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Geography is more important than life history in the recent

diversification of the tiger salamander complex

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ABSTRACT

The North American tiger salamander species complex, including its best-known species,

the Mexican axolotl, has long been a source of biological fascination. The complex exhibits a

wide range of variation in developmental life history strategies, including populations and

individuals that undergo metamorphosis, those able to forego metamorphosis and retain a larval,

aquatic lifestyle (i.e., paedomorphosis), and those that do both. This life history variation has

been assumed to lead to reproductive isolation and speciation, but the degree to which it has

shaped population- and species-level divergence is poorly understood. Using a large multi-locus

dataset from hundreds of samples across North America, we identified genetic clusters across the

geographic range of the tiger salamander complex. These clusters often contain a mixture of

paedomorphic and metamorphic taxa, indicating that geographic isolation has played a larger

role in lineage divergence than paedomorphosis in this system. This conclusion is bolstered by

geography-informed analyses indicating no effect of life history strategy on population genetic

differentiation and by model-based analyses demonstrating gene flow between adjacent

metamorphic and paedomorphic populations. This fine-scale genetic perspective on life-history

variation establishes a framework for understanding how plasticity, local adaptation, and gene

flow contribute to lineage divergence. Many members of the tiger salamander complex are

endangered, and the Mexican axolotl is an important model system in regenerative and

biomedical research. Our results chart a course for more informed use of these taxa in

experimental, ecological, and conservation research.

Keywords: *Ambystoma*, life history, phylogenetics, population genomics, salamanders

SIGNIFICANCE STATEMENT

Population structure and speciation are shaped by a combination of biotic and abiotic factors. The tiger salamander complex has been considered to be a key group where life history variation has led to a rapid rate of speciation, driven in large part by the evolution of obligate paedomorphosis–a condition where adults maintain an aquatic, larval phenotype. Using a large multi-locus dataset, we present evidence of gene flow between taxa with different life history strategies, suggesting that obligate paedomorphosis is not a strong driver of speciation in the tiger salamander complex. Many of these nominal taxa are listed as critically endangered, and our genetic results provide information and guidance that will be useful for their conservation.

Introduction

Life history—the complement of traits affecting survival and reproduction over an

organism's lifetime—affects ecology and dispersal, and therefore plays a potentially important

role in shaping population structure and speciation (1, 2). The Mexican axolotl (*Ambystoma*

mexicanum; hereafter, "axolotl") and related salamander species of the North American

Ambystoma tigrinum complex (Table 1) display enormous life history variation, particularly with

respect to the completion of metamorphosis (3). Some species are considered obligate

paedomorphs, where sexually mature adults retain a larval, aquatic body plan that includes

external gills and an enlarged tail fin (4). Others are considered obligate metamorphs,

transforming from aquatic larvae to terrestrial juveniles, eventually returning to the water to

breed as adults. Most populations are facultative, transforming under certain genetic and/or

environmental conditions (5–8). This lability in life history is believed to have played an

important role in the diversification of the *A. tigrinum* complex, particularly in the Trans-

Mexican Volcanic Belt (TMVB) region, where several paedomorphic species, including the

axolotl, are currently recognized (9–11). Obligate paedomorphosis is estimated to have evolved

in this region multiple times (11–13) in association with relatively large, permanent bodies of

water. Presumably, restricted gene flow between these isolated, paedomorphic populations led to

speciation as well as morphological adaptations to the aquatic lifestyle (14).

The term "species complex," while not a formal taxonomic category, is often used to

describe groups of closely related lineages early in the divergence process. The tiger salamander

species complex has been highlighted as a potentially valuable example of such a recent

radiation (15) that could provide insight into the early mechanisms initiating and/or maintaining

diversity (16–21). However, species complexes pose a number of challenges: phenotypic

differences among lineages may be subtle or absent (i.e., cryptic species), ancestral

polymorphisms may lead to incomplete lineage sorting that confounds molecular systematic

studies, and reproductive barriers may be incomplete. The reliance on phenotype for species

diagnosis in the *A. tigrinum* complex may require particular scrutiny, as several phenotypic traits,

including developmental state and adult color pattern, can be plastic and highly variable (22–24).

Given these challenges, fundamental questions remain concerning the reality of component

lineages within the tiger salamander complex, calling into question the current taxonomy as an

accurate reflection of its underlying evolutionary history.

Previous research has produced mixed results regarding levels of genetic differentiation

among lineages in the *A. tigrinum* complex, as well as the relative importance of paedomorphosis

as a driver of diversification. While population- and species-level structure is evident, an

important theme emerging across multiple studies has been that reproductive barriers are porous

(11, 25, 26). Shaffer and McKnight (11) noted "the striking lack of differentiation among the 14

species of the tiger salamander complex" (p. 425). Some studies have found an elevated degree

of population structure among paedomorphic populations relative to metamorphic populations

(12, 27). However, others have shown a lack of effect of paedomorphosis on population genetic

differentiation (28), and demonstrated that paedomorphic taxa and neighboring metamorphic

populations interbreed (29). Furthermore, crosses between metamorphic and paedomorphic taxa

can produce viable hybrid offspring under laboratory conditions (30, 31).

To understand the processes underlying diversification in the tiger salamander complex,

an important first step must be a range-wide assessment of population structure and the

clarification of population- and species-level boundaries. With this groundwork, insights into the

role of paedomorphosis and diversification can be addressed. For a study system of this

geographic and taxonomic scale, robust inference of population structure requires information

from multiple genes combined with thorough range-wide sampling. To meet these criteria, we

expand on a large multi-locus dataset containing 95 nuclear loci for 93 individuals (32), to

produce a data matrix for 347 individuals across the full geographic range of tiger salamanders

(Fig. S1). Given the complex and somewhat checkered history of the group's taxonomy, we

focus on a naïve approach, performing population structure analyses without *a priori*

identification of taxa to resolve geographic genetic clusters and characterize patterns of

admixture. We also overlay these results on the existing taxonomy to explore the correspondence

between current taxonomy and genetic differentiation. We then use these data for a phylogenetic

analysis to provide an updated working hypothesis for the evolution of the group. Finally, we test

the hypothesis that life history evolution has driven speciation in the tiger salamander complex,

particularly through the assumed isolation of paedomorphic species. In light of the life history

variation in this species complex, we use natural history information to place taxa into one of

five categories (Table 1) reflecting their propensity to metamorphose, then use model-based tests

of migration to ask whether paedomorphic populations (categories 4 and 5) exhibit greater levels

of genetic differentiation relative to those that regularly metamorphose (categories 1-3).

RESULTS AND DISCUSSION

Identification of genetic lineages and phylogenetic relationships

A principal components analysis (PCA) and a discriminant analysis of principal

components (DAPC; 33) both recovered *Ambystoma californiense* Gray 1853 as genetically

distinct from all other tiger salamanders (Fig. 1A, S2, S3), consistent with previous work (e.g.,

11, 34). Given that *A. californiense* is the only obligate metamorph in the complex (life history

category 1; Table 1) and is genetically divergent and geographically isolated from all other tiger

salamanders evaluated in this study, we focus the remaining analyses on the other members of

the complex. After removing *A. californiense* from the dataset, DAPC supported the recognition

of three primary genetic clusters: U.S. (including samples from southern Canada), northwestern

Mexico (primarily the Sierra Madre Occidental), and the central Mexican highlands (Fig. 1B,

S4). A PCA of these data produced similar clustering results, with ordination patterns largely

mirroring geographic sampling (Fig. S5). For a further discussion of the taxonomic implications

of our results, see the Supplementary Material.

Within central Mexico, we identified $K = 7$ as the best-fit number of clusters in both

DAPC and STRUCTURE analyses (Figs. 2A, S6). However, several of these clusters were only

represented as a small proportion of genetic assignments, rather than as meaningful groups of

individuals, and visual inspection at multiple levels of K revealed four clear geographic clusters

(hereafter referred to as CM1-CM4) that captured the primary patterns of genetic differentiation

and admixture across this region (Figs. 2A, S7-S8). Network analysis (35) also recovered groups

that generally correspond to CM1-CM4, but with evidence of many reticulations (Fig. 3). This

starburst-like pattern indicates the presence of conflicting topologies within the dataset (35),

which can be caused by biological processes including recent diversification with incomplete

lineage sorting, current or evolutionarily recent gene flow, or both. Phylogenetic analyses (36–

38) recovered monophyletic CM1, CM2, and CM4 clusters (see below for discussion of CM3),

but often with low statistical support (Fig. 3).

Based on type locality and range information, the CM1 cluster includes three stream-

breeding and facultatively paedomorphic taxa: *Ambystoma altamirani* Dugès 1895 (39)*, A.

leorae* (Taylor 1943) (40)*, and *A. rivulare* (Taylor 1940) (9). Subsequent analyses of this cluster

identified two admixed populations associated with eastern and western portions of its

geographic distribution (Figs. 2B, S9). Phylogenetically, CM1 was monophyletic and sister to all

other central Mexican groups (Fig. 3).

CM2 grouped *A. lermaense* (Taylor 1940) (9) with samples corresponding to the type

localities and range of *A. bombypellum* Taylor 1940 (9) and *A. granulosum* Taylor 1944 (41).

There was no evidence for genetic isolation among any of these three taxa (Figs. 2C, S9).

Within CM3, population-genetic analyses first separated samples of *A. ordinarium* Taylor

1940 (5) from the rest (with no admixture detected; Figs. 2D, S9). Other samples clustered into

northern and southern groups with some admixture in intermediate localities. The more southern

cluster includes two obligately paedomorphic (category 5) taxa, *A. andersoni* Krebs & Brandon

1984 (42) and *A. dumerilii* (Dugès 1870) (43), endemic to Lakes Zacapú and Pátzcuaro,

respectively. The samples from other CM3 localities likely represent what are currently

classified as *A. amblycephalum* Taylor 1940 (9) and *A. flavipiperatum* Dixon 1963 (44).

CM4 included the axolotl, *A. mexicanum* (Shaw & Nodder 1798) (45), and *A. taylori*

Brandon et al. 1981 (10)—both category 4 paedomorphs—as well as samples assigned to the

facultatively paedomorphic (category 3) *A. velasci* (Duges 1888) (46). Subsequent DAPC and

STRUCTURE analyses of CM4 placed all but one *A. mexicanum* individual sampled from Lakes

Xochimilco and Chapultepec (i.e., the remaining habitats of the axolotl) in a distinct cluster (Fig.

2E, S9), with evidence of some admixture in samples to the northwest of Mexico City.

Within the U.S. and Canada (excluding *A. californiense)*, DAPC and STRUCTURE

identified three genetic clusters ($K = 3$), with clear signs of admixed contact zones (Figs. 2F,

S10, S11). Geographically, these clusters are associated with the eastern U.S., the central +

western U.S., and the southwestern U.S. + Rocky Mountains (hereafter, we refer to the latter two

groups as central U.S. and Rocky Mountain U.S., respectively). Populations in the eastern U.S.

are assignable to *A. tigrinum* Green 1825, while central U.S. and Rocky Mountain U.S.

populations to *A. mavortium* Baird 1850. Phylogenetic analyses corroborated these results, albeit

with extensive reticulations among clusters in the phylogenetic network (Fig. 3) and mixed

support for species-tree relationships among the three major groups (Fig. 3 vs. Fig. S13). Further

exploration of population structure within each of the three U.S. clusters recovered additional

patterns of differentiation (Figs. S10, S11), including northern and southern clusters in the

eastern U.S. and central U.S., and an allopatric cluster restricted to the Pacific Northwest.

Overall, our U.S. results are similar to mtDNA-based results from previous studies, which

identified haplotype clades associated with the eastern U.S., Great Plains and Rocky Mountains,

and the Pacific Northwest (11, 47).

All analyses of the northern Mexico group identified two genetic clusters ($K = 2$; Figs.

2G, S12). The northern cluster corresponds to *A. rosaceum* Taylor 1941 (48), while the southern

cluster is likely *A. silvense* Webb 2004 (49, see also 50). Phylogenetic analyses recovered each

of these two northern Mexico clusters as monophyletic, but not sister to one another, and with

few reticulate nodes in the phylogenetic network (Fig. 3, S13).

Does paedomorphosis lead to increased genetic differentiation?

As a first step in addressing this question, we used Bayesian modeling to test for genetic

isolation of paedomorphic taxa (i.e., species scored as life history category 4 or 5 in Table 1). We

tested for gene flow between paedomorphs–*A. andersoni* and *A. dumerilii* in CM3, and *A.*

mexicanum and *A. taylori* in CM4–and their surrounding facultatively paedomorphic

populations. For both CM3 and CM4, the top-ranking models included migration among all

populations, regardless of their degree of paedomorphosis (Fig. 4). Alternative models of no

migration or restricted migration (Fig. S14) were rejected with decisive Bayes factors (Table S3).

In CM3, migration rates entering the *A. dumerilii* population were higher than those leaving that

population, while migration rates entering and exiting *A. andersoni* were approximately equal.

In CM4, migration rates exiting the *A. mexicanum* population were much higher than those

entering the population while migration rates entering and exiting *A. taylori* were approximately

equal (Fig. 4, Table S4). An important caveat to our migration and population structure analyses

is that divergence time is not included as a model parameter, which prevents distinguishing

between historical and contemporary gene flow. Thus, our ability to differentiate ongoing from

historical gene flow in these tests is limited.

We also assessed whether a paedomorphic life history is associated with greater

population genetic differentiation (calculated here as Φ_{ST}), which would be expected if

paedomorphosis is an important reproductive barrier (12). Plots of Φ_{ST} against geographic

distance showed that comparisons involving paedomorphic populations were largely overlapping

with those based on facultative-facultative comparisons, with no difference in slope or y-

intercept (Fig. 4). In both CM3 and CM4, partial Mantel tests found no significant correlation

between life history and pairwise Φ_{ST} after correcting for geographic distance (Fig. 4).

The influence of life history on diversification

A popular perspective on diversification in this species complex is that paedomorphosis

promotes reproductive isolation and, ultimately, speciation (10, 11). An important theme of our

results is that taxa with different life history strategies commonly cluster together by geography

in population genetic and phylogeographic analyses (Figs. 2, 3). Furthermore, genetic

differentiation is not significantly greater for paedomorphic populations, relative to facultative

populations, and there has been substantial gene flow among populations with different life

history strategies (Fig. 4). Thus, the evolution of life history extremes does not appear to be the

main driver of speciation in the tiger salamander complex.

It is also worth noting that facultative paedomorphosis is the inferred ancestral condition

(8) and nearly all extant lineages in the *Ambystoma tigrinum* complex are capable of

metamorphosis, either in the wild or in the lab. Only *A. dumerilii* is not known to survive

metamorphosis, even under laboratory conditions (51). That only one described taxon in the

complex is demonstrably fixed for paedomorphosis raises several questions: Are the rest of the

primarily paedomorphic populations *en route* to fixation, rather than actually fixed? Have

lineages fixed for paedomorphosis gone extinct? Do facultative populations fluctuate in

paedomorph frequency over time, depending on environmental and/or demographic conditions

(5)? Do transforming and non-transforming individuals interbreed in nature? While these remain

open questions, below we provide some relevant context for exploration and future research.

Theory predicts that paedomorphosis in salamanders is most likely to become fixed in a

population when ecological conditions are favorable in the larval, aquatic environment but

unfavorable on land (e.g., when terrestrial habitats are arid or resource-limited) (12, 52, 53).

However, it is not known how long fixed populations persist over evolutionary time; obligate

paedomorphic lineages may have arisen and gone extinct multiple times over tiger salamander

evolutionary history (54). One situation that would result in complete speciation is if

paedomorphosis became fixed in the population and migration with surrounding facultatively

paedomorphic populations ceased due to strong assortative mating and/or selection against

hybrids (55–57). However, assortative mating within life history morphs has not been

demonstrated in other salamanders (58–60) and different tiger salamander morphs can interbreed

(61). In fact, the only known instance of positive assortative mating in *A. tigrinum* is associated

with tail length (62, 63), which might favor metamorphosed males as paedomorphic individuals

can have relatively short tails (42). The evidence for selection against intermorph offspring is

also scarce. Captive breeding using artificial fertilization has produced *A. mexicanum* x *A.*

tigrinum, *A. dumerilii* x *A. tigrinum*, *A. andersoni* x *A. tigrinum*, and *A. dumerilii* x *A. rivulare*

hybrids (7, 31, 64–66). This suggests that postzygotic barriers are weak, although the extent to

which divergent lineages would interbreed in the wild remains a key question in need of

investigation.

If biological barriers to reproduction are limited or absent, there are several potential

mechanisms by which dispersal could mediate gene flow between paedomorphic and

metamorphic populations. First, metamorphosed individuals from facultatively paedomorphic

populations could move into bodies of water containing paedomorphic populations. Our

demographic analyses support this scenario, as they recovered immigration from facultative into

paedomorphic populations (Fig. 4). However, demographic analyses also estimated emigration

out of paedomorphic populations; thus, "obligate" paedomorphs might also occasionally

metamorphose and disperse. Occasional metamorphosis has been documented in several

putatively paedomorphic populations (9, 10, 42, 67–69), and may be more prevalent than

currently recognized because rare transformed individuals in nature are virtually impossible to

verify (70). Importantly, even a low frequency of transformed adults is likely sufficient to

maintain evolutionary cohesion with surrounding populations (10, 29, 71). Alternatively, the

dispersal of paedomorphs might be human-mediated; this could partially explain the elevated

emigration rates we observed in the axolotl (Fig. 4). Axolotls are known to be sold in local

markets as food, bait, or aquarium pets, and are sometimes moved from one area of central

Mexico to another (72). Finally, gene flow may not necessarily be mediated across a terrestrial

environment; genetic connectivity could also be maintained if populations have been forced into

contact by water level fluctuations in the groundwater system of a region. The dynamic

lacustrine history of the Cuenca Oriental (73), for instance, suggests that isolation of some

paedomorphic populations has been punctuated by broader aquatic connections over short

geological time scales, potentially facilitating gene flow. Collectively, these mechanisms may

explain the maintenance of geographic genetic groups, now or in the evolutionarily recent past,

containing a range of life histories.

The genetic cohesion we detected across wide swaths of the TMVB is particularly

striking given other evidence for local adaptation in this region. This scenario is perhaps best

exemplified in the Cuenca Oriental (Fig. 2E), where some lakes contain paedomorphic

salamanders adapted to saline conditions, a very rare trait among amphibians (74). At one site,

the saline Lake Alchichica, the population is currently considered a distinct species (*A. taylori*)

(10). However, our results indicate that *A. taylori* is not genetically distinct from surrounding,

putatively non-saline-adapted populations, a result consistent with a previous microsatellite-

based study that found significant gene flow to and from *A. taylori* (29). While we cannot rule

out the potential for localized selection and adaptation in the *A. taylori* genome, our results

highlight that even populations adapted to aquatic conditions intolerable to most other

Ambystoma have not reached a level of isolation identifying them as an independent evolutionary

lineage (75).

Comparisons to previous work

This study included extensive geographic sampling of the tiger salamander complex,

which provided a broad spatial context to more fully understand patterns of genetic variation.

However, it is important to consider the possibility that our data are simply not sensitive enough

to detect genetic differentiation associated with lineage divergence on an extremely recent time

scale. The common ancestor of *A. californiense* and the remainder of the species complex dates

to approximately five million years (11, 34), and phylogenomic data indicate that speciation

across the remaining tiger salamander lineages occurred within the last one million years (76).

Such recent timing renders lineage boundaries difficult to detect (15). Our markers, developed

from transcriptomic resources (32), may not have an overall substitution rate sufficient to detect

such subtle genetic differentiation. We recommend that future research uses a larger genomic

dataset or faster-evolving loci with particular focus on the sampling of paedomorphs in this

system. To this point, perhaps the strongest evidence for the genetic divergence of paedomorphic

populations comes from past microsatellite-based work indicating the genetic distinctiveness of

A. andersoni and *A. mexicanum* from a select set of populations across central Mexico (27) and

the genetic distinctiveness of *A. taylori* from neighboring populations in the Cuenca Oriental

(29). Both of these studies recovered signatures of gene flow, but identified greater overall

population structure compared to this study, which could be due to numerous factors, including a

faster microsatellite mutation rate and more limited locality sampling.

CONCLUSIONS

The extent to which populations within the tiger salamander complex exhibit phenotypic

plasticity in life history traits is remarkable and is believed to have played a role in the rapid

accumulation of lineages observed in the highlands of central Mexico (11, 57). While our results

suggest there is less species-level diversity in central Mexico than previously recognized, there is

clearly more diversity in this region compared to the US and Canada where there is (a) more

geographic space and (b) less life history variation within and between lineages. While we

cannot fully explain the greater diversity in central Mexico, our results suggest that major

patterns of diversification are related to a complex history of geographic isolation and secondary

contact, in which life history strategy has played a less important role, at least in the long

(evolutionary) run. We agree with previous work (12) that the complex geological history of the

TMVB, including montane uplift and fluctuating drainage connectivity since the Miocene, has

been the cornerstone of the evolutionary history of this species complex, and that the influences

of geographic isolation and paedomorphosis may work synergistically to lead to the

establishment of isolated populations. Lingering questions notwithstanding, this large-scale

genetic and geographic study establishes a framework for understanding the evolutionary history

of the *Ambystoma tigrinum* species complex. The results presented here will facilitate

comparative studies of the axolotl and its allies, provide direction for conservation prioritization

and management, and strengthen the use of the tiger salamander species complex as a model

system in biology.

MATERIALS & METHODS

Geographic sampling

We generated new data from 254 individuals sampled from across the range of the *A.*

tigrinum complex (Fig. S1; Table S1). These individuals were combined with 93 individuals

sampled in O'Neill et al. (32) to produce a data set comprising 347 individuals. We sampled a

large number of localities (188) with 1-9 individuals sampled per locality (mean = 1.8). This

sampling included 166 individuals from the US, 2 from Canada, and 178 from Mexico.

Ambystoma californiense samples were primarily included as a close outgroup to the remainder

of the species complex and were sampled from localities with limited to no impacts of

introgression from invasive *A. mavortium* (47). Additional outgroup data were generated for two

species outside of the *A. tigrinum* complex: *A. opacum* and *A. texanum*. Full details regarding the

generation of these data can be found in the Supplementary Materials. All *A. tigrinum* complex

species were assigned to a life history category based on review of the published literature and

private field records from authors of this study. These life history categories are as follow: 1 =

obligate metamorph (no paedomorphs have been documented in the field); 2 = strong bias

towards metamorphosis with rare paedomorphs found in the field; 3 = both metamorphosis and

paedomorphosis common in the field; 4 = strong bias towards paedomorphosis with rare

metamorphs found in the field; 5 = obligate paedomorph (no metamorphs have been documented

in the field). In the text, "paedomorph" describes taxa scored as 4 or 5, while "facultative

paedomorph" describes taxa scored as 2 or 3.

Data collection and sequencing

We generated DNA sequence data from a panel of 95 nuclear loci developed specifically

for the tiger salamander species complex. A more complete description of marker development

can be found in O'Neill et al. (32). Genomic DNA was extracted using a DNeasy Blood and

Tissue kit (Qiagen). For a small number of DNA extractions, we increased DNA quantities using

a Repli-g whole genome amplification kit (Qiagen). We used an initial round of PCR in 96-well

plate format to amplify all loci from an individual, followed by a smaller second round of PCR to

amplify loci that did not amplify in the initial PCR. See O'Neill et al. (32) for the details of PCR

conditions and primer sequences.

PCR products from all loci were pooled for each individual in roughly equal

concentrations based on the intensity of amplification as visualized on an agarose gel. Indexed

Illumina sequencing libraries were generated for each individual using an Illumina Nextera XT

DNA Library Preparation Kit (Illumina). Subsequent to library preparation, indexed libraries

were quantified using a Qubit Fluorometer, pooled in equimolar concentrations, and checked on

an Agilent Bioanalyzer to assure proper fragmentation.

Sequencing was performed in four rounds using an Illumina MiSeq. We performed an

initial round using a total of seven individuals to test the compatibility of the Illumina Nextera

XT library kit with our PCR amplicons. Three subsequent rounds of library preparation and

sequencing were performed on sets of 96 individuals each, and some individuals were sequenced

in multiple rounds due to initially low read counts. All sequencing was performed with paired-

end 150 bp reads. Overall, we generated a total of 29,426,894 PE reads across all newly

sequenced individuals, with an average of 309,757 PE reads, 1,148X coverage, and 7.6%

missing loci per individual (Table S2).

Bioinformatics and dataset generation

All sequence reads were processed using a newly developed bioinformatic pipeline

written for this project (https://doi.org/10.5281/zenodo.3585970) that produces multiple

sequence alignments for individual loci and genome-wide SNP matrices sampled from variable

sites. This pipeline was developed using the Snakemake workflow management system (77),

linking together multiple software tools to take sequence data from raw reads to phased sequence

alignments for each locus. Demultiplexed paired-end Illumina fastq files were used as input, with

separate forward and reverse read files for each individual. Sequence data were trimmed and

filtered in Trimmomatic (78) using a sliding window of 4 base-pairs and a minimum average

quality score of 15. Filtered sequence reads were then aligned to reference sequences from the

O'Neill et al. (2013) dataset (32); specifically, we used the clean sequences of an *A. ordinarium*

sample that had high coverage and low amounts of missing data. The resulting aligned contigs

were processed using SAMtools (79) to filter and prepare data for FreeBayes (80), which was

used to call variable sites. Variants were filtered with VCFTools (81) by removing indels and

setting a quality threshold of phred score > 20 and a minimum read depth of 30. The program

WhatsHap (82) was used to perform read-based phasing of the data for each locus x individual

contig. Finally, phased haplotypes from each individual (two copies, regardless of homo- or

heterozygosity) were combined into an alignment of all individuals using MAFFT with the

default auto parameter (83). We generated fasta files of SNPs using the SNP-sites program (84)

and created a SNP genotype matrix by sampling variable sites from a concatenated sequence

alignment of all loci. We identified three loci (E12G1, E6A11, and E7G8) as potential paralogs

based on high alignment error and high levels of heterozygosity for all individuals. These three

loci were excluded from all analyses.

Across the remaining 92 loci, alignment lengths ranged from 123 to 630 bp (avg. = 270

bp) with a total concatenated alignment of 24,788 bp. For the full dataset, including *A. texanum*

and *A. opacum* outgroups, single-locus alignments contained an average of 55 variable sites

(min. = 15, max. = 121) and an average of 41 parsimony informative sites (min. = 11, max. =

102). Population genetic analyses restricted to the *A. tigrinum* complex including *A.*

californiense—which were further filtered for non-biallelic SNPs and a minor allele count ≥ 3—

contained a total of 2,360 SNPs.

Population structure and lineage discovery

We developed hypotheses of population-level lineages across the range of the *A. tigrinum*

complex ignoring the existing taxonomy, starting with the identification of major geographic

patterns of differentiation, and then performing a recursive set of analyses on more

geographically restricted sets of individuals. In our initial round of analyses we used two non-

parametric methods: principal components analysis (PCA) and discriminant analysis of principal

components (DAPC) (33). While both analyses provide a multivariate summary of genetic data,

DAPC is also used to assess the fit of data to varying numbers of population clusters. These

analyses were applied to our full genotypic dataset including *A. californiense* and all remaining

individuals from the *A. tigrinum* complex. The PCA was calculated using the function 'prcomp'

in the R package stats (85), while the DAPC was calculated using the package adegenet (86).

The optimal number of principal components to retain for DAPC was identified using cross-

validation via the xvalDapc function with default parameter values. DAPC was performed

without prior assignment of individuals to groups across a range of cluster levels ($K = 1\text{-}20$). We

used two metrics to identify the best estimate of the primary splits in our data. First, we used the

Bayesian Information Criterion (BIC) calculated in the DAPC analysis to assess the fit of the

data to different levels of K. We note that the level of K with the absolute lowest BIC may not be

a better explanation of the data than a K with a slightly higher BIC (87); therefore, we applied

this measure for general guidance on a range of K that may describe the data well. We paired this

assessment with visualizations of the first and second principal components (Fig. S2, S5) and

DAPC ordination plots to identify the level of K at which similar clustering patterns could be

observed with minimal change at successively higher levels of K. DAPC of the complete tiger

salamander species complex identified a consistent pattern beginning at $K = 5$ for high

differentiation of all *A. californiense* samples (Fig. S3). Further DAPC analysis with *A.*

californiense removed identified a consistent pattern beginning at $K = 5$ for differentiation

between clusters of populations from northern and central Mexico, and three clusters of U.S.

populations (two from the Western U.S. and one from the Eastern U.S.; Fig. S4).

436   Using the clusters identified in the DAPC analysis of the total data set, we then used both

DAPC and STRUCTURE v.2.3.4 (88) to analyze subsequent data sets comprising smaller

numbers of individuals. Recursive rounds of DAPC analyses were stopped when BIC scores

showed little improvement (ΔBIC < 2) at values of $K > 1$. STRUCTURE analyses used an

admixture model and 500,000 generations following a burn-in of 100,000 generations. Analyses

were performed for $K = 1$-10 with 16 replicate analyses for each K. To help identify an optimal

value for K, we calculated ΔK using the Evanno method (89) via the CLUMPAK web tool (90).

A limitation of the Evanno method is that it cannot estimate the likelihood of $K = 1$ (89), thus,

we also visually inspected individual group assignments and concluded a value of $K = 1$ if the

corresponding DAPC cluster showed little improvement (ΔBIC < 2) at values of $K > 1$, or when

individuals were simply being split without any geographic or individual clustering. We also

evaluated models where K was equal to the number of currently recognized species in that

genetic cluster (e.g., $K = 3$ for CM1, which included the type localities for three described

species: *A. altamirani*, *A. leorae*, and *A. rivulare*). We mapped individual species assignments

onto these results to test the potential correspondence between naïve clustering and the existing

taxonomy (Fig. S9).

Phylogenetic reconstruction

Given the high levels of admixture among groups, we used SplitsTree v. 4.14.8 (35, 91) to generate four phylogenetic networks: one with all tiger salamander individuals, and one each for the U.S., central Mexico, and northern Mexico subgroups. Networks were constructed using uncorrected p-distances and the NeighborNet algorithm (92).

We also used three different analytical approaches to place hypothesized population lineages in a phylogenetic framework. For all analyses we used a reduced data set containing the concatenated data for 2-7 representative individuals from each population genetic cluster, which limited computation time and avoided violating the coalescent-model assumption of zero gene flow. We first inferred the phylogeny using Bayesian Inference in BEAST v.1.8.3 (37). Analyses were run for 5 million generations, sampling every 1,000 generations after the first 500,000 generations were removed as burn-in. Run convergence was assessed with Tracer v.1.6.0 (93). Next, we inferred a maximum-likelihood phylogeny using RAxML v.8 (38). Node support was assessed using a rapid bootstrap analysis with 1,000 replicates, which was summarized as a 95% rule consensus tree using the program SumTrees in the DendroPy python library (94). For both BEAST and RAxML, PartitionFinder (95) was used to identify the number of preferred gene partitions and their substitution models, and analyses were performed on the CIPRES Science Gateway server (96). Finally, we inferred phylogenetic relationships using SVDquartets (36) implemented in PAUP* version 4.0a164 (97), sampling all possible quartets and assessing node support with 1,000 bootstrap replicates. For all phylogenetic analyses, trees were visualized using FigTree v.1.4.2 (98).

Tests of migration and population differentiation of paedomorphic taxa

We used the coalescent-based program Migrate-N v.3.2 (99) to explicitly test for population structure and gene flow in each obligately paedomorphic species of *Ambystoma* (*A. andersoni, A. dumerilii, A. taylori,* and *A. mexicanum*). For each model (Fig. S14), we treated paedomorphic species as distinct populations, and the facultatively paedomorphic individuals from the same genetic cluster (CM3 or CM4) as another population. The best-fitting model was determined via Bayes factors (BF), which were calculated using Bézier-corrected marginal likelihoods (Table S3); the highest-ranking models have a BF = 0. Each Migrate-N analyses ran for 50 million steps, recording every 10 steps, with a burn-in of five million and the default heating scheme. Suitable upper bounds for priors on population size (Θ) and migration rate (M) were determined from an initial test run of 10 million steps for each analysis. Median values and 95% confidence intervals of Θ and M are in Fig. 4 and Table S4.

Within CM3 and CM4, we also calculated pairwise Φ_{ST} (a metric of population differentiation) among all populations using the function 'pairPhiST' in the R package haplotypes (100). These values were plotted against pairwise geographic distances (Fig. 4), which were calculated from locality latitude/longitude data (Table S1) and converted to kilometers using the function 'rdist.earth' in the R package fields (101). A life history matrix was also generated by assigning a value of 0 for pairwise comparisons of populations with the same life history and a value of 1 for populations with different life histories (i.e., facultatively paedomorphic vs. obligately paedomorphic). Paedomorph-paedomorph comparisons were omitted to avoid confounding them with facultative-facultative comparisons. Finally, we used these matrices in a partial mantel test [function 'mantel.partial' package vegan (102)] to test for a

correlation between Φ_{ST} and life history while correcting for geographic distance. Significance of

this test was determined using 999 permutations and an alpha level of 0.05.

DATA AVAILABILITY

Supplementary figures and tables are provided with the online version of this manuscript.

Input files for all population genetic and phylogenetic analyses are available via figshare

(https://figshare.com/projects/Life_history_strategy_does_not_reflect_genetic_differentiation_in

_the_tiger_salamander_species_complex/74115). Sequence data are available on the NCBI

Sequence Read Archive (BioProject accession PRJNA594660).

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 Figure Legends

Figure 1. Discriminant analysis of principal components (DAPC) on (A) the entire genomic
dataset (without *A. opacum* and *A. texanum* outgroups), and (B) the genomic dataset without *A*.
californiense. In both analyses, patterns of genetic differentiation stabilized at values of $K \geq 5$ (K
= 5 is shown in both plots, with colors representing genetic clusters). Points represent individuals
and ellipses show the groups identified by DAPC. The first and second principal components
from the DAPC are on the x and y axes, respectively. Results from additional values of K are
provided in Supplementary Figures 3 and 4.
753
Figure 2. Population genomic analyses of the *Ambystoma tigrinum* species complex. (A)
STRUCTURE plot of central Mexico samples. While the Evanno et al. (2005) method identified
$K = 7$, four major geographic genetic groups are apparent and were analyzed separately in
subsequent analyses (B-E). In each plot, vertical bars represent individuals, while the y-axis
shows the membership probability for each group. Pie charts mapped below the assignment plot
show the average group membership probability for that locality. Some localities were combined
to a single pie chart to reduce visual clutter; see Table S1 for specific coordinates of each sample.
The range of the Trans-Mexican Volcanic Belt (TMVB) is identified by shading and is modified
from Ferrari (2004). The range of the Cuenca Oriental is outlined in red and is modified from
Percino-Daniel et al. (2016). Two geographic outliers placed in CM2 are thought to represent
range introductions and are not shown here, but are discussed in supplementary text and shown
in Fig. S9. (F,G) STRUCTURE results from analyses of the U.S. and northern Mexico groups,
respectively. Detailed results are shown in Figs. S11-S12. Black squares on the maps denote the
type localities of all species included in this study. On each STRUCTURE membership plot,
salamander face symbols denote the presence of salamanders with a life history category of 1-3
(the cartoon lacks gills) or 4-5 (paedomorphic, cartoon shows gills); groups labeled with both
symbols contain a mixture of these life history categories.
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Figure 3. Results of phylogenetic network analyses (left) and the SVDquartets-based species
tree analysis (right) of the *Ambystoma tigrinum* species complex. Clades are colored according to
their cluster assignments in population genetic analyses, as in Fig. 2. In phylogenetic networks,
species names are only shown for paedomorphic taxa in categories 4 or 5 (Table 1). Life history
categories for all taxa are given in parentheses in the species tree. Note that SVDquartet analysis
used a subset of individuals, indicated by solid circles on the phylogenetic networks.
Phylogenetic results from Bayesian and maximum-likelihood concatenated analyses were largely
concordant with the quartets tree (Fig. S13), although two important differences are indicated by
dashed lines. Bootstrap support (BS) values are indicated on each node of the quartets tree (BS
values ≥ 95 are not shown), while tips are labeled with individual IDs (Table S1). Branch lengths
are not scaled to time or substitution rate.
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Figure 4. Maps (left) show localities for CM3 and CM4, the two genetic clusters containing
paedomorphic taxa (life history categories 4 and 5). The top-ranked migration model is shown to
the right of each map, with median mutation-scaled effective population sizes (Θ) shown inside
the shapes denoting populations. Arrows indicate the direction of migration ($M = m/\mu$, where m
is the fraction of immigrants in each generation and μ is the mutation rate per generation per site)
and median migration rates are provided next to each arrow. Full model test results are provided

in Table S3 and full parameter estimates are provided in Table S4. Scatterplots (right) show
pairwise geographic distances by pairwise ϕ_{ST} among all populations. Point colors and shapes
denote the life history strategies of the pairwise comparison being made: black squares = both
populations are facultatively paedomorphic; red circles = one population is facultatively
paedomorphic and one population is obligately paedomorphic; black stars = both populations are
obligately paedomorphic. Solid lines denote the line of best fit, calculated separately for each life
history comparison. Regressions were not performed on paedomorph-paedomorph comparisons
due to low sample sizes. Shaded areas denote the 95% confidence interval for the regression. P-
values to the right of each graph were calculated using a partial mantel test, which tested for a
correlation between ϕ_{ST} and life history while correcting for geographic distance (n.s. = not
significant).
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Table 1. Currently recognized species in the *Ambystoma tigrinum* complex. Habitat preference, range, conservation status ("Status"), and life history strategy were compiled from AmphibiaWeb (2019) and the IUCN (2019). Conservation status abbreviations: LC = least concern, V = vulnerable, E = endangered, CE = critically endangered, DD = data deficient. Life history codes: 1 = obligate metamorph (no paedomorphs documented in the field), 2 = Strong bias towards metamorphosis with rare paedomorphs found in the field, 3 = Both metamorphosis and paedomorphosis common in the field, 4 = Strong bias towards paedomorphosis with rare metamorphs found in the field, 5 = obligate paedomorph (no metamorphs documented in the field). In the text, "paedomorph" describes taxa scored as 4 or 5, while "facultative paedomorph" describes taxa scored as 2 or 3.

Species	Habitat Preference	Range	Status	Life History
Ambystoma altamirani Dugès, 1895	Streams, occasional ponds	High elevations in the west and south of the Valley of Mexico (Mexico State, Morelos, and Distrito Federal)	E	2
Ambystoma amblycephalum Taylor, 1940	Ponds	Small area of Tacicuaro (north and west Michoacan)	CE	2
Ambystoma andersoni Krebs & Brandon, 1984	Lakes / Ponds	Restricted to Lake Zacapu and its outflow streams (north-central Michoacan)	CE	5
Ambystoma bombypellum Taylor, 1940	Lakes / Ponds, occasional streams	Known only from type locality San Martin/Asunción, 15 km west of Morelia (Michoacan)	DD	2
Ambystoma californiense Gray 1853	Vernal pools	Great Central Valley and adjacent inner coast ranges, central California	V	1
Ambystoma dumerilii (Dugès, 1870)	Lakes / Ponds	Restricted to Lake Patzcauro (north-central Michoacan)	CE	5
Ambystoma flavipiperatum Dixon, 1963	Lakes / Ponds, occasional streams	Sierra de Quila (Jalisco). Historically known from the type locality in Santa Cruz, southwest of Guadalajara.	E	3
Ambystoma granulosum Taylor, 1944	Lakes / Ponds, occasional streams	Northwest of Toluca city (Mexico State and border of Michoacan)	CE	3
Ambystoma leorae (Taylor, 1943)	Streams	High elevations of Rio Frio in Iztaccihuatl -Popocatepetl Natl. Park (Mexico State and adjacent Puebla)	CE	2
Ambystoma lermaense (Taylor, 1940)	Lakes / Ponds	Highlands near Toluca, in Rio Lerma and Lake Lerma near Almoloya (Mexico State)	E	3
Ambystoma mavortium Baird, 1850*	Lakes / Ponds	Widespread distribution from west-central Canada through the USA to northern Mexico (Sonora, Chihuahua, Coahuila). Disjunct distribution in northwestern USA and Canada (Washington, British Columbia).	LC	3
Ambystoma mexicanum (Shaw and Nodder, 1798)	Lakes	Southern areas of Xochimilco (Mexico City). Historically distributed in Lakes of Xochimilco and Chalco.	CE	4
Ambystoma ordinarium Taylor, 1940	Streams	From south of Patzcuaro through Morelia City to eastern Michoacan	E	3
Ambystoma rivulare (Taylor, 1940)	Streams	Sierra Chinchua, Mariposa Monarca Biosphere Reserve, Michoacan, Valle de Bravo, Amanalco and Nevado de Toluca (Mexico State)	E	2
Ambystoma rosaceum Taylor, 1941	Streams	Sierra Madre Occidental (Sonora, Durango, Chihuahua, Sinaloa, Nayarit, Jalisco)	LC	3
Ambystoma silvense Webb, 2004	Lakes / Ponds	Sierra Madre Occidental (at least Chihuahua, Durango)	DD	3
Ambystoma taylori Brandon et al., 1982	Saline lake	Restricted to Lake Alchichica (Puebla)	CE	4
Ambystoma tigrinum Green, 1825	Lakes / Ponds	From southeast Canada south to east Texas and extending east of the Appalachian Mountains through the Atlantic Coastal Plain north to Long Island (New York)	LC	2
Ambystoma velasci (Dugès, 1888)	Lakes / Ponds, Streams	Sierra Madre Occidental (Chihuahua) and south across central Mexico (Durango, Jalisco, Nuevo León, Coahuila, Zacatecas, San Luis Potosi, Hidalgo, Queretaro, Tlaxcala, Puebla, Veracruz, Aguascalientes)	LC	3

A) DAPC of full dataset

B) DAPC without *A. californiense*

A) Central Mexico (all groups), *K* = 7

CM1 CM2 CM3 CM4

Trans-Mexican Volcanic Belt (TMVB)

B) Central Mexico 1 (CM1), *K* = 2

C) Central Mexico 2 (CM2), *K* = 2

D) Central Mexico 3 (CM3), *K* = 3

E) Central Mexico 4 (CM4), *K* = 2

Mexico City Cuenca Oriental

F) United States and Canada, *K* = 3

G) Northern Mexico, *K* = 2

Type Localities (■)

1) *Ambystoma flavipiperatum*
2) *A. andersoni*
3) *A. dumerilii*
4) *A. amblycephalum*
5) *A. ordinarium*
6) *A. rivulare*
7) *A. bombypellum*
8) *A. granulosum*
9) *A. lermaense*
10) *A. velasci*
11) *A. altamirani*
12) *A. mexicanum*
13) *A. leorae*
14) *A. taylori*
15) *A. mavortium diaboli*
16) *A. m. melanostictum*
17) *A. m. nebulosum*
18) *A. m. mavortium*
19) *A. m. stebbinsi*
20) *A. tigrinum tigrinum*
21) *A. rosaceum*
22) *A. silvense*

Paedomorph (category 4 or 5)

Facultative Paedomorph/ Metamorph (categories 1-3)

Central Mexico 4

A. mexicanum (4)

Central Mexico 3

A. taylori (4)

Central Mexico 1

Central Mexico 3

A. dumerilii (5)   *A. andersoni* (5)

Central Mexico 2

= Paedomorphic taxa (category 4 or 5)

Northern Mexico 1

Northern Mexico 2

Rocky mtn. U.S.

Eastern U.S.

Central U.S.

central Mex 4
A. taylori (4),
A. velasci (3)
3056
3087   64
1643   46   70

central Mex 4
A. mexicanum (4)
1523
1780   93
1643 60

1523
1780   93
3080   60

central Mex 2
A. bombypellum (2),
A. granulosum (3),
A. lermaense (3)
3101
1455   40
1680   57
1686   55
1347

central Mex 3
A. amblycephalum (2),
A. andersoni (5),
A. dumerilii (5),
A. flavipiperatum (3)
3136
3061   57
2024   69   81
1319
3069

central Mex 3
A. ordinarium (3)
2338
2069

central Mex 1
A. altamirani (2),
A. leorae (2),
A. rivulare (2)
1589
1578   58
1228   53
3115

northern Mex 2
A. silvense (3)
3066
3032

northern Mex 1
A. rosaceum (3)
3026
3216   48
3028   38

Rocky mtn. U.S.
A. m. nebulosum (3)
2910
2792   81
2809   83
1956

central U.S.
A. mavortium (3)
(incl. 4 subspp.)
2816
1929   61
1884   91
2765
2925   73
1993   68
2897

eastern U.S.
A. tigrinum (2)
374
2735
3134   90
2579

A. californiense (1)   1867
1842

A. texanum

A. opacum

A. talpoideum

Central Mexico Group 3 (CM3)

● *A. andersoni*
▲ *A. dumerilii*
□ Other taxa

Migration Model:

.0041
481 441 613 381
.0023 457 751 .0031

Population pair
■ Facultative-Facultative
● Facultative-Paedomorph
★ Paedomorph-Paedomorph

Partial Mantel Statistic: -0.158
p-value = 0.791 (n.s.)

Central Mexico Group 4 (CM4)

● *A. mexicanum*
▲ *A. taylori*
□ Other taxa

Migration Model:

.0047
209 891 609 679
.0023 217 841 .0033

Population pair
■ Facultative-Facultative
● Facultative-Paedomorph
★ Paedomorph-Paedomorph

Partial Mantel Statistic: 0.245
p-value = 0.18 (n.s.)