How will cold water fish respond to climate change: rapid and high

magnitude phenotypic responses in Arctic charr.

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ABSTRACT

Phenotypic plasticity is the ability of an organism to express multiple phenotypes in response to the prevailing environmental conditions without genetic change. This response may thus occur to anthropogenic modifications of that environment and may be asymmetric. Arguably, the most significant future anthropogenic modification of the environment is contemporary climate change. We tested the effect of a temperature differential of 4 ºC on the Arctic charr phenotypic response within a generation, mimicking the event of short term year-to-year variation and long-term change in temperature climate change models predicted for northern regions. We demonstrated that Arctic charr phenotype can respond rapidly and markedly to an environmental cue. The plastic response to different temperature regimes comprised a shift in the mean phenotype coupled with a reduction in the between-individual phenotypic variation in the expressed head shape. The magnitude of shape difference was cumulative over time but the rate of divergence diminished as fish became larger. Individuals raised in the elevated temperature treatment expressed a phenotype analogous to a benthivorous ecotype of this species rather than that of the parental pelagic feeding form. By confirming the potential of a rapid and marked variation with an increase temperature that swung from a pelagic toward a benthic phenotype, we provide evidence that a substantial portion of Arctic charr populations worldwide could be subjected to phenotype expression changes under climate change scenarios.

INTRODUCTION

In nature, the expression of intraspecific, discrete, alternative phenotypes can result from modulation by the environment through plasticity [1] or from the emergence of evolutionary groups with diverging gene pools [2], or a combination of both [3]. Phenotypic plasticity is the ability of a single genotype to express multiple alternative phenotypes in response to different environmental conditions [4]. Plasticity itself is a trait, present across a broad range of taxa [5-7]. It can be instantaneous, anticipatory or delayed, permanent or reversible, adaptive or non-adaptive, beneficial or harmful, passive, discrete, continuous, and generational [8]. It is known to facilitate the expression of novel phenotypic traits [9] upon which selection may then act [10]. Thus, phenotypic plasticity has an important role in the evolutionary processes of organisms that exhibit plasticity because variation fuels evolution [11].

Identifying pathways along which phenotypic plasticity can arise is still a major topic for inquiry in evolutionary ecology, since patterns and processes that underlie variation are multifaceted and highly variable. Despite the complexity of the nature of phenotypic plasticity, there are some abiotic factors that are known to be ubiquitous in driving variation in ecosystems, one of which is temperature [12, 13]. Many plants and animals have the capacity for a plastic response to temperature, as it is unlikely that rapid environmental change would induce a genetic response [14-16]. A rapid change in temperature could induce greater levels of phenotypic variation among individuals within a population by exposing previously hidden (cryptic) genetic variation or by inducing new epigenetic changes [15]. Accordingly, the response of an organism to temperature can include variation in the mean expressed phenotype itself but it can also include change in the variance of phenotypes expressed in a population [17].

As ectotherms, fish are particularly sensitive to change in temperature [18]. A phenotypic trait in fish that is strongly linked with temperature is growth [19, 20]. However, the subsequent effect of temperature and induced growth rate heterogeneity (e.g., early developmental and juvenile growth) on the modulation of morphology is not a mechanism fully understood, despite being observed to constraint or enhance morphology modulation in a number of species [21-23]. A few studies have looked at the potential for temperature change to cause heterochrony in the developmental and ontogenetic processes that modulate the expression of functional traits [24, 25], whilst others have attempted to predict the morphological outcome of temperature induced plasticity [12]. However, the ecological consequences that result from temperature induced phenotypic change has received little attention [26]. As the expression of some discrete functional phenotypes in some fish species is plastic, and by definition modulated by variation in the natural environment, it is presumed that they will also respond to anthropogenic modifications of that environment. The most obvious and far reaching anthropogenic modification of the contemporary environment is that of climate change. Climate change is predicted to raise the average temperature of the Earth's surface between 2-5 $^{\circ}$C in the next 85 years [27]. Change in the variability of climate can take place at different temporal scales (e.g. diurnally, intra-seasonally or inter-annually), whereas temperature extremes are now occurring more regularly and are predicted to increase even more in frequency in the future [28].

The Arctic charr (*Salvelinus alpinus*) is a species of freshwater fish with a highly variable phenotype [29]. The species is ideal to explore how phenotypic diversity is realized within and among generations as it expresses a high degree of phenotypic plasticity and rapid intra-specific divergence, and as a result of its range and habitat use, it experiences diverse selective environments, which facilitate tracking the link between ecological processes and evolutionary outcomes [1, 30-32]. In this experimental study we tested the effect of an elevated

water temperature of 4°C on the short-term, plastic, phenotypic expression in an Arctic charr

population. The induced temperature variation in this study is comparable in magnitude to year

on year of extreme temperature fluctuations predicted by climate change models [33]. We

predict that Arctic charr individuals reared in warmer water will express greater phenotypic

variation than fish at a control temperature (see Figure 1C) but that they will not display a

significant shift in the mean phenotype expressed (Figure 1B and D) [17]. Overall, we aimed

to: 1) quantify the among-individual morphological variation of the head of a unimodal Arctic

charr population exposed to different temperature regimes during early development, 2)

determine the allometry trajectories that result in any differences in expressed phenotype as

evidence of differing developmental pathways in groups exposed to different temperature and

3) examine if at different temperature treatments, head shape among individuals is more

analogous to the expressed phenotypes in plankton feeding (the parental phenotype) or macro-

benthos feeding ecotypes in the wild.

MATERIALS and METHODS

Fish collection and rearing

Nine full sibling crosses were produced from adult Arctic charr belonging to a

morphologically unimodal, plankton feeding population that inhabits Loch Clair, Scotland.

Fish were caught during spawning time from the in-flowing River Coulin (57°32.648'N,

5°19.125'W). Fertilised eggs were water hardened and transferred to incubation facilities at the

Scottish Centre for Ecology and the Natural Environment, Loch Lomond. Eggs were

acclimatised over a two-hour period to the new water supply and placed in mesh baskets

suspended in a holding tank in a constant temperature room maintaining a water temperature

of 4 °C (± 0.5 °C).

Experimental procedure

To control for differences in development due to different temperatures, the number of

degree-days (dd) were used as a measure of developmental rate. Degree-days are the

cumulative count of the water temperature for a known period of time in days. Eggs reached

the eyed stage after 212dd at which point they were raised to a temperature of 6 $^\circ$C ($\pm$ 0.5 $^\circ$C)

for a further 89dd, before being separated into two temperature treatment groups (n=480 per

group), an ambient and an elevated temperature treatment. Eggs were held in equal numbers in

eight replicate tanks per treatment group (n=60 per tank). Water temperatures were then either

lowered by 2 $^\circ$C to 4 $^\circ$C or raised by 2 $^\circ$C to 8 $^\circ$C ($\pm$ 0.5 $^\circ$C) respectively. Eggs exposed to the

ambient temperature (4 $^\circ$C) began hatching after 367dd, the hatching period lasted 73dd (total

developmental time to 100% hatch 440dd). Eggs exposed to the elevated temperature (8 $^\circ$C)

began hatching after 388dd, the hatching period lasted 50dd (total incubation time to 100%

hatch 438dd). When hatching was complete, temperatures were raised by 3 $^\circ$C; the ambient

temperature treatment to 7 $^\circ$C and the elevated temperature treatment to 11 $^\circ$C ($\pm$ 0.5 $^\circ$C). Fish

became partially dependant on exogenous food at 505dd for the ambient temperature treatment

and 504dd for the elevated temperature treatment. When the yolk sack was fully exhausted,

689dd for the ambient temperature treatment and 686dd for the elevated temperature treatment,

temperatures were raised a further 2 $^\circ$C for both treatments to 9 $^\circ$C (ambient temperature

treatment) and 13 $^\circ$C (elevated temperature treatment; $\pm$ 0.5 $^\circ$C). Fish were fed four times a day

to satiation at three hour intervals ($\pm$ 0.5 hours) using a standard commercial 3mm hatchery

sinking pellet.

Data collection

Adult Arctic charr used as brood stock were photographed in a lateral position on the left side

before spawning. Lateral view photographs of juveniles were taken at 700dd (N=130), 1000dd

(N=80) and 1400dd (N=60) using a Cannon EOS 350D digital camera, for geometric

morphometric analysis. Nine consistently identifiable landmarks on the head (Figure A1) were

digitised in two dimensions on each fish image using TPSdig2 (Rohlf, 2006a) and TPSutil

(Rohlf, 2006b).

Data analysis

Prior to geometric morphometric analysis, landmark data were subject to a Procrustes

superimposition using MorphoJ [34] to remove variation in the data created by size, position

and orientation [35]. The mean shape configuration was then computed and the variation

around this mean calculated [36].

Following this, a single, pooled within-group regression of Procrustes co-ordinates on

log centroid size was conducted in MorphoJ for samples collected at 700dd, 1000dd and

1400dd's. The residuals from this regression provide a measure of shape free from allometric

scaling [37] associated with early ontogeny. The residuals from this regression were

subsequently used for all further morphometric analysis.

A single Discriminant Function Analysis (1000 permutations) was conducted in

MorphoJ to compare geometric morphometric data from three developmental stages (700dd,

1000dd and 1400dd) to test if the degree of shape difference between groups changed over

time. Procrustes distance and Mahalanobis distance were used as pairwise measures of the

magnitude of shape difference. Within each temperature treatment (ambient vs warm), the

polynomial relationship between discriminant function scores and size (represented by centroid

size) were used to illustrate the presence of allometric trajectories. To decide whether

allometric trajectories between temperature treatments were parallel, convergent, divergent, or

common (see Figure A2), the slopes of the regressions were examined

Using the scores generated in the Discriminant Function Analysis as a measure of

shape, a generalised linear mixed effect model, fitted by maximum likelihood (using the

software R 3.1 for Windows [38] and the package lme4) was used to describe the effect of

temperature, exposure time (the number of dd at each developmental stage), fish size (measured

as centroid size) and replicate (tank) as a random effect, on the expression of shape. Model

simplification was conducted by removing the highest order, least significant terms,

sequentially [39]. Terms that were removed from the model without significantly increasing

model deviance using likelihood ratio tests (χ^2) were discarded. *Post hoc* pairwise comparisons

using T-test assessed if centroid size (related to body size) differed between ambient and

elevated treatments at each sampling time (700, 1000, and 1400dd).

A Canonical Variate Analysis (1000 permutations) executed in MorphoJ was used to

establish which group had been affected by the temperature treatment by comparing the head

shape of individuals raised in ambient or elevated temperature conditions in the laboratory to

their parents.

A variance ratio test was conducted using the software R 3.1 for Windows on all raw

Procrustes coordinates and used to compare within-group phenotypic variation between

groups.

To establish which temperature regime had stimulated expression of head shape closest

to the wild type, head shape of juveniles at 1400dd was compared against the head shape of

two known ecologically divergent sympatric polymorphic groups of Arctic charr that show

morphological and dietary specialisations on either plankton or macro-benthos feeding

resources, one from Loch Rannoch (Perthshire, Scotland) [40] and one from Loch Dughaill

(Strathcarron, Scotland) [41] using a Canonical Variate Analysis (1000 permutations) in

MorphoJ. The same nine consistently identifiable landmarks used for analysing laboratory

raised individuals were digitised in two dimensions on existing images of fish from Loch

Rannoch and Loch Dughaill.

RESULTS

The Discriminant Function Analysis (DFA) found head shape differed across all sampling periods between ambient and elevated temperature exposed fish (Procrustes distance = 0.028; P<0.01). The magnitude of shape differences however increased over time (at 700dd, Procrustes distance = 0.036; P<0.01; at 1000dd, Procrustes = 0.040; P<0.01; at 1400dd, Procrustes distance = 0.061; P<0.01; Figure 2).

Discriminant Function Analysis of shape only, correctly assigned 73.8% of ambient temperature fish and 82.8% of elevated temperature fish (from all sampling periods combined) to the correct temperature exposure. In general, elevated temperature fish had a more rounded head and sub-terminal mouth than ambient temperature fish (Figure A3). Individuals in both temperature treatments displayed allometric trajectories with polynomial relationships of head shape on size differing from 0 (P<0.05); centroid size explained 55% of variation in Discriminant Function Score of head shape in the ambient temperature treatment group (R^2 =0.55) and 57% in the elevated temperature treatment group (R^2=0.57). Regression slopes indicated that allometric trajectories were divergent (Figure 2 and Figure A2).

Using discriminant function scores as a measure of shape in a mixed model, temperature had a highly significant effect on head shape (likelihood ratio χ^2 = 39.81, p<0.01) (Table 1). There was a significant interaction between Exposure time and Centroid Size (likelihood ratio χ^2 = 7.23, p = 0.01) (Table 1). The interaction between exposure (number of degree-days) and fish size (centroid size) was negative. Thus the difference in morphology between fish raised on each temperature regime was greater in larger fish, but the rate of divergence decreased as fish became larger; indicating that speed of divergence caused by different temperature regimes is greater during early ontogenetic stages. Pairwise comparisons of centroid size (a measure of fish body size) between temperature treatments at each sampling time, showed a difference in size at 700dd (t = 1.98, p < 0.01) and at 1400dd (t = 2.0, p < 0.01),

but no size differences were found at 1000dd (P < 0.05). At 700dd and 1400dd, individuals

raised at ambient temperature were larger than individuals exposed to elevated temperature.

The variance ratio test found the phenotypic variation within treatment groups was

higher in ambient temperature fish compared with elevated temperature fish (F = <0.01, df =

269, p = <0.01). For ambient temperature, variance in Procrustes scores (SS distance/(n-1))

was 0.0046 at 700dd, 0.0032 at 1000dd, and 0.0018 at 1400dd. For elevated temperature,

variance of Procrustes scores (SS distance/(n-1)) was 0.0038 at 700dd, 0.0039 at 1000dd, and

0.0016 at 1400dd.

Canonical Variate Analysis (CVA) found that the head shape of fish exposed to the

ambient temperature treatment was not significantly different to that of their parents

(Procrustes distance = 0.014; *P*=0.38), however elevated temperature fish were significantly

different (Figure A4; Procrustes distance = 0.044; *P*=<0.001).

Using a CVA to compare experimental (Coulin) charr from both temperature groups to

benthic and pelagic foraging specialists from Loch Rannoch and Loch Dughaill, we found

ambient temperature fish expressed a phenotype closer to the pelagic ecotype and fish exposed

to elevated temperature displayed a phenotype closer to benthic ecotype for both comparisons

(Figure 3 and Table 2). CV1 mostly captures shape differences between fish of different origin,

experimental charr and Loch Rannoch charr (Figure 3A) and experimental and Loch Dughaill

charr (Figure 3B). CV2 however captures within group shape differences between fish raised

at ambient or elevated temperatures, benthic and pelagic ecotypes from Loch Rannoch (Figure

3A) and benthic and pelagic ecotypes Loch Dughaill (Figure 3B).

DISCUSSION

When studying contemporary climate warming effects, such as Arctic charr phenotypic

response, temporal scale matters. Temperature effects on aquatic organisms can manifest on

different temporal scales, from short term temporal fluctuations of temperature (e.g., hourly to inter-annual), which includes small oscillations to extreme events (e.g., large deviation of temperature from the typical mean), to long-term coarse-grained trend of variations [33, 42]. The temperature differential of 4 ℃ induced in this experiment can be compared to the scenario of an extreme year-to-year temperature variation that an Arctic charr cohort could experience, but also provide insights on the long-term temperature increase effect that Arctic charr populations will be subjected to in few decades [43]. Accordingly, in this study we demonstrate clear differences of an environmentally contingent expressed phenotype in a cold-water ectotherm under predicted climate change scenarios. Exposure to elevated water temperatures resulted in non-random, post-hatching morphological change in the head shape amongst Arctic charr that in the wild, display continuous variation (i.e., they express a unimodal phenotype). Contrary to our predictions, the observed plastic response to an elevated temperature regime within a single generation, comprised both a shift in the mean phenotype coupled with a reduction in the phenotypic variance (e.g., between-individual variation) in the expressed head shape. This observed effect on the phenotype was cumulative, with divergence between treatment groups increasing over time. However, the rate of divergence decreased with time, suggesting that the effect of temperature on development for phenotypic differences occurred predominant at early life stages. This indicates that the timing of exposure to different environmental conditions by individuals is important to the initiation of a plastic response and its magnitude.

Considering that temperature is one of the most widely reported environmental factors driving plastic responses in ectotherms in general, but fish in particular [44], Arctic charr develop at a faster rate at higher temperature [45]. Developmental pace in structurally important features, such as bone metabolism, are fundamentally linked to the form of morphological plasticity examined in this experiment [46, 47]. Elsewhere studies on plasticity

during development have shown that tissues can show responses of remarkable magnitude to changing environmental inputs, resulting in dramatic morphological changes among individuals [48, 49]. Our results demonstrated a strong and immediate influence of temperature on the direction, magnitude, and variation in the expression of phenotypic plasticity of a cold-water specialist species [29, 50]. Here Arctic charr offspring raised at elevated temperatures expressed a mean phenotype different to their siblings raised at an ambient temperature but also to that of their parents. The magnitude of this morphological response was mimicking the level of divergence observed in wild Arctic charr sympatric ecotypes. When compared to polymorphic populations that show distinct and stable foraging specialisms, individuals raised in the elevated temperature treatment expressed a phenotype analogous to a benthivorous ecotype rather than that of the parental pelagic form [40, 51]. The head of fish exposed elevated temperature was shorter, more robust, with a rounder snout; features that are typical of littoral macro-benthos feeding specialists (Figure A3) in several post-glacial fishes [52, 53]. In contrast, ambient temperature exposed fish expressed a phenotype elongated and pointed head, which is more typical of fish showing pelagic planktivory in the wild [54, 55].

Exposure to different environmental conditions is known to have asymmetrical impacts on the stability of developmental pathways during early life-stages [56-58]. Developmental stability is known to limit the phenotypic expression from a single genotype, thus it can modulate the magnitude and structure of phenotypic variation among individuals and cohorts [59, 60]. Several mechanisms, such as heterochrony, are known to drive the plastic expression of phenotypes through the effects of temperature exposure on developmental pathways [26, 61, 62]. In some cases, the expressed phenotype of an organism can be significantly altered by a single short-term environmental perturbation of the timing or rate during development; such events are thought to be a route to rapid phenotypic novelty [63]. Differences in temperature can induce heterochrony by disrupting the onset and termination of developmental processes

and the rate at which these processes occur. Consequently, temperature effects mediated

through heterochrony have the potential to result in skeletal development of different

craniofacial features in fishes [45, 64].

Early development represents a critical life-stage for phenotypic variation as

temperature can influence a variety of traits and the level of their variation in a population over

ontogeny [65-68]. The thermal history experienced by an individual during early development

is known to be a key factor defining adult phenotype [26, 61, 62]. Particularly during early

development, environmental signals can influence developmental processes through epigenetic

mechanisms, to influence the expressed phenotype [69]. Temperature exposure in early life is

known to result in epigenetic effects on traits such as growth, with consequences that can range

from short-term to transgenerational [70]. Our result suggested that temperature-induced

variation in developmental pathways resulted in divergent plastic allometric trajectories in the

early development of Arctic charr. Allometry, can been seen as a canalized process and an

interacting agent that can steer morphological variation [71, 72]. However, the opposite is also

true, whereas allometric trajectories can deviate from the "usual" shape-size relationship (e.g.,

as size increase, shape changes under specific allometric rules), which could serve as additional

measure of development instability [59]. Consequently, growth trajectories manifested as

allometric scaling provide the potential to express different phenotypes, which increase the

potential for novelty [73]. Altogether, both heterochrony and allometry are involved in

determining the degree of phenotypic variation generated during growth and morphological

development, and the expression of phenotypic variation observed in this study appears to be

influenced by temperature with concomitant developmental effects [59, 74].

The potential of Arctic charr to display phenotypic variation is considerable, whether

variation due to plasticity is continuous (e.g., unimodal) or discontinuous (e.g., polymodal)

[29]. Populations that express high level of variability in natural conditions show larger mean

and extended variance in plastic phenotypic expression in response to different temperatures [17, 75]. Although our study also showed a significant plastic response resulting in a shift in the mean phenotype expressed, there was also a decrease in variation of the phenotypes expressed among individuals raised at higher temperature treatment, which contrasts with previous work. Increased phenotypic variance in response to elevated temperatures has been suggested to be the reflection of a reduction in genotype precision (e.g., canalization and developmental stability) that will cause spreading in reaction norms [17, 45]. Our opposing result could be the result of a non-linear reaction norm of head shape on size. In this scenario a certain range of temperature exposure (either colder or warmer) has a different magnitude effect on phenotypic mean and variation [see description of how this may occur in Figure 1 in 26]. Another, not mutually exclusive possibility, is that despite both treatments being sampled after the same number of degree-days [a proxy for growth opportunity; 76] had elapsed, individual responses in the two temperature groups was partly the result of a size effect. In our experiment, growth opportunity and the actual realized body growth were uncoupled. Contrary to the predictions of Neuheimer and Taggart [76], the effect of time and temperature are thus not necessary equal in the promotion of body size and anatomical structures (e.g., heterochrony and allometry). Consequently, if variance is cumulative with the increase of size [77], this could partly explain the discrepancies between our predictions and findings because ambient temperature treatment individuals were generally larger by the end of the experiment.

Environmentally mediated variation in phenotypic expression is a fundamental attribute for many organisms, which can result in both short and long-term ecological evolutionary consequences for species that display plasticity. The disjunctive nature of the freshwater environment that landlocked Arctic charr occupy means that, in the absence of migratory routes for range adjustment, they will particularly need to adapt to changing novel conditions [15]. High levels of plasticity is one way in which they might do this. Yet, the role of phenotypic

plasticity with climate change is contentious, both positive and negative effects are possible.

To add to the complexity of climate change scenarios, positive and negative effects derived

from phenotypic plasticity can "act" on different scales, from year to year variations to coarse-

scale temporal trends. Organisms may have different plastic responses to changes in the

environment depending on the time and magnitude of scale of the exposure, which may have

different fitness consequences [16, 78]. Because we demonstrated that the expressed

phenotypic mean in Arctic charr can respond to an environmental cue rapidly and with a

magnitude that is highly likely to have functional consequences (i.e. temperature-mediated shift

from a pelagic towards a benthic phenotype), an important question arises when we are trying

to extrapolate our results into the wild. Would such plastic phenotypic change result in an

"overshoot" of the optimum phenotype or would it buffer against rapid fluctuations in the

environment?

In the context that even subtle differences in individual's morphology can influence

feeding efficiency and specialization [30], the potential of a fitness-buffering effect of plasticity

can easily fluctuate positively or negatively, depending if environmental cues and selective

filters become decoupled or not [15, 75]. Under climate change scenarios, decoupling between

phenotype and environment [79] might occur at different temporal scales. Short temporal

environmental oscillations, for instance extreme environmental events (e.g., a 4 ºC variation

scenario), will influence fine-grained phenotypic variation, at the cohort level [16]. If such

variation occurs over short-time periods, relative to the life-span of the organism, it can lead to

a temporary phenotype-environment mismatch, with resulting in fitness consequences for that

cohort specifically. If the year to year environmental fluctuations become less predictable with

an increase in the frequency of extreme events as is predicted by the climate scenarios, then

these temporarily phenotype-environment mismatches will become more frequent and have

greater impact on the population dynamics [75]. For longer-term temporal trends, if the overall

ecosystem doesn't change consistently in the same direction [e.g.,shift from benthic to pelagic

fuelled freshwater food webs; 80] as the expressed environmental canalized phenotype, the

phenotype-environment mismatches could lead to the emergence of an "evolutionary trap"

[i.e., the phenotype suddently become maladaptive; 81].

Alternatively, the effect of plasticity can provide the variation in expressed phenotype

upon which natural selection can act. Thus the scope for selection can provide the potential to

kick start evolutionary processes that could result in adaptive phenotypic change that would

lead to an "evolutionary rescue" [10, 82]. The probability of occurrence between these two

alternative effects of phenotypic plasticity on responses to temperature change is likely an

interaction between the capacity of the organism for phenotypic plasticity, the speed of mean

change in the environment (e.g., temperature) and the degree of short interval variation in the

environment.

The last component emerging from this experiment that can have evolutionary

consequences for a population in the face of changing selection regimes by affecting population

dynamic and probabilities of extinction is phenotypic variance [75, 83]. In this study, we

showed a decrease in variation in expressed phenotype at elevated temperature. The decrease

in the variation in the phenotypes expressed among individuals raised in the elevated

temperature treatment could reduce the phenotypic range for selection to act upon if this

scenario is applied in the wild. Accordingly, phenotypic variance can be a major component

that will lead a population to recovery from rapid (possibly transient) environmental change.

Consequently, a reduction or an increase of variance under climate change scenarios should

not be underestimated, as phenotypic variability has important consequences at the population

level.

CONCLUSION

The ecological and evolutionary consequence of rising temperature on Arctic charr will

depend on many factors [e.g., population specific lethal limit or growth optimum, interspecific

dynamics, and disease; 78]. However, our result clearly demonstrated the capacity of Arctic

charr for a rapid plastic response to temperature variation. The unimodal population shifted

their phenotypic mean within a generation, which had an impact on the phenotypic variability

through a major decline. Temperature change is not constant, rather it often varies in a

stochastic manner that introduces year-to year variation around a coarse-scale temporal trend

(e.g., contemporary increase). This can result in environmental variation that will correlate with

a cohort's phenotypic variation in a population, consolidating the importance of plasticity in

species resilience to rapid changes, as genetic changes are not expected to be capable of

responding over such short time scales [16].

Phenotypic plasticity can particularly aid in population persistence in new and changing

environments and where the pace of change is relatively rapid [48]. Contemporary temperature-

driven divergence in Nordic freshwater fishes have been reported [84]. Whether this flexible

response will be standard within and among cold-winter species is unknown. Nonetheless, the

fact remains that the majority of landlocked Arctic charr populations are currently

planktivorous [85]. By confirming the potential of a rapid and marked variation with an

increase temperature that swung from a pelagic toward a benthic phenotype, we provide

evidence that a substantial portion of Arctic charr populations worldwide (and most likely other

salmonid species) could be subjected to phenotype expression changes under climate change

scenarios.

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Table 1. Parameter estimates from the minimum adequate generalized linear mixed model describing the effect of temperature, exposure and centroid size on shape DF scores).

	Estimate	SE	X^2	P
Intercept (Ambient)	-6.84	1.91	-	-
Exposure time	0.002	0.002	-	-
Elevated temperature	2.36	0.21	39.81	<0.01
Centroid size	1.63	0.48	-	-
Exposure time : Centroid Size	-0.0012	0.0005	7.23	<0.01

Table 2 Results from the CVA comparing ambient and elevated temperature raised fish to wild benthic and pelagic ecotypes from Loch Rannoch and Loch Dughaill.

Comparison	Degree of difference	Location	Procrustes distance	P value
Pelagic - Ambient	Less	Rannoch	0.157	<0.01
Pelagic - Elevated	More	Rannoch	0.158	<0.01
				<0.01
Benthic - Ambient	More	Rannoch	0.111	<0.01
Benthic - Elevated	Less	Rannoch	0.095	<0.01
				<0.01
Pelagic - Ambient	Less	Dughaill	0.121	<0.01
Pelagic - Elevated	More	Dughaill	0.149	<0.01
				<0.01
Benthic - Ambient	More	Dughaill	0.152	<0.01
Benthic - Elevated	Less	Dughaill	0.129	<0.01

Figure 1. Hypothetical expressed phenotype frequency distributions shown as reaction norms resulting from an environmental change, such as temperature. Black lines represent expressed phenotypes in a population subjected to exisiting conditions and grey line represent population expression of phenotype subjected to the new environmental conditions. In panel a), the population doesn't express any change in phenotypic mean or in phenotypic variation when exposed to the new environmental conditions. In panel b), the population shows a shift in the expressed phenotypic mean but not in phenotypic variation when exposed to the new environmental regime. In panel c), population the phenotypic mean stays the same after the environmental condition changes, but the variation in expressed phenotypic increases in the population exposed to the new environmental regime. In panel d), the population (ecotype) subjected to the new environmental conditions shift both in phenotype mean and in phenotypic variation. Direction and magnitude of variation in panel b-d are not absolute, and will be species and location dependent as well of the strength and time of exposure to the environmental variables.

Figure 2. Individual Discriminant Function Scores (DF1) for head shape of Arctic charr of different sizes (represented by centroid size). Three sample periods are given, 700dd are represented by circles, 1000dd by triangles and 1400dd by squares. Ambient exposed fish are denoted with open symbols; elevated temperature fish are denoted with closed symbols. Polynomial regressions for both temperature treatments are showing allometric trajectories toward phenotypic divergence of head shape changes (see Figure A2).

Figure 3. Canonical Variate Analysis of 1400dd ambient and elevated temperature exposed
Arctic charr and sympatric polymorphic populations of Arctic charr from; A) Loch Rannoch,
B) Loch Dughaill. Elipses represent 95% confidence limits for means. Polymorphic charr
populations are represented by squares and experimental Arctic charr by circles. Open symbols
denote wild plankton feeding Arctic charr and Arctic charr raised at an ambient temperature.
Closed symbols denote benthic feeding Arctic charr and Arctic charr raised at an elevated
temperature. X and Y axis percentages denotes the amount of variation for each canonical
variate. Wire frames on CV1 are scaled at -8 and +8, wire frames for CV2 are scaled at -4 and
+4. C) and D) show mean (+/- 95% CL) for CV2 scores.

Appendix

Figure A1. Nine landmarks were used to characterise the shape of the head; Landmark (LM) 1, the tip of the nose; LM2, the most posterior part of the upper jaw; LM3, edge of cranium directly above the eye; LM4, edge of cranium at the central point between LM3 and LM1 at a 90 degree angle; LM5-8, most upper, posterior, lower and anterior parts of eye respectively; LM9, most posterior part of the gill operculum.

745 Figure A2. Possible pattern of allometric trajectories between temperature treatments of this
746 study, figure is modified from Simonsen, Siwertsson [72]. Allometric trajectory patterns can
747 be a) parallel, b) divergent, c) convergent, or d) common.

760
761   A

762
B

Figure A3. Individuals sampled at 1400dd, from the group a) raised at elevated temperature,
with a morphology analogous to benthic feeding wild ecotype and b) raised at ambient
temperature, with a morphology analogous to plankton feeding wild ecotype (parental
population).

Figure A4. CV1 and CV2 scores derived from Canonical Variate Analysis of: parent fish (grey
squares), ambient temperature exposed fish sampled at 1400dd (open circles) and elevated
temperature exposed fish sampled at 1400dd (closed circles) with 95% confidence elipses of
centroid. Percentage on X and Y axes denotes the amount of variation explained by each
canonical variate.