Thus, only complete single-copy loci were extracted from CDS datasets and

corresponding AA sequences for further phylogenetic analyses. Since loci were identified as true

orthologs if they score above expected bit-score and complete if their lengths lie within ~95% of

BUSCO group mean length, many partial erroneously assembled sequences were filtered out.

OrthoMCL v2.0.9 [80] was used to compute AOs in all species using predicted AA

sequences by TransDecoder. AA sequences were used in an all-vs-all BLASTP with an E-value

cutoff of 10^{-10} to find putative orthologs and paralogs. The Markov Cluster algorithm (MCL)

inflation point parameter was set to 2. Only 1:1 orthologs were used in further analyses. In order

to exclude false-positive homology clusters identified by OrthoMCL, we applied machine

learning filtering procedure [83] implemented in OGCleaner software v1.0 [84] using a

metaclassifier with logistic regression.

Finally, to identify additional clusters, we used Yang's tree-based orthology inference

pipeline [81] that was specifically designed for non-model organisms using transcriptomic data.

Yang's algorithm is capable of parsing paralogous gene families into "orthology" clusters that

can be used in phylogenetic analyses. It has been shown that paralogous sequences encompass

useful phylogenetic information [97]. First, the Transdecoder-predicted AA sequences were

trimmed using CD-HIT with the identity threshold of 0.995. Then, all-vs-all BLASTP with an E-

value cutoff of 10^{-5} search was implemented. The raw BLASTP output was filtered by hit

fraction of 0.4. Then, MCL clustering was performed with inflation point parameter of 2. Each

cluster was aligned using iterative algorithm of PASTA [85] and then was used to infer a

maximum-likelihood (ML) gene tree using IQ-TREE v1.5.2 [86] with an automatic model

selection. Tree tips that were longer than relative and absolute cutoffs of 0.4 and 1 respectively

were removed. Mono- and paraphyletic tips that belonged to the same species were masked as

well. To increase quality of homology clusters realignment, tree inference and tip masking steps

were iterated with more stringent relative and absolute masking cutoffs of 0.2 and 0.5

respectively. Finally, POs (AA sequences and corresponding CDSs) were extracted by rooted

ingroups (RI) procedure using *Ephemera danica* as an outgroup (for details see [81]).

Cluster alignment, trimming and supermatrix assembly

For most of the analyses only clusters with ≥ 42 (~50%) species present were retained. In total, we obtained five cluster types, namely DNA (CDS) and AA COs, AA AOs and DNA and AA POs. Each cluster was aligned using PASTA [85] for the DNA and AA alignments and PRANK v150803 [87, 88] for the codon alignments and alignments with removed either 1st and 2nd or 3rd codon positions. In order to reduce the amount of randomly aligned regions, we implemented ALISCORE v2.0 [89] trimming procedure (for PASTA alignments) followed by masking any site with ≥ 42 gap characters (for both PASTA and PRANK alignments). Also, sequence fragments with >50% gap characters were removed from clusters that were subjected to ASTRAL v4.10.12 [90] estimation since fragmentary data may have a negative effect on accuracy of gene and hence species tree inference [98]. For each of the cluster type, we assembled supermatrices from trimmed gene alignments. Additionally, completely untrimmed supermatrices were generated from DNA and AA COs with ≥ 5 species present.

Phylogenetic tree reconstruction

Four contrasting tree building methods (ML:IQTREE, Bayesian:ExaBayes, Supertree:ASTRAL, Alignment-Free (AF): Co-phylog) were used to infer odonate phylogenetic relationships using different input data types (untrimmed and trimmed supermatrices, codon supermatrices, codon supermatrices with 1st and 2nd or 3rd positions removed, gene trees and assembled transcriptomes). In total we performed 48 phylogenetic analyses and compared topologies to identify stable and conflicting relationships (Data S2).

We inferred phylogenetic ML trees from each supermatrix using IQTREE implementing two partitioning schemes: single partition and those identified by PartitionFinder v2.0 (three GTR models for DNA and a large array of protein models for AA) [91] with relaxed hierarchical clustering option [99]. In the first case, IQTREE was run allowing model selection and assessing nodal support with 1000 ultrafast bootstrap (UFBoot) [100] replicates. In the second case, IQTREE was run with a given PartitionFinder partition model applying gene and site resampling to minimize false-positives [101] for 1000 UFBoot replicates.

For Bayesian analyses implemented in ExaBayes [92], we used highly trimmed (retaining sites only with occupancy of ≤ 5 gap characters) and original DNA and AA CO supermatrices assuming a single partition. We initiated 4 independent runs with 4 Markov Chain Monte Carlo

(MCMC) coupled chains sampling every 500th iteration. Due to high computational demands of

the procedure, only the GTR and JTT substitution model priors were applied to DNA and AA

CO supermatrices respectively with the default topology, rate heterogeneity and branch lengths

priors. However, all supported protein substitution models as a prior was specified for the

trimmed AA CO supermatrix. For convergence criteria an average standard deviation of split

frequencies (ASDSF) [102], a potential scale reduction factor (PSRF) [103] and an effective

sample size (ESS) [104] were utilized. Values of 0% < ASDSF <1% and 1% < ASDSF < 5%

indicate excellent and acceptable convergence respectively; ESS >100 and PSRF ~ 1 represent

good convergence (see ExaBayes manual, [92]).

ASTRAL analyses were conducted using two input types: (i) gene trees obtained by IQ-

TREE allowing model selection for fully trimmed DNA and AA clusters and (ii) gene trees

obtained from the alignment-tree coestimation process in PASTA. Nodal support was assessed

by local posterior probabilities [105]. ASTRAL is a statistically consistent supertree method

under the multispecies coalescent model with better accuracy than other similar approaches [90].

In addition to standard phylogenetic inferential approaches, we applied an alignment-free

(AF) species tree estimation algorithm using Co-phylog [93]. Raw Transdecoder CDS outputs

were used in this analysis using k-mer size of 9 as the half context length required for Co-phylog.

Bootstrap replicate trees were obtained by running Co-Phylog with the same parameter settings

on each subsampled with replacement CDS Transdecoder libraries and were used to assess nodal

support.

Assessment of phylogenetic support via quartet sampling

As an additional phylogenetic support, we implemented quartet sampling (QS) approach [47].

Briefly, it provides three scores for internal nodes: (i) the quartet concordance (QC) score gives

an estimate of how sampled quartet topologies agree with the putative species tree; (ii) quartet

differential (QD) estimates frequency skewness of the discordant quartet topologies, which can

be indicative of introgression if a skewed frequency is observed and (iii) quartet informativeness

(QI) quantifies how informative sampled quartets are by comparing likelihood scores of

alternative quartet topologies. Finally, QS provides a quartet fidelity (QF) score for terminal

nodes that measures a taxon "rogueness". We performed QS analysis with all 48 putative species

phylogenies using SuperMatrix_50BUSCO_dna_pasta_ali_trim supermatrix, specifying an IQ-

TREE engine for quartet likelihood calculations with 100 replicates (i.e. number of quartet draws

per focal branch).

Fossil dating

A Bayesian algorithm of MCMCTREE [94] with approximate likelihood computation was

implemented to estimate divergence times within Odonata using 20 crown group fossil

constraints with corresponding prior distributions (Data S3). First, we estimated branch length by

ML and then the gradient and Hessian matrix around these ML estimates in MCMCTREE using

SuperMatrix_50BUSCO_dna_pasta_ali_trim supermatrix. Second, we used the gradient and

Hessian matrix which construct0 an approximate likelihood function by Taylor expansion [106]

to perform fossil calibration in MCMC framework. For this step we specified GTR+Γ

substitution model with 4 gamma categories; birth, death and sampling parameters of 1, 0.5 and

0.01, respectively. To ensure convergence, the analysis was run independently 10 times for 10^6

generations, logging every 50th generation and then removing 10% as burn-in. Visualization of

the calibrated tree was performed in R using MCMCtreeR package [95].

Analyses of introgression

In order to address the scope of possible reticulate evolution across odonate phylogeny, we used

various methods of introgression detection such as HyDe/D [96], D_{FOIL} [61], χ^2 goodness-of-fit

test, QuIBL [63] and PhyloNet [64, 65].

The HyDe framework allows detection of hybridization events which relies on

quantification of phylogenetic site patterns (invariants). HyDe estimates whether a putative

hybrid population (or taxon) H is sister to either population P1 with probability γ or to P2 with

probability 1-γ in a 4-taxon (quartet) tree ((P1,H,P2),O), where O denotes an outgroup. Then, it

conducts a formal statistical test of H_0: $\gamma = 0$ vs. H_1: $\gamma > 0$ using Z-test, where $\gamma = 0$ (=1) is

indicative of non-significant introgression. We applied HyDe to the concatenated supermatrix

SuperMatrix_50BUSCO_dna_pasta_ali_trim of 1603 BUSCO genes under default parameters

specifying *Ephemera danica* as an outgroup. Under this setup HyDe evaluates all possible taxa

quartets. Since HyDe only allows indication of a single outgroup taxon (i.e. *Ephemera danica*),

we excluded all quartets that contained the *Isonychia kiangsinensis* outgroup from the HyDe

output. Additionally, we calculated Patterson's D statistic [54] for every quartet from the

frequency (p) of ABBA-BABA site patterns estimated by HyDe as $D = \frac{p_{ABBA}-p_{BABA}}{p_{ABBA}+p_{BABA}}$. To test

significance of D statistics we used a χ^2 test to assess whether the proportions p_{ABBA} and p_{BABA}

were significantly different. To minimize effect of false positive cases (type I error) in the

output, we first applied a Bonferroni correction to the P values derived from Z- and χ^2 tests and

then filtered the results based on a significance level of 10^{-6} for both D and γ. Also, we noticed

that negative values of D were always associated with nonsignificant γ values, thus all of the D

values remained positive after filtering was applied.

D_{FOIL} is an alternative site pattern-based approach that detects introgression using

symmetric 5-taxon (quintet) trees, i.e. (((P1,P2),(P3,P4)),O). D_{FOIL} represents a collection of

similar to D statistics applied to quintet trees and, if considered simultaneously, they provide a

powerful approach to identify introgression including ancestral as well as donor and recipient

taxa (i.e. introgression directionality). Moreover, D_{FOIL} exhibits exceptionally low false positive

rates [61]. Since the number of possible quintet topologies for a phylogeny of 85 taxa is >

32×10^6, for analysis we extracted them only for every odonate suborder individually with the

custom R scripts. Note, that for Anisozygoptera we only considered quintets that can be formed

between Anisozygoptera, Anisoptera, and Zygoptera taxonomic groups. Analogously to HyDe,

we applied D_{FOIL} to the concatenated supermatrix SuperMatrix_50BUSCO_dna_pasta_ali_trim

of 1603 BUSCO genes under default parameters specifying *Ephemera danica* as an outgroup.

Also, since D_{FOIL} requires that every quintet has a symmetric topology we considered only those

quintets within our phylogeny that met this criterion (Figure 1). Additionally, D_{FOIL} requires that

the divergence time of P3 and P4 precedes divergence of P1 and P2, thus we switched species

assignments for those cases that violated this assumption, i.e. from (((P1,P2),(P3,P4)),O) to

(((P3,P4),(P1,P2)),O).

As an alternative test for introgression we performed a simple yet conservative χ^2

goodness-of-fit test on the gene tree count values for each triplet. First, we tested the three count

values to see whether their proportions significantly deviate from 1/3, and if the null hypothesis

is *not* rejected it will be indicative of extreme levels of ILS. Second, we tested the two count

values that originated from discordant gene trees to see whether their proportions significantly

deviate from 1/2, and if the null hypothesis is rejected, it will suggest a presence of introgression.

In order to correct the P values resulted from the χ^2 test for multiple testing, we applied the

Benjamini-Hochberg procedure at a false discovery rate cutoff (FDR) of 0.05.

QuIBL is based on the analysis of branch length distributions across gene trees to infer
putative introgression patterns. Briefly, under coalescent theory internal branches of rooted gene
trees for a set of 3 taxa (triplet) can be viewed as a mixture of two distributions with the
underlying parameters. Each mixture component generates branch lengths corresponding to
either ILS or introgression/speciation. Thus, estimated mixing proportions (π_1 for ILS and π_2 for
introgression/speciation; $\pi_1 + \pi_2 = 1$) of those distribution components show what fraction of the
gene trees were generated through ILS or non-ILS processes. For a given triplet, QuIBL
computes frequency of gene trees that support three alternative topologies. Then for every
alternative topology QuIBL estimates mixing proportions along with other relevant parameters
via Expectation-Maximization and computes Bayesian Information Criterion (BIC) scores for
ILS-only and introgression models. For concordant topologies elevated values of π_2 are expected
whereas for discordant ones π_2 can vary depending on the severity of ILS/intensity of
introgression. In extreme cases when the gene trees were generated exclusively under ILS, π_2
will approach to zero and the expected gene tree frequency for each alternative topology of a
triplet will be approximately 1/3. To identify significant cases of introgression here we used a
stringent cutoff of $\Delta BIC < -30$ [63]. We ran QuIBL on every triplet individually under default
parameters with number of steps (`numsteps` parameter) is equal to 50 and specifying one of
the Ephemeroptera species for triplet rooting. For computational efficiency we extracted triplets
only from the odonate superfamilies in a similar manner as we did for D_{FOIL} (see above). For this
analysis we used 1603 ML gene trees estimated from CO orthology clusters.
In order to estimate phylogenetic networks from the 1603 ML gene trees estimated from
CO orthology clusters, we used pseudolikelihood (InferNetwork_MPL) and likelihood
(CalGTProb) approaches implemented in PhyloNet. For both analyses we only selected gene
trees that had at least one of the outgroup species (*Isonychia kiangsinensis* and *Ephemera*
danica) and at least three ingroup taxa. We performed network searches for our focal clades,
namely Anisozygoptera, Aeshnidae, Gomphidae+Petaluridae and Libellulidae. For
pseudolikelihood analysis, we ran PhyloNet on every clade individually allowing a single
reticulation event, with the starting tree that corresponds to the species phylogeny (`-s` option),
100 iterations (`-x` option), 0.9 bootstrap threshold for gene trees (`-b` option) and optimization of
branch lengths and inheritance probabilities on the inferred networks (`-po` option). To ensure
convergence, the network searches were repeated 3 times for every clade. For the

Anisozygoptera, we explicitly indicated *Epiophlebia superstes* as a putative hybrid (–h option).

For the full likelihood estimation, we fixed the topology of a focal clade (equivalent to the

species tree topology) and calculated likelihood scores for possible networks with a single

reticulation (generated with a custom script) using CalGTProb. Additionally, to asses

significance of networks, we used difference of BIC scores (ΔBIC) derived from network

without reticulation (i.e. tree) and a network with a reticulation.

Dimensionality reduction and visualization

To uncover and visualize complex relationships between site pattern frequencies and Patterson's

D statistic and Hyde γ parameter we implemented a dimensionality reduction technique t-

distributed stochastic neighbor embedding (tSNE) [60] under default parameters in R.

Specifically we estimated tSNE maps from counts of 15 quartet site patterns calculated by HyDe

("AAAA" ,"AAAB" ,"AABA" , "AABB" , "AABC" , "ABAA" , "ABAB", "ABAC" , "ABBA"

, "BAAA" ,"ABBC" , "CABC" , "BACA" , "BCAA" , "ABCD").

QUANTIFICATION AND STATISTICAL ANALYSES

All statistical analyses were performed in R (https://www.R-project.org/). Statistical significance

was tested using the Wilcoxon Rank Sum test, Fisher Exact test and/or χ^2 test.

DATA AND SOFTWARE AVAILABILITY

All raw RNA-seq read libraries generated for this study have been uploaded to the Short Read

Archive (SRA) (SRR12086708-SRR12086765). All assembled transcriptomes, alignments,

inferred gene and species trees, fossil calibrated phylogenies (including paleontological record

assessed from PaleoDB used to plot the fossil distribution in Figure 1), PhyloNet results, QS

results and introgression results are available on figshare

(https://doi.org/10.6084/m9.figshare.12518660). Custom scripts that were used to create various

input files as well as the analysis pipeline are available on GitHub

https://github.com/antonysuv/odo_introgression.