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Favipiravir and severe acute respiratory syndrome coronavirus 2 in hamster model

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Summary

There is a need for safe and effective antiviral molecules with which to combat COVID-19 pandemics. Recently, *in vitro* inhibitory activity of favipiravir against SARS-CoV-2 was reported. Here, we used a Syrian hamster model to explore the pharmacokinetics of this molecule and its *in vivo* efficacy against SARS-CoV-2. Results revealed that high doses (700-1400mg/kg/day) significantly reduced virus replication in the lungs accompanied by clinical alleviation of the disease. However, these high doses were associated with significant toxicity in hamsters. Favipiravir pharmacokinetics displayed non-linear increase in plasma exposure between the doses and good lung penetration. Analysis of viral genomes *in vivo* showed that favipiravir induced a mutagenic effect. Whilst the plasma trough concentrations observed in this study were comparable with those previously found during human clinical trials, this potential toxicity requires further investigation to assess whether a tolerable dosing regimen can be found in humans that effectively reduces virus replication.

Keywords

COVID-19, SARS-CoV-2, antiviral therapy, favipiravir, animal model, preclinical research

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Introduction

In March 2020, the World Health Organization declared coronavirus disease 2019 (COVID-19) a pandemic (WHO, 2020). The COVID-19 outbreak was originally identified in Wuhan, China, in December 2019 and spread rapidly around the world within a few months. The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), the causative agent of COVID-19, belongs to the *Coronaviridae* family and is closely related to the SARS-CoV which emerged in China in 2002 (Zhu et al., 2020). After an incubation period of about 5 days, disease onset usually begins with an influenza-like syndrome associated with high virus replication in respiratory tracts (Huang et al., 2020, He et al., 2020). In some patients, a late acute respiratory distress syndrome, associated with high levels of inflammatory proteins, occurs within one to two weeks (Huang et al., 2020). As of 7 July 2020, more than 11.6 million cases of COVID-19 have resulted in more than 538,000 deaths (Dong et al., 2020). In the face of this ongoing pandemic and its unprecedented repercussions, not only on human health but also on society, ecology and economy, there is an urgent need for effective infection prevention and control measures.

Whilst host-directed and immune-based therapies could prove useful for the clinical management of critically ill patients, the availability of safe and effective antiviral molecules would represent an important step towards fighting the current pandemic. As conventional drug development is a slow process, repurposing of drugs already approved for any indication was extensively explored and led to the implementation of many clinical trials for the treatment of COVID-19 (Mercorelli et al., 2018). However, the development of effective antiviral drugs for the treatment of COVID-19, should, as much as possible, rely on robust pre-clinical *in vivo* data, not only on efficacy generated *in vitro*. Accordingly, rapid implementation of rodent and non-human primate animal models should help to assess more finely the potential safety and efficacy of drug candidates and to determine appropriated dose regimens in humans (Chan et al., 2020, Rockx et al., 2020).

Favipiravir (6-fluoro-3-hydroxypyrazine-2-carboxamine) is an anti-influenza drug approved in Japan that has shown broad-spectrum antiviral activity against a variety of other RNA viruses (Guedj et al., 2018, Yamada et al., 2019, Segura Guerrero et al., 2018, Tani et al., 2018, Jochmans et al., 2016, Takahashi et al., 2003, Rosenke et al., 2018). Favipiravir is a prodrug that is metabolized intracellularly into its active ribonucleoside 5'-triphosphate form that acts as a nucleotide analogue to selectively inhibit RNA-dependent RNA polymerase and induce lethal mutagenesis (Baranovich et al., 2013, Sangawa et al., 2013). Recently, several studies reported *in vitro* inhibitory activity of favipiravir against SARS-CoV-2 with 50% effective concentrations (EC_{50}) ranging from 62 to >500µM (10 to >78µg/mL) (Wang et al., 2020, Jeon et al., 2020, Shannon et al., 2020). Based on these results, more than 20 clinical trials on the management of COVID-19 by favipiravir are in progress (https://clinicaltrials.gov/). In the

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present study, a Syrian hamster model (*Mesocricetus auratus*) was implemented to explore the *in vivo* safety and efficacy and the pharmacokinetics (PK) of several dosing regimens of favipiravir.

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Results

In vitro efficacy of favipiravir

Using VeroE6 cells and an antiviral assay based on reduction of cytopathic effect (CPE), we recorded EC_{50} and EC_{90} of 32 and 52.5 µg/mL using a multiplicity of infection (MOI) of 0.001, 70.0 and >78.5µg/mL with an MOI of 0.01 (Figure S1) in accordance with previous studies (Wang et al., 2020, Jeon et al., 2020, Shannon et al., 2020). Infectious titer reductions (fold change in comparison with untreated cells) were ≥2 with 19.6µg/mL of favipiravir and ranged between 11 and 342 with 78.5µg/mL. Using CaCo2 cells, which do not exhibit CPE with SARS-CoV-2 BavPat1 strain, infectious titer reductions were around 5 with 19.6µg/mL of favipiravir and ranged between 144 and 7721 with 78.5µg/mL of the drug. 50% cytotoxic concentrations (CC_{50}) in VeroE6 and CaCo2 cells were >78.5µg/mL.

Infection of Syrian hamsters with SARS-CoV-2

Following Chan *et al.*, we implemented a hamster model to study the efficacy of antiviral compounds (Chan et al., 2020). Firstly, we intranasally infected four-week-old female Syrian hamsters with 10^6 $TCID_{50}$ of virus. Groups of two animals were sacrificed 2, 3, 4 and 7 days post-infection (dpi). Viral replication was quantified in sacrificed animals by RT-qPCR in organs (lungs, brain, liver, small/large bowel, kidney, spleen and heart) and plasma. Viral loads in lungs peaked at 2 dpi, remained elevated until 4 dpi and dramatically decreased at 7 dpi (Figure 1a). Viral loads in plasma peaked at 3 dpi and viral replication was detected in the large bowel at 2 dpi (Figure 1b and Table S1). No viral RNA was detected in almost all the other samples tested (Table S1). Subsequently, we infected animals with two lower doses of virus (10^5 and 10^4 $TCID_{50}$). Viral RNA was quantified in lungs, large bowel and plasma from sacrificed animals 2, 3, 4 and 7 dpi (Figure 1a and 1b). Viral loads in lungs peaked at 2 and 3 dpi with doses of 10^5 and 10^4 $TCID_{50}$ respectively. Maximum viral loads in lungs of animals infected with each dose of virus were comparable. Viral RNA yields in plasma and large bowel followed a similar trend but with more variability, with this two lower doses. In addition, clinical monitoring of animals showed no marked symptoms of infection but significant weight losses from 3 dpi when compared to animals intranasally inoculated with sodium chloride 0.9% (Figure 1c).

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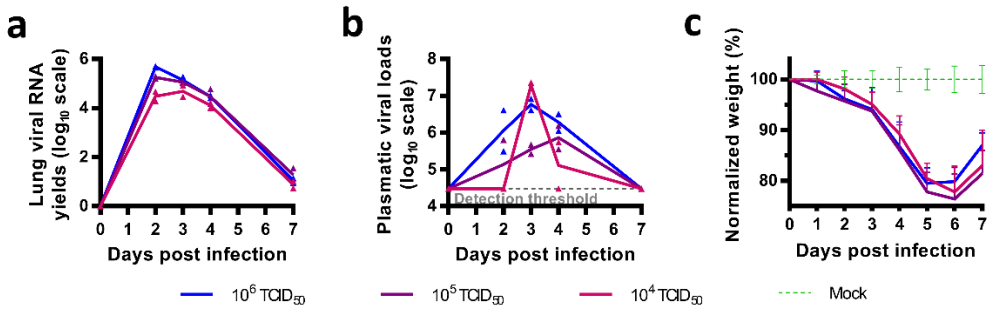

Figure 1: Implementation of hamster model

Hamsters were intranasally infected with 10^6, 10^5 or 10^4 $TCID_{50}$ of virus. Viral replication was quantified using an RT-qPCR assay. **a** Lung viral RNA yields. **b** Plasmatic viral loads. **c** Clinical course of the disease. Normalized weight at day *n* was calculated as follows: (% of initial weight of the animal at day *n*)/(mean % of initial weight for mock-infected animals at day *n*). Data represent mean ±SD (details in Table S1).

In vivo efficacy of favipiravir

To assess the efficacy of favipiravir, hamsters received the drug, intraperitoneally, three times a day (TID). We used three doses of favipiravir: 18.75, 37.5 and 75mg/day (corresponding to 340±36, 670±42 and 1390±126 mg/kg/day respectively).

In a first set of experiments, treatment was initiated at day of infection (preemptive antiviral therapy) and ended at 2 dpi. We infected groups of 6 animals intranasally with three doses of virus (10^6, 10^5 and 10^4 $TCID_{50}$) and viral replication was measured in lungs and plasma at 3 dpi (Figure 2a). When analysis of virus replication in clarified lung homogenates was based on infectious titers (as measured using $TCID_{50}$ assay), an inverse relationship was observed between infectious titers and the dose of favipiravir (Figure 2b). This trend was even more important when low doses of virus were used to infect animals. At each dose of virus, mean infectious titers for groups of animals treated with 75mg/day TID were significantly lower than those observed with untreated groups ($p \leq 0.0001$): reduction of infectious titers ranged between 1.9 and 3.7 log_{10}. For animals infected with 10^5 or 10^4 $TCID_{50}$, significant infectious titer reductions of around 0.8 log_{10} were also observed with the dose of 37,5mg/day TID ($p \leq 0.038$). Drug 90% and 99% effective doses (ED_{90} and ED_{99}) were estimated based on these results and ranged between 31-42mg/day and 53-70mg/day respectively (Table 2). When analysis of virus replication in clarified lung homogenates were assessed on viral RNA yields (as measured using quantitative real time RT-PCR assay), significant differences with groups of untreated animals, ranging between 0.7 and 2.5 log_{10}, were observed only with the higher dose of favipiravir ($p \leq 0.012$). Once again, this difference was more noticeable with lower doses of virus (Figure 2b). Since we found higher reductions of infectious titers than those observed with viral RNA yields, we estimated the relative infectivity of viral particle (*i.e.* the ratio of the number of infectious particles over the number of viral RNA molecules). Decreased infectivity was observed in all treated groups of animals. These differences

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were always significant with the higher dose of favipiravir ($p \leq 0.031$) and were significant with the dose of 37.5mg/day TID for animals infected with 10^5 or 10^4 $TCID_{50}$ of virus ($p \leq 0.041$). We then measured plasmatic viral loads using quantitative real time RT-PCR assay and found, with the higher dose of favipiravir and the groups of animals infected with 10^6 or 10^4 $TCID_{50}$, significant reductions of 2.1 and 2.62 log_{10}, respectively ($p \leq 0.022$) (Figure 2b).

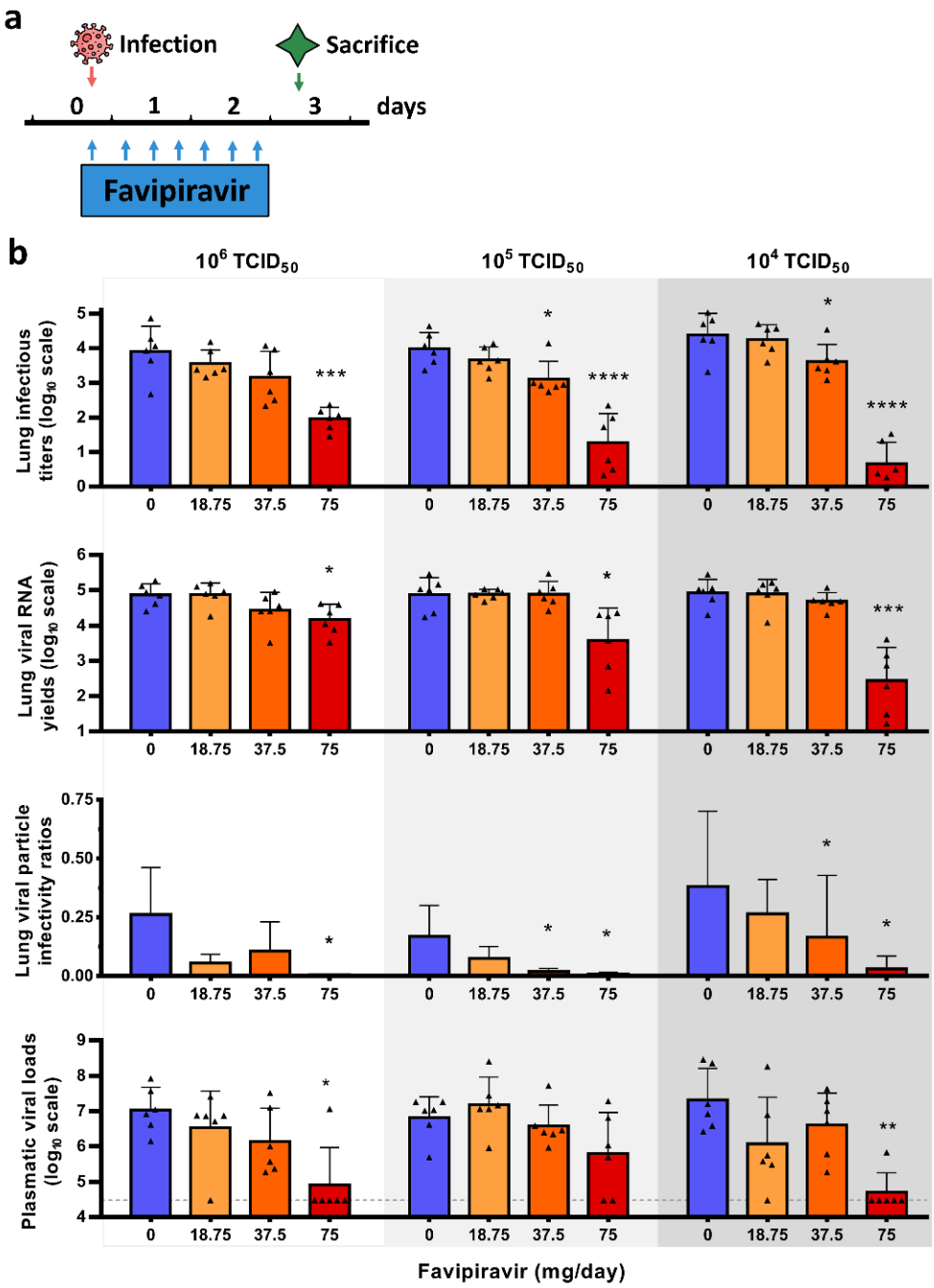

Figure 2: Virological results with preemptive favipiravir therapy

a Experimental timeline. **b** Viral replication in lungs and plasma. Hamsters were intranasally infected with 10^6, 10^5 or 10^4 $TCID_{50}$ of virus. Lung infectious titers (measured using a $TCID_{50}$ assay) and viral RNA yields were (measured using an RT-qPCR assay) expressed in $TCID_{50}$/copy of ɣ-actine gene and viral genome copies/copy of

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ɣ-actine gene respectively. Relative lung viral particle infectivities were calculated as follows: ratio of lung infectious titer over viral RNA yields. Plasmatic viral loads (measured using an RT-qPCR assay) are expressed in viral genome copies/mL of plasma (the dotted line indicates the detection threshold of the assay). Data represent mean ±SD. ****, ***, ** and * symbols indicate that the average value for the group is significantly lower than that of the untreated group with a p-value <0.0001, ranging between 0.0001-0.001, 0.001-0.01 and 0.01-0.05 respectively (details in Table S2 and S3).

Table 2: 50%, 90% and 99% drug effective doses

	ED_{50} mg/day (95%CI[1])	ED_{90} mg/day (95%CI[1])	ED_{99} mg/day (95%CI[1])
Preemptive therapy			
10^4 $TCID_{50}$	34 (30-37)	42 (38-46)	53 (48-58)
10^5 $TCID_{50}$	26 (21-30)	37 (31-44)	56 (46-65)
10^6 $TCID_{50}$	15 (10-20)	31 (21-41)	70 (48-93)
Preventive therapy			
10^4 $TCID_{50}$	27 (25-29)	35 (32-38)	47 (44-51)

[1]: 95% confidence interval

Dose-response curves are presented in Figure S2.

In a second set of experiments, we assessed, over a period of 7 days, the impact of treatment on the clinical course of the disease using weight loss as the primary criterion (Figure 3a). Beforehand, we evaluated the toxicity of the three doses of favipiravir with groups of four non-infected animals treated from day 0 to day 3 (Figure 3b). High toxicity was observed with the dose of 75mg/day TID with significant weight loss noticed from the first day of treatment (Table S4). We also found a constant, but moderate, toxicity with the dose of 37.5mg/day TID that was significant at day 4 and 5 only. No toxicity was detected with the lower dose of favipiravir. To assess if the toxicity observed with the highest dose of favipiravir was exacerbated by the infection, we compared weight losses of infected and non-infected animals treated with the dose of 75mg/day TID. Regardless of the dose of virus, no significant difference was observed at 1, 2 and 3 dpi (Figure S3). After this evaluation of favipiravir toxicity, we intranasally infected groups of 10 animals with two doses of virus (10^5 or 10^4 $TCID_{50}$). Treatment with a dose of 37.5mg/day TID was initiated on the day of infection (preemptive antiviral therapy) and ended at 3 dpi (Figure 3a). With both doses of virus, treatment was associated with clinical alleviation of the disease (Figure 3c-d). With the dose of 10^5 $TCID_{50}$, mean weights of treated animals were significantly higher than those of untreated animals at 5 and 6 dpi ($p \leq 0.031$). Similar observations were made with the dose of 10^4 $TCID_{50}$ at 5, 6 and 7 dpi ($p<0.0001$).

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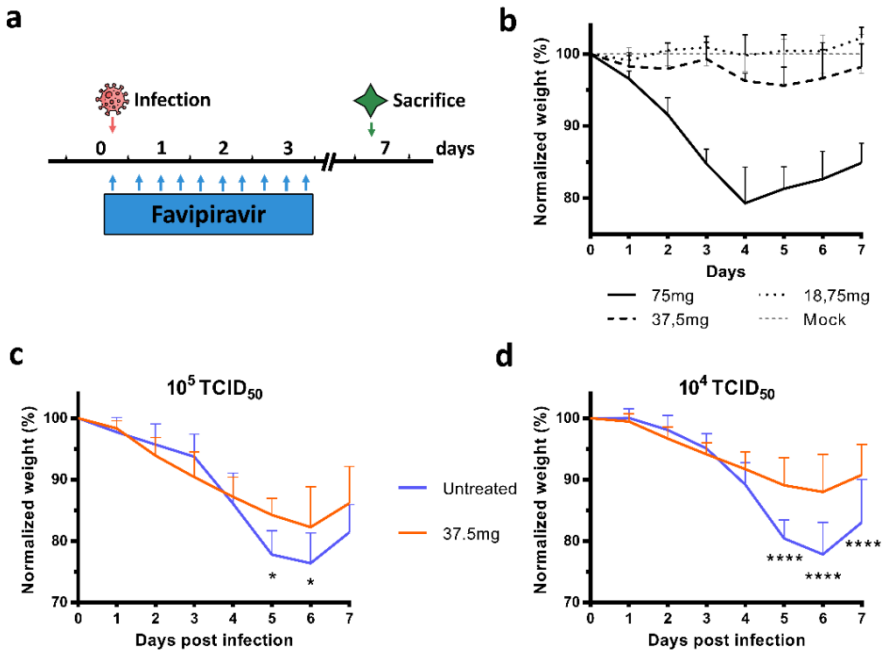

Figure 3: Clinical follow-up with preemptive favipiravir therapy

a Experimental timeline. **b** Evaluation of the toxicity of the three doses of favipiravir (mg/day TID) with uninfected animals following an identical experimental timeline without infection. **c-d** Clinical follow-up with animals infected respectively with 10^5 and 10^4 TCID$_{50}$ of virus and treated with a dose of favipiravir of 37.5mg/day TID. Normalized weight at day *n* was calculated as follows: (% of initial weight of the animal at day *n*)/(mean % of initial weight for mock-infected animals at day *n*). Data represent mean ±SD. **** and * symbols indicate a significant difference between treated and untreated animals with a p-value <0.0001 and ranging between 0.01-0.05 respectively (details in Table S2 and S4).

In a third set of experiments, treatment was started one day before infection (preventive antiviral therapy) and ended at 2 dpi. We intranasally infected groups of 6 animals with 10^4 TCID$_{50}$ of virus and viral replication was measured in lungs and plasma at 3 dpi (Figure 4a). Once again, an inverse relationship was observed between lung infectious titers and the dose of favipiravir (Figure 4b). Mean infectious titers for groups of animals treated with 37.5 and 75mg/day TID were significantly lower than those observed with untreated groups ($p \leq 0.002$). Of note, undetectable infectious titers were found for all animals treated with the higher dose. Estimated ED_{90} and ED_{99} were 35 and 47mg/day respectively (Table 2). Significant reductions of viral RNA yields of 0.9 and 3.3 log_{10}, were observed with animals treated with 37.5 and 75mg/day TID respectively ($p \leq 0.023$). Resulting infectivity of viral particle was decreased, with a significant reduction only for the higher dose of favipiravir ($p=0.005$). Finally, we found significantly reduced plasmatic viral loads with animals treated with 37.5 and 75mg/day TID ($p \leq 0.005$).

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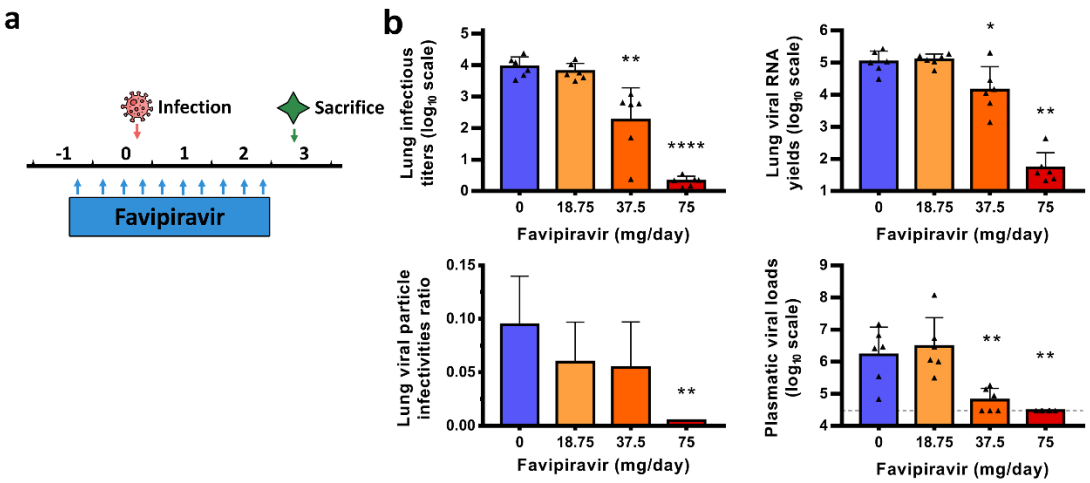

Figure 4: Virological results with preventive favipiravir therapy

a Experimental timeline. **b** Viral replication in lungs and plasma. Hamsters were intranasally infected with 10^4 $TCID_{50}$ of virus. Lung infectious titers (measured using a $TCID_{50}$ assay) and viral RNA yields awee (measured using an RT-qPCR assay). They are expressed in $TCID_{50}$/copy of ɣ-actine gene and viral genome copies/copy of ɣ-actine gene respectively. Relative lung virus infectivities were calculated as follows: ratio of lung infectious titer over viral RNA yields. Plasmatic viral loads (measured using an RT-qPCR assay) are expressed in viral genome copies/mL of plasma (the dotted line indicates the detection threshold of the assay). Data represent mean ±SD. ****, ** and * symbols indicate that the average value for the group is significantly different from that of the untreated group with a p-value <0.0001, ranging between 0.001-0.01 and 0.01-0.05 respectively (details in Table S2 and S3).

Favipiravir pharmacokinetics (PK) in a hamster model

We first assessed the PK and lung distribution of favipiravir in a subgroup of uninfected animals. Groups of animals were treated respectively with a single dose of favipiravir administrated intraperitoneally: 6.25mg, 12.5 mg and 25 mg. In each dose group, we sacrificed 3 animals at specific time points post-treatment (0.5, 1, 5 or 8 hours) for determination of favipiravir in plasma. Drug concentration in lung tissue was determined at 0.5 and 5 hours post-treatment. Subsequently, we assessed the favipiravir concentration after multiple dose in animals intranasally infected with 10^5 $TCID_{50}$ of virus. Groups of 9 animals received the three doses evaluated for 3 days (Figure 2a): 18.75mg/day, 37.5mg/day and 75mg/day TID and were sacrificed at 12-hours after the last treatment dose. Favipiravir was quantified in plasma (n=9) and lung tissue (n=3).

Results are presented in Table 3 and Figure S4. The single dose PK analysis showed that the maximum concentration of favipiravir was observed at 0.5 hour at all doses, then plasma drug concentrations decreased exponentially to reach concentrations below 10 µg/ml at 12 hours. Favipiravir PK exhibited a non-linear increase in concentration between the doses. After multiple doses, trough concentrations (12 hours) of favipiravir also exhibited a non-linear increase between doses. The extrapolated 12 hours post-treatment concentrations after a single dose were calculated in order to determine the

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accumulation ratio. Accumulation ratios were respectively 6, 16 and 21 at the 3 doses, confirming the non-proportional increase between doses. The average concentration after single dose administration over 0 to 12-hour intervals was calculated and the respective values obtained were 10.1 µg/mL, 38.7 µg/mL and 100.5 µg/mL for the 3 favipiravir doses.

Favipiravir lung concentrations were 1.6 to 2.7-fold lower than in plasma for both administration of single and multiple doses. After a single dose, the mean lung to plasma ratio ranged from 0.37 to 0.62 according to the time post-treatment and was similar between the 3 doses of favipiravir at 0.5 hours. A high ratio 5 hours post-treatment was observed at the highest dose (25 mg) with an increase by a factor 1.6 to 1.8 compared with the lower doses. After multiple doses, the lung penetration of favipiravir was confirmed with a mean lung to plasma ratio ranging from 0.35 to 0.44. Favipiravir was not detected in the lungs at the lowest dose (18.75 mg/day).

Table 3: Plasma and lung concentrations of favipiravir after administration of a single dose or multiple dose of favipiravir

	Single Dose			Multiple Dose[1] (Day 3)		
	Plasma (µg/mL)	Lung (µg/g)	L/p ratio	Plasma (µg/mL)	Lung (µg/g)	L/p ratio
Dose: 25 mg						
0.5 hr	372 ± 47.5	216 ± 39	0.58 ± 0,04			
1 hr	279 ± 49.9					
5 hr	135 ± 49.0	81,3 ± 24	0.62 ± 0,10			
8 hr	5.77 ± 1.34					
12 hr	*1.43*[2]			29.9 ± 9.83	16.0 ± 4.87	0.44 ± 0,07
Dose: 12.5 mg						
0.5 hr	166 ± 52.0	90.7 ± 12.7	0.58 ± 0.14			
1 hr	155 ± 20.6					
5 hr	10.7 ± 5.16	3.84 ± 1.49	0.37 ± 0.052			
8 hr	1.94 ± 0.06					
12 hr	*0.16*[2]			2.57 ± 1.22	1.36 ± 0.14	0.35 ± 0,03
Dose: 6.25 mg						
0.5 hr	86.3 ± 4.11	50.2 ± 16.4	0.58 ± 0.17			
1 hr	35.2 ± 27.8					
5 hr	2.90 ± 0.25	1.09 ± 0.05	0.38 ± 0.05			
8 hr	0.56 ± 0.16					
12 hr	*0.05*[2]			0.31 ± 0.14	not detected	*n.a.*

Data represent mean ±SD; Three animals for each condition except at multiple dose (n=9 for plasma; n=3 for lung); details in Table S5

[1]: PK realized after 3 days of favipiravir administered three times a day (18.75, 37.5 or 75mg/day TID)

[2]: extrapolated C_{12h}. *na*: not applicable

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Mutagenic effect of favipiravir

To understand which genomic modifications accompanied favipiravir treatment, direct complete genome sequencing of clarified lung homogenates from animals intranasally infected with 10^6 $TCID_{50}$ of virus and treated with the two highest doses of drug (preemptive antiviral therapy; Figure 2) was performed. Data were generated by next generation sequencing from lung samples of four animals per group (untreated, 37.5mg/day TID and 75mg/day TID). The mean sequencing coverage for each sample ranged from 10,991 to 37,991 reads per genomic position and we subjected substitutions with a frequency ≥1% to further analysis. The genetic variability in virus stock was also analyzed: 14 nucleotide polymorphisms were detected of which 5 recorded a mutation frequency higher than 10% (Table S6).

In order to study the mutagenic effect of favipiravir, we used the consensus sequence from virus stock as reference and all the mutations simultaneously detected in a lung sample and in virus stock were not considered in the further analysis (1 to 4 mutations per sample, see Table S6). Overall, no majority mutations were detected (mutation frequency >50%), mutations were distributed throughout the whole genome and almost all of them exhibited a frequency lower than 10% (Figure 5a and 5b).

Results revealed a relationship between the number of mutations detected per sample and the dose of favipiravir (Figure 5c): the mean number of mutations increased by a factor 2 and 4.8 with groups of animals treated with 37.5 and 75mg/day TID, respectively. The difference is significant only with a dose of 37.5mg/day TID (p=0.029). This increase of the number of mutations is mainly the consequence of the occurrence of a large number of G→A substitutions and, to a lesser extent, C→U substitutions. Consequently, regardless of the dose of favipiravir, mean frequency of G→A substitutions was significantly increased by a factor of 4.2 ($p \leq 0.009$). This rise of these transition mutations led to increased frequency of all transition mutations (significant only at dose of 37.5mg/day TID; p=0.037) and increased frequency of non-synonymous mutations (significant only at dose of 75mg/day TID; p=0.009) (Figure 5d). We investigated whether or not effectiveness in treated animals was linked with the characteristics of the mutations detected on viral populations and found that frequency of non-synonymous, synonymous and G→A mutations were associated with infectious titers in lungs ($p<0.03$; Figure 5e). Finally, our experiments revealed some parallel evolution events; 32 substitutions in viral sub-populations were detected in two independent animals. Notably, 18 of these shared mutations were detected only with treated animals, 14 of them being non-synonymous (Table S8). These mutations are located in nsp2, 3, 4, 5, 6, 14, N protein, Matrix, ORF 3a and 8. At this stage, one cannot conclude if these substitutions reflect the adaptation to the hamster model or are the result of the antiviral selection.

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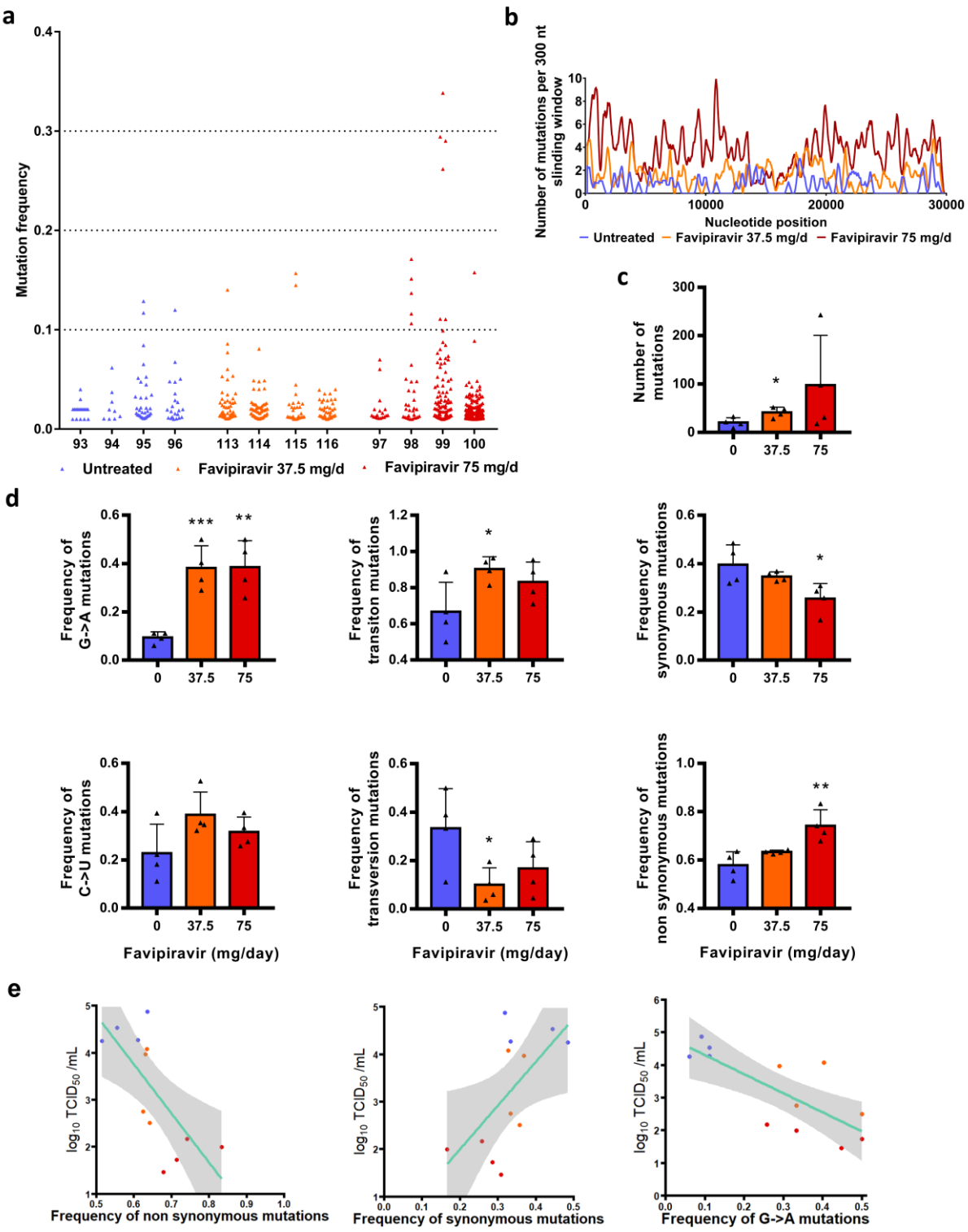

Figure 5: Mutagenic effect of favipiravir

a Viral genetic diversity in clarified lung homogenates. For each condition, four samples were analyzed. Each triangle represents a mutation (only substitutions with a frequency ≥1% were considered). **b** Patterns of mutation distribution on complete viral genome. Each variable nucleotide position was counted only once when found. The variability was represented using 75 nt sliding windows. For each condition, variable nucleotide positions were determined and represented using a 300 nt sliding window. **c** Mean number of mutations. Data represent

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mean ±SD. **d** Mutation characteristics. For each sample, the frequency of a given mutation was calculated as follows: number of this kind of mutation detected in the sample divided by the total number of mutations detected in this sample. Data represent mean ±SD. ** and * symbols indicate that the average value for the group is significantly different from that of the untreated group with a p-value ranging between 0.001-0.01 and 0.01-0.05 respectively (details in details in Table S6 and S7). **e** Association between lung infectious titers (measured using a $TCID_{50}$ assay) and frequency of non synonymous, synonymous and G→A mutations. Each dot represent data from a given animal.

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Discussion

In the current study, we used a hamster model to assess efficacy of the favipiravir against SARS-CoV-2. Following infection, viral RNA was mainly detected in lungs, blood, and, to a lesser extent, in the large bowel. Peak of viral replication was observed at 2-3 dpi followed by observation of significant weight losses, in line with recently reported investigations that involved 6-10 weeks old hamsters (Kaptein et al., 2020, Chan et al., 2020). Clinically, the main symptom was weight loss, observed from the first day of infection and followed by recovery at 6dpi. This confirmed that the *in vivo* model, with younger animals (4 weeks-old), is suitable for preclinical evaluation of antiviral compounds against SARS-CoV-2.

Using a preemptive strategy, we demonstrated that doses of favipiravir of around 700-1400mg/kg/day TID reduced viral replication in lungs of infected animals and allowed clinical alleviation of the disease. Reduction of viral replication was greater when estimated on the basis of infectious titers than on total viral RNA as previously observed in non-human primates treated with Remdesivir (Williamson et al., 2020). However, the effective doses of favipiravir were higher than those usually used in rodent models (≈100-400mg/kg/day) (Sidwell et al., 2007, Smither et al., 2014, Julander et al., 2009, Tani et al., 2018, Oestereich et al., 2016, Yamada et al., 2019). This can be correlated with the high favipiravir EC_{50} found *in vitro* for SARS-CoV-2. Moreover, effective doses were associated with significant toxicity in our hamster model. This observed toxicity reflected only the adverse effects of favipiravir and was not exacerbated during SARS-CoV-2 infection. Indeed, similar weight losses were measured among infected and non-infected animals treated with the highest dose of favipiravir at 1, 2 and 3dpi.

In the present study, reduction of viral replication was correlated with the dose of favipiravir administrated and inversely correlated with the dose of virus inoculated. In a recent study, favipiravir administrated *per os* twice daily (loading dose of 600mg/kg/day followed by 300mg/kg/day) revealed a mild reduction of lung viral RNA yields using a similar hamster model with high doses of virus ($2x10^6$ $TCID_{50}$) (Kaptein et al., 2020). These results are in accordance with ours at the lower dose of favipiravir (around 340mg/kg/day TID).

By characterizing the dose response curve, we estimated that the dose required to reduce by 90% (ED_{90}) the level of infectious titers in lungs is in the range of 570-780mg/kg/day. In the most favourable situation, where high doses were used as a preemptive therapy, favipiravir led to undetectable viral replication in lung and plasma. These results showed that the use of high doses of favipiravir could expand its *in vivo* spectrum against RNA viruses.

With influenza viruses, favipiravir acts as a nucleotide analogue. It is metabolized intracellularly to its active form and incorporated into nascent viral RNA strands. This inhibits RNA strand extension and

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294 induces abnormal levels of mutation accumulation into the viral genome (Baranovich et al., 2013,
295 Sangawa et al., 2013). Recently, it was shown *in vitro* that favipiravir has a similar mechanism of action
296 with SARS-CoV-2 through a combination of chain termination, reduced RNA synthesis and lethal
297 mutagenesis (Shannon et al., 2020). Our genomic analysis confirmed the mutagenic effect of favipiravir
298 *in vivo*. Indeed, we found that favipiravir treatment induced appearance of a large number of G→A
299 and C→U mutations into viral genomes. This was associated to a decrease of viral infectivity probably
300 because alteration of the genomic RNA disturb the replication capacity. Similar findings were described
301 *in vitro* and *in vivo* with other RNA viruses (Baranovich et al., 2013, Guedj et al., 2018, Escribano-
302 Romero et al., 2017, Arias et al., 2014). Of note, we also observed a strong inverse association between
303 infectious titers in lungs and the proportion of non-synonymous mutations detected in viral
304 populations. Because random non-synonymous mutations are more deleterious than synonymous
305 mutations (Cuevas et al., 2012), this suggests that they were randomly distributed over the three
306 positions of the codons and that no compensatory mechanism was triggered by the virus to eliminate
307 them (*i.e.* negative selection). Finally, the inverse correlation between lung infections titers and the
308 frequency of G→A substitutions showed that an increased proportion of these mutations beyond an
309 error threshold might be expected to cause lethal mutagenesis.

310 Genomic analyses revealed that 18 mutations detected in viral sub-populations were shared only with
311 treated animals. Two of them were located in the nsp14 coding region involved in the proof-reading
312 activity of the viral RNA polymerisation (Eckerle et al., 2007, Ferron et al., 2018). However, they were
313 located in the N7 MTase domain involved in viral RNA capping (Chen et al., 2013, Ma et al., 2015). By
314 comparison, resistance mutations selected against Remdesivir in β-coronavirus murine hepatitis virus
315 model were obtained in the RdRP (nsp12) coding sequence (Agostini et al., 2018). Further
316 investigations are needed to assess the impact of these mutations on the antiviral effect of favipiravir.

317 Favipiravir PK in our hamster model displayed a non-linear increase in plasma exposure between the
318 doses as already reported in nonhuman primates (Madelain et al., 2017). The observed favipiravir
319 concentration versus time profiles were in agreement with previous results of a PK study performed
320 in 7-8 week-old hamsters orally treated with a single dose of 100mg/kg of favipiravir (Gowen et al.,
321 2015). The maximum plasma drug concentration occurred at 0.5 h after oral administration, earlier
322 than in humans, and then decreased rapidly in agreement with its short half-life (Madelain et al., 2016).
323 After repeated doses, plasma exposure confirmed non-linear PK over the entire range of doses, further
324 emphasized by accumulation ratios. The important accumulation observed at the highest dose could
325 explain in part the toxicity observed in hamsters at this dose. Favipiravir undergoes an important
326 hepatic metabolism mainly by aldehyde oxidase producing an inactive M1 metabolite and inhibits
327 aldehyde oxidase activity in a concentration- and time-dependent manner. These properties explain

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the self-inhibition of its own metabolism as observed in our study in which the highest dose of favipiravir led to a greater increase in favipiravir concentrations (Madelain et al., 2020).

A good penetration of favipiravir in lungs was observed with lung/plasma ratios ranging from 35 to 44% after repeated doses, consistent with its physicochemical properties. Lung exposure was also in accordance with previous studies (Gowen et al., 2015).

How clinically realistic are these results? To address this question we compared the drug concentrations obtained in the hamster model with those obtained in patients. In 2016, a clinical trial evaluated the use of favipiravir in Ebola infected patients (Sissoko et al., 2016). The dose used in Ebola infected patients was 6000mg on day 0 followed by 1200mg BID for 9 days. The median trough concentrations of favipiravir at Day 2 and Day 4 were 46.1 and 25.9µg/mL, respectively. This is within the range observed here in hamsters treated with the highest dose (around 1400mg/kg/day), with a mean trough concentration of 29.9µg/mL. However, additional investigations are required to determine whether or not similar favipiravir plasma exposure in SARS-COV-2 infected patients are associated with antiviral activity. The major differences in PK between hamster and humans, and the toxicity observed at the highest doses in our animal model limits the extrapolation of our results. Therefore, whether safe dosing regimens in humans may achieve similar plasma exposure and recapitulate the profound effect on viral replication is unknown. Further, the intracellular concentration of the active metabolite was not determined and which parameter of the drug pharmacokinetics best drives the antiviral effect remains to be established.

In summary, this study establishes that high doses of favipiravir are associated with antiviral activity against SARS-CoV-2 infection in a hamster model. The better antiviral efficacy was observed using a preventive strategy, suggesting that favipiravir could be more appropriate for a prophylactic use. Our results should be interpreted with caution because high doses of favipiravir were associated with signs of toxicity in our model. It is required to determine if a tolerable dosing regimen could generate similar exposure in non-human primates, associated with significant antiviral activity, before testing a high dose regimen in COVID-19 patients. Furthermore, subsequent studies should determine if an increased antiviral efficacy can be reached using favipiravir in association with other effective antiviral drugs, since this strategy may enable to reduce the dosing regimen of favipiravir. Finally, this work reinforces the need for rapid development of animal models to confirm *in vivo* efficacy of antiviral compounds and accordingly, to determine appropriate dose regimens in humans before starting clinical trials.

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Author Contributions

Conceptualization, J.S.D., M.C., G.M. and A.N. ; Methodology, J.S.D., M.C., G.L., G.M. and A.N. ; Formal Analysis, J.S.D., M.C. and G.L. ; Investigation, J.S.D., M.C., G.M., F.T., P.R.P., G.P., K.B. and A.N. ; Resources, F.T., B.C., J.G., X.d.L., C.S. and A.N. ; Writing – Original Draft, J.S.D., M.C., J.G., C.S. and A.N. ; Writing – Review & Editing, J.G., X.d.L., C.S. and A.N. ; Visualization, J.S.D., M.C., G.L., F.T., P.R.P. and A.N. ; Supervision, A.N. ; Funding Acquisition, F.T., B.C., X.d.L. and A.N.

Declaration of Interests

J.G has consulted for F. Hoffman-La Roche. C.S has consulted for ViiV Healthcare, MSD and Gilead. The remaining authors declare no competing interests.

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Methods

Cells

VeroE6 cells (ATCC CRL-1586) and Caco-2 cells (ATCC HTB-37) were grown at 37°C with 5% CO_2 in minimal essential medium (MEM) supplemented with 7.5% heat-inactivated fetal bovine serum (FBS), 1% Penicillin/Streptomycin and 1% non-essential amino acids (all from ThermoFisher Scientific).

Virus

All experiments with infectious virus were conducted in biosafety level (BSL) 3 laboratory. SARS-CoV-2 strain BavPat1, supplied through European Virus Archive GLOBAL (https://www.european-virus-archive.com/), was provided by Christian Drosten (Berlin, Germany). Virus stocks were prepared by inoculating at MOI of 0.001 a 25cm2 culture flask of confluent VeroE6 cells with MEM medium supplemented with 2.5% FBS. The cell supernatant medium was replaced each 24h hours and harvested at the peak of infection, supplemented with 25mM HEPES (Sigma), aliquoted and stored at -80°C.

In vitro determination of EC_{50}, EC_{90}, CC_{50} and infectious titer reductions

One day prior to infection, 5×10^4 VeroE6 cells were seeded in 96-well culture plates (5×10^4 cells/well in 100µL of 2.5% FBS medium (assay medium). The next day, seven 2-fold serial dilutions of favipiravir (Courtesy of Toyama-Chemical; 0.61µg/mL to 78.5µg/mL, in triplicate) were added (25µL/well, in assay medium). Eight virus control wells were supplemented with 25µL of assay medium and eight cell controls were supplemented with 50µL of assay medium. After 15 min, 25µL of virus suspension, diluted in assay medium, was added to the wells at an MOI of 0.01 or 0.001 (except for cell controls). Three days after infection, cell supernatant media were collected to perform $TCID_{50}$ assay (at concentration of 78.5, 39.3, 19.6µg/mL), as described below, in order to calculate infectious titer reductions and cell viability was assessed using CellTiter-Blue reagent (Promega) following manufacturer's intructions. Fluorescence (560/590nm) was recorded with a Tecan Infinite 200Pro machine (Tecan). The 50% and 90% effective concentrations (EC_{50}, EC_{90}) were determined using logarithmic interpolation (% of inhibition were calculated as follows: $(OD_{sample}-OD_{virus\ control})/(OD_{cell\ control}-OD_{virus\ control})$). For the evaluation of CC_{50} (the concentration that induced 50% cytoxicity), the same culture conditions were set as for the determination of the EC_{50}, without addition of the virus, then cell viability was measured using CellTiter Blue (Promega). CC_{50} was determined using logarithmic interpolation.

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In vivo experiments

Approval and authorization

In vivo experiments were approved by the local ethical committee (C2EA—14) and the French 'Ministère de l'Enseignement Supérieur, de la Recherche et de l'Innovation' (APAFIS#23975) and performed in accordance with the French national guidelines and the European legislation covering the use of animals for scientific purposes. All experiments were conducted in BSL 3 laboratory.

Animal handling

Three-week-old female Syrian hamsters were provided by Janvier Labs. Animals were maintained in ISOcage P - Bioexclusion System (Techniplast) with unlimited access to water/food and 14h/10h light/dark cycle. Animals were weighed and monitored daily for the duration of the study to detect the appearance of any clinical signs of illness/suffering. Virus inoculation was performed under general anesthesia (isoflurane). Organs and blood were collected after euthanasia (cervical dislocation) which was also realized under general anesthesia (isofluorane).

Hamster Infection

Anesthetized animals (four-week-old) were intranasally infected with 50µL containing 10^6, 10^5 or 10^4 $TCID_{50}$ of virus in 0.9% sodium chloride solution). The mock group was intranasally inoculated with 50µL of 0.9% sodium chloride solution.

Favipiravir administration

Hamster were intra-peritoneally inoculated with different doses of favipiravir. Control group were intra-peritoneally inoculated with a 0.9% sodium chloride solution.

Organ collection

Organs were first washed in 10mL of 0.9% sodium chloride solution and then transferred to a 2mL or 50mL tube containing respectively 1mL (small/large bowel pieces, kidney, spleen and heart) or 10mL (lungs, brain and liver) of 0.9% sodium chloride solution and 3mm glass beads. They were crushed using a the Tissue Lyser machine (Retsch MM400) for 5min at 30 cycles/s and then centrifuged 5min à 1200g. Supernatant media were transferred to a 2mL tube, centrifuged 10 min at 16,200g and stored at -80°C. One milliliter of blood was harvested in a 2mL tube containing 100µL of 0.5M EDTA (ThermoFischer Scientific). Blood was centrifuged for 10 min at 16,200g and stored at -80°C.

Quantitative real-time RT-PCR (RT-qPCR) assays

To avoid contamination, all experiments were conducted in a molecular biology laboratory that is specifically designed for clinical diagnosis using molecular techniques, and which includes separate laboratories dedicated to perform each step of the procedure. Prior to PCR amplification, RNA extraction was performed using the QIAamp 96 DNA kit and the Qiacube HT kit and the Qiacube HT

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(both from Qiagen) following the manufacturer's instructions. Shortly, 100 µl of organ clarified homogenates, spiked with 10µL of internal control (bacteriophage MS2) (Ninove et al., 2011), were transferred into an S-block containing the recommended volumes of VXL, proteinase K and RNA carrier. RT-qPCR (SARS-CoV-2 and MS2 viral genome detection) were performed with the Express one step RT-qPCR Universal kit (ThermoFisher Scientific) using 3.5µL of RNA and 6.5µL of RT-qPCR mix that contains 250nmol of each primer and 75nmol of probe. Amplification was performed with the QuantStudio 12K Flex Real-Time PCR System (ThermoFisher Scientific) using the following conditions: 50°C for 10min, 95°C for 20s, followed by 40 cycles of 95°C for 3s, 60°C for 30s. qPCR (ɣ-actine gene detection) was perfomed under the same condition as RT-qPCR with the following modifications: we used the Express one step qPCR Universal kit (ThermoFisher Scientific) and the 50°C step of the amplification cycle was removed. Primers and probes sequences used to detect SARS-CoV-2, MS2 and ɣ-actine are described in Table S9.

Tissue-culture infectious dose 50 ($TCID_{50}$) assay

To determine infectious titers, 96-well culture plates containing confluent VeroE6 cells were inoculated with 150µL per well of serial dilutions of each sample (four-fold or ten-fold dilutions when analyzing lung clarified homogenates or cell supernatant media respectively). Each dilution was performed in sextuplicate. Plates were incubated for 4 days and then read for the absence or presence of cytopathic effect in each well. Infectious titers were estimated using the method described by Reed & Muench (REED and MUENCH, 1938).

Favipiravir pharmacokinetics

Animal handling, hamster infections and favipiravir administrations were performed as described above. A piece of left lung was first washed in 10mL of sodium chloride 0.9% solution, blotted with filter paper, weighed and then transferred to a 2mL tube containing 1mL of 0.9% sodium chloride solution and 3mm glass beads. It was crushed using the Tissue Lyser machine (Retsch MM400) during 10min at 30 cycles/s and then centrifuged 5min à 1200g. Supernatant media were transferred to 2mL tubes, centrifuged 10 min at 16,200g and stored at -80°C. One milliliter of blood was harvested in a 2mL tube containing 100µL of 0.5M EDTA (ThermoFischer Scientific). Blood was centrifuged for 10 min at 16,200g and stored at -80°C.

Quantification of favipiravir in plasma and lung tissues was performed by a validated sensitive and selective validated high-performance liquid chromatography coupled with tandem mass spectrometry method (UPLC-TQD, Waters, USA) with a lower limit of quantification of 0.1 µg/mL. Precision and accuracy of the 3 quality control samples (QCs) were within 15% over the calibration range (0.5 µg/mL to 100 µg/mL) (Bekegnran *et al.*, submitted). Favipiravir was extracted by a simple protein precipitation method, using acetonitrile for plasma and ice-cold acetonitrile for clarified lung homogenates. Briefly,

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50 µL of samples matrix was added to 500µL of acetonitrile solution containing the internal standard (favipiravir-13C,15N, Alsachim), then vortexed for 2min followed by centrifugation for 10min at 4°C. The supernatant medium was evaporated and the dry residues were then transferred to 96-well plates and 50 µL was injected. To assess the selectivity and specificity of the method and matrix effect, blank plasma and tissues homogenates from 2 control animals (uninfected and untreated) were processed at each run. Moreover, the same control samples spiked with favipiravir concentration equivalent to the QCs (0.75, 50 and 80 µg/mL) were also processed and compared to the QCs samples.

Noncompartemental analysis conducted using software Pkanalix2019R2 (www.lixoft.com). Areas under the plasma concentration time curve were computed using medians of favipiravir concentrations at 0.5, 1, 5 and 8 hours, and extrapolated until T=12h. C_{trough} were extrapolated at T=12h using lambda-z loglinear regression on the decreasing slope of concentrations.

Sequence analysis of the full-length genome

200µL of lung clarified homogenate or infectious cell supernatant (virus stock) was inactivated with an equal volume of VXL lysis buffer (Qiagen) and viral RNA was extracted using an EZ1 Advanced XL robot with the EZ1 mini virus 2.0 kit (both from Qiagen) and linear acrylamide (ThermoFisher Scientific) in place of carrier RNA. cDNA was generated in a final volume of 40µL using 14µL of nucleic acid extract, random hexamer and the Protoscript II First Strand cDNA Synthesis Kit (New England Biolabs). A specific set of primers (Table S10) was used to generate thirteen amplicons covering the entire genome with the Q5 High-Fidelity DNA polymerase (New England Biolabs). PCR mixes (final volume 25µL) contained 2.5µL of cDNA, 2µL of each primer (10µM) and 12.5 µL of Q5 High-Fidelity 2X Master Mix. Amplification was performed with the following conditions: 30 sec at 98°C, then 45 cycles of 15 sec at 98°C and 5 min à 65°C. Size of PCR products was verified by gel electrophoresis. For each sample, an equimolar pool of all amplicons was prepared and purified using Monarch PCR & DNA Cleanup Kit (New England Biolabs). After DNA quantification using Qubit dsDNA HS Assay Kit and Qubit 2.0 fluorometer (ThermoFisher Scientific), amplicons were fragmented by sonication into fragments of around 200bp long. Libraries were built by adding barcodes, for sample identification, and primers using AB Library Builder System (ThermoFisher Scientific). To pool equimolarly the barcoded samples a quantification step by real time PCR using Ion Library TaqMan Quantitation Kit (ThermoFisher Scientific) was performed. Then, emulsion PCR from pools and loading on 530 chip was performed using the automated Ion Chef instrument (ThermoFisher Scientific). Sequencing was performed using the S5 Ion torrent technology v5.12 (ThermoFisher Scientific) following manufacturer's instructions. Consensus sequence was obtained after trimming of reads (reads with quality score <0.99, and length <100pb were removed and the 30 first and 30 last nucleotides were removed from the reads). Mapping of the reads on a reference (determine following blast of De Novo contigs) was done using CLC genomics

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workbench software v.20 (Qiagen). A *de novo* contig was also produced to ensure that the consensus sequence was not affected by the reference sequence. Mutation frequency for each position was calculated as the number of reads with a mutation compared to the reference divided by the total number of reads at that site. Only substitutions with a frequency of at least 1% were taken into account for the analysis (Table S6).

ED_{50}, ED_{90} and ED_{99} determination

We conducted a nonlinear regression of infectious viral load against dose, using an E_{max} model, giving $VL = VL_0 \times \left(1 - \left(\frac{D^{\gamma}}{D^{\gamma}+D_{50}^{\gamma}}\right)\right)$ with VL_0 being infectious viral load of untreated animals. We estimated D_{50} the dose required to decrease viral load by 50%, using a coefficient γ to account for the high sigmoidicity of the relation between dose and titers. γ coefficient was chosen as the one maximizing likelihood of the model. We extrapolated the D_{90} and D_{99} using $D_{90} = \sqrt[\gamma]{9 \times D_{50}^{\gamma}}$ and $D_{99} = \sqrt[\gamma]{99 \times D_{50}^{\gamma}}$, as well as their 95% confidence interval using the delta method.

Statistical analysis

Graphical representations and statistical analyses were performed with Graphpad Prism 7 (Graphpad software) except linear/nonlinear regressions and their corresponding graphical representations that were performed using R statistical software (http://www.R-project.org). Statistical details for each experiments are described in the figure legends and in corresponding supplemental tables. P-values lower than 0.05 were considered statistically significant.

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Supplemental Data

Supplemental figure 1: In vitro efficacy of favipiravir

Supplemental figure 2: Dose-response curves

Supplemental figure 3: Evaluation of the toxicity for animals infected and treated with high doses of favipiravir

Supplemental figure 4: Plasma concentrations of favipiravir after administration of a single dose of favipiravir

Supplemental table 1: Implementation of hamster model

Supplemental table 2: Individual data from in vivo experiments

Supplemental table 3: Statistical analysis of in vivo experiments

Supplemental table 4: Statistical analysis of clinical monitoring

Supplemental table 5: Individual data of favipiravir pharmacokinetics

Supplemental table 6: Individual data for analysis of mutagenic effect of favipiravir

Supplemental table 7: Statistical analysis of mutagenic effect of favipiravir

Supplemental table 8: Shared mutations detected in lung clarified homogenates

Supplemental table 9: (RT)-qPCR systems

Supplemental table 10: Primer sequences used to produce overlapping amplicons for next generation sequencing

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References

AGOSTINI, M. L., ANDRES, E. L., SIMS, A. C., GRAHAM, R. L., SHEAHAN, T. P., LU, X., SMITH, E. C., CASE, J. B., FENG, J. Y., JORDAN, R., RAY, A. S., CIHLAR, T., SIEGEL, D., MACKMAN, R. L., CLARKE, M. O., BARIC, R. S. & DENISON, M. R. 2018. Coronavirus Susceptibility to the Antiviral Remdesivir (GS-5734) Is Mediated by the Viral Polymerase and the Proofreading Exoribonuclease. *mBio,* 9.

ARIAS, A., THORNE, L. & GOODFELLOW, I. 2014. Favipiravir elicits antiviral mutagenesis during virus replication in vivo. *Elife,* 3**,** e03679.

BARANOVICH, T., WONG, S. S., ARMSTRONG, J., MARJUKI, H., WEBBY, R. J., WEBSTER, R. G. & GOVORKOVA, E. A. 2013. T-705 (favipiravir) induces lethal mutagenesis in influenza A H1N1 viruses in vitro. *J Virol,* 87**,** 3741-51.

CHAN, J. F., ZHANG, A. J., YUAN, S., POON, V. K., CHAN, C. C., LEE, A. C., CHAN, W. M., FAN, Z., TSOI, H. W., WEN, L., LIANG, R., CAO, J., CHEN, Y., TANG, K., LUO, C., CAI, J. P., KOK, K. H., CHU, H., CHAN, K. H., SRIDHAR, S., CHEN, Z., CHEN, H., TO, K. K. & YUEN, K. Y. 2020. Simulation of the clinical and pathological manifestations of Coronavirus Disease 2019 (COVID-19) in golden Syrian hamster model: implications for disease pathogenesis and transmissibility. *Clin Infect Dis*.

CHEN, Y., TAO, J., SUN, Y., WU, A., SU, C., GAO, G., CAI, H., QIU, S., WU, Y., AHOLA, T. & GUO, D. 2013. Structure-function analysis of severe acute respiratory syndrome coronavirus RNA cap guanine-N7-methyltransferase. *J Virol,* 87**,** 6296-305.

CUEVAS, J. M., DOMINGO-CALAP, P. & SANJUAN, R. 2012. The fitness effects of synonymous mutations in DNA and RNA viruses. *Mol Biol Evol,* 29**,** 17-20.

DONG, E., DU, H. & GARDNER, L. 2020. An interactive web-based dashboard to track COVID-19 in real time. *Lancet Infect Dis,* 20**,** 533-534.

ECKERLE, L. D., LU, X., SPERRY, S. M., CHOI, L. & DENISON, M. R. 2007. High fidelity of murine hepatitis virus replication is decreased in nsp14 exoribonuclease mutants. *J Virol,* 81**,** 12135-44.

ESCRIBANO-ROMERO, E., JIMENEZ DE OYA, N., DOMINGO, E. & SAIZ, J. C. 2017. Extinction of West Nile Virus by Favipiravir through Lethal Mutagenesis. *Antimicrob Agents Chemother,* 61.

FERRON, F., SUBISSI, L., SILVEIRA DE MORAIS, A. T., LE, N. T. T., SEVAJOL, M., GLUAIS, L., DECROLY, E., VONRHEIN, C., BRICOGNE, G., CANARD, B. & IMBERT, I. 2018. Structural and molecular basis of mismatch correction and ribavirin excision from coronavirus RNA. *Proc Natl Acad Sci U S A,* 115**,** E162-E171.

bioRxiv preprint doi: https://doi.org/10.1101/2020.07.07.191775; this version posted July 7, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

GOWEN, B. B., SEFING, E. J., WESTOVER, J. B., SMEE, D. F., HAGLOCH, J., FURUTA, Y. & HALL, J. O. 2015. Alterations in favipiravir (T-705) pharmacokinetics and biodistribution in a hamster model of viral hemorrhagic fever. *Antiviral Res,* 121**,** 132-7.

GUEDJ, J., PIORKOWSKI, G., JACQUOT, F., MADELAIN, V., NGUYEN, T. H. T., RODALLEC, A., GUNTHER, S., CARBONNELLE, C., MENTRE, F., RAOUL, H. & DE LAMBALLERIE, X. 2018. Antiviral efficacy of favipiravir against Ebola virus: A translational study in cynomolgus macaques. *PLoS Med,* 15**,** e1002535.

HE, X., LAU, E. H. Y., WU, P., DENG, X., WANG, J., HAO, X., LAU, Y. C., WONG, J. Y., GUAN, Y., TAN, X., MO, X., CHEN, Y., LIAO, B., CHEN, W., HU, F., ZHANG, Q., ZHONG, M., WU, Y., ZHAO, L., ZHANG, F., COWLING, B. J., LI, F. & LEUNG, G. M. 2020. Temporal dynamics in viral shedding and transmissibility of COVID-19. *Nat Med,* 26**,** 672-675.

HUANG, C., WANG, Y., LI, X., REN, L., ZHAO, J., HU, Y., ZHANG, L., FAN, G., XU, J., GU, X., CHENG, Z., YU, T., XIA, J., WEI, Y., WU, W., XIE, X., YIN, W., LI, H., LIU, M., XIAO, Y., GAO, H., GUO, L., XIE, J., WANG, G., JIANG, R., GAO, Z., JIN, Q., WANG, J. & CAO, B. 2020. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet,* 395**,** 497-506.

JEON, S., KO, M., LEE, J., CHOI, I., BYUN, S. Y., PARK, S., SHUM, D. & KIM, S. 2020. Identification of antiviral drug candidates against SARS-CoV-2 from FDA-approved drugs. *Antimicrob Agents Chemother*.

JOCHMANS, D., VAN NIEUWKOOP, S., SMITS, S. L., NEYTS, J., FOUCHIER, R. A. & VAN DEN HOOGEN, B. G. 2016. Antiviral Activity of Favipiravir (T-705) against a Broad Range of Paramyxoviruses In Vitro and against Human Metapneumovirus in Hamsters. *Antimicrob Agents Chemother,* 60**,** 4620-9.

JULANDER, J. G., SHAFER, K., SMEE, D. F., MORREY, J. D. & FURUTA, Y. 2009. Activity of T-705 in a hamster model of yellow fever virus infection in comparison with that of a chemically related compound, T-1106. *Antimicrob Agents Chemother,* 53**,** 202-9.

KAPTEIN, S. J., JACOBS, S., LANGENDRIES, L., SELDESLACHTS, L., TER HORST, S., LIESENBORGHS, L., HENS, B., VERGOTE, V., HEYLEN, E., MAAS, E., DE KEYZER, C., BERVOETS, L., RYMENANTS, J., VAN BUYTEN, T., THIBAUT, H. J., DALLMEIER, K., BOUDEWIJNS, R., WOUTERS, J., AUGUSTIJNS, P., VEROUGSTRAETE, N., CAWTHORNE, C., WEYNAND, B., ANNAERT, P., SPRIET, I., VELDE, G. V., NEYTS, J., ROCHA-PEREIRA, J. & DELANG, L. 2020. Antiviral treatment of SARS-CoV-2-infected hamsters reveals a weak effect of favipiravir and a complete lack of effect for hydroxychloroquine. *bioRxiv***,** 2020.06.19.159053.

MA, Y., WU, L., SHAW, N., GAO, Y., WANG, J., SUN, Y., LOU, Z., YAN, L., ZHANG, R. & RAO, Z. 2015. Structural basis and functional analysis of the SARS coronavirus nsp14-nsp10 complex. *Proc Natl Acad Sci U S A,* 112**,** 9436-41.

bioRxiv preprint doi: https://doi.org/10.1101/2020.07.07.191775; this version posted July 7, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

MADELAIN, V., GUEDJ, J., MENTRE, F., NGUYEN, T. H., JACQUOT, F., OESTEREICH, L., KADOTA, T., YAMADA, K., TABURET, A. M., DE LAMBALLERIE, X. & RAOUL, H. 2017. Favipiravir Pharmacokinetics in Nonhuman Primates and Insights for Future Efficacy Studies of Hemorrhagic Fever Viruses. *Antimicrob Agents Chemother,* 61.

MADELAIN, V., MENTRE, F., BAIZE, S., ANGLARET, X., LAOUENAN, C., OESTEREICH, L., NGUYEN, T. H. T., MALVY, D., PIORKOWSKI, G., GRAW, F., GUNTHER, S., RAOUL, H., DE LAMBALLERIE, X. & GUEDJ, J. 2020. Modeling Favipiravir Antiviral Efficacy Against Emerging Viruses: From Animal Studies to Clinical Trials. *CPT Pharmacometrics Syst Pharmacol,* 9**,** 258-271.

MADELAIN, V., NGUYEN, T. H., OLIVO, A., DE LAMBALLERIE, X., GUEDJ, J., TABURET, A. M. & MENTRE, F. 2016. Ebola Virus Infection: Review of the Pharmacokinetic and Pharmacodynamic Properties of Drugs Considered for Testing in Human Efficacy Trials. *Clin Pharmacokinet,* 55**,** 907-23.

MERCORELLI, B., PALU, G. & LOREGIAN, A. 2018. Drug Repurposing for Viral Infectious Diseases: How Far Are We? *Trends Microbiol,* 26**,** 865-876.

NINOVE, L., NOUGAIREDE, A., GAZIN, C., THIRION, L., DELOGU, I., ZANDOTTI, C., CHARREL, R. N. & DE LAMBALLERIE, X. 2011. RNA and DNA bacteriophages as molecular diagnosis controls in clinical virology: a comprehensive study of more than 45,000 routine PCR tests. *PLoS One,* 6**,** e16142.

OESTEREICH, L., RIEGER, T., LUDTKE, A., RUIBAL, P., WURR, S., PALLASCH, E., BOCKHOLT, S., KRASEMANN, S., MUNOZ-FONTELA, C. & GUNTHER, S. 2016. Efficacy of Favipiravir Alone and in Combination With Ribavirin in a Lethal, Immunocompetent Mouse Model of Lassa Fever. *J Infect Dis,* 213**,** 934-8.

REED, L. J. & MUENCH, H. 1938. A SIMPLE METHOD OF ESTIMATING FIFTY PER CENT ENDPOINTS. *American Journal of Epidemiology,* 27**,** 493-497.

ROCKX, B., KUIKEN, T., HERFST, S., BESTEBROER, T., LAMERS, M. M., OUDE MUNNINK, B. B., DE MEULDER, D., VAN AMERONGEN, G., VAN DEN BRAND, J., OKBA, N. M. A., SCHIPPER, D., VAN RUN, P., LEIJTEN, L., SIKKEMA, R., VERSCHOOR, E., VERSTREPEN, B., BOGERS, W., LANGERMANS, J., DROSTEN, C., FENTENER VAN VLISSINGEN, M., FOUCHIER, R., DE SWART, R., KOOPMANS, M. & HAAGMANS, B. L. 2020. Comparative pathogenesis of COVID-19, MERS, and SARS in a nonhuman primate model. *Science,* 368**,** 1012-1015.

ROSENKE, K., FELDMANN, H., WESTOVER, J. B., HANLEY, P. W., MARTELLARO, C., FELDMANN, F., SATURDAY, G., LOVAGLIO, J., SCOTT, D. P., FURUTA, Y., KOMENO, T., GOWEN, B. B. & SAFRONETZ, D. 2018. Use of Favipiravir to Treat Lassa Virus Infection in Macaques. *Emerg Infect Dis,* 24**,** 1696-1699.

bioRxiv preprint doi: https://doi.org/10.1101/2020.07.07.191775; this version posted July 7, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

SANGAWA, H., KOMENO, T., NISHIKAWA, H., YOSHIDA, A., TAKAHASHI, K., NOMURA, N. & FURUTA, Y. 2013. Mechanism of action of T-705 ribosyl triphosphate against influenza virus RNA polymerase. *Antimicrob Agents Chemother,* 57**,** 5202-8.

SEGURA GUERRERO, N. A., SHARMA, S., NEYTS, J. & KAPTEIN, S. J. F. 2018. Favipiravir inhibits in vitro Usutu virus replication and delays disease progression in an infection model in mice. *Antiviral Res,* 160**,** 137-142.

SHANNON, A., SELISKO, B., LE, N., HUCHTING, J., TOURET, F., PIORKOWSKI, G., FATTORINI, V., FERRON, F., DECROLY, E., MEIER, C., COUTARD, B., PEERSEN, O. & CANARD, B. 2020. Favipiravir strikes the SARS-CoV-2 at its Achilles heel, the RNA polymerase. *bioRxiv***,** 2020.05.15.098731.

SIDWELL, R. W., BARNARD, D. L., DAY, C. W., SMEE, D. F., BAILEY, K. W., WONG, M. H., MORREY, J. D. & FURUTA, Y. 2007. Efficacy of orally administered T-705 on lethal avian influenza A (H5N1) virus infections in mice. *Antimicrob Agents Chemother,* 51**,** 845-51.

SISSOKO, D., LAOUENAN, C., FOLKESSON, E., M'LEBING, A. B., BEAVOGUI, A. H., BAIZE, S., CAMARA, A. M., MAES, P., SHEPHERD, S., DANEL, C., CARAZO, S., CONDE, M. N., GALA, J. L., COLIN, G., SAVINI, H., BORE, J. A., LE MARCIS, F., KOUNDOUNO, F. R., PETITJEAN, F., LAMAH, M. C., DIEDERICH, S., TOUNKARA, A., POELART, G., BERBAIN, E., DINDART, J. M., DURAFFOUR, S., LEFEVRE, A., LENO, T., PEYROUSET, O., IRENGE, L., BANGOURA, N., PALICH, R., HINZMANN, J., KRAUS, A., BARRY, T. S., BERETTE, S., BONGONO, A., CAMARA, M. S., MUNOZ, V. C., DOUMBOUYA, L., HAROUNA, S., KIGHOMA, P. M., KOUNDOUNO, F. R., LOLAMOU, R., LOUA, C. M., MASSALA, V., MOUMOUNI, K., PROVOST, C., SAMAKE, N., SEKOU, C., SOUMAH, A., ARNOULD, I., KOMANO, M. S., GUSTIN, L., BERUTTO, C., CAMARA, D., CAMARA, F. S., COLPAERT, J., DELAMOU, L., JANSSON, L., KOUROUMA, E., LOUA, M., MALME, K., MANFRIN, E., MAOMOU, A., MILINOUNO, A., OMBELET, S., SIDIBOUN, A. Y., VERRECKT, I., YOMBOUNO, P., BOCQUIN, A., CARBONNELLE, C., CARMOI, T., FRANGE, P., MELY, S., NGUYEN, V. K., PANNETIER, D., TABURET, A. M., TRELUYER, J. M., KOLIE, J., MOH, R., GONZALEZ, M. C., KUISMA, E., LIEDIGK, B., NGABO, D., RUDOLF, M., THOM, R., KERBER, R., GABRIEL, M., WLFEL, R., WOLFEL, R., BADIR, J., BENTAHIR, M., DECCACHE, Y., DUMONT, C., DURANT, J. F., EL BAKKOURI, K., UWAMAHORO, M. G., SMITS, B., TOUFIK, N., et al. 2016. Experimental Treatment with Favipiravir for Ebola Virus Disease (the JIKI Trial): A Historically Controlled, Single-Arm Proof-of-Concept Trial in Guinea. *Plos Medicine,* 13.

SMITHER, S. J., EASTAUGH, L. S., STEWARD, J. A., NELSON, M., LENK, R. P. & LEVER, M. S. 2014. Post-exposure efficacy of oral T-705 (Favipiravir) against inhalational Ebola virus infection in a mouse model. *Antiviral Res,* 104**,** 153-5.

bioRxiv preprint doi: https://doi.org/10.1101/2020.07.07.191775; this version posted July 7, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

TAKAHASHI, K., FURUTA, Y., FUKUDA, Y., KUNO, M., KAMIYAMA, T., KOZAKI, K., NOMURA, N., EGAWA, H., MINAMI, S. & SHIRAKI, K. 2003. In vitro and in vivo activities of T-705 and oseltamivir against influenza virus. *Antivir Chem Chemother,* 14**,** 235-41.

TANI, H., KOMENO, T., FUKUMA, A., FUKUSHI, S., TANIGUCHI, S., SHIMOJIMA, M., UDA, A., MORIKAWA, S., NAKAJIMA, N., FURUTA, Y. & SAIJO, M. 2018. Therapeutic effects of favipiravir against severe fever with thrombocytopenia syndrome virus infection in a lethal mouse model: Dose-efficacy studies upon oral administration. *PLoS One,* 13**,** e0206416.

WANG, M., CAO, R., ZHANG, L., YANG, X., LIU, J., XU, M., SHI, Z., HU, Z., ZHONG, W. & XIAO, G. 2020. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res,* 30**,** 269-271.

WHO 2020. World Health Organization. WHO Director-General's opening remarks at the media briefing on COVID-19 - 11 March 2020 (https://www.who.int).

WILLIAMSON, B. N., FELDMANN, F., SCHWARZ, B., MEADE-WHITE, K., PORTER, D. P., SCHULZ, J., VAN DOREMALEN, N., LEIGHTON, I., YINDA, C. K., PEREZ-PEREZ, L., OKUMURA, A., LOVAGLIO, J., HANLEY, P. W., SATURDAY, G., BOSIO, C. M., ANZICK, S., BARBIAN, K., CIHLAR, T., MARTENS, C., SCOTT, D. P., MUNSTER, V. J. & DE WIT, E. 2020. Clinical benefit of remdesivir in rhesus macaques infected with SARS-CoV-2. *Nature*.

YAMADA, K., NOGUCHI, K., KIMITSUKI, K., KAIMORI, R., SAITO, N., KOMENO, T., NAKAJIMA, N., FURUTA, Y. & NISHIZONO, A. 2019. Reevaluation of the efficacy of favipiravir against rabies virus using in vivo imaging analysis. *Antiviral Res,* 172**,** 104641.

ZHU, N., ZHANG, D., WANG, W., LI, X., YANG, B., SONG, J., ZHAO, X., HUANG, B., SHI, W., LU, R., NIU, P., ZHAN, F., MA, X., WANG, D., XU, W., WU, G., GAO, G. F., TAN, W., CHINA NOVEL CORONAVIRUS, I. & RESEARCH, T. 2020. A Novel Coronavirus from Patients with Pneumonia in China, 2019. *N Engl J Med,* 382**,** 727-733.