A Meta-analysis of the known Global Distribution and Host Range of the *Ralstonia* Species Complex

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Abstract

The *Ralstonia* species complex is a genetically diverse group of plant wilt pathogens. *Ralstonia* strains are classified by a "phylotype-sequevar" phylogenetic system. Since the development of the phylotype-sequevar system, over one hundred papers have described the genetic diversity of *Ralstonia* strains isolated from agriculturally important crops, ornamental plants, and plants in natural ecosystems. Our goal is to create a database that contains the reported global distribution and host range of *Ralstonia* sequevars. In this second release, we have catalogued information from 73 manuscripts that report one or more *Ralstonia* strain isolated from 88 geographic regions. The database is hosted as a GitHub repository (https://github.com/lowepowerlab/Ralstonia_Global_Diversity) that will be updated regularly.

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Introduction

Bacterial pathogens in the *Ralstonia* species complex are xylem pathogens on a broad range of agricultural and natural plant hosts. *Ralstonia* strains clog plant xylem vessels, leading plant hosts to wilt [1]. Historically, *Ralstonia* strains were classified based on carbon utilization patterns ("Biovar") and host range ("Race"). However, DNA sequence-based taxonomies more accurately reflect the evolutionary trajectories of *Ralstonia* (Fig 1).

Currently, the *Ralstonia* species complex is classified into three species: *R. solanacearum*, *R. pseudosolanacearum*, and *R. syzygii*. Strains are also classified into a phylotype system, which overlaps with the species boundaries. All *R. solanacearum* strains are within phylotype II, but phylotype II is divided into IIA and IIB groups. *R. pseudosolanacearum* strains are either in phylotype I or phylotype III. *R. syzygii* strains are in phylotype IV. Strains are further sub-classified into sequence variants, or "sequevars", based on the DNA sequence of *egl* and other reference genes. The phylotype-sequevar system developed and described by Prior and Fegan [2]. The separation of *Ralstonia* into three species was first proposed by [3], formalized by [4], and reinforced by [5]. Hundreds of papers have used the phylotype-sequevar system to describe the genetic diversity of *Ralstonia* isolates around the world.

The *Ralstonia* community typically states that *Ralstonia* strains infect over 250 plant species in over 50 botanical families. Is that an under-estimation? Our goal is to perform a meta-analysis that documents the known host range and global distribution of each sequevar in the *Ralstonia* species complex. We intend to update this preprint and the associated GitHub repository at regular intervals as we populate the database.

Figure 1. Taxonomic classification of the *Ralstonia* species complex at the species, phylotype and sequevar levels. Sequevars highlighted in red are assigned to strains in conflicting phylotypes.

Methods
Article Selection Criteria and Search Strategy:

To catalogue sequevar and geographic distributions for newly described *Ralstonia* strains, we performed a secondary analysis of the paper that cite "How Complex is the *Ralstonia solanacearum* species complex?" by Fegan and Prior 2005 [2]. Using Google Scholar, we identified the papers that both

cite Fegan and Prior, 2005 and include the word "sequevar". The most recent papers are prioritized for addition to the database in each release with the eventual goal to catalog all papers.

To be included in the database, the articles must have used the phylotype and/or phylotype-sequevar system to characterize the strains. Studies that used biovar and race identification methods without using a phylogenetic method were not included. Papers focusing on molecular biology of single or well-documented strains were also not included because they did not report information about new isolates.

Database access:

The full dataset is stored in a GitHub repository. We anticipate formal updates to the database once to twice per year. https://github.com/lowepowerlab/Ralstonia_Global_Diversity

Results and Discussion

We compiled 3395 strains from 73 papers published between 2012 and 2020 [4–77], including the common reference strains for *Ralstonia* sequevar studies. These strains represent over 64 sequevars isolated from 88 countries or territories (Table 1).

The full dataset is included as Table S1 and is hosted in the GitHub repository. For each strain, we recorded taxonomy (phylotype and sequevar), host (specific name and the host plant's taxonomic Family and Order), isolation year, isolation location, NCBI accessions (genome or partial sequences of *egl* and housekeeping genes like *mutS* and/or *rplB* genes) and the citation.

Table 1: Summary of the Sequevar database

Database Release Date	# Papers	Paper published range	# Strains	# Sequevars	# Countries or Territories	# Host plants
2020/07/03	35	2017-2020	1625	57	50	56
2021/03/23	73	2012-2020	3395	over 64[a]	88	124

[a]*As of this release, we have not investigated the sequevar assignment of strains classified into "unknown" or "new" sequevars.*

We investigated which sequevars have been isolated from multiple locations (Table 2). We classified the geographic location to either country level or to the regional level for island territories. Even with this incomplete dataset, several sequevars were identified in multiple locations. As expected [64], the IIB-1 sequevar is the most widely distributed group of *Ralstonia*. IIB-1 *Ralstonia* cause brown rot of potato and can be latently disseminated in both potato tubers and ornamental Geranium sp. [78]. This group of strains is under strict quarantine worldwide and are classified as Select Agent pathogens in the United States. An additional three phylotype II sequevars (IIB-4, IIA-6, and IIA-7 and six phylotype I sequevars (I-13, I-14, I-15, I-17, I-18, and I-31) were isolated from at least eight geographic areas.

Table 2: Widely distributed sequevars

sequevar	# countries or territories
IIB-1	41
IIB-4	12
IIA-6	9

IIA-7	8
I-13	9
I-14	10
I-15	8
I-17	8
I-18	11
I-31	10

We investigated which sequevars have been isolated from diverse host plants (Table 3). We classified the host to the species and family level, based on the information presented in the primary literature. Most of the sequevars that are widely distributed have broad host ranges. Ten sequevars have been isolated from at least 10 plant species in 5 to 17 botanical families.

Table 3: Broad host range sequevars

sequevar	# plant species	# plant families
IIB-1	15	5
IIB-4	20	7
I-13	18	12
I-14	27	17
I-15	26	14
I-17	21	10
I-18	13	8
I-31	13	5
I-44	17	13
I-48	10	5

Conclusion

Bacterial wilt pathogens in the *Ralstonia* species complex are high impact global pathogens. We created a strain database that we will regularly update to document the distribution and host range of *Ralstonia*. The second release of the database contains data from 73 papers.

References

1. Ingel B, Caldwell D, Duong F, Parkinson DY, McCulloh KA, Iyer-Pascuzzi AS, et al. Revisiting the source of wilt symptoms: X-ray microcomputed tomography provides direct evidence that Ralstonia biomass clogs xylem vessels. BioRxiv. 2021. p. 2021.03.19.436187. doi:10.1101/2021.03.19.436187

2. Fegan M, Prior P. How complex is the *Ralstonia solanacearum* species complex. Bacterial wilt disease and the *Ralstonia solanacearum* species complex. APS press St. Paul; 2005. pp. 449–461.

3. Remenant B, Coupat-Goutaland B, Guidot A, Cellier G, Wicker E, Allen C, et al. Genomes of three tomato pathogens within the Ralstonia solanacearum species complex reveal significant evolutionary divergence. BMC Genomics. 2010;11: 379.

4. Safni I, Cleenwerck I, De Vos P, Fegan M, Sly L, Kappler U. Polyphasic taxonomic revision of the Ralstonia solanacearum species complex: proposal to emend the descriptions of R. solanacearum and R. syzygii and reclassify current R. syzygii strains. Int J Syst Evol Microbiol. 1099;2014;64: 3087–103.

5. Prior P, Ailloud F, Dalsing BL, Remenant B, Sanchez B, Allen C. Genomic and proteomic evidence supporting the division of the plant pathogen *Ralstonia solanacearum* into three species. BMC Genomics. 2016;17: 90.

6. Abdurahman A, Parker ML, Kreuze J, Elphinstone JG, Struik PC, Kigundu A, et al. Molecular Epidemiology of Ralstonia solanacearum Species Complex Strains Causing Bacterial Wilt of Potato in Uganda. Phytopathology. 2019;109: 1922–1931.

7. Cho H, Song E-S, Lee YK, Lee S, Lee S-W, Jo A, et al. Analysis of Genetic and Pathogenic Diversity of Ralstonia solanacearum Causing Potato Bacterial Wilt in Korea. Plant Pathol J. 2018;34: 23–34.

8. Jimenez Madrid AM, Doyle VP, Ivey MLL. Characterization of Ralstonia solanacearum species complex strains causing bacterial wilt of tomato in Louisiana, USA. Can J Plant Pathol. 2019;41: 329–338.

9. Pardo JM, López-Alvarez D, Ceballos G, Alvarez E, Cuellar WJ. Detection of Ralstonia solanacearum phylotype II, race 2 causing Moko disease and validation of genetic resistance observed in the hybrid plantain FHIA-21. Trop Plant Pathol. 2019;44: 371–379.

10. Tan X, Qiu H, Li F, Cheng D, Zheng X, Wang B, et al. Complete Genome Sequence of Sequevar 14M Ralstonia solanacearum Strain HA4-1 Reveals Novel Type III Effectors Acquired Through Horizontal Gene Transfer. Front Microbiol. 2019;10: 1893.

11. Bergsma-Vlami M, van de Bilt JLJ, Tjou-Tam-Sin NNA, Westenberg M, Meekes ETM, Teunissen HAS, et al. Phylogenetic Assignment of Ralstonia pseudosolanacearum (Ralstonia solanacearum Phylotype I) Isolated from Rosa spp. Plant Dis. 2018;102: 2258–2267.

12. Ravelomanantsoa S, Vernière C, Rieux A, Costet L, Chiroleu F, Arribat S, et al. Molecular Epidemiology of Bacterial Wilt in the Madagascar Highlands Caused by Andean (Phylotype IIB-1) and African (Phylotype III) Brown Rot Strains of the Ralstonia solanacearum Species Complex. Front Plant Sci. 2018;8: 2258.

13. Thano P, Akarapisan A. Phylotype and sequevar of Ralstonia solanacearum which causes bacterial wilt in Curcuma alismatifolia Gagnep. Lett Appl Microbiol. 2018;66: 384–393.

14. Weibel J, Tran TM, Bocsanczy AM, Daughtrey M, Norman DJ, Mejia L, et al. A *Ralstonia solanacearum* strain from Guatemala infects diverse flower crops, including new asymptomatic hosts vinca and sutera, and causes symptoms in geranium, mandevilla vine, and new host African daisy (*Osteospermum ecklonis*). Plant Health Prog. 2016;17: 114–121.

15. Zhang YW, Chen Y, Hu CH, Li QQ, Lin W, Yuan GQ. Bacterial wilt caused by Ralstonia pseudosolanacearum (R. solanacearum phylotype I) on Luffa cylindrica in China. J Plant Pathol. 2018;100: 593–593.

16. Albuquerque GMR, Santos LA, Felix KCS, Rollemberg CL, Silva AMF, Souza EB, et al. Moko Disease-Causing Strains of Ralstonia solanacearum from Brazil Extend Known Diversity in Paraphyletic Phylotype II. Phytopathology. 2014;104: 1175–1182.

17. Du H, Chen B, Zhang X, Zhang F, Miller SA, Rajashekara G, et al. Evaluation of Ralstonia solanacearum Infection Dynamics in Resistant and Susceptible Pepper Lines Using Bioluminescence Imaging. Plant Dis. 2017;101: 272–278.

18. Freitas RG, Hermenegildo PS, Guimarães LMS, Zauza EAV, Badel JL, Alfenas AC. Detection and characterization of Ralstonia pseudosolanacearum infecting Eucalyptus sp. in Brazil. For Pathol. 2020;50: e12593.

19. Gutarra L, Herrera J, Fernandez E, Kreuze J, Lindqvist-Kreuze H. Diversity, Pathogenicity, and Current Occurrence of Bacterial Wilt Bacterium Ralstonia solanacearum in Peru. Front Plant Sci. 2017;8: 1221.

20. Kyaw HWW, Tsuchiya K, Matsumoto M, Iiyama K, Aye SS, Zaw M, et al. Genetic diversity of Ralstonia solanacearum strains causing bacterial wilt of solanaceous crops in Myanmar. J Gen Plant Pathol. 2017;83: 216–225.

21. Liu Y, Wu D, Liu Q, Zhang S, Tang Y, Jiang G, et al. The sequevar distribution of Ralstonia solanacearum in tobacco-growing zones of China is structured by elevation. Eur J Plant Pathol. 2017;147: 541–551.

22. Obrador-Sánchez JA, Tzec-Simá M, Higuera-Ciapara I, Canto-Canché B. Genetic diversity of Ralstonia solanacearum strains from Mexico associated with Moko disease. Eur J Plant Pathol. 2017;149: 817–830.

23. Patil VU, Girimalla V, Sagar V, Chauhan RS, Chakrabarti SK. Genome sequencing of four strains of Phylotype I, II and IV of Ralstonia solanacearum that cause potato bacterial wilt in India. Braz J Microbiol. 2017;48: 193–195.

24. Rossato M, Santiago TR, Mizubuti ESG, Lopes CA. Characterization and pathogenicity to geranium of Brazilian strains of Ralstonia spp. Trop Plant Pathol. 2017;42: 458–467.

25. She X, Yu L, Lan G, Tang Y, He Z. Identification and Genetic Characterization of Ralstonia solanacearum Species Complex Isolates from Cucurbita maxima in China. Front Plant Sci. 2017;8: 1794.

26. Wang L, Wang B, Zhao G, Cai X, Jabaji S, Seguin P, et al. Genetic and pathogenic diversity of Ralstonia solanacearum causing potato brown rot in China. Am J Potato Res. 2017;94: 403–416.

27. Pastou D, Chéron JJ, Cellier G, Guérin F, Poussier S. First Report of Ralstonia pseudosolanacearum Phylotype I Causing Bacterial Wilt in New Caledonia. Plant Dis. 2020;104: 278.

28. Guarischi-Sousa R, Puigvert M, Coll NS, Siri MI, Pianzzola MJ, Valls M, et al. Complete genome sequence of the potato pathogen Ralstonia solanacearum UY031. Stand Genomic Sci. 2016;11: 7.

29. Ramírez M, Moncada RN, Villegas-Escobar V, Jackson RW, Ramírez CA. Phylogenetic and pathogenic variability of strains of Ralstonia solanacearum causing moko disease in Colombia. Plant Pathol. 2020;69: 360–369.

30. Albuquerque GMR, Souza EB, Silva AMF, Lopes CA, Boiteux LS, Fonseca ME de N. Genome Sequence of Ralstonia pseudosolanacearum Strains with Compatible and Incompatible Interactions with the Major Tomato Resistance Source Hawaii 7996. Genome Announc. 2017;5. doi:10.1128/genomeA.00982-17

31. She X, He Z, Li H. Genetic structure and phylogenetic relationships ofRalstonia solanacearumstrains from diverse origins in Guangdong Province, China. J Phytopathol. 2018;166: 177–186.

32. Caruso P, Biosca EG, Bertolini E, Marco-Noales E, Gorris MT, Licciardello C, et al. Genetic diversity reflects geographical origin of Ralstonia solanacearum strains isolated from plant and water sources in Spain. Int Microbiol. 2017;20: 155–164.

33. Shutt VM, Shin G, van der Waals JE, Goszczynska T, Coutinho TA. Characterization of Ralstonia strains infecting tomato plants in South Africa. Crop Prot. 2018;112: 56–62.

34. Yahiaoui N, Chéron J-J, Ravelomanantsoa S, Hamza AA, Petrousse B, Jeetah R, et al. Genetic Diversity of the Ralstonia solanacearum Species Complex in the Southwest Indian Ocean Islands. Front Plant Sci. 2017;8: 2139.

35. Santiago TR, Lopes CA, Caetano-Anollés G, Mizubuti ESG. Genetic Structure of Ralstonia solanacearum and Ralstonia pseudosolanacearum in Brazil. Plant Dis. 2020;104: 1019–1025.

36. Sedighian N, Krijger M, Taparia T, Taghavi SM, Wicker E, Van Der Wolf JM, et al. Genome Resource of Two Potato Strains of Ralstonia solanacearum Biovar 2 (Phylotype IIB Sequevar 1) and Biovar 2T (Phylotype IIB Sequevar 25) Isolated from Lowlands in Iran. Mol Plant Microbe Interact. 2020;33: 872–875.

37. Vasconez IN, Besoain X, Vega-Celedón P, Valenzuela M, Seeger M. First report of bacterial wilt caused by Ralstonia solanacearum phylotype IIB sequevar 1 affecting tomato in different regions of Chile. Plant Dis. 2020;104: 2023.

38. Bocsanczy AM, Espindola AS, Norman DJ. Whole-Genome Sequences of Ralstonia solanacearum Strains P816, P822, and P824, Emerging Pathogens of Blueberry in Florida. Microbiol Resour Announc. 2019;8. doi:10.1128/MRA.01316-18

39. Hayes MM, MacIntyre AM, Allen C. Complete Genome Sequences of the Plant Pathogens Ralstonia solanacearum Type Strain K60 and R. solanacearum Race 3 Biovar 2 Strain UW551. Genome Announc. 2017;5. doi:10.1128/genomeA.01088-17

40. Izadiyan M, Taghavi SM. Characterization of Iranian Ralstonia solanacearum biovar 2 strains by partial sequencing of egl, mutS and pga genes. Australas Plant Pathol. 2019;48: 607–615.

41. Cellier G, Remenant B, Chiroleu F, Lefeuvre P, Prior P. Phylogeny and population structure of brown rot- and Moko disease-causing strains of Ralstonia solanacearum phylotype II. Appl Environ Microbiol. 2012;78: 2367–2375.

42. Hong JC, Norman DJ, Reed DL, Momol MT, Jones JB. Diversity Among Ralstonia solanacearum Strains Isolated from the Southeastern United States. Phytopathology. 2012;102: 924–936.

43. N'guessan CA, Abo K, Fondio L, Chiroleu F, Lebeau A, Poussier S, et al. So near and yet so far: the specific case of Ralstonia Solanacearum populations from Côte d'Ivoire in Africa. Phytopathology. 2012;102: 733–740.

44. Garcia AL, Lima WG, Souza EB, Michereff SJ, Mariano RLR. CHARACTERIZATION OF RALSTONIA SOLANACEARUM CAUSING BACTERIAL WILT IN BELL PEPPER IN THE STATE OF PERNAMBUCO, BRAZIL. J Plant Pathol. 2013;95: 237–245.

45. Izadiyan M, Taghavi SM. HOST RANGE VARIATION AND GENETIC DIVERSITY OF IRANIAN ISOLATES OF RALSTONIA SOLANACEARUM FROM POTATO AND TOMATO WITH RAPD AND (GTG)5-PCR. J Plant Pathol. 2013;95: 87–97.

46. Lebeau A, Gouy M, Daunay MC, Wicker E, Chiroleu F, Prior P, et al. Genetic mapping of a major dominant gene for resistance to Ralstonia solanacearum in eggplant. Theor Appl Genet. 2013;126: 143–158.

47. N'guessan CA, Brisse S, Le Roux-Nio A-C, Poussier S, Koné D, Wicker E. Development of variable number of tandem repeats typing schemes for Ralstonia solanacearum, the agent of bacterial wilt, banana Moko disease and potato brown rot. J Microbiol Methods. 2013;92: 366–374.

48. Pan ZC, Xu J, Prior P, Xu JS, Zhang H, Chen KY, et al. Development of a specific molecular tool for the detection of epidemiologically active mulberry causing-disease strains of *Ralstonia solanacearum* phylotype I (historically race 5-biovar 5) in China. Eur J Plant Pathol. 2013;137: 377–391.

49. Parkinson N, Bryant R, Bew J, Conyers C, Stones R, Alcock M, et al. Application of variable-number tandem-repeat typing to discriminate Ralstonia solanacearum strains associated with English watercourses and disease outbreaks. Appl Environ Microbiol. 2013;79: 6016–6022.

50. Waki T, Horita M, Kurose D, Mulya K, Tsuchiya K. Genetic Diversity of Zingiberaceae Plant Isolates of *Ralstonia solanacearum* in the Asia-Pacific Region. Japan Agricultural Research Quarterly: JARQ. 2013;47: 283–294.

51. Lin C-H, Tsai K-C, Prior P, Wang J-F. Phylogenetic relationships and population structure ofRalstonia solanacearumisolated from diverse origins in Taiwan. Plant Pathol. 2014;63: 1395–1403.

52. Fonseca NR, Guimarães LMS, Hermenegildo PS, Teixeira RU, Lopes CA, Alfenas AC. Molecular characterization ofRalstonia solanacearuminfectingEucalyptusspp. in Brazil. For Pathol. 2014;44: 107–116.

53. Horita M, Tsuchiya K, Suga Y, Yano K, Waki T, Kurose D, et al. Current classification of Ralstonia solanacearum and genetic diversity of the strains in Japan. J Gen Plant Pathol. 2014;80: 455–465.

54. Kumar A, Prameela TP, Panja B. Genetic characterization of an Indian isolate of *Ralstonia solanacearum* race 3/ biovar 2/ phylotype IIB from potato. 2014 [cited 24 Mar 2021]. Available: https://api.semanticscholar.org/CorpusID:82677467

55. Ramesh R, Achari GA, Gaitonde S. Genetic diversity of *Ralstonia solanacearum* infecting solanaceous vegetables from India reveals the existence of unknown or newer sequevars of Phylotype I strains. Eur J Plant Pathol. 2014;140: 543–562.

56. Sagar V, Jeevalatha A, Mian S, Chakrabarti SK, Gurjar MS, Arora RK, et al. Potato bacterial wilt in India caused by strains of phylotype I, II and IV of *Ralstonia solanacearum*. Eur J Plant Pathol. 2014;138: 51–65.

57. Siri MI, Sanabria A, Boucher C, Pianzzola MJ. New type IV pili-related genes involved in early stages of *Ralstonia solanacearum* potato infection. Mol Plant Microbe Interact. 2014;27: 712–724.

58. Sujeeun L. Rôle des effecteurs de type III dans l'interaction *Ralstonia solanacearum*/ aubergine: approches de génétique évolutive et fonctionnelle. Université de la Réunion. 2014. Available: https://agritrop.cirad.fr/573610/

59. Zulperi D, Sijam K, Ahmad ZAM, Awang Y, Mohd Hata E. Phylotype classification of *Ralstonia solanacearum* biovar 1 strains isolated from banana (*Musa* spp.) in Malaysia. Archives of Phytopathology and Plant Protection. 2014;47: 2352–2364.

60. Ajitomi A, Inoue Y, Horita M, Nakaho K. Bacterial wilt of three *Curcuma* species, *C. longa* (turmeric), *C. aromatica* (wild turmeric) and *C. zedoaria* (zedoary) caused by *Ralstonia solanacearum* in Japan. J Gen Plant Pathol. 2015;81: 315–319.

61. Albuquerque P, Marcal ARS, Caridade C, Costa R, Mendes MV, Tavares F. A quantitative hybridization approach using 17 DNA markers for identification and clustering analysis of *Ralstonia solanacearum*. Plant Pathol. 2015;64: 1270–1283.

62. Ayin CM, Schlub RL, Yasuhara-Bell J, Alvarez AM. Identification and characterization of bacteria associated with decline of ironwood (*Casuarina equisetifolia*) in Guam. Australas Plant Pathol. 2015;44: 225–234.

63. Cellier G, Moreau A, Chabirand A, Hostachy B, Ailloud F, Prior P. A duplex PCR assay for the detection of *Ralstonia solanacearum* phylotype II strains in *Musa* spp. PLoS One. 2015;10: e0122182.

64. Clarke CR, Studholme DJ, Hayes B, Runde B, Weisberg A, Cai R, et al. Genome-Enabled Phylogeographic Investigation of the Quarantine Pathogen Ralstonia solanacearum Race 3 Biovar 2 and Screening for Sources of Resistance Against Its Core Effectors. Phytopathology. 2015;105: 597–607.

65. Naranjo Feliciano E, Yglesia Lozano A, García A, Martínez Zubiaur Y. Diversidad genética de aislados de *Ralstonia solanacearum* de Cuba, mediante amplificación de las regiones repetitivas del genoma (REP-PCR). Rev Protección Veg. 2015;30: 52–59.

66. Gurjar MS, Sagar V, Bag TK, Singh BP, Sharma S, Jeevalatha A, et al. Genetic diversity of *Ralstonia solanacearum* strains causing bacterial wilt of potato in the Meghalaya state of India. J Plant Pathol. 2015;97: 135–142.

67. Gutarra L, Kreuze J, Lindqvist-Kreuze H, De Mendiburu F. Variation of Resistance to Different Strains of *Ralstonia Solanacearum* in Highland Tropics Adapted Potato Genotypes. Am J Potato Res. 2015;92: 258–265.

68. Lopes CA, Rossato M, Boiteux LS. The Host Status Of Coffee (*Coffea arabica*) To *Ralstonia solanacearum* Phylotype I Isolates. Trop Plant Pathol. 2015;40: 1–4.

69. Sikirou R, Zocli B, Paret ML, Deberdt P, Coranson-Beaudu R, Huat J, et al. First Report of Bacterial Wilt of Gboma (*Solanum macrocarpon*) Caused by *Ralstonia solanacearum* in Benin. Plant Dis. 2015;99: 1640–1640.

70. Stulberg MJ, Huang Q. A TaqMan-Based Multiplex qPCR Assay and DNA Extraction Method for Phylotype IIB Sequevars 1 & 2 (Select Agent) Strains of *Ralstonia solanacearum*. PLoS One. 2015;10: e0139637.

71. Stulberg MJ, Shao J, Huang Q. A Multiplex PCR Assay to Detect and Differentiate Select Agent Strains of *Ralstonia solanacearum*. Plant Dis. 2015;99: 333–341.

72. Wang H, Huang Y, Wang J, Wang M, Xia H, Lu H. Phenotypic fingerprints of *Ralstonia solanacearum* biovar 3 strains from tobacco and tomato in China assessed by phenotype MicroArray analysis. Plant Pathol J. 2015;14: 38–43.

73. Li Y, Feng J, Liu H, Wang L, Hsiang T, Li X, et al. Genetic Diversity and Pathogenicity of *Ralstonia solanacearum* Causing Tobacco Bacterial Wilt in China. Plant Dis. 2016;100: 1288–1296.

74. Ravelomanantsoa S, Robène I, Chiroleu F, Guérin F, Poussier S, Pruvost O, et al. A novel multilocus variable number tandem repeat analysis typing scheme for African phylotype III strains of the *Ralstonia solanacearum* species complex. PeerJ. 2016;4: e1949.

75. Sánchez JAO. Variabilidad genética de la bacteria *Ralstonia solanacearum* de cepas aisladas de plátano en México. Centro de Investigación Científica de Yucatán. 2016. Available: https://pdfs.semanticscholar.org/2a5f/21145faea3ad1b56ddb52117000a45c50268.pdf

76. Ayana G. Diversity and Phylotype Analysis of *Ralstonia solanacearum* Strains Causing Tomato and Potato Bacterial Wilt in Ethiopia. Pest Management Journal of Ethiopia. 2016 [cited 24 Mar 2021]. Available: https://www.researchgate.net/profile/Muluken_Goftishu/publication/315537001_Evaluation_of_Neem_Seed_and_Citrus_Peel_Powder_for_the_Management_of_Maize_Weevil_Sitophilus_zeamais_Motsch_Coleoptera_Curculionidae_in_Sorghum/links/59079aa20f7e9bc0d59a7d10/Evaluation-of-Neem-Seed-and-Citrus-Peel-Powder-for-the-Management-of-Maize-Weevil-Sitophilus-zeamais-Motsch-Coleoptera-Curculionidae-in-Sorghum.pdf

77. Waki T, Horita M. Grouping of *Ralstonia solanacearum* strains in Tochigi Prefecture based on pathogenicity to Solanum plants and discriminating pathogenicity groups by PCR. Jpn J Phytopathol. 2014;80: 229–234.

78. Swanson JK, Montes L, Mejia L, Allen C. Detection of Latent Infections of Ralstonia solanacearum Race 3 Biovar 2 in Geranium. Plant Dis. 2007;91: 828–834.