

NMD is required for timely cell fate transitions by fine-tuning gene expression and controlling translation

Elena Galimberti¹, Robert Sehlke^{2#}, Michelle Huth^{1#}, Marius Garmhausen², Merrit Romeike¹, Julia Ramesmayer¹, Sarah Stummer¹, Fabian Titz-Teixeira², Veronika Herzog³, Anastasia Chugunova⁴, Katrin Friederike Leesch⁴, Laurenz Holcik^{1,6}, Klara Weipoltshammer⁵, Laura Santini¹, Andreas Lackner¹, Arndt von Haeseler^{1,6,7}, Christa Bücken¹, Andrea Pauli⁴, Christian Schoefer⁵, Stefan L. Ameres³, Austin Smith⁸, Andreas Beyer², Martin Leeb^{1*}

Affiliations:

¹ Max Perutz Labs Vienna, University of Vienna, Vienna BioCenter, Dr.-Bohr-Gasse 9, 1030 Vienna, Austria

² CECAD Cologne, Joseph-Stelzmann-Str. 26 50931 Köln, Germany

³ Institute of Molecular Biotechnology, Vienna BioCenter, Dr.-Bohr-Gasse 3, 1030 Vienna, Austria

⁴ Research Institute of Molecular Pathology, Vienna BioCenter, Campus-Vienna-Biocenter 1, 1030 Vienna, Austria

⁵ Department for Cell and Developmental Biology, Medical University of Vienna, Schwarzspanierstrasse 17, 1090 Vienna, Austria

⁶ Center for Integrative Bioinformatics Vienna, Max Perutz Labs, University of Vienna and Medical University of Vienna, Dr.-Bohr-Gasse 9, 1030 Vienna, Austria

⁷ Bioinformatics and Computational Biology, Faculty of Computer Science, University of Vienna, Vienna, Austria

⁸ Wellcome - MRC Cambridge Stem Cell Institute, University of Cambridge, Cambridge CB2 0AW, UK

*corresponding author

equal contribution

ABSTRACT

Cell fate transitions depend on balanced rewiring of transcription and translation programmes to mediate ordered developmental progression. Components of the nonsense-mediated mRNA decay (NMD) pathway have been implicated in regulating embryonic stem cell (ESC) differentiation, but the exact mechanism is unclear. Here we show that NMD controls the translation initiation factor *Eif4a2* and its premature termination codon encoding isoform (*Eif4a2^{PTC}*). NMD deficiency leads to translation of a specific truncated Eif4a2 protein, which elicits increased translation rates and causes significant delays in mouse ESC differentiation. Thereby a previously unknown feedback loop between NMD and translation initiation is established. Our results illustrate a clear hierarchy between KOs in severity of target deregulation and differentiation phenotype (*Smg5* > *Smg6* > *Smg7*), which highlights heterodimer-independent functions for Smg5 and Smg7. Together, our findings expose an intricate link between mRNA stability and translation initiation control that must be maintained for normal dynamics of cell state transitions.

1 INTRODUCTION

2 Mouse embryonic stem cells (ESCs) capture the developmentally transient naïve pluripotent state *in vitro*.
3 ESC self-renewal is maintained by an interactive transcription factor (TF)-network, which has been
4 extensively characterized^{1,2}. This network is decommissioned between embryonic day (E)4.5 and E5.5 in
5 mouse development³. Similar kinetics are maintained in ESCs, which irreversibly commit to differentiation
6 24-36 h after 2i withdrawal⁴⁻⁶. Rex1 is a known marker of naïve pluripotency and high-resolution
7 dissection of the exit from naïve pluripotency is facilitated by the availability of Rex1 promoter-driven
8 destabilized GFP reporter ESC lines (Rex1::GFPd2)^{7,8}. Rex1-GFP downregulation is initiated within 24 h
9 after 2i withdrawal (N24) and completed after 48 h (N48). Extinction of Rex1-GFPd2 expression coincides
10 with functional commitment to differentiation.

11 Several genome-wide screens have uncovered drivers of the exit from pluripotency⁹, many of which are
12 involved in transcriptional regulation and epigenetic modification. These screens also identified a large
13 cohort of post-transcriptional regulators. RNA modifiers, such as m6A^{10,11}; negative regulators of mRNA
14 stability, such as Pum1¹²; and components of the nonsense-mediated mRNA decay (NMD) pathway¹²⁻¹⁴
15 have been implicated in regulating ESC differentiation. Nonetheless, how post-transcriptional regulatory
16 mechanisms contribute to cell fate changes remains poorly understood.

17 NMD is a translation coupled mechanism that promotes degradation of mRNAs containing a premature
18 termination codon (PTC)¹⁵. However, PTC-independent NMD activity has also been shown¹⁶⁻¹⁸. NMD is
19 triggered by phosphorylation of the RNA-helicase Upf1, which is essential for NMD. Degradation occurs
20 either by Smg6-mediated endonucleolytic cleavage, or by exonucleolytic cleavage, mediated by a Smg5-
21 Smg7 heterodimer¹⁵. Transcriptome-wide analysis demonstrated that Smg6 and Smg5-Smg7 have highly
22 overlapping mRNA targets¹⁹. There is also evidence that Smg factors regulate telomere maintenance²⁰⁻
23 ²². Although NMD components constitute some of the top hits in genome-wide screens studying exit from

24 naïve pluripotency, neither the contribution of individual NMD effector proteins, nor the key mRNA
25 targets of NMD that lead to delayed cell fate transition are known.

26 Here we identify a role for NMD in ensuring normal differentiation kinetics by facilitating establishment
27 of proper cell fate specific gene expression programmes and by controlling expression of Eif4a2, a key
28 translation initiation factor. We identify the resulting Eif4a2-dependent increased translational activity in
29 NMD-deficient ESCs as the main reason for their inability to properly commit to differentiation.

30

31 **RESULTS**

32 **Variable degree of defects in exit from naïve pluripotency in NMD-deficient ESCs**

33 To delineate the molecular function of NMD in the exit from naïve pluripotency, we generated Rex1-
34 GFPd2 reporter ESC lines^{4, 23} deficient for the three NMD downstream effectors *Smg5*, *Smg6* or *Smg7*
35 (NMD KO ESCs), and corresponding rescue cell lines in which the missing NMD-factor was re-expressed
36 (NMD rescue ESCs) (Extended Data Fig. 1a,b). NMD KO cells showed normal ESC morphology, cell cycle
37 profile and telomere maintenance (Extended Data Fig. 1c,d), but exhibited pronounced delays in the exit
38 from naïve pluripotency. This manifested in delayed downregulation of the Rex1-GFPd2 reporter 24 h
39 after the onset of differentiation (N24) and delayed entry into commitment, assayed 72 h after 2i
40 withdrawal (Fig. 1a,b). Both defects were restored in rescue cell lines, showing causality of NMD-factor
41 deficiency for delays in exit from naïve pluripotency (Fig 1a,b). The three NMD-factor KOs displayed
42 variable degrees of differentiation delays: the strongest effect was observed in the absence of *Smg5* and
43 the weakest in the absence of *Smg7*. This does not align with the proposed strictly heterodimer-
44 dependent function of *Smg5* and *Smg7*, which would predict similar phenotypes for *Smg5* and *Smg7* KOs
45²⁴⁻²⁶. The results therefore suggest a less strict heterodimer dependence than anticipated for the activity
46 of either *Smg5* or *Smg7*.

47 To monitor the effect of NMD deficiency on long-term differentiation potential, we performed 3D
48 aggregate embryoid body (EB) differentiation during a 10-day time-course. As expected, downregulation
49 of naïve and primed pluripotency markers was severely impaired in NMD KO EBs. Although NMD KO
50 aggregates upregulated the formative marker genes *Fgf5* and *Otx2* with similar kinetics to WT, the
51 subsequent shutdown of the formative program, observed in WT cells between day 4 (d4) and d6, was
52 impaired (Fig. 1c). Furthermore, expression of the endo-mesoderm defining TF *Brachyury* (*T*) was not
53 properly downregulated after a peak of expression at d4 to d6, suggesting a general function of NMD in
54 shaping transcriptomes during early cell fate decisions (Fig. 1c). Further supporting a role for NMD in
55 regulating differentiation, teratomas derived from *Smg6* KO ESCs showed a lower degree of
56 differentiation than WT and rescue controls (Extended Data Fig. 1e). Together, these data show a global
57 differentiation defect of Smg factor-deficient cells, with a variable degree of phenotypic strengths (*Smg5*
58 > *Smg6* > *Smg7*).

59

60 **Cooperativity between NMD factors regulates NMD and exit from naïve pluripotency**

61 To investigate potential cooperativity between NMD effectors, we performed siRNA-mediated
62 knockdowns of *Smg6* and *Smg7* in *Smg5* and *Smg6* KO ESCs (Fig. 1d-h and Extended Data Fig. 1f). Thereby,
63 we generated cells double- and triple-depleted for NMD downstream effectors. WT cells showed a near
64 complete loss of Rex1-GFPd2 expression at N48 (Fig. 1d). siRNA-mediated depletion of *Smg6* in *Smg5* KO
65 cells resulted in only a minor, non-synergistic increase in the differentiation delay assessed in Rex1-GFPd2
66 reporter assays (Fig. 1e), despite the strong defects observed in both single KOs. By contrast, the depletion
67 of *Smg7* in a *Smg6* KO background yielded a strong synergistic differentiation phenotype (Fig. 1f).
68 Similarly, co-depletion of both heterodimer members, *Smg5* and *Smg7*, exhibited a clear synergistic effect
69 (Fig. 1g), further highlighting heterodimer-independent functions for *Smg5* and *Smg7*. Combined

70 depletion of all the NMD effectors by double knockdown of *Smg6* and *Smg7* in *Smg5* KO ESCs resulted in
71 differentiation delays on par with *Smg6/7* or *Smg5/7* double depletion (Fig. 1h). To assess the impact of
72 NMD effector depletion and co-depletion on NMD-specific mRNA target gene expression, we assessed
73 levels of the known NMD target gene *Gadd45b* by RT-qPCR¹⁶. Consistent with observed differentiation
74 delays, we observed strong genetic interactions between *Smg6* and *Smg7* as well as between *Smg5* and
75 *Smg7*, and weaker interactions between *Smg5* and *Smg6* regarding *Gadd45b* transcript abundance
76 (Extended Data Fig. 1g). Together, this suggests that differentiation delays scale with the extent of NMD
77 impairment.

78 To further study the cooperativity of NMD factors we sought to generate stable NMD double KO (dKO)
79 ESCs. However, attempts using CRISPR/Cas9 to generate all possible dKO ESC lines yielded only
80 *Smg5/Smg6* (*Smg5/6*) dKO ESCs. Neither *Smg5/Smg7* nor *Smg6/Smg7* dKOs could be established, despite
81 multiple attempts using efficient gRNAs. This suggests an essential role of *Smg7* in ESCs in the absence of
82 its heterodimerization partner *Smg5* or in the absence of *Smg6*. *Smg5/6* dKO cells showed a deficiency in
83 downregulating naïve TF mRNAs similar to that observed in *Smg5* single KO cells, indicating a dominant
84 role of *Smg5* in regulating differentiation-relevant RNA-homeostasis programmes (Fig. 1i).

85 Taken together, *Smg5*, *Smg6* and *Smg7* KO ESCs exhibit variable degrees of differentiation defects and
86 NMD downstream factors act synergistically during the exit from pluripotency. *Smg7* possesses a central,
87 facultatively essential function: It can be depleted without resulting in strong effects on early
88 differentiation, but an essential role is revealed in the absence of either *Smg5* or *Smg6*.

89

90 ***Smg7* is necessary and sufficient for pUpf1 binding, independently of *Smg5***

91 Continuous NMD activity relies on cyclic phosphorylation and dephosphorylation of Upf1. We observed
92 increased pUpf1 levels in all NMD KO cells (Fig. 2a), in line with previous reports²⁵. Notably, pUpf1 levels

93 followed the same trend as the differentiation phenotype and deregulation of the NMD target *Gadd45b*
94 (pUpf1 levels in Smg5 KO > Smg6 KO > Smg7 KO > WT ESCs). Upf1 is phosphorylated by the kinase Smg1
95 ^{27, 28}. *Smg1* mRNA is itself an NMD target, but in contrast to pUpf1 is upregulated to a similar extent in all
96 three NMD KO ESCs (Extended Data Fig. 2a). Therefore, graded increase of pUpf1 levels in NMD KO is
97 unlikely to be a direct effect of higher *Smg1* expression. Impaired Upf1 dephosphorylation may therefore
98 underlie increased pUpf1 levels in NMD mutant cells. The three NMD effectors Smg5, Smg6 and Smg7
99 have all been implicated in the recruitment of PP2A to dephosphorylate Upf1 ^{25, 26}, but the strongest
100 increase of pUpf1 in Smg5 KO ESCs suggests that Smg5 acts as the main PP2A recruiter during NMD
101 (Extended Data Fig. 2b).

102 To identify the molecular basis for an apparent heterodimer-independent function of *Smg5* and *Smg7* and
103 for the observed genetic interactions between Smg factors, we performed co-immunoprecipitation (co-
104 IP) experiments. We precipitated Smg5 or Smg7 in WT and NMD KO ESCs, in the presence and absence of
105 RNA. Smg5 and pUpf1, but not Smg6, co-immunoprecipitated with Smg7 in an RNA-independent manner
106 in WT ESCs (Fig. 2b). Interestingly, Smg7 efficiently bound pUpf1 in the absence of its heterodimerization
107 partner Smg5 (Fig. 2b). The Smg7-pUpf1 interaction was also maintained in *Smg5/6* dKO ESCs, suggesting
108 that Smg7 can interact with pUpf1 independently of other downstream NMD effectors. Conversely,
109 however, Smg5 failed to bind pUpf1 in the absence of *Smg7* (Fig. 2c), indicating that Smg7 is the critical
110 docking site for pUpf1 during NMD and that pUpf1 binding to Smg7 is independent of Smg5. When we
111 performed the same co-IP experiments in the presence of RNA, we observed an interaction between Smg5
112 or Smg7 and Smg6 in the absence of the respective heterodimerization partner (Extended Data Fig. 2c,d).
113 This RNA-dependent interaction was not observed in WT (Fig. 2b,c).

114 Taken together, biochemical analysis and viability of *Smg5/6* dKO ESCs indicate that Smg7 can bind to
115 pUpf1 in the absence of Smg5 to sustain limited NMD function. Smg5, however, is unable to interact with
116 pUpf1 in the absence of Smg7 (Fig. 2c and Extended Data Fig. 2d). The strong differentiation phenotype

117 of *Smg5*-deficient cells can be explained by efficient binding of pUpf1 by Smg7 alone and subsequent
118 inefficient dephosphorylation and target degradation, resulting in a ‘poisoning’ of both branches of NMD
119 (Fig. 2d). By contrast, deficiency in Smg7 results in failure to recruit components of the exonucleolytic
120 branch, but allows potential recognition and degradation by Smg6, resulting in only a mild NMD target
121 deregulation and differentiation phenotype.

122

123 **Integrating transcriptome-wide gene expression with mRNA half-life analyses identifies relevant NMD** 124 **targets during the exit from naïve pluripotency**

125 To uncover the molecular mode of action by which NMD regulates the exit from naïve pluripotency, we
126 applied a combinatorial approach based on the following logic: an NMD target transcript relevant in the
127 context of differentiation must show elevated expression levels, following the same graded pattern as the
128 differentiation phenotype (*Smg5* > *Smg6* > *Smg7*) and a concomitant increased mRNA half-life.

129 To this end, we first performed RNA-seq of *Smg5* KO, *Smg6* KO, *Smg7* KO and WT cells in 2i and after 24
130 h of differentiation (N24) to assess the impact of NMD factor depletion on global gene expression (Fig. 3a-
131 d and Extended Data Table 1). In 2i, we identified 881 transcripts deregulated in NMD KO ESCs compared
132 to WT (adj. $p \leq 0.01$), 516 of which were significantly upregulated and 252 significantly downregulated in
133 all three NMD KO ESCs. This shows a strong overlap in deregulated genes between all three KO cell lines;
134 only 113 transcripts did not show the same directionality in transcriptional changes between all NMD KO
135 cells (Fig. 3a). Overall, upregulated transcripts (clusters 2 and 4), showed a graded increase in deregulation
136 (*Smg5* KO > *Smg6* KO > *Smg7* KO), similar to the KO phenotypes (Fig. 3b). 256 out of 564 transcripts
137 belonging to clusters 2 and 4 were significantly upregulated in all three NMD KO cell lines and showed a
138 graded response to Smg-factor depletion, suggesting that genes found within this group are responsible
139 for the observed differentiation delays.

140 GO analysis of the 256 graded upregulated transcripts in 2i revealed an enrichment for methyltransferase
141 and helicase activity. The remaining upregulated transcripts in 2i showed enrichment for telomere
142 maintenance and helicase activity. Within downregulated genes we detected an enrichment for
143 transcripts involved in neural development, somite development and other differentiation related
144 processes (Extended Data Fig. 3a and Extended Data Table 2). This suggests that leaky expression of the
145 differentiation programme in 2i is dampened in NMD-deficient cells.

146 At N24, we identified 1,174 deregulated transcripts in NMD KO cells compared to WT cells (adj. $p \leq 0.01$).
147 Among these, 727 were upregulated in all NMD KO cells and 336 were downregulated in all NMD KO cells
148 (Fig. 3c). Out of the 808 upregulated transcripts belonging to clusters 2 and 3, 434 transcripts were
149 upregulated following the graded pattern (*Smg5* KO > *Smg6* KO > *Smg7* KO) (Fig. 3d). GO analysis of these
150 434 transcripts revealed enrichment for pluripotency-related terms, such as LIF response and stem cell
151 maintenance (Extended Data Fig. 3b and Extended Data Table 2). This was not evident for the 293
152 transcripts not showing graded upregulation. Within downregulated transcripts, we detected an
153 enrichment of differentiation-related terms, such as neural tube development and pattern specification
154 (Extended Data Fig. 3b and Extended Data Table 2). This suggested that genes showing graded expression
155 are involved in and reflect the different levels of differentiation delays seen in NMD KO cells.

156 Previous reports showed increased c-Myc levels in NMD-deficient ESCs and proposed a causative
157 involvement of c-Myc in NMD KO induced differentiation delays¹³. However, we did not detect increased
158 c-Myc transcript levels by RNA-seq or protein levels on western blots in any of the three NMD KO ESCs
159 (Extended Data Fig. 3c,d). On the contrary, we reproducibly detected reduced levels of c-Myc upon NMD
160 disruption. Furthermore, genetic depletion of *c-Myc* in NMD KO ESCs did not rescue the differentiation
161 delays (Extended Data Fig. 3e,f) indicating that c-Myc is not relevant for the differentiation defects
162 observed in NMD KO ESCs.

163 Elevated transcript levels of direct NMD targets are expected to result from increased half-lives, based on
164 impairment of mRNA degradation. Therefore, we used SLAM-seq to measure transcriptome-wide half-
165 lives²⁹. We detected a global increase of transcript half-lives in *Smg5* and *Smg6* KO ESCs compared to WT
166 ESCs (Fig. 3e). We were able to calculate half-lives for 4,342 transcripts; 3,062 of which exhibited a longer
167 half-life in NMD KO cells (Fig. 3e). On average, mRNA half-life was significantly longer in NMD KOs than in
168 WT ($p < 10^{-4}$), increasing from 2.1 h in WT to 2.7 and 3 h in *Smg5* KO and *Smg6* KO, respectively (Fig. 3e).
169 As a cohort, pluripotency TF-encoding mRNAs exhibited a significant increase in half-lives (Fig. 3f) similar
170 to the overall increase of half-lives across the entire transcriptome. Both identity of transcripts and
171 amplitude of half-life changes showed a strong overlap between *Smg5* and *Smg6* KO ESCs (Extended Data
172 Fig. 3g). Transcriptome-wide comparisons showed that the vast majority of transcriptionally upregulated
173 genes showed a concomitant increase in half-lives (Fig. 3g and Extended Data Fig. 3h).

174 To identify bona-fide NMD targets with relevance for the differentiation delay phenotype, we investigated
175 a group of 250 genes that were significantly upregulated in all three *Smg* KO ESCs in 2i for which we could
176 also calculate half-lives. Of these, 136 showed an increase in half-lives in both *Smg5* and *Smg6* KOs, and
177 57 showed an expression pattern in accord with the degree of phenotype-strength (Fig. 3h, Extended Data
178 Fig. 3i and Extended Data Table 3). These 57 genes were involved in various cellular processes, such as
179 tRNA modification, TOR-signaling and calcium transport (Extended Data Table 3). None of the naïve
180 pluripotency factors was part of this group, suggesting that direct regulation of naïve pluripotency factors
181 by NMD is not the primary cause for the observed differentiation delays in NMD KO ESCs. Taken together,
182 we identified a cohort of 57 direct NMD target genes, with an expression pattern suggestive of an
183 involvement in controlling the exit from naïve pluripotency.

184

185

186 ***Eif4a2 is a bona-fide NMD target in ESCs and its NMD sensitivity is evolutionarily conserved***

187 Among the 57 NMD targets with potential phenotypic relevance, our attention was drawn to the RNA
188 helicase *Eif4a2*, a component of the Eif4F translation initiation complex, which mediates 5' cap recognition
189 by the small ribosomal subunit^{30, 31}. Several layers of evidence suggest *Eif4a2* as a bona-fide NMD target.
190 Firstly, the *Eif4a2* locus encodes for two splice-isoforms: one full-length protein, and one PTC-containing
191 isoform (Fig. 4a). Secondly, *Eif4a2* mRNA levels were significantly increased after translation inhibition by
192 cycloheximide (CHX) treatment, a hallmark of bona-fide NMD targets (Fig. 4b). CHX response was similar
193 to that observed for *Gadd45b*. The PTC-isoform (*Eif4a2^{PTC}*) showed an even more pronounced sensitivity
194 to CHX treatment. Thirdly, *Eif4a2^{PTC}* transcript levels were upregulated in a graded fashion (*Smg5* KO >
195 *Smg6* KO > *Smg7* KO) throughout a long-term EB differentiation time-course, consistent with phenotypic
196 relevance (Fig. 4c). Full length *Eif4a2* (*Eif4a2^{FL}*) protein levels were increased in NMD KO cells (Fig. 4d). The
197 *Eif4a2^{PTC}* isoform produced a protein that was weakly detected by western blot analysis in WT and NMD
198 rescue cells and strongly increased in all three NMD KO ESCs, both in 2i and at N24 (Fig. 4d and Extended
199 Data Fig. 4a). This indicates that while expression levels are tightly regulated by NMD, the *Eif4a2^{PTC}* protein
200 shows low levels of expression in WT cells. The upregulation of *Eif4a2* was unique among Eif4F complex
201 members, since the others, including the close homologue *Eif4a1*, showed neither increased expression
202 nor significantly longer half-lives in NMD KOs (Fig. 4d, Extended Data Fig. 4b,c and Table S3), nor did they
203 react to CHX treatment (Extended Data Fig. 4d). Taken together, this identifies *Eif4a2* as a bona-fide NMD
204 target in ESCs and shows that NMD disruption leads to the production of an *Eif4a2^{PTC}* protein.

205 The linkage of NMD to *Eif4a2* expression appears to be conserved in evolution. The gene structure of
206 *Eif4a2* is strikingly similar between mouse, dog and human (Fig. 4e). The mouse *Eif4a2* coding sequence
207 showed identities of 94%, 95% and 98.5% to human, dog and rat, respectively (Fig. 4f). Despite the PTC-
208 exon being under no apparent selective pressure to maintain protein-coding potential (PTC at nucleotide
209 position 8 out of 107 in this exon), we observed 96.4%, 98.2% and 99% nucleotide identity for human, dog

210 and rat, respectively, on par with the other well conserved *Eif4a2* exons. Conservation of UTRs and introns
211 was significantly lower (Fig. 4f). In a transcriptome-wide comparison, the *Eif4a2* PTC exon showed high
212 conservation at the nucleotide level: less than 5% of all exons and less than 3% of all UTRs showed higher
213 conservation rates between mouse and rat than the *Eif4a2*^{PTC} exon (Fig. 4g and Extended Data Fig. 4e-g).
214 In the comparisons with human and dog these percentages were even lower. Therefore, the potential of
215 *Eif4a2* to be regulated by NMD is a feature conserved in mammalian evolution.

216

217 ***Eif4a2* is causative for defects in exit from naïve pluripotency in NMD-deficient ESCs**

218 To delineate a potential causative relationship between increased *Eif4a2* levels and the differentiation
219 defect observed in NMD-deficient cells, we generated *Eif4a2* KO cells and NMD (*Smg5* or *Smg6*) / *Eif4a2*
220 double deficient cells (collectively referred to as NMD/*Eif4a2* dKO) by deleting all potential *Eif4a2* isoforms
221 or specifically *Eif4a2*^{PTC} (Fig. 5a and Extended Data Fig. 5a). All *Eif4a2* KO cell lines proliferated normally,
222 showed normal ESC morphology and could be maintained for more than 10 passages. Deletion of the PTC
223 exon resulted in an increase in full length *Eif4a2* protein levels in WT and NMD KO ESCs (Fig. 5a). Both the
224 complete absence of *Eif4a2* or specific deletion of *Eif4a2*^{PTC} accelerated Rex1-GFPd2 downregulation
225 kinetics in WT cells (Fig. 5b), suggesting that increased expression of *Eif4a2*^{PTC} and not full-length *Eif4a2* is
226 the major cause of the differentiation block. In NMD/*Eif4a2*^{FL} and NMD/*Eif4a2*^{PTC} dKO cells, differentiation
227 kinetics were substantially rescued in a Rex1-GFPd2 assay (Fig. 5b). Accordingly, *Klf4* and *Nanog*
228 expression levels were downregulated to near WT-levels at N30 (Fig. 5c). Similar effects were observed
229 for protein levels of the naïve TFs *Nanog* and *Esrrb* at N24 (Extended Data Fig. 5b). Phenotypic rescue in
230 NMD/*Eif4a2*^{PTC} double KO cells, in which *Eif4a2*^{FL} protein levels are even further increased, supports the
231 proposition that *Eif4a2*^{PTC} is the major mediator of the observed defect in exit from naïve pluripotency.

232 To assess the specificity of *Eif4a2* depletion in restoring differentiation potential in an NMD-deficient ESC
233 background, we compared differentiation potential between *Tcf7l1* KO and *Eif4a2/Tcf7l1* dKO ESCs. *Tcf7l1*
234 encodes for a HMG box transcription factor and is one of the strongest drivers of the exit from naïve
235 pluripotency. *Tcf7l1* KO ESCs showed defects in exit from naïve pluripotency on par with NMD KOs, but
236 exhibited an NMD-independent, Gsk3 inhibition-like phenotype (Extended Data Fig. 5c) ⁴. *Tcf7l1* KO cells
237 were insensitive to co-depletion of *Eif4a2* (Extended Data Fig. 5d), suggesting that *Eif4a2* is a specific
238 genetic interactor of NMD in regulating differentiation.

239 To test whether *Eif4a2* upregulation is sufficient to cause a differentiation delay, we overexpressed flag-
240 tagged *Eif4a2^{FL}* or *Eif4a2^{PTC}* in WT ESCs (Extended Data Fig. 5e). At N24, *Eif4a2^{PTC}* overexpressing cells
241 showed increased transcript levels for *Klf4*, *Esrrb*, *Tfcp2l1* and *Nanog* and increased protein levels for
242 *Nanog* at N24 (Fig. 5d,e). Cells overexpressing *Eif4a2^{FL}* exhibited a weaker effect and showed mild
243 upregulation only of *Nanog*. This accords with increase of *Eif4a2^{PTC}* rather than *Eif4a2^{FL}* having a major
244 impact on the exit from naïve pluripotency. Notably, the differentiation delays elicited by increasing
245 expression levels of *Eif4a2* isoforms did not reach the intensity observed in NMD KO ESCs, suggesting that
246 factors in addition to *Eif4a2* contribute to the failure to properly shut down naïve pluripotency in NMD
247 KO ESCs. However, double depletion experiments show that deregulation of *Eif4a2* is the major cause for
248 exit from naïve pluripotency defects.

249 To test whether upregulation of *Eif4a2^{PTC}* mRNA or *Eif4a2^{PTC}* protein was responsible for the delay in
250 differentiation of NMD KO ESCs, we introduced a frameshift mutation in the first exon of *Eif4a2^{PTC}*
251 (*Eif4a2^{PTC-mut}*) (Extended Data Fig. 5f). This mutation does not allow the generation of a functional *Eif4a2*
252 protein, abrogating in-frame coding potential. We overexpressed either *Eif4a2^{PTC}* or *Eif4a2^{PTC-mut}* in WT
253 ESCs (Extended Data Fig. 5g). At N16, cells overexpressing *Eif4a2^{PTC}* showed upregulation of *Nanog* and
254 *Klf4* mRNA levels, while cells overexpressing *Eif4a2^{PTC-mut}* did not (Fig. 5f). This suggests that upregulation
255 of the truncated protein encoded by *Eif4a2^{PTC}* is responsible for the differentiation delay observed in NMD

256 KO cells. In summary, these data show that Eif4a2 is the NMD target causative for the observed
257 differentiation delays in Smg-factor deficient ES cells.

258

259 **Eif4a2-mediated differentiation delay is caused by PTC isoform-dependent regulation of translation**

260 Since Eif4a2 functions as a translation initiation factor, we tested whether its upregulation in NMD KO
261 cells resulted in increased translation rates. To this end we performed radioactive S³⁵ and O-propargyl-
262 puromycin (OPP) based translation assays and detected an increase in translation rates in NMD-deficient
263 cells in 2i and at N12 (Fig. 6a-c and Extended Data Fig. 6a). The increase in translation rates observed in
264 NMD KO ESCs was similar in magnitude to that observed for *Tsc2* KOs, in which deregulated mTORC1
265 activity leads to an increase in translation³². Full Eif4a2 KOs, but also specific depletion of the PTC isoform
266 resulted in mild reductions of translation rates. Eif4a2^{FL} and Eif4a2^{PTC} depletion in NMD KO cells reduced
267 translation rates to WT levels, showing that increased translation rates in NMD KO cells are dependent on
268 the PTC isoform of Eif4a2 (Fig. 6b,c).

269 To identify transcripts whose translation was potentially most affected by Eif4a2, we performed RIP-Seq
270³³, using *Eif4a2* KO ESCs engineered to only express either flag-tagged Eif4a2^{PTC} or Eif4a2^{FL} (Extended Data
271 Fig. 6b). As expected for a translation factor, noncoding transcripts like *Meg3* were depleted in flag-
272 Eif4a2^{FL} RIP samples, indicating the specificity of the assay. Out of 9,812 expressed transcripts (CPM > 1 in
273 all input samples), we detected 362 transcripts significantly bound by Eif4a2 (Extended Data Table 4).
274 Several naïve pluripotency (*Esrrb*, *Tbx3*, and *Klf4*) and exit-factor (*Jarid2*, *Alg13* and *Fbxw7*)⁶ encoding
275 transcripts were enriched for Eif4a2 binding, as confirmed by RIP-qPCR (Fig. 6d and Extended Data Fig 6c).
276 RIP-seq of flag-Eif4a2^{PTC} resulted in only weak enrichments, possibly owing to the unstable nature of the
277 Eif4a2^{PTC} protein. RIP-RT-qPCR could not detect binding of Eif4a2^{PTC} to Eif4a2^{FL} targets identified above
278 (Extended Data Fig. 6d).

279 To understand the interaction landscape of both *Eif4a2* isoforms, we used KO cells expressing either flag-
280 tagged Eif4a2^{PTC} or Eif4a2^{FL} (Extended Data Fig. 6b). We also over-expressed Eif4a2^{PTC} in WT ESCs to
281 identify potential Eif4a2^{FL}-dependent interactions of Eif4a2^{PTC}. We then performed co-IP followed by
282 mass-spectrometry to identify interaction partners of the two different isoforms. To compensate for the
283 highly unstable nature of Eif4a2^{PTC}, we performed sample preparation after short-time treatment with the
284 irreversible proteasome inhibitor epoxomicin³⁴. Thereby, we identified 254 and 306 proteins specifically
285 bound by Eif4a2^{FL} or Eif4a2^{PTC} over flag-only control IPs, respectively (Extended Data Table 5 and Fig. 6e).
286 Among these, 75 proteins precipitated with both isoforms. Twenty-one of these showed near equal
287 interactions between the Eif4a2 isoforms, including the NMD factor Smg7 and the translation initiation
288 factor transporter Eif4enif1 (Extended Data Table 5), suggesting some preservation of normal interaction
289 profiles of the PTC-specific isoform. Of the remainder, 47 showed stronger binding by Eif4a2^{FL}. Among
290 these were several negative regulators of mRNA stability and regulation (e.g., Ago2, Igfbp1 and Igfbp3,
291 Pum1, Trim71, Cnot). We further detected physical association of Eif4a2^{FL} with several NMD components,
292 such as Upf1 and Smg6, consistent with previous reports³⁵. Together, this indicates an intricate link
293 between Eif4a2 and the mRNA destabilization machinery. We detected seven factors that showed
294 stronger co-precipitation with the PTC-isoform. These include the pluripotency TF Nr0b1 (Dax1)⁸, the
295 pluripotency regulator Ogt³⁶ and the GATOR complex member and mTOR regulator Wdr59 (Fig. 6f).
296 Among the 178 proteins specifically bound to Eif4a2^{FL} and not to the PTC isoform were various translation
297 initiation factors including all Eif4G isoforms and several components of the Eif3 complex (Fig. 6g and
298 Extended Data Table 5). This highlights the integration of Eif4a^{FL} (but not Eif4a2^{PTC}) into a functional
299 translation initiation complex. Eif4a1 was not detected, indicating a mutually exclusive presence in the
300 complex. Notably, Sox2 interacted with Eif4a2^{FL} (Fig. 6g), indicating a potentially functional interaction of
301 core pluripotency regulators and the translation initiation machinery.

302 Among the 231 proteins specifically associated with the PTC isoform and not the full-length protein, we
303 detected the Fgf/ERK pathway component Raf1, the mTORC1 regulator Tsc2 and the GATOR complex
304 member Mios (Fig. 6h and Extended Data Table 5). Increased mTORC1 activity in NMD is evident by
305 increased p-p70-S6K levels in NMD KO cells (Extended Data Fig. 6e). We further detected an interaction
306 of Eif4a2^{PTC} with the key pluripotency factor Oct4 (*Pou5f1*).

307 Taken together, these results indicate that Eif4a2^{FL} is mainly involved in translation initiation, but shows
308 significant interactions with a set of mRNA destabilizing proteins, including NMD-factors, CCR4-CNOT
309 components and the RISC member Ago2. The unstable Eif4a2^{PTC} protein showed little association with
310 translation initiation complex members, but still binds mRNA binding proteins, including the NMD factors
311 Smg6 and Smg7.

312 In summary, we show that NMD controls transcript levels of hundreds of genes in ESCs and during early
313 differentiation. This results in delayed exit from naïve pluripotency and a failure to shut down cell fate
314 programmes in later cell fate decisions. Deregulation of a feedback circuit between NMD and translation
315 initiation in NMD mutant cells, encoded in a PTC-containing isoform of Eif4a2, results in elevated
316 translation rates, which are the major cause of NMD-associated differentiation defects in ESCs.

317

318 **DISCUSSION**

319 Here we report a function of NMD in the dynamic regulation of a mammalian cell state transition, the exit
320 from naïve pluripotency. Our results highlight a role for NMD far beyond purging PTC containing
321 transcripts. We here show a major role in modulating gene expression profiles and also translation,
322 suggesting NMD as important component of the cellular machinery maintaining cell identity and proper
323 differentiation trajectories.

324 Without NMD function, increased levels of Eif4a2 and Eif4a2^{PTC} disrupt normal differentiation kinetics and
325 lead to an increase in translation levels, including those of naïve pluripotency TFs, which impedes

326 developmental transition. Basal leaky expression of differentiation-associated transcripts was reduced
327 already in 2i. Consistently, NMD depletion leads to increased similarities between ESCs and the naïve *in*
328 *vivo* epiblast⁶. Therefore, NMD acts already in self-renewal conditions to prune the differentiation
329 programme. Despite exhibiting delayed downregulation of the pluripotency TF-network, NMD KO EBs
330 eventually initiate formative and primed expression programmes. However, subsequent downregulation
331 of key markers of these states, normally observed around d4 to d6 of EB differentiation, does not occur.
332 This suggests a general role for NMD in dynamically shaping and fine-tuning transcript abundance to
333 facilitate rapid gene regulatory network remodeling during cell fate decisions.

334 Differentiation defects across *Smg5*, *Smg6* and *Smg7* KOs scale with deregulation of known NMD target
335 genes and levels of phospho-Upf1, suggesting that NMD defects directly translate to differentiation
336 delays. The striking difference in NMD target regulation and extent of differentiation delays between
337 *Smg5* and *Smg7* KOs (*Smg5* >> *Smg7*) together with the strong synergistic effects after co-depletion are
338 difficult to reconcile with the proposed obligatory heterodimer dependence of these two factors^{24,26}. We
339 show that a division of labor between *Smg5* and *Smg7* is causative for the different effects of depletion in
340 the respective KOs. *Smg* factors have a dual role in triggering RNA degradation and in mediating Upf1
341 dephosphorylation; pUpf1 increase in all three *Smg* KOs accords with such a role of *Smg* factors in PP2A
342 recruitment^{25,26}. Our data suggest that *Smg7* acts as the main adapter or recruiter for pUpf1. However,
343 by itself it is unable to efficiently dephosphorylate Upf1, as evidenced by high pUpf1 levels in *Smg5* KO
344 ESCs, where *Smg7* binding to pUpf1 is unaffected. We propose that *Smg7* binding to Upf1 without
345 subsequent Upf1 dephosphorylation results in jamming of the dephosphorylation cycle, and ensuing
346 stalling of the mRNA degradation circuit. Aberrant recruitment of *Smg6* to mRNA targets already bound
347 by *Smg7* and pUpf1 cannot properly restore NMD function in *Smg5* KOs. *Smg5* alone is unable to bind
348 pUpf1 in our assays. This suggests that the main role of *Smg5* lies in its dephosphorylation activity, but

349 recruitment to its substrate is Smg7-dependent. This is consistent with the strongest increase of pUpf1
350 levels and consequently the strongest differentiation delays in *Smg5* KO ESCs.

351 Absence of *Smg6* causes a strong differentiation delay. This suggests that the exonucleolytic pathway
352 cannot completely compensate for loss of the endonucleolytic activity in *Smg6* KOs. *Smg7* KO leads to
353 very weak phenotypes and low levels of NMD deregulation. Our data support a model in which, in the
354 absence of Smg7, Smg5 cannot be recruited to pUpf1-bound mRNAs, enabling Smg6 to recognize these
355 potential targets and initiate their degradation. Thereby Smg6 can largely compensate for loss of Smg7,
356 resulting in only mild NMD defects in Smg7 KO cells. However, Smg6 is unable to compensate for the
357 complete absence of the exonucleolytic branch of NMD, suggested by the apparent loss of viability in a
358 *Smg5/7* dKO situation. This indicates a function of Smg5 and/or Smg7 in the endonucleolytic decay
359 pathway. Our data are consistent with a function of Smg5 or Smg7 in Upf1 dephosphorylation, which is a
360 key step also in the endonucleolytic Smg6-mediated mRNA decay axis. Despite the weak defects in single
361 *Smg7* KO cells, facultative essentiality for *Smg7* is revealed in the absence of either *Smg5* or *Smg6*.
362 Accordingly, only *Smg5/Smg6* dKO ESCs could be established, showing that Smg7 is sufficient to sustain
363 minimal NMD activity required for survival. This suggests that Smg7 can either interact with potentially
364 novel interactors in the absence of its normal interaction partners, or that Smg7 acts as a jack of all trades,
365 fulfilling minimal roles in target recognition, recruitment of the degradation machinery and Upf1
366 dephosphorylation. Taken together, the mode of action described above explains both the graded pUpf1
367 levels as well as graded impact on NMD of Smg5, Smg6 and Smg7 KOs.

368 A combination of RNA-seq and SLAM-seq based half-life analyses allowed us to compile a list of high-
369 confidence NMD targets during the exit from pluripotency. Our data show partial redundancy and highly
370 overlapping target gene sets between NMD downstream effectors in ESCs, consistent with previous
371 results showing shared NMD targets between Smg6 and Smg7¹⁹. However, while target mRNA identity

372 appears identical, the amplitude of mRNA deregulation scales with both the phenotype and the increase
373 in pUpf1 levels.

374 NMD targets showing a graded upregulation pattern (Smg5 > Smg6 > Smg7) include the translation
375 initiation factor *Eif4a2*. In contrast to previous reports, *c-myc* was not identified as an NMD target and
376 showed no relevance for the differentiation defect in our experiments¹³. We believe that our use of state-
377 of-the-art ESC culture conditions facilitated the identification of differentiation-relevant NMD targets,
378 that are obscured in heterogeneous FCS/LIF culture conditions.

379 In mammals, two homologs of the Eif4a helicase, Eif4a1 and Eif4a2, can be integrated into the eIF4F
380 complex^{30,31}. In our datasets only Eif4a2, and not Eif4a1, was found to be regulated by NMD. In addition
381 to encoding for a full-length protein, the *Eif4a2* locus also produces a distinct PTC-containing isoform. In
382 the truncated Eif4a2^{PTC} isoform 45 aa of the C-terminal helicase domain are missing, without affecting the
383 ATP binding domain. Transcripts of both isoforms showed increased half-lives in NMD KO cells and reacted
384 strongly to translation inhibition, a hallmark of bona fide NMD targets.

385 Control of protein synthesis rates is fundamental for maintenance of self-renewal and differentiation^{37,38}
386 and for maintaining an ESC-specific chromatin state^{39,40}. ESCs require a downregulation of translation
387 rates and ribosome biogenesis to successfully differentiate^{40,41}. We detected a marked increase in
388 translation rates in NMD KO ESCs, which is not solely an effect of increased mRNA abundance due to
389 increased mRNA stability, but directly dependent on increased Eif4a2 levels. This is evident from double-
390 depletion experiments, in which *Eif4a2* depletion in NMD KOs reduces translation and partly rescues
391 differentiation kinetics without restoring NMD function.

392 Increased protein levels of the Eif4a2^{PTC} isoform and not the full-length protein disrupt the normal
393 translation programme and the exit from naïve pluripotency. This became evident in NMD KO cells in
394 which the PTC exon was excised. In these cells, differentiation kinetics were restored despite increased

395 expression of Eif4a2^{FL}. Furthermore, only Eif4a2^{PTC} overexpression elicits clear differentiation delays. The
396 function of Eif4a2^{PTC} appears dependent on the presence of Eif4a2^{FL} because overexpression of Eif4a2^{PTC}
397 alone in Eif4a2 KO ESCs has no detectable impact on differentiation. This constitutes evidence that the
398 translation initiation potential of Eif4a2 can be increased by its PTC isoform, which in turn requires the
399 full-length protein to exert its function.

400 Eif4a2^{PTC} protein is detectably expressed in ESCs, but its physiological role remains to be determined. The
401 observed interaction between Eif4a2^{FL} and Eif4a2^{PTC} with key pluripotency TFs is of special note. This
402 interaction with the pluripotency circuitry remains to be explored, but could provide a direct link between
403 translation initiation and cell fate control. Such a function accords with recent reports of Sox2 binding to
404 the translation initiation factor transporter Eif4enif1, and Sox2 binding to RNA in mouse ESCs ^{44, 45}.

405 NMD is triggered during translation. NMD-mediated regulation of Eif4a2 provides a link back from NMD
406 to translation initiation, thereby establishing a feedback circuit (Fig. 6i). We propose that such a
407 mechanism can be used to balance NMD activation with translational activity. We propose Eif4a2^{PTC} to act
408 as a rheostat: Its upregulation upon NMD dysfunction increases translation initiation activity, which in
409 turn directly increases the chances of triggering NMD in successive rounds of Eif4a2-initiated translation.
410 In line with this hypothesis, our data confirm a reported physical interaction between Eif4a2 and Smg
411 factors ³⁵. Underscoring the importance of the Eif4a2^{PTC} mediated feedback loop to translation, the PTC-
412 containing exon shows an unexpectedly high level of evolutionary conservation in various mammalian
413 species, including human.

414 In conclusion, we show that NMD is required for proper restructuring of GRNs during differentiation by
415 targeting multiple transcripts, most prominently the translation initiation factor Eif4a2. Our results place
416 NMD as a central player in shaping transcriptomes to maintain proper cellular identity. We provide
417 evidence for an intricate feedback circuit between NMD and translation. NMD susceptibility is hardwired

418 in an evolutionarily conserved fashion into the *Eif4a2* gene structure and sequence. This direct link
419 between NMD and translation initiation will need to be considered when studying NMD associated
420 phenotypes. Our results are also relevant for improving therapeutic interventions and provide a rational
421 for choosing the appropriate Smg-factor in pharmacological approaches to inhibit NMD activity,
422 depending on the desired strength of NMD inactivation¹⁵.

423

424

425

426 **AUTHOR CONTRIBUTIONS**

427 EG performed experiments and analyzed data. RS, MR, FTT, LH and MG performed bioinformatic analysis,
428 supervised by AB, AvH and CB. MH, JR, SS and LS performed experiments. CW and CS analyzed teratoma
429 samples. AC, KFL and AP supported translation measurements. VH and SLA performed half-life
430 measurements. EG, AS and ML designed experiments. ML supervised the study and wrote the manuscript
431 together with EG with feedback from all authors.

432

433 **ACKNOWLEDGEMENTS**

434 We thank Thomas Sauer and Johanna Stranner at the Max Perutz Labs FACS facility for expert support,
435 Markus Hartl and the Massspec facility for mass-spec analysis and Irmgard Fischer for help with
436 microscopy. NGS was performed at the VBCF. This work was supported by the Austrian Science Fund (FWF;
437 P31334). Martin Leeb is a WWTF VRG group leader (VRG14-006) and was supported by an FWF
438 Schroedinger return fellowship.

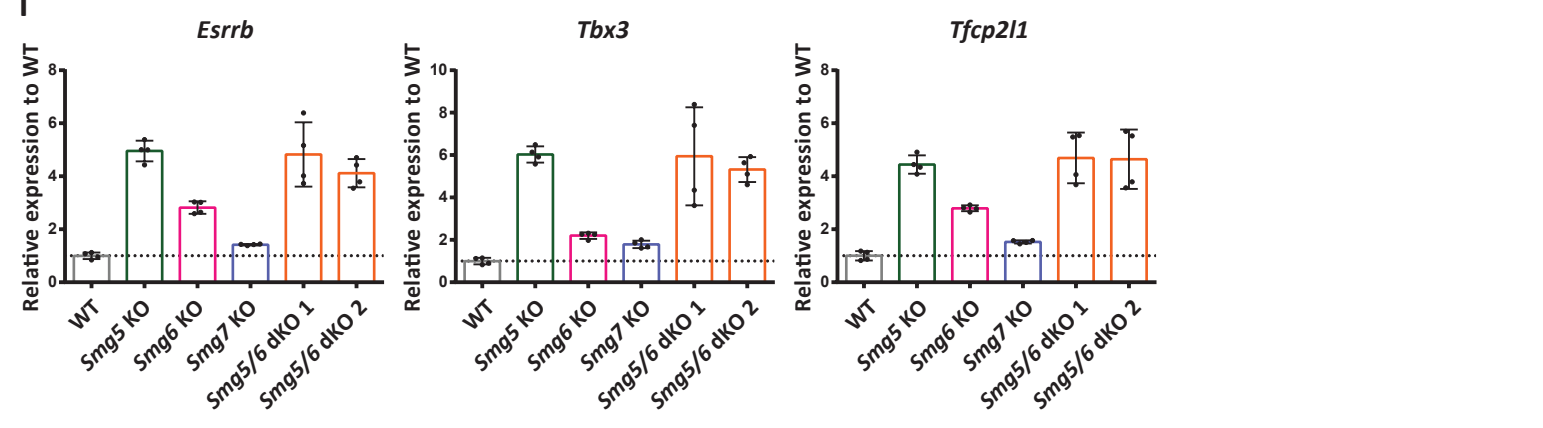
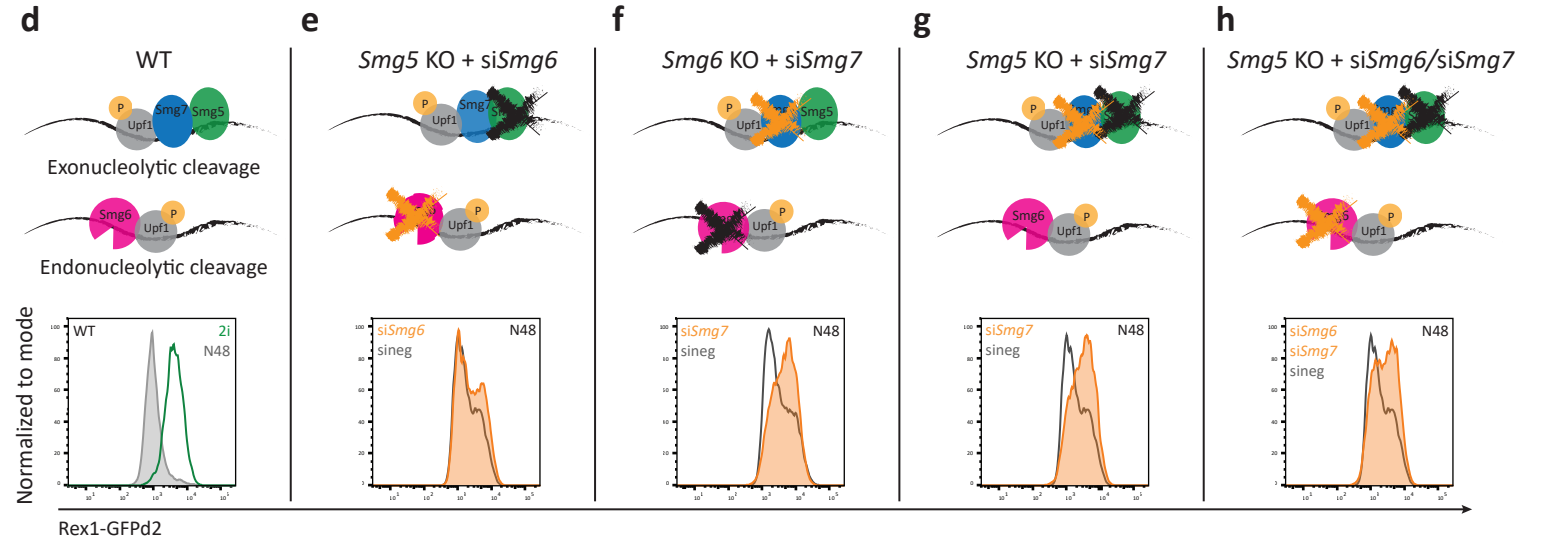
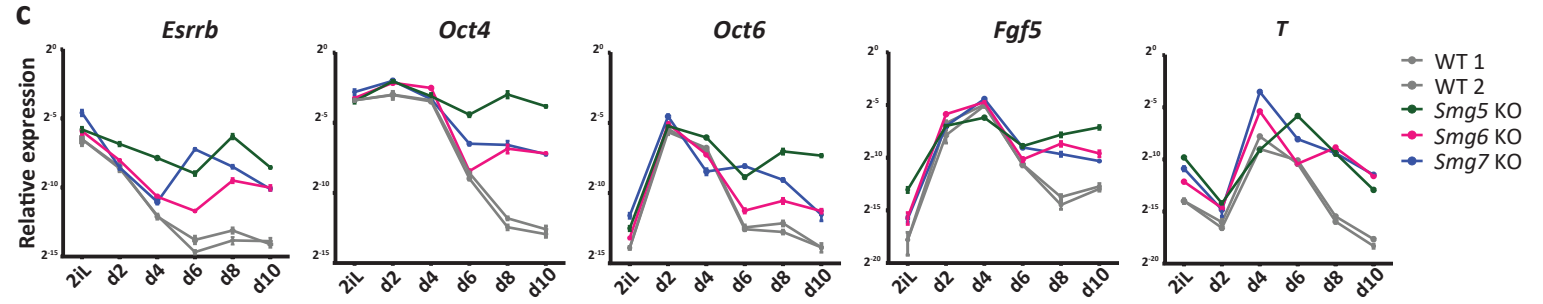
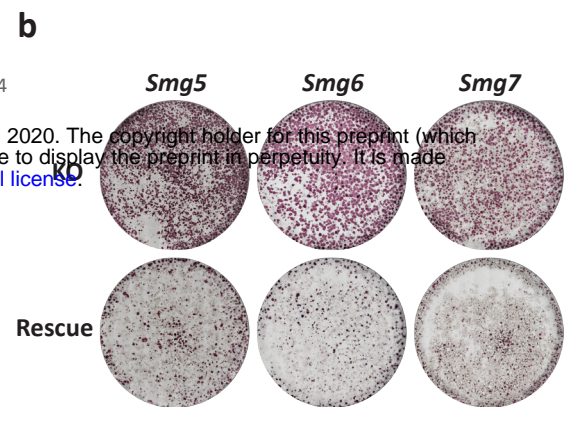
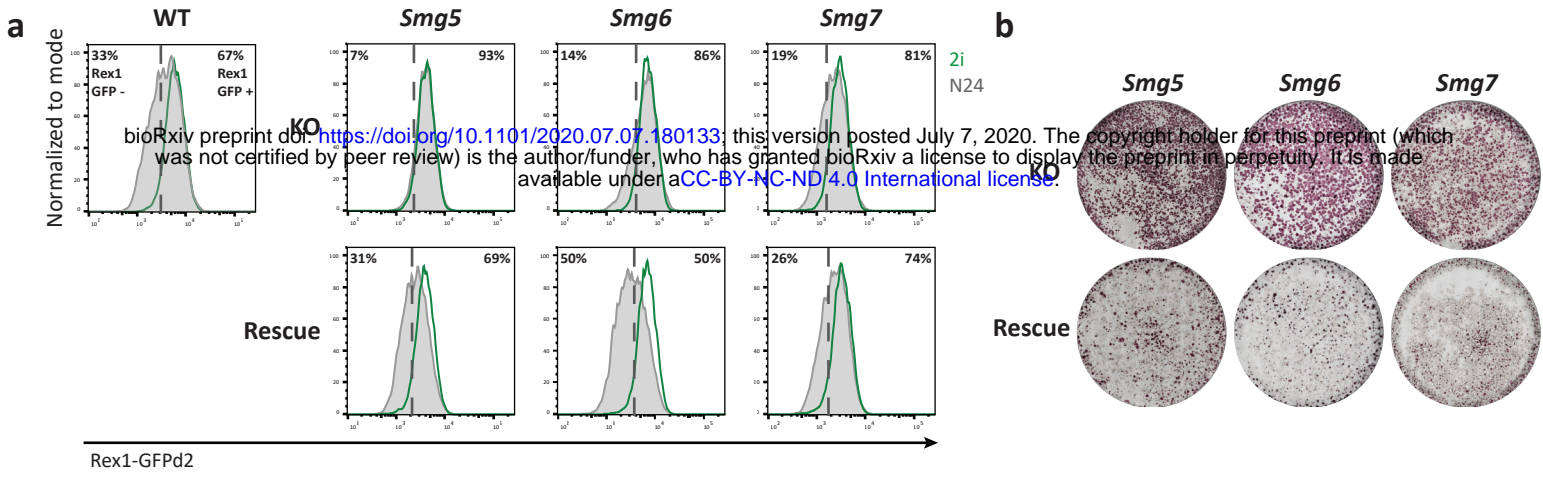
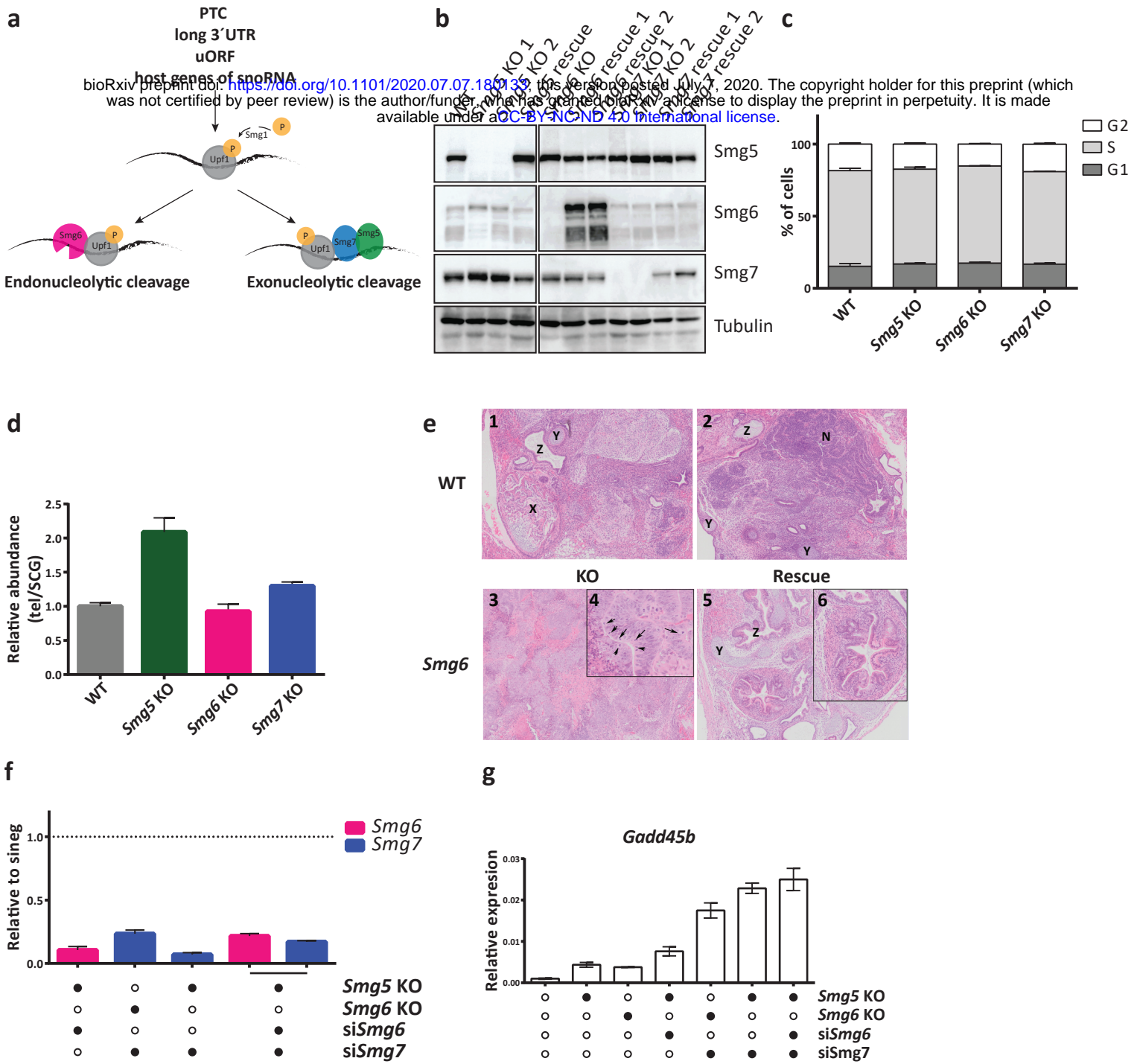


Figure 1

439 **Fig. 1 | Hierarchy of defects in exit from naïve pluripotency in NMD-deficient ESCs. a**, Rex1-GFPd2
440 analysis by FACS 24 h after induction of differentiation by withdrawing 2i in N2B27 medium (N24) in NMD
441 KO, NMD rescue and WT ESCs. Green indicates profiles in 2i and grey indicates N24 profiles. **b**,
442 Commitment assay in NMD KO and NMD rescue ESCs confirms commitment defects. Cells were re-plated
443 in 2i medium after 72 h of differentiation. Only non-committed cells can reinitiate ESC self-renewal.
444 Alkaline phosphatase staining was used to visualize ESC colonies. **c**, Representative experiment for
445 expression kinetics of the indicated genes during a 10-day embryoid body (EB) differentiation assay. Mean
446 and SD of technical replicates are plotted for each time point. Expression levels were normalized to 60S
447 ribosomal protein L32 Rpl32 (*L32*). **d-h**, Rex1-GFPd2 analysis at N48 of WT cells (**d**) and NMD KO and NMD
448 dKO cells after siRNA KD of *Smg6* or *Smg7* (**e-h**). **i**, Expression of the indicated genes at N24 in NMD KO,
449 *Smg5/Smg6* dKO and WT ESCs. Mean and SD are plotted for each cell line, n=2 biological replicates.
450 Expression was normalized first to *Actin* and results are shown relative to WT. Unpaired t-test was used
451 to calculate p-values (**** p < 0.02 unless indicated in the graphs).



453 **Extended Data Fig. 1 | Hierarchy of defects in exit from naïve pluripotency in NMD-deficient ESCs. a,**
454 Illustration of the endo- and exonucleolytic branches of NMD. **b,** Western blot analysis in WT, NMD KO
455 and NMD rescue ESCs using the indicated antibodies. Tubulin was used as loading control. **c,** Cell cycle
456 analysis of NMD KO and WT ESCs. Propidium iodide was used to stain the cells and the Watson Pragmatic
457 model was used to calculate the percentage of cells in each cell cycle phase. Mean and SEM are plotted
458 for each cell cycle phase, n=3 biological replicates. **d,** Telomere length calculation in NMD KO and WT
459 ESCs. Mean and SD of technical replicates are plotted for each cell line. Results were normalized to the
460 36B4 single copy locus. **e,** Teratomas derived from WT, *Smg6* KO and *Smg6* rescue ESCs. **1-2:** areas of WT-
461 derived teratoma showing well differentiated tissues of mesodermal (e.g., X enchondral ossification; Y
462 cartilage), endodermal (e.g., Z), and ectodermal (neuronal rosettes; N) origin. **3:** Area of *Smg6* KO-derived
463 teratomas showing abundant poorly differentiated neuronal tissue; poorly differentiated endodermal and
464 mesodermal tissues are present at lower abundancy (not shown). Abundant mitotic figures present in
465 neuronal tissue (arrows) demonstrate high proliferative capacity (**4**). **5:** Area of *Smg6* rescue-derived
466 teratomas showing examples of well differentiated endodermal (Z) and mesodermal (Y: cartilage) tissues.
467 Higher power magnification shows an endodermal duct with different, well developed epithelial cell types
468 (**6**). **f,** qPCR analysis confirming the siRNA-mediated knockdown of NMD components. Mean and SD of
469 technical replicates are plotted for each cell line. Expression was normalized first to *L32*. Expression levels
470 after sineg transfection were set to 1. **g,** Expression levels of *Gadd45b* in the indicated cells lines.
471 Expression was normalized to *Actin*. Mean and SD of technical replicates are plotted for each cell line.

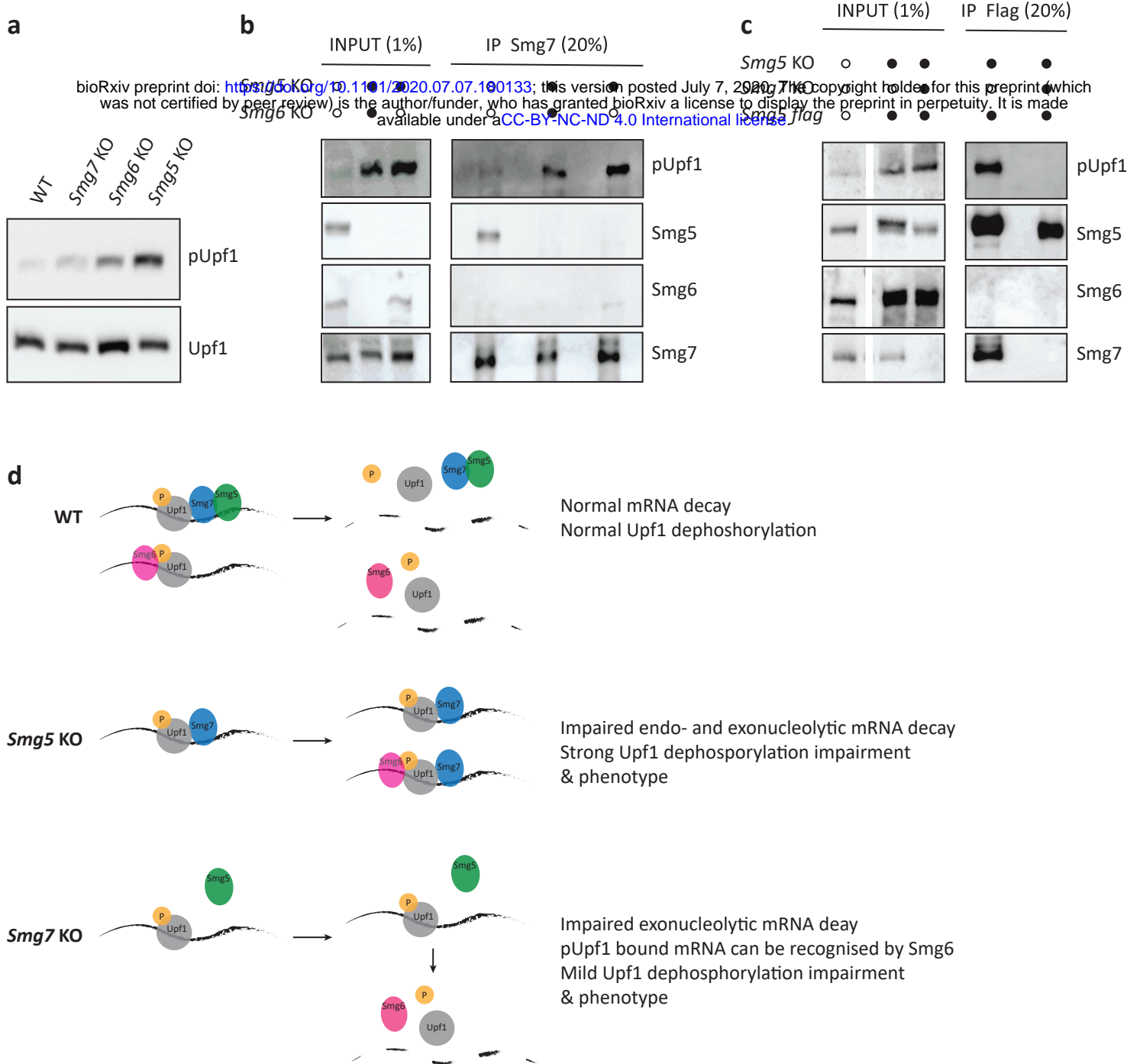
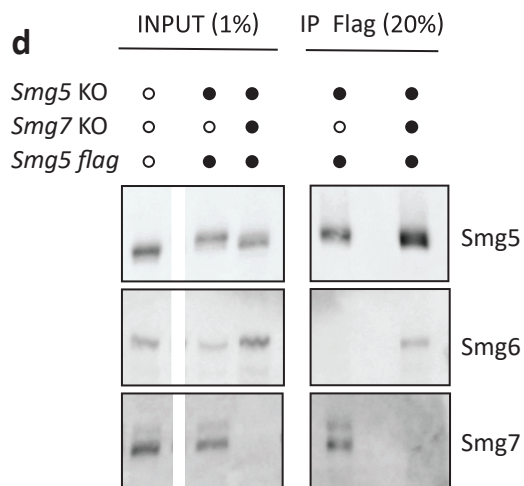
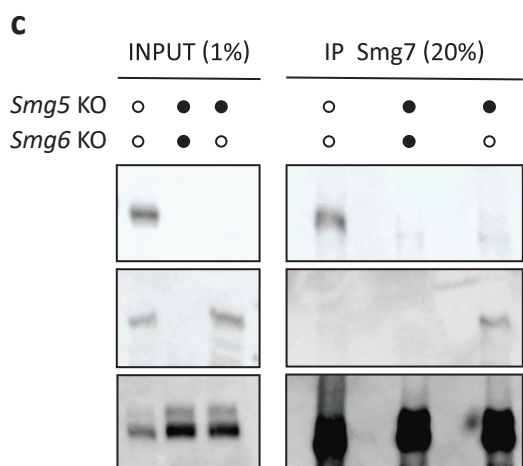
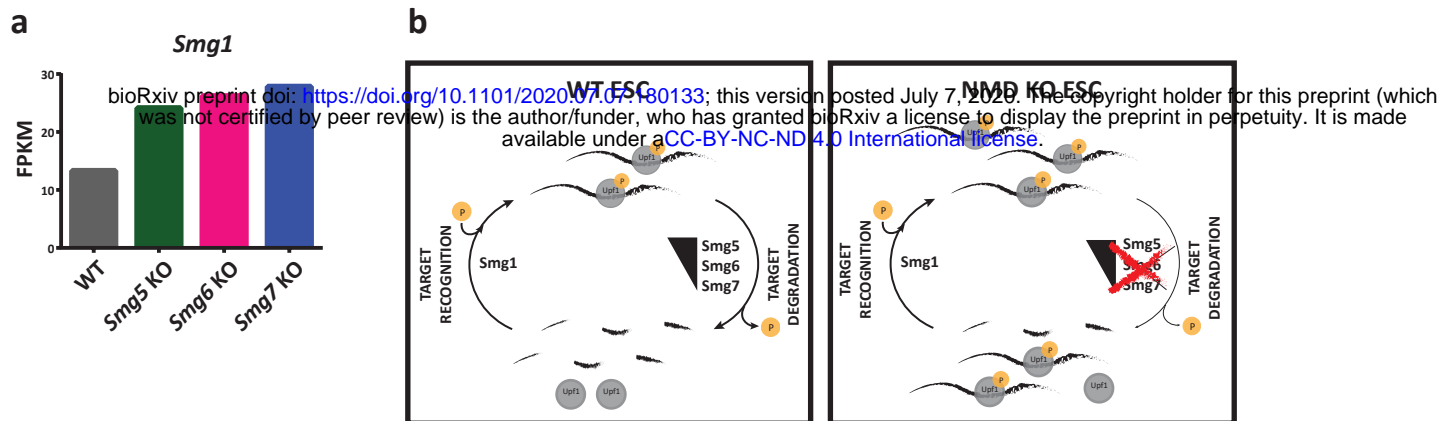


Figure 2

473 **Fig. 2 | Smg7 is necessary and sufficient for pUpf1 binding, independently of Smg5.** **a**, Western blot
474 analysis for Upf1 and pUpf1 expression in NMD KO and WT ESCs. **b**, Western blot showing results of Smg7
475 co-IP in the indicated cell lines after cyanase treatment for RNA removal. Antibodies used are indicated in
476 the figure. **c**, Western blot analysis of Smg5 co-IP (flag antibody co-IP in Smg5KO^{Smg5flagrescue} cells) in the
477 indicated cell lines after cyanase treatment. Antibodies used are indicated in the figure. **d**, Schematic
478 illustration of the proposed interactions between NMD effectors and pUpf1 dephosphorylation in WT,
479 *Smg5* and *Smg7* KO ESCs that lead to a graded deficiency in NMD and exit from naïve pluripotency.



481 **Extended Data Fig. 2 | Smg7 is necessary and sufficient for pUpf1 binding, independently of Smg5. a,**
482 Expression levels (FPKM) of *Smg1* derived from RNA-seq in WT and NMD KO ESCs in 2i. Mean values
483 between biological duplicates are plotted. **b,** Schematic representation of phosphorylation and
484 dephosphorylation cycle of Upf1 in WT and NMD KO ESCs. **c,** Western blot of Smg7 co-IP in the indicated
485 cell lines without cyanase treatment. Antibodies used are indicated in the figure. **d,** Western blot analysis
486 of Smg5 co-IP (flag antibody) in the indicated cell lines without cyanase treatment. Antibodies used are
487 indicated in the figure. **c** and **d** are parallel experiments to those shown in Figure 2b and c.

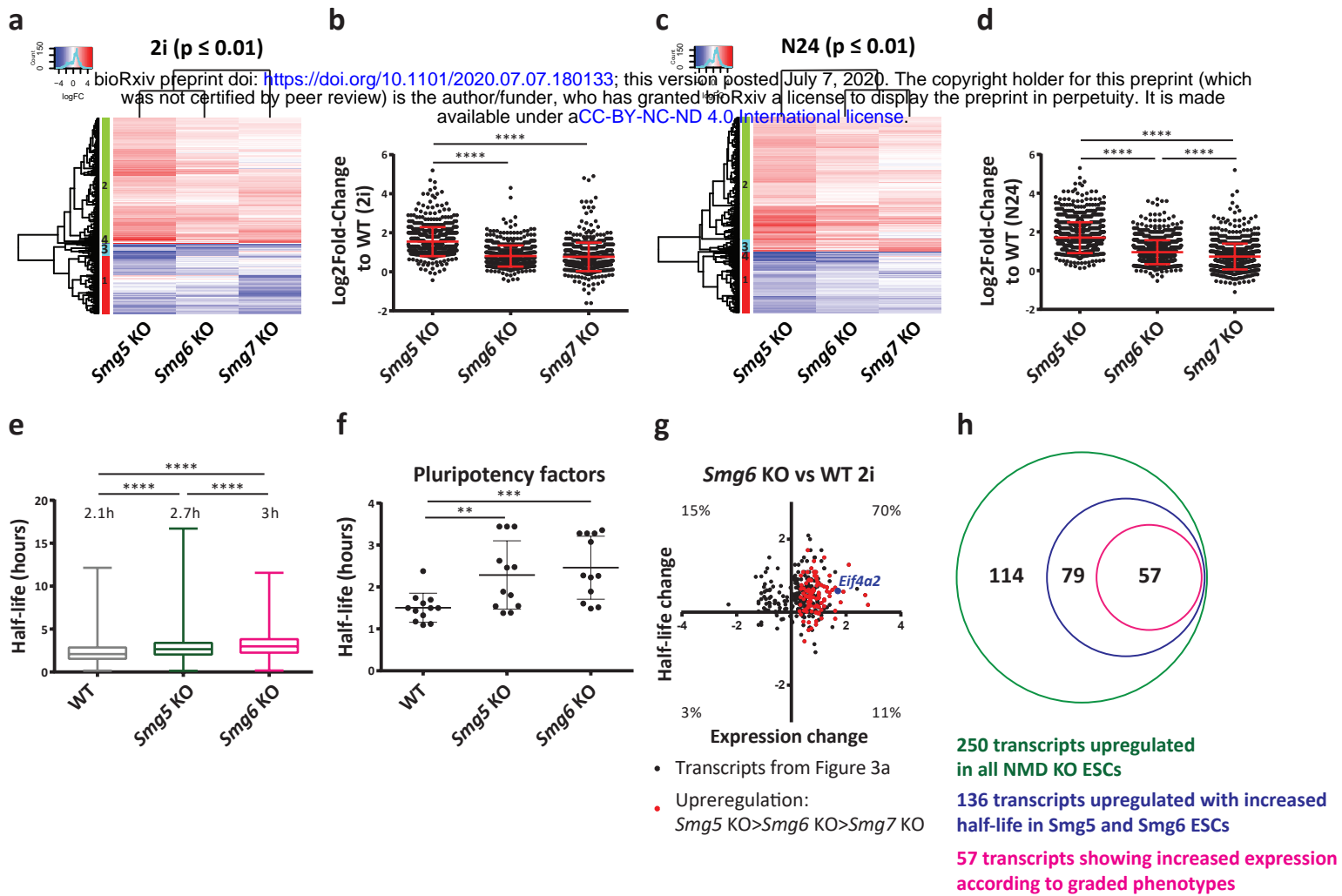
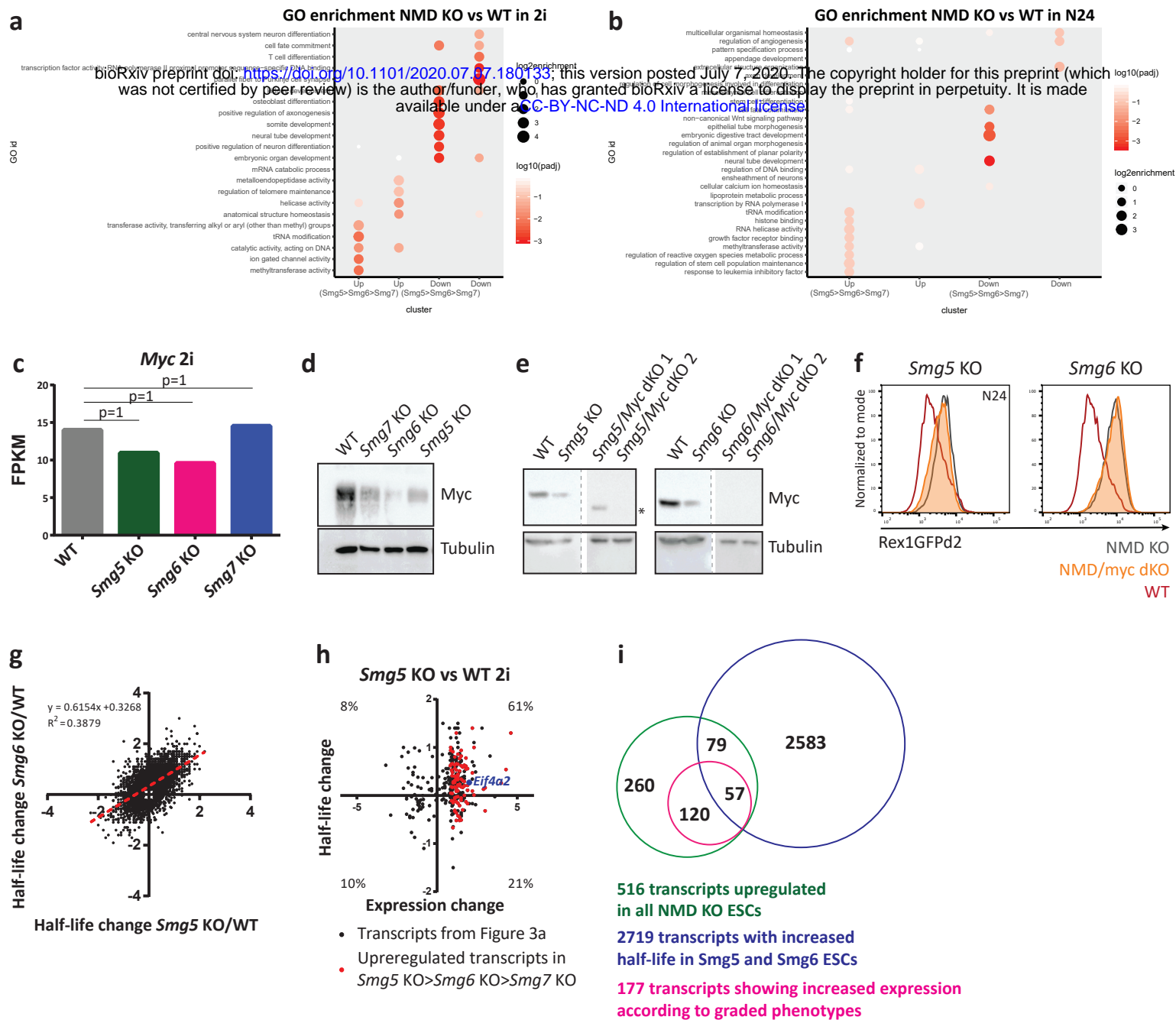


Figure 3

489 **Fig. 3 | Integrating transcriptome-wide gene expression with mRNA half-life analyses identifies relevant**
490 **NMD targets during the exit from naïve pluripotency.** **a**, Differentially expressed genes between NMD
491 KO and WT ESCs in self-renewal (2i) (p-adj. ≤ 0.01). p-adj. values were calculated testing against the null
492 hypothesis of a fold change $< |1.5|$. **b**, Log2fold change of deregulated transcripts in 2i belonging to
493 clusters 2 and 4 (564 transcripts). Mean and SD are indicated in the graph. Unpaired t-test was used to
494 calculate p-values. (**** p-value < 0.0001). **c**, Differentially expressed genes between NMD KO and WT
495 ESCs at N24 (p-adj. ≤ 0.01). p-adj. values were calculated testing against the null hypothesis of a fold
496 change $< |1.5|$. **d**, Log2fold change of deregulated transcripts at N24 belonging to clusters 2 and 3 (808
497 transcripts). Mean and SD are indicated in the graph. Unpaired t-test was used to calculate p-values. (****
498 p-value < 0.0001). **e**, Half-life analysis in WT, *Smg5* KO and *Smg6* KO ESCs. Half-life was calculated in hours
499 based on a pulse chase time course. Mean and SD are indicated in the graph. Paired t-test was used to
500 calculate p-values (**** p-value < 0.0001). **f**, Half-life of pluripotency factors (*Esrrb*, *Klf2*, *Klf5*, *Nanog*,
501 *Pou5f1*, *Sall4*, *Sox2*, *Stat3*, *Tbx3*) in WT, *Smg5* KO and *Smg6* KO ESCs. Half-lives were calculated in hours.
502 Mean and SD are indicated in the graph. Paired t-test was used to calculate p-values (** p-value = 0.006,
503 *** p-value = 0.0006). **g**, Transcriptome-wide comparison of changes in mRNA expression and half-life in
504 *Smg6* KO. Half-life changes between WT and *Smg6* KO for the 321 out of 516 transcripts upregulated in
505 NMD KO ESCs for which half-lives could be calculated were calculated (see Figure 3a). 150 of these 321
506 transcripts showed an increase in half-life ($\log_2FC \geq 0.2$) in both *Smg5* and *Smg6* KOs. Transcripts behaving
507 in accordance with the graded phenotype (*Smg5* > *Smg6* > *Smg7*) are depicted in red. **h**, Schematic
508 illustration of the approach used to identify NMD targets relevant for the differentiation phenotype in
509 ESCs. First significantly upregulated transcripts (see Figure 3a) with measurable half-lives in WT and *Smg*
510 KOs were selected (250). 136 transcripts showed a concomitant increase in half-life in *Smg5* and *Smg6*
511 KOs ($\log_2FC \geq 0.2$). 57 of these transcripts were upregulated according to the graded phenotypes.

512



514 **Extended Data Fig. 3 | Integrating transcriptome-wide gene expression with mRNA half-life analyses**
515 **identifies relevant NMD targets during the exit from naïve pluripotency. a**, Significantly enriched GO
516 terms of differentially expressed transcripts in NMD KO in 2i. **b**, Similar to **(a)** for differentially expressed
517 transcripts at N24. **c**, Expression levels of *c-myc* in NMD KO and WT ESCs in RNA-seq. Mean values between
518 biological duplicates are plotted. p-adj. values are indicated above each bar. **d**, Western blot analysis in
519 NMD KO and WT ESCs using the antibodies indicated. Tubulin was used as loading control. **e**, Western blot
520 analysis confirming absence of myc protein in NMD/myc dKO ESCs. Antibodies are indicated in the figure.
521 Tubulin was used as loading control. * band corresponds to truncated protein. **f**, Rex1-GFPd2 FACS profile
522 at N24 in NMD KO and NMD/myc dKO cells. FACS profile of NMD KO cells is grey, orange profile shows
523 NMD/myc dKO cells and red profile shows WT. **g**, Global analysis of NMD-induced half-life change in *Smg5*
524 KO and *Smg6* KO ESCs (all transcripts with a half-life change $\geq |0.2|$ in *Smg5* or *Smg6* KO ESCs are plotted.
525 Half-life change was calculated for 7,451 transcripts. Slope and R2 values are shown in the figure. **h**,
526 Transcriptome-wide analysis of changes in mRNA expression and half-lives in *Smg5* KO. Half-life changes
527 could be calculated for 321 out of the 516 transcripts upregulated in NMD KO ESCs (Figure 3a). 182 of the
528 516 transcripts showed a concomitant increase in half-life (half-life change ≥ 0.2). Transcripts behaving
529 according to the graded phenotype are depicted in red. **i**, Venn diagram showing 516 transcripts
530 upregulated in all NMD KO ESCs intersected with 177 transcripts showing graded increase in expression
531 in NMD factor KOs (*Smg5* > *Smg6* > *Smg7*) and 2,719 transcripts with increased half-lives in both *Smg5*
532 and *Smg6* KO ESCs.

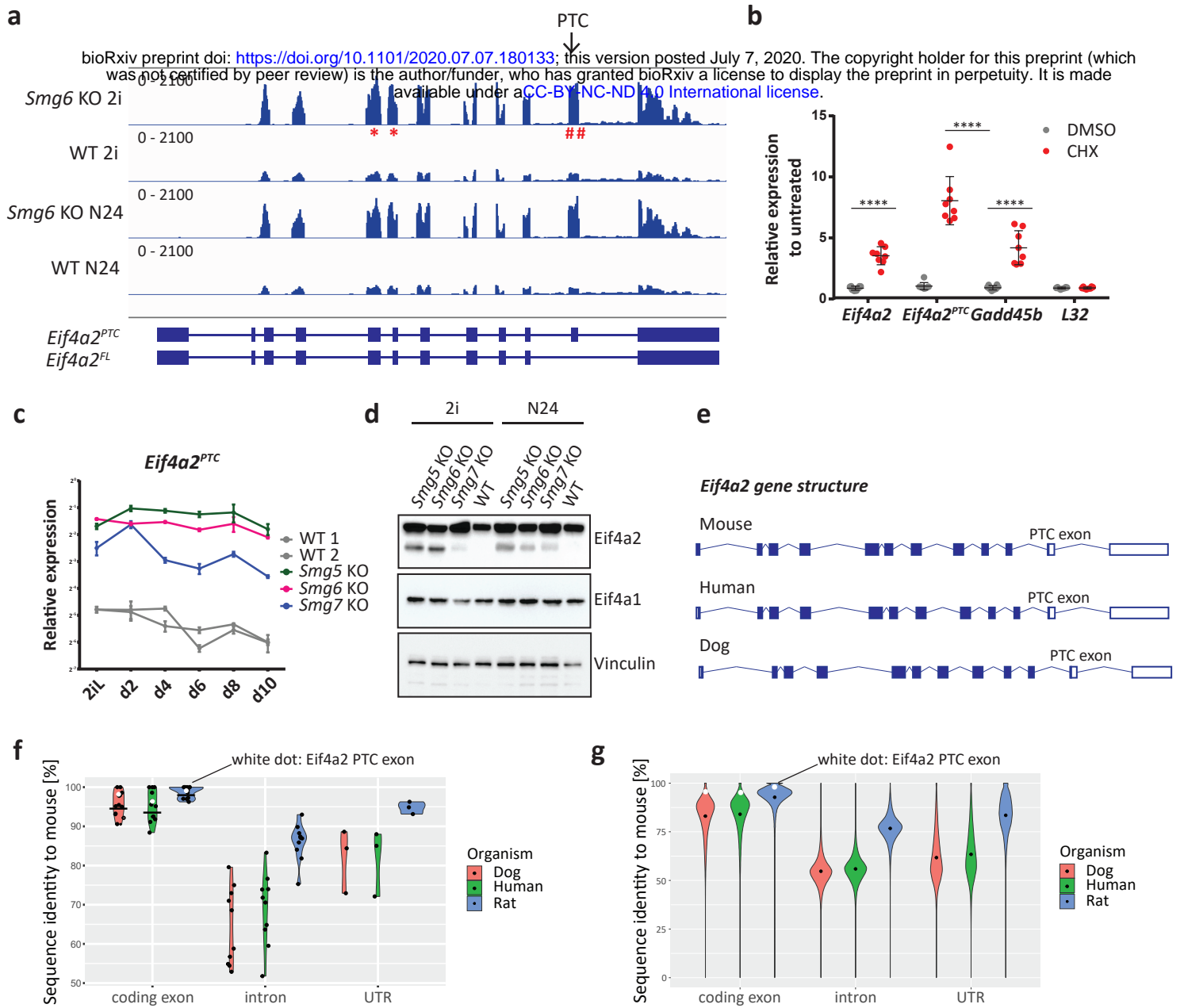
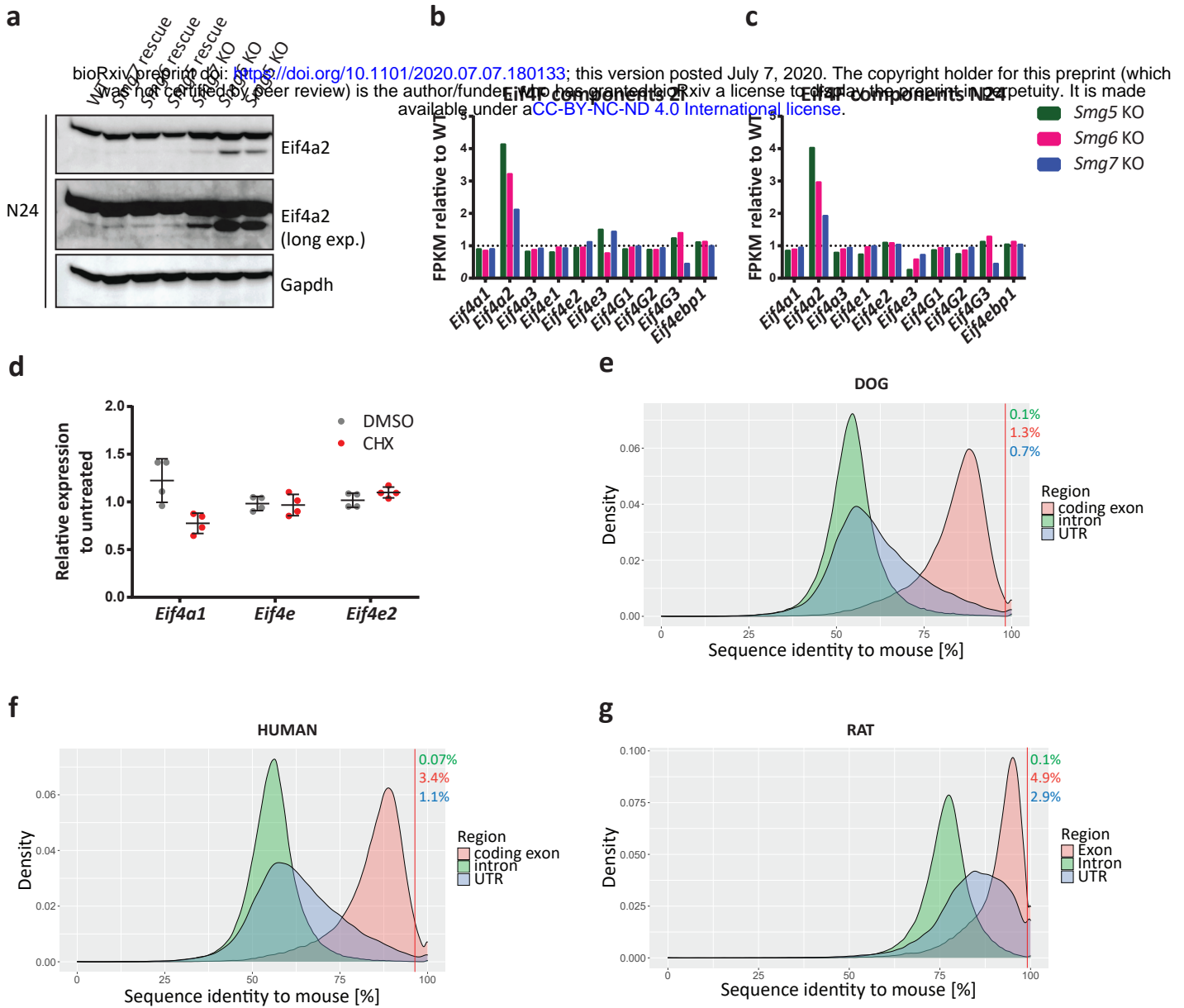


Figure 4

534 **Fig. 4 | Eif4a2 is a bona-fide NMD target in ESCs. a**, Genome browser view of the *Eif4a2* locus visualizing
535 RNA-seq results in *Smg6* KO and WT ESCs in 2i and at N24. * indicates qPCR primers amplifying all *Eif4a2*
536 isoforms. # indicates qPCR primers amplifying the *Eif4a2*^{PTC} isoform. **b**, Expression levels of the indicated
537 genes after treatment with CHX or DMSO in WT ESCs. Mean and SD are plotted, n=4 biological replicates.
538 Unpaired t-test was used to calculate p-values (**** p < 0.0001). **c**, Representative experiment for
539 expression kinetics of the indicated genes during an EB differentiation assay as in Figure 1c. Mean and SD
540 from technical replicates are plotted for each time point. Expression levels were normalized to *L32*. **d**,
541 Western blot analysis of Eif4a2 and Eif4a1 protein levels in NMD KO and WT ESCs in self-renewal (2i) and
542 at N24. Vinculin was used as loading control. **e**, Schematic representation of the gene structure of *Eif4a2*
543 in mouse, human and dog. **f**, Percentage of sequence identity to mouse of *Eif4a2* between dog, rat and
544 human. White dots indicate the PTC-containing exon of *Eif4a2* and black lines indicate the mean. **g**,
545 Percentage of sequence identity to mouse of the transcriptome between dog, rat and human. White
546 asterisks indicate the PTC-containing exon of *Eif4a2* and the black dots indicate the mean.



548 **Extended Data Fig. 4 | Eif4a2 is a bona-fide NMD target in ESCs.** **a**, Western blot analysis for Eif4a2
549 protein levels in NMD KO, NMD rescue and WT cells at N24. Gapdh was used as loading control. **b**,
550 Expression of Eif4F complex components and regulators in NMD KO ESCs relative to WT ESCs in self-
551 renewal (2i). FPKM values from RNA-seq analysis are shown, see Figure 3a. **c**, Expression of Eif4F complex
552 components and regulators in NMD KO cells relative to WT cells at N24. Values from RNA-seq analysis,
553 see Figure 3c. **d**, Expression levels of the indicated genes after treatment with CHX or DMSO in WT ESCs,
554 assayed by RT-qPCR. Mean and SD are plotted, n=2 biological replicates. **e-g**, Densities of the distributions
555 of the sequence identities of indicated regions of the mouse transcriptome compared to dog (**e**), human
556 (**f**) and rat (**g**). Red line indicates identity of PTC-containing *Eif4a2* exon. Percentage of genomic sequences
557 with higher conservation than PTC-containing exon of *Eif4a2* are indicated in the graphs.

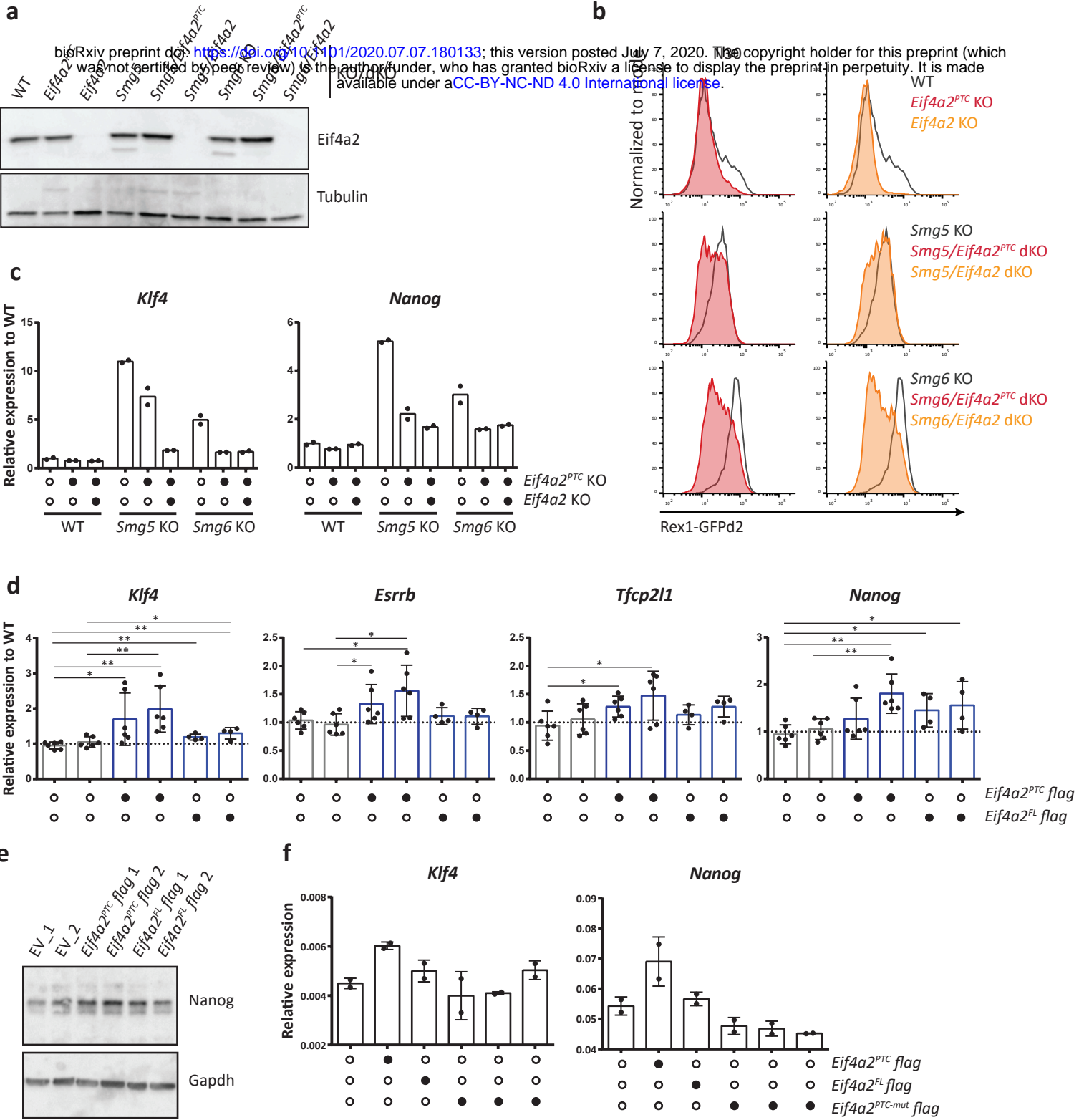
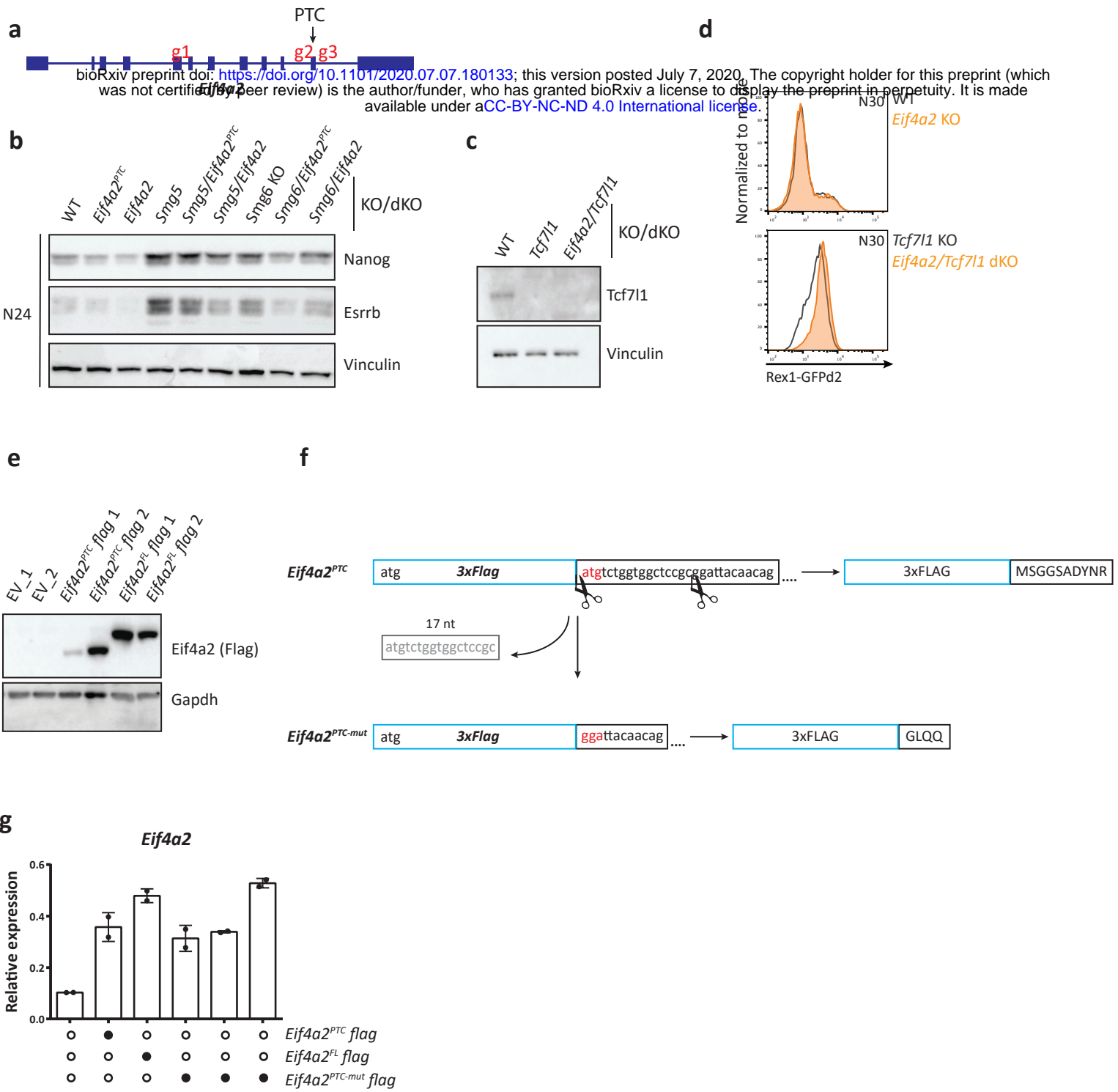


Figure 5

559 **Fig. 5 | *Eif4a2* is causative for defects in exit from naïve pluripotency in NMD-deficient ESCs.** **a**, Western
560 blot analysis of *Eif4a2* in the indicated cell lines. Tubulin was used as a loading control. **b**, Rex1-GFPd2
561 analysis at N30 in NMD KO and NMD/*Eif4a2* dKO ESCs. Grey indicates FACS-profiles of WT cells, red
562 indicates profiles of *Eif4a2*^{PTC} deficient (*Eif4a2*^{PTC} and NMD/*Eif4a2*^{PTC}) cells and orange indicates profiles of
563 *Eif4a2* deficient (*Eif4a2* KO and NMD/*Eif4a2* dKO) cells. **c**, Representative experiment for the expression
564 of indicated genes at N30 in NMD KO and NMD/*Eif4a2* dKO cells. Mean of technical replicates plotted for
565 each cell line. Expression was normalized first to *L32* and then to WT cells. **d**, Expression of the indicated
566 genes at N30 in *Eif4a2* overexpressing and WT cells. Mean and SD are plotted for each cell line, n=3
567 biological replicates. Expression was normalized first to housekeeping genes (*actin* or *L32*). The expression
568 level in WT cells was set to 1. Unpaired t-test was used to calculate p-values (* p < 0.05, ** p < 0.005). **e**,
569 Western blot analysis for Nanog in *Eif4a2* overexpressing and WT cells at N24. Gapdh was used as loading
570 control. **f**, Expression of the indicated genes at N16 in *Eif4a2* overexpressing and WT cells. Mean and SD
571 of technical replicates are plotted for each cell line.



573 **Extended Data Fig. 5 | *Eif4a2* is causative for defects in exit from naïve pluripotency in NMD-deficient**
574 **ESCs. a**, Schematic view of the *Eif4a2* gene. gRNAs (g1 to g3) used to generate *Eif4a2* and *Eif4a2^{PTC}*
575 depletions are indicated. g1 and g3 were used together to delete all *Eif4a2* isoforms. g2 and g3 were used
576 together to delete the *Eif4a2^{PTC}* isoform. **b**, Western blot analysis at N24 in the annotated cell lines for
577 pluripotency markers *Esrrb* and *Nanog*. Vinculin was used as loading control. **c**, Western blot analysis for
578 *Tcf7l1* expression in the indicated cell lines. Vinculin was used as loading control. **d**, Rex1-GFPd2 analysis
579 at N30 in WT (grey), *Eif4a2* KO (orange), *Tcf7l1* KO (grey) and *Eif4a2/Tcf7l1* dKO (orange) cells. **e**, Western
580 blot analysis of *Eif4a2* (flag antibody) in *Eif4a2* overexpressing *Eif4a2* KO cells. EV transfection shows no
581 signal. *Gapdh* was used as loading control. **f**, Schematic illustration of the mutation introduced into the
582 *Eif4a2^{PTC}* isoform. **g**, Expression of *Eif4a2* at N16 in *Eif4a2* overexpressing pools and WT cells. Mean and
583 SD of technical replicates are plotted for each cell line.

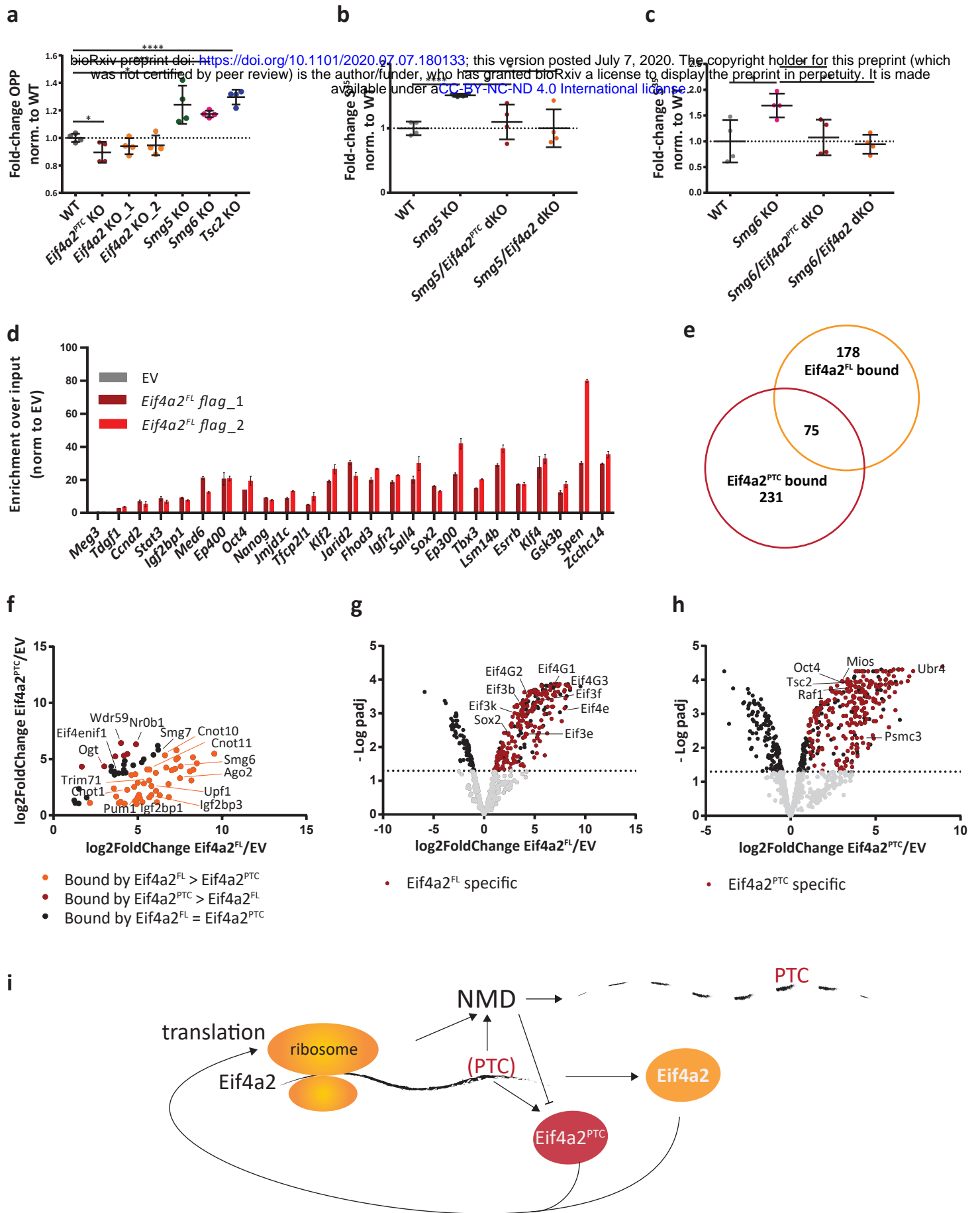
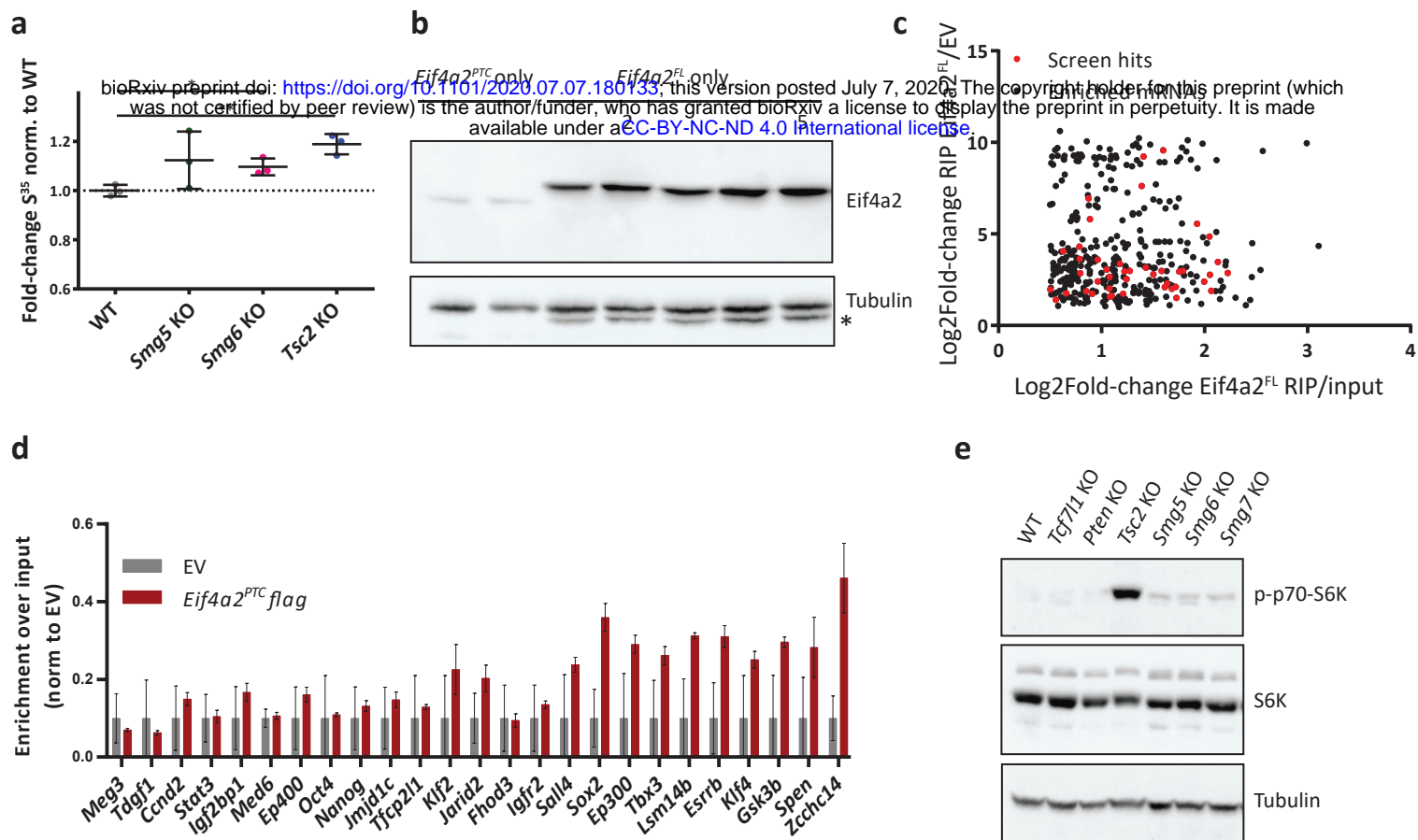


Figure 6

585 **Fig. 6 | Eif4a2-mediated differentiation delay is caused by PTC isoform-dependent regulation of**
586 **translation. a**, OPP incorporation in NMD KO, *Eif4a2* KO, *Eif4a2^{PTC}* KO, *Tsc2* KO and WT cells. Cells after 12
587 h (N12) of differentiation were pulsed for 20 s with OPP, followed by a 10 min chase. Mean and SD are
588 plotted, n=4 biological replicates. Unpaired t-test was used to calculate p-values (**** p < 0.0001, *** p
589 = 0.0001, * p < 0.05) **b**, S^{35} incorporation in *Smg5* KO, *Smg5/Eif4a2^{PTC}* dKO, *Smg5/Eif4a2* dKO relative to
590 WT. Cells in ES-DMEM-2i were pulsed with 40 μ Ci of S^{35} for 30 min before harvesting. Mean and SD are
591 plotted; n=4 biological replicates. Unpaired t-test was used to calculate p-values (**** p < 0.0001, * p \leq
592 0.05). **c**, S^{35} incorporation in *Smg6* KO, *Smg6/Eif4a2^{PTC}* dKO, *Smg6/Eif4a2* dKO relative to WT. Experiment
593 was performed as in 6b. Mean and SD are plotted, n=4 biological replicates. Unpaired t-test was used to
594 calculate p-values (** p = 0.002, * p \leq 0.05). **d**, Transcripts enriched by *Eif4a2^{FL}* RIP, detected by RT-qPCR.
595 The non-protein-coding gene *Meg3* was used as negative control. Values were normalized to empty vector
596 (EV)-RIP and to input. Mean and SD are plotted. Biological duplicates of *Eif4a2^{FL}* RIP-samples are shown
597 separately in shades of red. **e**, Significantly bound proteins (log2fold enrichment over EV \geq 1; p-adj. \leq 0.05)
598 to *Eif4a2^{FL}* and *Eif4a2^{PTC}* identified by co-IP followed by mass-spec analysis. **f**, Comparison of
599 enrichment levels of proteins significantly bound by both *Eif4a2^{FL}* and *Eif4a2^{PTC}* (75 proteins, see Figure
600 6e). **g**, Volcano plot showing proteins detected in mass-spectrometry after *Eif4a2^{FL}* immunoprecipitation.
601 Grey dots represent detected proteins, black dots represent significantly enriched or depleted proteins
602 and red dots represent proteins specifically enriched only in *Eif4a2^{FL}* co-IP (178 proteins, see Figure 6e).
603 **h**, Volcano plot showing proteins detected in mass-spectrometry after *Eif4a2^{FL}* immunoprecipitation. Grey
604 dots represent detected proteins, black dots represent significantly enriched or depleted proteins and red
605 dots represent proteins specifically enriched only in *Eif4a2^{PTC}* co-IP (231 proteins, see Figure 6e). **i**,
606 Summary model of NMD regulation indicating a proposed feedback circuit in which translation triggers
607 NMD and NMD in turn controls transcript levels of a key translation initiation factor.
608



610 **Extended Data Fig. 6 | Eif4a2-mediated differentiation delay is caused by PTC isoform-dependent**
611 **regulation of translation a**, S³⁵ incorporation levels in NMD KO, *Tsc2* KO and WT ESCs. Experiment was
612 performed independent from, but as described for Figure 6b. Mean and SD are plotted, n=2 biological
613 replicates. Unpaired t-test was used to calculate p-values (* p-value = 0.02, ** p-value = 0.002). **b**,
614 Western blot analysis for Eif4a2 in *Eif4a2* KO cells in which Eif4a2^{PTC} or Eif4a2^{FL} were re-expressed from
615 transposon vectors. Tubulin was used as a loading control. For Eif4a2^{FL}, only clones 2 and 5 were used
616 for subsequent analysis. * corresponds to Eif4a2 band due to insufficient stripping. **c**, Significantly
617 enriched transcripts in Eif4a2^{FL} RIP-Seq (log2fold change Eif4a2^{FL} RIP/input ≥ 0.5 , $p \leq 0.05$ and log2fold
618 change Eif4a2^{FL} RIP/EV RIP ≥ 1 , $p \leq 0.05$). Transcripts of genes with shown function as drivers of the exit
619 from naïve pluripotency⁶ are depicted in red. **d**, Transcripts levels of the indicated genes after Eif4a2^{PTC}
620 RIP-qPCR. The non-protein coding gene *Meg3* was used as negative control. Values were first
621 normalized to EV RIP and then to input. Mean and SD are plotted, n=2 biological replicates. **e**, Western
622 blot analysis in the indicated cells line in 2i for mTOR targets. Antibodies used are indicated in the figure.
623 Tubulin was used as loading control.

624 **Material and Methods:**

625 **Cloning**

626 gRNAs were designed using <http://crisprscan.org>. Annealed oligos (Extended Data Table 5) were cloned,
627 using a BsaI site, in a gRNA expressing vector (Addgene #41824)^{46 6}. For generating rescue cell lines, the
628 coding sequence of the gene of interest was amplified by PCR and cloned into a pCAG-3xFLAG-empty-pgk-
629 hph vector⁵ using a BamHI site. The sequence of Eif4a2^{FL} was *in vitro* synthesized (IDT) and then cloned
630 into the pCAG-3xFLAG-empty-pgk-hph vector. For the mutagenesis of Eif4a2^{PTC} the pCAG-3xFLAG-
631 Eif4a2^{PTC}-pgk-hph vector was cut using a BamHI and a SacII site. This generated an Eif4a2^{PTC} fragment that
632 was missing the first 17 nucleotides, thus generating an out of frame truncated protein product. The
633 mutated insert was blunt ended and re-cloned into the pCAG-3xFLAG-empty-pgk-hph vector cut with
634 SmaI. The correct insertion was verified for all the constructs by restriction digest and Sanger sequencing
635 with the 3xFlag_seq primer (Extended Data Table 6).

636 **Teratoma assay**

637 Paraffin-embedded teratoma tissue blocks were cut on a rotary microtome RM2255 (Leica). The 3 µm
638 sections were then stained for hematoxylin and eosin in the automated slide stainer Gemini AS (Histocom)
639 and mounted in Eukitt. Slides were scanned on a VS120 (Olympus) slide scanner.

640 **Cell culture**

641 Mouse embryonic stem cells (ESCs) containing a Rex1-GFPd2-IRES-BSD Cas9 (Rex1-GFPd2)^{4, 23} were used
642 as parental cell line for all the knockout cells generated in this study.

643 ESCs were cultured on gelatin (Sigma-Aldrich, G1890) coated plates in DMEM high-glucose (Sigma-Aldrich,
644 D5671) supplemented with 15% FBS (Gibco, 10270-106), 2 mM L-Glutamine (Sigma-Aldrich, G7513), 0.1
645 mM NEAA (Sigma-Aldrich, M7145), 1 mM Sodium Pyruvate (Sigma-Aldrich, S8636), 10 µg/ml penicillin-

646 streptomycin (Sigma-Aldrich, P4333), 55 μM β -mercaptoethanol (Fisher-Scientific, 21985-023), 2i (1.5 μM
647 CHIR99021 and 0.5 μM PD0325901) and 10 ng/ml LIF (batch tested, in-house) (ES DMEM-2i medium).

648 **Monolayer differentiation**

649 For differentiation, ESCs were plated on gelatin-coated plates at a density of 1×10^4 cells/cm² in basal
650 medium (N2B27), which is composed by 1:1 ratio of DMEM/F12 (Gibco, 21331020) and Neurobasal
651 medium (Gibco, 21103049) supplemented with 0.5x N2 (homemade), 1x B-27 Serum-Free Supplement
652 (Gibco, 17504-044), 2 mM L-Glutamine (Sigma-Aldrich, G7513), 0.1 mM NEAA (Sigma-Aldrich, M7145), 10
653 $\mu\text{g/ml}$ penicillin-streptomycin (Sigma-Aldrich, P4333), 55 μM β -mercaptoethanol (Fisher-Scientific, 21985-
654 023) and 2i (3 μM CHIR99021 and 1 μM PD0325901) (N2B27-2i). The following day 2i were withdrawn and
655 cells were differentiated for the indicated time in N2B27.

656 **Commitment assay**

657 For commitment assay, ESCs were plated on gelatin-coated plates in N2B27-2i medium at a density of 2 x
658 10^3 cells/cm². The following day 2i were withdrawn. After 48 h of differentiation medium was changed
659 and selection for Rex1-GFPd2 expressing cells was carried out by adding 5 ng/ml blasticidin ((BSD) Gibco,
660 R210-01) to the N2B27 medium. The following day medium was changed to N2B27-2i + BSD medium, and
661 medium was refreshed every two days. Alkaline-phosphatase staining was performed after four days in
662 N2B27-2i + BSD according to the manufacturer's protocol (Sigma-Aldrich, 86R). In brief, cells were fixed
663 using the Citrate-Acetone-Formaldehyde solution. Cells were then stained for 30 min at 4°C using a
664 Sodium Nitrite-FRV-Alkaline-Naphthol AS-BI Alkaline solution. Plates were then imaged using an Olympus
665 Cell-Sense microscope (OLYMPUS).

666 **EB differentiation assay**

667 For embryoid body differentiation, 1×10^5 cells were used and plated in ES DMEM without 2i and LIF as
668 hanging drops. After two days cells were collected and plated in a 10 cm petri dish, and medium was

669 changed every two days. Cells were harvested every two days and RNA was extracted using the ExtractME
670 kit (LabConsulting, EM09).

671 **Generation of knockout cell lines**

672 2×10^5 cells were transfected in a gelatin-coated 6-well plate in ES DMEM-2i using Lipofectamine 2000
673 (Fisher Scientific, 11668-027). Two pairs of gRNA were used for each gene (1 μg each) together with 0.5
674 μg of pCAG-dsRed (see full list of gRNAs in Extended Table 6). After 48 h to enrich for transfected cells,
675 dsRED/GFP double positive cells were sorted using a BD FACS Aria III. Sorted cells were plated at clonal
676 density in ES DMEM-2i. One week after sorting 48 colonies were picked (96 in case of the Smg5/Smg6 dKO
677 cells). Colonies were then trypsinized and half of the cell suspension was plated in a 96-well plate for
678 expansion. The remaining cells were lysed and DNA was extracted for PCR genotyping ⁶.

679 **PCR genotyping**

680 For DNA lysis half of a picked colony was pelleted in a PCR plate at 500g. After two PBS washes, cell pellets
681 were boiled in water at 95°C for 5 min. 3 $\mu\text{g}/\text{ml}$ of proteinase K was added after cooling down the plate
682 and incubated at 65°C for 1 h. Reaction was inactivated for 10 min at 95°C. To genotype the KO cells we
683 used a triple-primer strategy. One reverse primer specific for the deletion and one specific for the WT
684 allele were used in combination with a common forward primer (Extended Data Table 6). PCR results were
685 verified by Sanger sequencing and KO was confirmed by western-blot analysis. For genotyping PCR
686 JumpStart RedTaq PCR master mix (Sigma-Aldrich) was used following the manufacturer's protocol for
687 cycling with an annealing temperature of 55°C and 35 cycles.

688 **RNAi assay**

689 1.5×10^4 cells/cm² were transfected in a gelatin-coated 12-well plate in N2B27-2i using DharmaFECT 1
690 (Fisher Scientific, T-2001). For RNAi assays esiRNA for Eif4a2 (Sigma-Aldrich) and FlexiTube siRNAs for
691 Smg6 and Smg7 (Qiagen) were used. For siRNAs 20 ng siRNAs/4 x 10⁴ cells were used, whereas for esiRNAs

692 200 ng/6 x 10⁴ cells were used. The following day, after two PBS washes, medium was changed to N2B27.
693 Cells were harvested at the indicated time for either flow cytometry to determine Rex1-GFPd2 activity
694 (see flow cytometry) or for RNA extraction (see RNA analysis). Primers used for qPCR are annotated in
695 Extended Data Table 6.

696 **Cell cycle analysis**

697 ESCs were plated in gelatin-coated plates in ES DMEM-2i at a density of 1 × 10⁴ cells/cm². After two days
698 cells were harvested and fixed overnight in 70% ethanol. Cells were then incubated with 100 µg/ml RNase
699 A solution (Qiagen, 19101). 50 mg/liter Propidium Iodide solution (Sigma-Aldrich, P4170) was then added
700 to the cells. Cell cycle profiles were recorded on the LSRFortessa flow cytometer (BD bioscience).

701 **Flow Cytometry**

702 Cells were harvested using 0.25% trypsin/EDTA and trypsin was neutralized using ES DMEM. Rex1-GFPd2
703 was measured with LSRFortessa flow cytometer (BD bioscience). High-throughput-measurements were
704 acquired using a 96-well plate HTS unit on the LSRFortessa flow cytometer. Data analysis was performed
705 using flowJo software (BD bioscience).

706 **NMD inhibition assay**

707 ESCs were plated in gelatin-coated plates in ES DMEM-2i at a density of 2 × 10⁴ cells/cm². The following
708 day cells were treated for 8 h with either ES DMEM-2i + DMSO or ES DMEM- 2i + CHX (100 µg/ml (Sigma-
709 Aldrich, 01810)). RNA was then extracted using ExtractME kit (LabConsulting, EM09) according to the
710 manufacturer's protocol.

711 **Telomere quantification**

712 For telomere quantification genomic DNA was extracted from cells using the Puregene core Kit A (Qiagen).
713 DNA was quantified using PicoGreen Assay for dsDNA (Fisher Scientific, P11496) on a NanoDrop 3300

714 Fluorospectrometer (Fisher Scientific). PCR reactions were performed on a CFX384 Touch Real-Time PCR
715 Detection System (BioRad) using telomere specific primers and primers for the single-copy gene 36B4
716 (Extended Data Table 6). For each primer pair, a standard curve was created with known amounts of DNA
717 to determine primer efficiency. The telomere signal was normalized with the single-copy gene.

718 **RNA analysis**

719 RNA was extracted using the ExtractMe kit (LabConsulting, EM09) according to the manufacturer's
720 protocol. cDNA was retrotranscribed from 0.3 µg to 1 µg using the SensiFAST cDNA Synthesis Kit (Bioline,
721 BIO-65054). Real-time PCR was performed on the CFX384 Touch Real-time PCR Detection System (Bio-
722 Rad) using the Sensifast SYBR No Rox-Kit (Bioline, BIO-98020). Expression levels were normalized to either
723 L32 or Actin, as indicated in the figure legends (see full list of primers used for qPCR In Extended Data
724 Table 6). Results are shown as mean and standard deviation.

725 **Protein analysis**

726 Proteins were extracted using RIPA buffer (Sigma-Aldrich, 20-188) supplemented with Complete Mini
727 EDTA-free Protease inhibitor cocktail (Roche, 04693159001) and PhosSTOP (Phosphatase Inhibitor
728 Cocktail (Roche, 04906845001)). 5% milk was used for blocking. Primary antibodies were incubated
729 overnight at 4°C. Washes were performed using PBS-T (Sigma-Aldrich, P4417). Secondary antibodies were
730 incubated for 1 h at room-temperature. Primary antibodies were used at a dilution of 1:250 for anti-Smg5
731 (Abcam, ab33033, rabbit), 1:1000 for anti-Smg6 (Abcam, ab87539, rabbit), 1:2000 for anti Smg7
732 (NovusBio, NBP1-22967, rabbit), 1:1000 for anti-Phosho-Upf1 (Millipore, 07-1016, rabbit), 1:1000 for anti-
733 Upf1 (D15G6) (Cell Signaling, 12040S, rabbit), 1:1000 for anti-flag M2 (Sigma-Aldrich, F1804, mouse),
734 1:1000 for anti-c-Myc (D84C12) (Cell Signaling, 5605T, rabbit), 1:1000 for anti-Tcf7l1 (Fisher Scientific,
735 PA5-40327, rabbit), 1:1000 for anti-Eif4a2 (Abcam, ab31218, rabbit), 1:1000 for anti-Eif4a1 (Cell Signaling,
736 2490T, rabbit), 1:1000 for anti-Nanog (NovusBio, NB100-58842, rabbit), 1:1000 for anti-Esrrb (RND

737 systems, PP-H6705-00, mouse), 1:1000 for anti-p-p70-S6K (Thr389) (108D2) (Cell Signaling, 9234T, rabbit),
738 1:1000 for anti-S6K (49D7) (Cell Signaling, 2708T, rabbit), 1:5000 for anti-Tubulin (Sigma-Aldrich, T8203,
739 mouse), 1:5000 for anti-Gapdh (Sigma-Aldrich, G8795, mouse) and 1:5000 for anti-Vinculin (E1E9V) (Cell-
740 Signaling, 13901T, rabbit). Secondary antibodies were used at a dilution of 1:10000 for anti-rabbit IgG
741 (Amersham, NA934), 1:15000 for goat anti-mouse IgG (Santa Cruz, sc-2064). Chemiluminescence signal
742 from antibody binding was detected using ECL Select detection kit (GE healthcare, GERPN2235) with a
743 ChemiDoc system (BioRad).

744 **Immunoprecipitation**

745 Cells were plated in ES DMEM-2i, after two days they were harvested in IP lysis buffer (10 mM Tris Base,
746 10 mM NaCl, 2 mM EDTA, 0,5% Triton X-100) supplemented with Complete Mini EDTA-free Protease
747 inhibitor cocktail (Roche, 04693159001) and PhosSTOP (Phosphatase Inhibitor Cocktail (Roche,
748 04906845001)). 1 mg of lysate was used for the immunoprecipitation. Briefly, Dynabeads (Fisher
749 Scientific, 10004D) were coated with 5 µg of anti-flag M2 antibody (Sigma-Aldrich, F1804, mouse) for 1 h.
750 In case of RNA-free IP, lysates were treated with 50 U/ml Cyanase (Süd-Laborbedarf SLG, CY1000) and 0.5
751 ul/ml of 2 M MnSO₄ and incubated on ice for 10 min. Lysates were then cleared out by centrifugation at
752 16000 g for 15 min at 4°C, 1% of each lysate was kept as input. NaCl was added to a final concentration of
753 150 mM. Dynabeads were washed three times with wash buffer (137 mM NaCl, 20 mM Tris Base, 0,5%
754 (v/v) Tergitol-type NP-40) and one time with lysis buffer and then were incubated with the lysates for 3 h
755 at 4°C. Dynabeads were washed three times with wash buffer supplemented with Complete Mini EDTA-
756 free Protease inhibitor cocktail (Roche, 04693159001) and PhosSTOP (Phosphatase Inhibitor Cocktail
757 (Roche, 04906845001)). Samples were eluted in 2X sample buffer.

758 For the immunoprecipitation coupled with mass-spec cells were plated in ES DMEM-2i, the following day
759 they were treated with 1 µM epoxomicin (Gentaur, 607-A2606) for 3 h. Cells were harvested and lysed in

760 IP lysis buffer. 1 mg of lysate was used for the immunoprecipitation. Immunoprecipitation protocol was
761 followed (see above) with the addition of the cross-linking of the anti-flag M2 antibody to the Dynabeads
762 using dimethyl pimelimidate (DMP) (Sigma-Aldrich, D-8388). Briefly, after antibody coupling, Dynabeads
763 were washed three times with 200 mM Sodium Borate (pH=9) and then incubated for 30 min with DMP-
764 Sodium Borate solution. They were washed with the following buffers: three times with 250 mM Tris
765 (pH=8.0), two times with 100 mM glycine (pH=2), three times with TBS-T and one time with lysis buffer.
766 Beads were then incubated with the lysates for 2 h at 4°C. Five washes were performed with wash buffer
767 (137 mM NaCl, 20 mM Tris Base) and samples were submitted for mass-spec.

768 **Sample preparation for mass spectrometry analysis**

769 Beads with cross-linked antibody were transferred to new tubes and resuspended in 30 µL of 2 M urea in
770 50 mM ammonium bicarbonate (ABC). Disulfide bonds were reduced with 10 mM dithiothreitol for 30
771 min at room temperature before adding 25 mM iodoacetamide and incubating for 15 min at room
772 temperature in the dark. Remaining iodoacetamide was quenched by adding 5 mM DTT and the proteins
773 were digested with 150 ng trypsin (Trypsin Gold, Promega) at room temperature for 90 min. The
774 supernatant was transferred to a new tube, the beads were washed with another 30 µL of 2 M urea in 50
775 mM ABC and the wash combined with the supernatant. After diluting to 1 M urea with 50 mM ABC,
776 additional 150 ng trypsin were added and incubated overnight at 37°C in the dark. The digest was stopped
777 by addition of trifluoroacetic acid (TFA) to a final concentration of 0.5 %, and the peptides were desalted
778 using C18 Stagetips⁴⁷. Peptides were separated on an Ultimate 3000 RSLC nano-flow chromatography
779 system (Thermo-Fisher), using a pre-column for sample loading (Acclaim PepMap C18, 2 cm × 0.1 mm, 5
780 µm, Thermo-Fisher), and a C18 analytical column (Acclaim PepMap C18, 50 cm × 0.75 mm, 2 µm, Thermo-
781 Fisher), applying a segmented linear gradient from 2% to 35% and finally 80% solvent B (80% acetonitrile,
782 0.1% formic acid; solvent A 0.1% formic acid) at a flow rate of 230 nL/min over 120 min. Eluting peptides
783 were analyzed on a Q Exactive HF-X Orbitrap mass spectrometer (Thermo Fisher), which was coupled to

784 the column with a customized nano-spray EASY-Spray ion-source (Thermo-Fisher) using coated emitter
785 tips (New Objective).

786 **mRNA half-life measurement:**

787 ESCs were plated in N2B27-2i at a density of 20×10^4 cells/cm². The day after medium was changed to
788 N2B27-2i + 100 μ M 4SU (Carbosynth, NT06186) and they were cultured in this condition for 12 h. After a
789 PBS wash, cells were incubated with N2B27-2i medium + 10 mM Uridine (Sigma-Aldrich, U6381) for 3 h.
790 RNA extraction was carried out using Trizol (Fisher-Scientific, 10296-010) following the manufacturer's
791 instruction with the addition of 0.1 mM of DTT during the isopropanol precipitation. RNA was resuspended
792 in 1 mM DTT. 5 μ g of RNA were treated with 10 mM iodoacetamide (Sigma-Aldrich, I1149) followed by
793 ethanol precipitation²⁹. 2 ng of RNA were used for library prep. Libraries were prepared using the
794 QuantSeq 3'mRNA-seq Library Prep Kit for Illumina FWD (Lexogen, R3142) and were analyzed on a
795 HiSeqV4 SR100.

796 **RNA-Immunoprecipitation**

797 3×10^6 cells were plated in ES-DMEM. After two days cells were harvested and lysed using the MAGNA-
798 RIP kit (Millipore 17-700) according to the manufacturer's protocol. Briefly, cells were lysed by a freeze
799 and thaw cycle and lysates were stored at -80°C. Immunoprecipitations were performed at 4°C for 3 h
800 using 5 μ g of an anti-FLAG M2 antibody (Sigma-Aldrich, F1804, mouse). For RIP-Seq, libraries were
801 prepared using the QuantSeq 3'mRNA-seq Library Prep Kit for Illumina FWD (Lexogen, R3142) and
802 analyzed on a HiSeqV4 SR50. For RIP-qPCR, reverse transcription and qPCR were carried out as above.
803 Relative binding to input and empty vector control was calculated. Error bars show the standard deviation
804 between technical duplicates (Eif4a2^{FL} RIP) and between biological duplicates (Empty vector and Eif4a2^{PTC}
805 RIP).

806

807 **Translation rates measurement with S³⁵ incorporation**

808 ESCs were cultured in ES DMEM and 33 μ Ci S³⁵ (Hartmann, IS-103) was added to the culture media for 30
809 min. Cells were lysed in RIPA buffer (Sigma-Aldrich, 20-188) supplemented with Complete Mini EDTA-free
810 Protease inhibitor cocktail (Roche, 04693159001) as described above. Protein extracts were then spotted
811 on a nitrocellulose membrane. Membranes were stained with Ponceau. The membranes were wrapped
812 in saran wrap and exposed to a BAS Storage Phosphor Screen (GE Healthcare). After two days signal was
813 acquired using a Typhoon scanner (GE Healthcare). Radioactive signal was quantified using Fiji. S³⁵ signal
814 was normalized to ponceau staining. Results are shown as mean and standard deviation.

815 **Translation rates measurement with OPP incorporation**

816 ESCs were cultured in N2B27 2i medium, the following day 2i were withdrawn. After 12 h of differentiation
817 the medium was changed to N2B27 supplemented with OPP (Fisher Scientific) for a 20 s pulse. Cells were
818 then washed two times in N2B27 medium and then N2B27 medium was added for a 10 min chase. Cells
819 were then harvested. Fixation, permeabilization and Click-IT reaction were performed using the Click-iT
820 Plus OPP Alexa Fluor 647 Protein Synthesis Assay Kit (Fisher Scientific, C10458), according to the
821 manufacturer's protocol. Fluorescent signal was acquired with LSRFortessa flow cytometer (BD
822 bioscience). Median OPP-647 signal was used for quantification. Results are shown as mean and standard
823 deviation.

824

825

826

827

828

829 **Data analysis:**

830 **RNA-seq differential analysis**

831 RNA-seq samples and analysis used in this study were taken from Lackner and colleagues ⁶

832 **GO enrichment analysis**

833 GO analysis annotation were taken from the R package org.Mm.eg.db (Version 3.4.0). For the enrichment
834 of GO Terms in the different gene lists we only considered terms with 5 to 500 genes assigned to them.
835 Significance of the enrichment was determined using Fisher's exact test with all expressed genes as
836 background (Lackner et al., 2020). GO terms that did not differ in more than 5 genes were clustered and
837 one representative term for each cluster was defined (using the R base function hclust on the L1-distance
838 of the binary membership matrix). The representative term for each cluster was the term with the least
839 total annotated genes. Multiple hypothesis testing was executed using the Benjamini-Hochberg method
840 on all representative terms to calculate adjusted p-values.

841 **mRNA half-lives calculation**

842 QuantSeq data was analyzed using SLAM-DUNK (Version 0.2.4). For calculation of RNA half-lives,
843 background (no 4SU treatment) was subtracted to T>C conversion rates of transcripts with CPM ≥ 2 . Half-
844 lives were calculated by single exponential fit to a decay model ²⁹.

845 **mRNA half-lives differential analysis**

846 T>C conversion rates were used to model the half-life changes. Background (no 4SU treatment) was
847 subtracted from T>C conversion rates and then T>C conversion rates of transcripts with CPM ≥ 2 were
848 considered for the differential analysis. Differential analysis to quantify the change in T>C conversion rates
849 was calculated with a beta regression model (betareg package, version 3.1-3) of the form: T>C conversion
850 rates \sim time * genotype. In brief, firstly the T>C conversion rates after 3 h of Uridine chase were

851 normalized to the T>C conversion rates of the 4SU pulse. Secondly, the beta regression model quantified
852 the difference between T>C conversion rates of WT and NMD KO cells after 3 h of Uridine chase. p-values
853 were determined by a partial Wald test. To determine transcripts with a significant change in expression
854 and half-life we considered the 516 upregulated transcripts from Figure 3a. We could calculate a half-life
855 change for 250 out of 516 transcripts. To obtain direct targets, we postulated that upregulated transcripts
856 should have a concomitant increase in half-life (half-life change ≥ 0.2). Finally, to select NMD targets
857 relevant in ESCs out of these 136 transcripts we isolated the 57 that were upregulated in expression
858 according to the graded phenotypes.

859 **Exon conservation**

860 Pairwise whole genome alignments of mouse (GRCm38/mm10) against dog (canFam3), rat (RGSC6.0/rn6)
861 and human (GRCh38/hg38) from UCSC (<http://hgdownload.cse.ucsc.edu/downloads.html>) were used for
862 the analysis. With the Ensembl annotation of the mouse genome we extracted the respective regions of
863 the genes (introns, exons and UTRs) and calculated their sequence identities compared to the other
864 organism in the pairwise alignment as (number of matching nucleotide pairs in the
865 alignment)*100/length(alignment). As gene region annotation can be ambiguous due to variations in
866 transcript splicing, we used the following definitions. Everything that is annotated as coding sequence in
867 any protein coding transcript we count as coding exon. UTR is everything that is annotated as UTR in any
868 protein coding transcript except if it is also annotated as coding sequence in which case it is counted as
869 coding exon. Regions not covered by the above definitions are counted as introns.

870 **RIP-Seq differential analysis**

871 Quality control of the transcripts from input and RIP samples was performed using fastQC. Reads were
872 trimmed using bbduk (Version 38.57). Reads were mapped to the mm10 mouse reference genome with
873 STAR (Version 2.5.3a). Afterwards indexing was performed using samtools (Version 1.5) and reads in

874 transcripts were counted with HTSeq-count (Version 0.11.2). Transcripts that had cpm more or equal to
875 one in all the input samples were considered for the differential analysis. Differential analysis was
876 performed using DESeq2 (Version 1.24.0). To identify significantly enriched transcripts in Eif4a2^{FL} RIP we
877 considered only the 1,231 transcripts with cpm ≥ 30 in Eif4a2^{FL} RIP and either log-fold-change Eif4a2^{FL} input
878 / Eif4a2^{FL} RIP ≥ 1 and ≤ -1 (p-value ≤ 0.05) or log-fold-change Eif4a2^{FL} RIP / EV RIP ≥ 1 and ≤ -1 (p-value ≤ 0.05).
879 Out of those we considered as enriched only the transcripts that had Eif4a2^{FL} input / Eif4a2^{FL} RIP ≥ 0.5 and
880 log-fold-change Eif4a2^{FL} RIP / EV RIP ≥ 1 (362 transcripts).

881 **Mass spectrometry data acquisition and analysis**

882 The mass spectrometer was operated in data-dependent acquisition mode (DDA), survey scans were
883 obtained in a mass range of 375-1500 m/z with lock mass activated, at a resolution of 120k at 200 m/z
884 and an AGC target value of 3E6. The 8 most intense ions were selected with an isolation width of 1.6 m/z,
885 fragmented in the HCD cell at 28% collision energy and the spectra recorded for max. 250 ms at a target
886 value of 1E5 and a resolution of 30k. Peptides with a charge of +1 or $>+7$ were excluded from
887 fragmentation, the peptide match feature was set to preferred, the exclude isotope feature was enabled,
888 and selected precursors were dynamically excluded from repeated sampling for 30 s.

889 Raw data were processed using the MaxQuant software package (version 1.6.0.16, ⁴⁸) and the Uniprot
890 mouse reference proteome (January 2019, www.uniprot.org), target sequences, as well as a database of
891 most common contaminants. The search was performed with full trypsin specificity and a maximum of
892 two missed cleavages at a protein and peptide spectrum match false discovery rate of 1%.
893 Carbamidomethylation of cysteine residues were set as fixed, oxidation of methionine and N-terminal
894 acetylation as variable modifications. For label-free quantification the “match between runs” feature and
895 the LFQ function were activated - all other parameters were left at default.

896 MaxQuant output tables were further processed in R (R Core Team, 2018, <https://www.R-project.org/>).

897 Reverse database identifications, contaminant proteins, protein groups identified only by a modified

898 peptide, protein groups with less than three quantitative values in one experimental group, and protein

899 groups with less than 2 razor peptides were removed for further analysis. Due to differences in overall

900 contaminant levels between samples, LFQ values were re-normalized using the sample median of the

901 “background” protein subset (as identified in controls). Missing values were replaced by randomly

902 drawing data points from a normal distribution modeled on the whole dataset (data mean shifted by -1.8

903 standard deviations, width of distribution of 0.3 standard deviations). Differences between groups were

904 statistically evaluated using the LIMMA package ⁴⁹ at 5% FDR (Benjamini-Hochberg). The mass

905 spectrometry proteomics data have been deposited to the ProteomeXchange Consortium via the PRIDE

906 partner repository ⁵⁰ under the following accession number PXD019588. NGS data have been deposited

907 on GEO (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE153457>, reviewer token:

908 uvglmauwnfufpkp)

909 **REFERENCES**

- 910 1. Dunn, S.J., Martello, G., Yordanov, B., Emmott, S. & Smith, A.G. Defining an essential
911 transcription factor program for naive pluripotency. *Science* **344**, 1156-1160 (2014).
- 912 2. Young, R.A. Control of the embryonic stem cell state. *Cell* **144**, 940-954 (2011).
- 913 3. Boroviak, T., Loos, R., Bertone, P., Smith, A. & Nichols, J. The ability of inner-cell-mass cells to
914 self-renew as embryonic stem cells is acquired following epiblast specification. *Nature cell*
915 *biology* **16**, 516-528 (2014).
- 916 4. Wray, J. *et al.* Inhibition of glycogen synthase kinase-3 alleviates Tcf3 repression of the
917 pluripotency network and increases embryonic stem cell resistance to differentiation. *Nature*
918 *cell biology* **13**, 838-845 (2011).
- 919 5. Betschinger, J. *et al.* Exit from pluripotency is gated by intracellular redistribution of the bHLH
920 transcription factor Tfe3. *Cell* **153**, 335-347 (2013).
- 921 6. Lackner, A. *et al.* Cooperative molecular networks drive a mammalian cell state transition.
922 *bioRxiv*, 2020.2003.2023.000109 (2020).
- 923 7. Mulas, C., Kalkan, T. & Smith, A. NODAL Secures Pluripotency upon Embryonic Stem Cell
924 Progression from the Ground State. *Stem Cell Reports* **9**, 77-91 (2017).
- 925 8. Kalkan, T. *et al.* Tracking the embryonic stem cell transition from ground state pluripotency.
926 *Development (Cambridge, England)* **144**, 1221-1234 (2017).
- 927 9. Li, Q.V., Rosen, B.P. & Huangfu, D. Decoding pluripotency: Genetic screens to interrogate the
928 acquisition, maintenance, and exit of pluripotency. *WIREs Systems Biology and Medicine* **12**,
929 e1464 (2020).
- 930 10. Batista, Pedro J. *et al.* m6A RNA Modification Controls Cell Fate Transition in Mammalian
931 Embryonic Stem Cells. *Cell stem cell* **15**, 707-719 (2014).
- 932 11. Geula, S. *et al.* m⁶A mRNA methylation facilitates resolution of naïve pluripotency
933 toward differentiation. *Science* **347**, 1002-1006 (2015).
- 934 12. Leeb, M., Dietmann, S., Paramor, M., Niwa, H. & Smith, A. Genetic exploration of the exit from
935 self-renewal using haploid embryonic stem cells. *Cell Stem Cell* **14**, 385-393 (2014).
- 936 13. Li, T. *et al.* Smg6/Est1 licenses embryonic stem cell differentiation via nonsense-mediated mRNA
937 decay. *The EMBO journal* **34**, 1630-1647 (2015).
- 938 14. Lou, C.-H. *et al.* Nonsense-Mediated RNA Decay Influences Human Embryonic Stem Cell Fate.
939 *Stem cell reports* **6**, 844-857 (2016).

- 940 15. Kurosaki, T., Popp, M.W. & Maquat, L.E. Quality and quantity control of gene expression by
941 nonsense-mediated mRNA decay. *Nature Reviews Molecular Cell Biology* **20**, 406-420 (2019).
- 942 16. Mendell, J.T., Sharifi, N.A., Meyers, J.L., Martinez-Murillo, F. & Dietz, H.C. Nonsense surveillance
943 regulates expression of diverse classes of mammalian transcripts and mutes genomic noise.
944 *Nature Genetics* **36**, 1073-1078 (2004).
- 945 17. Wittmann, J., Hol, E.M. & Jäck, H.-M. hUPF2 silencing identifies physiologic substrates of
946 mammalian nonsense-mediated mRNA decay. *Mol Cell Biol* **26**, 1272-1287 (2006).
- 947 18. Tani, H. *et al.* Identification of hundreds of novel UPF1 target transcripts by direct determination
948 of whole transcriptome stability. *RNA Biology* **9**, 1370-1379 (2012).
- 949 19. Colombo, M., Karousis, E.D., Bourquin, J., Bruggmann, R. & Mühlemann, O. Transcriptome-wide
950 identification of NMD-targeted human mRNAs reveals extensive redundancy between SMG6-
951 and SMG7-mediated degradation pathways. *RNA* **23**, 189-201 (2017).
- 952 20. Reichenbach, P. *et al.* A Human Homolog of Yeast Est1 Associates with Telomerase and Uncaps
953 Chromosome Ends When Overexpressed. *Current Biology* **13**, 568-574 (2003).
- 954 21. Azzalin, C.M., Reichenbach, P., Khoriauli, L., Giulotto, E. & Lingner, J. Telomeric Repeat-
955 Containing RNA and RNA Surveillance Factors at Mammalian Chromosome Ends. *Science* **318**,
956 798 (2007).
- 957 22. Chawla, R. & Azzalin, C.M. The telomeric transcriptome and SMG proteins at the crossroads.
958 *Cytogenetic and Genome Research* **122**, 194-201 (2008).
- 959 23. Li, M. *et al.* Genome-wide CRISPR-KO Screen Uncovers mTORC1-Mediated Gsk3 Regulation in
960 Naive Pluripotency Maintenance and Dissolution. *Cell Rep* **24**, 489-502 (2018).
- 961 24. Jonas, S., Weichenrieder, O. & Izaurralde, E. An unusual arrangement of two 14-3-3-like domains
962 in the SMG5-SMG7 heterodimer is required for efficient nonsense-mediated mRNA decay.
963 *Genes Dev* **27**, 211-225 (2013).
- 964 25. Okada-Katsuhata, Y. *et al.* N- and C-terminal Upf1 phosphorylations create binding platforms for
965 SMG-6 and SMG-5:SMG-7 during NMD. *Nucleic acids research* **40**, 1251-1266 (2012).
- 966 26. Ohnishi, T. *et al.* Phosphorylation of hUPF1 Induces Formation of mRNA Surveillance Complexes
967 Containing hSMG-5 and hSMG-7. *Molecular Cell* **12**, 1187-1200 (2003).
- 968 27. Yepiskoposyan, H., Aeschmann, F., Nilsson, D., Okoniewski, M. & Mühlemann, O.
969 Autoregulation of the nonsense-mediated mRNA decay pathway in human cells. *RNA* **17**, 2108-
970 2118 (2011).

- 971 28. Huang, L. *et al.* RNA homeostasis governed by cell type-specific and branched feedback loops
972 acting on NMD. *Molecular cell* **43**, 950-961 (2011).
- 973 29. Herzog, V.A. *et al.* Thiol-linked alkylation of RNA to assess expression dynamics. *Nature Methods*
974 **14**, 1198-1204 (2017).
- 975 30. Yoder-Hill, J., Pause, A., Sonenberg, N. & Merrick, W.C. The p46 subunit of eukaryotic initiation
976 factor (eIF)-4F exchanges with eIF-4A. *Journal of Biological Chemistry* **268**, 5566-5573 (1993).
- 977 31. Li, W., Belsham, G.J. & Proud, C.G. Eukaryotic Initiation Factors 4A (eIF4A) and 4G (eIF4G)
978 Mutually Interact in a 1:1 Ratio in Vivo. *Journal of Biological Chemistry* **276**, 29111-29115 (2001).
- 979 32. Grabole, N. *et al.* Genomic analysis of the molecular neuropathology of tuberous sclerosis using
980 a human stem cell model. *Genome medicine* **8**, 94 (2016).
- 981 33. Keene, J.D., Komisarow, J.M. & Friedersdorf, M.B. RIP-Chip: the isolation and identification of
982 mRNAs, microRNAs and protein components of ribonucleoprotein complexes from cell extracts.
983 *Nature Protocols* **1**, 302-307 (2006).
- 984 34. Meng, L. *et al.* Epoxomicin, a potent and selective proteasome inhibitor, exhibits in
985 vivo antiinflammatory activity. *Proceedings of the National Academy of Sciences* **96**,
986 10403 (1999).
- 987 35. Schweingruber, C., Soffientini, P., Ruepp, M.D., Bachi, A. & Mühlemann, O. Identification of
988 Interactions in the NMD Complex Using Proximity-Dependent Biotinylation (BioID). *PLoS One* **11**,
989 e0150239 (2016).
- 990 36. Jang, H. *et al.* O-GlcNAc Regulates Pluripotency and Reprogramming by Directly
991 Acting on Core Components of the Pluripotency Network. *Cell stem cell* **11**, 62-74 (2012).
- 992 37. Bornelöv, S., Selmi, T., Flad, S., Dietmann, S. & Frye, M. Codon usage optimization in pluripotent
993 embryonic stem cells. *Genome Biology* **20**, 119 (2019).
- 994 38. Di Stefano, B. *et al.* The RNA Helicase DDX6 Controls Cellular Plasticity by Modulating P-Body
995 Homeostasis. *Cell stem cell* **25**, 622-638.e613 (2019).
- 996 39. Bulut-Karslioglu, A. *et al.* The Transcriptionally Permissive Chromatin State of Embryonic Stem
997 Cells Is Acutely Tuned to Translational Output. *Cell stem cell* **22**, 369-383.e368 (2018).
- 998 40. Corsini, N.S. *et al.* Coordinated Control of mRNA and rRNA Processing Controls Embryonic Stem
999 Cell Pluripotency and Differentiation. *Cell stem cell* **22**, 543-558.e512 (2018).
- 1000 41. You, K.T., Park, J. & Kim, V.N. Role of the small subunit processome in the maintenance of
1001 pluripotent stem cells. *Genes & development* **29**, 2004-2009 (2015).

- 1002 42. Shen, R., Weng, C., Yu, J. & Xie, T. eIF4A controls germline stem cell self-renewal by directly
1003 inhibiting BAM function in the Drosophila ovary. *Proc Natl Acad Sci U S A* **106**, 11623-11628
1004 (2009).
- 1005 43. Tuck, A.C. *et al.* Mammalian RNA Decay Pathways Are Highly Specialized and Widely Linked to
1006 Translation. *Molecular Cell* (2020).
- 1007 44. Holmes, Z.E. *et al.* The Sox2 transcription factor binds RNA. *Nature Communications* **11**, 1805
1008 (2020).
- 1009 45. Gao, Z. *et al.* Determination of protein interactome of transcription factor Sox2 in embryonic
1010 stem cells engineered for inducible expression of four reprogramming factors. *The Journal of*
1011 *biological chemistry* **287**, 11384-11397 (2012).
- 1012 46. Mali, P. *et al.* RNA-guided human genome engineering via Cas9. *Science* **339**, 823-826 (2013).
- 1013 47. Rappsilber, J., Mann, M. & Ishihama, Y. Protocol for micro-purification, enrichment, pre-
1014 fractionation and storage of peptides for proteomics using StageTips. *Nature Protocols* **2**, 1896-
1015 1906 (2007).
- 1016 48. Tyanova, S., Temu, T. & Cox, J. The MaxQuant computational platform for mass spectrometry-
1017 based shotgun proteomics. *Nature Protocols* **11**, 2301-2319 (2016).
- 1018 49. Ritchie, M.E. *et al.* limma powers differential expression analyses for RNA-sequencing and
1019 microarray studies. *Nucleic Acids Research* **43**, e47-e47 (2015).
- 1020 50. Perez-Riverol, Y. *et al.* The PRIDE database and related tools and resources in 2019: improving
1021 support for quantification data. *Nucleic Acids Research* **47**, D442-D450 (2018).

1022