

A Meta-analysis of the known Global Distribution and Host Range of the *Ralstonia* Species Complex

Tiffany Lowe-Power* and Kyle Chipman

University of California Davis

Abstract

The *Ralstonia* species complex is a genetically diverse group of plant wilt pathogens. *Ralstonia* strains are classified by a “phyloptype-sequevar” phylogenetic system. Since the development of the phylotype-sequevar system, over one hundred papers have described the genetic diversity of *Ralstonia* strains isolated from agriculturally important crops, ornamental plants, and plants in natural ecosystems. Our goal is to create a database that contains the reported global distribution and host range of *Ralstonia* sequevars. In this first release, we have catalogued information from 35 manuscripts that report one or more *Ralstonia* strain isolated from 50 geographic regions. The database is hosted as a GitHub repository (https://github.com/lowepowerlab/Ralstonia_Global_Diversity) that will be updated regularly.

* Corresponding author: tlowepower@ucdavis.edu

Introduction

Bacterial pathogens in the *Ralstonia* species complex cause wilting diseases on a broad range of agricultural and natural plant hosts. Historically, *Ralstonia* strains were classified based on carbon utilization patterns (Biovar) and host range (Race). However, DNA sequence-based taxonomies better reflect the evolutionary trajectories of *Ralstonia* (Fig 1).

Currently, the *Ralstonia* species complex is divided into three species: *R. solanacearum*, *R. pseudosolanacearum*, and *R. syzygii*. Strains are also classified into a phylotype system, which overlaps with the species boundaries. All *R. solanacearum* strains are within phylotype II, but phylotype II is divided into IIA and IIB groups. *R. pseudosolanacearum* strains are either in phylotype I or phylotype III. *R. pseudosolanacearum* strains are in phylotype IV. Strains are further sub-classified into sequence variants, or “sequevars”, based on the DNA sequence of *egl* and other reference genes. The phylotype-sequevar system developed and described by Prior and Fegan [1]. The separation of *Ralstonia* into three species was first proposed by [2], formalized by [3], and reinforced by [4]. Hundreds of papers have used the phylotype-sequevar system to describe the genetic diversity of *Ralstonia* isolates around the world.

The *Ralstonia* community typically states that *Ralstonia* strains infect over 250 plant species in over 50 botanical families. Is that an under-estimation? Our goal is to perform a meta-analysis that documents the known host range and global distribution of each sequevar in the *Ralstonia* species complex. We intend to update this preprint and the associated GitHub repository at regular intervals as we populate the database.

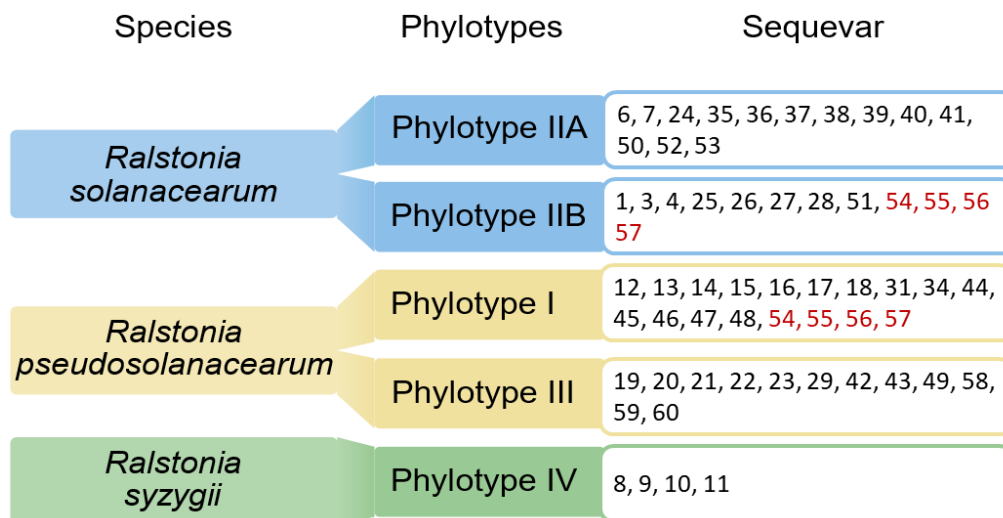


Figure 1. Taxonomic classification of the *Ralstonia* species complex at the species, phylotype and sequevar levels. Sequevars highlighted in red are assigned to strains in conflicting phylotypes.

Methods

Article Selection Criteria and Search Strategy:

To catalogue sequevar and geographic distributions for newly described *Ralstonia* strains, we performed a secondary analysis of the paper that cite “How Complex is the *Ralstonia solanacearum* species complex?” by Fegan and Prior 2005 [1]. Using Google Scholar, we identified the papers that both

cite Fegan and Prior, 2005 and include the word “sequevar”. The most recent papers are prioritized for addition to the database in each release with the eventual goal to catalog all papers.

To be included in the database, the articles must have used the phylotype and/or phylotype-sequevar system to characterize the strains. Studies that used biovar and race identification methods without using a phylogenetic method were not included. Papers focusing on molecular biology of single or well-documented strains were also not included because they did not report information about new isolates.

Database access:

The full dataset is stored in a GitHub repository. We anticipate formal updates to the database once to twice per year. https://github.com/lowepowerlab/Ralstonia_Global_Diversity

Results and Discussion

We compiled 1625 strains from 35 papers mostly published between 2017 and 2020 [5–39], including the common reference strains for *Ralstonia* sequevar studies. These strains represent 57 sequevars isolated from 50 countries or territories (Table 1).

The full dataset is included as Table S1 and is hosted in the GitHub repository. For each strain, we recorded taxonomy (phylotype and sequevar), host (specific name and the host plant’s taxonomic Family and Order), isolation year, isolation location, NCBI accessions (genome or partial sequences of *egl* and housekeeping genes like *mutS* and/or *rpIB* genes) and the citation.

Table 1: Summary of the Sequevar database

Database	Paper published	# Countries or Territories	# Host plants ^a			
Release Date	# Papers	range	# Strains	# Sequevars		
2020/07/03	35	2017-2020	1625	57	50	56

^a As of this release, we have not guaranteed that host plants are listed in a systematic format

We investigated which sequevars have been isolated from multiple locations (Table 2). We classified the geographic location to either country level or to the regional level for island territories. Even with this incomplete dataset, several sequevars were identified in multiple locations. As expected [40], the IIB-1 sequevar is the most widely distributed group of *Ralstonia*. IIB-1 *Ralstonia* cause brown rot of potato and can be latently disseminated in both potato tubers and ornamental Geranium sp. [41]. This group of strains is under strict quarantine worldwide and are classified as Select Agent pathogens in the United States. An additional six phylotype I sequevars (I-13, I-14, I-15, I-18, I-31, and I-33) and a phylotype IIA sequevar (IIA-6) were isolated from at least five geographic areas.

Table 2: Broadly distributed sequevars

sequevar	# countries or territories
IIB-1	19
IIA-6	6
I-13	6
I-14	5

I-15	8
I-18	9
I-31	11
I-33	5

Conclusion

Bacterial wilt pathogens in the *Ralstonia* species complex are high impact global pathogens. We created a strain database that we will regularly update to document the distribution and host range of *Ralstonia*. The first release of the database contains data from 35 papers.

References

1. Fegan M, Prior P. How complex is the “*Ralstonia solanacearum* species complex”? In: Allen C, Prior P, Hayward AC, editors. Bacterial Wilt Disease and the *Ralstonia solanacearum* species complex. St. Paul: APS Press; 2005. pp. 449–461.
2. Remenant B, Coupat-Goutaland B, Guidot A, Cellier G, Wicker E, Allen C, et al. Genomes of three tomato pathogens within the *Ralstonia solanacearum* species complex reveal significant evolutionary divergence. BMC Genomics. 2010;11: 379. doi:10.1186/1471-2164-11-379
3. Safni I, Cleenwerck I, De Vos P, Fegan M, Sly L, Kappler U. Polyphasic taxonomic revision of the *Ralstonia solanacearum* species complex: proposal to emend the descriptions of *R. solanacearum* and *R. syzygii* and reclassify current *R. syzygii* strains. Int J Syst Evol Microbiol. 2014;64: 3087–103. doi:10.1099/ijs.0.066712-0
4. Prior P, Ailloud F, Dalsing BL, Remenant B, Sanchez B, Allen C. Genomic and proteomic evidence supporting the division of the plant pathogen *Ralstonia solanacearum* into three species. BMC Genomics. BMC Genomics; 2016;17: 90. doi:10.1186/s12864-016-2413-z
5. Abdurahman A, Parker ML, Kreuze J, Elphinstone JG, Struik PC, Kigundu A, et al. Molecular Epidemiology of *Ralstonia solanacearum* Species Complex Strains Causing Bacterial Wilt of Potato in Uganda. Phytopathology. 2019;109: 1922–1931. doi:10.1094/PHYTO-12-18-0476-R
6. Cho H, Song ES, Lee YK, Lee S, Lee SW, Jo A, et al. Analysis of genetic and pathogenic diversity of *Ralstonia solanacearum* causing potato bacterial wilt in Korea. Plant Pathol J. 2018;34: 23–34. doi:10.5423/PPJ.FT.09.2017.0203
7. Jimenez Madrid AM, Doyle VP, Ivey MLL. Characterization of *Ralstonia solanacearum* species complex strains causing bacterial wilt of tomato in Louisiana, USA. Can J Plant Pathol. Taylor & Francis; 2019;41: 329–338. doi:10.1080/07060661.2019.1584588
8. Pardo JM, López-Alvarez D, Ceballos G, Alvarez E, Cuellar WJ. Detection of *Ralstonia solanacearum* phylotype II, race 2 causing Moko disease and validation of genetic resistance observed in the hybrid plantain FHIA-21. Trop Plant Pathol. Tropical Plant Pathology; 2019;44: 371–379. doi:10.1007/s40858-019-00282-3
9. Tan X, Qiu H, Li F, Cheng D, Zheng X, Wang B, et al. Complete genome sequence of sequevar 14 *Ralstonia solanacearum* strain HA4-1 reveals novel type III effectors acquired through horizontal gene transfer. Front Microbiol. 2019;10: 1–16. doi:10.3389/fmicb.2019.01893

10. Bergsma-Vlami M, van de Bilt JIJ, Tjou-Tam-Sin NNA, Westenberg M, Meekes ETM, Teunissen HAS, et al. Phylogenetic Assignment of *Ralstonia pseudosolanacearum* (*Ralstonia solanacearum* Phylotype I) Isolated from *Rosa* spp. *Plant Dis.* 2018;102: 2258–2267. doi:10.1094/PDIS-09-17-1345-RE
11. Ravelomanantsoa S, Vernière C, Rieux A, Costet L, Chiroleu F, Arribat S, et al. Molecular epidemiology of bacterial wilt in the Madagascar highlands caused by Andean (Phylotype IIB-1) and African (Phylotype III) brown rot strains of the *Ralstonia solanacearum* species complex. *Front Plant Sci.* 2018;8: 1–17. doi:10.3389/fpls.2017.02258
12. Thano P, Akarapisan A. Phylotype and sequevar of *Ralstonia solanacearum* which causes bacterial wilt in *Curcuma alismatifolia* Gagnep. *Lett Appl Microbiol.* 2018;66: 384–393. doi:10.1111/lam.12857
13. Weibel J, Tran TM, Bocsanczy AM, Daughtrey M, Norman DJ, Mejia L, et al. A *Ralstonia solanacearum* strain from Guatemala infects diverse flower crops, including new asymptomatic hosts vinca and sutera, and causes symptoms in geranium, mandevilla vine, and new host African Daisy (*Osteospermum ecklonis*). *Plant Heal Prog.* 2016;17: 114–121. doi:10.1094/PHP-RS-16-0001
14. Zhang YW, Chen YY, Hu CH, Li QQ, Lin W, Yuan GQ. Bacterial wilt caused by *Ralstonia pseudosolanacearum* (*R. solanacearum* phylotype I) on *Luffa cylindrica* in China. *J Plant Pathol. Journal of Plant Pathology*; 2018;100: 593. doi:10.1007/s42161-018-0098-7
15. Albuquerque GMR, Souza EB, Silva AMF, Lopes CA, Boiteux LS, Fonseca ME de N. Genome sequence of *Ralstonia pseudosolanacearum* strains with compatible and incompatible interactions with the major tomato resistance source Hawaii 7996. *Genome Announc.* 2017;5: 5–6. doi:10.1128/genomeA.00982-17
16. Du H, Chen B, Zhang X, Zhang F, Miller SA, Rajashekara G, et al. Evaluation of *Ralstonia solanacearum* infection dynamics in resistant and susceptible pepper lines using bioluminescence imaging. *Plant Dis.* 2017;101: 272–278. doi:10.1094/PDIS-05-16-0714-RE
17. Freitas RG, Hermenegildo PS, Guimarães LMS, Zauza EAV, Badel JL, Alfenas AC. Detection and characterization of *Ralstonia pseudosolanacearum* infecting *Eucalyptus* sp. in Brazil. *For Pathol.* 2020; 1–9. doi:10.1111/efp.12593
18. Gutarra L, Herrera J, Fernandez E, Kreuze J, Lindqvist-Kreuzer H. Diversity, pathogenicity, and current occurrence of bacterial wilt bacterium *Ralstonia solanacearum* in Peru. *Front Plant Sci.* 2017;8: 1–12. doi:10.3389/fpls.2017.01221
19. Kyaw HWW, Tsuchiya K, Matsumoto M, Iiyama K, Aye SS, Zaw M, et al. Genetic diversity of *Ralstonia solanacearum* strains causing bacterial wilt of solanaceous crops in Myanmar. *J Gen Plant Pathol. Springer Japan*; 2017;83: 216–225. doi:10.1007/s10327-017-0720-0
20. Liu Y, Wu D, Liu Q, Zhang S, Tang Y, Jiang G, et al. The sequevar distribution of *Ralstonia solanacearum* in tobacco-growing zones of China is structured by elevation. *Eur J Plant Pathol. European Journal of Plant Pathology*; 2017;147: 541–551. doi:10.1007/s10658-016-1023-6

21. Obrador-Sánchez JA, Tzec-Simá M, Higuera-Ciapara I, Canto-Canché B. Genetic diversity of *Ralstonia solanacearum* strains from Mexico associated with Moko disease. *Eur J Plant Pathol. European Journal of Plant Pathology*; 2017;149: 817–830. doi:10.1007/s10658-017-1228-3
22. Patil VU, Girimalla V, Sagar V, Chauhan RS, Chakrabarti SK. Genome sequencing of four strains of Phylotype I, II and IV of *Ralstonia solanacearum* that cause potato bacterial wilt in India. *Brazilian J Microbiol. Sociedade Brasileira de Microbiologia*; 2017;48: 193–195. doi:10.1016/j.bjm.2016.10.016
23. Rossato M, Santiago TR, Mizubuti ESG, Lopes CA. Characterization and pathogenicity to geranium of Brazilian strains of *Ralstonia* spp. *Trop Plant Pathol. Tropical Plant Pathology*; 2017;42: 458–467. doi:10.1007/s40858-017-0177-x
24. She X, Yu L, Lan G, Tang Y, He Z. Identification and genetic characterization of *Ralstonia solanacearum* species complex isolates from *Cucurbita maxima* in China. *Front Plant Sci.* 2017;8. doi:10.3389/fpls.2017.01794
25. Wang L, Wang B, Zhao G, Cai X, Jabaji S, Seguin P, et al. Genetic and Pathogenic Diversity of *Ralstonia solanacearum* Causing Potato Brown Rot in China. *Am J Potato Res. American Journal of Potato Research*; 2017;94: 403–416. doi:10.1007/s12230-017-9576-2
26. Pastou D, Chéron JJ, Cellier G, Guérin F, Poussier S. First Report of *Ralstonia pseudosolanacearum* Phylotype I Causing Bacterial Wilt in New Caledonia. *Plant Dis.* 2020;104: 278. doi:10.1094/PDIS-05-19-1068-PDN
27. Guarischi-Sousa R, Puigvert M, Coll NS, Siri MI, Pianzola MJ, Valls M, et al. Complete genome sequence of the potato pathogen *Ralstonia solanacearum* UY031. *Stand Genomic Sci. Standards in Genomic Sciences*; 2016;11: 1–8. doi:10.1186/s40793-016-0131-4
28. Ramírez M, Moncada RN, Villegas-Escobar V, Jackson RW, Ramírez CA. Phylogenetic and pathogenic variability of strains of *Ralstonia solanacearum* causing Moko disease in Colombia. *Plant Pathol.* 2020;69: 360–369. doi:10.1111/ppa.13121
29. Albuquerque GMR, Santos LA, Felix KCS, Rollemberg CL, Silva AMF, Souza EB, et al. Moko Disease-Causing Strains of *Ralstonia solanacearum* from Brazil Extend Known Diversity in Paraphyletic Phylotype II. *Phytopathology.* 2014;104: 1175–1182. doi:10.1094/PHYTO-12-13-0334-R
30. She X, He Z, Li H. Genetic structure and phylogenetic relationships of *Ralstonia solanacearum* strains from diverse origins in Guangdong Province, China. *J Phytopathol.* 2018;166: 177–186. doi:10.1111/jph.12674
31. Caruso P, Biosca EG, Bertolini E, Marco-Noales E, Gorris MT, Licciardello C, et al. Genetic diversity reflects geographical origin of *Ralstonia solanacearum* strains isolated from plant and water sources in Spain. *Int Microbiol.* 2017;20: 155–164. doi:10.2436/20.1501.01.298
32. Shutt VM, Shin G, van der Waals JE, Goszczynska T, Coutinho TA. Characterization of *Ralstonia* strains infecting tomato plants in South Africa. *Crop Prot. Elsevier*; 2018;112: 56–62. doi:10.1016/j.cropro.2018.05.013

33. Yahiaoui N, Chéron JJ, Ravelomanantsoa S, Hamza AA, Petrousse B, Jeetah R, et al. Genetic diversity of the *Ralstonia solanacearum* species complex in the Southwest Indian Ocean islands. *Front Plant Sci.* 2017;8. doi:10.3389/fpls.2017.02139
34. Santiago TR, Lopes CA, Caetano-Anollés G, Mizubuti ESG. Genetic Structure of *Ralstonia solanacearum* and *Ralstonia pseudosolanacearum* in Brazil. *Plant Dis.* 2020;104: 1019–1025. doi:10.1094/PDIS-09-19-1929-RE
35. Sedighian N, Krijger M, Taparia T, Taghavi SM, Wicker E, Van Der Wolf JM, et al. Genome resource of two potato strains of *Ralstonia solanacearum* biovar 2 (phylotype IIB/sequovar 1) and biovar 2T (phylotype IIB/sequovar 25) isolated from lowlands in Iran. *Mol Plant-Microbe Interact.* 2020;33: 872–875. doi:10.1094/MPMI-02-20-0026-A
36. Vasconez IN, Besoain X, Vega-Celedón P, Valenzuela M, Seeger M. First Report of Bacterial Wilt Caused by *Ralstonia solanacearum* Phylotype IIB Sequovar 1 Affecting Tomato in Different Regions of Chile. *Plant Dis.* 2020; PDIS-01-20-0181. doi:10.1094/pdis-01-20-0181-pdn
37. Bocsanczy AM, Espindola AS, Norman DJ. Whole-Genome Sequences of *Ralstonia solanacearum* Strains P816, P822, and P824, Emerging Pathogens of Blueberry in Florida. Maresca JA, editor. *Microbiol Resour Announc.* 2019;8: 17–18. doi:10.1128/MRA.01316-18
38. Hayes MM, MacIntyre AM, Allen C. Complete Genome Sequences of the Plant Pathogens *Ralstonia solanacearum* Type Strain K60 and *R. solanacearum* Race 3 Biovar 2 Strain UW551. *Genome Announc.* 2017;5: 1–2. doi:10.1128/genomeA.01088-17
39. Izadiyan M, Taghavi SM. Characterization of Iranian *Ralstonia solanacearum* biovar 2 strains by partial sequencing of *egl*, *mutS* and *pga* genes. *Australas Plant Pathol.* 2019;48: 607–615. doi:10.1007/s13313-019-00664-w
40. Clarke CR, Studholme DJ, Hayes B, Runde B, Weisberg A, Cai R, et al. Genome-Enabled Phylogeographic Investigation of the Quarantine Pathogen *Ralstonia solanacearum* Race 3 Biovar 2 and Screening for Sources of Resistance Against Its Core Effectors. *Phytopathology.* 2015;105: 597–607. doi:10.1094/PHYTO-12-14-0373-R
41. Swanson JK, Montes L, Mejia L, Allen C. Detection of latent infections of *Ralstonia solanacearum* race 3 biovar 2 in geranium. *Plant Dis.* 2007;91: 828–834. doi:10.1094/PDIS-91-7-0828