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8

Ovariectomy as a Management Technique in Suburban Deer Populations

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ABSTRACT Overabundant suburban deer (*Odocoileus* spp.) are a source of human-wildlife

conflict in many communities throughout the United States. Deer-vehicle collisions, tick-borne

pathogens, impacts on local vegetation, and other negative interactions are the typical reasons

cited for initiating a deer management program. Social attitudes, legal constraints, and perceived

safety concerns lead many communities to examine nonlethal management options. Surgical

sterilization is currently the only nonlethal method available to permanently sterilize females with

a single treatment. There are limited data demonstrating methods and outcomes in management

programs that sterilize a high percentage (>90%) of the local population, particularly regarding

the impact of immigration on non-isolated populations. We present data from 6 surgical

sterilization sites with geographically open populations in California, Maryland, Michigan, New

York, Ohio, and Virginia, USA. From 2012–2020, we sterilized 493 deer primarily via

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ovariectomy. We conducted annual or periodic population estimates using camera surveys, road-

based distance sampling, and intensive field observations to assess population trends. Initial

densities ranged from about 6–63 deer/km^2. Study sites ranged from 1.2 km^2 to 16.5 km^2, and

initial populations ranged from ~47 to 169 individuals. For our 6 study sites, we noted an average

reduction in deer abundance of approximately 25% (range: 16.2%–36.2%) from Year 1 to Year 2.

Four years after the first treatment, at monitored sites ($n = 4$) using this management method, we

noted an average total population reduction of about 45% (range: 28%–56%). During the first

year, the average cost per deer handled was $1,221 (range: $864–$1,998). These projects

demonstrate that significant reductions in local deer densities using high percentage surgical

sterilization programs can be achieved in non-insular locations. Sustained sterilization efforts are

necessary, as is the case with all deer management programs in open landscapes.

KEY WORDS deer management, human-wildlife conflicts, nonlethal, *Odocoileus* spp.,

ovariectomy, population management

In many urban and suburban situations, lethal management techniques for deer (*Odocoileus* spp.)

such as controlled hunting and sharpshooting are not feasible or practical because of legal, social,

public safety, or economic considerations (Williams et al. 2013, Kilpatrick et al. 2014). As a

result, various non-lethal management techniques such as trap and relocation or fertility control

have been explored (DeNicola et al. 2000). Fertility control options have been prioritized because

trap and relocation has been demonstrated to be stressful and result in unacceptably high post-

release mortality (Beringer et al. 2002).

Extensive research has been conducted on injectable or implantable wildlife fertility

control agents for nearly 5 decades, with numerous studies conducted in the 1970s and 1980s

(Harder and Peterle 1974, Bell and Peterle 1975, Matschke 1977, Kirkpatrick and Turner 1985,

Plotka and Seal 1989). Recent research has transitioned focus to vaccines and surgical

sterilization as the most promising approaches (Patten et al. 2007). Given the scope of research,

there has been relatively little improvement in the function of vaccines tested in the late-1980s on

horses (Liu et al. 1989, Kirkpatrick et al. 1990) or deer in the 1990s (Turner et al. 1992, McShea

et al. 1997, Miller et al. 1997, Miller et al. 2000). For long-lived species, at least one booster is

necessary to predictably maintain >90% effectiveness for >2 years, even for vaccines formulated

as a single dose (Gionfriddo et al. 2011, Rutberg et al. 2013a, Roelle et al. 2017).

Wildlife professionals have discussed the potential of fertility control as a wildlife

management technique for as long as efficacy trials have been conducted (McCullough 1987,

Bomford 1992, Garrott 1995, Warren 1995, Miller et al. 1998). Most studies have focused on

delivery methods and treatment efficacy, with very few assessments of management outcomes. In

addition, research to date has primarily described the limitations of fertility control versus its

potential to effectively manage deer overabundance in geographically open environments. In fact,

several studies have concluded that for fertility control to be successful, its use must be limited to

small, insular, or fenced areas (Seagle and Close 1996, Merrill et al. 2006, Boulanger et al. 2012,

Boulanger and Curtis 2016). The conclusion that the method is not viable in open populations is

founded on population-level field studies conducted at the Cornell University Campus, Ithaca,

New York, USA (Boulanger and Curtis 2016), Fripp Island, South Carolina, USA (Rutberg et al.

2013b), National Institute of Standards and Technology, Gaithersburg, Maryland, USA (Rutberg

and Naugle 2008), and Fire Island National Seashore, Patchogue, New York, USA (Naugle et al.

2002), as well as various modeling efforts (Seagle and Close 1996, Merrill et al. 2006). One

modeling effort projected a reduced local population using surgical sterilization at Cumberland

Island, Georgia, USA (Boone and Wiegert 1994).

Conclusions on the potential management benefits of fertility control have been based on

compromised methods and incorrect assumptions regarding immigration rates. In the Cornell

University study that used tubal ligation as the surgical method, it was concluded that surgical

sterilization would not work in geographically open environments without a lethal component

(Boulanger and Curtis 2016). In this study, adult female and fawn numbers declined

significantly, reflecting an effective program, but male abundance increased 9-fold, preventing an

overall population decline. Given the sterilization method used caused extended estrous cycles,

males were potentially attracted to females during later estrus periods. Antlered males may also

have sought refuge in the non-hunted study site, given the earn-a-buck hunting program that

occurred around the site (Williams et al. 2008, Boulanger et al. 2014). Another study found

limited immigration and no increase of adult males due to sterilization of adult females via tubal

ligation in an open suburban environment without proximate hunting (MacLean et al. 2006).

The first assessment of surgical sterilization on deer was conducted in the early 1990s

(Frank and Sajdak 1993) and additional research on surgical methods occurred in the 2000s

(MacLean et al. 2006, Boulanger et al. 2012). These efforts were initiated in recognition that

immunocontraceptive vaccines were limited in their long-term effectiveness, and an alternative

was needed in areas where treating the same animal more than once was too difficult or costly.

As a result, we decided to conduct additional fertility control research, using surgical sterilization

via ovariectomy, to reduce population abundance while eliminating breeding behavior associated

with tubal ligations. Our interest was to identify a method that could be used as an infertility

treatment and have population-level impacts on white-tailed deer (*O. virginianus*). Our research

focused on the potential of surgical sterilization by ovariectomy of female deer as a management

tool in geographically open suburban environments and suburban environments with security

fences permeable to deer. Given the strong philopatry of female white-tailed deer, immigration

could be minimal (Porter et al. 2004). We selected research locations based on fundamental

needs for extensive access and approachable deer (McCullough 1987, Garrott 1995, Rudolph et

al. 2000). Most researchers have concluded that 60–80% or greater of a deer population needs to

be rendered infertile to stabilize or begin to reduce a local population (McCullough 1987, Hone

1992, Garrott 1995, Grund 2011, Boulanger et al. 2012). Our objectives in each designated study

area were to 1) capture and permanently sterilize >95% of females; 2) assess immigration rates,

mortality, and changes in population abundance; and 3) assess effort.

STUDY AREAS

We applied surgical sterilization treatments to free-ranging deer on 6 study sites. White-tailed

deer were the target species at every site except The Villages Golf and Country Club (VGCC),

where we captured and treated black-tailed deer (*O. hemionus columbianus*). Deer capture efforts

were conducted between August and March from 2012–2020 to minimize stress on deer and

maximize baiting success. The justification for attempting to reduce deer abundance included

increasing deer-vehicle collisions, landscape damage, forest diversity impacts, or tick-borne

infection concerns. Decision-makers at each location opted to use a non-lethal approach because

of either social or legal constraints.

Our study sites were typified by a mixture of developed urban or suburban landscapes

with intermingled wetlands or other small to moderate open spaces. Regional differences in

climate among our various study sites affected the local natural plant communities. Temperate,

mesic hardwood forests predominantly composed the open areas at 5 study sites. Xeric

shrublands with dry oak savanna surrounded VGCC. Except for the National Institutes of Health

(NIH) campus, the sites were primarily single-family residential communities with property sizes

ranging from 0.4–2.0 ha with some larger private properties interspersed, such as golf courses.

Only the NIH campus and the VGCC community had security fencing. Neither fence prevented

deer from entering or exiting with documented emigration from both locations and annual

immigration at NIH. The other study sites were not isolated from the surrounding areas that

contained resident deer populations. The study sites did not permit hunting because of safety

concerns associated with high-density development and significant human activity. Hunting or

deer management activities occurred in communities adjacent to 4 of our study sites.

The study site in Ann Arbor, Michigan, USA (N42.2709, W83.7263; AA) consisted of 4.3

km^2, and most of its land area was covered by single-family residential communities surrounded

by wooded corridors (Fig. 1). The community opted to incorporate culling into the broader

program scope because of the disparity in development patterns. Sharpshooting occurred adjacent

to the northern and eastern borders of the study site. Sterilized deer were not culled during

sharpshooting operations. The City's management goal was to reduce deer densities to the point

where 75% of surveyed residents reported satisfaction with the deer abundance.

The Clifton neighborhood of Cincinnati, Ohio, USA (N39.1531, W84.5248; CLIF), was

located in the north-central section of the city and the 2.4 km^2 study site contained Mt. Storm,

Rawson Woods, and Edgewood Grove parks, as well as the surrounding neighborhoods

consisting of single-family residential communities and apartment complexes. The parks were

generally undeveloped and included steep wooded hillsides and some mowed open space.

Because deer were not limited to the parks under consideration, we delineated the study area as

depicted in Fig. 2. Deer densities had reached a level that was considered incompatible with the

Cincinnati Park Board's goal of a healthy park ecosystem.

140   Fairfax City, Virginia, USA (N38.8462, W77.3064; FC) was located in the western

141 suburbs of Washington, D.C., and is ~16.5 km^2 (Fig. 3). The area consisted of single-family

142 residential communities, apartment complexes, and commercial-use properties with wooded

143 corridors. The use of bait for capture or camera survey population estimates was not permitted

144 during the first year. After we demonstrated that baiting would be instrumental to the project's

145 success and improved accuracy of camera surveys, Virginia Department of Game and Inland

146 Fisheries (VA DGIF) amended the research permit to allow the use of bait.

147   The NIH site (N38.9992, W77.1025) was an office park complex located in Bethesda,

148 Maryland, USA (Fig. 4). The study area was 1.2 km^2 and consisted of more than 95 office

149 buildings in a campus-like environment. The site contained small natural wooded and landscaped

150 areas interspersed with the dense commercial development. Landscape damage and deer-vehicle

151 collisions were of concern.

152   The Village of Cayuga Heights, New York, USA (N42.4687, W76.4856; VCH) was

153 located adjacent to and contiguous with the City of Ithaca and contained 4.6 km^2 (Fig. 5). The

154 area consisted of single-family residential communities, apartment complexes, and commercial-

155 use properties with wooded corridors. At the start of this study, New York State law prevented

156 discharge of a weapon such as a bow, muzzleloader, or firearm within 152 m of an occupied

157 dwelling without owner permission. As a result, and given local opposition to a lethal

158 management program, there were no legal locations to cull deer in VCH. Therefore, elected

159 officials voted to implement a surgical sterilization initiative starting in 2012, given the

160 abundance of deer and associated conflicts. In 2014, the law changed reducing archery discharge

161 to 76 m, which permitted a cull that was conducted from 2015–2017. The community leaders

162 decided to incorporate archery culling to accelerate the population decline. The State of New

York then prohibited baiting within 91 m of a public road in 2017, and only 2 properties remained

legal given the high road density. As a result, VCH decided to transition to a remote

immobilization program followed by euthanasia from 2018–2019 to obtain even lower deer

densities. We included data and discussion on these subsequent culling activities to allow for

immigration insights as the population declined. However, we only included data collected prior

to culling in our analysis.

The VGCC was located in San Jose, California, USA (N37.2878, W121.7466) and

consisted of a 2.7 km^2 private community with 2,536 homes and an 18-hole golf course (Fig. 6).

Landscape damage and replacement costs exceeded $250,000 annually because of deer impacts

(B. Barncord, personal communication), and the community elected to execute a non-lethal

management program as a result.

METHODS

Licensed and trained veterinarians conducted all surgical procedures. White Buffalo Inc.

employees, trained by a wildlife veterinarian in humane capture and chemical restraint methods,

conducted all capture efforts. We performed all field procedures under Michigan Department of

Natural Resources Scientific Collectors Permit #1600, Ohio Department of Natural Resources

Scientific Collection Permit #18-48, VA DGIF Scientific Collection Permit #058581, Maryland

Department of Natural Resources Deer Cooperator Permit 55832-NIH-2014-2019, and NIH

Office of Research Services Animal Study Proposal Animal Care and Use Committee #ORS-54,

New York State Department of Environmental Conservation License to Collect and Possess

#2558, and California Department of Fish and Wildlife Scientific Collection Permit #SC-012522.

Deer Capture

We conducted deer capture and sterilization activities between August and March from 2012–

2020. We immobilized female white-tailed deer of all age classes using projectors with 2-ml

transmitter darts (Pneu-Dart, Inc., Williamsport, PA, USA) to administer tiletamine/zolazepam

(4.4 mg/kg) and xylazine hydrochloride (2.2 mg/kg; Kilpatrick et al. 1997). We approached deer

in a vehicle on public roadways and on private roadways and properties where permission was

granted. In AA, FC, and VCH, capture professionals were accompanied by a police officer

during mobile darting efforts (Evans et al. 2016). We also darted deer over 20–25 kg of whole-

kernel corn bait in the late afternoon and early evening where state regulations permitted. Ten

minutes after a dart was deployed, we located deer via radio telemetry, if not visibly recumbent.

In VCH, we also captured deer with drop nets during the first year. We chemically immobilized

adult deer under drop nets with an intramuscular injection of 400 mg ketamine hydrochloride

(HCl) and 150 mg xylazine HCl <2 minutes after capture. We administered 300 mg ketamine

HCl and 100 mg xylazine HCl to fawns. We fitted all deer captured with ear tags for individual

identification (Destron Fearing Duflex X-Large Cattle Tag, Dallas, TX, USA) and the backplate

of each labeled "Call Before Consumption" and a phone number. All individuals were

transported to temporary surgical facilities, except for NIH and 2 years in CLIF, where we used a

veterinary facility. We followed previous deer handling, tagging, very high frequency (VHF)

radio-collaring, and transport methods (Evans et al. 2016).

Surgical Sterilization Procedure

Initially, we conducted bilateral ovariectomies using a combination of clamping, electrocautery,

and excision for removal and coagulation to prevent hemorrhage (Evans et al. 2016). We

transitioned to a vessel sealer in fall 2016 to expedite the procedure and minimize risks associated

with post-surgical bleeding (Valleylab LigaSure, Medtronic, Minneapolis, MN, USA). We

conducted fallopian tubal ligectomies ($n = 26$) only when deemed necessary by veterinarians due

to extensive adhesions that complicated the extraction of the ovaries or during late gestation.

We also followed previous post-surgical release and monitoring procedures (Evans et al.

2016). The one exception was that we relocated 30 deer outside the fence at VGCC to adjacent

VGCC-owned property to assess whether these deer would return or remain outside and increase

the rate of population decline. To differentiate these relocated deer from the rest of the herd, they

received one yellow and one white extra-large ear tag.

Effort

We recorded start and stop times to assess the person-hours required for capture, handling,

transport, and sterilization surgery. Trained biologists and volunteers conducted capture,

handling and transportation to and from surgical locations. For each study site, we divided the

number of person-hours devoted to capture, handling, and transport by the number of deer

processed to derive an estimate of effort per deer. Each surgical team consisted of 2 people,

including trained and licensed veterinarians and veterinary technicians. We had two veterinary

teams performing surgeries during the first year at all project sites. Each capture night's surgical

effort was recorded starting when the first deer was darted and ending when the last deer was

recovered. We reported person-hours per deer for surgical effort.

Population Estimates

We conducted population monitoring using camera data from baited locations, driving distance

transects, and intensive field observations. We noted immigrants, antlered males, fawns, and both

tagged and untagged females. We defined immigrants as untagged deer that were not identified

the previous year.

At AA, CLIF and FC, we conducted baited camera surveys using tagged deer to estimate

deer abundance (Jacobson et al. 1997, Koerth et al. 1997, Curtis et al. 2009). We used game

cameras (D-80 White Flash Trail Cameras, Moultrie, Alabaster, AL, USA) set on motion

activated single shot with a 5-minute delay to optimize capture rates relative to photo storage. We

used camera coverage of ~1/0.61 km^2 with one camera placed in each block. Project staff pre-

baited sites for at least 7 days. We placed each camera 5–6 m from the center of bait, elevated 0.6

m, oriented north, and left for about 2 weeks. We retrieved cameras once we obtained about 300

photos from each baited location. There were some situations where we retrieved cameras with

<300 photos due to low deer activity. In the first year of FC, baiting was not permitted. To

conduct this unbaited camera survey, we divided FC into 12 quadrants and allocated 1 camera to

each quadrant. We placed cameras on public property in each quadrant at locations proximate to

heavily traveled deer trails and pinch points through which deer were forced to pass.

Each image was studied, all legible ear tag numbers were documented, and images of

insufficient quality were excluded due to lens flares or poor image quality. We recorded the total

number of deer, tagged and untagged individuals, unidentifiable tagged deer, identifiable and

unidentifiable antlered males, and untagged fawns in each photo. If a tagged deer was not

observed on camera or directly for 2 years, we eliminated the individual from that year's

population estimate.

We entered the number of tagged, untagged, and tagged but unidentifiable observations

from each photograph into Program NOREMARK (White 1996). Our tagged individuals were

adult females, with occasional female fawns and incidental male fawns. We used the Bowden

estimator that assumes sightability is not influenced by tag status, many animals are sighted

repeatedly, and the population is closed (Bowden 1993, Bowden and Kufel 1995). Our study

areas did not have fully closed populations, but there was likely minimal outside exchange given

deer philopatry, relatively small suburban home ranges, and the camera survey's short duration.

The resulting population estimate and confidence intervals for this method were for antlerless

deer (95% CI). We used the buck:doe ratio (BDR) method to estimate the number of individual

antlered males (Jacobson et al. 1997). We derived a total estimate using the BDR method when

we did not utilize other methods. We then added the number of individual antlered males

identified using the BDR method (BDR M) to the antlerless estimates from Program

NOREMARK to obtain a population estimate noted as NR-BDR. For a second estimate, we used

camera data to determine the ratio of tagged to untagged adult females photographed to estimate

the total number of adult females and fawns, assuming equal sightability (Eberhardt 1969). We

then added BDR M to the photographic ratio estimates of adult females and fawns to obtain a

population estimate, noted as LPE-BDR method. Finally, we used a census approach to derive

the population size. We determined the number of adult females and fawns, given the high

percentage capture status, intensive field observations (e.g., capture observations, bait site camera

data, population estimate data), known mortality and known dispersal data. This census also

included BDR M and field observations. We then averaged the methods used to derive a final

estimate. We also used field observations and photographic data to determine the percentage of

females sterilized after each annual capture session, antlerless immigration, and the fawn:adult

female ratio.

In VCH, the municipality conducted independent baited camera surveys and population

estimates using NOREMARK from 2013–2016, after capture efforts were completed in 2013–

2014), and prior to culling efforts in 2015–2017 (Curtis et al. 2013, 2014; Curtis and Ashdown

2015, 2016, 2017). Two NOREMARK estimates were made, one using all deer identifiable in

photographs and a second using the maximum number of marked deer potentially available in the

study area. These two estimates were averaged to obtain a final estimate. In 2018, we used a

census through field observations and camera data to estimate the population.

In VGCC, we established a non-overlapping 14.6-km driving transect and surveyed 3–5

times, either during the morning or evening, from 7–8 September 2011 and 9–11 October 2012.

We conducted surveys starting 2 hours before sunset and 1 hour after sunrise. While driving 15–

20 kph in a golf cart, spotters searched their respective side of the road. Upon sighting deer, we

recorded the number in each social group, age, and sex of the individuals, whether deer were

tagged or untagged, and we measured the perpendicular distance the group's center using a

rangefinder. These data were entered into the software program Distance (Version 6.0; Thomas et

al. 2010) to determine a population estimate and a 95% confidence interval.

In VGCC, beginning in fall 2014, 10–12 volunteers were assigned to separate

neighborhoods and recorded observations of every deer they encountered, including tag numbers

and antler patterns. We tallied the number and age class of individual animals observed as an

estimate of the population because the deer were very visible. Observers were active for 2–3

hours before sunset for 5 days.

Given the small area and relatively open landscape at NIH, we walked the available

habitat areas 2–3 hours before sunset and recorded tagged individuals, males based on antler

configuration, and untagged antlerless animals, noting any fawns. We walked the facility twice

during each estimate and drove intensively over the 2-day field operation. We used individual

animals observed as a census of the population because the deer were highly visible.

RESULTS

Capture and Surgical Sterilization

We handled 570 deer, including 493 surgical sterilization procedures, incidental male captures (n

$= 61$), recaptures ($n = 10$), and animals euthanized due to pre-existing conditions ($n = 2$) or

capture related injuries ($n = 4$) at the 6 study sites (Table 1). We experienced 15 capture and

handling related mortalities, 4 were individuals euthanized prior to surgery due to capture related

injuries, and 11 were post-surgical mortalities, resulting in a 2.6% mortality rate. During one

field session at VCH, we captured 29 incidental males under the drop net. Otherwise, incidental

males accounted for 32 captures across all other sites and sessions. We observed 6 failed

surgeries where treated females were later detected with fawns. When surgeries failed, single

fawns were recruited, and in all cases, animals were recaptured and retreated. One deer with a

failed surgery in VCH had an ovary remaining that was extracted upon recapture. Veterinarians

were unable to locate the uterus in one deer in AA, a suspected previous prolapse, and the

individual was later recaptured and successfully sterilized. We documented four surgical failures

in CLIF due to a deficient surgical method that left remnant ovarian tissue. The entire ovarian

capsule was not removed, allowing for regrowth of tissue. At CLIF, we recaptured 2 females to

confirm lactational status and check for failed surgeries, and at VCH, 2 treated females were

incidentally recaptured with the drop net. We treated an average of 84% (range: 31–100%) of the

females in the first year, and 94% (range: 89–100%) in the second year at each project site.

Surgical and Capture Effort and Cost

Effort by field staff in Year 1 averaged 4.9 hours/deer captured ($SD = 3.4$; range: 2.7–10.9) and

veterinary effort averaged 2.5 hours/deer sterilized ($SD = 2.0$; range: 1.3–6.1) (Table 2). Work

effort varied among sites and years. Local volunteers participated at AA, CLIF, FC, and NIH,

and as appropriate, volunteers handled tasks such as pre-baiting, site preparations, transportation,

and surgery when qualified. We could not accurately account for volunteer hours and our time

spent training and providing oversight to volunteers. We used effort per deer and not total

person-hours, which included pre-capture preparations and travel. Annual increases in capture

effort per deer occurred as fewer untreated deer were available in each study area. The total cost

for Year 1, including all projects, was $434,440, with 423 deer captured and 368 female deer

sterilized (Table 3). If FC Year 1 is removed from the analysis because VA DGIF did not permit

bait use, then the average cost per deer sterilized was $1,185.

Population Estimates and Immigration

For all 6 sites combined, we noted a mean reduction in deer abundance of about 25% (range:

16.2%–36.2%) from Year 1 to Year 2 (Table 4, Table 5). Four years after the first treatment at

sites managed with only surgical sterilization (CLIF, FC, NIH, VGCC), we noted a mean total

population reduction of 45% (range: 28%–56%), and a mean reduction of 90% in fawn to adult

female ratio. Overall, we demonstrated annual population declines of 3–36%, with an average

annual reduction of 16%. This does not include the last two years at NIH where very small

changes in abundance with few deer remaining resulted in relatively large percentage swings

(e.g., -8.7%, 44%). We detected 85 antlerless immigrants across all study sites between 2012–

2020, with 52 immigrant individuals detected at VCH. From 2015–2018 in VCH, 130 deer were

culled using archery and remote immobilization equipment resulting in a decline from 137 to 9

deer (Table 6).

DISCUSSION

Improvements in efficacy and longevity of non-surgical wildlife infertility methods have

stagnated, and under some circumstances, surgical sterilization has been shown to be more cost-

effective with longer-term efficacy (Evans et al. 2016). Cost for vaccine treatments exceeded

$700, demonstrating high costs, >20 years ago (Curtis et al. 1998) with minimal improvements in

vaccine administrative methods since that time. We documented that the average price per deer

treated using a surgical approach and bait was $1,185, similar to the amount reported by others

(Boulanger and Curtis 2016, Mathews et al. 2005). Present vaccine treatment options have

changed minimally since early summaries of fertility control technology (Bomford 1992, Warren

1995, Miller et al. 1998). Reduced efficacy to ~50% after one year with single-dose vaccines is

still a significant limitation for both gonadotropin-releasing hormone-based and porcine zona

pellucida-based preparations (Gionfriddo 2011, Rutberg et al. 2013a). Therefore, we concluded

that the permanence and 99% effectiveness of surgical methods (MacLean et al. 2006) were

necessary to assess localized impacts of deer management via fertility control.

We have demonstrated that permanently sterilizing ≥85% of female deer in 6 study sites

resulted in average annual population declines of 16%, with the largest local population decline

occurring between Year 1 and Year 2. This 25% average decline between Year 1 and Year 2 was

the result of mortality, a substantial reduction in fawn recruitment, and dispersal of male fawns as

they aged to yearlings. There were few yearling males present during Year 2 capture efforts,

relative to the abundance of male fawns during Year 1 capture efforts, reflecting that a high

percentage dispersed the spring and fall after the first field session. Given nearly 20% of non-

hunted population are composed of male fawns (DeNicola et al. 2008), this behavior contributed

substantially to the initial population reduction. This level of reduction is contrary to most

previous study projections when using fertility control treatments for managing deer in

geographically open environments (Seagle and Close 1996, Merrill et al. 2006, Boulanger et al.

2012). Other fertility control programs in geographically closed island or fenced environments

have documented similar rates of annual population decline (Naugle et al. 2002, Rutberg and

Naugle 2008, Rutberg et al. 2013b). Of our study sites, only NIH and VGCC had security

fencing, and neither fence prevented deer from entering or exiting. Our tagged and VHF collared

deer at VGCC were often observed outside of the property. We reduced the local deer population

by ~73% over 7 years at VGCC by sterilization alone. The program has successfully met

community's deer population goals, and some residents are now concerned that the population

will be reduced too low (A. DeNicola, personal communication). This is an improbable outcome

based on previous expectations of fertility control methods for managing deer populations.

Nearly all females need to be treated in populations with annual survival rates >90% to achieve a

population decline (Grund 2011). With nearly 90% treatment rates, we obtained about an 8%

population reduction from Year 3 to Year 4 in CLIF, and about 28% over 4 years, where we

documented 2 VHF collared female mortalities out of 17 collared over the first 3 years.

We selected research locations based on the fundamentals that emphasized the need for

extensive access and approachable deer (Garrott 1995, Rudolph et al. 2000). Mobile darting from

a vehicle on public roads and use of bait was critical in these case studies. By using an approach

method that deer were exceptionally accustomed to, we were able to capture nearly all the females

before they fully recognized the threat and began to avoid our capture efforts (Kilpatrick et al.

1997). If we had used vaccines that required repeated treatments and only afforded 90-95%

efficacy, our ability to capture and effectively prevent conception in enough females to achieve

the significant population reductions we documented would have been compromised.

We captured 74% of the total females sterilized in the first year at each study site. As a

result, fawn to adult female ratios diminished rapidly after the first year of treatments,

significantly changing the local herd demographics. As a result of reduced fawn recruitment,

there was an initial increase in the percentage of adult males after Year 1. This change was not

the result of immigration, but a proportional increase as the fawn age class diminished, increasing

the percentages of both adult sex classes. We did not observe an increase in the number of adult

males in any of our study sites. The abundance of adult males remained the same or decreased

over time in all locations. This outcome contrasts with the Cornell University campus tubal

ligation research findings. They observed a nine-fold increase in male abundance due to

immigration, and therefore, a minimal population reduction (Boulanger and Curtis 2016). We

demonstrated a 39% population reduction over 2 years in VCH using surgical sterilization.

At each study site, we had treated at least 89% of the adult females by the second year. The

reduced number of untreated deer in later years also facilitates the rapid transition to local

volunteers conducting field operations, if this is a program objective.

To maintain capture efficiency when we attained high percentage capture rates, we used

VHF collars intentionally placed on matriarchal females to track yet untreated females (Rudolph

et al. 2000). We experienced minimal diminishing returns even after >80% of the females were

treated. The collars allowed us to readily locate untagged females and maintain engagement rates.

These engagement rates contrast with previous modeling projections for suburban fertility control

treatments that depicted increased effort per deer treated as higher percentages are handled

(Rudolph et al. 2000). Radio-collar tracking, used in conjunction with mobile darting from public

roads, and the use of bait were critical to the success of all research projects. The effectiveness of

mobile darting from the road was further demonstrated with all but about 9 deer lethally removed

from VCH in April 2018.

Behavioral changes associated with surgical sterilization treatment via tubal ligation in

Highland Park, Illinois, USA were noted with increased dispersal of treated females and increased

mortality (Gilman et al. 2010). We had many female deer become unaccounted for, both collared

and uncollared, but we could not always determine if they dispersed or died locally because they

were never detected or recovered. We only tracked VHF collared females each year during

subsequent capture sessions. We could not validate all mortalities of collared females, and

therefore, could not differentiate our inability to locate collared female deer due to mortality or

dispersal. Consequently, it was not possible to accurately quantify dispersal rates.

Overall, we experienced 15 mortalities while handling 570 deer, resulting in a 2.6%

mortality rate for capture and post-surgery combined. NIH's relatively high handling mortality

rate resulted from training new veterinarians during the first 3 years and led to extended surgical

times of 75–100 minutes. These surgical efforts were 3–4 times longer than the typical time with

experienced veterinarians. There was 1 post-surgical death at NIH in Year 1, 1 in Year 2, 1 in

Year 3, and none in Years 4 and 5. If we removed the NIH training-related deaths, our mortality

rate was 2.0% (11/525). Acceptable mortality rates are suggested to be <2% for chemical

immobilization of large mammals, so when our capture efforts are combined with a surgical

procedure, we are confident in our results' acceptability (Arnemo et al. 2006).

Although fertility control programs can be costly, there are actual costs from doing

nothing as deer-related impacts increase. In Fairfield County, Connecticut, it was estimated that

each household incurred an average of ~$900/year associated with DVCs, landscape damage,

environmental impacts, tick treatments, and Lyme disease-associated medical costs (Arno and

Viola 2010). Also, the cost of management options is relative, given that lethal methods may be

severely constrained in their effectiveness in the same location because of development density

(AA, VCH, VGCC), social or political resistance (FC, NIH), logistical and technical limits of

archery hunting (CLIF), or legal constraints (VCH). Therefore, in situations where lethal

methods are limited or not an option, fertility control methods can reduce deer densities as low or

lesser than archery hunting (Williams et al. 2013) if deer are approachable by vehicles. We

demonstrated deer reductions at or below the archery threshold of ~17 deer/km^2 in AA (11.6

deer/km^2), FC (2.4 deer/km^2), NIH (11.7 deer/km^2), and VGCC (17.0 deer/km^2) (Williams et al.

2013). Longer-term data will allow us to assess whether culling or hunting in adjoining open

spaces reduces immigration rates at lower densities. In AA, we documented similar population

decline rates in the first few years to the other study sites that lack nearby culling programs. In

some unique locations, when politics permit, the most cost-effective management approach might

be a hybrid program using lethal methods in conjunction with fertility control treatments (Garrott

1995).

MANAGEMENT IMPLICATIONS

There are an increasing number of developed areas where lethal deer management options

cannot meet the stated objectives. Although not a deer management panacea, high-percentage

treatment via ovariectomy can significantly reduce local deer populations, especially in areas with

necessary vehicle access and approachable deer, given most legal and social obstacles are not

present. As with any management technique, the project scale and relative deer abundance will

affect the cost and feasibility of meeting stated objectives. Social attitudes in rural and developed

landscapes align with lethal and nonlethal methods' relative applicability, respectively. It may be

necessary to incorporate lethal methods combined with this nonlethal approach, if possible, when

immigration precludes meeting management objectives, or a shorter management timeline is

desired. Finally, it is important to note that fertility control treatments may be a necessary co-

management technique with hunting or culling programs when local populations cannot be

reduced by >30-40% annually because of legal and development constraints.

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LITERATURE CITED

Arnemo, J. M., P. Ahlqvist, R. Andersen, F. Berntsen, G. Ericsson, J. Odden, S. Brunberg, P.
    Segerström, and J. E. Swenson. 2006. Risk of capture-related mortality in large free-
    ranging mammals: experiences from Scandinavia. Wildlife Biology 12:109-113.

Arno, P. S., and D. Viola. 2010. Economic impacts of deer overpopulation. Department of Health
    Policy and Management, School of Health Sciences and Practice, New York Medical
    College.

Bell, R. L., and T. J. Peterle. 1975. Hormone implants control reproduction in white-tailed deer.
    Wildlife Society Bulletin 3:152–156.

Beringer, J., L. P. Hansen, J. A. Demand, J. Sartwell, M. Wallendorf, and R. Mange. 2002.

Efficacy of translocation to control urban deer in Missouri: costs, efficiency, and outcome.

Wildlife Society Bulletin 30:767–774.

Bomford, M. 1992. A role for fertility control in wildlife management? Bureau of rural resources.

Bulletin No.7. 50pp.

Boone, J. L., and R. G. Wiegert. 1994. Modeling deer herd management: sterilization is a viable

option. Ecological Modeling 72:175–186.

Boulanger, J. R., and P. D. Curtis. 2016. Efficacy of surgical sterilization for managing

overabundant suburban deer. Wildlife Society Bulletin 6:273–282.

Boulanger, J. R., P. D. Curtis, and B. Blossey. 2014. An integrated approach for managing white-

tailed deer in suburban environments: the Cornell University study. Cornell Cooperative

Extension, Department of Natural Resources, Ithaca, New York, USA.

Boulanger, J. R., P. D. Curtis, E. G. Cooch, and A. J. DeNicola. 2012. Sterilization as an

alternative deer control technique: a review. Human-Wildlife Interactions 6:273–282.

Bowden, D. C. 1993. A simple technique for estimating population size. Department of Statistics,

Colorado State University, Fort Collins, USA.

Bowden, D. C., and R. C. Kufeld. 1995. Generalized mark–resight population size estimation

applied to Colorado moose. Journal of Wildlife Management 59:840–851.

Curtis, P. D., and M. L. Ashdown. 2015. Population Analysis for White-tailed Deer in the Village

of Cayuga Heights, New York. Department of Natural Resources Cornell University,

Ithaca, New York. 17 pp.

Curtis, P. D., and M. L. Ashdown. 2016. Population Analysis for White-tailed Deer in the Village

of Cayuga Heights, New York. Department of Natural Resources Cornell University,

Ithaca, New York. 11 pp.

Curtis, P. D., and M. L. Ashdown. 2017. Population Analysis for White-tailed Deer in the Village

of Cayuga Heights, New York. Department of Natural Resources Cornell University,

Ithaca, New York. 11 pp.

Curtis, P. D., M. L. Ashdown, and J. R. Boulanger. 2013. Population Analysis for White-tailed

Deer in the Village of Cayuga Heights, New York. Department of Natural Resources

Cornell University, Ithaca, New York. 17 pp.

Curtis, P. D., M. L. Ashdown, and J. R. Boulanger. 2014. Population Analysis for White-tailed

Deer in the Village of Cayuga Heights, New York. Department of Natural Resources

Cornell University, Ithaca, New York. 11 pp.

Curtis, P. D., B. Bazartseren, P. M. Mattison, and J. R. Boulanger. 2009. Estimating deer

abundance in suburban areas with infrared-triggered cameras. Human–Wildlife

Conflicts 3:116–128.

Curtis, P. D., A. N. Moen, and M. E. Richmond. 1998. When should wildlife fertility control be

applied? Pages 1–4 in P. D. Curtis, editor. Workshop on the status and future of wildlife

fertility control. The Wildlife Society, Buffalo, New York, USA.

DeNicola, A. J., K. C. VerCauteren, P. D. Curtis, S. E. Hygnstrom. 2000. Managing white-tailed

deer in suburban environments: a technical guide. Cornell Cooperative Extension, Ithaca,

NY, USA.

DeNicola, A. J., D. Etter, and T. Almendinger. 2008. Demographics of non-hunted white-

tailed deer populations in suburban areas. Human-Wildlife Conflicts 2:102-109.

Eberhardt, L. L. 1969. Population estimates from recapture frequencies. Journal of Wildlife

Management 33:28–39.

Evans, C. S., A. J. DeNicola, and R. J. Warren. 2016. Immunocontraceptive vaccines or surgical

sterilization for deer? A question of effort and efficacy. Wildlife Society Bulletin 40:593–

598.

Frank, E. S., and S. L. Sajdak. 1993. Sterilization as a method of controlling an urban white-tailed

deer population. American Association of Zoological Parks and Aquariums Regional

Conference Proceedings 1:485–490.

Garrott, R. A. 1995. Effective management of free-ranging ungulate populations using

contraception. Wildlife Society Bulletin 23:445–452.

Gilman, R. T., N. E. Mathews, B. G. Skinner, V. L. Julis, E. S. Frank, and J. Paul-Murphy. 2010.

Effects of maternal status on the movement and mortality of sterilized female white-tailed

deer. Journal of Wildlife Management 74:1484–1491.

Gionfriddo, J. P., A. J. DeNicola, L. A. Miller, and K. A. Fagerstone. 2011. Efficacy of GnRH

immunocontraception of wild white-tailed deer in New Jersey. Wildlife Society Bulletin

35:142–148.

Grund, M. D. 2011. Survival analysis and computer simulations of lethal and contraceptive

management strategies for urban deer. Human-Wildlife Interactions 5:23–31.

Harder, J. D., and T. J. Peterle. 1974. Effect of diethylstilbestrol on reproductive performance of

white-tailed deer. Journal Wildlife Management 38:183–196.

Hone, J. 1992. Rate of increase and fertility control. Journal Applied Ecology 29:695-698.

Jacobson, H. A., J. C. Kroll, R. W. Browning, B. H. Koerth, and M. H. Conway. 1997.

Infrared-triggered cameras for censusing white-tailed deer. Wildlife Society Bulletin

25:547–556.

Kilpatrick, H. J., A. M. LaBonte, K. C. Stafford, III. 2014. The relationship between deer density,

tick abundance, and human cases of Lyme disease in a residential community. Journal of

Medical Entomology 51:777–784.

Kilpatrick, H. J., S. M. Spohr, and A. J. DeNicola. 1997. Darting urban deer: techniques

and technology. Wildlife Society Bulletin 25:542–545.

Kirkpatrick, J. F., I. K. M. Liu, and J. W. Turner, Jr. 1990. Remotely-delivered

immunocontraception in feral horses. Wildlife Society Bulletin 18:326–330.

Kirkpatrick, J. F., and J. W. Turner, Jr. 1985. Chemical fertility control and wildlife management.

Bioscience 35:485–491.

Koerth, B. H., C. D. McKown, and J. C. Kroll. 1997. Infrared-triggered camera versus helicopter

counts of white-tailed deer. Wildlife Society Bulletin 25:557–562.

Liu, I. K. M., M. Bernoco, and M. Feldman. 1989. Contraception in mares heteroimmunized with

pig zonae pellucidae. Journal Reproduction Fertility 85:19–29.

MacLean, R. A., N. E. Mathews, D. M. Grove, E. S. Frank, and J. Paul-Murphy. 2006. Surgical

technique for tubal ligation in white-tailed deer (*Odocoileus virginianus*). Journal of Zoo

and Wildlife Medicine 37:354–360.

Mathews, N. E., J. Paul-Murphy, and E. S. Frank. 2005. Evaluation of a trap-sterilize-release

program for white-tailed deer management in Highland Park, Illinois, 2002-2005. Gaylord

Nelson Institute for Environmental Studies, University of Wisconsin-Madison, Madison,

Wisconsin. 52 pp.

Matschke, G. H. 1977. Fertility control in white-tailed deer by steroid implants. Journal Wildlife

Management 41:731–735.

McCullough, D. R. 1987. Demography and management of wild populations by reproductive

intervention. Contraceptives in Wildlife Conference, 13 - 14 November 1987.

Philadelphia, Pennsylvania, USA. 29 pp.

McShea, W. J., S. L. Monfort, S. Hakim, J. F. Kirkpatrick, I. K. M. Liu, J. W. Turner Jr., L.

Chassy, and L. Munson. 1997. The effect of immunocontraception on the behavior and

reproduction of white-tailed deer. Journal of Wildlife Management 61:560–569.

Merrill, J. A., E. G. Cooch, and P. D. Curtis. 2006. Managing an overabundant deer population by

sterilization: effects of immigration, stochasticity and the capture process. Journal of

Wildlife Management 70:268–277.

Miller, L. A., B. E. Johns, and D. J. Elias. 1998. Immunocontraception as a wildlife management

tool: some perspectives. Wildlife Society Bulletin 26:237-243.

Miller, L. A., B. E. Johns, D. J. Elias, and K. A. Crane. 1997. Comparative efficacy of two

immunocontraceptive vaccines. Vaccine 15:1858-1862.

Miller, L. A., B. E. Johns, and G. J. Killian. 2000. Immunocontraception of white-tailed deer with

GnRH vaccine. American Journal of Reproductive Immunology 44:266–274.

Naugle, R. E., A. T. Rutberg, H. B. Underwood, J. W. Turner, and I. K. M. Liu. 2002. Field

testing of immunocontraception on white-tailed deer (*Odocoileus virginianus*) on Fire

Island National Seashore, New York, USA. Reproduction Supplement 60:143–153.

Patten, M. L., W. Jochle, and L. M. Penfold. 2007. Review of contraception in ungulate species.

Zoo Biology 26:311–326.

Plotka, E. D., and U. S. Seal. 1989. Fertility control in female white-tailed deer. Journal of

Wildlife Diseases 25:643–646.

Porter, W. F., H. B. Underwood, and J. L. Woodard. 2004. Movement behavior, dispersal, and the

potential for localized management of deer in a suburban environment. Journal of Wildlife

Management 68:247–256.

Roelle, J. E., S. S. Germaine, A. J. Kane, and B. S. Cade. 2017. Efficacy of SpayVac as a

596   contraceptive in feral horses. Wildlife Society Bulletin 41:107–115.

597   Rudolph, B. A., W. F. Porter, and H. B. Underwood. 2000. Evaluating immunocontraception for

598   managing suburban white-tailed deer in Irondequoit, New York. Journal of Wildlife

599   Management 64:463–473.

600   Rutberg A. T., and R. E. Naugle. 2008. Population-level effects of immunocontraception in

601   white-tailed deer (*Odocoileus virginianus*). Wildlife Research 35:494–501.

602   Rutberg A. T., R. E. Naugle, J. W. Turner, Jr., M. A. Fraker, and D. R. Flanagan. 2013a. Field

603   testing of single-administration porcine zona pellucida contraceptive vaccines in white-

604   tailed deer (*Odocoileus virginianus*). Wildlife Research 40:281–288.

605   Rutberg A. T., R. E. Naugle, and F. Verret. 2013b. Single-treatment porcine zona pellucida

606   immunocontraception associated with reduction of a population of white-tailed deer

607   (*Odocoileus virginianus*). Journal of Zoo and Wildlife Medicine 44:75–83.

608   Seagle, S. W., and J. D. Close. 1996. Modeling white-tailed deer population control by

609   contraception. Biological Conservation 76:87-91.

610   Thomas, L., S. T. Buckland, E. A. Rexstad, J. L. Laake, S. Strindberg, S. L. Hedley, J. R. Bishop,

611   T. A. Marques, and K. P. Burnham. 2010. Distance software: design and analysis of

612   distance sampling surveys for estimating population size. Journal of Applied Ecology

613   47:5–14.

614   Turner, J. W., I. K. M. Liu, and J. F. Kirkpatrick. 1992. Remotely delivered immunocontraception

615   in captive white-tailed deer. Journal of Wildlife Management 56:154–157.

616   Warren, R. J. 1995. Should wildlife biologists be involved in wildlife contraception research and

617   management? Wildlife Society Bulletin 23:441–444.

White, G. C. 1996. NOREMARK: Population estimation from mark-resighting surveys. Wildlife

Society Bulletin 24:50–52.

Williams, S. C., A. J. DeNicola, T. Almendinger, and J. Maddock. 2013. Evaluation of

organized hunting as a management technique for overabundant white-tailed deer

in suburban landscapes. Wildlife Society Bulletin 37:137–145.

Williams, S. C., A. J. DeNicola, and I. M. Ortega. 2008. Behavioral responses of white-

tailed deer subjected to lethal management. Canadian Journal of Zoology 86:1358–

1366.

Associate Editor:

Figure Captions

Figure 1. Delineation of focal white-tailed deer surgical sterilization study area in Ann Arbor,

Michigan, USA.

Figure 2. Delineation of white-tailed deer surgical sterilization study area in the Clifton area of

Cincinnati, Ohio, USA.

Figure 3. Delineation of white-tailed deer surgical sterilization study area in City of Fairfax,

Virginia, USA.

Figure 4. Delineation of white-tailed deer surgical sterilization study area in National Institutes of

Health, Bethesda, Maryland, USA.

Figure 5. Delineation of white-tailed deer surgical sterilization study area in the Village of

Cayuga Heights, New York, USA.

Figure 6. Delineation of black-tailed deer surgical sterilization study area in the Villages Golf and

Country Club, San Jose, California, USA.

Tables

Table 1. White-tailed deer surgical sterilization study site locations, field implementation dates,

quantity of females sterilized, capture and handling related mortalities, and the associated number

of deer captured during each field session from 2012–2020. AA = Ann Arbor, Michigan, USA;

CLIF = Clifton area, Cincinnati, Ohio, USA; FC = City of Fairfax, Virginia, USA; NIH =

National Institutes of Health, Bethesda, Maryland, USA; VCH = Village of Cayuga Heights, New

York, USA; VGCC = Villages Golf and Country Club, San Jose, California, USA

Study sites and program dates	Females sterilized	Mortality	Captured[a]
AA			
22–29 Jan 2017	49	0	52
2–6 Jan 2018	10	0	14
28–30 Nov 2018	1	0	1
22 Jan–20 Feb 2020	0	0	0
CLIF			
1–7 Dec 2015	41	2	44
16–19 Jan 2017	10	0	10
8–14 Jan 2018	8	0	11
24–27 Aug 2018	4	0	6
24–28 Jan 2019	7	0	9
1 Nov 2019–8 Feb 2020	9	1	10
FC			
31 Jan–6 Feb 2014	18	0	21

26–31 Jan 2015	18	0	20
14–16 Dec 2015	6	1	8
6–7 Dec 2016	5	0	5
16–18 Jan 2018	4	0	4
NIH			
14–16 Dec 2014	24	2	27
2 Dec 2015	5	1	5
3 Dec 2016	8	1	8
4 Dec 2017	2	0	2
8 Dec 2018	3	0	3
VCH			
1–15 Dec 2012	137	3	172
2–6 Dec 2013	11	0	13
26–27 Mar 2016	5	0	8
VGCC			
25 Jan–4 Feb 2013	99	4	107
8–12 Oct 2013	9	0	10
Total	493	15	570

[a]Includes females sterilized, recaptures, incidental male captures, and capture/pre-existing

conditions related euthanasia.

Table 2. Field staff and veterinary effort for 5 surgical sterilization study sites. We did not track

staff time in AA. Hours accrued for veterinarians are summarized as veterinary person-hours/deer

sterilized (VT H/S), and hours accrued for field staff are summarized as field staff person-

hours/deer captured (FS H/C). In CLIF and AA, volunteers participated in pre-baiting, site

preparations, surgery, and transport of animals. Volunteers participated in pre-baiting and site

preparations at FC. Volunteers participated in surgeries at NIH. Volunteer hours are not included,

but overall represent minimal impact given the training and support needed to train volunteers.

CLIF = Clifton area, Cincinnati, Ohio, USA; FC = City of Fairfax, Virginia, USA; NIH =

National Institutes of Health, Bethesda, Maryland, USA; VCH = Village of Cayuga Heights, New

York, USA; VGCC = Villages Golf and Country Club, San Jose, California, USA

Study sites	Year 1		Year 2		Year 3		Year 4		Year 5	
	VT H/S	FS H/C	VT H/S	FS H/C	VT H/S	FS H/C	VT H/S	FS H/C	VT H/S	FS H/C
CLIF	1.6	3.0	3.3	6.6	6.8	9.8	9.1	13.3	NA	15.0
FC	6.1	10.9	6.8	12.3	5.2	7.7	5.0	9.8	5.0	10.1
NIH	2.0	4.3	2.2	4.4	1.5	2.8	2.0	3.0	1.3	2.7
VCH	1.3	3.7	2.2	9.1						
VGCC	1.6	2.7	6.0	9.0						
Mean	2.5	4.9	4.1	8.3	4.5	6.8	5.4	8.7	3.2	9.3

667   Table 3. Cost per white-tailed deer handled and cost per deer sterilized in Year 1 at 6 surgical

668   sterilization study sites. AA = Ann Arbor, Michigan, USA; CLIF = Clifton area, Cincinnati,

669   Ohio, USA; FC = City of Fairfax, Virginia, USA; NIH = National Institutes of Health, Bethesda,

670   Maryland, USA; VCH = Village of Cayuga Heights, New York, USA; VGCC = Villages Golf

671   and Country Club, San Jose, California, USA

Study sites	Program cost	Cost/deer handled	Cost/deer sterilized
AA	$77,036	$1,481	$1,572
CLIF	$38,000	$864	$927
FC[a]	$41,959	$1,998	$2,331
NIH	$33,000	$1,222	$1,375
VCH	$148,945	$866	$1,087
VGCC	$95,500	$893	$965
Mean	-	$1,221	$1,376
Baited Site Mean	-	$1,065	$1,185

[a] Decreased Year 1 efficiency due to no bait use and low initial densities.

Table 4. White-tailed deer population estimates at 3 study sites derived by NOREMARK, BDR,

LPE and Census. The final estimate was an average of the methods. NR-BDR = NOREMARK

estimate for antlerless deer and antlered males found using BDR method; LPE-BDR = LPE

method for antlerless and antlered males found using BDR method; AA = Ann Arbor, Michigan,

USA; CLIF = Clifton area, Cincinnati, Ohio, USA; FC = City of Fairfax, Virginia, USA

	NOREMARK				LPE-	
Survey dates	(95% CI)	BDR M	NR-BDR	BDR	BDR	Census

AA						
4–12 Feb 2017	95 (81–113)	6	101	-	91	90
7–22 Jan 2018	60 (55–65)	11	71	-	69	70
1–13 Dec 2018	43 (40–47)	13	56	-	56	54
22 Jan –20 Feb 2020	39 (38–41)	11	50	-	51	50
CLIF						
8–26 Dec 2015		31	-	102	99	99
19–29 Jan 2017	53 (51–56)	29	82	-	81	83
16–27 Jan 2018	58 (53–63)	23	81	-	81	78
3–24 Feb 2019	64 (57–72)	14	78	-	-	70
13–28 Feb 2020	56 (54–58)	16	72	-	71	72
FC						
20–30 Jan 2014			-	91	-	-
1–11 Feb 2015			-	66	-	-
29 Nov–13 Dec 2015			-	53	-	-
7–20 Dec 2016	34 (32–38)	15	49	-	47	48
17–26 Jan 2018	27 (25–30)	13	40	-	41	40

679
Table 5. White-tailed deer population demographics derived from population estimates and field

operations for 6 surgical sterilization study sites from 2012–2020. BCS = Baited camera survey;

C = Census; DS = Distance sampling; NBCS = Non-baited camera survey; AA = Ann Arbor,

Michigan, USA; CLIF = Clifton area, Cincinnati, Ohio, USA; FC = City of Fairfax, Virginia,

USA; NIH = National Institutes of Health, Bethesda, Maryland, USA; VCH = Village of Cayuga

Heights, New York, USA; VGCC = Villages Golf and Country Club, San Jose, California, USA

Survey dates	Survey type	Population estimate (95% CI)	Deer/km²	Adult F sterilized (%)	Fawn:adult F	Antlered M	Antlerless immigrants
AA							
4–12 Feb 2017	BCS, C	94	21.9	90%	1.1:1	6	–
7–22 Jan 2018[a]	BCS, C	70	16.3	96%	0.2:1	11	1
1–13 Dec 2018[b]	BCS, C	55	12.8	97%	0.1:1	13	1
22 Jan–20 Feb 2020	BCS, C	50	11.6	88%	0.1:1	11	2
CLIF							
8–26 Dec 2015	BCS, C	99	41.3	86%	0.6:1	32	–
19–29 Jan 2017	BCS, C	83	34.6	89%	0.1:1	29	1
16–27 Jan 2018	BCS, C	80	33.3	91%	0.1:1	23	1
3–24 Feb 2019	BCS, C	74	30.8	94%	0.2:1	14	2
13–28 Feb 2020	BCS, C	72	29.9	98%	0.1:1	16	1
FC							
20–30 Jan 2014	NBCS	91	5.5	31%	1.3:1	6	–
1–11 Feb 2015	BCS, C	66	4	91%	0.5:1	9	–
29 Nov–13 Dec 2015	BCS, C	53	3.2	93%	0.2:1	16	0
7–20 Dec 2016	BCS, C	48	2.9	84%	0.1:1	15	1
17–26 Jan 2018	BCS, C	40	2.4	91%	0.2:1	13	0
NIH							
14–16 Dec 2014	C	47	39.2	100%	1.4:1	11	–

12–13 Dec 2015	C	30	25.0	100%	0.1:1	7	5
2–3 Dec 2016	C	24	20.0	100%	0.1:1	3	8
1–2 Dec 2017	C	23	19.2	100%	0:1	5	1
7–8 Dec 2018	C	25	20.8	100%	0.1:1	4	5
21 Nov 2019	C	14	11.7	89%	0:1	4	1
VCH							
4–12 Jan 2013	BCS	225	48.9	99%	0.6:1	-	-
17–24 Jan 2014	BCS	160	34.8	98%	0.03:1	-	3
6–13 Jan 2015	BCS	137	29.8	100%	0.04:1	-	14
18–25 Jan 2016[c]	BCS	68	14.8	100%	0.1:1	-	9
12–19 Dec 2016[c]	BCS	45	9.8	91%	0.02:1	-	7
2–8 Apr 2018[c]	C	9	2.0	71%	0.3:1	-	19
VGCC							
7–8 Sep 2010	DS	80 (48–95)	29.6	-	1.0:1	-	-
9–11 Oct 2012	DS	169 (105–223)	62.6	-	0.9:1	-	-
1–4 Feb 2013[d]	C	140	51.9	100%	0.9:1	40	0
8–12 Oct 2013	C	128	47.4	100%	0.2:1	37	2
5–9 Nov 2014	C	102	37.8	100%	0:1	30	0
20 Sep–4 Oct 2015	C	86	31.9	100%	0:1	23	0
17–26 Oct 2016	C	75	27.8	100%	0:1	18	0
20–28 Aug 2017	C	59	21.9	100%	0:1	10	0
1–17 Oct 2018	C	56	20.7	100%	0:1	6	0

		Total	Adult F	Adult F		Antlered

Cull dates	Cull type	Total culled	Adult F (tagged)	Adult F (untagged)	Fawns	Antlered M
1–31 Aug 2019	C	46	17.0	98%	0.02:1	6	1

[a] Population estimate is pre-cull and includes the 7 adult males culled on nearby properties.

[b] Population estimate is pre-cull and includes the 8 adult males culled on nearby properties.

[c] Population estimate is post-cull efforts.

[d] Population estimate does not include 30 deer relocated to outside of the fenced area. Fawn:adult female ratio based on initial population of ~169 deer.

Table 6. Annual white-tailed deer cull data segregated by tag status, sex, and age at Village of Cayuga Heights, New York, USA, from 2015 to 2018.

Cull dates	Cull type	Total culled	Adult F (tagged)	Adult F (untagged)	Fawns	Antlered M
6–14 Mar 2015	Archery	48	26	10	5	7
4–25 Mar 2016	Archery	24	13	5	4	2
2–23 Mar 2017	Archery	13	8	3	1	1
2–8 Apr 2018	Dart, Euthanasia	45	18	15	9	3