

1 **Identifying loci under selection via**  
2 **explicit demographic models**

3

4 **Hirzi Luqman<sup>1\*</sup>, Alex Widmer<sup>1</sup>, Simone Fior<sup>1†</sup>, Daniel**  
5 **Wegmann<sup>2,3†\*</sup>**

6

7 <sup>1</sup> Institute of Integrative Biology, ETH Zurich, Universitätstrasse 16, 8092 Zürich,  
8 Switzerland

9 <sup>2</sup> Department of Biology, University of Fribourg, Fribourg, Switzerland

10 <sup>3</sup> Swiss Institute of Bioinformatics, Fribourg, Switzerland

11 † denotes joint senior authors

12

13 Corresponding authors: [hirzi.luqman@env.ethz.ch](mailto:hirzi.luqman@env.ethz.ch) and [daniel.wegmann@unifr.ch](mailto:daniel.wegmann@unifr.ch)

14

## 15 **Abstract**

16 Adaptive genetic variation is a function of both selective and neutral forces. In  
17 order to accurately identify adaptive loci, it is hence critical to account for  
18 demographic history. Theory suggests that signatures of selection can be inferred  
19 using the coalescent, following the premise that the genealogies of selected loci  
20 deviate from neutral expectations. Here, we build on this theory to develop an  
21 analytical framework to identify Loci under Selection via explicit Demographic  
22 models (LSD). Under this framework, signatures of selection are inferred by  
23 demographic parameters, rather than through isolated summary statistics, and  
24 demographic history is accounted for explicitly. Given that demographic models can  
25 incorporate directionality, we show that LSD can provide information on the  
26 environment in which selection acts on a population. This can prove useful in  
27 dissecting the genomics of local adaptation, by characterising genetic trade-offs and  
28 extending the concepts of antagonistic pleiotropy and conditional neutrality from  
29 ecological theory to practical application in genomic data. We implement LSD via  
30 Approximate Bayesian Computation and demonstrate, via simulations, that LSD has  
31 high power to identify selected loci across a large range of demographic-selection  
32 regimes, including complex demographies, and that the directionality of selection  
33 can be inferred accurately for identified candidates. Using the same simulations, we  
34 further characterise the behaviour of isolation-with-migration models conducive to  
35 the study of local adaptation under regimes of selection. Finally, we apply LSD to the

36 detection and characterisation of loci underlying floral guides in *Antirrhinum majus*,  
37 and find consistent results with previous studies.

38

### 39 **Keywords**

40 Approximate Bayesian computation, demography, genetic trade-offs, genome scan,  
41 local adaptation, selection

42

## 43 **1 INTRODUCTION**

44 Elucidating the genetic basis of adaptation and identifying genetic determinants  
45 of population and species divergence are key foci in evolutionary biology. In natural  
46 systems, genetic variation is shaped by the demographic history, driven by the  
47 neutral processes of mutation, migration and drift, together with natural selection on  
48 loci underlying adaptive traits. Conceptually, while all gene genealogies are  
49 constrained by the demographic history of the population, the genealogies of loci  
50 affected by selection are perturbed and may differ in key characteristics compared to  
51 those evolving under neutrality, though converging patterns can arise (Bierne,  
52 Welch, Loire, Bonhomme, & David, 2011; Edmonds, Lillie, & Cavalli-Sforza, 2004;  
53 Excoffier, Foll, & Petit, 2009; J. Li et al., 2012; Montgomery Slatkin & Excoffier, 2012).  
54 Disentangling the genomic signatures of these two processes, e.g. correctly  
55 identifying adaptive loci, remains a prevailing challenge in the field of population

56 genetics (Biswas & Akey, 2006; Horscroft, Ennis, Pengelly, Sluckin, & Collins, 2019;  
57 Luikart, England, Tallmon, Jordan, & Taberlet, 2003).

58

59 A multitude of methods have been developed that identify loci under selection as  
60 those whose summary statistics deviate from the genome-wide distribution. These  
61 “outlier” approaches can generally be grouped into three classes: those that 1)  
62 highlight regions of elevated differentiation between populations (via e.g.  $F_{ST}$ -related  
63 statistics), 2) highlight regions of perturbed site frequency spectrum (SFS) via  
64 diversity or diversity-related estimators (e.g.  $\pi$ , Tajima’s  $D$ ) and 3) highlight regions  
65 of extensive linkage disequilibrium (LD) via haplotype statistics (e.g. EHH, iHS)  
66 (Beaumont & Nichols, 1996; Biswas & Akey, 2006; Luikart et al., 2003; Oleksyk,  
67 Smith, & O’Brien, 2010; Vitti, Grossman, & Sabeti, 2013). While in empirical studies  
68 inference of selection is often achieved through corroborating evidence from multiple  
69 measures, each approach generally builds upon a single statistic that individually  
70 captures only a partial aspect of the effect of selection on the underlying genealogies.  
71 Additionally, reliance on statistics that describe the sample rather than on  
72 parameters that define the population imply that these approaches provide an  
73 incomplete description of the system. Combining the information from multiple  
74 statistics, either indirectly to inform underlying parameters via demographic  
75 modelling or directly via composite statistics, has the potential to provide insight and

76 increase power in the localisation of targets of selection (Grossman et al., 2010;  
77 Sugden et al., 2018; Vitti et al., 2013; Zeng, Shi, & Wu, 2007).

78

79 Under the premise that adaptive genetic variation is a function of both selective  
80 and neutral forces, accounting for the demographic history of the study system is  
81 critical for the correct identification of selected loci (François, Martins, Caye, &  
82 Schoville, 2016; Hoban et al., 2016). To achieve this, empirical studies generally  
83 employ a demographic null model which describes the neutral distribution of the  
84 statistics used to infer selection (e.g.  $F_{ST}$ ; Beaumont & Nichols, 1996; Eckert et al.,  
85 2010; Excoffier et al., 2009; Hofer, Ray, Wegmann, & Excoffier, 2009), or estimates of  
86 sample relatedness or covariance to correct for neutral population structure  
87 (Bonhomme et al., 2010; Engelhardt & Stephens, 2010; Gautier, 2015; Günther &  
88 Coop, 2013; Price et al., 2006). These approaches can be successful in controlling the  
89 confounding influence of demography, however they generally suffer from one or  
90 more of the following caveats; relying on post-hoc treatment (Eckert et al., 2010;  
91 Excoffier et al., 2009; Hofer et al., 2009), not providing direct estimates for underlying  
92 demographic parameters (sample covariance approaches) or assuming a set of rather  
93 simple demographic models (Foll & Gaggiotti 2008, Beaumont & Nichols 1996).  
94 Alternatively, a (complex) demographic model might be inferred from putatively  
95 neutral sites and loci under selection may be identified as those for which the an  
96 additional selection parameter is required (e.g. Williamson et al., 2005) or as those for

97 which locus-specific estimates of demographic parameters differ from genome-wide  
98 estimates (e.g. Sousa, Carneiro, Ferrand, & Hey, 2013). Such an approaches may not  
99 only account for demography explicitly and concurrently with the inference of  
100 selection, but may also directly estimate the probability of a locus being under  
101 selection if performed in a Bayesian framework, and hence avoid arbitrary  
102 thresholds.

103

104 Coalescent theory provides a powerful framework to infer the demographic  
105 processes that drive observed genetic variation (Kingman, 1982; Wakeley, 2001).  
106 While the coalescent was derived to model neutral variation, theory suggests that it  
107 can be exploited to infer signatures of selection following the premise that the  
108 genealogies of selected loci are expected to deviate from neutral expectations (Barton  
109 & Bengtsson, 1986; Charlesworth, 2009; Charlesworth, Nordborg, & Charlesworth,  
110 1997; Fusco & Uyenoyama, 2011; Galtier, Depaulis, & Barton, 2000; Gossmann,  
111 Woolfit, & Eyre-Walker, 2011; Petry, 1983; Sousa, Carneiro, Ferrand, & Hey, 2013).  
112 Under coalescent theory, demographic models are parametrised by effective  
113 population size(s) ( $N_E$ ) and in the case of multiple populations additionally by  
114 effective migration rate(s) ( $m_E$ ), which respectively describe the level of drift and  
115 gene flow within and between populations (Charlesworth, 2009; Petry, 1983).  
116 Importantly, both  $N_E$  and  $m_E$  may change through time. Different modes of selection  
117 and adaptive processes can be expected to alter these demographic parameters in

118 different ways. In a single population, imprints of selection may be reflected in  
119 variations in  $N_E$ , which is expected to be reduced in regions undergoing selective  
120 sweeps and increased in regions undergoing diversifying selection (Galtier et al.,  
121 2000; Gossmann et al., 2011). In the case of two or more populations connected by  
122 gene flow, divergent selection is expected to reduce  $m_E$  at selected and linked sites  
123 (Petry, 1983), while balancing selection and adaptive introgression may be expected  
124 to increase  $m_E$  at affected regions. Such theory implies that information can be  
125 acquired not just on the strength (via the magnitude of deviation of  $N_E$  or  $m_E$  from  
126 neutral expectations) and mode of selection (i.e. whether these deviations in  $N_E$  or  $m_E$   
127 are characterised by a reduction or elevation relative to neutral expectations), but  
128 also potentially of the population (environment) in which selection acts (i.e. whether  
129 the reduction in  $m_E$  is in a particular direction, in a multi-population model). This  
130 paradigm presents a unique opportunity to address the genetic basis of local  
131 adaptation, a key concept in ecological genetics.

132

133 The framework to describe the genetic basis of local adaptation derives from  
134 ecological theory that relates the fitness advantage of alternate alleles under different  
135 environmental conditions (Savolainen, Lascoux, & Merilä, 2013). Specifically,  
136 alternate alleles may confer higher fitness in their respective local environment but  
137 reduced fitness in the foreign environment, i.e. antagonistic pleiotropy (AP), or  
138 alleles may confer higher fitness in their local environment but have no differential

139 effect relative to the alternative allele in the foreign environment, i.e. conditional  
140 neutrality (CN) (Anderson, Lee, Rushworth, Colautti, & Mitchell-Olds, 2013;  
141 Kawecki & Ebert, 2004; Savolainen et al., 2013). Such genetic trade-offs are generally  
142 investigated via estimates of fitness in genetic crosses grown in reciprocal transplant  
143 experiments (Kawecki & Ebert, 2004; Savolainen et al., 2013). In such experiments, as  
144 well as in natural populations inhabiting different environments and connected by  
145 gene flow, directional selection acting on phenotypic traits modulates fitness and  
146 purges individuals that carry maladaptive alleles, e.g. via hybrid (Naisbit, Jiggins, &  
147 Mallet, 2001; Rundle & Whitlock, 2001; Schluter, 2000) or immigrant (Nosil, Vines, &  
148 Funk, 2005) inviability, or lower fecundity. This effectively reduces  $m_E$  at these  
149 (selected) loci between these populations, with the reduction (from neutral  
150 expectations) proportional to the strength of selection imposed on the alternative  
151 alleles in each environment. Such genetic trade-offs have long been assumed to  
152 underlie phenotypic trade-offs in populations connected by gene flow and the  
153 modelling of selection in terms of  $m_E$  via the coalescent provides a means to explicitly  
154 describe and disentangle such processes.

155

156 In this paper, we outline a conceptual and methodological framework for  
157 identifying Loci under Selection via explicit Demographic models called LSD, that  
158 scales to genomic data. Our approach explicitly accounts for demography in the  
159 identification of candidate loci, avoids reliance on singular summary statistics, and



160 elucidates the driving parameters underlying differentiation at putative selected loci.  
161 Furthermore, applied in a probabilistic Bayesian framework, our approach does not  
162 rely on arbitrary thresholds to delimit candidate loci, addressing an inherent  
163 limitation of many outlier approaches. While LSD is flexible regarding the choice of  
164 demographic model and can in principle accommodate any discrete population  
165 model (including single population and stepping-stone models) as well as detect  
166 different modes of selection, we here demonstrate LSD's utility in studies of local  
167 adaptation by focusing on the detection of loci under divergent selection under  
168 isolation-with-migration (IM) models. We validate and assess the performance of  
169 LSD via extensive simulations, and apply the method to the detection of functionally  
170 validated loci underlying floral guides in two parapatric subspecies of *Antirrhinum*  
171 *majus* (common snapdragon) (Schwinn et al., 2006; Tavares et al., 2018).

172

## 173 2 MATERIALS AND METHODS

### 174 2.1 Model

175 We begin by outlining the conceptual framework underlying LSD. Consider a  
176 demographic model  $\mathcal{M}$ , parametrised by demographic parameters  $\theta$ , that generates  
177 genetic data  $\mathbf{D}$ . To quantify deviations from neutrality, LSD first estimates the  
178 demographic parameters  $\hat{\theta}$  from a collection of loci assumed to be neutral (Figure  
179 S1). In a second step, LSD performs demographic inference on all loci and

180 determines the posterior distribution  $\pi_l = \pi(\boldsymbol{\theta}|\mathbf{D}_l)$  for each locus. Finally, LSD  
181 assesses the concordance of  $\hat{\boldsymbol{\theta}}$  with  $\pi_l$  by determining  $p_l$ , the highest posterior  
182 density interval (HPDI) of  $\pi_l$  that contains  $\hat{\boldsymbol{\theta}}$ . Here,  $q_l = 1 - p_l$  corresponds to the  
183 false-discovery-rate (FDR) of identifying locus  $l$  as incompatible with  $\hat{\boldsymbol{\theta}}$ . The joint  
184 posterior distribution  $\pi_l$  may further provide information on the magnitude and  
185 directionality of selection.

186

187         Given that the evaluation of the likelihood is non-trivial and may be  
188 intractable under more complex models, we resort to an approximate approach  
189 (Marjoram & Tavaré, 2006) (Figure 1). Under an Approximate Bayesian Computation  
190 (ABC) framework, the likelihood is approximated by simulations, the outcomes of  
191 which are compared with observed data in terms of summary statistics. That is, we  
192 find the set of parameters  $\boldsymbol{\theta}$  that minimise the distance between the observed data  $\mathbf{D}$   
193 and the simulated data  $\mathbf{D}'$ . To efficiently evaluate this, we reduce the dimensionality  
194 of the data via summarising them into a set of lower-dimensional summary statistics  
195  $\mathbf{S}$  and  $\mathbf{S}'$ , which are selected to capture the relevant information in  $\mathbf{D}$  and  $\mathbf{D}'$ ,  
196 respectively (Joyce & Marjoram, 2008; Peter, Huerta-Sanchez, & Nielsen, 2012).

197

198         An appropriate model for generating simulated genetic data is provided by  
199 coalescent theory (Kingman, 1982; Wakeley, 2001), parametrised by population  
200 demographic parameters  $\boldsymbol{\theta} = \{\mathbf{N}_E, \mathbf{M}_E, \mu\}$ , where  $\mathbf{N}_E$  refers to the vector of effective

201 population sizes,  $M_E$  to the vector of effective migration rates, and  $\mu$  to the mutation  
202 rate. We stress that population sizes and migration rates may vary through time. To  
203 minimise the distance  $D$  from  $D'$ , model space in addition to parameter space should  
204 be explored.

205

## 206 **2.2 Implementation**

207 The analytical framework described above is practically implemented as  
208 follows (Figure 1). Coalescent simulations are generated by a coalescent simulator,  
209 e.g. msms (Ewing & Hermisson, 2010), under a user-defined demographic model.  
210 Definition and choice of the demographic model should i) be informed by  
211 knowledge of the study system, ii) be motivated by the model's capacity to provide a  
212 useful approximation of a biological process of interest, and iii) be sufficiently simple  
213 to remain computationally tractable. Additionally, given that we condition the  
214 inference of selection on demographic parameters, the model should be formulated  
215 according to whether deviation in  $N_E$  or  $M_E$  is desired for the inference of selection.  
216 Importantly, the model should be validated by demonstrating that the observed data  
217 can be accurately and sufficiently captured (Figure S7).

218

219 The processing, format and final output of observed genetic data will often  
220 differ from that of raw coalescent simulations, given that observed genetic data may  
221 be subject to various pre-sequencing (e.g. pooling), sequencing (e.g. sequencing

222 errors, stochastic sampling of reads) and post-sequencing (e.g. filters) events that  
223 perturb and reformat the data from the original source. We thus implemented two  
224 complimentary programs that interface with coalescent simulators to replicate  
225 observed sequencing pipelines and generate simulated sequencing data: *LSD-High*  
226 can accommodate and simulate both individual and pooled data and assumes mid to  
227 high coverage (>10x) data, while *LSD-Low* accepts individual data and can  
228 additionally accommodate low coverage (>2x) data by utilising genotype likelihoods  
229 via msToGLF and ANGSD (Korneliussen, Albrechtsen, & Nielsen, 2014). A suite of  
230 summary statistics is then calculated for the simulated and observed data via the  
231 same programs. Summary statistics currently implemented include the number of  
232 segregating sites ( $S$ ), private  $S$ , nucleotide diversity ( $\pi$ ), Watterson's estimator ( $\theta_w$ ),  
233 Tajima's  $D$  ( $\theta$ ), relative divergence ( $F_{ST}$ ), absolute divergence ( $D_{XY}$ ), and site  
234 frequencies, though in principle any summary statistic can be included, contingent  
235 on the data and appropriate additions to the programs' scripts. To account for  
236 potential correlation between summary statistics and to retain only their informative  
237 components, we apply a Partial Least Squares transformation (Wegmann,  
238 Leuenberger, & Excoffier, 2009).

239

240 The estimation of demographic parameters is performed using ABCtoolbox  
241 (Wegmann, Leuenberger, Neuenschwander, & Excoffier, 2010), via the ABC-GLM  
242 algorithm using the subset of  $n$  simulations closest to the observed summary

243 statistics, separately for the putative neutral regions and for a sliding window across  
244 the genome. Here, windows may be interpreted as loci, and for modelling simplicity  
245 we assume that recombination is free between loci and fixed within. To acquire a set  
246 of putative neutral regions, we assume that sites belonging to a particular structural  
247 or functional class are selectively neutral (Williamson et al., 2005). We may for  
248 instance rely on genomic regions outside all structural annotations unlinked to all  
249 structural genomic elements (i.e. with a conservative flanking distance, informed by  
250 the linkage disequilibrium decay distance). Alternatively, a more naïve  
251 approximation may rely on the whole genome to provide genome-wide expectations,  
252 which may in some cases, but not always (Begun et al., 2007; Fay, Wyckoff, & Wu,  
253 2002; H. Li & Stephan, 2006), reflect the neutral case. In order to remain  
254 computationally tractable, departure from neutrality is evaluated only for that subset  
255 of demographic parameters informative of selection, while assuming the others to be  
256 shared among neutral and selected loci.

257

## 258 **2.3 Simulations**

259 **Demography** To test the performance of the LSD implementation, we  
260 simulated pseudo-observed genomes using the program *msms* under different  
261 demographic and selection parameter values (regimes), focusing on isolation-with-  
262 migration (IM) models relevant for the characterisation of local adaptation. We

263 simulated three models representing different levels of complexity in terms of  
264 population structure and evolutionary history (Figure 2): i) A simple 2-deme IM  
265 model (model  $\mathcal{M}_1$ ), with effective population sizes  $N_1 = N_2 = 10,000$  and symmetric  
266 migration rates  $M_{12} = M_{21} = M$ . ii) A 6-deme IM model comprising 2 contrasting  
267 environments of 3 demes each (model  $\mathcal{M}_2$ ), structured as 'islands' with effective  
268 population size  $N_i = 1,000$  connected via migration  $M_{ic} = 100$  to meta-population  
269 'continents' of size  $N_1 = N_2 = 100,000$ . These continents exchange migrants at  
270 symmetric rates  $M_{12} = M_{21} = M$ . iii) A 2-deme divergence with bottleneck and  
271 exponential growth model (model  $\mathcal{M}_3$ ). Under this model, two demes split from an  
272 ancestral population of size  $N_A = 10,000$  at  $T_D = 200,000$  generations ago. Following  
273 divergence, deme 2 stays constant at  $N_2 = 10,000$ , while deme 1 undergoes a sudden  
274 bottleneck, immediately followed by exponential growth with rate  $\alpha = 2$  until  
275  $T_G = 160,000$  generations ago, at which  $N_1 = 10,000$  is reached and thereafter  
276 remains constant. Demes 1 and 2 are initially separated with no gene flow between  
277 demes (isolation period), after which secondary contact is established at  $T_C = 20,000$   
278 generations ago with symmetric migration rates  $M_{12} = M_{21} = M$ . In all models, we  
279 used neutral migration rates  $M = 0.5, 5$  and  $50$  migrants per generation and inferred  
280 selection as deviations from these rates. We use model  $\mathcal{M}_1$  to represent a simplified,  
281 generalised model of local adaptation, model  $\mathcal{M}_2$  to represent a more complex case of  
282 local adaptation comprising multiple, structured populations and model  $\mathcal{M}_3$  to  
283 reflect a scenario typical of glacial-induced secondary contact population dynamics.

284

285           **Selection** Each simulated pseudo-genome comprised  $n_n = 1,000$  neutral loci

286 and  $n_s = 50$  selected loci of 5kb length, for a total (pseudo-genome) size of 5.25Mb.

287 We assumed a diploid system and all loci to be biallelic with ancestral allele  $a$  and

288 derived allele  $A$ . In order to generate a certain fraction of segregating sites per locus

289 ( $\sim 2\%$ ), the mutation  $\mu$  varied from  $2.5 \times 10^{-3}$  (model  $\mathcal{M}_1$ ) to  $2.5 \times 10^{-4}$  (model  $\mathcal{M}_3$ ) and

290  $5 \times 10^{-5}$  (model  $\mathcal{M}_2$ ) per locus per generation.

291

292           To simulate genetic trade-offs, selection was simulated on alternate alleles in

293 the contrasting environments. Specifically, we assumed the beneficial alleles to be

294 dominant such that the relative fitness was  $1 + s_1$ ,  $1, 1$  and  $1, 1, 1 + s_2$  for the three

295 genotypes  $AA$ ,  $Aa$  and  $aa$  in the demes or meta-populations occupying the two

296 environments, respectively. For the selection coefficients  $s_1$  and  $s_2$ , we used all

297 combinations of coefficient values  $0, 0.001, 0.01$  and  $0.1$  and thus included cases of

298 conditional neutrality (CN) in which either  $s_1 > 0, s_2 = 0$  or  $s_1 = 0, s_2 > 0$  as well as

299 cases of antagonistic pleiotropy (AP) with  $s_1 > 0, s_2 > 0$  with both symmetric ( $s_1 =$

300  $s_2$ ) and asymmetric ( $s_1 \neq s_2$ ) regimes. CN regimes are by definition always

301 asymmetric and AP regimes are defined such that both alleles confer higher fitness in

302 their respective local environments but reduced fitness in the other, both with respect

303 to the alternate allele and to the fitness conferred in their home environments. We

304 further varied the time of the onset of selection from  $T_S = 400, 4,000, 40,000$  and  
305 400,000 generations ago.

306

307 For all models, we considered selection on standing variation with the initial  
308 frequency of the derived allele at  $f_1 = f_2 = 0.1$  in all demes. For model  $\mathcal{M}_1$ , we  
309 additionally investigated the case of *de-novo* mutations with initial frequencies  
310  $f_1 = 1/2N_1$  and  $f_2 = 0$ . These two cases represent the often-considered starting points  
311 for local adaptation (Peter et al., 2012). Dependent on the selection regime and due to  
312 the stochasticity of drift, the derived allele  $A$  may sometimes be lost and hence be  
313 absent in the simulation of selected loci (especially in the *de-novo* case). Because such  
314 a scenario contains no signal for detection of selection, we excluded such simulations  
315 (via the -SFC parameter in msms).

316

317 **Assessing accuracy** We inferred selection by contrasting the locus-specific  
318 migration rates  $m_{12}$  and  $m_{21}$  against their neutral estimates  $\hat{m}_{12}$  and  $\hat{m}_{21}$  (Figure 3).  
319 We evaluated the performance of our LSD implementation at identifying selected  
320 loci under these simulations by plotting the true positive rate (TPR) against the false  
321 positive rate (FPR) under the choice of HDPI thresholds from 0 to 1, and reporting  
322 the area under the curve (AUC) of the resultant receiver operating characteristic  
323 (ROC) curve. To evaluate the accuracy of the inferred symmetry of the joint posterior  
324 (of reciprocal migration rates), we compared this to the true underlying selection



325 coefficients, under the expectation that deviations from symmetry in the joint  
326 posterior should reflect asymmetry in selection regimes. Specifically, we determined  
327 for each locus  $l$  the posterior mass

$$\sigma_l = \int I\left(\frac{m_{21}}{m_{12}} < \frac{\hat{m}_{21}}{\hat{m}_{12}}\right) \pi_l d\theta,$$

328 where the indicator function  $I(\cdot)$  limits the integral to cases in which the deviation of  
329 one of the migration rates has reduced more than the reciprocal migration rate  
330 compared to a proportional deviation of both migration rates from their neutral  
331 estimates  $\hat{m}_{12}$  and  $\hat{m}_{21}$ . From this, we calculate the asymmetry as

$$a = \log \frac{\sigma}{1 - \sigma},$$

332 where  $\sigma = \frac{1}{n_s} \sum \sigma_l$  across loci simulated under selection.

333

## 334 **2.4 Case study**

335 To evaluate the performance of LSD on real data, we applied it to the detection of  
336 loci underlying floral guides in two parapatric subspecies of *Antirrhinum majus*.  
337 *A. majus* is an herbaceous, perennial, flowering plant native to the western  
338 Mediterranean. Owing to its diploid inheritance, relatively short generation time,  
339 ability for both self- and cross-pollination and rich and varied flower morphology,  
340 *A. majus* has lent itself as a model organism for over a century, with several key floral  
341 genes being first identified within this genus (Schwarz-Sommer, Davies, & Hudson,  
342 2003; Schwinn et al., 2006). Two subspecies, *A. m. striatum* and *A. m. pseudomajus*, differ

343 in the flower colouration that signposts the pollinator entry point, and form a natural  
344 hybrid zone in the Pyrenees that constitutes a benchmark example of divergent  
345 selection driven by assortative mating (Whibley et al., 2006). Several genetic loci have  
346 been shown to control the differences in these floral patterns (Bradley et al., 2017;  
347 Schwinn et al., 2006), and recently, Tavares et al. (2018) produced evidence of  
348 genomic signatures of selection at the ROS and EL loci, of which the former has been  
349 further functionally validated. Here, we apply LSD to sequencing data from this  
350 study to isolate the ROS and EL loci and to characterise their underlying selection  
351 signal.

352

353 We modelled this study system via a simple representation (model  $\mathcal{M}_1$ ) of one  
354 population on either side of the hybrid zone (YP1 (*A.m.striatum*) vs MP2  
355 (*A.m.pseudomajus*); populations 2.5km apart) and via a more inclusive island-  
356 continent model (model  $\mathcal{M}_2$ ) comprising three (distant) populations each per  
357 subspecies (CAM, ML, YP1 (*A.m.striatum*) vs MP2, CHI, CIN (*A.m.pseudomajus*);  
358 Figure S5), allowing all  $N_E$  and  $m_E$  parameters to be free. Given that these populations  
359 were previously sequenced using pool-seq (Tavares et al., 2018), we simulated  
360 pooling of individuals in-silico by pooling twice the amount of msms coalescent  
361 (haploid) samples as (diploid) individuals in the pooled populations via *LSD-High*.  
362 Samples were drawn from a parametric (negative binomial) distribution fitted to the  
363 empirical coverage distribution using the '*fitdistrplus*' package (Delignette-Muller &

364 Dutang, 2015) and *LSD-High*. We focused our analysis on chromosome 6 on which  
365 the ROS and EL loci lie. To acquire empirical estimates of neutral demographic  
366 parameters, we excluded all genomic regions present in the structural annotation  
367 plus 10kb flanking regions to generate a subset of putatively neutral regions on that  
368 chromosome. The inference of selection was then performed in sliding windows of  
369 size 10kb with a 1kb step-size. The mutation rate for the simulations ( $1.7 \times 10^{-8}$  per site  
370 per generation) and the filtering of the empirical and simulated data followed those  
371 reported in the original study, though we mapped on a more recent and complete  
372 version of the *Amajus* reference (version 3.0; M. Li et al., 2019).

373

### 374 **3 RESULTS**

#### 375 **3.1 Two-deme IM case (model $\mathcal{M}_1$ )**

376 **Power to identify selected loci** Our LSD implementation demonstrated a high  
377 diagnostic ability to discriminate between neutral and selected loci ( $AUC \gg 0.9$ )  
378 across a large range of migration-selection regimes (Figure 4). Notably, our results  
379 point towards an optimal, intermediate rate of migration ( $M=5$ ) at which selection is  
380 best detectable with high AUC values across a large set of selection coefficients. As  
381 migration rates increase ( $M=50$ ), migration from the foreign deme where selection  
382 acts on the alternate allele increasingly inhibits the build-up of beneficial  
383 polymorphisms in the local deme, in which case the power to detect selected loci

384 becomes limited to scenarios under longer regimes of strong selection. At lower  
385 migration rates ( $M=0.5$ ), long regimes of selection permit the detection of loci under  
386 the lowest selection coefficients, but power decreases for younger times compared to  
387 scenarios simulated under intermediate migration rates. This owes to LSD relying on  
388 the reduction of effective migration relative to neutral or genome-wide expectations,  
389 which in this case is already at a low level.

390

391       The power to detect selection increased with increasing selection coefficients  
392 in the case of AP if selection coefficients were similar ( $s_1 \approx s_2$ , cells along diagonal of  
393 sub-panels in Figure 4). In such cases, stronger selection coefficients on alternate  
394 alleles increasingly polarise and ultimately maintain larger allele frequency  
395 differences between the two environments. In tandem, the power to detect selection  
396 also generally increased with the time since the onset of selection  $T_S$ . However, in  
397 most cases of CN or when  $s_1 \gg s_2$  or  $s_1 \ll s_2$ , one of the two alleles may proceed to  
398 fixation, in which case the power to detect selection decays or is lost (e.g. cells along  
399 bottom row and left-most column of sub-panels in Figure 4). This is particularly  
400 evident when the onset of selection is more distant in the past and implies that cases  
401 of CN may be harder to detect than AP as their signatures of selection are often more  
402 transient and decay more rapidly.

403

404 To evaluate the false discovery rate (FDR) of LSD at different migration rates,  
405 we also conducted simulations in the absence of selection. For intermediate  
406 migration rates ( $M=5$ ), LSD performed better than FDR expectations (Figure S2B); at  
407 a defined threshold of 0.95, LSD resulted in a false positive rate of 2%. For the low  
408 and high migration rates ( $M=0.5, 50$ ) however, LSD performed worse than FDR  
409 expectations, producing false positive rates higher than expected with false positive  
410 rates of 44% and 8% respectively at a threshold of 0.95 (Figures S2A and S2C). This  
411 optimal, intermediate rate of migration for minimising FDR is consistent with the  
412 optimal migration rate for detecting selection.

413

414 **Power to characterise (a)symmetry** A benefit of LSD over classic outlier  
415 approaches is that it can provide insight into trade-offs underlying selection, by  
416 identifying cases in which selection acts at equal strength in the two demes or  
417 metapopulations (symmetric AP), or whether selection coefficients differ  
418 considerably (CN or asymmetric AP). As shown in Figure 5, the inferred  
419 (a)symmetry generally reflected the true (a)symmetry of the underlying selection  
420 coefficients well, particularly for regimes with high power to correctly identify  
421 selected loci ( $AUC > 0.95$ ). In lower powered regimes, we observe a few cases where  
422 the inferred asymmetry does not reflect the underlying asymmetry of the selection  
423 coefficients accurately (e.g. blue cells along diagonals in sub-panels at  $T_S = 4,000$ ;  
424 Figures 5 and S3). We treat these exceptional cases further in the discussion.

425

426           **Standing variation vs *de-novo*** A lower initial frequency of the derived allele  
427 may be expected to affect LSD's power to identify selected loci and its power to  
428 capture the underlying (a)symmetry of selection coefficients. However, we find that  
429 results for simulations building on selection from the *de-novo* and standing variation  
430 cases showed generally very similar patterns (Figures 4, 5 and S3). One notable  
431 exception however was the inaccurate inference of (a)symmetry in a few regimes  
432 with high power ( $AUC > 0.95$ ) in the *de-novo* case (e.g. blue cells along diagonals in  
433 sub-panels at  $T_S = 4,000$  in Figure S3B). This we attribute to the lower initial  
434 frequency of the derived allele  $A$  and consequently longer time needed to reach drift-  
435 migration-selection equilibrium for the *de-novo* cases. This is explored further in the  
436 discussion.

437

### 438 **3.2 More complex cases (models $\mathcal{M}_2$ and $\mathcal{M}_3$ )**

439           A key feature of LSD is its potential for explicit accommodation of complex  
440 demographies, which when not properly accounted for can lead to an inflation in  
441 false positives (De Villemereuil, Frichot, Bazin, François, & Gaggiotti, 2014; Foll &  
442 Gaggiotti, 2008; Lotterhos & Whitlock, 2014). Despite the added complexity of  
443 models  $\mathcal{M}_2$  and  $\mathcal{M}_3$ , results were generally very similar to that of model  $\mathcal{M}_1$ , with  
444 high power to identify selected loci ( $AUC \gg 0.9$ ) across a large range of migration-  
445 selection regimes, an optimal migration rate at an intermediate value ( $M=5$ ), a similar

446 dependence of power to detect selection on  $s_1$ ,  $s_2$  and  $T_{S_i}$  and inferences of  
447 (a)symmetry that reflected well the underlying (a)symmetry of selection coefficients  
448 (Figures 6 and S4). A notable difference compared to model  $\mathcal{M}_1$  was the longer  
449 amount of time needed to generate a high power to detect selection across a range of  
450 migration values in models  $\mathcal{M}_2$  (Figures 6A) and especially  $\mathcal{M}_3$  (Figure S4A), which  
451 we attribute to the lower mutation rates used in these models compared to  $\mathcal{M}_1$ .

452

### 453 **3.3 Case study results**

454 We identified a region of reduced effective migration between 52.9-53.2MB on  
455 chromosome 6 (Figure 7), consistent with the location of the ROS and EL loci  
456 (Tavares et al., 2018). Under model  $\mathcal{M}_1$ , this region is characterised by a set of  
457 smaller, multiple peaks (posterior probability of being divergent from neutral  
458 expectations  $> 99.9\%$ ) reflecting signatures identified by previous authors, with the  
459 left-most peaks (shaded in red) corresponding to ROS1 and ROS2 and the right peaks  
460 (shaded in green) to EL (Figure 7B). The joint posterior probability distributions  
461 reveal symmetric selection acting on both regions, implying that selection acts with  
462 similar strength in the two populations. Under Model  $\mathcal{M}_2$ , we find fewer outliers in  
463 the ROS-EL region than in model  $\mathcal{M}_1$ , with the left-most peak in this region  
464 corresponding to ROS2 and the right peaks consistent with EL. ROS1 appears to be  
465 less of an outlier than in model  $\mathcal{M}_1$  (posterior probability of being divergent  $\sim 99.8\%$ ).

466 In contrast to model  $\mathcal{M}_1$ , the ROS2 and EL peaks in model  $\mathcal{M}_2$  are characterised by  
467 asymmetry, specifically with stronger selection acting in the populations of  
468 *A.m.pseudomajus* than in the populations of *A.m.striatum*.

469

## 470 **4 DISCUSSION**

471 The trajectory of selected loci depends on demographic and selection  
472 parameters that define the system, namely the effective population sizes, effective  
473 migration rates and selection coefficients, as well as the intrinsic properties of  
474 mutation and recombination. Despite well-developed theory which relates the effect  
475 of population parameters on the trajectory of selected alleles, few methods or  
476 empirical studies have combined estimates of differential selection with explicit  
477 quantification of migration rates and effective population sizes to examine the  
478 conditions under which local adaptation can arise. In this study, we condition the  
479 identification of candidate loci on divergent population parameters using explicit  
480 demographic models, and demonstrate that under certain demographic-selection  
481 regimes, we can both detect and elucidate the processes underlying signatures of  
482 selection. While LSD is flexible regarding the choice of demographic models  
483 employed, we focus here specifically on those processes that are expected to lead to  
484 selection against gene flow, namely local adaptation and extrinsic reproductive  
485 barriers, that can be inferred via their expectation to reduce effective migration rates.

486



## 487 **4.1 Identifying selection**

488           In our simulations, we demonstrate that LSD has high diagnostic power (AUC  
489 >> 0.9) to identify selected loci across a large range of demographic-selection regimes.  
490 This power relies upon two fundamental aspects that contribute to generating  
491 observable patterns. First, selection must effectively be realised, i.e. result in a  
492 frequency shift of the beneficial allele. This requires that the strength of selection and  
493 initial frequency of the beneficial allele be sufficient to both counter the  
494 homogenising effect of migration (Felsenstein, 1976; Haldane, 1930; Lenormand,  
495 2002; M. Slatkin, 1973; Yeaman, 2015) and the eroding effect of drift (Wright, 1931).  
496 Secondly, the genomic data must contain signatures of selection that can be detected.  
497 In the case of LSD, this requires that the signatures of selection are discernible from  
498 the underlying noise (drift and migration) that characterises the system, as well as  
499 requires sufficient time for said signatures to be reflected in the employed statistics  
500 and hence in the inferred parameters  $N_E$  or  $mE$ . A lack of power in LSD must be  
501 interpreted considering these two conceptually different perspectives. Notably, the  
502 lack of discrimination power for high migration rates and low selection coefficients  
503 can be attributed to selection failing to realise as a consequence of local, beneficial  
504 alleles being swamped by immigrant, maladaptive alleles. In contrast, the lack of  
505 signal under low migration rates constitutes a methodological limitation of our  
506 implemented model, as it becomes increasingly difficult to detect reductions in  
507 effective migration when neutral or genome-wide migration rates are already at a

508 low level; even when selection is effectively being realised in the demes. This is  
509 analogous in effect to the loss of power to detect selection in highly differentiated  
510 populations in  $F_{ST}$  outlier tests (Hoban et al., 2016). Under the same principle, we  
511 argue that the converse expectation can be assumed to hold for loci underlying  
512 adaptive introgression or balancing selection. That is, we expect power to detect such  
513 loci to be low when populations are minimally differentiated, as the detection of  
514 candidate loci in these cases is informed by increased effective migration.

515

516       The power of LSD to correctly identify selected loci generally increased with  
517 stronger selection coefficients and longer time since the onset of selection, though  
518 with exceptions related to differential selection on alternate alleles in multiple  
519 populations. Specifically, when selection is of similar or equal strength in both demes  
520 or meta-populations, we observed a strong correlation between the power to detect  
521 selection and the true underlying selection coefficients. This follows theory which  
522 states that the reduction in effective migration is proportional to the strength of  
523 selection (Petry, 1983). However, we defer from translating these changes to explicit  
524 selection coefficients because in addition to the strength of selection, changes in  
525 effective migration are also a function of the recombination rate between linked and  
526 selected loci (Cutter & Payseur, 2013; Lotterhos, 2019; Petry, 1983). On the other  
527 hand, if selection is highly divergent in strength ( $s_i \gg s_j$ ) between the demes or meta-  
528 populations or when the onset of selection is sufficiently distant in the past, one of

529 the two selected alleles may have fixed in the system. In such a case, the signal to  
530 detect selection rapidly decays (Huber, DeGiorgio, Hellmann, & Nielsen, 2016;  
531 Przeworski, 2002). Finally, we observed little power to detect very recent selection,  
532 intrinsically related to our choice of summary statistics (Hohenlohe, Phillips, &  
533 Cresko, 2010). While a signal of selection necessarily requires time to build up, we  
534 note that extending LSD to include additional statistics sensitive to linkage  
535 disequilibrium such as extended haplotype homozygosity (EEH) (Szpiech &  
536 Hernandez, 2014) or single density score (SDS) (Field et al., 2016) may increase the  
537 power to detect more recent selection.

538

539 From our simulations, we find that the effect of the tested selection regimes on  
540 the power to detect selection is similar between the *de-novo* and standing genetic  
541 variation cases (model  $\mathcal{M}_1$ ; Figures 4 and S3A). This result relies on the fact that we  
542 only kept simulations of selected loci (comprising the pseudo-genomes) in which  
543 derived allele *A* was not lost. This particularly affected the *de-novo* case, where most  
544 simulations of selected loci were observed to result in the loss of allele *A*. This  
545 implies firstly, that in most observed cases, signals of selection are most likely to  
546 arise from standing variation (Jones et al., 2012; Lai et al., 2019; Reid et al., 2016), and  
547 secondly, that to distinguish selection arising *de-novo* to that from standing variation,  
548 additional information such as allele age, mutation rate or supplementary  
549 phylogenetic information is likely required (Peter et al., 2012).

550

## 551 **4.2 Revealing trade-offs underlying selection**

552         A major benefit of genome scans performed under a demographic framework  
553 is the capacity to infer the directionality of selection. In our simulations, we observe  
554 that the (a)symmetry in the reduction of reciprocal migration rates between demes or  
555 meta-populations as inferred by LSD reflects the (a)symmetry of underlying selection  
556 coefficients accurately for older onsets of selection ( $T_S > 4,000$ , red cells inhabiting  
557 left-upper triangle, blue in right-lower triangle; Figures 5, 6B, S3B, S4B), but less so  
558 for more recent onsets of selection ( $T_S \leq 4,000$ ; Figures 5, 6B, S3B, S4B). This is  
559 because prior to reaching drift-migration-selection equilibrium, estimated  
560 asymmetries in effective migration rates are also affected by asymmetry in allele  
561 frequencies (Figure 8). This is highlighted when contrasting the standing variation  
562 and *de-novo* simulation results for model  $\mathcal{M}_1$ , where incorrectly inferred  
563 (a)symmetries are more evident in the *de-novo* case due to the lower initial frequency  
564 of the derived allele ( $T_S \leq 4,000$ ; Figures 5 and S3B). A direct link between the  
565 (a)symmetry in inferred migration rates and selection coefficients is only established  
566 through time as the beneficial allele increases in frequency towards an equilibrium.  
567 From this, we deduce that if a strong asymmetry between effective migration rates is  
568 inferred, the system may still be in the process of evolving towards an equilibrium  
569 state, which may include cases in which one allele will ultimately be lost. If  
570 symmetry between effective migration rates is inferred on the other hand, the system

571 is likely near-equilibrium and we can expect both alleles to be maintained in the  
572 system. We note that in practice, however, the interpretation of the results is  
573 straightforward. This is because the inference of directionality is only relevant if the  
574 targets of selection can be detected accurately (e.g. in regimes with  $AUC > 0.9$ ). For  
575 these cases, we generally find the inferred (a)symmetries in migration rates to reflect  
576 the true (a)symmetry in selection coefficients (Figure 5, 6B, S3B, S4B) accurately,  
577 mostly because the power to detect selection is generally low if selection started  
578 acting only very recently.

579

580       The ability of LSD to infer the directionality of selection directly from genomic  
581 data can greatly facilitate investigations of genetic trade-offs underlying adaptation,  
582 which are seldom performed due to the considerable effort required to set up field  
583 trials of recombinant lines. As shown above, the inference of symmetry in LSD-  
584 identified candidates accurately reflects cases of AP with equal strength of selection  
585 on alternate alleles in the contrasting environments. The inference of asymmetry on  
586 the other hand can either indicate AP with stronger selection in one environment  
587 than the other, or CN. From our simulations, we find that scenarios reflecting AP are  
588 generally more readily detected than those reflecting CN. Given that selection acts  
589 only upon one of the two alleles in the latter case, fixation becomes likely and the  
590 ability to detect selection is transient. This implies that there may be an observation  
591 bias between AP and CN; such that the inference of CN may be comparatively

592 under-represented. This bias appears to contrast with that reported in ecological  
593 literature, where instances of AP are more rarely detected compared to CN due to  
594 the additional power required to detect variance in fitness concurrently in two  
595 environments (Anderson et al., 2013). LSD may further complement field trials as  
596 such experiments typically test genetic trade-offs under contemporary selective  
597 environments, which may not reflect past conditions driving the observed adaptive  
598 responses, but whose signature may still be inferred from genomic data. Using LSD  
599 to formulate expectations about fitness effects and to inform the choice of  
600 environmental conditions under which to validate identified candidate genes can  
601 thus greatly aid such experiments.

602

### 603 **4.3 Real-world application**

604 We demonstrate a real-world application of LSD by successfully isolating and  
605 characterising the selection signal of loci underlying an extrinsic reproductive barrier  
606 in *A. majus*. Our results from contrasting a single population (model  $\mathcal{M}_1$ ) and three  
607 populations (model  $\mathcal{M}_2$ ) per subspecies both identified the ROS and EL loci which  
608 were previously reported to underlie differences in floral patterns between these  
609 subspecies (Tavares et al., 2018). Interestingly however, our results show a different  
610 signal of (a)symmetry between the tested models  $\mathcal{M}_1$  and  $\mathcal{M}_2$ , with the peaks  
611 corresponding to the ROS and EL loci characterised by symmetry under model  $\mathcal{M}_1$ ,  
612 in contrast to a pattern of asymmetry under model  $\mathcal{M}_2$ , specifically with stronger

613 selection inferred to act in the populations of *A.m.pseudomajus* than in the  
614 populations of *A.m.striatum*. We stress that the interpretation of LSD genome scans is  
615 conditional on the model and populations used, such that in our example model  $\mathcal{M}_1$   
616 uncovers population-pair specific differences at the contact zone (YP1 vs MP2) while  
617 model  $\mathcal{M}_2$  reveals common (global) differences between the two subspecies. We do  
618 not necessarily expect these two signals to be identical, and indeed, Tavares et al.  
619 (2018) also found differences between distant and close *A.m.striatum*-*A.m.pseudomajus*  
620 population pairs in terms of observed  $\theta_w$  and  $F_{ST}$  summary statistics. Given that there  
621 is no evident difference in environment or pollinators on opposite sides of the hybrid  
622 zone, reproductive barriers in this system have often been proposed to be maintained  
623 through assortative mating mediated by pollinator preference for the dominant  
624 (most common) flower phenotype in a given area and the subspecies' distinct flower  
625 colouration. However, whether selection on alternate alleles follows the same  
626 positive frequency-dependence across the broader scale including more distant  
627 populations is currently unknown. The difference in signal between local pairs at the  
628 contact zone ( $\mathcal{M}_1$ ) and the global set ( $\mathcal{M}_2$ ) may be generated by different frequency-  
629 dependent selection curves for the alternate alleles and potentially loss of AP away  
630 from the contact zone (Figure S6).

631

632 **5 CONCLUSION**

633           When selection occurs in the presence of gene flow, selected sites are  
634 predicted to exhibit gene genealogies with demographic parameters divergent from  
635 those of neutral non-linked sites, leading to heterogeneity in demography across the  
636 genome. In this study, we condition the identification of candidate loci on divergent  
637 population parameters using explicit demographic models, and demonstrate that  
638 under certain conditions of migration, selection strength and onset time, we can both  
639 detect and elucidate the underlying processes driving signatures of selection.  
640 Incorporating and utilising the inference of demographic parameters in the  
641 identification of candidate loci address some key issues and assumptions that prevail  
642 in the discrimination of selected variants, namely 1) the explicit consideration of  
643 demography, 2) heterogeneity in drift and gene flow across the genome, 3)  
644 information synthesis of multiple, complementary summary statistics, and 4)  
645 transparency towards underlying driving mechanisms.

646

647           Our power analysis using simulations shows that LSD, and our  
648 implementation of it, represents a powerful method for detecting selection that is  
649 robust to different and complex demographies. Furthermore, given that certain  
650 demographic parameters e.g. migration are not inherently commutative, we show  
651 that the directionality or population-specificity in selection can be inferred. This can  
652 facilitate identifying in which environment selection acts and hence elucidate genetic  
653 trade-offs; bridging an analytical divide between experimental ecology and



654 population genomics. Importantly, the proposed approach as well as our  
655 implementation is not limited to the demographic models investigated here, nor the  
656 explicit choice of simulation programs or summary statistics used. This flexibility  
657 and customisability of LSD can facilitate e.g. more realistic accommodation of  
658 recombination (via different coalescent simulators), improved detection of more  
659 recent selection (via linkage-informative statistics), and inference of other modes of  
660 selection (e.g. balancing selection) and adaptive introgression by conditioning the  
661 detection of selection on e.g. increase (rather than reduction) of  $m_E$  or changes in  $N_E$   
662 relative to neutral expectations.

663

## 664 **ACKNOWLEDGEMENTS**

665 We would like to thank Niklaus Zemp for providing IT support. This work  
666 was supported by the Swiss National Science Foundation (SNSF) grants  
667 31003A\_160123 and 31003A\_182675 awarded to AW and grant 31003A\_173062  
668 awarded to DW.

669

670 **REFERENCES**

671

672 Anderson, J. T., Lee, C. R., Rushworth, C., Colautti, R., & Mitchell-Olds, T. (2013).

673 Genetic tradeoffs and conditional neutrality contribute to local adaptation.

674 *Molecular Ecology*, 22(3), 699–708. <https://doi.org/10.1038/jid.2014.371>

675 Barton, N., & Bengtsson, B. O. (1986). The barrier to genetic exchange between

676 hybridising populations. *Heredity*, 57(3), 357–376.

677 <https://doi.org/10.1038/hdy.1986.135>

678 Beaumont, M. A., & Nichols, R. A. (1996). Evaluating loci for use in the genetic

679 analysis of population structure. *Proceedings of the Royal Society B: Biological*

680 *Sciences*, 263(1377), 1619–1626. <https://doi.org/10.1098/rspb.1996.0237>

681 Begun, D. J., Holloway, A. K., Stevens, K., Hillier, L. W., Poh, Y. P., Hahn, M. W., ...

682 Langley, C. H. (2007). Population genomics: Whole-genome analysis of

683 polymorphism and divergence in *Drosophila simulans*. *PLoS Biology*, 5(11),

684 2534–2559. <https://doi.org/10.1371/journal.pbio.0050310>

685 Bierne, N., Welch, J., Loire, E., Bonhomme, F., & David, P. (2011). The coupling

686 hypothesis: Why genome scans may fail to map local adaptation genes.

687 *Molecular Ecology*, 20(10), 2044–2072. <https://doi.org/10.1111/j.1365->

688 294X.2011.05080.x

689 Biswas, S., & Akey, J. M. (2006). Genomic insights into positive selection. *Trends in*

690 *Genetics*, 22(8), 437–446. <https://doi.org/10.1016/j.tig.2006.06.005>

691 Bonhomme, M., Chevalet, C., Servin, B., Boitard, S., Abdallah, J., Blott, S., &

692 SanCristobal, M. (2010). Detecting selection in population trees: The Lewontin

693 and Krakauer test extended. *Genetics*, 186(1), 241–262.

694 <https://doi.org/10.1534/genetics.110.117275>

695 Bradley, D., Xu, P., Mohorianu, I. I., Whibley, A., Field, D., Tavares, H., ... Coen, E.

696 (2017). Evolution of flower color pattern through selection on regulatory small

697 RNAs. *Science*, 358(6365), 925–928. <https://doi.org/10.1126/science.aao3526>

698 Charlesworth, B. (2009). Fundamental concepts in genetics: Effective population size

699 and patterns of molecular evolution and variation. *Nature Reviews Genetics*, 10(3),

700 195–205. <https://doi.org/10.1038/nrg2526>

701 Charlesworth, B., Nordborg, M., & Charlesworth, D. (1997). The effects of

702 background and interference selection on patterns of genetic variation in

703 subdivided populations. *Genetics Research*, 70, 155–174.

704 <https://doi.org/10.1534/genetics.115.178558>

705 Cutter, A. D., & Payseur, B. A. (2013). Genomic signatures of selection at linked sites:

706 Unifying the disparity among species. *Nature Reviews Genetics*, 14(4), 262–274.

707 <https://doi.org/10.1038/nrg3425>

708 De Villemereuil, P., Frichot, É., Bazin, É., François, O., & Gaggiotti, O. E. (2014).

709 Genome scan methods against more complex models: When and how much

710 should we trust them? *Molecular Ecology*, 23(8), 2006–2019.

- 711 <https://doi.org/10.1111/mec.12705>
- 712 Delignette-Muller, M. L., & Dutang, C. (2015). fitdistrplus: An R package for fitting  
713 distributions. *Journal of Statistical Software*, 64(4), 1–34.  
714 <https://doi.org/10.18637/jss.v064.i04>
- 715 Eckert, A. J., Van Heerwaarden, J., Wegrzyn, J. L., Nelson, C. D., Ross-Ibarra, J.,  
716 González-Martínez, S. C., & Neale, D. B. (2010). Patterns of population structure  
717 and environmental associations to aridity across the range of loblolly pine (*Pinus*  
718 *taeda L.*, Pinaceae). *Genetics*, 185(3), 969–982.  
719 <https://doi.org/10.1534/genetics.110.115543>
- 720 Edmonds, C. a, Lillie, A. S., & Cavalli-Sforza, L. L. (2004). Mutations arising in the  
721 wave front of an expanding population. *Proceedings of the National Academy of*  
722 *Sciences of the United States of America*, 101(4), 975–979.  
723 <https://doi.org/10.1073/pnas.0308064100>
- 724 Engelhardt, B. E., & Stephens, M. (2010). Analysis of population structure: A unifying  
725 framework and novel methods based on sparse factor analysis. *PLoS Genetics*,  
726 6(9). <https://doi.org/10.1371/journal.pgen.1001117>
- 727 Ewing, G., & Hermisson, J. (2010). MSMS: A coalescent simulation program  
728 including recombination, demographic structure and selection at a single locus.  
729 *Bioinformatics*, 26(16), 2064–2065. <https://doi.org/10.1093/bioinformatics/btq322>
- 730 Excoffier, L., Foll, M., & Petit, R. J. (2009). Genetic Consequences of Range  
731 Expansions. *Annual Review of Ecology, Evolution, and Systematics*, 40(1), 481–501.  
732 <https://doi.org/10.1146/annurev.ecolsys.39.110707.173414>
- 733 Fay, J. C., Wyckoff, G. J., & Wu, C. I. (2002). Testing the neutral theory of molecular  
734 evolution with genomic data from *Drosophila*. *Nature*, 415(6875), 1024–1026.  
735 <https://doi.org/10.1038/4151024a>
- 736 Felsenstein, J. (1976). The theoretical population genetics of variable selection and  
737 migration. *Annu. Rev. Genet*, 10, 253–280.
- 738 Field, Y., Boyle, E. A., Telis, N., Gao, Z., Gaulton, K. J., Yengo, L., ... Pritchard, J. K.  
739 (2016). Detection of human adaptation during the past 2000 years, 354(6313),  
740 760–764. <https://doi.org/10.1126/science.aag0776>.Detection
- 741 Foll, M., & Gaggiotti, O. (2008). A genome-scan method to identify selected loci  
742 appropriate for both dominant and codominant markers: A Bayesian  
743 perspective. *Genetics*, 180(2), 977–993. <https://doi.org/10.1534/genetics.108.092221>
- 744 François, O., Martins, H., Caye, K., & Schoville, S. D. (2016). Controlling false  
745 discoveries in genome scans for selection. *Molecular Ecology*, 25(2), 454–469.  
746 <https://doi.org/10.1111/mec.13513>
- 747 Fusco, D., & Uyenoyama, M. K. (2011). Sex-specific incompatibility generates locus-  
748 specific rates of introgression between species. *Genetics*, 189(1), 267–288.  
749 <https://doi.org/10.1534/genetics.111.130732>
- 750 Galtier, N., Depaulis, F., & Barton, N. H. (2000). Detecting bottlenecks and selective  
751 sweeps from DNA sequence polymorphism. *Genetics*, 155(2), 981–987.

- 752 Gautier, M. (2015). Genome-wide scan for adaptive divergence and association with  
753 population-specific covariates. *Genetics*, 201(4), 1555–1579.  
754 <https://doi.org/10.1534/genetics.115.181453>
- 755 Gossmann, T. I., Woolfit, M., & Eyre-Walker, A. (2011). Quantifying the variation in  
756 the effective population size within a genome. *Genetics*, 189(4), 1389–1402.  
757 <https://doi.org/10.1534/genetics.111.132654>
- 758 Grossman, S. R., Shylakhter, I., Karlsson, E. K., Byrne, E. H., Morales, S., Frieden, G.,  
759 ... Sabeti, P. C. (2010). A Composite of Multiple Signals Distinguishes Causal  
760 Variants in Regions of Positive Selection. *Science*, 327, 883–886.
- 761 Günther, T., & Coop, G. (2013). Robust identification of local adaptation from allele  
762 frequencies. *Genetics*, 195(1), 205–220. <https://doi.org/10.1534/genetics.113.152462>
- 763 Haldane, J. B. S. (1930). A mathematical theory of natural and artificial selection.  
764 (Part VI, Isolation.). *Math. Proc. Cambridge Philos. Soc.*, 26, 220–230.
- 765 Hoban, S., Kelley, J. L., Lotterhos, K. E., Antolin, M. F., Bradburd, G., Lowry, D. B., ...  
766 Whitlock, M. C. (2016). Finding the genomic basis of local adaptation: Pitfalls,  
767 practical solutions, and future directions. *American Naturalist*, 188(4), 379–397.  
768 <https://doi.org/10.1086/688018>
- 769 Hofer, T., Ray, N., Wegmann, D., & Excoffier, L. (2009). Large allele frequency  
770 differences between human continental groups are more likely to have occurred  
771 by drift during range expansions than by selection. *Annals of Human Genetics*,  
772 73(1), 95–108. <https://doi.org/10.1111/j.1469-1809.2008.00489.x>
- 773 Hohenlohe, P. A., Phillips, P. C., & Cresko, W. A. (2010). Using population genomics  
774 to detect selection in natural populations: Key concepts and methodological  
775 considerations. *International Journal of Plant Sciences*, 171(9), 1059–1071.  
776 <https://doi.org/10.1086/656306>
- 777 Horscroft, C., Ennis, S., Pengelly, R. J., Sluckin, T. J., & Collins, A. (2019). Sequencing  
778 era methods for identifying signatures of selection in the genome. *Briefings in*  
779 *Bioinformatics*, 20(6), 1997–2008. <https://doi.org/10.1093/bib/bby064>
- 780 Huber, C. D., DeGiorgio, M., Hellmann, I., & Nielsen, R. (2016). Detecting recent  
781 selective sweeps while controlling for mutation rate and background selection.  
782 *Molecular Ecology*, 25(1), 142–156. <https://doi.org/10.1111/mec.13351>
- 783 Jones, F. C., Grabherr, M. G., Chan, Y. F., Russell, P., Mauceli, E., Johnson, J., ...  
784 Kingsley, D. M. (2012). The genomic basis of adaptive evolution in threespine  
785 sticklebacks. *Nature*, 484(7392), 55–61. <https://doi.org/10.1038/nature10944>
- 786 Joyce, P., & Marjoram, P. (2008). Approximately sufficient statistics and bayesian  
787 computation. *Statistical Applications in Genetics and Molecular Biology*, 7(1).  
788 <https://doi.org/10.2202/1544-6115.1389>
- 789 Kawecki, T. J., & Ebert, D. (2004). Conceptual issues in local adaptation. *Ecology*  
790 *Letters*, 7(12), 1225–1241. <https://doi.org/10.1111/j.1461-0248.2004.00684.x>
- 791 Kingman, J. F. C. (1982). The coalescent. *Stochastic Processes and Their Applications*,  
792 13(3), 235–248. [https://doi.org/10.1016/0304-4149\(82\)90011-4](https://doi.org/10.1016/0304-4149(82)90011-4)

- 793 Korneliussen, T. S., Albrechtsen, A., & Nielsen, R. (2014). ANGSD: Analysis of Next  
794 Generation Sequencing Data. *BMC Bioinformatics*, 15(1), 356.  
795 <https://doi.org/10.1186/s12859-014-0356-4>
- 796 Lai, Y. T., Yeung, C. K. L., Omland, K. E., Pang, E. L., Hao, Y., Liao, B. Y., ... Li, S. H.  
797 (2019). Standing genetic variation as the predominant source for adaptation of a  
798 songbird. *Proceedings of the National Academy of Sciences of the United States of*  
799 *America*, 116(6), 2152–2157. <https://doi.org/10.1073/pnas.1813597116>
- 800 Lenormand, T. (2002). Gene flow and the limits to natural selection. *Trends in Ecology*  
801 *& Evolution*, 17(4), 183–189. <https://doi.org/10.1111/j.1365-2184.1976.tb01248.x>
- 802 Li, H., & Stephan, W. (2006). Inferring the demographic history and rate of adaptive  
803 substitution in *Drosophila*. *PLoS Genetics*, 2(10), 1580–1589.  
804 <https://doi.org/10.1371/journal.pgen.0020166>
- 805 Li, J., Li, H., Jakobsson, M., Li, S., Sjödin, P., & Lascoux, M. (2012). Joint analysis of  
806 demography and selection in population genetics: Where do we stand and  
807 where could we go? *Molecular Ecology*, 21(1), 28–44.  
808 <https://doi.org/10.1111/j.1365-294X.2011.05308.x>
- 809 Li, M., Zhang, D., Gao, Q., Luo, Y., Zhang, H., Ma, B., ... Xue, Y. (2019). Genome  
810 structure and evolution of *Antirrhinum majus* L. *Nature Plants*, 5(2), 174–183.  
811 <https://doi.org/10.1038/s41477-018-0349-9>
- 812 Lotterhos, K. E. (2019). The effect of neutral recombination variation on genome  
813 scans for selection. *G3: Genes, Genomes, Genetics*, 9(6), 1851–1867.  
814 <https://doi.org/10.1534/g3.119.400088>
- 815 Lotterhos, K. E., & Whitlock, M. C. (2014). Evaluation of demographic history and  
816 neutral parameterization on the performance of FST outlier tests. *Molecular*  
817 *Ecology*, 23(9), 2178–2192. <https://doi.org/10.1111/mec.12725>
- 818 Luikart, G., England, P. R., Tallmon, D., Jordan, S., & Taberlet, P. (2003). The power  
819 and promise of population genomics: From genotyping to genome typing.  
820 *Nature Reviews Genetics*, 4(12), 981–994. <https://doi.org/10.1038/nrg1226>
- 821 Marjoram, P., & Tavaré, S. (2006). Modern computational approaches for analysing  
822 molecular genetic variation data. *Nature Reviews Genetics*, 7(10), 759–770.  
823 <https://doi.org/10.1038/nrg1961>
- 824 Naisbit, R. E., Jiggins, C. D., & Mallet, J. (2001). Disruptive sexual selection against  
825 hybrids contributes to speciation between *Heliconius cydno* and *Heliconius*  
826 *melpomene*. *Proceedings of the Royal Society B: Biological Sciences*, 268(1478), 1849–  
827 1854. <https://doi.org/10.1098/rspb.2001.1753>
- 828 Nosil, P., Vines, T. H., & Funk, D. J. (2005). Perspective: Reproductive Isolation  
829 Caused By Natural Selection Against Immigrants From Divergent Habitats.  
830 *Evolution*, 59(4), 705. <https://doi.org/10.1554/04-428>
- 831 Oleksyk, T. K., Smith, M. W., & O'Brien, S. J. (2010). Genome-wide scans for  
832 footprints of natural selection. *Philosophical Transactions of the Royal Society B:*  
833 *Biological Sciences*, 365(1537), 185–205. <https://doi.org/10.1098/rstb.2009.0219>

- 834 Peter, B. M., Huerta-Sanchez, E., & Nielsen, R. (2012). Distinguishing between  
835 Selective Sweeps from Standing Variation and from a De Novo Mutation. *PLoS*  
836 *Genetics*, 8(10). <https://doi.org/10.1371/journal.pgen.1003011>
- 837 Petry, D. (1983). The effect on neutral gene flow of selection at a linked locus.  
838 *Theoretical Population Biology*, 23(3), 300–313. [https://doi.org/10.1016/0040-](https://doi.org/10.1016/0040-5809(83)90020-5)  
839 5809(83)90020-5
- 840 Price, A. L., Patterson, N. J., Plenge, R. M., Weinblatt, M. E., Shadick, N. A., & Reich,  
841 D. (2006). Principal components analysis corrects for stratification in genome-  
842 wide association studies. *Nature Genetics*, 38(8), 904–909.  
843 <https://doi.org/10.1038/ng1847>
- 844 Przeworski, M. (2002). The Signature of Positive Selection at Randomly Chosen Loci.  
845 *Genetics*, 162(4), 1179–1189.
- 846 Reid, N. M., Proestou, D. A., Clark, B. W., Warren, W. C., Colbourne, J. K., Shaw, J.  
847 R., ... Whitehead, A. (2016). The genomic landscape of rapid repeated  
848 evolutionary adaptation to toxic pollution in wild fish. *Science*, 354(6317), 1305–  
849 1308. <https://doi.org/10.1126/science.aah4993>
- 850 Rundle, H. D., & Whitlock, M. C. (2001). A genetic interpretation of ecologically  
851 dependent isolation. *Evolution*, 55(1), 198–201.
- 852 Savolainen, O., Lascoux, M., & Merilä, J. (2013). Ecological genomics of local  
853 adaptation. *Nature Reviews Genetics*, 14(11), 807–820.  
854 <https://doi.org/10.1038/nrg3522>
- 855 Schluter, D. (2000). *The Ecology of Adaptive Radiation*. Oxford University Press, Oxford,  
856 U.K. Retrieved from [https://global.oup.com/academic/product/the-ecology-of-](https://global.oup.com/academic/product/the-ecology-of-adaptive-radiation-9780198505228?cc=ch&lang=en&)  
857 [adaptive-radiation-9780198505228?cc=ch&lang=en&](https://global.oup.com/academic/product/the-ecology-of-adaptive-radiation-9780198505228?cc=ch&lang=en&)
- 858 Schwarz-Sommer, Z., Davies, B., & Hudson, A. (2003). An everlasting pioneer: The  
859 story of *Antirrhinum* research. *Nature Reviews Genetics*, 4(8), 655–664.  
860 <https://doi.org/10.1038/nrg1127>
- 861 Schwinn, K., Venail, J., Shang, Y., Mackay, S., Alm, V., Butelli, E., ... Martin, C.  
862 (2006). A Small Family of MYB-Regulatory Genes Controls Floral Pigmentation  
863 Intensity and Patterning in the Genus *Antirrhinum*. *The Plant Cell*, 18(April),  
864 831–851. <https://doi.org/10.1105/tpc.105.039255.1>
- 865 Slatkin, M. (1973). Gene flow and selection in a cline. *Genetics*, 75(4), 733–756.
- 866 Slatkin, Montgomery, & Excoffier, L. (2012). Serial founder effects during range  
867 expansion: A spatial analog of genetic drift. *Genetics*, 191(1), 171–181.  
868 <https://doi.org/10.1534/genetics.112.139022>
- 869 Sousa, V. C., Carneiro, M., Ferrand, N., & Hey, J. (2013). Identifying loci under  
870 selection against gene flow in isolation-with-migration models. *Genetics*, 194(1),  
871 211–233. <https://doi.org/10.1534/genetics.113.149211>
- 872 Sugden, L. A., Atkinson, E. G., Fischer, A. P., Rong, S., Henn, B. M., &  
873 Ramachandran, S. (2018). Localization of adaptive variants in human genomes  
874 using averaged one-dependence estimation. *Nature Communications*, 9(1).

- 875 <https://doi.org/10.1038/s41467-018-03100-7>
- 876 Szpiech, Z. A., & Hernandez, R. D. (2014). Selscan: An efficient multithreaded  
877 program to perform EHH-based scans for positive selection. *Molecular Biology*  
878 *and Evolution*, 31(10), 2824–2827. <https://doi.org/10.1093/molbev/msu211>
- 879 Tavares, H., Whibley, A., Field, D. L., Bradley, D., Couchman, M., & Copsey, L.  
880 (2018). Selection and gene flow shape genomic islands that control floral guides,  
881 1–6. <https://doi.org/10.1073/pnas.1801832115>
- 882 Vitti, J. J., Grossman, S. R., & Sabeti, P. C. (2013). Detecting Natural Selection in  
883 Genomic Data. *Annual Review of Genetics*, 47(1), 97–120.  
884 <https://doi.org/10.1146/annurev-genet-111212-133526>
- 885 Wakeley, J. (2001). The coalescent in an island model of population subdivision with  
886 variation among demes. *Theoretical Population Biology*, 59(2), 133–144.  
887 <https://doi.org/10.1006/tpbi.2000.1495>
- 888 Wegmann, D., Leuenberger, C., & Excoffier, L. (2009). Efficient approximate Bayesian  
889 computation coupled with Markov chain Monte Carlo without likelihood.  
890 *Genetics*, 182(4), 1207–1218. <https://doi.org/10.1534/genetics.109.102509>
- 891 Wegmann, D., Leuenberger, C., Neuenschwander, S., & Excoffier, L. (2010).  
892 ABCtoolbox: a versatile toolkit for approximate Bayesian computations. *BMC*  
893 *Bioinformatics*, 11, 116. <https://doi.org/10.1186/1471-2105-11-116>
- 894 Whibley, A. C., Langlade, N. B., Andalo, C., Hanna, A. I., Bangham, A., Thébaud, C.,  
895 & Coen, E. (2006). Evolutionary paths underlying flower color variation in  
896 *Antirrhinum*. *Science*, 313(5789), 963–966. <https://doi.org/10.1126/science.1129161>
- 897 Williamson, S. H., Hernandez, R., Fledel-alon, A., Zhu, L., Nielsen, R., & Bustamante,  
898 C. D. (2005). Simultaneous inference of selection and population growth from  
899 patterns of variation in the human genome. *PNAS*, 102(22), 7882–7887. Retrieved  
900 from  
901 [http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1142382&tool=pmce](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1142382&tool=pmcentrez&rendertype=abstract)  
902 [ntrez&rendertype=abstract](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1142382&tool=pmcentrez&rendertype=abstract)
- 903 Wright, S. (1931). Evolution in Mendelian Populations. *Genetics*, 16(2), 97–159.
- 904 Yeaman, S. (2015). Local adaptation by alleles of small effect. *American Naturalist*,  
905 186(october), S74–S89. <https://doi.org/10.1086/682405>
- 906 Zeng, K., Shi, S., & Wu, C. I. (2007). Compound tests for the detection of hitchhiking  
907 under positive selection. *Molecular Biology and Evolution*, 24(8), 1898–1908.  
908 <https://doi.org/10.1093/molbev/msm119>

909  
910

## 911 DATA ACCESSIBILITY

912  
913  
914  
915

We provide scripts to perform LSD genome scans at the GitHub repository:  
<https://github.com/hirzi/LSD>.

916

917 **AUTHOR CONTRIBUTIONS**

918

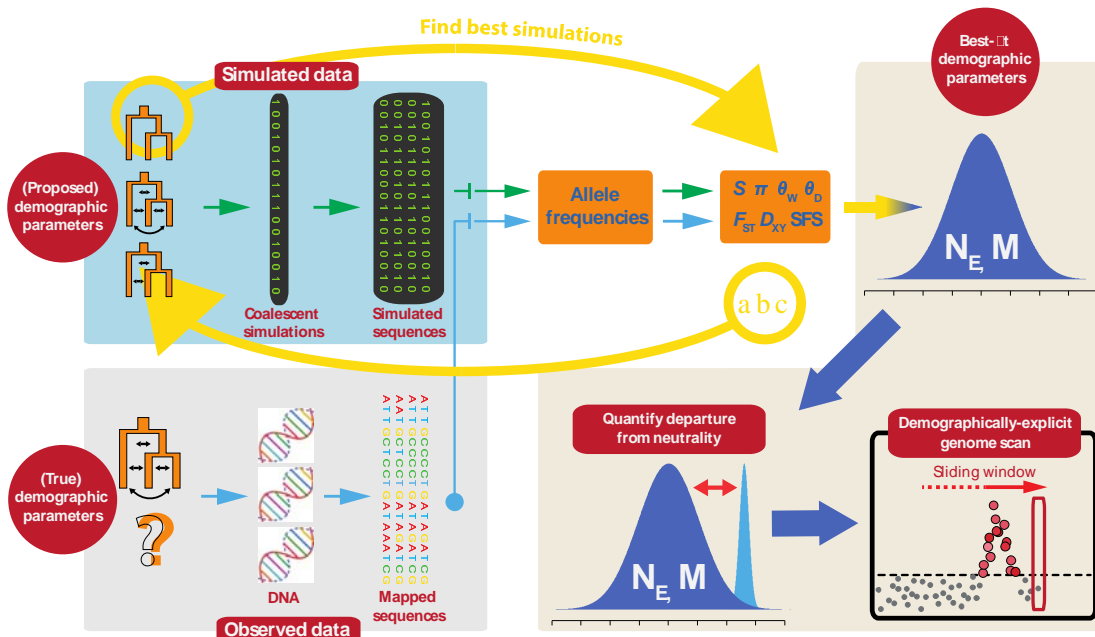
919 HL, DW, AW and SF designed the study. HL wrote the LSD scripts and performed the  
920 simulations and analyses. HL and DW developed the methods. HL wrote the manuscript,  
921 which all authors critically revised.

922

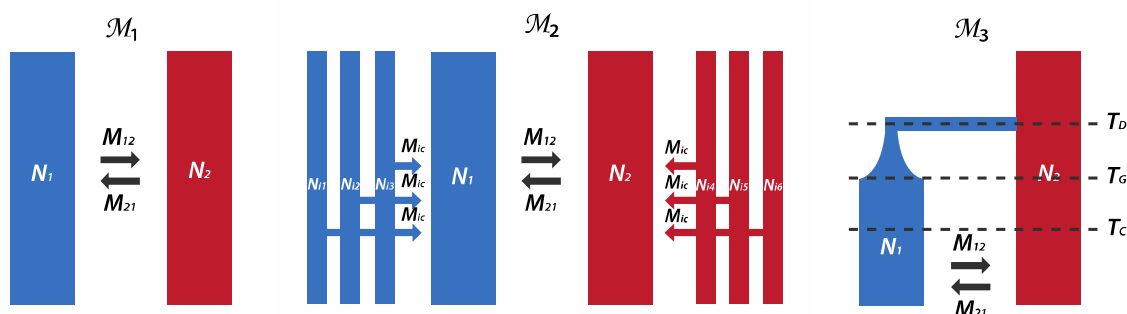
923



924 FIGURES  
925



926  
927 **Figure 1.** Identifying Loci under Selection via explicit Demographic models (LSD).  
928 LSD identifies loci under selection by first estimating demographic parameters and  
929 then quantifying the departure of these parameters from neutral expectations. Our  
930 specific implementation of LSD employs Approximate Bayesian Computation (ABC)  
931 for parameter estimation, and is performed in a genome scan approach.  
932



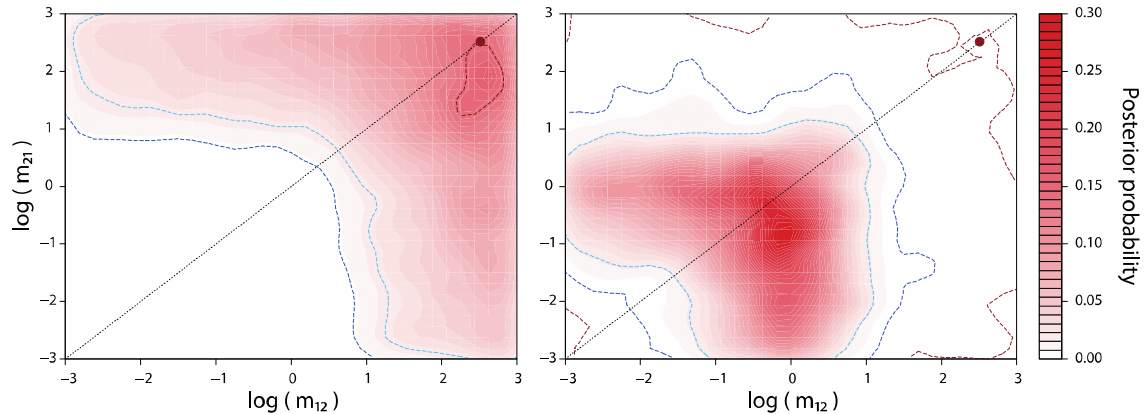
933  
934 **Figure 2.** Models used in the simulations and case study. Model  $\mathcal{M}_1$  represents a  
935 simple 2-deme isolation-with-migration (IM) model with reciprocal migration.  
936 Model  $\mathcal{M}_2$  represents a 6-deme island-continent model where common differences  
937 between environments are modelled by connecting the sampled demes (i.e. islands)  
938 to respective meta-population continents via gene flow. Model  $\mathcal{M}_3$  represents a 2-  
939 deme divergence with bottleneck and exponential growth model. Different deme  
940 colours reflect contrasting environments. In all models, selection is inferred from the

941 deviation from neutrality of the reciprocal migration rates between the two  
942 contrasting environments.

943

944

945



946

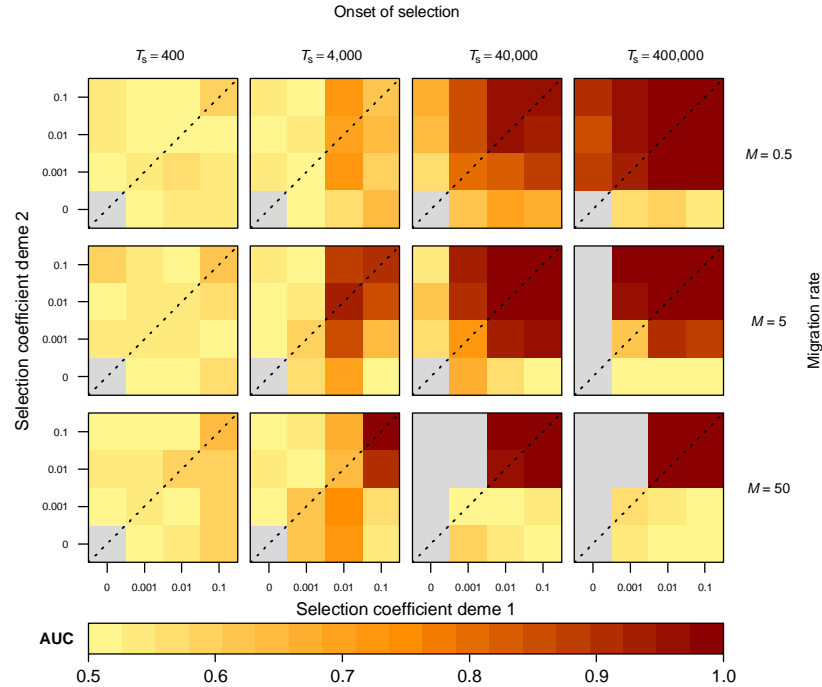
947 **Figure 3.** Joint posterior distribution of the reciprocal migration parameters,  $m_{12}$  and  
948  $m_{21}$ . The neutral joint parameter estimate, as informed by the global posterior  
949 distribution of all neutral regions (Fig. S1), is indicated by the red dot in the top right  
950 corner. The red contours represent the joint posterior distribution of a genomic  
951 region (i.e. window), with the blue contours representing the 95% (light blue) and  
952 99% (dark blue) highest density region (HDR) credible intervals. Left – a window not  
953 significantly divergent from the neutral estimate; right – a window significantly  
954 divergent from the neutral estimate, with slightly higher relative reduction in  $m_{12}$   
955 than in  $m_{21}$ .

956

957

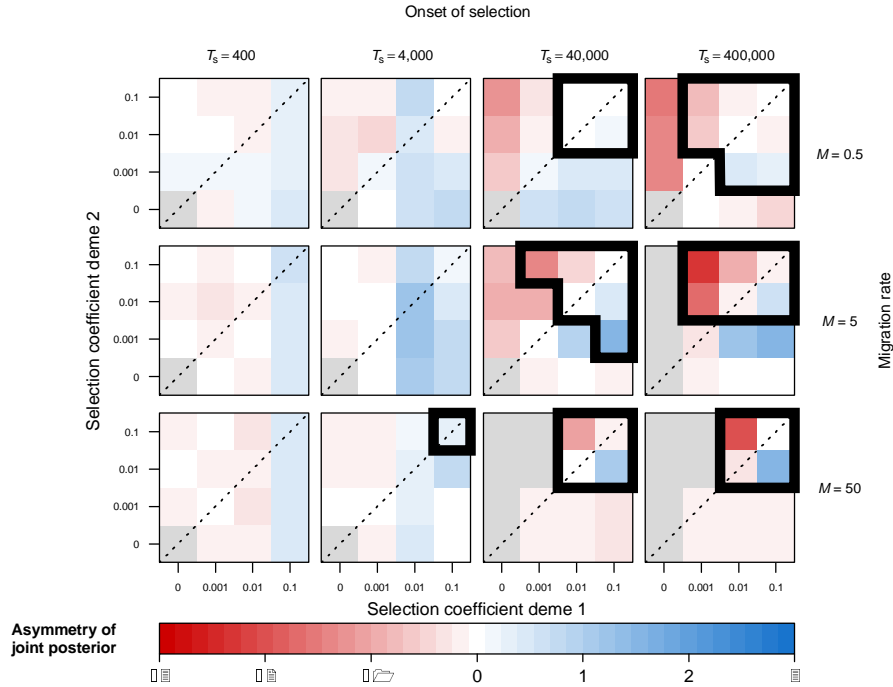
958

959



960  
 961  
 962  
 963  
 964  
 965  
 966  
 967  
 968  
 969  
 970

**Figure 4.** Simulation results showing the effect of migration rate, time of onset of selection and deme-specific selection coefficients on LSD diagnostic performance (AUC), for the 2-deme IM model (model  $\mathcal{M}_1$ ; standing genetic variation case). Each cell represents a pseudo-genome simulated under a specific selection regime. The cell colours reflect the AUC calculated by the correct discrimination of 1000 neutral loci and 50 selected loci in the 1050 loci simulated pseudo-genomes. An AUC value of 0.5 reflects random assignment while that of 1 reflects perfect classification (i.e. TPR=1, FPR=0). Grey cells indicate selection regimes where the derived allele is always lost.



971

972

**Figure 5.** Simulation results showing the effect of migration rate, time of onset of

973

selection and deme-specific selection coefficients on LSD inferred (a)symmetry of

974

selection, for the 2-deme IM model (model  $\mathcal{M}_1$ ; standing genetic variation case).

975

Each cell represents a pseudo-genome simulated under a specific selection regime.

976

The cell colours reflect the (a)symmetry values inferred by LSD, where a value of 0

977

reflects perfect symmetry of the joint posterior while values divergent from this

978

reflect asymmetry. Cells surrounded by thick lines indicate the values of

979

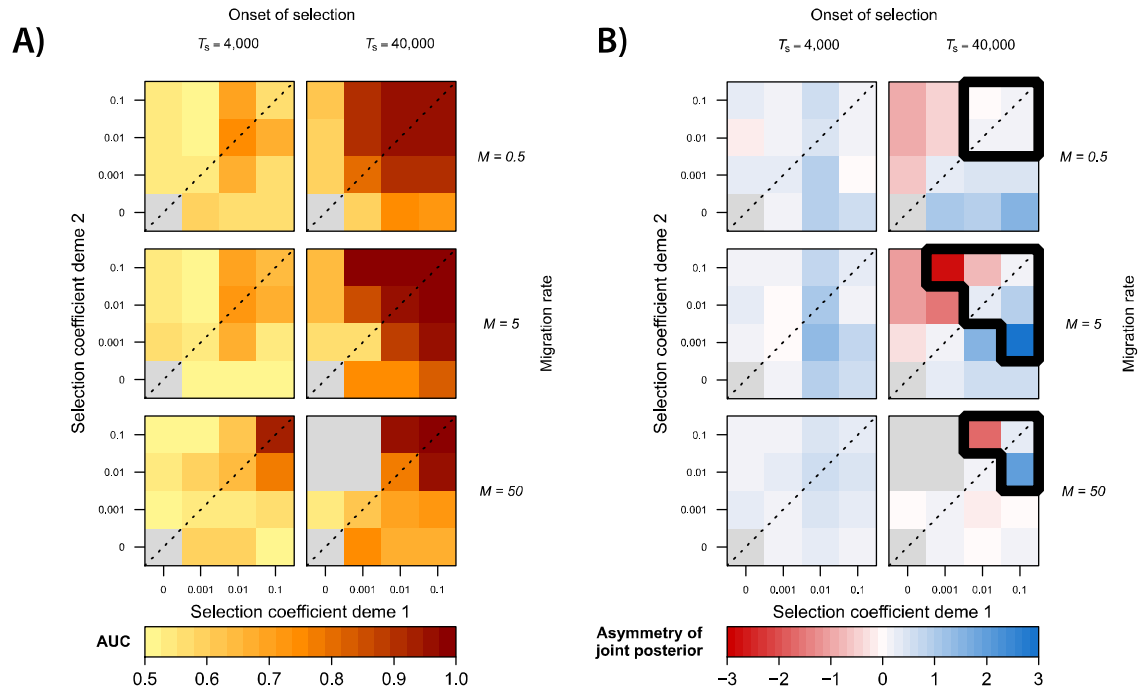
(a)symmetry for regimes expected to generate meaningful signal (i.e.  $AUC > 0.95$ ).

980

Grey cells indicate selection regimes where the derived allele is always lost.

981

982



983

984

**Figure 6.** Simulation results for a 2-deme divergence with bottleneck and exponential growth model (model  $\mathcal{M}_3$ ; standing genetic variation case) showing the effect of migration rate, time of onset of selection and deme-specific selection coefficients on A) LSD diagnostic performance (AUC) and B) LSD inferred (a)symmetry of selection. Divergence time of the two populations,  $T_D$ , is 200,00 generations ago. Each coloured cell represents a pseudo-genome simulated under a specific selection regime. Grey cells indicate selection regimes where the derived allele is always lost. B) Cells surrounded by thick lines indicate the values of (a)symmetry for regimes expected to generate meaningful signal (i.e. AUC>0.95).

986

987

988

989

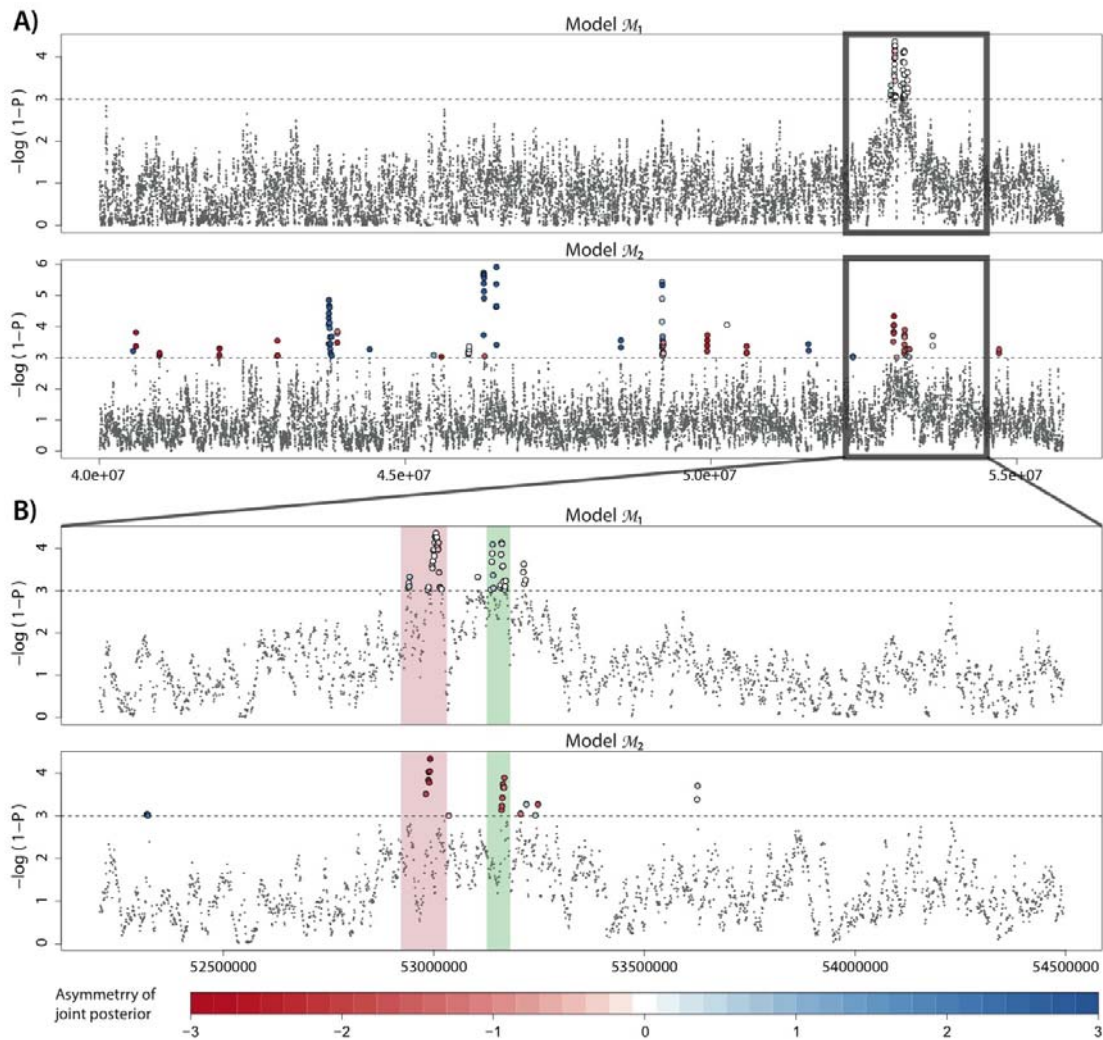
990

991

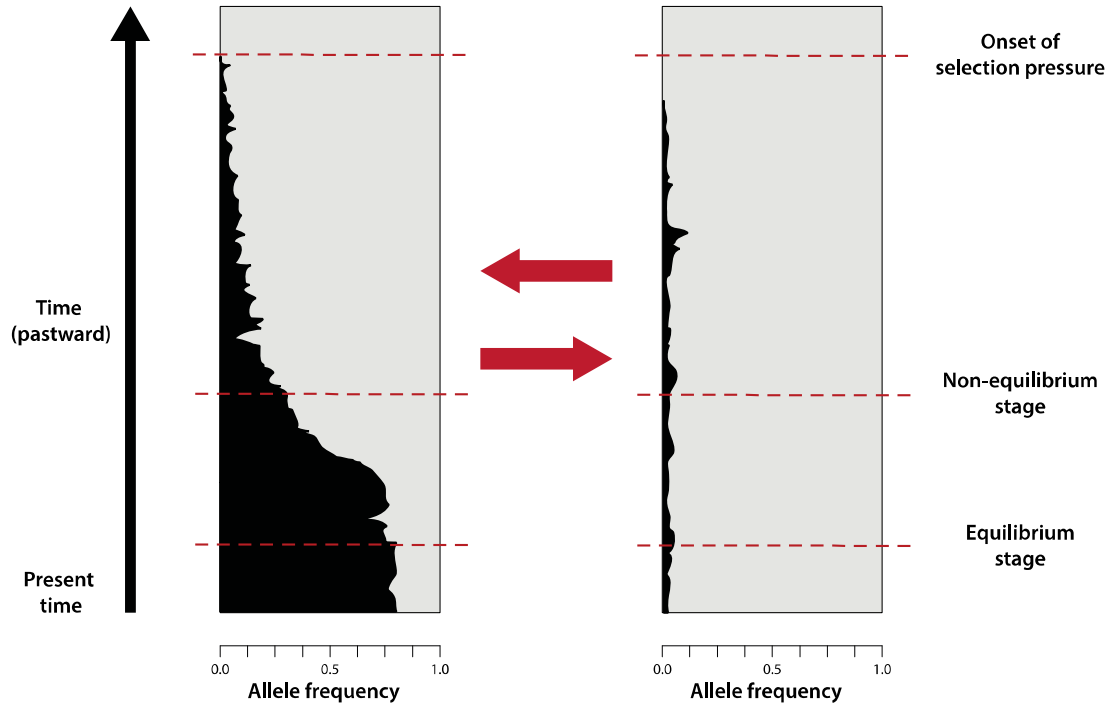
992

993

994



995  
996 **Figure 7.** Manhattan plot for the LSD scan of the *A.m.striatum*-*A.m.pseudomajus*  
997 system. The posterior probability of observing the neutral estimate for 10kb windows  
998 (1kb step-size) is plotted for A) a 16Mb region of chromosome 6, under models  $\mathcal{M}_1$   
999 and  $\mathcal{M}_2$ , and B) a 2Mb zoomed-in region of chromosome 6 focusing on the ROS-EL  
1000 region, under the same two models. The horizontal dashed line indicates a 99.9%  
1001 posterior probability of deviating from neutral expectations. Colour for loci above  
1002 this threshold denotes the joint ( $m_{12}$ - $m_{21}$ ) posterior (a)symmetry, and reflects the  
1003 relative strengths of selection in the two divergent demes or subspecies. A large  
1004 divergent peak centered around the ROS-EL region (A) is composed of a set of  
1005 smaller peaks (B), consistent with the ROS (red) and EL (green) loci.  
1006  
1007



1008

1009

1010

1011

1012

1013

1014

1015

1016

**Figure 8.** Conceptual illustration of allele frequency trajectories over time in a 2-deme IM model ( $\mathcal{M}_1$ ), for an example *de-novo* case and antagonistic pleiotropic selection regime. The frequency of derived allele *A* is indicated in black and that of ancestral allele *a* in grey. Red arrows represent migration. Prior to reaching drift-migration-selection equilibrium, estimated asymmetries in effective migration rates are also affected by asymmetry in allele frequencies.