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Evolutionary dynamics of sex-biased genes expressed in cricket brains and gonads

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Evolutionary dynamics of sex-biased genes expressed in cricket brains and gonads

Abstract

Sex-biased gene expression, particularly male-biased expression in the gonad, has often been linked to rapid protein sequence evolution (nonsynonymous to synonymous substitutions, dN/dS) in animals. This evolutionary trend may arise from adaptive evolution and/or from relaxed purifying selection due to low pleiotropy. In insects, research on sex-biased transcription and dN/dS remains largely focused on a few holometabolous species, with variable findings on male and female gonadal effects. The brain provides the neurological foundation for male and female mating behaviors. However, little is known about sex-biased expression of brain genes and their rates of protein sequence evolution. Here, we studied sex-biased gene expression in a hemimetabolous insect, the cricket *Gryllus bimaculatus*. We generated novel male and female RNA-seq data for two sexual tissue types, the gonad and somatic reproductive system, and for two core components of the nervous system, the brain and ventral nerve cord. From a genome-wide analysis of genes expressed in these tissues, we report the accelerated evolution of testis-biased gene, as compared to ovary-biased and unbiased genes, which was associated with an elevated frequency of positive selection events. With respect to the brain, while sex-biased brain genes were much less common than for the gonads, they exhibited a striking tendency for rapid evolution, an effect that was stronger for the female than for the male brain. Certain sex-biased brain genes were predicted to be involved in mating or sex-related functions, which we suggest may cause exposure to sexual selection. A tendency for narrow cross-tissue expression breadth, suggesting low pleiotropy, was observed for sex-biased brain genes. We speculate that this feature may permit relaxed purifying selection, which in turn, may allow enhanced freedom for adaptive protein functional changes in these sex-biased genes. Our results demonstrate that sex-biased expression in the male gonad, and sex-biased expression in the brain, especially the female brain, are associated with a propensity for rapid protein sequence evolution in a cricket model system. We discuss the results with respect to our findings on pleiotropy and positive selection, and consider the plausible roles of the mating biology of this cricket in shaping the observed patterns.

Keywords: *Gryllus*, sex-biased expression, dN/dS, pleiotropy, gonad, brain, mating biology

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1 INTRODUCTION

Sexual dimorphism in animals is thought to be driven by differential gene expression, as most genes are common to both sexes (Ellegren & Parsch, 2007; Ingleby et al., 2014; Grath & Parsch, 2016). Sex-biased gene expression, and particularly male-biased gene expression, has been widely linked to rapid protein sequence evolution in studied animals (reviewed by (Ellegren & Parsch, 2007; Ingleby et al., 2014; Grath & Parsch, 2016)). In the insects, studies have largely focused on the holometabolous insect *Drosophila*, and have repeatedly shown the rapid evolution (high nonsynonymous to synonymous substitution rates, dN/dS) of male-biased genes, particularly those from the male sex cells or gonads, as compared to their female counterparts and/or to sexually unbiased genes (Jagadeeshan & Singh, 2005; Ellegren & Parsch, 2007; Haerty et al., 2007; Zhang et al., 2007; Jiang & Machado, 2009; Meisel, 2011; Grath & Parsch, 2012; Perry et al., 2015; Whittle & Extavour, 2019) (but see also (Dorus et al., 2006)). This pattern was also recently observed for the gonads of red flour beetles (*T. castaneum*) (Whittle et al., 2020). The rapid divergence of male-biased genes has been proposed to be due to adaptive changes in amino acids arising from sexual selection pressures including male-male and sperm competition (Swanson et al., 2001; Zhang et al., 2004; Proschel et al., 2006; Haerty et al., 2007), but could also reflect low pleiotropy that may relax purifying selection (Zhang et al., 2007; Mank & Ellegren, 2009; Assis et al., 2012; Dean & Mank, 2016; Whittle & Extavour, 2019). Nonetheless, the pattern of accelerated evolution of male-biased genes is not universal, as an opposite pattern of rapid evolution of female-biased, including ovary-biased, genes has been found in some holometabolous insects, namely mosquitoes (*Aedes*, *Anopheles*) (Papa et al., 2017; Whittle & Extavour, 2017). This difference from flies may reflect variation in their mating biology, whereby female-female competition for suitable males or male-mate choice may be more common in mosquitoes than in flies, and/or reflect variation in male- and female-related purifying selection among insects (Whittle & Extavour, 2017). At present however, given the narrow scope of insects studied to date, further investigation of sex-biased expression in the reproductive system and protein evolution is warranted, particularly in models outside the Holometabola.

While studies of sex-biased expression and its link to protein sequence evolution have largely focused on the reproductive system, a major, and markedly understudied structure, in terms of molecular evolution, is the brain. The brain is a major tissue type providing the

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neurological basis for the mating behaviors of courtship, intrasex competition, mate-choice, and post-mating male-female responses (Mank et al., 2007; Dalton et al., 2010; Naurin et al., 2011; Wright & Mank, 2013). Male and female differences in gene expression *per se* in the brain have been examined in some insects and vertebrates (Jagadeeshan & Singh, 2005; Mank et al., 2007; Santos et al., 2008; Small et al., 2009; Naurin et al., 2011; Catalan et al., 2012; Wright & Mank, 2013; Ingleby et al., 2014; Tomchaney et al., 2014; Huylmans & Parsch, 2015; Shi et al., 2016; Yang et al., 2016; Khodursky et al., 2020). Further, in *Drosophila*, analyses of a small number of neural genes showed a direct connection to mating functions and behaviors (Drapeau et al., 2003; Kadener et al., 2006; Dauwalder, 2008). However, there is a striking paucity of data on whether and how sex-biased expression in the brain is associated with protein sequence evolution (Mank et al., 2007; Wright & Mank, 2013). Moreover, the minimal research available from birds, humans and flies has suggested that male and female expression may have different effects on the rates of protein evolution, depending on the system (Mank et al., 2007; Shi et al., 2016; Khodursky et al., 2020) (see also some brain-related (Biswas et al., 2016) and composite-tissue analyses (Catalan et al., 2018; Congrains et al., 2018)), and the causes of those patterns remain poorly understood. It is therefore evident that additional study of sex-biased expression in the brain is needed, particularly with respect to its relationship to molecular evolution.

An insect model system that offers significant opportunities to address these problems is the cricket *Gryllus* (Order Orthoptera). *Gryllus* is a hemimetabolous insect, and thus in an outgroup order to the Holometabola (Misof et al., 2014). The two-spotted cricket *G. bimaculatus* in particular has emerged as a significant insect model in biology, including for genetics, neuroscience and germ line establishment and development (Kulkarni & Extavour, 2019). In fact, many of the developmental mechanisms of *G. bimaculatus* appear more typical of arthropods than the widely studied, and relatively derived, model *Drosophila melanogaster* (Mito & Noji, 2008; Donoughe & Extavour, 2016). Moreover, many aspects of its mating biology are currently well understood. *G. bimaculatus* exhibits intense male-male and sperm competition, including aggressive male-male fighting and mate guarding (Vedenina & Shestakov, 2018; Gee, 2019), increased rates of male transfer of spermatophores to females in the presence of other males (Lyons & Barnard, 2006), and the complete mixing of sperm from multiple males in the storage organ of the female reproductive tract, the spermatheca (Simmons, 1986; Morrow & Gage, 2001). In addition, females have shown preferences for novel and young

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mating partners (Zhemchuzhnikov et al., 2017), and for males with larger body size and higher quality auditory signals (Bateman et al., 2001; Zhemchuzhnikov et al., 2017). Females also exhibit a post-mating behaviour of removing spermatophores of non-favored males from their reproductive tract (Simmons, 1986), suggesting a propensity for female mate choice in this organism. Moreover, in terms of the brain, experiments in *G. bimaculatus* have shown that the brain is directly involved in male mating behaviors such as courtship, copulation, spermatophore protrusion, mating intervals and male-female auditory mating signalling (Matsumoto & Sakai, 2000; Haberkern & Hedwig, 2016; Sakai et al., 2017). The study of *Gryllus* therefore provides a valuable avenue to advance our knowledge of sex-biased expression in reproductive and brain tissues, including relationships to dN/dS and pleiotropy, in a taxon having well-studied mating biology.

Here, we rigorously assess sex-biased gene expression for two tissue types from the reproductive system (gonad and somatic reproductive system) and from the nervous system (brain and ventral nerve cord) in *G. bimaculatus*, and evaluate their relationships to protein sequence evolution. We report that male-biased gene expression in the gonad is linked to rapid protein sequence evolution (dN/dS), as compared to unbiased and female-biased genes. However, we observed no consistent effect of sex-biased expression in the somatic reproductive system (non-germ line tissues) on dN/dS, despite the roles of these sexual tissues in male-female interaction, mating and fertilization, and their potential exposure to sexual selection pressures (Swanson & Vacquier, 2002; Swanson et al., 2004; Clark & Swanson, 2005; Panhuis & Swanson, 2006; Haerty et al., 2007). With respect to the brain, we demonstrate that sex-biased genes are uncommon as compared to the gonad, and that these genes typically evolve very rapidly, especially the female-biased brain genes. Further, sex-biased brain genes are conspicuously linked to predicted sex-related functions. The sex-biased brain genes exhibit especially low cross-tissue expression, a proxy for pleiotropy (Mank & Ellegren, 2009), which may in itself accelerate protein sequence evolution due to relaxed purifying constraint. We propose that this low pleiotropy may also comprise a mechanism potentially allowing greater freedom for these brain-expressed proteins to evolve adaptive functional changes, an evolutionary dynamic that has been suggested in some studies (Otto, 2004; Larracuente et al., 2008; Mank et al., 2008; Mank & Ellegren, 2009; Meisel, 2011). We consider the putative roles of the male and female mating biology of *G. bimaculatus* in shaping the present finings.

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2 RESULTS AND DISCUSSION

2.1 Identification of sex-biased genes

The CDS of our main target species *G. bimaculatus* were obtained from its recently available genome (Ylla et al., 2021) . The annotated genome had 17,714 predicted transcripts (after selecting the longest CDS per gene; (Ylla et al., 2021)). For this gene set, we extracted the CDS with a start codon, no ambiguous nucleotides, and at least 150bp in length, yielding 15,539 CDS for study (mean length=417.0 codons/CDS ±3.5 (standard error (SE))) for *G. bimaculatus*. For analysis of sex-biased gene expression in *G. bimaculatus* we isolated and obtained RNA-seq data for four paired male and female tissue types from adult virgins (biological replicates (Congrains et al., 2018) and read counts in Table S1). The tissues included the gonad (testis for males, ovaries for females), somatic reproductive system, brain and ventral nerve cord (shown in Fig. 1A-F). The somatic (non-germ line related) reproductive system herein for males included the pooled vasa deferentia, seminal vesicles and ejaculatory duct, and for females included the spermatheca, common and lateral oviducts, and bursa (Fig. 1A,B; note that a ninth, unpaired, tissue type, the male accessory glands was also isolated and was used in pleiotropy analysis and its own supplementary dN/dS analysis (Fig. 1G), as detailed in Methods). For each sample, reads were mapped to the entire *G. bimaculatus* CDS list to determine FPKM. We found FPKM was strongly correlated between the biological replicates, with Spearman's $R \geq 0.92$ ($P<0.05$; Fig. S1; one exception being the male somatic reproductive system, $R=0.71$, $P<0.05$), indicating high reproducibility of expression profiles. Sex-biased gene expression was then determined separately for each of the four male-female paired tissue types using a cut-off of two-fold higher expression in one sex versus the other and a P value of <0.05 (see Methods). All genes per tissue that were not sex-biased in expression were defined as unbiased, such that all studied genes were assigned to one of three categories (male-biased, female-biased, unbiased; see Methods for details).

As shown in Fig. 2, sex-biased gene expression was most common in the gonadal tissues, where 4,822 (31.0%) of all *G. bimaculatus* genes under study were sex-biased in expression: 2,698 (17.4%) and 2,124 (13.7%) genes had ovary-biased and testis-biased expression respectively, and a total of 10,717 (69.0%) were unbiased in expression. By comparison, sex-biased gene expression was markedly less common in the somatic reproductive system, where

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only 5.6% of genes were sex-biased, with 353 (2.3%) and 520 (3.3%) genes showing female- and male-bias respectively. As compared to the gonad, markedly fewer genes exhibited female-biased and male-biased expression in the nervous system tissues, where 4.5% of 15,539 *G. bimaculatus* genes had sex-biased expression in the ventral nerve cord: 279 (1.8%) and 425 (2.7%) were female- and male-biased respectively (Fig. 2). For the brain, only 1.0% of genes were sex-biased in expression, with 51 (0.33%) and 106 (0.68%) being female- and male-biased respectively, an uncommonness that notably has also been suggested for brains of *D melanogaster* (Huylmans & Parsch, 2015). Together, using the present criteria, it is evident that sex-biased gene expression is most common in the gonad, which is consistent with high phenotypic and transcriptional dimorphism of these sex organs in animals (Arbeitman et al., 2004; Parisi et al., 2004; Zhang et al., 2004; Small et al., 2009; Oliver et al., 2010; Meisel, 2011; Harrison et al., 2015; Whittle & Extavour, 2017; Whittle & Extavour, 2019; Whittle et al., 2020). In contrast, sex-biased gene expression is markedly less common in the somatic reproductive system and ventral nerve cord, and least common in the brain of *G. bimaculatus*.

2.2 Molecular evolution of sex-biased genes

2.2.1 Rates of evolution

We aimed to assess whether and how evolutionary pressures on protein sequence divergence, measured as dN/dS, varied with sex-biased gene expression. Unlike *Drosophila*, *Gryllus* is currently an emerging model genus with limited genomic resources outside the recent *G. bimaculatus* genome (Ylla et al., 2021). Thus, to measure dN/dS, we generated and assembled novel RNA-seq data for its sister species *G. assimilis* to obtain a CDS list for that organism (Table S2). Two-species assessments of dN/dS have been repeatedly shown to be an effective means to study divergence of sex-biased genes (*cf.* (Mank et al., 2007; Baines et al., 2008; Meisel, 2011; Assis et al., 2012; Whittle & Extavour, 2017; Jaquiery et al., 2018) including for organisms with few available genomes, as is the case with *Gryllu*s. Details of the *G. assimilis* assembly, including BUSCO scores (Seppey, Manni et al. 2019), and ORF predictions (Min et al., 2005) are provided in Text File S1. Following reciprocal BLASTX (Altschul et al., 1990) between *G. bimaculatus* and *G. assimilis* CDS and retention of genes with unsaturated dN and dS values (<1.5) after alignment, we identified 7,220 high confidence *G. bimaculatus*-*G. assimilis* orthologs that were used for all dN/dS analyses. Across all 7,220 orthologs under study,

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we found that the alignments with gaps removed were on average 68.0% (standard error=0.3%) of the original *G. bimaculatus* CDS length, and that the median dN/dS was 0.1152. The median dN was 0.0042 and median dS was 0.0396, values that were substantially <1, consistent with unsaturated substitution rates and a close phylogenetic relatedness between these two sister *Gryllus* species. Notably, the 90th percentile of dN values was 0.042 and 95th percentile was 0.094, also each well below 1, which facilitates precise ortholog detection (by a protein similarity search, reciprocal BLASTX (Altschul et al., 1997)), and indicates the studied ortholog gene set does not exclude relatively rapidly evolving genes in the genome(s) (that is, includes those with 22-fold higher dN than the median). Further, we found that the percent of all male-biased and female-biased *G. bimaculatus* genes (shown in Fig. 2) respectively that had high confidence orthologs between the two *Gryllus* species was 57.7% and 75.7% for the gonads, 55.2% and 52.1% for the somatic reproductive system, 42.4% and 39.2% for the brain, and 50.3% and 64.2% for the ventral nerve cord. Of note, the fact that we detected the fewest orthologs for the brain is suggestive of rapid protein sequence evolution of sex-biased genes in that tissue, which typically limits ortholog detection between divergent sequences (and/or sometimes may reflect gene losses/gains) (, while the highest detection in ovary-biased genes suggests putatively relatively slow protein sequence evolution. The ortholog datasets were subjected to dN/dS analyses as described below.

To precisely reveal the relationship between sex-biased gene expression for each individual tissue type and dN/dS, we identified genes that were sex-biased in expression in only one of the four female-male paired tissues (gonad, somatic reproductive system, brain or ventral nerve cord) and unbiased in all three remaining tissues in *G. bimaculatus*. These genes are hereafter denoted as tissue-specific sex-biased, or TSSB genes (N_{TSSB} values provided in Table S3). We emphasize that the TSSB status of a gene indicates that there is a tissue-specific sex difference in expression (has female-biased or male-biased status) that is not observed in other tissues (unbiased status in all other tissues), and does not imply that this gene is not expressed in any other tissue. Further, we identified those genes with universally unbiased expression in all four tissues types as a control (N=3,449; Table S3). The vast majority of the 7,220 genes (with orthologs in both species) fell into one of these two categories (94.5% had TSSB or universally unbiased status, while the remainder had mixed statuses among tissues).

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2.2.2 dN/dS of sex-biased genes in the four tissue types and pleiotropy

The dN/dS values of sex-biased$_{TSSB}$ genes for each of the four paired *Gryllus* tissue types under study, and for universally unbiased genes, are shown in Fig. 3A. In turn, for completeness, the dN/dS values of all sex-biased genes for each tissue, regardless of status in other tissues (sex-biased$_{ALL}$), are shown in Fig. 3B. The results show that in this cricket model, testis-biased$_{TSSB}$ genes evolved faster than ovary-biased$_{TSSB}$ and universally unbiased genes (Mann-Whitney U (MWU)-tests P<0.001 and 0.05 respectively, Fig. 3A). Further, sex-biased brain genes, while uncommon (Fig. 2, Table S3), evolved exceptionally rapidly. In particular, we noted faster evolution of the female-biased$_{ALL}$ brain genes than of unbiased$_{ALL}$ genes (MWU-test P=0.047, Fig. 3B). Given the P value is near the cutoff of 0.05, we analysed dN/dS of each brain gene set on a gene-by-gene basis for further scrutiny (see below section "*2.3 Rapid evolution of sex-biased genes from the brain*"). In turn, no statistically significant differences in dN/dS were observed among male-biased, female-biased, or unbiased genes from the somatic reproductive system or ventral nerve cords (using the TSSB genes and universally unbiased genes in Fig. 3A, or using ALL genes per tissue type in Fig. 3B (MWU-tests P>0.05)). In this regard, it is evident that the primary molecular evolutionary patterns in this cricket system include the rapid evolution of testis-biased genes and of sex-biased brain genes, particularly female-biased brain genes.

We next assessed the expression breadth across tissues (using nine tissues, the four paired female and male tissues and the male accessory glands, see Methods), as a proxy for pleiotropy, or multifunctionality of a gene, which is thought to strengthen purifying selection and in turn may restrict adaptive evolutionary potential (Otto, 2004; Larracuente et al., 2008; Mank et al., 2008; Mank & Ellegren, 2009; Meisel, 2011; Assis et al., 2012; Dean & Mank, 2016; Whittle & Extavour, 2017). Genes were categorized into bins based on expression at >5 FPKM in 1-2, 3-4, 5-6, and 7-9 tissues (note: we choose a direct determination of expression breadth, rather than an index (Duret & Mouchiroud, 2000; Haerty et al., 2007; Meisel, 2011), see also (Yanai et al., 2005)). As shown in Fig. 4A, when studying all 7,220 genes with high confidence orthologs, we found that the rate of evolution of *Gryllus* genes was strongly inversely correlated with expression breadth. The lowest dN/dS values were found in genes transcribed in 7-9 tissues under study (median dN/dS=0.096), and the highest in genes expressed in 1-2 tissues (median=0.221, Ranked ANOVA and Dunn's paired contrasts P<0.05). Further, as indicated in

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Fig. 4B, with respect to sex-biased gene expression, we found that testis-biased$_{TSSB}$ genes had markedly lower expression breadth than ovary-biased$_{TSSB}$ genes and than universally unbiased genes (MWU-tests P<0.001). Female-biased$_{TSSB}$ brain genes had the smallest median expression breadth of all studied categories, which despite their low N value (Table S3), was statistically significantly lower than that of the universally unbiased genes (MWU-test P=0.021, Fig. 4B). Thus, this suggests a plausible connection between rapid protein sequence evolution and pleiotropy for sex-biased genes from the brain and gonad, either due to relaxed constraint in itself, and/or due to an associated freedom to evolve functional changes under low purifying constraint (see below section "*2.7 Evidence of A History of Positive Selection in Sex-Biased Gonadal and Brain Genes"*). In the following sections, we focus in detail on the dN/dS patterns for sex-biased genes in the brain and the reproductive system in Fig. 3 and Fig. 4, and consider further the putative roles of pleiotropy and positive selection in affecting their molecular evolution.

2.3 Rapid evolution of sex-biased genes from the brain

With respect to the brain, female-biased$_{TSSB}$ genes had markedly higher median dN/dS values (median=0.295) than male-biased$_{TSSB}$ genes (0.203, Fig. 3A), although that contrast was not statistically significant (MWU-test P=0.739). This may reflect the low statistical power of this comparison due to the rarity of genes with sex-biased$_{TSSB}$ brain status (Table S3). When studying all genes with sex-biased$_{ALL}$ expression in the brain, regardless of their expression status in other tissues (Fig. 3B), we found that the 20 female-biased$_{ALL}$ brain genes had substantially higher median dN/dS values (median= 0.245) than the 45 male-biased$_{ALL}$ (0.169) and the unbiased$_{ALL}$ brain-expressed genes (0.115), wherein its contrast to the unbiased set was, as aforementioned, statistically significant (MWU-test P=0.047). Thus, the statistical tests suggest there are significant patterns in the brain (Fig. 3AB). Nonetheless, given the growing recognition that P-values alone may not always provide a full perspective to discern important biological patterns (Amrhein et al., 2019), particularly for samples with small sizes (such as for sex-biased brain genes studied here, Fig. 2), and given the close proximity of the core P value to 0.05, we aimed to further assess these findings by examining the sex-biased brain$_{ALL}$ genes on a gene-by-gene basis, including their rates of evolution and their putative functions, as shown in Table 1. Using this approach, we show that 11 of the 20 female-biased$_{ALL}$ brain genes (Fig. 3B)

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and 19 of 45 male-biased$_{ALL}$ brain genes had dN/dS values more than two-fold higher (>0.236) than the median observed for universally unbiased genes (median=0.118; this value is shown in Fig. 3A; median across the whole genome=0.115). This close examination of individual genes within each gene set, combined with the observed P-values (Fig. 3), taken together indicate that the sex-biased brain genes share a striking propensity to evolve rapidly as compared to universally unbiased genes and the genome as a whole, with the effect being particularly elevated in the female brain (Fig. 3A, Table 1). While the study of protein evolution of sex-biased brain genes (brains *sensu stricto*, rather than simply heads, or pooled brain-eye tissues as considered by some previous studies (Catalan et al., 2018; Congrains et al., 2018)) remains rare, rapid evolution of female-biased brain genes has been reported in some bird embryos (Mank et al., 2007), and in some autosomal genes in flies (Khodursky et al., 2020). However, an opposite pattern of rapid evolution of male-biased brain genes for several stages of development was reported in humans (Shi et al., 2016). The avian result was interpreted as possibly reflecting selective pressures arising from brain-regulated mating behaviors (Mank et al., 2007). We suggest that this may also be a main factor contributing to the trend of rapid evolution of sex-biased brain genes here for crickets.

We examined the putative GO functions for the sex-biased brain genes (Fig. 3). For this, we used single-direction BLASTX (Altschul et al., 1990) of the *G. bimaculatus* entire CDS list to the CDS of well-studied insect model *D. melanogaster* (Gramates et al., 2017) to identify its putative orthologs, which were assessed in the GO tool DAVID (Huang da et al., 2009) (note that single direction BLASTX was used for functional analysis, rather than the reciprocal BLASTX approach that was used for *G. bimaculatus* and *G. assimilis* contrasts for dN/dS, as we considered the latter approach overly conservative for predictive functional analysis, and that it might limit detection of potential paralogs, in *G. bimaculatus*; see details in Methods). First, we conducted enrichment analyses using all *G. bimaculatus sex*-biased brain genes, regardless of the two-species *Gryllus* ortholog status (N values in Fig. 2). We found that female-biased brain genes were enriched for transcriptional functions and sensory perception, while male-biased brain genes were enriched for proteolysis and neuron remodelling (Table S4). We then identified putative functions of those genes with orthologs that were used in our dN/dS analyses, including ALL (sex-biased) genes and the subset of genes that had TSSB status (Fig. 3AB, Table S3) on a gene-by-gene basis as shown in Table 1 (note: any brain genes that had the same sex-biased

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expression status in the gonad are also shown as gonad sex bias "GSB"). We observed that the predicted functions of female-biased brain genes included involvement in neurotransmission (*AP-1-2β*), apoptosis (*D. melanogaster* ID number CG2681), and DNA binding (CG11403) (Table 1). Remarkably, certain brain-expressed genes were predicted to be involved in sexual processes or organs, including multicellular reproduction (CG10407), inter-male aggressive behavior (*tramtrack*) (Yamamoto et al., 1998) and the ejaculatory bulb (*EbpIII*) (Table 1). Each of these genes had exceptionally elevated dN/dS values of 0.460, 0.384 and 0.244 respectively (Table 1), as compared to the median for universally unbiased genes (median=0.118, Fig. 3A). The fastest evolving female-biased brain gene (dN/dS=0.970) was a putative ortholog of *kekkon-3*, a member of a *kekkon* gene family known to be involved in neuron function and differentiation of the central nervous system in flies (Musacchio & Perrimon, 1996), that is conserved in flies and mosquitoes (MacLaren et al., 2004). Collectively, the genes that are upregulated in the cricket female brain may play significant roles in female behaviors, such as mating functions, possibly contributing to their rapid divergence.

Despite a tendency for accelerated evolution, not every female-biased *G. bimaculatus* brain gene evolved rapidly (Table 1). For instance, one highly constrained gene (GBI_02686-RA, (dN/dS=0 (dN=0 dS=0.041)) was an ortholog match to *D. melanogaster crinkled*, which is involved in hearing (vibration sensing) in both flies and vertebrates (Todi et al., 2005; Boekhoff-Falk & Eberl, 2014). We speculate that a history of strong constraint reflected in dN/dS of this female-biased brain gene could indicate an essential role of negative phonotaxis (potentially relevant to avoiding predators (Schneider et al., 2017)), perhaps an effect enhanced in females. However, the sex-biased expression of this putative *crinkled* gene may also suggest it has a sexual role. A fundamental factor underlying male-female attraction in *G. bimaculatus* is song, which is used by males to attract females (positive phonotaxis), and is thought to be regulated by the auditory neural pathways involving the brain (Lankheet et al., 2017; Sakai et al., 2017). Thus, it is tempting to speculate that the strong purifying selection on this female-biased gene could reflect an important role in receiving male auditory signals for courtship and mating. Further studies in crickets should assess sex-biased gene expression in the brain of males and females from mixed mating populations (virgin males and females were studied herein, see Methods) to identify brain-related auditory genes potentially involved in mating. Questions of interest for future work include whether they tend to be highly conserved in sequence, and/or whether some

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may exhibit adaptive changes possibly due to neural-related mating behaviors. Studies in related crickets (*Teleogryllus*) have suggested that neural genes involved in mating, including those involved in acoustics, may have key roles in early stages of male or female development (Kasumovic et al., 2016), and be associated with sex-related behavioral plasticity and abrupt adaptive evolutionary changes (Pascoal et al., 2020). Thus, acoustics, mating, and neural gene evolution may often be intrinsically tied. Additional valuable future directions could include study of sex-biased expression in the male and female auditory organs located on the tibia of the forelegs in crickets (Lankheet et al., 2017; Schneider et al., 2017), in the antennae, which are involved in male-female attraction and male-male aggression and contain neurons involved in sex-related pheromonal signalling (Murakami & Itoh, 2003; Yoritsune & Aonuma, 2012; Boekhoff-Falk & Eberl, 2014), and in the terminal abdominal ganglion, which has been linked to mating behaviors (Sakai et al., 2017). These types of follow-up studies in *G. bimaculatus* will help further identify and evaluate the evolutionary roles of brain and neural genes linked to mating and sex-related auditory and pheromonal signalling in this taxon.

With regard to the male-biased brain genes, a range of predicted functions were observed. For instance, multiple genes were associated with phagocytosis (six of 45 genes), and early-stage development (three genes). In addition, some genes had predicted sexual roles. In particular, a putative *G. bimaculatus* ortholog (GBI_17358-RA) of a *D. melanogaster* ejaculatory bulb protein *EbpIII* had a dN/dS value of 0.449, which was nearly four-fold higher than the median for universally unbiased genes (0.118, Table 1). This same *EbpIII* related gene (GBI_17358-RA) was also found to be testis-biased in expression (Table 1), which is consistent with putatively significant roles in both brain and testicular functions in *G. bimaculatus*. As described above, a different *G. bimaculatus* gene (GBI_17348-RA) that was also an ortholog match to *D. melanogaster EbpIII* was sex-biased in the female-brain (dN/dS=0.243, Table 1), suggesting the possibility that there are two distinct paralogs to this gene, which may have different roles in male and female brains in crickets (note that the *G. bimaculatus* to *D. melanogaster* BLASTX used for functional analysis was one-directional, and thus allowed more than one cricket gene to match a single *D. melanogaster* gene, see Methods). These two genes matching *EbpIII*, one biased in the male-brain and the other in the female brain, are candidates to be involved in male-female attraction, mating or sexual behaviors. In *D. melanogaster*, while the exact functions of *EbpIII* remain under assessment, its key predictive classifications include olfactory function,

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post-mating behavior, and mating plugs (flybase.org, (Gramates et al., 2017)), further suggesting a possible function in male-female brain mediated sexual behaviors in *G bimaculatus*. We also discovered that the male-biased brain genes included a putative ortholog of *Angiotensin converting enzyme*, a gene whose functions include involvement in *D. melanogaster* spermatid nucleus differentiation and sperm individualization (Hurst et al., 2003). This gene had a dN/dS value of 0.236, which is double the median of universally unbiased genes (Table 1). In this regard, multiple male-biased brain genes exhibit rapid protein-level divergence and thus are candidates to have potential sex-related roles in this taxon.

While the tendency for rapid protein sequence evolution of sex-biased brain genes in Table 1 could largely result from relaxed purifying constraint and neutral protein sequence changes, as suggested by their low pleiotropy (Fig. 4B), the low pleiotropy could in principle also act to accelerate protein changes by more readily allowing adaptive functional changes (Otto, 2004; Larracuente et al., 2008; Mank et al., 2008; Mank & Ellegren, 2009; Meisel, 2011; Assis et al., 2012; Dean & Mank, 2016; Whittle & Extavour, 2017). We suggest here that several features of the mating biology of *G. bimaculatus* might cause episodic adaptive evolution and underlie the high dN/dS values observed herein (see also below section "*2.7 Evidence of A History of Positive Selection in Sex-Biased Gonadal and Brain Genes*"). For instance, *G. bimaculatus* exhibits aggressive male-male fighting and mate guarding (Vedenina & Shestakov, 2018; Gee, 2019) and males transfer larger spermatophores to females when in the company of rival males (Lyons & Barnard, 2006). Such behaviors are likely mediated by the male brain. This could, in principle, lead to sexual selection pressures on the male-biased brain genes, which might give rise to adaptive changes in dN/dS. It is also feasible that inter-locus sexual conflict could contribute to the tendency for rapid evolution of both sets of male- and female-biased brain genes (Koene et al., 2013; Mank et al., 2013; Pennell et al., 2016). In other words, it is possible that aggressive male-male behaviors in *G. bimaculatus* (Vedenina & Shestakov, 2018; Gee, 2019), directed by male-biased brain genes, may negatively affect female fitness. This might be predicted to lead to an adaptive response in female-biased brain genes (e.g., genes regulating the behavior of removal of spermatophores of certain males by females after mating (Bateman et al., 2001)), causing an evolutionary "arms race" that could in theory accelerate evolution of proteins of both types of genes (Ellegren & Parsch, 2007; Mank et al., 2013). Taken together, we suggest that there are several plausible mechanisms related to mating biology of this taxon that may

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underlie the observed patterns for sex-biased brain genes (Table 1), mediated by low pleiotropy and, in turn, an enhanced potential for adaptive evolution.

A key aspect of future research should include studies of male and female brains in courtship and mating environments, given that the brain likely regulates these sex-related behaviors in *Gryllus* including song, sexual attraction, copulation and aggression (Matsumoto & Sakai, 2000; Haberkern & Hedwig, 2016; Sakai et al., 2017), and that brain expression has been found to differ between sexes under mating conditions in other insects such as *Drosophila* (based on expression analysis of combined whole head-thorax expression in males and females in that study (Fowler et al., 2019)). We anticipate that in crickets under courtship and mating environments, more genes, in addition to those identified in for virgins (Table 1), may exhibit sex-biased expression given the intense male competition (Vedenina & Shestakov, 2018; Gee, 2019) and the propensity for female-choice in this taxon (Bateman et al., 2001; Zhemchuzhnikov et al., 2017). In turn, future research in these crickets may allow further testing of the notion that mating behaviors may underlie the rapid protein sequence evolution of brain genes, and thus ultimately possibly contribute to processes such as reproductive isolation and speciation.

It should be recognized that while sex-biased brain genes, by definition, exhibit differences in gene expression between the female and male brain, these sex biases may reflect differences in cellular expression and/or allometric scaling differences in male and female brains. As an example, the female-biased brain gene *crinkled* (Table 1) may be more highly expressed in all female than male brain cells, the female brain may typically contain more cells that express this gene (Montgomery & Mank, 2016), and/or the gene may be more highly expressed in cells from a particular sub-region(s) of the brain (Tuller et al., 2008), whereby the size or cell composition of the subsections may vary between females and males (Montgomery & Mank, 2016). Further studies of gene expression, and the allometry of subsections of the brain in males and females, would be needed to distinguish among these possibilities, and to better understand the factors underlying differences in male and female brain expression.

2.4 Rates of Evolution of Sex-biased Genes from the Reproductive System

2.4.1 Rapid evolution of testis-biased genes

With respect to sex-biased expression in the gonads and dN/dS, which has been more commonly studied as compared to the brain in insects, we observed marked differences in rates

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of protein sequence evolution among sex-biased$_{TSSB}$ genes. First, dN/dS decreased progressively from testis-biased$_{TSSB}$ (median=0.128), to universally unbiased genes (median=0.118) to ovary-biased genes (median=0.097, each paired MWU-test P<0.05; see also Fig. 3B). Thus, the rate differences were most marked between testis-biased$_{TSSB}$ and ovary-biased$_{TSSB}$ genes, with intermediate values for those with universally unbiased expression. The tendency for rapid evolution of testis-biased genes in this cricket concurs with patterns observed for *Drosophila* (Zhang et al., 2004; Proschel et al., 2006; Ellegren & Parsch, 2007; Zhang et al., 2007; Jiang & Machado, 2009; Meisel, 2011; Assis et al., 2012; Perry et al., 2015; Grath & Parsch, 2016; Whittle & Extavour, 2019) (see results in a related fly (Congrains et al., 2018)), and recent findings in beetles (*Tribolium castaneum*) (Whittle et al., 2020). However, the results are opposite to the rapid evolution of ovary-biased (or ovary-specific) genes previously reported in the mosquitoes *Aedes* and *Anopheles* (Papa et al., 2017; Whittle & Extavour, 2017). In this regard, it is worth considering possible reasons for variation in the effects of sex-biased gonadal expression among these insect taxa.

Given that *Gryllus* (Orthoptera) is a distant outgroup to the two Diptera groups (*Drosophila* and *Aedes/Anopheles*) and the Coleoptera (*Tribolium*) (Misof et al., 2014) it may be suggested, based on the collective anecdotal evidence, that there could be a shared ancestral effect of testis-biased expression in *Drosophila-Tribolium-Gryllus* (Zhang et al., 2004; Ellegren & Parsch, 2007; Harrison et al., 2015; Whittle et al., 2020)) and a derived effect of rapid evolution of ovary-biased (or ovary-specific) genes in *Aedes/Anopheles* (Papa et al., 2017; Whittle & Extavour, 2017). Under this hypothesis, the pattern observed for studied *Aedes* and *Anopheles* species would be a derived feature, and could reflect variation in mating biology among these insects. For example, although both *Drosophila* and *Aedes aegypti* (the *Aedes* species studied in (Whittle & Extavour, 2017)) are polyandrous and thus prone to sperm competition, the polyandry is thought to be relatively weak in the mosquitoes (Helinski et al., 2012). Further, this mosquito can exhibit intensive male swarming during courtship that may involve female-female mosquito competition and/or male-mate choice (Oliva et al., 2014; Whittle & Extavour, 2017). In addition, nonporous mating plugs are formed in the female mosquito reproductive tract after mating, which prevent sperm competition (Oliva et al., 2014) and thus differ both from the mating plugs formed in *Drosophila*, which allows sperm transfer from competitor males (Manier et al., 2010; Avila et al., 2015), and from observations of

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complete sperm mixing from multiple males in *Gryllus* (Simmons, 1986). Any of these mating-related features could in principle give rise to sexual selection and the relatively faster evolution of ovary-biased than testis-biased genes in mosquitoes (Whittle & Extavour, 2017), and not in the other studied insects. In addition, relaxed purifying selection, possibly due to low pleiotropy, may be more common for ovary-biased genes in the mosquitoes (Whittle & Extavour, 2017), as inferred for testis-biased (or male-biased) genes in some organisms, including flies (Allen et al., 2018; Ghiselli et al., 2018) , and suggested for the crickets studied here (Fig. 4B). Studies in even more insect models, particularly in monogamous versus polyandrous species (Harrison et al., 2015), and in additional insects with various degrees of male-male or female-female competition and with and without impermeable mating plugs (Whittle & Extavour, 2017), would help elucidate whether and how and why the effects of sex-biased transcription on protein evolution vary among insects.

Functional predictions of testis-biased$_{TSSB}$ and ovary-biased$_{TSSB}$ genes in *G. bimaculatus* are shown in Table 2 (using *D. melanogaster* orthologs and GO clustering). Testis-biased$_{TSSB}$ genes were predicted to be preferentially involved in cilium functions, potentially reflecting roles in sperm motility (Trotschel et al., 2019). Ovary-biased$_{TSSB}$ genes were particularly involved in fundamental processes such as transcription functions. Thus, the former may be linked to specialized functions of the male gonad, and sperm functionality, while the latter may include genes involved in broader functions in addition to their roles in the female gonad. In terms of GO functions of the universally unbiased genes, these genes were preferentially involved in core cellular and nuclear functions including protein structure (coiled coil), nucleotide binding and splicing (Table S5), differing from more specialized functions of testis-biased genes.

It is worth mentioning that in Fig. 3A, while testis-biased$_{TSSB}$ genes had higher dN/dS values than ovary-biased$_{TSSB}$ genes and than the universally unbiased genes, they did not exhibit any statistically significant differences with respect to the male-biased genes from the three other tissues, including from the brain (MWU-tests $P>0.05$). Significantly, however, given the much greater abundance of testis-biased$_{TSSB}$ genes than male-biased$_{TSSB}$ genes from other tissues (8- to 65- fold more common, Fig. 2, Table S3), it may be inferred that testis-biased gene expression plays a substantial role in shaping the portion of the genome that is rapidly evolving in *G. bimaculatus*.

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2.4.2 Sex-biased gonadal expression in G. assimilis

While our main target for expression analyses was *G. bimaculatus*, and *G. assimilis* was used primarily as a reference point to measure rates of protein divergence, we considered the degree of conservation of gene expression between the two species for the 7,220 genes with orthologs for the gonads (which had the largest N values of all tissues, Table S3). The results are shown in Fig. S2 and are described in Text File S1. We observed that the finding of elevated dN/dS of testis-biased versus ovary-biased genes was robust to whether the sex-biased status (testis-biased, ovary-biased) was observed in one species or was conserved in both of these species. Thus, testis-biased expression in one species (i.e., *G. bimaculatus* or *G. assimilis*, Fig. S2) is sufficient to predict elevated pairwise dN/dS.

2.4.3 *Possible influence of the faster-X effect*

The faster-X theory contends that genes located on the X-chromosome evolve faster than those on autosomes in male heterogametic XY systems due to rapid fixation of recessive beneficial mutations in hemizygous males (or the Z-chromosome in WZ systems) (Charlesworth et al., 1987). A faster-X effect could also possibly result from relaxed selection on the X-chromosome as compared to autosomes due to lower effective population size (Parsch & Ellegren, 2013). The former cause of a faster-X effect may be evidenced by rapid evolution of male-biased (or typically testis-biased) genes as compared to female-biased and unbiased genes, while the absence of this relationship among sex-biased genes may suggest relaxed selection (Mank et al., 2010a; Parsch & Ellegren, 2013). Given that the recently available and large (1.66 Gbp) *G. bimaculatus* genome remains on scaffolds in this non-traditional model (Ylla et al., 2021), that hypothesis cannot yet be explicitly tested, unlike in insect taxa with widely available and intensively studied genomes (e.g. *Drosophila, Tribolium* (Mank et al., 2010b; Whittle et al., 2020)). Nonetheless, it is worthwhile to consider whether the faster-X effect could contribute to any of the results herein. A recent study of the faster-X effect in beetles (*Tribolium*, an X/Y system) found weak or absent male dosage compensation in the gonads of that taxon, which was associated with an excess of female-biased gonadal genes on the X-chromosome, and the X-chromosome exhibited lower dN/dS than the autosomes (Whittle et al., 2020). These observations suggested an absence of a faster-X effect in *Tribolium*, possibly mediated by low gonadal dosage compensation and rarity of X-linked male-biased genes. A weak faster-X effect

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has been suggested in *Drosophila* (Mank et al., 2010b; Meisel & Connallon, 2013; Avila et al., 2014; Charlesworth et al., 2018), possibly due to poor dosage compensation in gonads of that taxon (Gu & Walters, 2017; Argyridou & Parsch, 2018). In the XX (female) and X0 (male) system of aphids, a faster-X effect was observed, believed to arise under the selective non-neutral model (Jaquiery et al., 2018), and thus presumably male dosage compensation. Thus, this faster-X pattern could in principle also occur in the XX and X0 system of *G. bimaculatus* (Yoshimura et al., 2006). In this context, given that studied crickets and locusts (Camacho et al., 2015; Pascoal et al., 2020) including *G. bimaculatus* (Yoshimura et al., 2006) have cytologically relatively large X-chromosomes compared to the autosomes, we suggest that under specific circumstances, a faster-X effect could possibly give rise to the rapid evolution of testis-biased genes (as compared to ovary-biased and universally unbiased) found herein. Specifically, if there is full gonadal dosage compensation (or overcompensation) on the X chromosome in males in this cricket species then that may cause a high concentration of male-biased gonadal genes on the X chromosome. If there are few testis-biased genes on autosomes, then a faster-X effect could contribute at least partly to the observed patterns of highest dN/dS in testis-biased genes, with lower values for ovary-biased and unbiased genes (Fig. 3), a pattern expected under a selection-based faster-X effect (Parsch & Ellegren, 2013). Importantly, however, as here we have sex-biased expression data from the brain, we also suggest from our findings (Fig. 3, Table 1) that if brain genes are preferentially linked to the X chromosome and exhibit full dosage compensation, this could contribute to rapid evolution of male-biased brain genes (relative to unbiased genes), but could not give rise to the rapid evolution of female-biased brain genes, given that those genes are not monosomic (not X0) in females, excluding a putative role of a faster-X effect. Further studies will thus be valuable to deciphering whether the faster-X effect, and gonadal and brain dosage compensation, may contribute in some manner towards the observed rapid evolution of the testis-biased genes and male-biased brain genes in the cricket model.

2.5 Sex-biased genes from the somatic reproductive system

In contrast to the gonad, the lack of differences in dN/dS of male-biased$_{TSSB}$ and female-biased$_{TSSB}$ genes, and between those groups and the universally unbiased genes, for the somatic reproductive system (MWU-tests P>0.05, Fig. 3A; and when using ALL genes, Fig. 3B) is surprising, given the roles of these sexual tissues in reproductive success and fitness, including

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for the female tissues (oviducts, spermathecae, and bursa). Few comparable insect data of sex-biases in somatic reproductive system tissues are available. Some specific genes involved in the female reproductive tract in *Drosophila* have been linked to rapid and/or adaptive evolution, which may be due to their dynamic roles in receiving and maintaining sperm after mating (Swanson & Vacquier, 2002; Swanson et al., 2004) (note: see section "*2.7 Evidence of A History of Positive Selection in Sex-Biased Gonadal and Brain Genes*" which suggests a small number of female somatic reproductive system genes evolve adaptively). However, a separate assessment of genes broadly defined as female reproductive tract proteins in *D. melanogaster* (based on expression data from mixed or mated flies) showed those genes exhibited slow protein evolution (dN/dS), below the genome-wide average (Haerty et al., 2007). Our results from unmated *Gryllus* suggest no differences in dN/dS between female-biased$_{TSSB}$ somatic reproductive system genes and the universally unbiased genes or the genome as a whole (Fig. 3).

It is also notable that markedly fewer genes were sex-biased in expression in the somatic reproductive system as compared to the gonads (Fig. 2). One possible reason is that there may be an inherent variation in expression among individuals for the male somatic reproductive system (which had the least strongly correlated FPKM among replicates of all nine tissue types, Fig S1H), such that a consistent male to female difference in expression may be less apt to be observed for those tissues. Another possibility is that the gonads in adults are continuously supporting the dynamic process of gametogenesis (Pauli & Mahowald, 1990; Williamson & Lehmann, 1996) causing high female and male expression differentiation (Fig. 2), while the somatic reproductive system, particularly in unmated tissues as studied here, may be less dynamic, and thus exhibit less potential for differential transcription between males and females.

2.6 Rapid divergence of genes from the male accessory glands and seminal fluid proteins

For thoroughness in the study of reproductive structures, given that genes from the male accessory glands, including seminal fluid protein (SFPs), have been linked to rapid evolution in species of *Drosophila* (Haerty et al., 2007; Sepil et al., 2019), and in some identified cricket SFPs based on partial gene sets attained from assembled reproductive transcriptome sequences for species such as *G. firmus*, *G. pennsylvanicus* and *Allonemobius fasciatus* (Andres et al., 2006; Braswell et al., 2006; Andres et al., 2013), we assessed expression and evolution of such genes in *G. bimaculatus*. The findings for the male accessory glands (described in detail in Text

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File S1 and Table S6) showed that *G. bimaculatus* genes that had expression solely in the male accessory glands rarely had a high confidence ortholog in its sister species *G. assimilis*. Thus, this suggests a history of rapid evolution potentially so extensive that it prevents protein similarity detection by these methods, and/or a history of lineage-specific gene losses or gains of genes involved in this particular sexual tissue (Haerty et al., 2007; Tautz & Domazet-Loso, 2011).

For the study of SFPs, we used the recently available gene list of 134 SFPs from the species *D. melanogaster* as the reference, as this species has the most intensively-studied insect genome and expression/proteomics data for the identification of these genes (Sepil et al., 2019). The results are described in Text File S1 and Table S7. We found that only 20 *D. melanogaster* SFP genes had identifiable putative orthologs in *G. bimaculatus* (14.9%). Seven of those were included among the subset of 7,220 genes with between-species orthologs in the two species of *Gryllus* (note the stringent criteria used for the intra-*Gryllus* ortholog matches, see Methods). The dN/dS values of these seven genes are shown in Table 3; all were above the genome-wide median dN/dS value (0.115). Positive selection was indicated for the gene matching an odorant binding SFP protein *Obp56g*, with dN/dS>1 (Table 3). Together, we conclude that the putative SFPs in the crickets studied herein have evolved very rapidly, a feature shared with SFPs of *D. melanogaster* (Haerty et al., 2007; Sepil et al., 2019), and that could be due to their potential subjection to sex-related selection pressures. For instance, in flies SFPs may enhance sperm competitive ability in the female reproductive tract or egg release from the ovary (Heifetz et al., 2000; Fedorka et al., 2011), and males may alter relative production of different SFPs when exposed to male rivals (Fedorka et al., 2011). If similar types of mechanisms of sexual selection exist in crickets, then they could contribute to fast evolution of SFP genes. Another potentially significant behavioural factor in *G. bimaculatus*, is the tendency of females to preferentially retain deposited spermatophores of certain (larger) males (Simmons, 1986; Bateman et al., 2001), which comprises a mechanism of female-choice in this species (Bateman et al., 2001), potentially accelerating SFP evolution.

2.7 Evidence of A History of Positive Selection in Sex-Biased Gonadal and Brain Genes

Finally, we considered the incidences of positive selection among the 7,220 genes with between-species *Gryllus* orthologs. Gene-wide dN/dS>1 was taken as evidence of positive

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selection (Swanson et al., 2001; Torgerson et al., 2002; Nielsen et al., 2005; Clark et al., 2006; Yang, 2007; Hunt et al., 2011; Buschiazzo et al., 2012; Ghiselli et al., 2018; Hill et al., 2019)). The use of dN/dS>1 across a gene is a conservative means to identify positive selection (Swanson et al., 2001; Buschiazzo et al., 2012), as nonsynonymous codon changes should be sufficiently common to cause the ratio to exceed 1. We found that 1.63% of all the 7,220 *G. bimaculatus*-*G. assimilis* gene orthologs (N=118 genes) showed dN/dS>1.

We then considered whether dN/dS values of the sex-biased$_{TSSB}$ genes from the gonad (Table 4), which had the highest N values of all tissues analysed (Table S3), were consistent with the aforementioned hypothesis that reduced gene pleiotropy, or expression breadth (and thus purifying selection), may lead to an enhanced opportunity for functional evolution of genes (Otto, 2004; Larracuente et al., 2008; Mank et al., 2008; Mank & Ellegren, 2009; Meisel, 2011; Assis et al., 2012; Whittle et al., 2020). We found that the percent of genes with positive selection increased from ovary-biased$_{TSSB}$ genes (1.02%, 19 of 1,858) to universally unbiased genes (1.91%, 66 of 3,449) and testis-biased$_{TSSB}$ genes (2.09%, 22 of 1,055; Chi^2 P with Yates' correction was <0.05 for each paired contrast to ovary-biased$_{TSSB}$ genes, Table 4). In turn, expression breadth of these genes decreased from all ovary-biased$_{TSSB}$ (average expression breadth of 7.97±0.04 (standard error)), to universally unbiased (6.95±0.05) and to testis-biased$_{TSSB}$ genes (5.90±0.18 tissues; (MWU-tests P<0.001 for each of three paired contrasts (Fig. 4B). Strikingly, the differences were even more magnified in the subset of genes with dN/dS>1 shown in Table 4, with markedly higher average expression breadth (2.5 fold) for ovary-biased$_{TSSB}$ (6.74±0.74) than for testis-biased$_{TSSB}$ (2.73±0.72) genes (MWU-test P<0.05, Table 4). These patterns observed using whole-gene dN/dS values in this cricket system provide empirical data consistent with the theoretical proposition that that the fewer tissues a gene is expressed in, the more its adaptive evolutionary potential may be enhanced, likely by relaxing purifying selection that may be imposed by multiple cross-tissue functions (Otto, 2004; Larracuente et al., 2008; Mank et al., 2008; Mank & Ellegren, 2009; Meisel, 2011). Our data thus specifically suggest that this hypothesis can apply to sex-biased genes (Mank & Ellegren, 2009). We note nonetheless that given the close relatedness between the two *Gryllus* species studied here, this may potentially elevate observed dN/dS for some genes (Mugal et al., 2014) (see below section "*2.8 Close relatedness of Gryllus taxa*"), and thus further studies of dN.dS using additional

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Gryllus species as data becomes available will help test the rigor of these patterns across the genus.

We further assessed whether there was evidence of positive selection for sex-biased brain genes, which were much less common than those from the gonad (Table S3, Fig. 2). The only gene with whole-gene dN/dS >1 (=3.675, GBI_19557-RB, Table 1) was of unknown function and was expressed primarily in the male brain (number tissues with >5 FPKM =1 tissue). Thus, this result is also concordant with adaptive evolution facilitated by low pleiotropy. The female-biased brain gene with the highest dN/dS of 0.9735 matched *D. melanogaster kekkon3*. This value (near one) could suggest a history of neutral evolution, but may also reflect positive selection at multiple codon sites in that gene; we cannot distinguish between these two possibilities using gene-wide dN/dS.

As a follow-up supplemental analysis to gene-wide dN/dS, we examined positive selection among species at specific codon sites using branch-site analysis (with *G. bimaculatus* as the target branch) (Yang, 2007), based on three-way alignments of *G. bimaculatus*, *G. assimilis* and an available cricket outgroup species *Laupala kohalensis* (Blankers et al., 2018; Ylla et al., 2021). The results are described in Text File S1 and Table S8. It should be emphasized the assessment is inherently very conservative given it only includes the subset of genes with high confidence three-way reciprocal orthologs among the three species (that is, only 26,7% of the 7,220 genes with orthologs in the two *Gryllus* species had three-species orthologs, see Methods, and Text File S1). Nonetheless, we found that a non-negligible portion of the male- and female-biased$_{TSSB}$ gonadal genes showed positive selection (≥9.6%), and that only minor variation was observed between groups, perhaps due to the conserved nature of the analysis (Table S8). Three sex-biased brain genes that were studied in Table 1 (among ten of the 65 in Table 1 that had three-species orthologs available for analysis, Table S8) showed positive selection using branch-site analysis (GBI_05906-RA, GBI_09477-RB, GBI_05452-RB, Table S8). This result is consistent with the hypothesis of a history of adaptive evolution in the brain, possibly elevating dN/dS (Fig. 3AB).

It is worth noting that for the branch-site analysis, we found that a small subset of genes that were female-biased in the somatic reproductive system (six of 33 genes (18.2%) with three-species orthologs), which includes the reproductive tract and/or spermathecae, tended to evolve adaptively using branch-site analysis (Table S8). In this context, the result suggests that a small

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number of female-biased reproductive system genes may evolve adaptively, potentially in response to sexual selection pressures, as suggested in flies (Swanson et al., 2004; Prokupek et al., 2008), in this cricket taxon. Further studies using more powerful branch-site positive selection tests (Yang, 2007) as genomic data emerge in even more crickets, and/or population genetics analysis of frequencies of codon mutations (McDonald & Kreitman, 1991), will further reveal the scale of positive selection at specific codon sites in the sex-biased genes from various tissues. Such analyses will also allow further evaluation of the link between positive selection (dN/dS>1) and gene pleiotropy that was suggested for gonads using the gene-wide dN/dS herein (Table 4), and permit additional evaluation of this relationship for the brain, which had relatively few sex-biased genes with which to consider this specific relationship (of dN/dS>1 and pleiotropy) using gene-wide dN/dS (Fig. 2, Table 1).

2.8 Close relatedness of *Gryllus* taxa

The study of closely related species such as *G. bimaculatus* and *G. assimilis* as conducted herein allows for examination of genes with unsaturated substitutions and thus accurate measures of dN/dS (see section "*2.2.1 Rates of evolution*" for median dN and dS values*)*, as applied in other studies within insect genera (Zhang et al., 2007; Baines et al., 2008; Meisel, 2011; Assis et al., 2012; Whittle & Extavour, 2017; Jaquiery et al., 2018). We note that very close relationships have been proposed in theory for some unicellular and viral systems (Rocha et al., 2006; Kryazhimskiy & Plotkin, 2008), and possibly some multicellular eukaryotes (Mugal et al., 2020), to potentially affect dN/dS due to a short time periods to fix or remove polymorphic mutations (see also counterevidence from (Gibson & Eyre-Walker, 2019)). In the present study, we propose that our core results are apt to be minimally influenced by any potential such effect, given that all genomic analyses were conducted in an identical manner for sex-biased genes from all tissues and for the same two species, and thus the time of divergence is the same throughout the two genomes. Nonetheless, follow-up studies in more species of *Gryllus* should consider the degree of relatedness in potentially shaping dN/dS among taxa. Further, the combined analyses of interspecies dN/dS data with polymorphism-level genomics data will allow discernment of whether any degree of nonsynonymous mutations may remain polymorphic (yet unfixed) between closely related cricket species. Unlike widely studied insects such as *Drosophila* that have vast available polymorphism and species genomic datasets (Wang et al., 2015; Gramates et

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al., 2017), studies testing hypotheses on the relationship between time since divergence and dN/dS in *Gryllus* will become feasible as more species genomes, as well as genome-wide population level datasets in multiple species, become available in the future.

3 CONCLUSIONS

Here we have conducted comprehensive assessment of sex-biased gene expression in reproductive and nervous system tissues, and revealed their relationships to potential pressures on protein sequence evolution, in a cricket model system. We have demonstrated the consistent tendency for rapid evolution of sex-biased brain genes, particularly female-biased brain genes, (Fig. 3, Table 1), and of male-biased genes from the gonad, in *G. bimaculatus*. Further, our data suggest a direct link between low pleiotropy and elevated dN/dS of sex-biased genes in the brain and the gonad (Fig. 3, Fig. 4) that may reflect relaxed purifying selection, which in turn may permit elevated instances of positive selection (Table 4) (Otto, 2004; Larracuente et al., 2008; Mank et al., 2008; Mank & Ellegren, 2009; Meisel, 2011). We speculate that the features of this cricket's mating biology may give rise to sexual selection and thus contribute at least partly towards the accelerated evolution of the sex-biased brain genes, and male-biased gonadal genes, in this taxon.

Suggested significant directions for future studies include the following approaches: First, research on sex-biased gene expression from different brain regions may further decipher its relationship to protein evolution (Tuller et al., 2008), and the possible roles of allometric scaling (Montgomery & Mank, 2016). Second, investigation of the involvement of sex-biased brain genes in gene pathways and networks, and their expression breadth across even more tissue types than those studied herein, may help elucidate why they often evolve rapidly. Third, similar studies as conducted herein in more divergent *Gryllus* species and in other genera such as *Drosophila* may help reveal whether the relationships between sex-biased expression and dN/dS varies over evolutionary time (Mugal et al., 2014), and/or is affected by the turnover in sex-biased expression status (Zhang et al., 2007; Whittle & Extavour, 2019). Fourth, additional studies should consider potential differences in sex-biased expression of alternately spliced mRNAs among taxa, as high confidence genome-wide splicing variants are further refined for the recent *G. bimaculatus* genome (Ylla et al., 2021) and as whole genome and large-scale RNA-seq data (allowing splicing predictions) emerge in other comparable *Gryllus* species, some

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variants of which may be involved in sexual differentiation (Nagoshi et al., 1988; Wexler et al., 2019). Fifth, refinement of the *G. bimaculatus* genome to discern the sex chromosomes and autosomes and gene localizations, combined with expression data, will allow further testing of any putative role of dosage compensation and faster-X effect on rapid evolution of sex-biased genes from the brains and gonads (Parsch & Ellegren, 2013; Whittle et al., 2020). Sixth, the sequencing of additional *Gryllus* genomes and/or generation of population sequence data for *G. bimaculatus* may allow MacDonald-Kreitman tests and more powerful positive branch-site selection tests (McDonald & Kreitman, 1991; Yang, 2007) than available herein, particularly for those with small sample sizes of sex-biased genes such as the brain. Seventh, assessment of sex-biased gene expression in *G. bimaculatus* adult males and females should be conducted in a courtship environment with male-male rivals, and/or with multiple females exposed to few males (female-female competition), and include assessments of the putative roles of acoustics-related genes (*cf.* (Kasumovic et al., 2016; Pascoal et al., 2018; Pascoal et al., 2020). Given that mating behaviors may be largely mediated by gene expression in male and female brains in *Gryllus* (Matsumoto & Sakai, 2000; Haberkern & Hedwig, 2016; Sakai et al., 2017), and in other insects such as *Drosophila* (Fowler et al., 2019), such follow-up research in the brain will be valuable to better understand the potential ties between mating behaviors, sex-biased expression, and protein sequence evolution. Finally, the study of sex-biased expression in brains and gonads among insects that have known differences in their mating biology (including for example variation in testis size, sperm mixing, degree of female-female competition, mate choice (cf. Harrison et al., 2015)), including among additional species of *Gryllus*, will help further decipher whether and how protein sequence evolutionary rates may be shaped by these various mechanisms of sexual selection across a phylogeny.

4 METHODS

4.1 Biological samples and RNA-seq

For our RNA-seq assessment of *G. bimaculatus* we isolated the male and female gonad, somatic reproductive system, brain and ventral nerve cord (shown in Fig. 1, Table S1; Fig. 1A,B schematic is based on (Kumashiro & Sakai, 2001) and simplified from Fox 2001; http://lanwebs.lander.edu/faculty/rsfox/invertebrates/acheta.html)). A ninth, unpaired

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reproductive tissue type, the male accessory gland, was also extracted, as its gene expression has been linked to protein sequence changes (Swanson et al., 2001; Clark & Swanson, 2005; Haerty et al., 2007), and it provides an additional sexual tissue type for the analysis of cross-tissue expression breadth. Further, we considered that its inclusion in the male somatic reproductive system sample might overwhelm the transcript population of that tissue type upon RNA-seq, and make it incomparable to the female reproductive system.

The rearing of specimens for tissue sampling was as follows: post hatching, wild type *G. bimaculatus* nymphs from an existing laboratory colony inbred for at least 14 years were grown at 29°C until adulthood in well-ventilated plastic cages on a 12 hour day/ 12 hour night cycle (Kainz et al., 2011). Plastic cages were provided with egg cartons for shelter, and the animals were fed with ground cat food (Purina item model number 178046) and water. Prior to the final nymphal molt, animals were sexed based on the presence (female) or absence (male) of an ovipositor and separated into male and female cages to avoid any mating and thus obtain virgin samples. Dissections were then performed on the unmated adults within a week after their final molt, by briefly anesthetizing the animals on ice for 5-10 minutes prior to dissection. Different tissue types (gonad, somatic reproductive system, brain, ventral nerve cord, male accessory reproductive glands) were dissected per animal using sterile equipment wiped with ethanol and RNaseZap (Ambion, catalog number AM9780), in ice-cold 1x Phosphate Buffer Saline (PBS), and the tissue cleaned of any unwanted contaminating material. Each tissue was then transferred immediately into individual 1.5ml Eppendorf tubes containing 500µl of pre-frozen Trizol (Thermo Fisher, catalog number 15596018) on dry ice, and stored at -80°C until further use. RNA extractions, library processing and RNA-seq were then performed as described previously (Whittle et al., 2020). The same procedure was conducted for specimens of *G. assimilis*, which was used to obtain RNA-seq data for an assembly to be used for dN/dS analysis (Table S2; which also included a carcass tissue type). The *G. assimilis* eggs were obtained from the Hedwig lab (University of Cambridge, UK) and reared to adulthood, using the same animal husbandry protocols as published previously for *G. bimaculatus* (Mito & Noji, 2008; Kainz et al., 2011; Kochi et al., 2016).

The complete RNA-seq data are available at the Short Read Archive (SRA) under the project identifier PRJNAXXXXXX (under species name and Study ID SRPXXXXXX, Tables S1, S2). The RNA-seq reads (76bp in length) for each sample were trimmed of adapters and poor

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quality bases using the program BBduk available from the Joint Genome Institute (https://jgi.doe.gov/data-and-tools/bbtools/) using default parameters.

4.2 CDS of *G. bimaculatus* and sex-biased gene expression

The expression level for each of 15,539 *G. bimaculatus* genes was determined by mapping reads from each RNA-seq dataset per tissue to the full CDS list using Geneious Read Mapper (Kearse et al., 2012), a program previously found to be as effective as other common read mappers (cf. Whittle et al., 2020). We compared expression between males and females for the gonad, somatic reproductive system, brain, and ventral nerve cord using the program DESeq2, which uses the mapped reads across biological replicates and the negative binomial distribution to quantify the P-values of expression differences (Love et al., 2014). In addition, the degree of sex-biased expression per gene was determined using the ratio of average in FPKM across the replicates for female and male tissues. Any gene that had a two-fold or greater ratio in average expression in one sex (as compared to the other) and a statistically significant P-value from DESeq ($P<0.05$) as well as a FPKM of at least 1 in one tissue type was defined as sex-biased (*cf.* on a two-fold cutoff (Proschel et al., 2006; Assis et al., 2012; Whittle & Extavour, 2017; Parker et al., 2019; Whittle & Extavour, 2019; Whittle et al., 2020). Given the use of two biological replicates for the large-scale RNA-seq (Table S1) and our high threshold cutoff (two-fold), the identification of sex-biased genes herein is conservative. All genes not defined as sex-biased per tissue type were defined as unbiased (Zhang et al., 2010; Whittle & Extavour, 2017; Darolti et al., 2018; Parker et al., 2019; Whittle & Extavour, 2019; Whittle et al., 2020), which herein includes all genes with less than two-fold sex-biased expression or with <1 FPKM (including undetectable expression, and apt not to play sex-related roles) in both females and males (Whittle & Extavour, 2017; Whittle & Extavour, 2019; Whittle et al., 2020). Thus, all 15,539 genes belonged to one of these three categories per tissue (Fig. 2). We note that 95.4% of the 15,539 *G. bimaculatus* genes were expressed in at least one of the nine tissues (Additional file 1; Table S1), suggesting the vast majority of genes have putative roles in some or all of these studied tissues.

4.3 Assembly of *G. assimilis* RNA-seq data and protein sequence divergence analysis

4.3.1 Assembly of reads

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To study dN/dS, we generated and assembled RNA-seq CDS for the *G. bimaculatus* sister species *G. assimilis*. We assembled all the trimmed reads for the RNA-seq datasets of *G. assimilis* shown in Table S2. For this, the *G. assimilis* reads were *de novo* assembled into contigs using Trinity (Grabherr et al., 2011) set to default parameters using Galaxy (https://usegalaxy.org/). We then identified CDS using the PlantTribes pipeline tools (Wall et al., 2008). To assess the completeness of the assembled transcriptome, we used BUSCO 3.0.1 (Seppey et al., 2019) to reveal the percentage of the single-copy CDS that was observed in the standardized Arthropod conserved gene set, and as employed in gVolante ((Nishimura et al., 2017) https://gvolante.riken.jp/analysis.html). To refine the CDS for *G. assimilis* we then assessed each CDS in ORF predictor, using its downloadable Perl script (Min et al., 2005), to identify the highest quality reading frame per sequence. In ORF predictor, we used the option to include the best-hit (lowest e-value) BLASTX alignment (conducted in BLAST+ v2.7.1, https://blast.ncbi.nlm.nih.gov) (Altschul et al., 1997) of *G. assimilis* versus the reference *G. bimaculatus* protein database (i.e., its translated 15,539 CDS) to define reading frames, and retained all *G. assimilis* CDS that were at least 150bp long and had a start codon.

It is worth noting that while paired-end reads have often been used for RNA-seq assembly, transcriptome assemblies from single-end reads have been successfully employed to obtain CDS (not requiring isoforms) as studied herein (Gongora-Castillo & Buell, 2013; Hibsh et al., 2015). Further to this point, single-end reads have even been applied for *de novo* assemblies in non-traditional model systems (Gongora-Castillo & Buell, 2013; Hibsh et al., 2015). Here, we have the additional advantage of a closely related reference genome to *G. assimilis*, namely *G. bimaculatus* (Ylla et al., 2021), to identify and confirm orthologs.

4.3.2 Ortholog identification and dN/dS

Gene ortholog matches between *G. bimaculatus* and *G. assimilis* were identified using reciprocal BLASTX of the full CDS list between the two species in the program BLAST+ v2.7.1 (https://blast.ncbi.nlm.nih.gov) (Altschul et al., 1997). Genes having an identical best match sequence (lowest e-value) in both forward and reverse contrasts and $e<10^{-6}$ were defined as putative orthologs. The identified orthologous gene sequences in *G. bimaculatus* and *G. assimilis* were aligned by codons using MUSCLE (Edgar, 2004) set to default parameters in the program Mega-CC v7 (Kumar et al., 2012) and gaps removed. Removal of divergent regions from

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alignments, despite partial loss of sequence regions, improves quantification of protein divergence; thus, highly divergent segments were removed using the program GBlocks v. 0.91b set at default parameters (Castresana, 2000; Talavera & Castresana, 2007).

Using the aligned *G. bimaculatus* and *G. assimilis* CDS, we employed yn00 of PAML using the Yang and Nielson 2000 substitution model, which comprises a maximum likelihood method that accounts for codon usage biases (Yang & Nielsen, 2000; Yang, 2007), to measure dN, dS, and dN/dS (Yang, 2007) (note that dN/dS measures using Yang and Neilson 2000 (Yang & Nielsen, 2000) were strongly correlated to those using other models; e.g., values from the Pamilo and Bianchi 1993 method (Pamilo & Bianchi, 1993) had Spearman's R=0.95 $P<2X10^{-7}$). Values of dN/dS reflect the standardized rate of protein sequence evolution (dN to dS), whereby values >1, =1, and <1 suggest a prevalent history of positive selection, neutral evolution and purifying selection respectively (Yang, 2007). However, even when <1 for gene-wide measures of dN/dS, elevated values suggest greater roles of positive selection and/or relaxed purifying selection (Swanson & Vacquier, 2002; Buschiazzo et al., 2012). Genes that were best matches by reciprocal BLASTX, and for which both values of dN and dS values were <1.5 (and thus were unsaturated (Castillo-Davis et al., 2004; Treangen & Rocha, 2011)), were defined as high confidence orthologs (N=7,220) between *G. bimaculatus* and *G. assimilis* for dN/dS analysis. The low dN values between these two cricket species, which were typically well below 1 as described in the Results and Discussion, should allow precise detection of orthologs, not only for single copy genes but also for duplicated genes, which can evolve rapidly (Demuth et al., 2006; Hahn et al., 2007). Overall, given the strict criteria we used for identification of high confidence orthologs, the paired alignments and dN, dS, and dN/dS measures herein are conservative.

4.4 Positive selection tests

In our core assessments of gene-wide dN/dS using paired contrasts of *G. bimaculatus* and *G. assimilis* from the same genus, any values >1 were interpreted as an indicator of a potential history of positive selection (Swanson et al., 2001; Torgerson et al., 2002; Nielsen et al., 2005; Clark et al., 2006; Yang, 2007; Hunt et al., 2011; Buschiazzo et al., 2012; Ghiselli et al., 2018). For analysis of genes with dN/dS >1 (Table 4), we included only those genes with both dN and dS>0.

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In addition to this assessment, we examined positive selection at specific codon sites for the *G. bimaculatus* branch using branch-site analysis in codeml of PAML (Yang, 2007). As an outgroup species was required for this assessment, we used the recently available assembled and annotated *Laupala kohalensis* genome (Blankers et al., 2018; Ylla et al., 2021). Three-way orthologs between *G. bimaculatus*, *G. assimilis*, and *L. kohalensis* were identified using reciprocal BLASTX ($e<10^{-6}$) among each of the three paired species contrasts (our criterion was that for each *G. bimaculatus*-*G. assimilis* paired ortholog, the same matching *L. kohalensis* CDS must be found using reciprocal BLASTX to *G. bimaculatus* CDS and to *G. assimilis* CDS). Genes were aligned by codons using all three-species CDS and filtered using GBlocks (Castresana, 2000; Talavera & Castresana, 2007) and gaps removed as described in "*Ortholog identification and dN/dS*" (note: alignments using this relatively distant outgroup were conducted independently of the paired *Gryllus* alignments). The phylogeny was ((*G. bimaculatus*, *G. assimilis*), *L. kohalensis*) and was unrooted for the PAML free-ratio analysis (Model=1, NSsites=0 in codeml) that was used to determine dN and dS per branch. Only those genes with dN<3, and with dS<3 (Mank et al., 2007) in the *L. kohalensis* branch were defined as high confidence orthologs and used for branch-site analysis (unlike the two-species contrasts within *Gryllus*, which were more closely related and had a cut-off of 1.5). For genes meeting these criteria, branch-site positive selection was assessed on the *G. bimaculatus* branch using Chi-square values for 2XΔln Likelihood ($P<0.05$) between models without (null hypothesis) and with (alternate hypothesis) positive selection (Model=2, NSsites=2, omega fixed versus estimated) as described in the PAML manual (Yang, 2007). We note that our stringent approach to defining three-way orthologs, and the distance of the outgroup, favors study of the more conservative portion of the genome for branch-site analysis. Further, some studies have suggested that branch-site analysis can lack sensitivity to detect functional changes (Nozawa et al., 2009; Toll-Riera et al., 2011), and/or may generate false positives (Nozawa et al., 2009; Wisotsky et al., 2020), the latter likely being sensitive to the stringency of alignment. We thus aimed to control this factor by our strict approach to this assessment (excluding genes with any signs of dN or dS saturation).

4.5 Sex-biased expression between *G. bimaculatus* and *G. assimilis*

As a supplementary analysis to our core assessment of sex-biased expression in our main target taxon *G. bimaculatus*, we also examined sex-biased transcription of genes in *G. assimilis*.

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For this, expression was determined using its assembled CDS list (described in Text file S1 and in the Results and Discussion) and the RNA-seq data (Table S2), as was described for *G. bimaculatus*. We focused on the gonads, which had the highest number of sex-biased genes among tissues in *G. bimaculatus* (Fig. 2). We assessed the correlation in expression for orthologs between the two species using Spearman's ranked correlations. In turn, we determined those genes with conserved and variable sex-biased expression status in the gonads between species, and their relationships to dN/dS.

4.6 Gene ontology

Gene ontology (GO) was characterized using the tool DAVID (Huang da et al., 2009). For this, we identified orthologs to *G. bimaculatus* in the insect model *D. melanogaster*, which has the most well-studied insect genome to date (CDS v6.24 available from www.flybase.org (Gramates et al., 2017)), using BLASTX (https://blast.ncbi.nlm.nih.gov) (Altschul et al., 1997) and the best match (lowest e-value with cut off of $e<10^{-3}$ of *D. melanogaster*). Single direction BLASTX (Altschul et al., 1997) with *G. bimaculatus* CDS as the query to the *D. melanogaster* protein database was used for these assessments (unlike for the more rigorous reciprocal BLASTX analysis used to identify orthologs between the two *Gryllus* species for dN/dS analysis), as we considered that the latter would be overly conservative between these insects from different orders for the purpose of functional characterization and analysis, and might prevent detection of putative paralogs in the crickets. *D. melanogaster* gene identifiers were input into DAVID (Huang da et al., 2009) to obtain gene putative GO functions and/or classifications. The *D. melanogaster* BLASTX searches were used solely for identification of putative orthologs to ascertain putative GO functions for our sex-biased and unbiased genes (and for putative SFP identification), and were not used for any dN/dS analysis, which was restricted to genes aligned within the crickets.

4.7 Seminal fluid proteins

As a complementary reproductive assessment in *G. bimaculatus*, we examined seminal fluid proteins (SFPs). We used *D. melanogaster* as our reference for SFP identification given this species has the most well-studied insect genome, transcriptome, and proteome to date, thus providing a more complete profile than the currently available smaller and likely partial SFP lists

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for crickets, which were obtained using transcriptomics and/or proteomics from reproductive tissues (Andres et al., 2013; Simmons et al., 2013). A recent proteome analysis of sexual structures in *D. melanogaster* confirmed functions for 134 SFPs (Sepil et al., 2019). Thus, we identified potential orthologs in *G. bimaculatus* to these SFPs in *D. melanogaster* (using single direction BLASTX as conducted for GO analysis) and assessed whether those genes had high confidence orthologs (and their dN/dS values) between *G. bimaculatus* and *G. assimilis*.

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Conflict of Interest

Authors declare no conflict of interest.

Author contributions

XX, XX and XX designed the study. XX reared G. *bimaculatus* and *G. assimilis* and sampled tissues for RNA-seq. XX analyzed the data and wrote the manuscript with contributions by XX and XX. All authors read and approved the final manuscript.

References

Allen, S. L., Bonduriansky, R., & Chenoweth, S. F. (2018). Genetic constraints on microevolutionary divergence of sex-biased gene expression. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences,* **373**.

Altschul, S. F., Gish, W., Miller, W., Myers, E. W., & Lipman, D. J. (1990). Basic local alignment search tool. *Journal of Molecular Biology,* **215,** 403-410.

Altschul, S. F., Madden, T. L., Schaffer, A. A., Zhang, J., Zhang, Z., Miller, W., & Lipman, D. J. (1997). Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Research,* **25,** 3389-402.

Amrhein, V., Greenland, S., & McShane, B. (2019). Scientists rise up against statistical significance. *Nature,* **567,** 305-307.

bioRxiv preprint doi: https://doi.org/10.1101/2020.07.07.192039; this version posted April 26, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Andres, J. A., Larson, E. L., Bogdanowicz, S. M., & Harrison, R. G. (2013). Patterns of transcriptome divergence in the male accessory gland of two closely related species of field crickets. *Genetics,* **193,** 501-13.

Andres, J. A., Maroja, L. S., Bogdanowicz, S. M., Swanson, W. J., & Harrison, R. G. (2006). Molecular evolution of seminal proteins in field crickets. *Molecular Biology and Evolution,* **23,** 1574-84.

Arbeitman, M. N., Fleming, A. A., Siegal, M. L., Null, B. H., & Baker, B. S. (2004). A genomic analysis of *Drosophila* somatic sexual differentiation and its regulation. *Development,* **131,** 2007-21.

Argyridou, E., & Parsch, J. (2018). Regulation of the X Chromosome in the Germline and Soma of *Drosophila melanogaster* Males. *Genes (Basel),* **9,** 242 (doi.org/10.3390/genes9050242).

Assis, R., Zhou, Q., & Bachtrog, D. (2012). Sex-biased transcriptome evolution in *Drosophila*. *Genome Biology and Evolution,* **4,** 1189-200.

Avila, F. W., Wong, A., Sitnik, J. L., & Wolfner, M. F. (2015). Don't pull the plug! the *Drosophila* mating plug preserves fertility. *Fly (Austin),* **9,** 62-7.

Avila, V., Marion de Proce, S., Campos, J. L., Borthwick, H., Charlesworth, B., & Betancourt, A. J. (2014). Faster-X effects in two *Drosophila* lineages. *Genome Biology and Evolution,* **6,** 2968-82.

Baines, J. F., Sawyer, S. A., Hartl, D. L., & Parsch, J. (2008). Effects of X-linkage and sex-biased gene expression on the rate of adaptive protein evolution in *Drosophila*. *Molecular Biology and Evolution,* **25,** 1639-50.

Bateman, P. W., Giliston, L. N., & Ferguson, J. W. H. (2001). Male size and sequential mate preference in the cricket *Gryllus bimaculatus*. *Animal Behavior,* **61,** 631-637.

Biswas, K., Chakraborty, S., Podder, S., & Ghosh, T. C. (2016). Insights into the dN/dS ratio heterogeneity between brain specific genes and widely expressed genes in species of different complexity. *Genomics,* **108,** 11-7.

Blankers, T., Oh, K. P., Bombarely, A., & Shaw, K. L. (2018). The Genomic Architecture of a Rapid Island Radiation: Recombination Rate Variation, Chromosome Structure, and Genome Assembly of the Hawaiian Cricket Laupala. *Genetics,* **209,** 1329-1344.

Boekhoff-Falk, G., & Eberl, D. F. (2014). The *Drosophila* Auditory System. *Wiley Interdiscip Rev Dev Biol,* 179–191.

Braswell, W. E., Andres, J. A., Maroja, L. S., Harrison, R. G., Howard, D. J., & Swanson, W. J. (2006). Identification and comparative analysis of accessory gland proteins in Orthoptera. *Genome,* **49,** 1069-80.

Buschiazzo, E., Ritland, C., Bohlmann, J., & Ritland, K. (2012). Slow but not low: genomic comparisons reveal slower evolutionary rate and higher dN/dS in conifers compared to angiosperms. *BMC Evolutionary Biology,* **12,** 8.

Camacho, J. P., Shaw, M. W., Cabrero, J., Bakkali, M., Ruiz-Estevez, M., Ruiz-Ruano, F. J., Martin-Blazquez, R., & Lopez-Leon, M. D. (2015). Transient Microgeographic Clines during B Chromosome Invasion. *American Naturalist,* **186,** 675-81.

Castillo-Davis, C. I., Bedford, T. B., & Hartl, D. L. (2004). Accelerated rates of intron gain/loss and protein evolution in duplicate genes in human and mouse malaria parasites. *Molecular Biology and Evolution,* **21,** 1422-7.

Castresana, J. (2000). Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Molecular Biology and Evolution,* **17,** 540-52.

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Catalan, A., Hutter, S., & Parsch, J. (2012). Population and sex differences in *Drosophila melanogaster* brain gene expression. *BMC Genomics,* **13,** 654.

Catalan, A., Macias-Munoz, A., & Briscoe, A. D. (2018). Evolution of Sex-Biased Gene Expression and Dosage Compensation in the Eye and Brain of *Heliconius* Butterflies. *Molecular Biology and Evolution,* **35,** 2120-2134.

Charlesworth, B., Campos, J. L., & Jackson, B. C. (2018). Faster-X evolution: Theory and evidence from *Drosophila*. *Molecular Ecology,* **27,** 3753-3771.

Charlesworth, B., Coyne, J., & Barton, N. (1987). The relative rates of evolution of sex chromosomes and autosomes. *American Naturalist,* **130,** 113-146.

Clark, N. L., Aagaard, J. E., & Swanson, W. J. (2006). Evolution of reproductive proteins from animals and plants. *Reproduction,* **131,** 11-22.

Clark, N. L., & Swanson, W. J. (2005). Pervasive adaptive evolution in primate seminal proteins. *PLoS Genetics,* **1,** e35.

Congrains, C., Campanini, E. B., Torres, F. R., Rezende, V. B., Nakamura, A. M., Oliveira, J. L. d., Lima, A. L. A., Chahad-Ehlers, S., Sobrinho, I. S., & Brito, R. A. d. (2018). Evidence of Adaptive Evolution and Relaxed Constraints in Sex-Biased Genes of South American and West Indies Fruit Flies (Diptera: Tephritidae). *Genome Biology and Evolution,* **10,** 380-395.

Dalton, J. E., Kacheria, T. S., Knott, S. R., Lebo, M. S., Nishitani, A., Sanders, L. E., Stirling, E. J., Winbush, A., & Arbeitman, M. N. (2010). Dynamic, mating-induced gene expression changes in female head and brain tissues of *Drosophila melanogaster*. *BMC Genomics,* **11,** 541.

Darolti, I., Wright, A. E., Pucholt, P., Berlin, S., & Mank, J. E. (2018). Slow evolution of sex-biased genes in the reproductive tissue of the dioecious plant Salix viminalis. *Molecular Ecology,* **27,** 694-708.

Dauwalder, B. (2008). Systems behavior: of male courtship, the nervous system and beyond in *Drosophila*. *Current Genomics,* **9,** 517-24.

Dean, R., & Mank, J. E. (2016). Tissue Specificity and Sex-Specific Regulatory Variation Permit the Evolution of Sex-Biased Gene Expression. *The American Naturalist,* **188,** E74-E84.

Demuth, J. P., De Bie, T., Stajich, J. E., Cristianini, N., & Hahn, M. W. (2006). The evolution of mammalian gene families. *PloS One,* **1,** e85.

Donoughe, S., & Extavour, C. G. (2016). Embryonic development of the cricket *Gryllus bimaculatus*. *Developmental Biology,* **411,** 140-56.

Dorus, S., Busby, S. A., Gerike, U., Shabanowitz, J., Hunt, D. F., & Karr, T. L. (2006). Genomic and functional evolution of the *Drosophila melanogaster* sperm proteome. *Nature Genetics,* **38,** 1440-5.

Drapeau, M. D., Radovic, A., Wittkopp, P. J., & Long, A. D. (2003). A gene necessary for normal male courtship, yellow, acts downstream of fruitless in the *Drosophila melanogaster* larval brain. *Journal of Neurobiology,* **55,** 53-72.

Duret, L., & Mouchiroud, D. (2000). Determinants of substitution rates in mammalian genes: expression pattern affects selection intensity but not mutation rate. *Molecular Biology and Evolution,* **17,** 68-74.

Edgar, R. C. (2004). MUSCLE: a multiple sequence alignment method with reduced time and space complexity. *BMC Bioinformatics,* **5,** 113.

Ellegren, H., & Parsch, J. (2007). The evolution of sex-biased genes and sex-biased gene expression. *Nature Reviews Genetics,* **8,** 689-98.

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Fedorka, K. M., Winterhalter, W. E., & Ware, B. (2011). Perceived sperm competition intensity influences seminal fluid protein production prior to courtship and mating. *Evolution,* **65,** 584-90.

Fowler, E. K., Bradley, T., Moxon, S., & Chapman, T. (2019). Divergence in Transcriptional and Regulatory Responses to Mating in Male and Female Fruitflies. *Scientific Reports,* **9,** 16100.

Gee, D. (2019). The effects of weaponry and mating experience on the level and outcome of agonistic interactions in male field crickets, *Gryllus bimaculatus* (orthoptera: Gryllidae). *PhD Thesis,* University of Derby, UK.

Ghiselli, F., Iannello, M., Puccio, G., Chang, P. L., Plazzi, F., Nuzhdin, S. V., & Passamonti, M. (2018). Comparative Transcriptomics in Two Bivalve Species Offers Different Perspectives on the Evolution of Sex-Biased Genes. *Genome Biology and Evolution,* **10,** 1389-1402.

Gibson, B., & Eyre-Walker, A. (2019). Investigating Evolutionary Rate Variation in Bacteria. *Journal of Molecular Evolution,* **87,** 317-326.

Gongora-Castillo, E., & Buell, C. R. (2013). Bioinformatics challenges in de novo transcriptome assembly using short read sequences in the absence of a reference genome sequence. *Natural Product Reports,* **30,** 490-500.

Grabherr, M. G., Haas, B. J., Yassour, M., Levin, J. Z., Thompson, D. A., Amit, I., Adiconis, X., Fan, L., Raychowdhury, R., Zeng, Q., Chen, Z., Mauceli, E., Hacohen, N., Gnirke, A., Rhind, N., di Palma, F., Birren, B. W., Nusbaum, C., Lindblad-Toh, K., Friedman, N., & Regev, A. (2011). Full-length transcriptome assembly from RNA-Seq data without a reference genome. *Nature Biotechnology,* **29,** 644-52.

Gramates, L. S., Marygold, S. J., Santos, G. D., Urbano, J. M., Antonazzo, G., Matthews, B. B., Rey, A. J., Tabone, C. J., Crosby, M. A., Emmert, D. B., Falls, K., Goodman, J. L., Hu, Y., Ponting, L., Schroeder, A. J., Strelets, V. B., Thurmond, J., & Zhou, P. (2017). FlyBase at 25: looking to the future. *Nucleic Acids Research,* **45,** D663-D671.

Grath, S., & Parsch, J. (2012). Rate of amino acid substitution is influenced by the degree and conservation of male-biased transcription over 50 myr of *Drosophila* evolution. *Genome Biology and Evolution,* **4,** 346-359.

Grath, S., & Parsch, J. (2016). Sex-Biased Gene Expression. *Annual Review of Genetics,* **50,** 29-44.

Gu, L., & Walters, J. R. (2017). Evolution of Sex Chromosome Dosage Compensation in Animals: A Beautiful Theory, Undermined by Facts and Bedeviled by Details. *Genome Biology and Evolution,* **9,** 2461-2476.

Haberkern, H., & Hedwig, B. (2016). Behavioural integration of auditory and antennal stimulation during phonotaxis in the field cricket *Gryllus bimaculatus*. *Journal of Experimental Biology,* **219,** 3575-3586.

Haerty, W., Jagadeeshan, S., Kulathinal, R. J., Wong, A., Ram, K. R., Sirot, L. K., Levesque, L., Artieri, C. G., Wolfner, M. F., Civetta, A., & Singh, R. S. (2007). Evolution in the fast lane: rapidly evolving sex-related genes in *Drosophila*. *Genetics,* **177,** 1321-1335.

Hahn, M. W., Han, M. V., & Han, S. G. (2007). Gene family evolution across 12 Drosophila genomes. *PLoS Genetics,* **3,** e197.

Harrison, P. W., Wright, A. E., Zimmer, F., Dean, R., Montgomery, S. H., Pointer, M. A., & Mank, J. E. (2015). Sexual selection drives evolution and rapid turnover of male gene

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expression. *Proceedings of the National Academy of Sciences of the United States of America,* **112,** 4393-8.

Heifetz, Y., Lung, O., Frongillo, E. A., Jr., & Wolfner, M. F. (2000). The *Drosophila* seminal fluid protein Acp26Aa stimulates release of oocytes by the ovary. *Current Biology,* **10,** 99-102.

Helinski, M. E., Valerio, L., Facchinelli, L., Scott, T. W., Ramsey, J., & Harrington, L. C. (2012). Evidence of polyandry for *Aedes aegypti* in semifield enclosures. *American Journal of Tropical Medicine and Hygiene,* **86,** 635-41.

Hibsh, D., Schori, H., Efroni, S., & Shefi, O. (2015). De novo transcriptome assembly databases for the central nervous system of the medicinal leech. *Sci Data,* **2,** 150015.

Hill, T., Koseva, B. S., & Unckless, R. L. (2019). The Genome of Drosophila innubila Reveals Lineage-Specific Patterns of Selection in Immune Genes. *Molecular Biology and Evolution,* **36,** 1405-1417.

Huang da, W., Sherman, B. T., & Lempicki, R. A. (2009). Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nature Protocols,* **4,** 44-57.

Huber, F. (1963). The role of the central nervous system in orthoptera during the co-ordination and control of stridulation. *Acoustic Behaviour of Animals, Elsevier Publishing Company, Amsterdam London New York; R.G. Busnel (Ed.),* 440-488.

Hunt, B. G., Ometto, L., Wurm, Y., Shoemaker, D., Yi, S. V., Keller, L., & Goodisman, M. A. (2011). Relaxed selection is a precursor to the evolution of phenotypic plasticity. *Proceedings of the National Academy of Sciences of the United States of America,* **108,** 15936-41.

Hurst, D., Rylett, C. M., Isaac, R. E., & Shirras, A. D. (2003). The drosophila angiotensin-converting enzyme homologue Ance is required for spermiogenesis. *Developmental Biology,* **254,** 238-47.

Huylmans, A. K., & Parsch, J. (2015). Variation in the X:Autosome Distribution of Male-Biased Genes among Drosophila melanogaster Tissues and Its Relationship with Dosage Compensation. *Genome Biology and Evolution,* **7,** 1960-71.

Ingleby, F. C., Flis, I., & Morrow, E. H. (2014). Sex-biased gene expression and sexual conflict throughout development. *Cold Spring Harbor Perspectives in Biology,* **7,** a017632.

Jacob, P. F., & Hedwig, B. (2016). Acoustic signalling for mate attraction in crickets: Abdominal ganglia control the timing of the calling song pattern. *Behavioural Brain Research,* **309,** 51-66.

Jagadeeshan, S., & Singh, R. S. (2005). Rapidly evolving genes of *Drosophila*: differing levels of selective pressure in testis, ovary, and head tissues between sibling species. *Molecular Biology and Evolution,* **22,** 1793-801.

Jaquiery, J., Peccoud, J., Ouisse, T., Legeai, F., Prunier-Leterme, N., Gouin, A., Nouhaud, P., Brisson, J. A., Bickel, R., Purandare, S., Poulain, J., Battail, C., Lemaitre, C., Mieuzet, L., Le Trionnaire, G., Simon, J. C., & Rispe, C. (2018). Disentangling the Causes for Faster-X Evolution in Aphids. *Genome Biology and Evolution,* **10,** 507-520.

Jiang, Z. F., & Machado, C. A. (2009). Evolution of sex-dependent gene expression in three recently diverged species of *Drosophila*. *Genetics,* **183,** 1175-1185.

Kadener, S., Villella, A., Kula, E., Palm, K., Pyza, E., Botas, J., Hall, J. C., & Rosbash, M. (2006). Neurotoxic protein expression reveals connections between the circadian clock and mating behavior in *Drosophila*. *Proceedings of the National Academy of Sciences of the United States of America,* **103,** 13537-42.

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Kainz, F., Ewen-Campen, B., Akam, M., & Extavour, C. G. (2011). Notch/Delta signalling is not required for segment generation in the basally branching insect *Gryllus bimaculatus*. *Development,* **138,** 5015-26.

Kasumovic, M. M., Chen, Z., & Wilkins, M. R. (2016). Australian black field crickets show changes in neural gene expression associated with socially-induced morphological, life-history, and behavioral plasticity. *BMC Genomics,* **17,** 827.

Kearse, M., Moir, R., Wilson, A., Stones-Havas, S., Cheung, M., Sturrock, S., Buxton, S., Cooper, A., Markowitz, S., Duran, C., Thierer, T., Ashton, B., Meintjes, P., & Drummond, A. (2012). Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics,* **28,** 1647-9.

Khodursky, S., Svetec, N., Durkin, S. M., & Zhao, L. (2020). The evolution of sex-biased gene expression in the Drosophila brain. *Genome Research,* **30,** 874-884.

Kochi, Y., Miyashita, A., Tsuchiya, K., Mitsuyama, M., Sekimizu, K., & Kaito, C. (2016). A human pathogenic bacterial infection model using the two-spotted cricket, *Gryllus bimaculatus*. *FEMS Microbiology Letters,* **363**.

Koene, J. M., Liew, T. S., Montagne-Wajer, K., & Schilthuizen, M. (2013). A syringe-like love dart injects male accessory gland products in a tropical hermaphrodite. *PloS One,* **8,** e69968.

Kryazhimskiy, S., & Plotkin, J. B. (2008). The population genetics of dN/dS. *PLoS Genetics,* **4,** e1000304.

Kulkarni, A., & Extavour, C. G. (2019). The Cricket *Gryllus bimaculatus*: Techniques for Quantitative and Functional Genetic Analyses of Cricket Biology. *Results and Problems in Cell Differentiation,* **Volume 68,** 183-216.

Kumar, S., Stecher, G., Peterson, D., & Tamura, K. (2012). MEGA-CC: computing core of molecular evolutionary genetics analysis program for automated and iterative data analysis. *Bioinformatics,* **28,** 2685-6.

Kumashiro, M., & Sakai, M. (2001). Reproductive behavior in the male cricket *Gryllus bimaculatus* DeGeer: I. Structure and function of the genitalia. *Journal of Experimental Biology,* 1123–1137.

Lankheet, M. J., Cerkvenik, U., Larsen, O. N., & van Leeuwen, J. L. (2017). Frequency tuning and directional sensitivity of tympanal vibrations in the field cricket *Gryllus bimaculatus*. *J R Soc Interface,* **14,** 2017003.

Larracuente, A. M., Sackton, T. B., Greenberg, A. J., Wong, A., Singh, N. D., Sturgill, D., Zhang, Y., Oliver, B., & Clark, A. G. (2008). Evolution of protein-coding genes in *Drosophila*. *Trends in Genetics,* **24,** 114-23.

Love, M. I., Huber, W., & Anders, S. (2014). Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biology,* **15,** 550.

Lyons, C., & Barnard, D. (2006). A learned response to sperm competition in the field cricket, *Gryllus bimaculatus* (de Geer). *Animal Behavior,* **72,** 673-680.

MacLaren, C. M., Evans, T. A., Alvarado, D., & Duffy, J. B. (2004). Comparative analysis of the Kekkon molecules, related members of the LIG superfamily. *Development Genes and Evolution,* **214,** 360-6.

Manier, M. K., Belote, J. M., Berben, K. S., Novikov, D., Stuart, W. T., & Pitnick, S. (2010). Resolving mechanisms of competitive fertilization success in *Drosophila melanogaster*. *Science,* **328,** 354-7.

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Mank, J. E., & Ellegren, H. (2009). Are sex-biased genes more dispensable? *Biology Letters,* **5,** 409-12.

Mank, J. E., Hultin-Rosenberg, L., Axelsson, E., & Ellegren, H. (2007). Rapid evolution of female-biased, but not male-biased, genes expressed in the avian brain. *Molecular Biology and Evolution,* **24,** 2698-706.

Mank, J. E., Hultin-Rosenberg, L., Zwahlen, M., & Ellegren, H. (2008). Pleiotropic constraint hampers the resolution of sexual antagonism in vertebrate gene expression. *American Naturalist,* **171,** 35-43.

Mank, J. E., Nam, K., & Ellegren, H. (2010a). Faster-Z evolution is predominantly due to genetic drift. *Molecular Biology and Evolution,* **27,** 661-70.

Mank, J. E., Vicoso, B., Berlin, S., & Charlesworth, B. (2010b). Effective population size and the Faster-X effect: empirical results and their interpretation. *Evolution,* **64,** 663-74.

Mank, J. E., Wedell, N., & Hosken, D. J. (2013). Polyandry and sex-specific gene expression. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences,* **368,** 20120047.

Matsumoto, Y., & Sakai, M. (2000). Brain control of mating behavior in the male cricket *Gryllus bimaculatus* DeGeer: brain neurons responsible for inhibition of copulation actions. *Journal of Insect Physiology,* **46,** 539-552.

McDonald, J. H., & Kreitman, M. (1991). Adaptive protein evolution at the Adh locus in *Drosophila*. *Nature,* **351,** 652-4.

Meisel, R. P. (2011). Towards a more nuanced understanding of the relationship between sex-biased gene expression and rates of protein-coding sequence evolution. *Molecular Biology and Evolution,* **28,** 1893-900.

Meisel, R. P., & Connallon, T. (2013). The faster-X effect: integrating theory and data. *Trends in Genetics,* **29,** 537-44.

Min, X. J., Butler, G., Storms, R., & Tsang, A. (2005). OrfPredictor: predicting protein-coding regions in EST-derived sequences. *Nucleic Acids Research,* **33,** W677-80.

Misof, B., Liu, S., Meusemann, K., Peters, R. S., Donath, A., Mayer, C., Frandsen, P. B., Ware, J., Flouri, T., Beutel, R. G., Niehuis, O., Petersen, M., Izquierdo-Carrasco, F., Wappler, T., Rust, J., Aberer, A. J., Aspock, U., Aspock, H., Bartel, D., Blanke, A., Berger, S., Bohm, A., Buckley, T. R., Calcott, B., Chen, J., Friedrich, F., Fukui, M., Fujita, M., Greve, C., Grobe, P., Gu, S., Huang, Y., Jermiin, L. S., Kawahara, A. Y., Krogmann, L., Kubiak, M., Lanfear, R., Letsch, H., Li, Y., Li, Z., Li, J., Lu, H., Machida, R., Mashimo, Y., Kapli, P., McKenna, D. D., Meng, G., Nakagaki, Y., Navarrete-Heredia, J. L., Ott, M., Ou, Y., Pass, G., Podsiadlowski, L., Pohl, H., von Reumont, B. M., Schutte, K., Sekiya, K., Shimizu, S., Slipinski, A., Stamatakis, A., Song, W., Su, X., Szucsich, N. U., Tan, M., Tan, X., Tang, M., Tang, J., Timelthaler, G., Tomizuka, S., Trautwein, M., Tong, X., Uchifune, T., Walzl, M. G., Wiegmann, B. M., Wilbrandt, J., Wipfler, B., Wong, T. K., Wu, Q., Wu, G., Xie, Y., Yang, S., Yang, Q., Yeates, D. K., Yoshizawa, K., Zhang, Q., Zhang, R., Zhang, W., Zhang, Y., Zhao, J., Zhou, C., Zhou, L., Ziesmann, T., Zou, S., Li, Y., Xu, X., Zhang, Y., Yang, H., Wang, J., Wang, J., Kjer, K. M., & Zhou, X. (2014). Phylogenomics resolves the timing and pattern of insect evolution. *Science,* **346,** 763-7.

Mito, T., & Noji, S. (2008). The Two-Spotted Cricket *Gryllus bimaculatus*: An Emerging Model for Developmental and Regeneration Studies. *CSH Protoc,* **2008,** pdb emo110.

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Montgomery, S. H., & Mank, J. E. (2016). Inferring regulatory change from gene expression: the confounding effects of tissue scaling. *Molecular Ecology,* **25,** 5114-5128.

Morrow, E. H., & Gage, M. G., H (2001). Sperm competition experiments between lines of crickets producing different sperm lengths. *Proceedings of the Royal Society of London. Series B: Biological Sciences,* **268,** 2281-6.

Mugal, C. F., Kutschera, V. E., Botero-Castro, F., Wolf, J. B. W., & Kaj, I. (2020). Polymorphism Data Assist Estimation of the Nonsynonymous over Synonymous Fixation Rate Ratio omega for Closely Related Species. *Molecular Biology and Evolution,* **37,** 260-279.

Mugal, C. F., Wolf, J. B., & Kaj, I. (2014). Why time matters: codon evolution and the temporal dynamics of dN/dS. *Molecular Biology and Evolution,* **31,** 212-31.

Murakami, S., & Itoh, M. T. (2003). Removal of both antennae influences the courtship and aggressive behaviors in male crickets. *Journal of Neurobiology,* **57,** 110-8.

Musacchio, M., & Perrimon, N. (1996). The Drosophila kekkon genes: novel members of both the leucine-rich repeat and immunoglobulin superfamilies expressed in the CNS. *Developmental Biology,* **178,** 63-76.

Nagoshi, R. N., McKeown, M., Burtis, K. C., Belote, J. M., & Baker, B. S. (1988). The control of alternative splicing at genes regulating sexual differentiation in D. melanogaster. *Cell,* **53,** 229-36.

Naurin, S., Hansson, B., Hasselquist, D., Kim, Y. H., & Bensch, S. (2011). The sex-biased brain: sexual dimorphism in gene expression in two species of songbirds. *BMC Genomics,* **12,** 37.

Nielsen, R., Bustamante, C., Clark, A. G., Glanowski, S., Sackton, T. B., Hubisz, M. J., Fledel-Alon, A., Tanenbaum, D. M., Civello, D., White, T. J., J, J. S., Adams, M. D., & Cargill, M. (2005). A scan for positively selected genes in the genomes of humans and chimpanzees. *PLoS Biology,* **3,** e170.

Nishimura, O., Hara, Y., & Kuraku, S. (2017). gVolante for standardizing completeness assessment of genome and transcriptome assemblies. *Bioinformatics,* **33,** 3635-3637.

Nozawa, M., Suzuki, Y., & Nei, M. (2009). Reliabilities of identifying positive selection by the branch-site and the site-prediction methods. *Proceedings of the National Academy of Sciences of the United States of America,* **106,** 6700-5.

Oliva, C. F., Damiens, D., & Benedict, M. Q. (2014). Male reproductive biology of *Aedes* mosquitoes. *Acta Tropica,* **132 Suppl,** S12-9.

Oliver, T. A., Garfield, D. A., Manier, M. K., Haygood, R., Wray, G. A., & Palumbi, S. R. (2010). Whole-genome positive selection and habitat-driven evolution in a shallow and a deep-sea urchin. *Genome Biology and Evolution,* **2,** 800-14.

Otto, S. P. (2004). Two steps forward, one step back: the pleiotropic effects of favoured alleles. *Proceedings: Biological Sciences,* **271,** 705-14.

Pamilo, P., & Bianchi, N. O. (1993). Evolution of the Zfx and Zfy genes: rates and interdependence between the genes. *Molecular Biology and Evolution,* **10,** 271-81.

Panhuis, T. M., & Swanson, W. J. (2006). Molecular evolution and population genetic analysis of candidate female reproductive genes in *Drosophila*. *Genetics,* **173,** 2039-47.

Papa, F., Windbichler, N., Waterhouse, R. M., Cagnetti, A., D'Amato, R., Persampieri, T., Lawniczak, M. K., Nolan, T., & Papathanos, P. A. (2017). Rapid evolution of female-biased genes among four species of *Anopheles* malaria mosquitoes. *Genome Research,* **27,** 1536-1548.

bioRxiv preprint doi: https://doi.org/10.1101/2020.07.07.192039; this version posted April 26, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Parisi, M., Nuttall, R., Edwards, P., Minor, J., Naiman, D., Lü, J., Doctolero, M., Vainer, M., Chan, C., Malley, J., Eastman, S., & Oliver, B. (2004). A survey of ovary-, testis-, and soma-biased gene expression in *Drosophila melanogaster* adults. *Genome Biology,* **5,** R40.

Parker, D. J., Bast, J., Jalvingh, K., Dumas, Z., Robinson-Rechavi, M., & Schwander, T. (2019). Sex-biased gene expression is repeatedly masculinized in asexual females. *Nat Commun,* **10,** 4638.

Parsch, J., & Ellegren, H. (2013). The evolutionary causes and consequences of sex-biased gene expression. *Nature Reviews Genetics,* **14,** 83-7.

Pascoal, S., Liu, X., Fang, Y., Paterson, S., Ritchie, M. G., Rockliffe, N., Zuk, M., & Bailey, N. W. (2018). Increased socially mediated plasticity in gene expression accompanies rapid adaptive evolution. *Ecology Letters,* **21,** 546-556.

Pascoal, S., Risse, J. E., Zhang, X., Blaxter, M., Cezard, T., Challis, R. J., Gharbi, K., Hunt, J., Kumar, S., Langan, E., Liu, X., Rayner, J. G., Ritchie, M. G., Snoek, B. L., Trivedi, U., & Bailey, N. W. (2020). Field cricket genome reveals the footprint of recent, abrupt adaptation in the wild. *Evol Lett,* **4,** 19-33.

Pauli, D., & Mahowald, A. P. (1990). Germ-line sex determination in *Drosophila melanogaster*. *Trends in Genetics,* **6,** 259-64.

Pennell, T. M., de Haas, F. J., Morrow, E. H., & van Doorn, G. S. (2016). Contrasting effects of intralocus sexual conflict on sexually antagonistic coevolution. *Proceedings of the National Academy of Sciences of the United States of America,* **113,** E978-86.

Perry, J. C., Harrison, P. W., & Mank, J. E. (2015). The Ontogeny and Evolution of Sex-Biased Gene Expression in *Drosophila melanogaster*. *Molecular Biology and Evolution,* **31,** 1206-1219.

Prokupek, A., Hoffmann, F., Eyun, S. I., Moriyama, E., Zhou, M., & Harshman, L. (2008). An evolutionary expressed sequence tag analysis of *Drosophila* spermatheca genes. *Evolution,* **62,** 2936-47.

Proschel, M., Zhang, Z., & Parsch, J. (2006). Widespread adaptive evolution of *Drosophila* genes with sex-biased expression. *Genetics,* **174,** 893-900.

Rocha, E. P., Smith, J. M., Hurst, L. D., Holden, M. T., Cooper, J. E., Smith, N. H., & Feil, E. J. (2006). Comparisons of dN/dS are time dependent for closely related bacterial genomes. *Journal of Theoretical Biology,* **239,** 226-35.

Sakai, M., Kumashiro, M., Matsumoto, Y., Ureshi, M., & Otsubo, T. (2017). Reproductive Behavior and Physiology in the Cricket *Gryllus bimaculatus*. *The Cricket as a Model Organism* 245-269.

Santos, E. M., Kille, P., Workman, V. L., Paull, G. C., & Tyler, C. R. (2008). Sexually dimorphic gene expression in the brains of mature zebrafish. *Comparative Biochemistry and Physiology. Part A: Molecular and Integrative Physiology,* **149,** 314-24.

Schneider, E. S., Romer, H., Robillard, T., & Schmidt, A. K. D. (2017). Hearing with exceptionally thin tympana: Ear morphology and tympanal membrane vibrations in eneopterine crickets. *Scientific Reports,* **7,** 15266.

Sepil, I., Hopkins, B. R., Dean, R., Thezenas, M. L., Charles, P. D., Konietzny, R., Fischer, R., Kessler, B. M., & Wigby, S. (2019). Quantitative Proteomics Identification of Seminal Fluid Proteins in Male *Drosophila melanogaster*. *Molecular and Cellular Proteomics,* **18,** S46-S58.

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Seppey, M., Manni, M., & Zdobnov, E. M. (2019). BUSCO: Assessing Genome Assembly and Annotation Completeness. *Methods in Molecular Biology,* **1962,** 227-245.

Shi, L., Zhang, Z., & Su, B. (2016). Sex Biased Gene Expression Profiling of Human Brains at Major Developmental Stages. *Scientific Reports,* **6,** 21181.

Simmons, L. W. (1986). Female choice in the field cricket *Gryllus bimaculatus (*De Geer). *Animal Behavior,* **34,** 1463-1470.

Simmons, L. W., Tan, Y. F., & Millar, A. H. (2013). Sperm and seminal fluid proteomes of the field cricket Teleogryllus oceanicus: identification of novel proteins transferred to females at mating. *Insect Molecular Biology,* **22,** 115-30.

Small, C. M., Carney, G. E., Mo, Q., Vannucci, M., & Jones, A. G. (2009). A microarray analysis of sex- and gonad-biased gene expression in the zebrafish: evidence for masculinization of the transcriptome. *BMC Genomics,* **10,** 579.

Swanson, W. J., Clark, A. G., Waldrip-Dail, H. M., Wolfner, M. F., & Aquadro, C. F. (2001). Evolutionary EST analysis identifies rapidly evolving male reproductive proteins in *Drosophila*. *Proceedings of the National Academy of Sciences of the United States of America,* **98,** 7375-7379.

Swanson, W. J., & Vacquier, V. D. (2002). The rapid evolution of reproductive proteins. *Nature Reviews: Genetics,* **3,** 137-44.

Swanson, W. J., Wong, A., Wolfner, M. F., & Aquadro, C. F. (2004). Evolutionary expressed sequence tag analysis of *Drosophila* female reproductive tracts identifies genes subjected to positive selection. *Genetics,* **168,** 1457-65.

Talavera, G., & Castresana, J. (2007). Improvement of phylogenies after removing divergent and ambiguously aligned blocks from protein sequence alignments. *Systematic Biology,* **56,** 564-577.

Tautz, D., & Domazet-Loso, T. (2011). The evolutionary origin of orphan genes. *Nature Reviews Genetics,* **12,** 692-702.

Todi, S. V., Franke, J. D., Kiehart, D. P., & Eberl, D. F. (2005). Myosin VIIA defects, which underlie the Usher 1B syndrome in humans, lead to deafness in *Drosophila*. *Current Biology,* **15,** 862-8.

Toll-Riera, M., Laurie, S., & Alba, M. M. (2011). Lineage-specific variation in intensity of natural selection in mammals. *Molecular Biology and Evolution,* **28,** 383-98.

Tomchaney, M., Mysore, K., Sun, L., Li, P., Emrich, S. J., Severson, D. W., & Duman-Scheel, M. (2014). Examination of the genetic basis for sexual dimorphism in the *Aedes aegypti* (dengue vector mosquito) pupal brain. *Biology of Sex Differences,* **5,** 10.

Torgerson, D. G., Kulathinal, R. J., & Singh, R. S. (2002). Mammalian sperm proteins are rapidly evolving: evidence for positive selection in functionally diverse genes. *Molecular Biology and Evolution,* **19,** 1973-1980.

Treangen, T. J., & Rocha, E. P. (2011). Horizontal transfer, not duplication, drives the expansion of protein families in prokaryotes. *PLoS Genetics,* **7,** e1001284.

Trotschel, C., Hamzeh, H., Alvarez, L., Pascal, R., Lavryk, F., Bonigk, W., Korschen, H. G., Muller, A., Poetsch, A., Rennhack, A., Gui, L., Nicastro, D., Strunker, T., Seifert, R., & Kaupp, U. B. (2019). Absolute proteomic quantification reveals design principles of sperm flagellar chemosensation. *EMBO Journal,* e102723.

Tuller, T., Kupiec, M., & Ruppin, E. (2008). Evolutionary rate and gene expression across different brain regions. *Genome Biology,* **9,** R142.

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Vedenina, V. Y., & Shestakov, L. S. (2018). Loser in Fight but Winner in Love: How Does Inter-Male Competition Determine the Pattern and Outcome of Courtship in Cricket *Gryllus bimaculatus*? *Frontiers in Ecology and Evolution,* **27,** https://doi.org/10.3389/fevo.2018.00197.

Wall, P. K., Leebens-Mack, J., Muller, K. F., Field, D., Altman, N. S., & dePamphilis, C. W. (2008). PlantTribes: a gene and gene family resource for comparative genomics in plants. *Nucleic Acids Research,* **36,** D970-6.

Wang, F., Jiang, L., Chen, Y., Haelterman, N. A., Bellen, H. J., & Chen, R. (2015). FlyVar: a database for genetic variation in *Drosophila melanogaster*. *Database: The Journal of Biological Databases and Curation,* doi 10.1093/database/bav079.

Wexler, J., Delaney, E. K., Belles, X., Schal, C., Wada-Katsumata, A., Amicucci, M. J., & Kopp, A. (2019). Hemimetabolous insects elucidate the origin of sexual development via alternative splicing. *Elife,* **8**.

Whittle, C. A., & Extavour, C. G. (2017). Rapid Evolution of Ovarian-Biased Genes in the Yellow Fever Mosquito (*Aedes aegypti*). *Genetics,* **206,** 2119-2137.

Whittle, C. A., & Extavour, C. G. (2019). Selection shapes turnover and magnitude of sex-biased expression in *Drosophila* gonads. *BMC Evolutionary Biology,* **19,** 60 (doi: 10.1186/s12862-019-1377-4).

Whittle, C. A., Kulkarni, A., & Extavour, C. G. (2020). Absence of a faster-X effect in beetles (*Tribolium*, Coleoptera). *G3 (Bethesda),* **10,** 1125–1136.

Williamson, A., & Lehmann, R. (1996). Germ cell development in Drosophila. *Annual Review of Cell and Developmental Biology,* **12,** 365-91.

Wisotsky, S. R., Kosakovsky Pond, S. L., Shank, S. D., & Muse, S. V. (2020). Synonymous site-to-site substitution rate variation dramatically inflates false positive rates of selection analyses: ignore at your own peril. *Molecular Biology and Evolution,* **https://doi.org/10.1093/molbev/msaa037**.

Wright, A. E., & Mank, J. E. (2013). The scope and strength of sex-specific selection in genome evolution. *Journal of Evolutionary Biology,* **26,** 1841-53.

Yamamoto, D., Fujitani, K., Usui, K., Ito, H., & Nakano, Y. (1998). From behavior to development: genes for sexual behavior define the neuronal sexual switch in *Drosophila*. *Mechanisms of Development,* **73,** 135-46.

Yanai, I., Benjamin, H., Shmoish, M., Chalifa-Caspi, V., Shklar, M., Ophir, R., Bar-Even, A., Horn-Saban, S., Safran, M., Domany, E., Lancet, D., & Shmueli, O. (2005). Genome-wide midrange transcription profiles reveal expression level relationships in human tissue specification. *Bioinformatics,* **21,** 650-9.

Yang, L., Zhang, Z., & He, S. (2016). Both Male-Biased and Female-Biased Genes Evolve Faster in Fish Genomes. *Genome Biology and Evolution,* **8,** 3433-3445.

Yang, Z. (2007). PAML 4: phylogenetic analysis by maximum likelihood. *Molecular Biology and Evolution,* **24,** 1586-91.

Yang, Z., & Nielsen, R. (2000). Estimating synonymous and nonsynonymous substitution rates under realistic evolutionary models. *Molecular Biology and Evolution,* **17,** 32-43.

Ylla, G., Nakamura, T., Itoh, T., Kajitani, R., Toyoda, A., Tomonari, S., Bando, T., Ishimaru, Y., Watanabe, T., Fuketa, M., Matsuoka, Y., Barnett, A. A., Noji, S., Mito, T., & Extavour, C. G. (2021). Insights into the genomic evolution of insects from cricket genomes. *Communications Biology,* **in press**.

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Yoritsune, A., & Aonuma, H. (2012). The anatomical pathways for antennal sensory information in the central nervous system of the cricket, *Gryllus bimaculatus*. *Invertebrate Neuroscience,* **12,** 103-17.

Yoshimura, A., Nakata, A., Mito, T., & Noji, S. (2006). The characteristics of karyotype and telomeric satellite DNA sequences in the cricket, Gryllus bimaculatus (Orthoptera, Gryllidae). *Cytogenetic and Genome Research,* **112,** 329-36.

Zhang, Y., Sturgill, D., Parisi, M., Kumar, S., & Oliver, B. (2007). Constraint and turnover in sex-biased gene expression in the genus *Drosophila*. *Nature,* **450,** 233-7.

Zhang, Y. E., Vibranovski, M. D., Krinsky, B. H., & Long, M. (2010). Age-dependent chromosomal distribution of male-biased genes in *Drosophila*. *Genome Research,* **20,** 1526-33.

Zhang, Z., Hambuch, T. M., & Parsch, J. (2004). Molecular evolution of sex-biased genes in *Drosophila*. *Molecular Biology and Evolution,* **21,** 2130-9.

Zhemchuzhnikov, M. K., Kutcherov, D. A., Kymre, J. H., & Knyazev, A. N. (2017). Louder Songs can Enhance Attractiveness of Old Male Crickets (*Gryllus Bimaculatus*). *Journal of Insect Behavior,* **30,** 211-219.

Table 1. The dN/dS values of all female-biased brain genes and male-biased brain genes among the 7,220 genes with *G. bimaculatus* and *G. assimilis* orthologs. Tissue-specific sex bias (TSSB) indicates genes that have sex-biased expression in the brain and are unbiased in all other paired tissues (shown by "*"). Gonad sex bias (GSB) indicates the gene has the same female- or male-biased expression status in the gonad as in the brain and is unbiased in other tissues ("**"). The best matching *D. melanogaster* (Dmel) ortholog is shown with identifiers and gene names from FlyBase (Gramates et al., 2017). Genes are listed by highest to lowest dN/dS values per category.

***G. bimaculatus* ID**	**dN/dS**	**TSSB**	**GSB**	**Matching Dmel ID**	**Dmel gene name**
Female-biased in brain (N=20)					
GBI_10990-RA	0.9739	*		FBgn0028370	*kekkon-3 (kek3)*
GBI_06557-RA	0.8282			FBgn0035082	CG2811
GBI_06507-RA	0.5640			FBgn0035951	CG5068
GBI_00147-RA	0.5270			No match	
GBI_11079-RA	0.5226			FBgn0031265	CG2794
GBI_14015-RA	0.4598			FBgn0038395	CG10407
GBI_14708-RA	0.3835	*		FBgn0003870	*tramtrack (ttk)*
GBI_01688-RA	0.3273	*		FBgn0011604	*Imitation SWI (Iswi)*
GBI_16251-RA	0.2633	*		FBgn0052432	CG32432
GBI_04158-RA	0.2452			FBgn0027582	CG6230
GBI_17348-RA	0.2439			FBgn0011695	*Ejaculatory bulb protein III (EbpIII)*
GBI_05906-RA	0.2258			FBgn0033215	CG1942
GBI_13745-RB	0.1525	*		FBgn0010380	*Adaptor protein (AP-1-2β)*
GBI_09497-RB	0.1433		**	No match	
GBI_00160-RA	0.0692			FBgn0026876	CG11403
GBI_07457-RC	0.0558			FBgn0037659	*Lysine (K)- demethylase 2 (Kdm2)*
GBI_04405-RA	0.0451		**	FBgn0024997	CG2681
GBI_06070-RA	0.0357			FBgn0035724	CG10064
GBI_02686-RA	0		**	FBgn0000317	*crinkled (ck)*
GBI_09453-RB	0	*		FBgn0031550	*Intraflagellar transport 57 (IFT57)*
Male-biased in brain (N=45)					
GBI_19557-RB	3.6750	*		FBgn0030947	CG6696
GBI_01683-RA	0.7988	*		FBgn0039590	CG10011
GBI_10265-RB	0.6262			FBgn0035132	*methuselah-like 10 (mthl10)*
GBI_09477-RB	0.6208			FBgn0004364	*18-wheeler (18w)*
GBI_01684-RA	0.5977	*		FBgn0031473	CG3104
GBI_17358-RA	0.4488		**	FBgn0011695	*Ejaculatory bulb protein III (EbpIII)*

GBI_03471-RA	0.4445			FBgn0019972	*Death rel. ICE-like caspase (Drice)*
GBI_07016-RA	0.4422	*		FBgn0053196	*dumpy (dpy)*
GBI_08544-RB	0.3989	*		No match	
GBI_09470-RA	0.3951		**	FBgn0039478	*Neprilysin 5 (Nep5)*
GBI_01935-RB	0.3929	*		FBgn0012051	*Calpain-A (CalpA)*
GBI_17696-RA	0.3765			No match	
GBI_05452-RB	0.3402			FBgn0036877	CG9452
GBI_07279-RA	0.3265			FBgn0025874	*Meiotic central spindle (Meics)*
GBI_11920-RB	0.3100	*		FBgn0000083	*Annexin B9 (AnxB9)*
GBI_04818-RB	0.2852			FBgn0051217	*modular serine protease (modSP)*
GBI_14462-RA	0.2756	*		No match	
GBI_04545-RA	0.2414			FBgn0012051	*Calpain-A (CalpA)*
GBI_12729-RA	0.2362			FBgn0012037	*Angiotensin converting enzyme (Ance)*
GBI_11067-RA	0.2248			FBgn0033250	CG14762
GBI_15926-RA	0.2248		**	FBgn0030778	CG4678
GBI_04544-RA	0.2013			FBgn0012051	*Calpain-A (CalpA)*
GBI_17460-RA	0.1685			FBgn0038047	CG5245
GBI_01710-RA	0.1497			FBgn0004638	*downstream of receptor kinase (drk)*
GBI_03557-RA	0.1337			FBgn0037802	*Sirtuin 6 (Sirt6)*
GBI_07735-RA	0.1300			FBgn0041713	*yellow-c*
GBI_00231-RA	0.1299	*		FBgn0259736	CG42390
GBI_08685-RA	0.1260	*		FBgn0036454	CG17839
GBI_10295-RA	0.0921			No match	
GBI_15959-RA	0.0902			FBgn0013348	*Troponin C at 41C (TpnC41C)*
GBI_01504-RC	0.0890			FBgn0037665	*Sulfotransferase 2* (*St2*)
GBI_14634-RB	0.0721	*		FBgn0032979	*Chromatin-linked adaptor (Clamp)*
GBI_09694-RB	0.0652		**	FBgn0032768	CG17564
GBI_07712-RA	0.0492	*		FBgn0263025	CG43320
GBI_08082-RA	0.0489			FBgn0030304	*Cytochrome P450 (Cyp4g15)*
GBI_11047-RB	0.0435		**	FBgn0264907	CG44098
GBI_07069-RB	0.0430			FBgn0002524	CG4162
GBI_14322-RA	0.0227			FBgn0243514	*eater*
GBI_00965-RA	0		**	FBgn0034909	CG4797
GBI_02270-RA	0		**	FBgn0260439	*Protein phosphatase 2A (Pp2A-29B)*
GBI_03078-RA	0	*		FBgn0002789	*Muscle protein 20 (Mp20)*
GBI_06961-RA	0	*		FBgn0031800	CG9497
GBI_07963-RA	0	*		FBgn0036316	CG10960
GBI_14909-RA	0	*		FBgn0038385	*F-box and leucine repeat 7 (Fbxl7)*
GBI_15287-RA	0			FBgn0034267	CG4984

Table 2. Top GO functional groups for testis-biased$_{TSSB}$ and ovary-biased$_{TSSB}$ genes identified in *G. bimaculatus* (those with orthologs in *G. assimilis*). Genes were sex-biased only in the gonads and not in the somatic reproductive system, brain or ventral nerve cords (tissue-specific sex-biased, TSSB). The top clusters with the greatest enrichment (abundance) scores are shown per category. *P*-values are derived from a modified Fisher's test, where lower values indicate greater enrichment. Data are from DAVID software (Huang da et al., 2009) using those *G. bimaculatus* genes with predicted *D. melanogaster* orthologs.

Ovary-biased genes (N=1,858)		**Testis-biased genes (N=1,055)**	
GO Function	**P-value**	**GO Function**	**P-value**
Cluster 1: Enrichment Score 10.31		**Cluster 1: Enrichment Score: 5.38**	
nucleotide-binding	1.00E-15	ubiquitin-protein transferase activity	1.20E-07
ATP-binding	2.00E-14	**Cluster 2: Enrichment Score: 3.66**	
Cluster 2: Enrichment Score 7.19		cilium assembly	3.70E-06
WD40/YVTN repeat-like-containing domain	7.70E-09	cilium morphogenesis	6.90E-05
Cluster 3: Enrichment Score 5.41		**Cluster 3: Enrichment Score: 3.28**	
transcription, DNA-templated	5.70E-03	Nucleotide-binding	2.50E-04
		ATP binding	6.40E-04

Table 3. The *D. melanogaster* seminal fluid proteins (SFPs) (Sepil et al., 2019) that were found to have putative orthologs in *G. bimaculatus* (GB) among the subset of 7,220 genes with intra-*Gryllus* orthologs used for dN/dS analysis. Expression levels (FPKM) for each gene are shown for the three male sexual tissues under study.

SFP gene in *D. melanogaster*	**Gene name or ID**	**Gene match in *G. bimaculatus***	**dN/dS in *Gryllus***	**Male sexual tissue expression** (FPKM)		
				Accessory glands	Testis	Male somatic reproductive system
FBgn0034474	*Obp56g*	GBI_14450-RA	2.4819	41.495	0	0.32
FBgn0028986	*Spn38F*	GBI_05353-RD	0.3435	0.565	4.13	43.58
FBgn0028987	*Spn28F*	GBI_00301-RB	0.2866	36.84	270.87	94.46
FBgn0030362	*regucalcin*	GBI_08029-RA	0.2496	37.63	15.08	23.19
FBgn0030932	*Ggt-1*	GBI_03406-RA	0.2302	9.845	8.60	21.73
FBgn0038198	*Npc2b*	GBI_06029-RA	0.2197	7.5	0.50	793.96
FBgn0283509	*Phm*	GBI_06121-RA	0.1496	71.82	32.28	86.185

Table 4. The proportion of genes with sex-biased$_{TSSB}$ gonadal and universally unbiased expression in *G. bimaculatus* that had dN/dS>1, and their expression breadth across tissues (average number of nine tissues with expression >5 FPKM).

Gene category	**N dN/dS >1**	**N Genes**	**Percent of genes**	**Chi^2 P[a]**	**Ave. exp. breadth**	**SE**	**MWU- test P (expression breadth)[a]**
Ovary-biased$_{TSSB}$	19	1,858	1.02	a	6.74	0.74	a
Testis-biased$_{TSSB}$	22	1,055	2.09	b	2.73	0.72	b
Universally unbiased	66	3,449	1.91	b	5.62	0.76	a

[a] Different letters in the columns with P values indicate a statistically significant difference between categories with P<0.05. SE=standard error.

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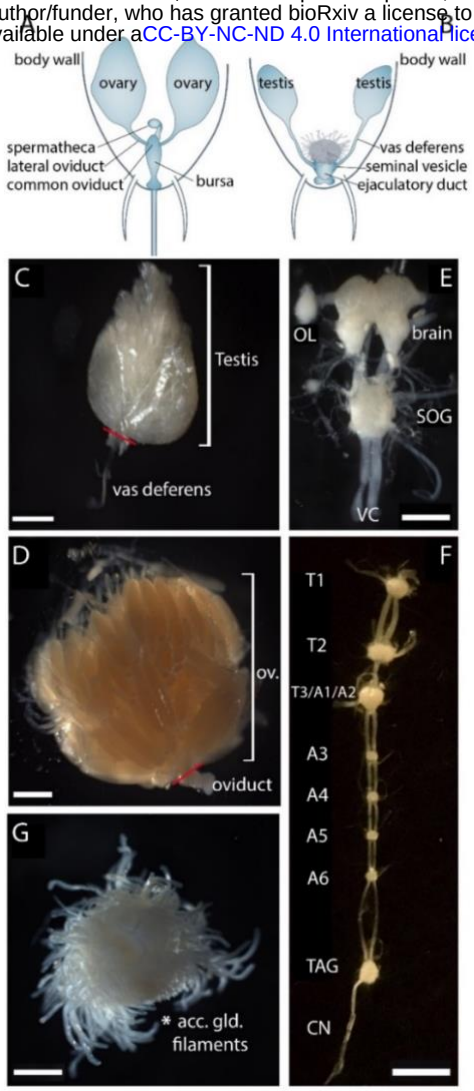

Fig. 1. *Gryllus bimaculatus* reproductive and nervous system tissues studied herein. A) Schematic diagram of the female reproductive system showing the gonads and the somatic tissues included in the somatic reproductive system under study. B) Schematic diagram for the male gonads and male somatic reproductive system. C-G provide micrographs of various tissue types studied herein. C) the testis (one testis shown here; both testes from a given male were used for sampling), including a part of its attached vas deferens (boundary indicated by red line; the vasa deferentia were included in the male somatic reproductive system libraries, and not in testis libraries). D) the ovary (ov; one ovary shown here; both ovaries from a given female were used for sampling) and an immediately attached segment of oviduct (boundary indicated by red line; the oviducts were included in the female somatic reproductive system libraries, and not in ovary libraries). E) the brain, including an optic lobe (OL) (one OL shown here; both OLs from a given individual were included in brain samples). For context, the attached suboesophageal ganglion (SOG) and upper portion of the ventral nerve cord (VC) are also shown; these structures were included in the ventral nerve cord libraries and not in brain libraries. F) the ventral nerve cord including the three thoracic ganglia (T1: prothoracic, T2: mesothoracic, T3/A1/A2: metathoracic ganglion complex), and five abdominal ganglia (A3-A6 and the terminal abdominal ganglion TAG) (Huber, 1963; Jacob & Hedwig, 2016). The cercal nerve (CN) of one side is also shown. For the ventral nerve cord samples, all tissues in F and the SOG were pooled. G) The male accessory gland consisting of numerous accessory gland filaments (asterisk; also shown schematically as filamentous structure in B). Scale bars: 500μm in C and E, 1000μm in D and G, 2500μm in F.

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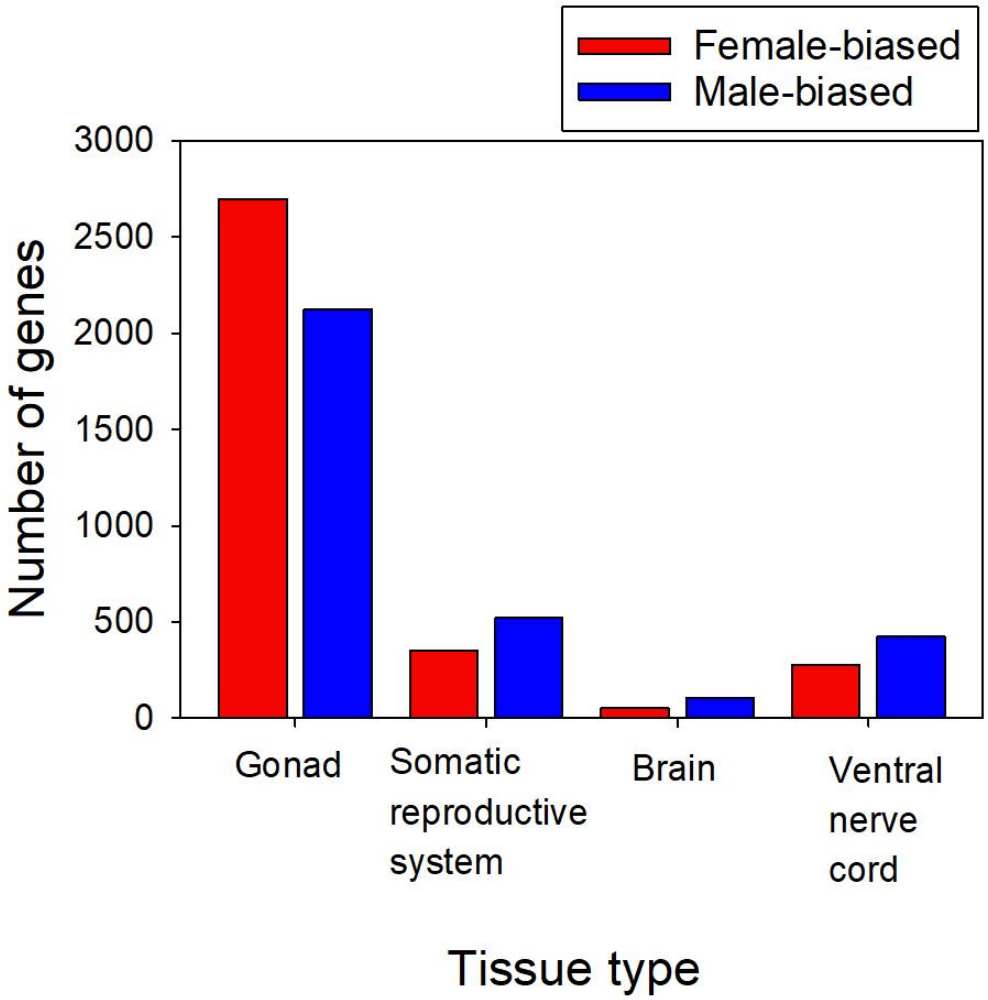

Fig. 2. The number of male-biased and female-biased genes identified in the gonad, somatic reproductive system, brain, and ventral nerve cord across all 15,539 *G. bimaculatus* genes under study (sex-biased indicates a two-fold difference in expression and P<0.05). All remaining genes not shown per tissue type had unbiased status as follows: gonad (N=10,717), somatic reproductive system (N=14,666), brain (N=15,382) and ventral nerve cord (N=14,835).

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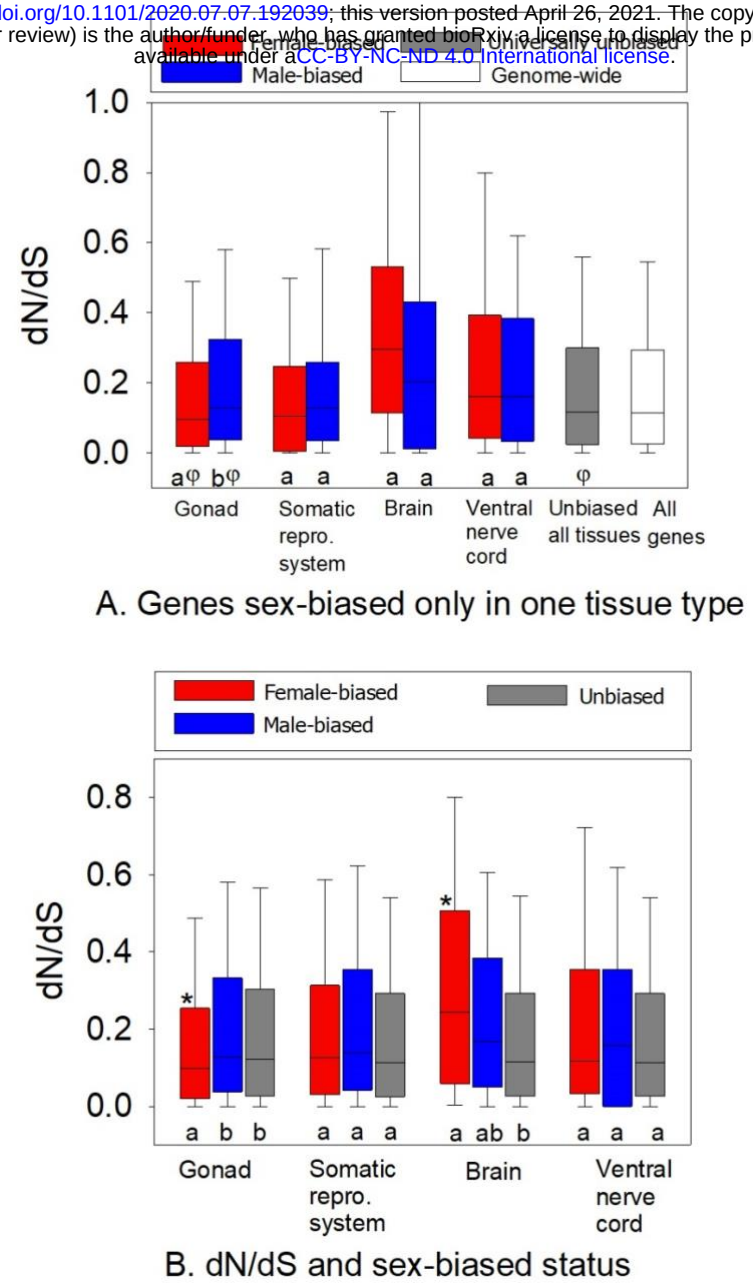

Fig. 3. Box and whisker plots of the dN/dS values of genes with female- or male-biased expression in *G. bimaculatus*, and attained using the genes with high confidence orthologs its sister species *G. assimilus*. A) Genes with female- or male-biased gene expression in only one tissue type and unbiased in three remaining paired tissues, that is, with tissue-specific sex bias (TSSB). In addition, genes with universally unbiased expression in all four paired tissue types and the genome-wide dN/dS are shown. B) dN/dS of all (ALL) genes with sex-biased expression in each of four tissue types regardless of status in other tissues. In panel A, different letters (a, b) under the two bars within each tissue type indicate a statistically significant difference (MWU-test P<0.05), and φ indicates the difference in dN/dS in with respect to universally unbiased genes (MWU-tests P<0.05). For panel B, different letters among the three bars within each tissue type indicates MWU-test P<0.05 (note that "ab" for the brain indicates no difference of male-biased to female-biased or unbiased genes) and *indicates a difference in dN/dS between female-brain biased and ovary-biased genes. N values of genes per category are provided in Table S3. repro. = reproductive. Outliers above the 95th percentile, including dN/dS >1, were excluded from the figure to allow visualizations on the Y-axis.

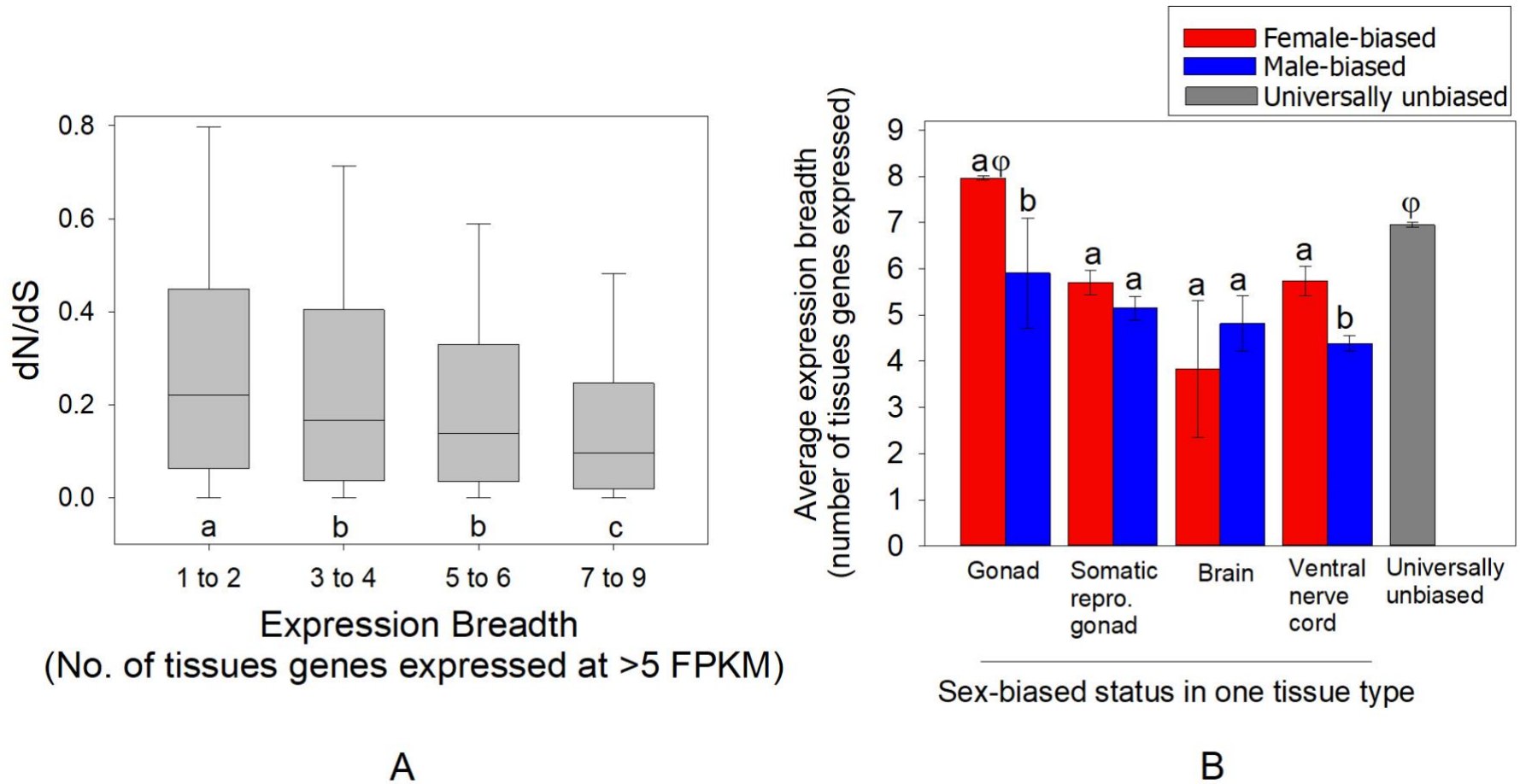

Fig. 4. A) Box and whisker plots of the dN/dS values of all studied genes with respect to their expression breadth, or pleiotropy, in *G. bimaculatus* (N=7,220 genes). B) The average expression breadth (number of tissues with expression of a gene ≥5FPKM) of genes with sex-biased expression in only one tissue type, that is, with female- or male-biased$_{TSSB}$ expression. In A, different letters below bars indicate a statistically significant difference using ranked ANOVA with Dunn's paired contrast (P<0.05). In B, different letters in each pair of bars indicate a difference using MWU-tests. φ above ovary-biased and universally unbiased genes indicates a statistically significant difference from each other and from all other bars. Error bars in B indicate standard errors. repro. = reproductive. For panel A, outliers above the 95th percentile, including dN/dS>1 were excluded for visualization on the Y-axis.