

Bright Near-infrared Anti-Stokes Fluorescence of ICG under Low Power CW Laser Excitation and its Applications in Bioimaging

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Abstract: Anti-Stokes fluorescence was observed in ICG, a molecule approved by the FDA for clinical use. The wavelengths of its fluorescence are mainly located in the near-infrared band of 800 nm~900 nm, with a high quantum yield up to 8%. In order to know its generation mechanism, based on multi-photon absorption (MPA) theory, thermally activated delayed fluorescence (TADF) theory and hot band absorption theory, its power dependence, temperature dependence of absorption spectra and fluorescence spectra, and fluorescence lifetime were measured. Its generation mechanism was finally determined to be hot band absorption process. Since ICG showed bright anti-Stokes fluorescence in near-infrared region, which offers substantially longer penetration depth in biological tissues than visible light, excellent photostability and biosafety, we applied it to in vivo imaging and compared it with upconversion nanoparticles (UCNPs). The result is that ICG exhibited much stronger fluorescence than UCNPs, providing more anatomical information of samples. This contributes to a better choice for anti-Stokes fluorescence bioimaging.

Keywords: anti-Stokes fluorescence; FDA-approved molecule; hot-band absorption; near-infrared; high quantum yield

1 Introduction

Anti-Stokes luminescence is a special optical process, which converts long-wavelength excitation to short-wavelength emission. The generation mechanism of anti-stokes fluorescence can be divided into four classes, multi-photon absorption (MPA) process, upconversion process, thermally activated delayed fluorescence (TADF) process and hot band absorption process^{1,2}.

The MPA process refers to the material simultaneously absorbing multiple long-wavelength excited photons to generate one short-wavelength emitted photon³. This process usually occurs in several kind of materials such as dye molecules, AIE nanoparticles, quantum dots, etc⁴⁻⁶. The occurrence of MPA emission requires extremely high excitation light energy, so the femtosecond pulse laser is usually used as the excitation light source, which is quite expensive.

During the upconversion process, the upconversion nanoparticle (UCNP) composed of sensitizers and emitters absorbs multiple long-wavelength photons in succession to produce one short-wavelength photon. This process can be achieved by using a continuous wave (CW) laser, which is low cost. Upconversion process is usually divided into lanthanide (Ln)-based upconversion and triplet-triplet annihilation (TTA)-based upconversion⁷. The nanoparticles used for the lanthanide-based upconversion process are mainly composed of lanthanides, the absorption cross section of which is small, resulting in weak upconversion signals⁸. Meanwhile, such inorganic nanoparticles usually exhibit some deficiencies including low solubility and long metabolism time in the organism, which limit their applications in vivo^{9,10}. Another upconversion process, TTA-based upconversion, appears mainly in the nanoparticles containing organometallic complex sensitizers and organic emitters, seldom in the nanoparticles including only organic sensitizers and organic emitters¹¹. Such nanoparticles have large absorption coefficients and high quantum efficiency, so their upconversion signals are stronger than those of the lanthanide nanoparticles^{12,13}. However, their photostability is relatively low because of the quenching caused by molecular oxygen¹⁴.

Thermally activated delayed fluorescence (TADF) is mainly produced by organic molecules. In anti-Stokes luminescence process, the molecule only needs to absorb one long-wavelength photon, and then under thermal activation, it emits one short-wavelength photon. TADF can also be excited by the CW laser.

The generation mechanism of anti-Stokes fluorescence of dye molecules induced by hot band absorption is similar to

that of TADF, both of which involve the thermal activation. But in the hot band absorption process, the dye molecule is thermally activated at first, then absorbs one long-wavelength photon, and finally emit one short-wavelength photon. This anti-Stokes fluorescence can be excited by the CW laser. Compared with nanoparticles, the dye molecules can be excreted in a shorter time, imposing less side effects on the living body, which are more suitable for in vivo experiments. To date, most studies have focused on the commercial dyes with high quantum efficiency in the visible range, which are not suitable for in vivo imaging because of severe biological tissue scattering effect. The method of synthesizing molecules that can produce this anti-Stokes fluorescence is not clear, so the research on the hot band absorption materials is still in its infancy.

In our study, an FDA-approved dye molecule^{15, 16}, ICG, was found that it can produce bright near-infrared anti-Stokes fluorescence. To explore the potential mechanism about how it produces anti-Stokes fluorescence, a series of confirmatory experiments based on theoretical models were carried out. Moreover, imaging at in vivo level with ICG and comparison with the lanthanide upconversion nanoparticles were further performed to highlight the promising near-infrared anti-Stokes fluorophore. Advantages such as clinical availability, bright near-infrared anti-Stokes fluorescence with reduced scattering effect¹⁷⁻¹⁹ have suggested ICG's strong potential for medical application.

Theoretical models

Unlike the lanthanide nanoparticle or dye nanoparticle, as a dye molecule, ICG's anti-Stokes fluorescence does not belong to upconversion fluorescence, but belongs to one type of the following three, MPA fluorescence, TADF and hot band absorption fluorescence. The schematic illustration for the three theoretical models has been shown in Fig. 1. There are obvious differences between them, which can be used to determine the generation mechanism of anti-Stokes fluorescence of ICG.

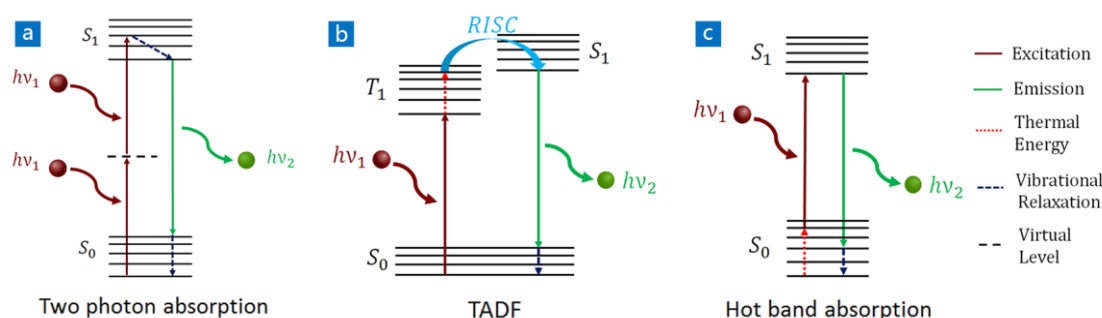


Fig.1 | Schematic illustration of different organic molecular anti-Stokes shift luminescence processes.

MPA process

The schematic of the energy level transition of the molecule in the MPA process has been shown in Fig. 1(a). A molecule in the ground state S_0 absorbs multiple photons simultaneously to the excited state S_1 (red solid line arrows). The total energy of the absorbed photons is equal to the molecular transition energy. Then the molecule relaxes to the lowest vibrational energy level of S_1 (the blue dotted arrow), and eventually transitions back to S_0 , emitting anti-Stokes fluorescence (the green solid arrow).

This is a non-linear optical process. The optical response of a non-linear medium can be determined by the polarization intensity P and the electric field intensity E of the incident light²⁰. The formula of the total polarization intensity is shown as follow:

$$P = P^{(1)} + P^{(2)} + P^{(3)} + \dots = \epsilon_0 (\chi^{(1)} E + \chi^{(2)} E^2 + \chi^{(3)} E^3) \quad (1)$$

ϵ_0 is the vacuum permittivity and $\chi^{(n)}$ is the n th-order polarizability of the dielectric material. Here $\chi^{(1)}$ is the linear polarization; $\chi^{(2)}$ is the second-order polarization; $\chi^{(3)}$ is the third-order polarization, $\chi^{(1)} \gg \chi^{(2)} \gg \chi^{(3)}$. The first term $\epsilon_0 \chi^{(1)} E$ corresponds to the absorption, scattering and reflection of light in a linear optical response; the second term $\epsilon_0 \chi^{(2)} E^2$ reflects second-order nonlinear optical effects such as second harmonic (SHG); the third term $\epsilon_0 \chi^{(3)} E^3$ describes two-photon and three-photon absorption, third harmonic (THG). When the incident light intensity is weak, the second and subsequent terms can be ignored, and only the first term is retained, which is a linear process. When the incident light intensity is high enough, the higher-order terms can no longer be ignored, so multi-photon absorption process can occur. This is why the MPA process is usually implemented with the femtosecond pulsed laser, rather than the CW laser.

The frequency-upconverted fluorescence intensity I_u is proportional to power n of the excitation intensity I_e , as formula 2 shows,

$$I_u \propto I_e^n \quad (2)$$

In the case of single-photon absorption process, $n=1$. If it is a non-linear process, for the two-photon absorption process, $n=2$, and for the three-photon absorption process, $n=3$.

For the MPA process, the absorption and fluorescence would both be predominantly independent of temperature.

TADF

In this model, as shown in Fig. 1(b), a molecule in the ground state S_0 absorbs a long-wavelength photon and directly reaches the triplet state T_1 (the red solid line arrow), and then undergoes a reverse intersystem crossing (RISC) process from the lowest triplet excited state to the lowest singlet excited state (the blue solid arrow) through thermal activation (the red dotted line arrow), emitting a short-wavelength photon (the green solid arrow). There is also triplet participation in the Stokes luminescence process.

Since the thermal activation process occurs in the triplet state, both anti-Stokes and Stokes fluorescence intensity is positively correlated with temperature. In addition, the molecular absorption coefficients are mainly independent of temperature.

Due to the participation of long-lived triplet states, the lifetime of this type of fluorescence is relatively long ($\sim \mu s$), which is several orders of magnitude longer than that of singlet fluorescence without triplet participation²¹⁻²⁴.

Hot band absorption process

The thermal activation in the hot band absorption process acts on the molecule in the ground state. As shown in Fig. 1(c), a molecule in the ground state with lower energy reaches the ground state with higher energy after thermal activation (the red dotted line arrow), then absorbs a long-wavelength photon to reach the excited state (the red solid line arrow), and finally emits a short-wavelength photon (the green solid arrow)²⁵⁻³⁰.

The occurrence of this type of anti-Stokes fluorescence is closely related to the vibrational energy level of the ground state S_0 . The population of vibrational energy levels in the ground state satisfies the Boltzmann distribution³¹. As formula 3 shows,

$$\frac{n_i}{n_0} = e^{-E_i/k_B T} \quad (3)$$

n_0 is the molecular populations of the lowest vibrational energy level in the ground state, while n_i corresponds to the the molecular populations of the higher vibrational energy level E_i in the ground state, k_B is the Boltzmann constant, and T is the temperature of the system. The higher the temperature is, the more molecules will be at the higher vibrational energy levels in the ground state, and the fewer molecules will be at the lowest energy level. Molecules in higher ground state vibrational levels will to produce anti-Stokes fluorescence. It can be seen that the absorption coefficients are temperature dependent, therefore, the fluorescence intensity should also be temperature dependent. Besides, the lifetime of anti-Stokes fluorescence is almost the same as that of Stokes fluorescence, both in the order of nanoseconds.

In order to determine which mechanism is responsible for ICG's anti-Stokes fluorescence, experiments were performed to distinguish these theoretical models.

Experimental details

Materials

ICG was purchased from DanDong Pharmaceutical Factory (Liaoning, China). DMSO was purchased from Sinopharm Chemical Reagent Co., Ltd., China. Fresh serum and bile were obtained from rats in our laboratory. The UCNP (NaYF₄: Yb³⁺, Tm³⁺) were acquired from Jilin University (Changchun, China). Phosphate buffered saline (PBS) was purchased from Sinopharm Chemical Reagent Co., Ltd., China.

Animal preparation

Sprague Dawley rats (female, 180g) were used for in vivo experiments. They were provided by the Zhejiang Academy of Medical Sciences and kept at the Experimental Animal Center of Zhejiang University. The room temperature of the rearing environment was maintained at 24 °C with a 12 h light/dark cycle, and the rats were continuously supplied with water and standard laboratory chow. All in vivo experiments were approved by the Institutional Ethical Committee of Animal Experimentation of Zhejiang University and strictly abided by "The National Regulation of China for Care and Use of Laboratory Animals". The rat was anesthetized at first. Then it was fixed on the imaging platform in the supine position, and shaving and laparotomy were performed to completely expose the imaging area of interest. Finally it was injected with materials through its inferior vena cava.

Experimental device for measuring absorption spectra, fluorescence spectra and power dependence

(915nm CW laser excitation)

The absorption spectra of ICG in DMSO were measured by UV-VIS-NIR spectrophotometer (CARY 5000, Agilent). The fluorescence spectra were acquired from a home-built system based on the PG2000 spectrometer (Ideaoptics Instruments). And the power dependence (915nm CW laser excitation) was derived from the fluorescence spectra excited by different

powers.

Experimental device for measuring lifetime and power dependence (915nm fs pulsed laser excitation)

The femtosecond pulsed laser (PHAROS PH1-10W, LIGHT CONVERSION; repetition rate: 1 MHz; pulse width: 200 fs) beam was introduced into a commercial inverted microscope (IX83, Olympus) as the excitation light. After the laser beam reflected by the dichroic mirror and passing through the objective (PLN20X, Olympus), the sample was excited. By using a turnover mirror, the emitted fluorescence signals passing through the dichroic mirror and filter were either delivered to a spectrometer (Andor, 193i + iXon DU-897U) or detected by the APD (τ -SPAD, PICOQUANT). Fluorescence spectra excited by different powers were measured with the spectrometer and used to analyze power dependence. The computer with integrated time-correlated single photon counting module system (TCSPC, DPC-230 Photon Correlator, Becker & Hickl GmbH) realized the measurement of the fluorescence lifetime of the sample based on the synchronous signals output by the laser and electrical signals from the APD.

Optical setup for anti-Stokes fluorescence wide-field imaging

The laser beam was coupled to a collimator, and then expanded by a lens with a ground glass sheet to provide a large illumination area. The ground glass sheet was used to eliminate laser speckles. The sample was placed in the position illuminated by the excitation light, and fluorescence signals were captured by a wide spectral response camera (GA1280, 1280 pixels \times 1024 pixels, TEKWIN SYSTEM, China) after passing through the prime lens (focal length: 35 mm, TEKWIN SYSTEM, China) coated with antireflection (AR) coating at 900-1700 nm and optical filters which were used to filter away the excitation light and extract the fluorescence signals.

Results and discussion

Molecular structure, absorption spectra, fluorescence spectra and the anti-Stokes fluorescence image

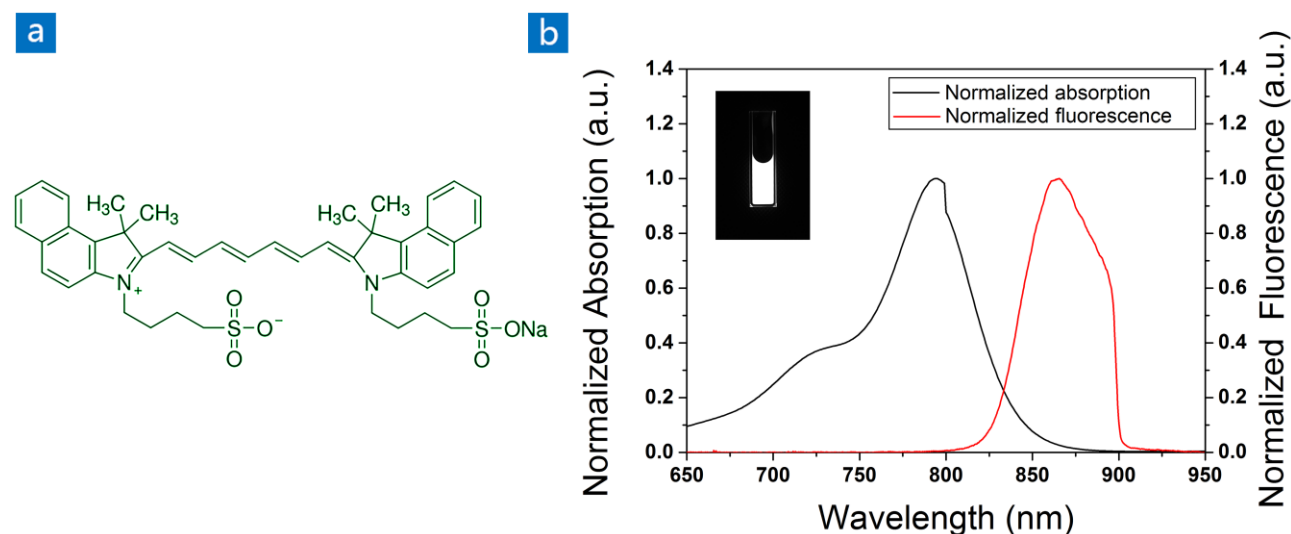


Fig.2 | Characterization of ICG molecule. (a) Molecular structure of ICG. (b) Normalized absorption spectra and normalized fluorescence spectra (900 nm short-pass) of ICG in DMSO. **Insert:** The anti-stokes fluorescence image of ICG (0.1 mg/mL, in DMSO) at the spectral region below 900 nm, under 915 nm CW laser irradiation (26.8 mW/cm²), exposure time: 25 ms.

The molecular structure of the ICG is shown in Fig. 2(a). The black curve in Fig. 2(b) is the absorption spectra (650nm-950nm) of 0.01mg/ml ICG in DMSO. The principle absorption band ($\lambda_{\max} = 794$ nm) in Fig. 2(b) corresponds to the transition from the lowest vibrational energy level (v_0) of the ground state (S_0) to the lowest vibrational energy level (v_0') of the excited state (S_1). The blue edge shoulder at 720 nm is associated with the $v_0 \rightarrow v_1'$ of the same $S_0 \rightarrow S_1$ electronic transition, where v_1' is a higher vibrational energy level of S_1 ³². 915nm is at the tail of the absorption spectra. The red curve in Fig. 2(b) is the anti-Stokes fluorescence spectra (< 900nm) of 0.1mg/ml ICG in DMSO under 915nm CW laser excitation. A 900 nm short-pass filter was used. Most of the fluorescence signals are located in the near infrared band of 800nm-900nm. Insert shows the strong anti-Stokes fluorescence from ICG. As previously reported, the full-spectra quantum efficiency of ICG dissolved in DMSO is 13%³³, so it can be concluded that the quantum efficiency between 800nm-900nm band should reach 8% proportionally.

Power dependence

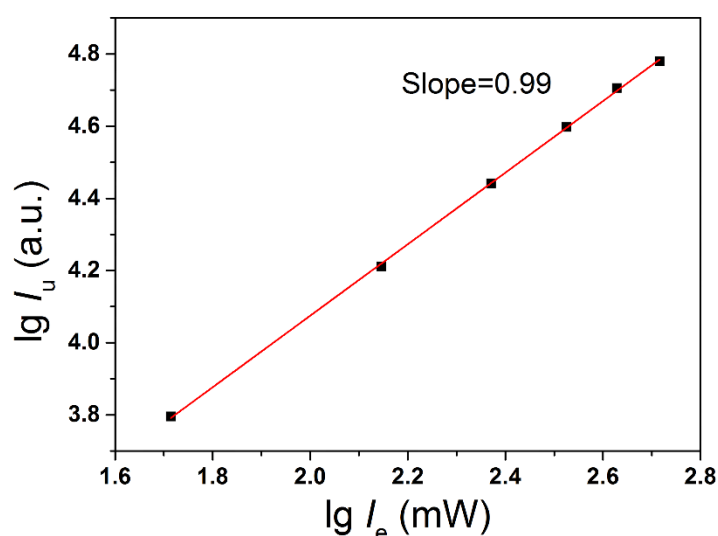


Fig.3 | Logarithmic plot of power dependence of fluorescence signal at 865 nm with excitation light (915 nm CW laser), showing a linear dependence of slope 0.99.

Since this anti-Stokes fluorescence can be excited by a CW laser, unlike the MPA fluorescence that must be excited by a pulsed laser with high peak power, the first step was to measure the power dependence of ICG in DMSO under 915nm CW laser excitation. The fluorescence spectra of 0.1mg/ml ICG in DMSO excited by different excitation powers were measured with a home-built system. According to the change of the fluorescence intensity at 865nm with the excitation power, a logarithmic plot of power dependence is drawn in Fig. 3. It can be seen that the value of n in formula 2 is 0.99, which is close to 1, indicating a linear anti-Stokes luminescence process, excluding the MPA mechanism.

Temperature dependence of the absorption spectra and the fluorescence spectra

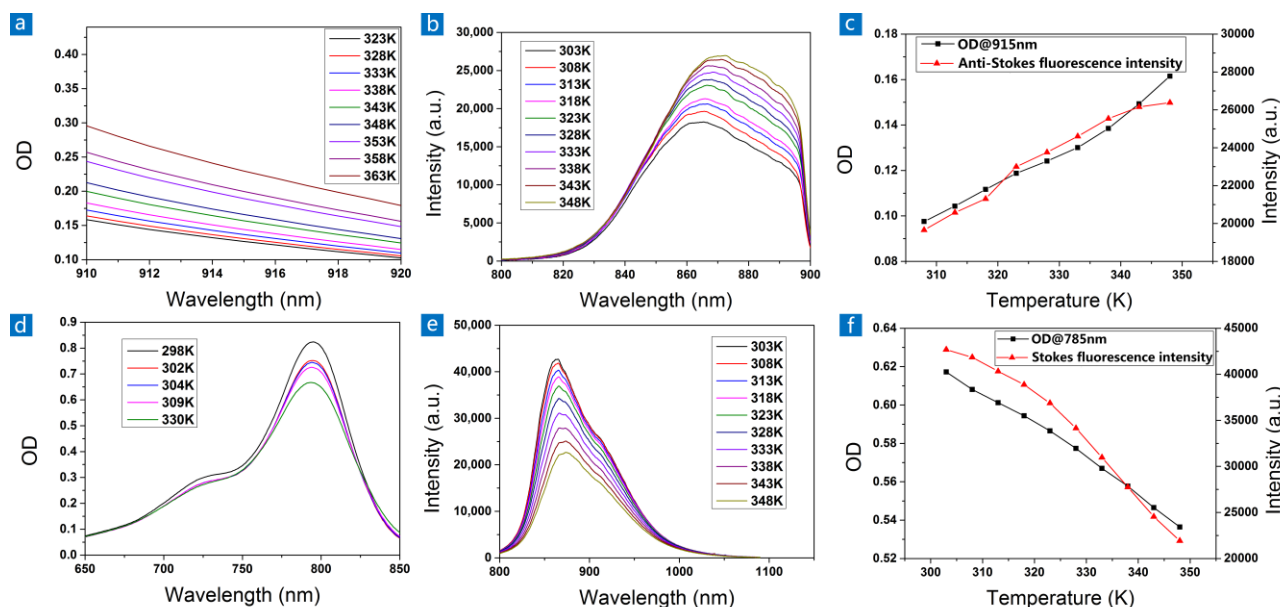


Fig. 4 | (a) Dependence of the absorption spectra (910 nm-920 nm) on temperature. (b) The variation of anti-Stokes fluorescence spectra with temperature under the excitation of 915 nm CW laser. (c) The black line is the variation of OD at 915 nm with temperature. The red line is the variation of anti-Stokes fluorescence intensity excited by 915 nm CW laser with temperature. (d) Dependence of the absorption spectra (650 nm-850 nm) on temperature. (e) The variation of Stokes fluorescence spectra with temperature under the excitation of 785 nm CW laser. (f) The black line is the

variation of OD at 785 nm with temperature. The red line is the variation of Stokes fluorescence intensity excited by 785 nm CW laser with temperature.

The next step was to determine whether the mechanism of anti-Stokes fluorescence is TADF or hot band absorption. The variations of absorption spectra and fluorescence spectra with temperature were measured, as shown in Fig. 4. As the temperature increases, the absorption at long wavelengths increase, while the absorption at short wavelengths decrease. The Boltzmann distribution function shown in formula 3 is a reasonable explanation of this phenomenon. When the temperature rises, the number of molecules at higher vibrational levels in the ground state increases, and the number of molecules at the lowest energy level decreases accordingly. Therefore, with the increasement of the absorption at long wavelengths in Fig 4. (a), both the main band and the higher energy shoulder shown in Fig. 4 (d) decreases at the same time. The anti-Stokes fluorescence spectra excited by 915 nm CW laser in Fig 4. (b) rises correspondingly due to the increased absorption at 915nm as the temperature rises from 303K to 348K, and the Stokes fluorescence spectra excited by 785 nm CW laser in Fig 4. (e) decreases accordingly. Fig 4. (c) and Fig 4. (f) intuitively show that the trends of the absorption at excitation wavelength and the fluorescence intensity excited by it changing with temperature are consistent.

Although the temperature dependence of the anti-Stokes fluorescence signals is also consistent with that of TADF, the temperature dependence of the Stokes fluorescence signals and absorption is not. Therefore, the generation mechanism of this anti-Stokes fluorescence is likely to be “hot band absorption”.

Lifetime of anti-Stokes and Stokes fluorescence

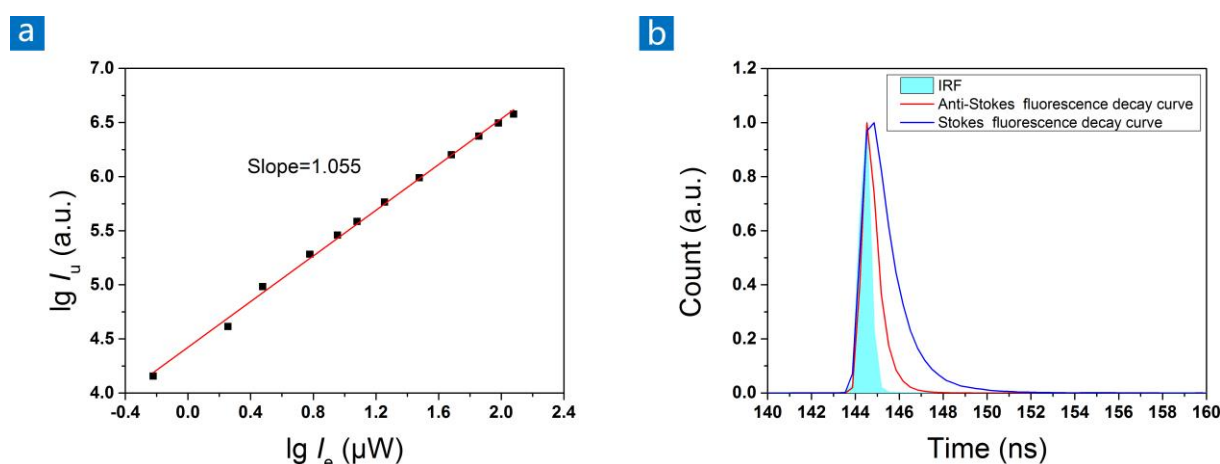


Fig.5 | Lifetime of ICG's anti-Stokes and Stokes fluorescence at a concentration of 2 mg/ml. (a) The power dependence of the anti-Stokes fluorescence signal with input power produced by a 915 nm femtosecond pulsed laser. **(b)** The cyan area is the impulse response function (IRF) of the system. The red line is the Anti-Stokes fluorescence decay curve of ICG under 915 nm femtosecond pulsed laser excitation. The blue line is the Stokes fluorescence decay curve of ICG under 750 nm femtosecond pulsed laser excitation.

To further determine the mechanism, a time-correlated single-photon counting (TCSPC) equipment was used to measure the lifetime of anti-Stokes and Stokes fluorescence under the femtosecond pulsed laser excitation at 915 nm and 750 nm, respectively. In order to ensure that the measured anti-Stokes fluorescence lifetime did not involve non-linear processes, the appropriate excitation power was explored at first. The power dependence of anti-Stokes fluorescence was measured under the excitation of 915 nm femtosecond pulse laser. As shown in Fig. 5(a), when the power of 915 nm lasers is less than 120 μW after passing through the objective, only linear optical process is involved. So the 915 nm laser power after passing through the objective was set to 4.4 μW , and the 750 nm laser power was set to 49 nW. Then the anti-Stokes and Stokes fluorescence decay curves were measured. The results are shown in Fig. 5(b). The cyan area in Fig. 5(b) is the IRF of this system, whose decay time is 0.25 ns, shorter than that of fluorescence. So this system could be used to measure the fluorescence lifetime of ICG in DMSO. The anti-Stokes and Stokes fluorescence lifetime of 2 mg/ml ICG in DMSO can be obtained as 0.67 ns and 1.21 ns respectively, both on the order of nanoseconds. Therefore, “hot band absorption” should be the mechanism by which this anti-Stokes fluorescence is produced.

Comparison of anti-Stokes fluorescence intensity between ICG and UCNP in vitro

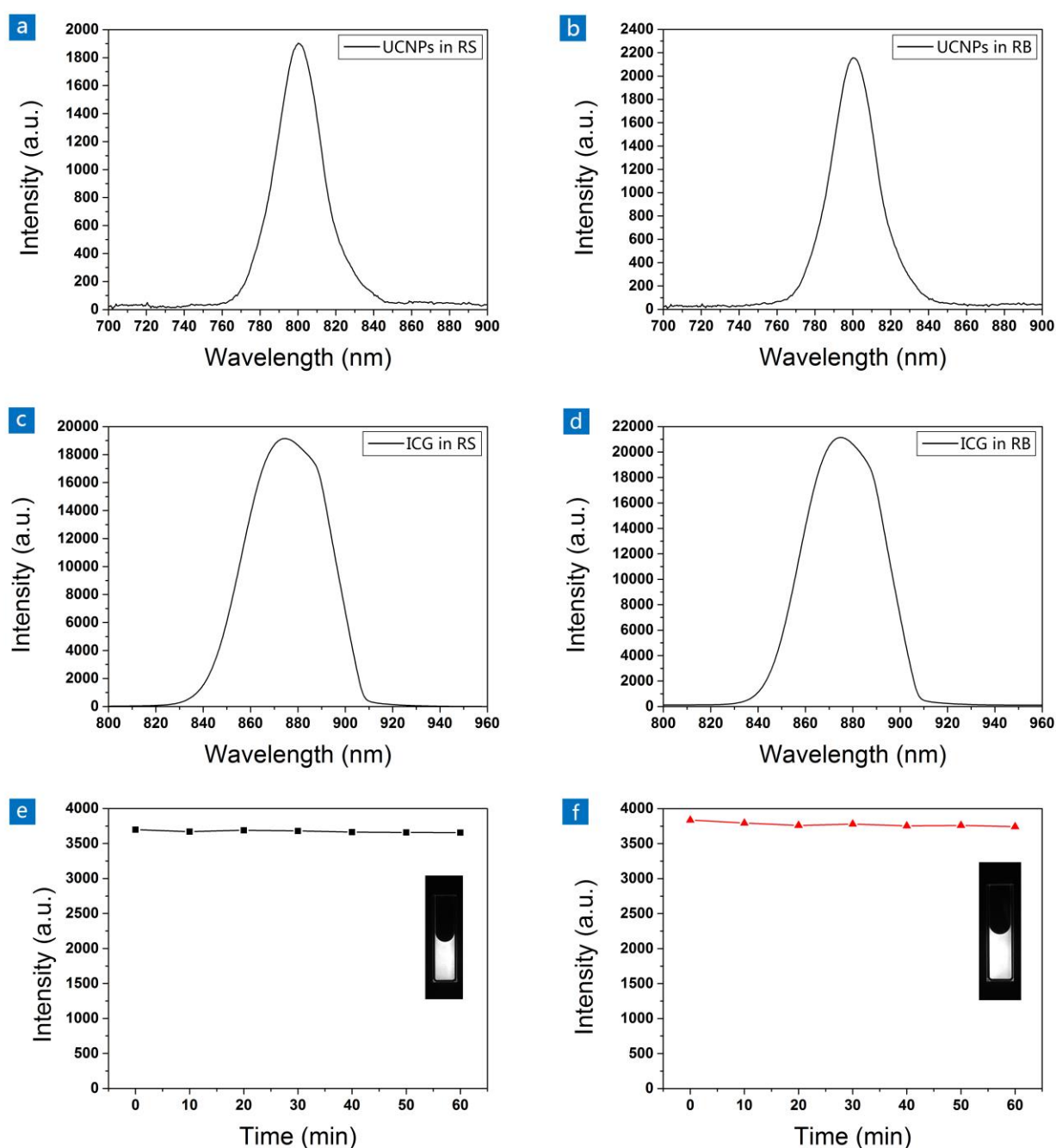


Fig.6 | (a) Anti-Stokes fluorescence spectra of NaYF₄: Yb³⁺, Tm³⁺ (1 mg/ml) in rat serum under 980 nm CW laser irradiation (1.8 W/cm²). Integration time: 4 s. (b) Anti-Stokes fluorescence spectra of NaYF₄: Yb³⁺, Tm³⁺ (1 mg/ml) in rat bile under 980 nm CW laser irradiation (1.8 W/cm²). Integration time: 4 s. (c) Anti-Stokes fluorescence spectra of ICG (0.1 mg/ml) in rat serum under 915 nm CW laser irradiation (0.9 W/cm²). Integration time: 0.5 s. (d) Anti-Stokes fluorescence spectra of ICG (0.1 mg/ml) in rat bile under 915 nm CW laser irradiation (0.9 W/cm²). Integration time: 0.5 s. (e) Photostability of ICG (0.1 mg/ml) in rat serum under 915 nm CW laser irradiation (68 mW/cm²). (f) Photostability of ICG (0.1 mg/ml) in rat bile under 915 nm CW laser irradiation (68 mW/cm²). **Insert:** The anti-Stokes fluorescence images of 0.1 mg/ml ICG in rat serum (e) and rat bile (f) at the spectral region below 900 nm, under 915 nm CW laser irradiation (68 mW/cm²), exposure time: 25 ms.

The generation mechanism of ICG's anti-Stokes fluorescence has been determined. Considering that it is an FDA-approved clinical fluorescent contrast agent, it was used for bioimaging and compared with UCNPs. UCNPs are a popular class of anti-Stokes luminescent materials that are often used in bioimaging. NaYF₄: Yb³⁺, Tm³⁺, a type of UCNPs, was selected to do this work. Comparison of ICG and UCNPs on anti-Stokes fluorescence intensity was performed at in vitro level firstly. The anti-Stokes fluorescence spectra of 1 mg/ml NaYF₄: Yb³⁺, Tm³⁺ in rat serum and bile are shown in Fig. 6(a) and Fig. 6(b). It can be seen that the peak fluorescence intensity at 800 nm is only around 2000. However, as shown

in Fig. 6(c) and Fig. 6(d), under the conditions of lower excitation power and less integration time, the peak fluorescence intensity at 865 nm of 0.1 mg/ml ICG can reach 20000. Therefore, ICG is much brighter than NaYF₄: Yb³⁺, Tm³⁺ both in rat serum and bile.

Since dyes often have the disadvantage of poor photostability^{34, 35}, photo-bleaching resistance analysis was also performed. As shown in Fig. 6(e) and (f), after the continuous irradiation of 915 nm CW laser whose power (68 mW/cm²) was sufficient for in vivo imaging for one hour, ICG showed almost no attenuation of the fluorescence signals, suggesting its excellent photostability under the irradiation of 915 nm CW laser.

In vivo wide-field imaging

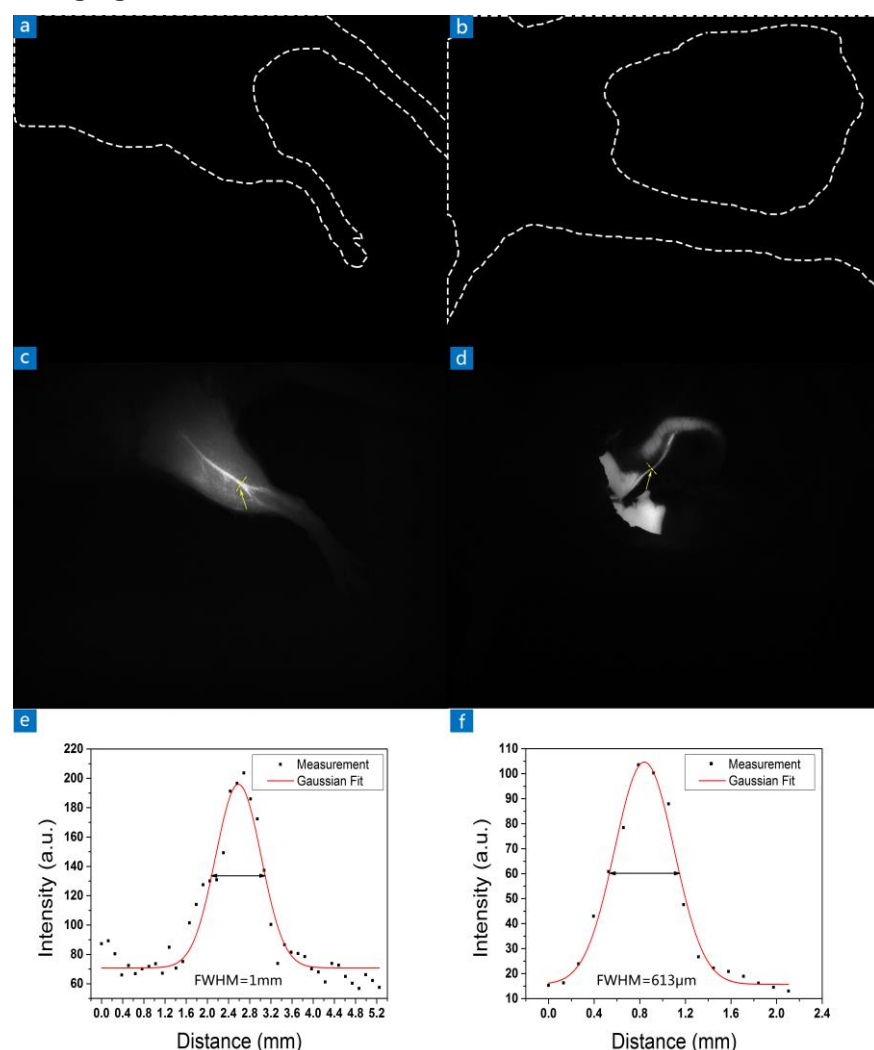


Fig.7 | (a) In vivo anti-Stokes fluorescence wide-field imaging of blood vessels of a rat after receiving injection of NaYF₄: Yb³⁺, Tm³⁺ (9.6 mg/ml, 500μl) under 980 nm CW laser irradiation (57 mW/cm²). (b) In vivo anti-Stokes fluorescence wide-field imaging of the biliary tract of a rat after receiving injection of NaYF₄: Yb³⁺, Tm³⁺ (9.6 mg/ml, 500μl) under 980 nm CW laser irradiation (57 mW/cm²). (c) In vivo anti-Stokes fluorescence wide-field imaging of blood vessels of a rat after receiving injection of ICG (1 mg/ml, 200μl) under 915 nm CW laser irradiation (16.5 mW/cm²). (d) In vivo anti-Stokes fluorescence wide-field imaging of the biliary tract of a rat after receiving injection of ICG (1 mg/ml, 300ul) under 915 nm CW laser irradiation (4.5 mW/cm²). Optical filter, 800 nm long-pass and 900 nm short-pass; exposure time, 25 ms. (e) The FWHM of the blood vessel pointed by the arrow in (c). (f) The FWHM of the biliary tract pointed by the arrow in (d).

Benefiting from ICG's bright near-infrared anti-Stokes fluorescence signals and less scattering of near-infrared light in biological tissues which is conducive for biological imaging, ICG was used to visualize blood vessels and biliary tracts in rats. As shown in Fig. 7(c) and Fig. 7(d), the blood vessels in the hind limb of the rat injected with ICG and its biliary tract can be clearly identified, while no fluorescence signal could be detected on the blood vessels and biliary tract of the rat injected with NaYF₄: Yb³⁺, Tm³⁺ in Fig. 7(a) and Fig. 7(b). Fig. 7(e) and Fig. 7(f) show the FWHM of the hind limb blood vessel and biliary tract of the ICG-injected rat, indicating that the width of the hind limb large blood vessel and

biliary tract of the rat are 1 mm and 613 μm respectively. Compared with $\text{NaYF}_4: \text{Yb}^{3+}, \text{Tm}^{3+}$ with the absorption peak at 980nm, another advantage of using ICG's anti-Stokes fluorescence under excitation at 915 nm for bioimaging is that 980nm is also a water absorption peak, but the absorption of water at 915nm is quite low³⁶, which could effectively eliminate the photothermal effect and avoid thermal damage on biological tissues.

Conclusions

In summary, we found that ICG can produce bright near-infrared anti-Stokes fluorescence, outperforming the conventional UCNPs, and further elucidated the potential luminescence mechanism- "hot band absorption". Advantages including high quantum efficiency in the near-infrared window, deeper penetrating ability in biological tissues, excellent photostability and FDA-approved suggest ICG's promising application in bioimaging field. Besides, it also provides a new direction for the design of hot band absorbent materials, which could possibly be excellent candidates for different application areas such as OLED, imaging, sensing and even medical diagnostics and treatment.

References

1. Zhu X, Su Q, Feng W, Li F. Anti-Stokes shift luminescent materials for bio-applications. *Chem Soc Rev* **46**, 1025-1039 (2017).
2. Uoyama H, Goushi K, Shizu K, Nomura H, Adachi C. Highly efficient organic light-emitting diodes from delayed fluorescence. *Nature* **492**, 234-238 (2012).
3. Carriles R, Schafer DN, Sheetz KE, Field JJ, Cisek R, *et al.* Invited review article: Imaging techniques for harmonic and multiphoton absorption fluorescence microscopy. *Rev Sci Instrum* **80**, 081101 (2009).
4. He T, Sreejith S, Gao Y, Grimsdale AC, Zhao Y, *et al.* Superior optical nonlinearity of an exceptional fluorescent stilbene dye. *Appl Phys Lett* **106**, 111904 (2015).
5. Zheng Z, Li D, Liu Z, Peng H-Q, Sung HHY, *et al.* Aggregation-Induced Nonlinear Optical Effects of AIEgen Nanocrystals for Ultradeep In Vivo Bioimaging. *Adv Mater* **31**, 1904799 (2019).
6. Larson DR, Zipfel WR, Williams RM, Clark SW, Bruchez MP, *et al.* Water-soluble quantum dots for multiphoton fluorescence imaging in vivo. *Science* **300**, 1434-6 (2003).
7. Liu Q, Feng W, Yang T, Yi T, Li F. Upconversion luminescence imaging of cells and small animals. *Nat Protoc* **8**, 2033-44 (2013).
8. Auzel F. Upconversion and Anti-Stokes Processes with f and d Ions in Solids. *Chem Rev* **104**, 139-174 (2004).
9. Zhou J, Liu Z, Li F. Upconversion nanophosphors for small-animal imaging. *Chem Soc Rev* **41**, 1323-49 (2012).
10. Longmire M, Choyke PL, Kobayashi H. Clearance properties of nano-sized particles and molecules as imaging agents: considerations and caveats. *Nanomedicine* **3**, 703-717 (2008).
11. Zhao J, Ji S, Guo H. Triplet-triplet annihilation based upconversion: from triplet sensitizers and triplet acceptors to upconversion quantum yields. *RSC Adv* **1**, 937-950 (2011).
12. Liu Q, Yang T, Feng W, Li F. Blue-Emissive Upconversion Nanoparticles for Low-Power-Excited Bioimaging in Vivo. *J Am Chem Soc* **134**, 5390-5397 (2012).
13. Balushev S, Miteva T, Yakutkin V, Nelles G, Yasuda A, *et al.* Up-Conversion Fluorescence: Noncoherent Excitation by Sunlight. *Phys Rev Lett* **97**, 143903 (2006).
14. Askes SHC, Leeuwenburgh VC, Pomp W, Arjmandi-Tash H, Tanase S, *et al.* Water-Dispersible Silica-Coated Upconverting Liposomes: Can a Thin Silica Layer Protect TTA-UC against Oxygen Quenching? *ACS Biomater Sci Eng* **3**, 322-334 (2017).
15. Treger Jeremy S, Priest Michael F, Iezzi R, Bezanilla F. Real-Time Imaging of Electrical Signals with an Infrared FDA-Approved Dye. *Biophys J* **107**, L09-L12 (2014).
16. Yu X, Feng Z, Cai Z, Jiang M, Xue D, *et al.* Deciphering of cerebrovasculatures via ICG-assisted NIR-II fluorescence microscopy. *J Mater Chem B* **7**, 6623-6629 (2019).
17. Jin H-G, Zhong W, Yin S, Zhang X, Zhao Y-H, *et al.* Lesson from Nature: Biomimetic Self-Assembling Phthalocyanines for High-Efficient Photothermal Therapy within the Biological Transparent Window. *ACS Appl Mater Interfaces* **11**, 3800-3808 (2019).
18. Hong G, Antaris AL, Dai H. Near-infrared fluorophores for biomedical imaging. *Nat Biomed Eng* **1**, 0010 (2017).
19. Frangioni JV. In vivo near-infrared fluorescence imaging. *Curr Opin Chem Biol* **7**, 626-634 (2003).
20. Panoui NC, Sha WEI, Lei DY, Li GC. Nonlinear optics in plasmonic nanostructures. *J Opt* **20**, 083001 (2018).
21. Chen Z, Wu Z, Ni F, Zhong C, Zeng W, *et al.* Emitters with a pyridine-3,5-dicarbonitrile core and short delayed fluorescence lifetimes of about 1.5 μs : orange-red TADF-based OLEDs with very slow efficiency roll-offs at high luminance. *J Mater Chem C* **6**, 6543-6548 (2018).
22. Skaisgirius R, Serevičius T, Kazlauskas K, Geng Y, Adachi C, *et al.* Origin of dual emission in σ -bridged donor-acceptor TADF compounds. *J Mater Chem C* **7**, 12601-12609 (2019).
23. Yuan J, Tang Y, Xu S, Chen R, Huang W. Purely organic optoelectronic materials with ultralong-lived excited states under ambient conditions. *Sci Bull* **60**, 1631-1637 (2015).
24. Chen J-X, Tao W-W, Chen W-C, Xiao Y-F, Wang K, *et al.* Red/Near-Infrared Thermally Activated Delayed Fluorescence OLEDs with Near 100% Internal Quantum Efficiency. *Angew Chem*, 10.1002/ange.201906575 (2019).
25. Erickson LE. On anti-stokes luminescence from Rhodamine 6G in ethanol solutions. *J Lumin* **5**, 1-13 (1972).
26. Chang M, Elliott S, Gustafson T, hu C, Jain RK. Observation of Anti-Stokes Fluorescence in Organic Dye Solutions. *IEEE J Quantum*

Electron **8**, 527-528 (1972).

27. Bloor D, Cross G, Sharma P, Elliott J, Rumbles G. Frequency up-conversion in fluid and solid solutions of the oxazine dye, Rhodamine B. *J CHEM SOC FARADAY TRANS* **89**, 4013-4015 (1993).
28. Clark J, Rumbles G. Laser Cooling in the Condensed Phase by Frequency Up-Conversion. *Phys Rev Lett* **76**, 2037-2040 (1996).
29. Clark J, Miller P, Rumbles G. Red Edge Photophysics of Ethanol Rhodamine 101 and the Observation of Laser Cooling in the Condensed Phase. *J Phys Chem A* **102**, 4428-4437 (1998).
30. Liu Y, Su Q, Zou X, Min C, Feng W, *et al.* Near-Infrared in Vivo Bioimaging by Molecular Upconversion Probe. *Chem Commun* **52**, 7466-7469 (2016).
31. Treanor CE, Rich JW, Rehm RG. Vibrational Relaxation of Anharmonic Oscillators with Exchange-Dominated Collisions. *J Chem Phys* **48**, 1798-1807 (1968).
32. Pickett LW. A Vibrational Analysis of the Absorption Spectrum of Furan in the Schumann Region. *J Chem Phys* **8**, 293-297 (1940).
33. Benson R, Kues H. Absorption and fluorescence properties of cyanine dyes. *J CHEM ENG DATA* **22**, 379-383 (1977).
34. Ahmad M, King T, Ko D-K, Cha B, Lee J. Performance and photostability of xanthene and pyrromethene laser dyes in sol-gel phases. *J Phys D* **35**, 1473 (2002).
35. Knobbe ET, Dunn B, Fuqua PD, Nishida F. Laser behavior and photostability characteristics of organic dye doped silicate gel materials. *Appl Opt* **29**, 2729-2733 (1990).
36. Palmer KF, Williams D. Optical properties of water in the near infrared. *J Opt Soc Am* **64**, 1107-1110 (1974).

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Author contributions

All authors commented on the manuscript.

Jun Qian proposed this project and provided guidance and assistance to the project. Jing Zhou is the main executor of the project, exploring the mechanism of ICG's anti-Stokes fluorescence and using it for bioimaging. Di Wu extracted the serum and bile of rats, and provided assistance for shaving, dissecting and injecting materials to the rats in the in vivo imaging experiments. Zikang Ye helped to measure the fluorescence lifetime of ICG. Dingwei Xue and Mubin He provided some help for the in vivo imaging experiments during the project.

Competing interests

The authors declare no competing financial interests.

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