

1 **Combinatorial CRISPR screening reveals functional buffering** 2 **in autophagy**

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24 **ABSTRACT**

25 Functional genomics studies in model organisms and human cell lines provided important insights
26 into gene functions and their context-dependent role in genetic circuits. However, our functional
27 understanding of many of these genes and how they combinatorically regulate key biological
28 processes, remains limited. To enable the SpCas9-dependent mapping of gene-gene interactions
29 in human cells, we established 3Cs multiplexing for the generation of combinatorial gRNA libraries
30 in a distribution-unbiased manner and demonstrate its robust performance. The optimal number
31 for combinatorial hit calling was 16 gRNA pairs and the skew of a library's distribution was
32 identified as a critical parameter dictating experimental scale and data quality. Our approach
33 enabled us to investigate 247,032 gRNA-pairs targeting 12,736 gene-interactions in human
34 autophagy. We identified novel genes essential for autophagy and provide experimental evidence
35 that gene-associated categories of phenotypic strengths exist in autophagy. Furthermore, circuits
36 of autophagy gene interactions reveal redundant nodes driven by paralog genes. Our
37 combinatorial 3Cs approach is broadly suitable to investigate unexpected gene-interaction
38 phenotypes in unperturbed and diseased cell contexts.

39

40 **INTRODUCTION**

41 Combinatorial gRNA expression (gRNA multiplexing) for related or orthogonal CRISPR
42 applications enable the comprehensive characterization of genetic interactions in human cells.
43 Several methods are available to support the generation of combinatorial gRNA expression
44 systems, 1) restriction enzyme-based¹⁻⁴, 2) golden-gate assembly⁵⁻⁸, 3) gateway-dependent^{9,10},
45 as well as 4) recombination-dependent gRNA cloning techniques¹¹. These systems are widely
46 used to clone gRNA sequences in combination with RNA polymerase III promoters, resulting in
47 arrayed gRNA-expression cassettes. In contrast, RNA polymerase II promoters generate RNA
48 transcripts that can contain multiple gRNA sequences, although these transcripts require post-
49 transcriptional processing to yield functional gRNA sequences and are currently limited to Cas12
50 applications^{12,13}. The most widely used Cas-nuclease thus far is SpCas9, though a cloning-free
51 gRNA multiplexing concept for Cas9 gRNAs is currently lacking, because repetitive and
52 homologous sequences are unstable in lentiviral vectors¹⁴⁻¹⁶, rendering them less suited for large-
53 scale combinatorial screening.

54 Concomitant mutations in two genes can yield unexpected phenotypes with respect to each
55 gene's individual phenotype¹⁷. Synthetic lethality as the most extreme combinatorial phenotype
56 has clinical applications and is under therapeutic exploration¹⁸⁻²⁰. A prominent example of a

57 synthetic lethal gene pair with clinical relevance is poly (ADP-ribose) polymerase (PARP)
58 inhibition in the context of defective *BRCA1* or *BRCA2* genes^{15,21,22}. Additionally to this DNA-
59 damage repair-related example, synthetic lethal interactions have been identified in combinatorial
60 gRNA CRISPR screens, including the apoptotic genes *BCL2L1* and *MCL1* or *BCL2L1* and
61 *BCL2L2*^{16,23,24}, the mitogen-activated protein kinases 1 and 3 (*MAPK1* and *MAPK3*)¹, as well as the
62 PIP₃ phosphatase *PTEN* and the mammalian target of rapamycin *MTOR*^{25,26}. To perform pairwise
63 hit calling in CRISPR screens, two methods are currently established: 1) a variational Bayes
64 approach (GEMINI)²⁷, and 2) the difference of expected to measured log₂-fold-changes (dLFC) in
65 which the expected log₂-fold-change of a gRNA combination is the sum of the log₂-fold-changes
66 of each individual gRNA when partnered with control gRNAs^{16,24}. These computational
67 approaches have been applied to negative CRISPR screens. However, with combinatorial gRNA
68 CRISPR screens being mostly performed in drop-out conditions, we lack knowledge of their
69 performance for Cas9-based combinatorial gRNAs in positive or FACS-based phenotypic
70 enrichment screens.

71 Bulk and selective autophagy are tightly regulated processes that target cellular material for
72 lysosomal degradation and their misregulation culminates in abnormal cell growth and cell death
73 with implications in various human diseases^{28,29}. Rationally-engineered fluorescent reporter
74 systems in combination with high-throughput CRISPR screens facilitated the systematic
75 categorization of genes based on their essentiality in autophagy. As such, the transmembrane
76 protein 41B, as well as the ubiquitin-activating enzyme UBA6, and the hybrid ubiquitin-conjugating
77 enzyme/ubiquitin ligase BIRC6 were recently identified as important players in the autophagy
78 network³⁰⁻³⁵. Despite the mapping of essential genes, CRISPR screens paired with fluorescent
79 autophagic reporters revealed the stress-dependent regulation of autophagy-related genes³⁶⁻³⁸,
80 and provided mechanistic insights into the gene specificity in bulk and selective autophagy by
81 identifying PARKIN regulators and the ANT complex as essential components for mitophagy^{39,40}.
82 In combination with proximity biotinylation-coupled mass spectrometry, CRISPR screens also
83 contributed to a spatial proteogenomic understanding of PARK2-dependent mitophagy⁴¹. Thus,
84 unbiased CRISPR approaches coupled to fluorescent reporters and mass spectrometry are
85 valuable approaches to identify vulnerabilities in bulk and selective autophagy for the treatment
86 of neurodegenerative diseases and cancer⁴²⁻⁴⁴.

87 Here, we add to our previous work and describe the SpCas9-based 3Cs multiplexing technology
88 for the generation of dual combinatorial (multiplexed) gRNA libraries with diversities of up to
89 several hundreds of thousands of gRNA combinations⁴⁵. In addition to identifying critical technical
90 parameters, we generated a combinatorial library targeting the human autophagy network and

91 performed a fluorescent reporter-based FACS enrichment screen to identify hitherto
92 uncharacterized single genes and gene interactions essential for autophagy. Our 3Cs multiplexing
93 technology is widely applicable and can serve as a general tool for the identification of context-
94 dependent gene interactions at any scale.

95

96 **RESULTS**

97 **3Cs multiplexed Cas9 gRNA libraries**

98 To expand on our 3Cs technology and enable Cas9 gRNA multiplexing, we generated a lentiviral
99 Cas9-gRNA expression plasmid (pLenti-Multiplex) by placing a human 7SK (h7SK) promoter
100 upstream of a previously engineered Cas9-tracrRNA followed by a human U6 (hU6) promoter
101 upstream of a wildtype Cas9-tracrRNA sequence^{46,47}. Both gRNA cassettes contain a gRNA
102 placeholder sequence encoding for I-CeuI or I-SceI restriction enzyme sites, respectively (Figure
103 1A)⁴⁵. Furthermore, in addition to a puromycin selection cassette, the plasmid contains an f1
104 bacteriophage origin of replication sequence in sense direction, supporting the CJ236 bacteria
105 and M13KO7 bacteriophage-dependent generation of dU-containing single stranded (ss) DNA
106 (Figure 1A, Supp. Figure 1A). In contrast to single gRNA 3Cs reactions, 3Cs multiplexing is
107 performed in the presence of two gRNA-encoding oligonucleotide pools of which each contains
108 unique 5' and 3' homology sequences for annealing to the h7SK or hU6 gRNA cassettes, thereby
109 generating hetero-duplex dU-containing double stranded (ds) DNA that contains all possible
110 gRNA combinations of the two oligo pools (Figure 1A). A coverage-based electroporation into
111 *dut/ung*-positive bacteria results in template-strand depletion and combinatorial gRNA-containing
112 dsDNA. To test this concept, we designed two gRNA-encoding oligo pools each containing 50
113 gRNA sequences targeting GFP (pool-1) or mCherry (pool-2) and used them to generate a
114 combinatorial gRNA library, in addition to the respective single gRNA libraries (Figure 1B). Based
115 on the typical three-band pattern of 3Cs dsDNA^{45,48}, we observed similar yields and quality of dU-
116 containing hetero-duplex dsDNA after applying two pools in a single 3Cs reaction when compared
117 to either pool alone (Supp. Figure 1A). Digesting the final libraries with I-CeuI and I-SceI enzymes
118 confirmed the exclusive presence of gRNA-containing plasmids (Figure 1C). Paired-end next-
119 generation sequencing (NGS) revealed all three libraries to be complete with distribution skews
120 of 1.56, 1.17, and 1.53 for GFP, mCherry, and GFP+mCherry, respectively (Figure 1D, Supp.
121 Figure 1B-C). Most importantly, when packaged into lentiviral particles and transduced into GFP
122 and mCherry-expressing hTERT-RPE1(Cas9) cells, the combinatorial GFP+mCherry library
123 induced the simultaneous depletion of GFP and mCherry fluorescence while both single libraries

124 selectively depleted either GFP or mCherry fluorescence (Figure 1E). Thus, 3Cs multiplexing
125 generates tightly distributed combinatorial gRNA libraries that are functional in human cells.

126

127 **3Cs multiplexing decouples sequence distribution and diversity**

128 Current combinatorial gRNA libraries contain 9 to 18 gRNA pairs per gene-pair^{3,15,24}, thus, a robust
129 technology must generate large pairwise gRNA diversities without compromising reagent quality.
130 We therefore investigated whether 3Cs multiplexing, similar to single gRNA 3Cs⁴⁵, would
131 decouple library quality from library size. To this end, we designed four oligonucleotide pools per
132 gRNA cassette in which one, two, three, or four nucleotide positions within a non-human-target
133 (NHT) gRNA sequence were randomized to mimic increasing combinatorial gRNA diversities (1N,
134 4*4=16 combinations; 2N, 16*16=256 combinations; 3N, 64*64=4,096, 4N, 256*256=65,536
135 combinations). NGS confirmed the libraries to be complete and evenly distributed with areas
136 under the curve (AUC) between 0.59 and 0.69 (Figure 2A, Supp. Figure 2A-B). Notably, we
137 identified the distribution skew to be very narrow with ranges from 1.1 (1N) to 1.49 (4N), values
138 mostly unmatched even with single gRNA libraries (Supp. Figure 2A-B). Thus, 3Cs multiplexing
139 is a highly robust method to generate combinatorial gRNA libraries with large sequence
140 diversities.

141

142 **Library distribution skew influences experimental scale**

143 Higher replicate correlation was computationally demonstrated to correlate with smaller library
144 distribution skew and higher library representation⁴⁹. Thus, we aimed at experimentally
145 investigating to what extent a combinatorial gRNA library's distribution skew contributes to hit
146 calling accuracy by generating combinatorial libraries with artificially distorted gRNA
147 representations and screening them in different coverages. We selected a panel of 20 genes
148 equally divided in 10 core essential (CE) and 10 tumor-suppressor genes (TSGs) and designed
149 4 gRNAs per gene. As internal controls, we choose 80 pre-validated NHT sequences^{50,51}. We
150 designed two 3Cs oligo pools: pool-1 contained CE and TSG-targeting gRNAs for h7SK and hU6
151 cassettes in equal ratios (2*80 gRNAs), while pool-2 consisted exclusively of NHT gRNA
152 sequences in equal ratios for h7SK and hU6 cassettes (2*80 gRNAs). To generate artificially
153 distorted library skews, we mixed both pools in equimolar ratios (1:1), and in ratios of increasing
154 NHT sequence-molarity (1:10 and 1:100) and applied them to 3Cs multiplexing. NGS confirmed
155 an increased fraction of NHT reads in the 1:10 and 1:100 libraries (Figure 2B, Supp. Figure 2C),
156 and revealed an AUC value for the 1:1 library of 0.65, while the 1:10 and 1:100 libraries contained
157 increased AUC values of 0.85 and 0.9, respectively (Figure 2C). Most importantly, library

158 distribution skews increased from 1.2 (1:1) to 2.4 (1:10) and 13.46 (1:100) (Supp. Figure 2D),
159 demonstrating that final 3Cs library quality is coupled to an oligo pool's distribution.

160 Using these libraries in representations of 20- and 200-fold, we evaluated a total of 153,600
161 pairwise gRNA-knockouts in human hTERT-RPE1(Cas9) cells in biological replicates. Pairwise
162 guide and gene-level counts correlated well for individual libraries and biological replicates (gene
163 level, 20x - 1:1, Pearson $r = 0.96$; 1:10, Pearson $r = 1$; 1:100, Pearson $r = 0.79-1$; 200x - 1:1,
164 Pearson $r = 0.91-0.98$; 1:10, Pearson $r = 0.96-0.97$; 1:100, Pearson $r = 0.98-1$; Supp. Figure 3A-
165 B). We next analyzed the difference of \log_2 fold changes (\log_2FC) in our screens by means of
166 separating essential genes from non-targeting controls and computed Cohen's D statistics (quality
167 score (QS)), a value recently introduced to quantify screen quality⁵². Quality scores for the 1:1
168 library in both tested coverages were very high with 2.95 and 3.25 for 20- and 200-fold,
169 respectively (Figure 2D). However, a decline in screen quality appeared when library distribution
170 skews increased above 2 (1:10 library, 20x - QS: 1.5, 200x - QS: 2.68; 1:100 library, 20x - -0.44,
171 200x - -1.17; Figure 2D).

172 We next assessed if higher experimental coverage was able to rescue wider library distribution
173 skews by identifying essential gene pairs with MAGeCK analyses and cut-off filters set to
174 $FDR \leq 10\%$ and $\log_2FC < -0.5$. Strikingly, the number of retrieved gene pairs per library distribution
175 skew was nearly identical between 20- and 200-fold coverages (Figure 2E), suggesting that
176 CRISPR libraries with narrow distribution skews can be screened at minimal experimental
177 coverage without compromising data quality. Furthermore, these analyses also suggest that
178 higher experimental coverage is likely insufficient to rescue wider library distribution skews.
179 Therefore, a library's distribution skew is a critical parameter that should be considered when
180 designing combinatorial CRISPR libraries and screens.

181

182 **The optimal number of pairwise gRNAs for statistical gene-interaction calling**

183 The number of gRNAs per gene has a profound impact on statistical hit calling^{53,54}, thus we
184 analyzed the concordance of essential pairwise genes identified across the two experimental
185 coverages with increasing numbers of pairwise gRNAs. We down-sampled the 1:1 data sets and
186 generated a series of read count tables containing between 1 and 16 randomly chosen gRNA
187 pairs for each gene combination and performed MAGeCK analyses with cut-off filters at
188 $FDR \leq 10\%$ and $\log_2FC \leq -0.5$ ^{53,54}. A total of 100 essential gene interactions exist within both
189 libraries (10*10 essential genes), and as expected, 16 pairwise gRNAs per gene interaction
190 consistently retrieved the largest number of statistically significant essential gene interactions
191 (200x - 70%, 20x - 72%) (Figure 2F). However, performance differences became evident in the

192 down-sampled libraries containing up to 4 pairwise gRNAs with 13% and 20% paired gene
193 interactions identified, for 200 and 20-fold library representations, respectively (Figure 2F).
194 Strikingly, the performance increased noticeably in both coverages up until 13 pairwise gRNAs
195 from which on the performance plateaued (Figure 2F). These results are consistent with previous
196 observations for single gRNA SpCas9-dependent CRISPR screens, in which 4 to 6 gRNAs per
197 gene have been identified for robust statistical hit calling⁵⁵. Assuming the generation of gRNA
198 number-balanced libraries, our observations identify 16 pairwise gRNAs (4*4) as the optimal
199 number for statistical gene-interaction calling.

200

201 **Single and inherent-single gRNA enrichment screens are complementary**

202 Pairwise gRNA screens have been applied to phenotypes related to cell viability (drop-outs), but
203 their performance in enrichment-based screens remains elusive. To address this question, we
204 identified parameters to investigate pairwise gene interactions in the self-degrading process of
205 human autophagy by LC3-coupled fluorescent-dependent enrichment screening. We used the
206 established GFP-LC3-RFP reporter construct to generate a monoclonal hTERT-RPE1 reporter
207 cell line and confirmed reporter functionality by mTOR inhibition (Torin1 treatment) alone and in
208 combination with the autophagy inhibitor Bafilomycin (Supp. Figure 4A)⁵⁶. Next, we assembled a
209 literature-curated list of 192 “extended autophagy” genes that included, among others, autophagy-
210 related genes (ATGs), autophagy receptors, as well as transcription factors and deubiquitinases,
211 plus 7 core essential genes as negative controls. To generate a combinatorial gRNA library, we
212 designed a second oligo pool, named “core autophagy”, consisting of 64 commonly considered
213 core autophagy genes (Figure 3A, core autophagy, Supp. Table 1). For each gene, we designed
214 4 gRNA sequences using rule set 2 and added 10% NHT sequences to each pool⁵¹, resulting in
215 a total of 876 gRNAs in the “extended autophagy” pool and 282 gRNAs in the “core autophagy”
216 pool, together generating 247,032 pairwise gRNAs (Figure 3A). We applied the extended
217 autophagy pool alone and in combination with the core autophagy pool to 3Cs reactions and
218 generated the single gRNA and pairwise gRNA libraries, respectively. NGS revealed narrow
219 distribution skews (single library, skew ratio=1.22; multiplex library, skew ratio=1.69) and AUC
220 values of 0.65 and 0.68 (Supp. Figure 4B-D), supporting our previous observation that 3Cs
221 multiplexing is a highly robust method for the generation of combinatorial gRNA libraries.

222 To benchmark our screening approach, we first applied the single autophagy library to screen for
223 genes essential for Torin1-induced autophagy (Figure 3B). In order to reduce the number of false-
224 positive hits, we chose stringent FACS-gating criteria that only enriched cells in which autophagy
225 was completely blocked (Figure 3C, single screen). Importantly, proliferative effects as a

226 consequence of gene knockouts are a source of false-positive hit calling in enrichment screens,
227 especially in TP53-positive cells. Thus, we collected a coverage-based cell sample at the day of
228 FACS sorting to correct for positive or negative proliferative effects (Figure 3B). Guide RNA and
229 gene level correlations between pre- and post-FACS samples were high among biological
230 replicates (guide level, Pearson $r=0.87$; gene level, Pearson $r=0.97$; Supp. Figure 5A-B), thus we
231 applied MAGeCK and retrieved 12 significantly enriched genes with \log_2FC between 3.76 and
232 9.12 with $FDR \leq 10\%$ (Supp. Table 2). Importantly, our benchmark screen successfully retrieved
233 previously identified core autophagy genes required for mTOR-induced autophagy and correctly
234 assigned their function to bulk autophagy by means of a positive \log_2FC between pre- and post-
235 FACS conditions (Figure 3D)^{32,33,35,39,40,57,58}.

236 Next, we dramatically scaled up to perform the autophagy multiplex screen at a 20-fold coverage
237 at pre- and post-FACS time points, investigating a total of 1,235,160 pairwise gRNAs. Guide RNA
238 and gene level correlations between pre- and post-FACS samples were high among biological
239 triplicates: gRNA level, pre-FACS, Pearson $r=0.99$; post-FACS, Pearson $r=0.58-0.69$; gene level,
240 pre-FACS, Pearson $r=0.99$; post-FACS, Pearson $r=0.77-0.84$ (Supp. Figure 5A, C-D). During the
241 course of FACS sorting, we noticed a population of cells that was absent from single gRNA
242 transduced cells and extended our sorting strategy to also investigate cells from this “high-gate”
243 (Figure 3C). We first assessed the nature of the high-gate cell population and identified the
244 majority of reads to account for ATG4B-dependent gRNA pairs (post-FACS 1, 86.94%; post-
245 FACS 2, 78.03%; post-FACS 3, 75.53%) (Supp. Figure 5E). To permit its function, ATG4 cleaves
246 the LC3-reporter and paired ATG4 gene knockouts likely interfere with reporter functionality and
247 served as additional controls to validate our library and screening strategy⁵⁶.

248 Within the combinatorial gRNA library, four sets of gRNA interactions occur, 1) control with control
249 (NHT:NHT), 2) extended autophagy with control (gene:NHT), 3) control with core autophagy
250 (NHT:gene), and 4) extended autophagy with core autophagy (gene:gene) (Figure 3A). Thus, we
251 next assessed the concordance of identified single genes essential for autophagy induction
252 between our single autophagy benchmark screen and the extended autophagy gRNAs paired
253 with control guides in the combinatorial screen. We applied MAGeCK to compare pre- and post-
254 FACS time points and identified 10 significantly enriched genes with \log_2FC between 1.53 and
255 2.75 and $FDR \leq 10\%$ (Figure 3E). Screen concordance was high with 7 jointly identified genes,
256 though each experiment successfully identified unique genes essential for bulk autophagy
257 induction (Figure 3D-E, unique hits in blue-bold). Importantly, the jointly identified genes ATG9A,
258 RB1CC1, ATG12, ATG7, ATG3, and ATG16L1 as well as the unique hit genes ATG101, ATG10,
259 ATG13, ATG5, ATG14 and TMEM41B could be successfully validated by targeting each gene

260 with a newly designed single gRNA, for which gRNA performance during the course of validation
261 was quantified by TIDE analysis (Figure 3F). Screen concordance could be further improved to
262 11 jointly identified genes by filtering for p-values below 0.05 and simultaneously relaxing the
263 FDR, although the number of uniquely identified genes per screen increased accordingly to 8 and
264 7 for dedicated single and inherent single screens, respectively (Supp. Figure 6A-B). This
265 demonstrates that dedicated single and inherent-single autophagy screens are complementary
266 and that either one alone was insufficient to identify all true-positive hits.

267

268 **Synergistic gene pairs essential for autophagy**

269 Targeting single genes with two gRNAs has been shown to improve knockout rates⁵⁹, we
270 therefore investigated the consistency of this phenotype within our combinatorial data set. As
271 expected, we repeatedly observed a higher log₂FC for combinatorial-gRNA targeted genes
272 (Supp. Figure 7A), particularly for genes that we identified and validated as essential for
273 autophagy (Supp. Figure 7B). Next, we assessed pairwise gene interactions essential for
274 autophagy induction by applying MAGeCK to pre- and post-FACS samples. This identified a total
275 of 3645 significant gene pairs, of which 187 gene pairs accounted for pairs in which both genes
276 were jointly or uniquely identified as single essential for autophagy (Figure 4A, D). 3245 gene
277 pairs accounted for gene pairs in which one or the other gene was identified as being essential
278 for autophagy (Figure 4B, D), and 213 gene pairs consisted of genes for which neither partner
279 gene was identified as being essential for autophagy (Figure 4C, D). Interactions between the
280 majority of identified gene pairs are driven by a single gene that is essential for autophagy and
281 interactions between non-essential autophagy genes are rare, which is well in agreement with
282 previous work that mapped genetic interactions in *Saccharomyces cerevisiae*⁶⁰. To corroborate
283 our findings, we set up arrayed dual gene-targeting validations in which either one gene or both
284 genes were essential for autophagy induction. As expected, targeting two essential autophagy
285 genes consistently increased the fraction of cells in which autophagy was blocked (Figure 4E),
286 and pairing an essential autophagy gene with an autophagy-related but non-essential gene also
287 increased the fraction of cells in which autophagy was blocked (Figure 4F). Interestingly, among
288 the non-essential:non-essential hits, we identified ULK1, AMBRA1, WIPI2, and BECN1 in
289 combinations with control gRNAs, suggesting these genes to be essential for autophagy on their
290 own, a conclusion supported by the literature^{33,57}. They have likely missed our attention in the
291 single gRNA CRISPR screens due to their relatively mild phenotype compared to the other
292 autophagy essential genes. Indeed, in arrayed single gRNA validations, the depletion of ULK1
293 and WIPI2 alone blocked autophagy, although the phenotype was milder when compared to

294 ATG9A depletion that we identified as a strong hit (Figure 4G). This suggests, in the context of
295 hTERT-RPE1 cells and Torin1-induced autophagy, that gene-associated categories of
296 phenotypic strengths exist in autophagy. Most interestingly, among others, we identified several
297 gene pairs capable to block autophagy induction (Figure 4C). We noted several gene pairs in
298 which one gene is either of the Ras-related protein family (RABs) as well as several pairs that
299 included ATG2A. Among these interactions we identified ATG2A-ATG2B for which the validation
300 in arrayed dual-gene knockout conditions confirmed the blockage of autophagy only when both
301 genes are interfered with, experimentally verifying these genes as functional redundant
302 homologues (Figure 4G).

303 Four mathematical definitions of genetic interactions have been proposed previously (Product
304 (MULT), Additive (SUM), Log (LOG), and Min (MIN))¹⁷. We also included a Max (MAX) definition
305 and applied all models to identify combinatorial phenotypes that are surprising with respect to
306 each gene's single phenotype. Genetic interactions are rare⁶⁰, we thus computed the deviation of
307 observed paired-knockout phenotypes from their expectation (delta log₂FC, $\Delta\log_2FC$) and
308 identified the MAX model to fit our combinatorial autophagy data set best (Figure 4H), especially
309 when only considering genes that we identified to be essential for autophagy for which the
310 deviation from the expectation is low (Supp. Figure 7C). In total, we identified 3665 genetic
311 interactions that we filtered by significance ($p \leq 0.05$) and $\Delta\log_2FC$ s above their single standard
312 deviation, resulting in a total of 57 (1.6%) high confidence synergistic gene pairs (Figure 4I, yellow
313 dots, Supp. Table 3). To better depict relations among these gene interactions, we performed a
314 network analysis and identified 5 network components (1, 36 nodes; 2, 10 nodes; 3, 5 nodes; 4,
315 3 nodes; and 5, 4 x 2 nodes) (Figure 4J). Most importantly, we identified functional redundancy in
316 autophagosome assembly (ATG2A/B), phosphatidylinositol phosphorylation (PIK3CA/PI4K2A),
317 phagosome-lysosome fusion (RAB7A/5A and RAB7B/5A), and selective autophagy (PARK2 with
318 PLEKHM1 and CALCOCO2, OPTN with PINK1, and PARK7 with FAM134B) (Figure 4J).
319 Furthermore, we identified DNA Damage-Regulated Autophagy Modulator Protein 1 (DRAM1)-
320 dependent interactions with mitochondrial PINK1 and PARK2 for which the DRAM1-dependent
321 induction of autophagy was shown to be dependent on mitochondrial protein synthesis
322 inhibition⁶¹. Thus, our identified gene interactions likely resemble functional nodes within the
323 human autophagy circuit in which redundancy exists and point towards gene paralogs as a critical
324 factor in generating redundancy in autophagy.

325

326 **DISCUSSION**

327 Several technologies are available to generate combinatorial gRNA-containing plasmids and
328 libraries. They mostly depend on open plasmid DNA and PCR amplified gRNA-encoding
329 oligonucleotide pools, resulting in cloning-artefacts and sequence biases. Our newly developed
330 3Cs multiplexing technology functions with single stranded template plasmids and oligonucleotide
331 pools, thereby circumventing additional and unintended sequence representation dispersions.
332 Passaging coverage and CRISPR library distribution skew have recently been computationally
333 predicted to be critical factors for data quality⁴⁹. Indeed, our experimental analysis of combinatorial
334 libraries with varying distribution skews, applied with different passaging coverages, supports this
335 prediction and identifies a distribution skew of below 2 as a threshold enabling passaging
336 coverages below 100-fold. Furthermore, we provide experimental evidence that combinatorial
337 libraries with distribution skews above 2 should be screened with passaging coverages ≥ 200 to
338 compensate for the library's sequence dispersion. However, rescuing libraries with large
339 distribution skews (≥ 10) by applying high passaging coverage is likely going to fail, as the
340 sequence dispersion is too big to be compensated for due to sequence underrepresentation at
341 screen initiation.

342 Combinatorial gRNA screens are more frequently performed to identify gene pairs essential for
343 cell viability. However, as of today, no combinatorial gRNA screen in combination with pathway-
344 specific fluorescence reporters has been performed. While the inherent single gRNA phenotypes
345 appear to be sufficient in the context of cell viability screenings, we show that dedicated and
346 inherent single gRNA screens are complementary for combinatorial enrichment screens. This is
347 likely due to the strong enrichment of combinatorial gene phenotypes and the associated
348 underrepresentation of single-gene effects. However, this can be compensated for by including a
349 dedicated single gene screen when planning combinatorial enrichment screens. Furthermore, we
350 note that genes associated with strong phenotypes are sufficiently identified by this approach, but
351 genes causing milder phenotypes will likely miss attention. This obstacle, however, is
352 circumvented by carefully examining gene-interactions in the class of phenotype-associated non-
353 essential gene pairs. Indeed, we demonstrate that AMBRA1, ULK1, WIPI2, and BECN1 are
354 essential for autophagy, even though both single gRNA screens failed to identify them.

355 Genome-wide or combinatorial CRISPR screens demand large numbers of cells. Thus, current
356 efforts aim at minimizing cell culture demands by providing combinatorial-gRNA minimized
357 CRISPR libraries^{59,62}. Supporting this notion, we identify log₂FC of dual-gRNA targeted genes to
358 be larger than their single-gRNA targeted counterpart. Interestingly, this observation was limited
359 to genes that we identified to be essential for autophagy, supporting the notion that dual-gRNA

360 gene targeting also induces stronger phenotypes in enrichment screens and that minimized
361 libraries will also be beneficial for these applications.

362 Functional buffering by paralogs has recently been shown to be largely absent from single gRNA
363 CRISPR screens⁶³, suggesting paralogs to contribute to network redundancy. Indeed, we identify
364 paralogs as functional buffers in autophagy acting in autophagosome assembly (ATG2A/B),
365 phosphatidylinositol phosphorylation (PIK3CA/PI4K2A), and phagosome-lysosome fusion
366 (RAB7A/5A and RAB7B/5A). This is profoundly important when therapeutically targeting
367 autophagy induction in cells that are dependent on high basal autophagy levels such as acute
368 myeloid leukemia (AML) cells⁶⁴. Lastly, we note that our analysis failed to identify functional
369 buffering within the mammalian ATG8 family of proteins (LC3s and GABARAPs), as well as within
370 the protein class of autophagy receptors (FUNDC1, SQSTM1, OPTN, PLEKHM1, PEX13,
371 CALCOCO2, FAM134B). Furthermore, the lack of buffering gene interactions between ATG8s
372 and autophagy receptor proteins, together, supports the notion of their cargo selectivity that
373 prevents these genes to contribute to autophagy redundancy⁶⁵.

374

375 **MATERIAL AND METHODS**

376 **3Cs multiplex template plasmid DNA and cloning**

377 pLentiCRISPRv2 (Addgene: 98290) was enzymatically digested with AelI and BsiWI and gel
378 purified to remove the hU6 gRNA- and SpCas9-expressing cassettes. Likewise, the combinatorial
379 gRNA-expressing cassette of pKLV2.2 (Addgene: 72666) was digested with AelI and BsiWI. The
380 2030 bp fragment that encodes the combinatorial gRNA-expressing cassettes and a PGK
381 promoter was gel purified and cloned into the reduced, purified backbone of pLentiCRISPRv2. In
382 order to generate unique annealing homology for the 3Cs oligonucleotides and enable template
383 plasmid removal, the h7SK promoter-associated tracrRNA was replaced by a previously
384 engineered tracrRNA sequence (tracrRNA v2) and h7SK and hU6 promoter-associated gRNA
385 cloning sites were modified to contain placeholder sequences encoding for I-CeuI and I-SceI
386 homing endonuclease restriction sites, respectively⁶⁶.

387

388 **3Cs oligonucleotide design rules**

389 All oligonucleotides that were used for multiplexed 3Cs gRNA library generation are listed in 'DNA
390 oligonucleotides'. DNA oligonucleotides were purchased from Sigma-Aldrich, from Integrated
391 DNA Technologies (IDT) as single or pooled oligonucleotides in o-pool formats, and from Twist
392 Bioscience as oligonucleotide pools.

393 To discriminate between h7SK and hU6 and enable exclusive annealing to only one expression
394 cassette, the 3Cs oligonucleotides were designed with two specific homology regions flanking the
395 intended 20-nt gRNA sequence for either the h7SK or hU6 expression cassettes. The 3Cs h7SK-
396 oligonucleotides were 57 nts in length (T_m above 50°C) and matched the 3' end of the h7SK
397 promoter region and the 5' start of the tracrRNA v2, while the 3Cs hU6-oligonucleotides were 59
398 nts in length (T_m above 50°C) and matched the 3' end of the hU6 promoter region and the 5' start
399 of the tracrRNA v1 in the template plasmids.

400

401 **Generation of sequence distorted 3Cs libraries**

402 For the generation of biased multiplex 3Cs gRNA libraries, two 3Cs oligonucleotide pools were
403 designed for each expression cassette of the 3Cs multiplex template plasmid following the 3Cs
404 oligonucleotide design rules. The first pool was composed of tumor suppressor and essential
405 gene-targeting gRNAs (target pool), while the second pool only included non-human targeting
406 gRNAs (control pool). To generate three different libraries that represent libraries of different
407 quality regarding their distribution, the two oligonucleotide pools were mixed in a different ratio for
408 each of the three libraries. For the first library, the target and control pool were mixed in a 1:1
409 ratio, to resemble an evenly distributed gRNA library. For the second library a 1:10 ratio of target
410 to control pool was applied. The third library was generated with a 1:100 ratio, to resemble a
411 library with highly underrepresented gRNA sequences. The mixed oligonucleotide pools were
412 phosphorylated, annealed to purified dU-ssDNA of the 3Cs multiplex template plasmid, and the
413 3CS synthesis reactions were performed as described above.

414

415 **Generation of multiplexed 3Cs- gRNA libraries**

416 *1. Equipment*

417 Desktop microcentrifuge, shaking incubator at 37°C, 1.5 ml collection tubes, filtered sterile pipette
418 tips, thermoblocks at 90°C and 50°C (e.g., Thermo Fisher, 88870004), an ultracentrifuge capable
419 of spinning 50 ml falcon tubes at 10,000 rpm (Beckman Coulter Avanti J-30 I ultracentrifuge and
420 a Beckman JA-12 fixed angle rotor), falcon tubes (polypropylene, 50 ml (Corning 352070)), a Bio-
421 Rad Gene Pulser electroporation system (BioRad 164–2076), electroporation cuvettes Plus (2
422 mm, Model no. 620 (BTX)), a gel electrophoresis chamber, erlenmeyer flasks (glass, 200 ml and
423 500 ml), 10 cm dishes 10 cm plastic culture dishes (Corning, CLS430591), 14 ml round-bottom
424 polystyrene tubes (e.g. Thermo Fisher, 10568531).

425

426 *2. dU-ssDNA template amplification*

427 *KCM transformation*

428 KCM competent bacteria (*Escherichia coli* strain K12 CJ236, NEB, E4141) were transformed with
429 3Cs multiplex template plasmid template by mixing 100 ng of DNA with 2 μ l of 5x KCM buffer
430 (0.5M KCl, 0.15M CaCl₂, 0.25M MgCl₂) and water in a 10 μ l reaction. After 10 min of incubation
431 on ice, an equal volume of CJ236 bacteria was added to the DNA/KCM mixture, gently mixed,
432 and chilled on ice for 15 min. The bacteria–DNA mixture was then incubated at room temperature
433 for 10 min and subsequently inoculated into 200 μ l of prewarmed SOC media (ThermoFisher
434 Scientific, 15544034). Bacteria were incubated at 37°C and 200 rpm for 1 hr and then streaked
435 on LB-agar plates with ampicillin (100 μ g/ml) and chloramphenicol (34 μ g/ml) for incubation
436 overnight at 37°C.

437

438 *Phage amplification and ssDNA purification*

439 The morning after transformation, a single colony of transformed *E. coli* CJ236 was picked into 1
440 ml of 2YT media (Roth, 6676.2) supplemented with M13KO7 helper phage (NEB, N0315) to a
441 final concentration of 1×10^8 pfu/ml and ampicillin (final concentration 100 μ g/ml) to maintain the
442 host F' episome and the phagemid, respectively. After 2 hrs of shaking at 200 rpm and 37°C,
443 kanamycin (Roth, T832.3) was added to a final concentration of 25 μ g/ml to select for bacteria
444 that have been infected with M13KO7 helper phage. Bacteria were kept at 200 rpm and 37°C for
445 6 to 8 hrs. Afterwards, the culture was transferred to 30 ml of 2YT media supplemented with
446 ampicillin (final concentration 100 μ g/ml) and kanamycin (final concentration 25 μ g/ml). After an
447 additional 20 hrs of shaking at 200 rpm and 37°C, the bacterial culture was centrifuged for 10 min
448 at 10,000 rpm and 4°C in a Beckman JA-12 fixed angle rotor. The supernatant was subsequently
449 transferred to 6 ml (1/5 of culture volume) PEG/NaCl (20% polyethylene glycol 8,000, 2.5 M NaCl)
450 and incubated for 1 hr at room temperature to precipitate phage particles. After 10 min of
451 centrifugation at 10,000 rpm and 4°C in a Beckman JA-12 fixed angle rotor, the phage pellet was
452 resuspended in 1.5 ml Dulbecco's phosphate-buffered saline (PBS, Sigma, D8662) and
453 centrifuged at 13,000 rpm for 5 min, before the phage-containing supernatant was transferred to
454 a clean 1.5 ml microcentrifuge tube and stored at 4°C. Circular ssDNA was purified from the
455 resuspended phages with the E.Z.N.A. M13 DNA Mini Kit (Omega Bio-Tek, D69001-01) according
456 to the manufacturer's protocol. Purity of the isolated ssDNA was ensured by agarose gel
457 electrophoresis and purified ssDNA was stored at 4°C.

458

459 3. *Multiplexed 3Cs-DNA synthesis*

460 The protocol for multiplexed 3Cs-DNA synthesis was adapted from Wegner et al., 2019 and
461 optimized for reactions on the 3Cs multiplex template plasmid with two specific annealing sites.
462 The oligonucleotides that were used for 3Cs reactions and the suppliers are listed separately (see
463 '3Cs oligonucleotide design rules' and 'DNA oligonucleotides').

464
465 *Oligonucleotide phosphorylation and annealing*
466 600 ng of oligonucleotides per annealing site (both, 3Cs h7SK- and hU6-oligonucleotides) were
467 phosphorylated in two separate 20 µl reactions by mixing them with 2 µl 10x TM buffer (0.1 M
468 MgCl₂, 0.5 M Tris-HCl, pH 7.5), 2 µl 10 mM ATP (NEB, 0756), 1 µl 100 mM DTT (Cell Signaling
469 Technology Europe, 7016), 20 units of T4 polynucleotide kinase (NEB, M0201) and water to a
470 total volume of 20 µl. The mixture was incubated for 1 hr at 37°C. Phosphorylated oligonucleotides
471 were immediately annealed to purified multiplex dU-ssDNA template by adding both 20 µl
472 phosphorylation products to 25 µl 10x TM buffer, 20 µg of dU-ssDNA template and water to a total
473 volume of 250 µl. The mixture was denatured for 5 min at 95°C, annealed for 5 min at 55°C and
474 cooled down for 10 min at room temperature.

475
476 *Multiplexed 3Cs-DNA reaction*
477 3Cs-DNA was generated by adding 10 µl of 10 mM ATP, 10 µl of 100 mM dNTP mix (Roth,
478 0178.1/2), 15 µl of 100 mM DTT, 2000 ligation units of T4 DNA ligase (NEB, M0202), and 30 units
479 of T7 DNA polymerase (NEB, M0274) to the annealed oligonucleotide-ssDNA mixture. The 3Cs
480 synthesis mix was incubated for 12 hrs (overnight) at room temperature. Afterwards the 3Cs
481 synthesis product was purified and desalted using a GeneJET Gel Extraction Kit (Thermo Fisher,
482 K0692) according to the following protocol: 600 µl of binding buffer and 5 µl 3M sodium acetate
483 (Sigma-Aldrich, 71196) were added to the synthesis product, mixed and applied to two purification
484 columns, which were centrifuged for 3 min at 460 g. The flow-through was applied a second time
485 to the same purification column to maximize yield. After two wash steps and 3 min of centrifugation
486 at maximum speed, the DNA was eluted in 50 µl prewarmed water. The 3Cs reaction product was
487 analyzed by gel electrophoresis alongside the dU-ssDNA template on a 0.8% TAE/agarose gel
488 (100 V, 30 min).

489
490 *4. Multiplexed 3Cs-DNA library amplification, clean-up and quality control*

491 *Electroporation of 3Cs synthesis product*

492 To amplify the multiplex 3Cs libraries, the 3Cs-DNA synthesis product was electroporated. To do
493 so, 400 µl of electrocompetent E. coli (10-beta, NEB, C3020K) were thawed on ice and mixed

494 with 6 μg of purified 3Cs-DNA. For electroporation, the DNA must be eluted in water or a low salt
495 solution. After that, the mixture was incubated on ice for 15 min and then transferred into a cold
496 2 mm cuvette (BTX, 45–0125) that was then inserted into a Bio-Rad Gene Pulser with the
497 following settings: resistance 200 Ω , capacity 25 F, voltage 2.5 kV. After electroporation, cells
498 were rescued in 25 ml of pre-warmed SOC media and incubated for 30 min at 37°C and 200 rpm.
499 After 30 min the culture was transferred into 400 ml of LB media supplemented with 100 $\mu\text{g}/\text{ml}$
500 ampicillin.

501

502 *Determination of transformation efficiency*

503 To ensure library representation during and after amplification, the number of transformants was
504 determined. After 30 min of incubation at 37°C and 200 rpm subsequently to the electroporation,
505 a series of 10-fold dilutions of the 10 μl of bacterial culture in sterile Dulbecco's phosphate-
506 buffered saline (PBS, Sigma, D8662) was prepared. Dilutions were plated in triplicates on LB-
507 agar containing 100 $\mu\text{g}/\text{ml}$ ampicillin and incubated overnight at 37 °C. The next morning, the
508 obtained colonies were counted. The number of transformants must be at least 100-fold higher
509 than the library complexity to ensure maintenance of library diversity.

510

511 *I-CeuI and I-SceI clean-up and quality control*

512 Plasmid DNA of overnight liquid cultures was purified using the Qiagen Plasmid Maxi Kit (Qiagen,
513 12163), according to the manufacturer's protocol to obtain the pre-library (P1). For removal of
514 residual 3Cs template plasmid from the multiplex pre-library, 3 μg of purified DNA was digested
515 with 10 units I-SceI (NEB, R0694), 10 units I-CeuI (NEB, R0699) and 5 μl NEB CutSmart buffer
516 (NEB, B7204) in a reaction volume of 50 μl for 3 hrs at 37°C. After 3 hrs, an additional 10 units of
517 I-SceI and I-CeuI and 5 μl NEB CutSmart buffer were added, as well as water to a final volume
518 of 100 μl . After further incubation for additional 3 hours, the digestion reaction was subjected to
519 gel electrophoresis on a 0.8% TAE/agarose gel (125 V, 40 min) to separate undigested 3Cs
520 synthesis product from linearized template plasmid. The band resembling the undigested correct
521 3Cs synthesis product was purified using a Thermo Fisher Scientific GeneJET Gel Extraction Kit,
522 according to the manufacturer's protocol. Then, the purified 3Cs synthesis product was
523 electroporated, according to the electroporation protocol described above. The next day, the
524 resulting final 3Cs multiplex library preparation (P2) was purified from liquid culture using a Qiagen
525 Plasmid Maxi Kit, according to the manufacturer's protocol and quality controlled by analytical
526 restriction enzyme digests, SANGER sequencing and by Next Generation Sequencing.

527

528 **Next-generation sequencing (NGS)**

529 *NGS sample preparation of 3Cs multiplex plasmid libraries*

530 3Cs multiplex plasmid libraries were prepared for NGS as follows: 250 ng of plasmid DNA was
531 used per PCR reaction and used in a volume of 50 μ l using Next High-Fidelity 2x PCR Master
532 Mix (NEB, M0541) (according to the manufacturer's protocol), containing 2.5 μ l of 10 μ M primers
533 each of forward and reverse primers. Depending on the library complexity up to four 50 μ l
534 reactions were performed. Primer sequences are listed separately (see 'DNA oligonucleotides').
535 Thermal cycler parameters were set as follows: initial denaturation at 98°C for 5 min, 15 cycles of
536 denaturation at 98°C for 30 s, annealing at 65°C for 30 s, extension at 72°C for 40 s, and final
537 extension at 72°C for 5 min. PCR products were purified from a 1.5 % TAE/agarose gel using a
538 GeneJet Gel Extraction Kit (Thermo Fisher Scientific), according to manufacturer's protocol.
539 Purified PCR products were denatured and diluted according to Illumina the guide lines and set
540 to a final concentration of 2.6 pM in a total volume of 2.2 ml and 15% PhiX control and loaded
541 onto a MiSeq, NextSeq 500 or NovaSeq sequencer (Illumina), according to manufacturer's
542 protocol. Sequencing was performed with single- or paired-end reads, 75 or 150 cycles, plus 8
543 cycles of index reading.

544

545 *NGS sample preparation of 3Cs multiplex screening samples*

546 To prepare 3Cs multiplex screening samples, the required amount of genomic DNA for sufficient
547 coverage was calculated first: for the autophagy single and multiplex FACS screening samples,
548 the required genomic DNA was calculated as *number of FACS sorted cells* \times
549 *screening coverage* \times 6.6 pg. For the autophagy multiplex proliferation control screen, the
550 required genomic DNA was determined by calculating *library complexity* \times
551 *screening coverage* \times 6.6 pg.

552 The NGS sample preparation of all samples from screening with the biased libraries was
553 performed with *library complexity* \times 200 (*maximum screening coverage*) \times 6.6 pg DNA. The
554 calculated amount of genomic DNA was used in a first PCR (PCR1) reaction with 2 to 4 μ g of
555 genomic DNA in a 50 μ l reaction using the Next High-Fidelity 2x PCR Master Mix (NEB, M0541)
556 (according to the manufacturer's protocol) and 2.5 μ l of 10 μ M PCR1 primers, each of forward
557 and reverse. Thermal cycler parameters were set as follows: initial denaturation at 98°C for 5 min,
558 15 cycles of denaturation at 98°C for 55 s, annealing at 65°C for 55 s, extension at 72°C for 110
559 s, and final extension at 72°C for 7 min. After PCR 1, 25 μ l of PCR 1 product was transferred to
560 a second PCR reaction (PCR2) in a 100 μ l reaction with 50 μ l High-Fidelity 2x PCR Master Mix
561 and 2.5 μ l of 10 μ M PCR 2 primers that contain Illumina adaptors. Primer sequences for PCR1

562 and PCR2 are listed separately (see 'DNA oligonucleotides'). Thermal cycler parameters were
563 set as follows: initial denaturation at 98°C for 5 min, 10 cycles of denaturation at 98°C for 30 s,
564 annealing at 65°C for 30 s, extension at 72°C for 40 s, and final extension at 72°C for 5 min. PCR
565 products were purified from a 1.5 % TAE/agarose gel and processed for NGS sequencing as
566 described for plasmid libraries.

567

568 **NGS data quality control and read count table generation**

569 Raw next generation sequencing data were processed and demultiplexed with bcl2fastq
570 v2.19.1.403 (Illumina). Read counts of individual gRNAs and gRNA combinations were
571 determined using cutadapt 2.8, Bowtie2 2.3.0, and custom Python 3 scripts^{67,68}. In brief, reads
572 were trimmed with cutadapt using 5' adapter sequences, truncated to 20 nucleotides, and aligned
573 to the respective gRNA library using Bowtie2 with no mismatches allowed. The uniformity of each
574 library distribution was assessed by plotting the cumulative distribution of all sequencing reads as
575 a Lorenz curve and determining the area under the curve. The library distribution skew (skew
576 ratio) of each library was determined by plotting the density of read counts and dividing the top
577 10 quantiles by the bottom 10 quantile. Cohen's D statistics were applied to assess the quality of
578 the biased libraries by comparing the distributions of non-targeting sequences and sequences
579 targeting core essential genes⁵². Pairwise sample correlations were determined with Pearson's
580 correlation of the normalized read counts and visualized with hierarchically clustered heat maps
581 (Seaborn library 0.10.1)⁶⁹.

582

583 **Enrichment analyses**

584 All enrichment analyses using MAGeCK were performed with median normalization of read
585 counts and gRNAs with zero counts in the control samples were removed. Down-sampling of the
586 1:1 dataset was performed by randomly choosing 1 to 16 gRNA combinations per gene
587 combination without replacement followed by individual MAGeCK analyses. gRNA combinations
588 with an $FDR \leq 10\%$ and $\log_2FC \leq -0.5$ were counted as statistically significant hits.

589

590 **Genetic interaction models**

591 Interactions of gene pairs were computed according to five different models: SUM, MIN, LOG,
592 MULT were used according to their definition in¹⁷, the MAX model defines the expected phenotype
593 of a double gene-knockout as the maximal phenotype of the individual single gene-knockouts.
594 The expected phenotypes of all double gene-knockouts were computed based on the phenotype
595 of the respective single gene knockouts which were defined as the median \log_2FC (as provided

596 by the MAGeCK analysis output) of all combinations of NHTs and gRNAs targeting the respective
597 gene. For each model, the deviation of observed paired-knockout phenotypes from their
598 expectation were expressed as their difference: $\Delta\log_2FC = \text{observed} - \text{expected}$. Assuming that
599 genetic interactions were rare, density plots of the dLFCs for each model were used to identify
600 the model with the highest number of neutral interactions, indicated by a single large peak around
601 0 on the x-axis. Using the MAX model, we kept only combinations with $p \leq 0.05$ and a $\Delta\log_2FC$
602 larger than the standard deviation of all $\Delta\log_2FC$ s.

603

604 **Autophagy gene interaction network**

605 To generate a network visualization based on our derived gene-gene interactions in autophagy,
606 we exported MAX model-dependent delta \log_2FC per gene-gene interaction and imported them
607 into the open source software platform for visualizing complex networks, Cytoscape (3.8.0)⁷⁰. The
608 style of the derived network was manually curated with the layout being set to a circular one.

609

610 **Cell culture**

611 Cell culture was performed as described previously⁴⁵. In brief, HEK293T cells (ATCC, CRL-3216)
612 were maintained in Dulbecco's Modified Eagle's Medium (DMEM, Thermo Fisher Scientific,
613 41965-039) and puromycin-sensitive hTERT-RPE1 cells (provided by Andrew Holland) in
614 DMEM: Nutrient Mixture F-12 (DMEM/F12, Thermo Fisher Scientific, 11320-074), each
615 supplemented with 10% fetal bovine serum (FBS, Thermo Fisher Scientific, 10270) and 1%
616 penicillin-streptomycin (Sigma-Aldrich, P4333) at 37°C with 5% CO₂. In addition, hTERT-RPE1
617 cells were supplemented with 0.01 mg/ml hygromycin B (Capricorn Scientific, HYG-H). No
618 method to ensure the state of authentication has been applied. Mycoplasma contamination testing
619 was performed immediately after the arrival of the cells and multiple times during the course of
620 the experiments. The hTERT-RPE1 GFP-LC3-RFP reporter cell line was generated by
621 transducing hTERT-RPE1(Cas9) cells with retroviral particles generated with the transfer plasmid
622 pMRX-IP-GFP-LC3-RFP (Addgene: 84573). Single cell clones were isolated and reporter
623 functionality was tested by Torin1 and Bafilomycin A1 treatments.

624

625 **Genomic DNA extraction**

626 Genomic DNA of cells was purified by resuspending PBS washed pellets of 40-50 million cells in
627 12 ml of TEX buffer (10 mM Tris-HCl, pH 7.5, 1 mM EDTA, pH 7.9, 0.5% SDS). Then 300 μ l of
628 proteinase K (10 mg/ml) and 300 μ l of Ribonuclease A (90 U/mg, 20 mg/ml) were added to the
629 resuspended cells. The tubes were incubated overnight at 37°C at constant shaking. After

630 complete cell lysis, 4 ml of 5 M NaCl was added, the solution was mixed and incubated at 4 °C
631 for 40 min. After that the tubes were centrifuged at 14,000xg for 1 hr. The supernatant was
632 transferred to a fresh tube and 24 ml of ice-cold 96% ethanol was added before the mixture was
633 placed at -20°C overnight. The next day, the tubes were centrifuged at 14,000xg for 1 hr.
634 Afterwards the supernatant was removed and the precipitated DNA was washed with ice-cold
635 70% ethanol. After further centrifugation 14,000xg for 1 hr the supernatant was removed and the
636 DNA pellet was dried at room temperature and then dissolved in 5 ml of sterile water.

637

638 **GFP and mCherry knockouts**

639 To examine the expression of gRNAs from both expression cassettes of the 3Cs multiplex
640 template plasmid, two oligonucleotide pools were designed following the 3Cs oligonucleotide
641 design rules. One oligonucleotide pool was designed for the h7SK cassette with 50 gRNAs
642 targeting eGFP, the second pool was designed for the hU6 expression cassette with 50 gRNA
643 targeting the mCherry gene. The two pools were used to generate three 3Cs libraries to selectively
644 target either eGFP (GFP single library) or mCherry (mCherry single library) or both simultaneously
645 (eGFP-mCherry multiplex library) following the protocol for generation of multiplexed 3Cs- gRNA
646 libraries described above. Lentiviral supernatant of the three libraries was generated. Monoclonal
647 hTERT–RPE1 cells with stable SpCas9, eGFP and mCherry expression were plated at 40%
648 confluency. The next day, the cells were transduced with viral supernatant of one of the three
649 libraries. After 48 hrs of transduction, cells were selected with puromycin (2.5µM) for 10 days.
650 Then, eGFP and mCherry ratios were quantified by FACS analysis.

651

652 **Generation, quantification and transduction of lentiviral particles**

653 Generation, quantification and transduction of lentiviral particles was performed as described
654 previously⁴⁵. In brief, the day before transfection, HEK293T cells were seeded to 2.5×10^5
655 cells/ml. To transfect HEK293T cells, transfection media containing 1/10 of culture volume Opti-
656 MEM I (Thermo Fisher Scientific, 31985–047), 10.5 µl Lipofectamin 2000 (Thermo Fisher
657 Scientific, 11668019), 1.65 µg/ml transfer vector, 1.35 µg/ml pPAX2 (Addgene: 12260) and 0.5/ml
658 µg pMD2.G (Addgene: 12259) was prepared. The mixture was incubated for 30 min at room
659 temperature and added dropwise to the media. Lentiviral supernatant was harvested 48 hr after
660 transfection and stored at -80°C.

661 To determine the lentiviral titer, hTERT–RPE1 cells were plated in a 6-well plate with 50,000 cells
662 per well. The following day, cells were transduced in the presence of 8 µg/ml polybrene (Sigma,

663 H9268) and a series of 0.5, 1, 5, and 10 μ l of viral supernatant. After 2 days of incubation at 37°C,
664 cells were subjected to puromycin selection for a total duration of 2 weeks, after which established
665 colonies were counted per viral dilution. The number of colonies in the highest dilution was then
666 volume normalized to obtain the final lentiviral titer.

667 To transduce hTERT–RPE1 cells, they were seeded at an appropriate density for each
668 experiment with a maximal confluency of 60–70%. On the day of transduction, polybrene was
669 added to the media to a final concentration of 8 μ g/ml. The volume of lentiviral supernatant was
670 calculated on the basis of the diversity of the respective library and of the desired coverage and
671 multiplicity of infection (MOI) of the experiment. A MOI of 0.5 was applied to all screens. The
672 number of cells that were transduced at the beginning of an experiment was calculated by
673 multiplying the diversity of the library with the desired coverage and needed MOI.

674

675 **3Cs CRISPR screening**

676 *Library distribution and experimental coverage interdependency screening*

677 To explore the interdependency of multiplexed CRISPR library distribution and experimental
678 screening coverage, three distorted 3Cs multiplex libraries were generated (see ‘generation of
679 distorted libraries’) that represented libraries of different gRNA distributions. All three libraries
680 were screened with a 20-fold and 200-fold coverage, each in triplicates. For the 20-fold screening,
681 for each replicate, 1.1 million SpCas9 expressing hTERT–RPE1 cells were plated (0.37 million
682 cells per flask) and transduced with the respective library with a MOI of 0.5. After 48 hrs the cells
683 were selected with 2 μ M puromycin and kept in growing conditions for 14 days. At day 14, the
684 cells were harvested, pooled and stored at -20°C until their genomic DNA was extracted and
685 processed for NGS. For the 200-fold screening a total of 11 million (0.5 million cells per flask)
686 SpCas9 expressing hTERT–RPE1 cells were plated and transduced with the respective library
687 with a MOI of 0.5. Further screening was performed identically to the 20-fold screen.

688

689 *Combinatorial 3Cs-gRNA autophagy screening*

690 Autophagy single and combinatorial gRNA screens for single or synergistic autophagy inhibition
691 were performed in biological replicates and triplicates in the monoclonal hTERT–RPE1 cell line
692 that stably expresses *Streptococcus pyogenes* Cas9 (SpCas9) and the autophagic flux probe
693 (GFP-LC3-RFP)⁵⁶, respectively. For each replicate 20 million cells (10 million for each, end time
694 point and day 2 control) were transduced with lentiviral supernatant of the autophagy multiplex
695 library with an MOI of 0.5 and a 1000- or 20-fold library coverage for single or combinatorial
696 autophagy library screening, respectively. The control time points were harvested 2 days post-

697 transduction. All remaining cells were kept in growing conditions until day 7, at what point the cells
698 were passaged, pooled and reseeded at library-diversity-maintaining density. After 13, 14 and 15
699 days the cells were treated with the mTOR inhibitor Torin1 (250 nM, InvivoGen, 1222998-36-8)
700 for 24 hrs in three batches to induce autophagy. After 24 hrs of Torin1 treatment, cells were
701 collected and 50,000 to 100,000 cells for single or 1.5 to 2.25 million cells for combinatorial
702 screening of each batch were FACS sorted to enrich for cells with blocked autophagy. The sorted
703 cells were reseeded and expanded for seven days before they were harvested, pooled and stored
704 at -20°C until their genomic DNA was extracted and processed for NGS.

705

706 **FACS**

707 Cell sorting was carried out with the FACS core facility of the Georg-Speyer Haus on a BD
708 FACSAria Fusion, and CRISPR screening hit validation analysis on a FACSCanto II flow
709 cytometer (BD Biosciences). Data was processed by FlowJo (FlowJo, LLC). Gating was carried
710 out on the basis of viable and single cells that were identified on the basis of their scatter
711 morphology.

712

713 **Arrayed autophagy candidate validation**

714 The validation of single and combinatorial autophagy screening hits was performed in arrayed
715 conditions (one knockout per well). To do so, single and dual gene-targeting CRISPR constructs
716 were designed and generated. For each gene, the top scoring guide sequence was selected with
717 Azimuth 2.0 of the GPP sgRNA Designer ([https://portals.broadinstitute.org/gpp/public/analysis-
718 tools/sgrna-design](https://portals.broadinstitute.org/gpp/public/analysis-tools/sgrna-design)) and purchased as forward and reverse oligonucleotide with compatible
719 overhangs for restriction enzyme cloning (see 'DNA oligonucleotides'). The two oligonucleotides
720 containing the gRNA target site were annealed and cloned into a restriction-enzyme digested and
721 gel purified CRISPR vector. In more detail, single gRNA constructs were cloned into lenti-sgRNA
722 blast vector (Addgene: 104993) by BsmBI restriction enzyme cloning. For combinatorial hit
723 validation the dual CRISPR gRNA expression cassettes of pKLV2.2-h7SKgRNA5(SapI)-
724 hU6gRNA5(BbsI)-PGKpuroBFP-W (Addgene: 72666) was cloned into the lenti-sgRNA blast
725 plasmid to enable blasticidin selection of dual gRNA constructs. A silent point-mutation was
726 introduced to remove the BbsI recognition site within the blasticidin sequence to allow the
727 subsequent insertion of one gRNA by SapI (NEB, R0569) cloning into the h7SK expression
728 cassette and the second gRNA by BbsI (NEB, R0539) cloning into the hU6 expression cassette.
729 After cloning and sequence verification by SANGER sequencing, lentiviral supernatant was
730 generated for each construct as described. Monoclonal hTERT-RPE1 cells with stable SpCas9

731 and GFP-LC3-RFP reporter expression were plated in 6-well plates with 50,000 cells per well.
732 The following day, cells were transduced in the presence of 8 µg/ml polybrene (Sigma, H9268)
733 with lentiviral supernatant. After 48 hrs the cells were selected with 10 µg/ml blasticidin
734 (InvivoGen, ant-bl) for 7 days, passaged and cultivated at 40-60% confluency under constant
735 blasticidin selection for an additional 7 days. At day 14, cells were treated with Torin1 to induce
736 autophagy for 24 hrs until they were collected at day 15 and subject to FACS cell sorting to
737 measure single or dual gene-knockout-induced autophagy blockage.

738

739 **gRNA performance and TIDE assay**

740 Guide RNA performance was evaluated by TIDE assay, as described previously^{45,71}. In short, for
741 each gRNA sequence, +/- 400 nucleotides from the gRNA annealing site, PCR primers were
742 designed to result in a PCR product of 800 to 1000 nts in length. The gRNA-locus is then PCR
743 amplified with OneTaq DNA polymerase (NEB, M0480) using 1 µg of genomic DNA, 40 µM dNTPs
744 (final concentration), 0.2 µM of each forward and reverse amplification primer, 10x OneTaq
745 standard buffer, and 2.5 units of OneTaq DNA polymerase. PCR cycles were set up as follows:
746 initial denaturation at 94°C for 3 min, 39 cycles of denaturation at 94°C for 20 s, annealing at 55°C
747 for 30 s, strand extension at 68°C for 2 min, and final strand extension at 68°C for 5 min. The
748 PCR products were analyzed on a 0.8% TAE/agarose gel (100 V, 30 min) and purified using a
749 Thermo Fisher Scientific GeneJET Gel Extraction Kit according to the manufacturer's protocol.
750 The purified PCR product was pre-mixed with forward amplification primer and processed by
751 SANGER sequencing, after which wildtype and gRNA-treated SANGER chromatograms were
752 analyzed by TIDE and the percentage of unedited DNA extracted (<https://tide.deskgen.com/>).

753

754 **Data availability**

755 NGS data are provided as raw read count tables as Supplementary Table 4. Plasmids encoding
756 for extended- and combinatorial-autophagy libraries will be available through the Goethe
757 University Depository (<http://www.innovectis.de/INNOVECTIS-Frankfurt/Technologieangebote/Depository>).

759

760 **Code availability**

761 Custom software is publicly available from GitHub, <https://github.com/GEG-IBC2/3Cs-MPX>
762 (GEG-IBC2, 2019; copy archived at <https://github.com/elifesciences-publications/3Cs>).

763

764 **DNA oligonucleotides and oligo pools**

765 Sequences of used DNA oligonucleotides and for 3Cs libraries are provided in Supplementary
766 Table 2.

767

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788

789 **COMPETING INTERESTS**

790 The Goethe University Frankfurt has filed a patent application related to this work on which
791 Valentina Diehl, Martin Wegner, Ivan Dikic and Manuel Kaulich are inventors (WO2017EP84625).
792 The Goethe University provides an exclusive license of the 3Cs technology to Vivlion GmbH for
793 which Ivan Dikic and Manuel Kaulich are co-founders, shareholders and chief officers.

794

795 **FIGURE LEGENDS**

796 **Figure 1. 3Cs multiplexing for combinatorial CRISPR gRNA libraries. A)** 3Cs
797 multiplexing workflow. Two gRNA-encoding oligonucleotide pools are selectively

798 annealed to one of the gRNA-expression cassettes of the dU-containing ssDNA template
799 plasmid for T7 DNA Pol-dependent generation of heteroduplex dU-dsDNA. Amplification
800 in dut/ung-positive bacteria amplifies the combinatorial gRNA plasmid. See Material and
801 Methods for a more details. **B)** Cas9 GFP/mCherry multiplex library design. Combinatorial
802 gRNA constructs target GFP and mCherry genes simultaneously; each gene is
803 individually targeted by 50 gRNAs, both genes are simultaneously targeted with 2601
804 gRNA combinations. The single guide GFP-targeting library contains the wildtype gRNA
805 placeholder in the hU6 cassette (mCherry), and vice versa. **C)** Gel-electrophoresis after
806 analytical restriction enzyme digest of final 3Cs single and combinatorial libraries. **D)**
807 Area-under-the-curve (AUC) determination of single and combinatorial library
808 representation. As a reference, a perfectly distributed library (ideal) is shown in grey.
809 Percentages indicate a library's representation at 90% of cumulative reads. AUC values
810 are indicated next to each library's identifier. **E)** FACS analysis of GFP- and mCherry-
811 positive hTERT-RPE1 cells after transduction with single or combinatorial libraries. Error
812 bars represent standard deviations (SDs) over three biological replicates (n = 3).

813

814 **Figure 2. Library distribution skew dictates experimental scale and data quality. A)**
815 Area-under-the-curve (AUC) determination of the nucleotide-randomized libraries. As a
816 reference, a perfectly distributed library (ideal) is shown in grey. Percentages indicate
817 library representations at 90% of cumulative reads. AUC values are indicated next to each
818 library's identifier. **B)** Analysis of NHT-fraction in the biased 3Cs libraries. Median and
819 quartiles of the distributions are shown as red straight and black dotted line, respectively.
820 **C)** Area-under-the-curve (AUC) determination of the biased libraries. As a reference, a
821 perfectly distributed library (ideal) is shown in grey. Percentages indicate a library's
822 representation at 90% of cumulative reads. AUC values are indicated next to each
823 library's identifier. **D)** Density plots showing the log₂FC separation of combinatorial gRNA
824 constructs targeting core essential genes (blue) and non-essential controls (green) in
825 experimental coverages of 20x (dotted) and 200x (straight). QS: quality score. **E)** Analysis
826 of the number of depleted gene-pairs detected with MAGeCK at the indicated FDR and
827 log₂FC cutoffs. **F)** Determination of the number of depleted gene-pairs at the indicated

828 FDR and log2FC cutoffs from sub-sampled read-count tables of replicates for each
829 coverage containing 1 to 16 randomly chosen gRNA pairs.

830

831 **Figure 3. Single and multiplex-inherent single gRNA enrichment screens are**
832 **complementary. A)** Cas9 autophagy single and multiplex library design. Combinatorial
833 gRNA constructs target extended and core autophagy genes; each gene is targeted by 4
834 gRNAs, 192 extended autophagy genes and 64 core autophagy genes with 10% of NHT
835 controls per cassette generating 247,032 gRNA combinations. Single gRNA extended
836 autophagy library contains the wildtype gRNA placeholder in the hU6 cassette (core
837 autophagy). **B)** Single (s) and combinatorial (mpx) autophagy screening workflow. Cov.:
838 coverage (library representation); MOI: multiplicity of infection; FACS: fluorescence-
839 activated cell sorting; NGS: next-generation sequencing. **C)** Cell sorting representations
840 of single and combinatorial (MPX) screen of post-FACS gate (#) and high-gate (*). **D)**
841 MAGeCK analysis of dedicated single gRNA autophagy screen of pre- and post-FACS
842 samples with hit genes in red when matching cutoff criteria of $FDR \leq 10\%$ and p -
843 value ≤ 0.05 . Screen-selective hits in blue. **E)** MAGeCK analysis of multiplex-inherent
844 single gRNA autophagy screen of pre- and post-FACS samples with hit genes in red when
845 matching cutoff criteria of $FDR \leq 10\%$ and p -value ≤ 0.05 . Screen-selective hits in blue. **F)**
846 Analysis of single hit genes derived from D) and E) in arrayed autophagy blockage
847 validations (red). Evaluation of gRNA activity by TIDE analysis (grey). Error bars
848 represent standard error of mean (SEM) over three biological replicates per autophagy
849 blockage (n=3). ND: not determined.

850

851 **Figure 4. Paralogs are redundant in autophagy. A)** MAGeCK analysis of combinatorial
852 gRNAs targeting gene pairs in which both genes were identified as essential for
853 autophagy of pre- and post-FACS samples. Hit gene pairs are shown in red when
854 matching cutoff criteria of p -value ≤ 0.05 . **B)** MAGeCK analysis of combinatorial gRNAs
855 targeting gene pairs in which one gene was identified as essential for autophagy of pre-
856 and post-FACS samples. Hit gene pairs are shown in blue when matching cutoff criteria
857 of p -value ≤ 0.05 . **C)** MAGeCK analysis of combinatorial gRNAs targeting gene pairs with
858 neither gene being identified as essential for autophagy of pre- and post-FACS samples.

859 Hit gene pairs are shown in yellow when matching cutoff criteria of $p\text{-value} \leq 0.05$. Ras-
860 related protein family (RAB) and ATG2 gene pairs are shown in blue and green,
861 respectively. **D)** Global view on identified gene pairs per category in percent. Ess:
862 essential; non-ess: non-essential. Color code adapted from A) to C). **E-G)** Arrayed
863 analysis of hit gene pairs and the induced blockage of autophagy per gene knockout of
864 each category, color code of A) to C). Control genes and sequences are shown in grey.
865 Error bars represent standard error of mean (SEM) over three biological replicates ($n=3$).
866 **H)** Density plots of delta $\log_2\text{FC}$ ($\Delta\log_2\text{FC}$) analyses computed by MAX, SUM, MIN,
867 MULT, and LOG models. **I)** Correlation between observed and expected $\log_2\text{FC}$ values,
868 derived from MAX model, for combinatorial gene-targeting. Data points above standard
869 deviation and with $p\text{-values} \leq 0.05$, derived from C), are highlighted in yellow, representing
870 synergistic gene interactions in autophagy. **J)** Network analysis of synergistic autophagy
871 gene pairs, derived from I). Edge color and width set to $\Delta\log_2\text{FC}$ values derived from
872 MAX model. Edges and nodes of paralog gene pairs are highlighted in pink.

873

874 **Supp. Figure 1. A)** Analysis of single and multiplex dU-containing hetero-duplex 3Cs
875 DNA by gel-electrophoresis. **B)** NGS sequencing depth of single and combinatorial GFP,
876 mCherry, and GFP+mCherry 3Cs libraries. A sample's median and quartiles are shown
877 as red straight and black dotted line, respectively. **C)** Analysis of distribution skew (skew)
878 and completeness (compl.) per library, based on read counts derived from B).

879

880 **Supp. Figure 2. A)** NGS sequencing depth of nucleotide-randomized combinatorial 3Cs
881 libraries (1-4N). A library's median and quartiles are shown as red straight and black
882 dotted line, respectively. **B)** Analysis of distribution skew (skew) and completeness
883 (compl.) per library, based on read counts derived from A). **C)** NGS sequencing depth of
884 distribution skew biased combinatorial 3Cs libraries (1:1, 1:10, 1:100). A sample's median
885 and quartiles are shown as red straight and black dotted line, respectively. **D)** Analysis of
886 distribution skew (skew) and completeness (compl.) per library, based on read counts
887 derived from C).

888

889 **Supp. Figure 3. A-B)** Pairwise sample correlation (Pearson's correlation coefficient),
890 visualized as hierarchically clustered heatmaps (n), library distributions skews (1:1, 1:10,
891 1:100) and coverages (20x, 200x) on gRNA-pair (A) and gene-pair (B) levels. Color code
892 based on Pearson's correlation coefficient (ρ) of normalized gRNA read counts.

893
894 **Supp. Figure 4. A)** FACS analysis of monoclonal hTERT-RPE1 GFP-LC3B-RFP reporter
895 cell line under conditions of basal autophagy (Basal), Torin1-induced autophagy
896 (+Torin1), and Torin1-induced but Bafilomycin A1-blocked autophagy (+Torin1 +
897 Bafilomycin A1). Gating is based on Torin1-induced reduction of GFP signal; percentage
898 (%) of cells in gate. **B)** Analysis of NGS sequencing depth per 3Cs library (single, mpX)
899 and replicate post-FACS sample (1-3). A sample's median and quartiles are shown as
900 red straight and black dotted line, respectively. **C)** Analysis of distribution skew (skew)
901 and completeness (compl.) per autophagy library (single, mpX), based on read counts
902 derived from B). **D)** Area-under-the-curve (AUC) determination of the single and
903 combinatorial (multiplex) autophagy libraries. As a reference, a perfectly distributed library
904 (ideal) is shown in grey. AUC values are indicated next to each library's identifier.

905
906 **Supp. Figure 5. A-D)** Pairwise sample correlation (Pearson's correlation coefficient),
907 visualized as hierarchically clustered heatmaps (1-3) of pre- and post-FACS samples of
908 autophagy enrichment screens with single (B) and combinatorial gRNA-targeting in
909 autophagy blockage (C) and high-gates (D) on gRNA and gene level. Color code based
910 on Pearson's correlation coefficient (ρ) of normalized gRNA read counts. **E)** Analysis of
911 ATG4B-associated guide pairs in high-gate post-FACS samples.

912
913 **Supp. Figure 6. A-B)** MAGeCK analysis of dedicated single (A) and multiplex-inherent
914 single (B) gRNA autophagy screens between pre- and post-FACS samples with genes in
915 red when matching cutoff criteria of $p\text{-value} \leq 0.05$. Screen-selective hits in blue.

916
917 **Supp. Figure 7. A)** Log₂FC-analysis of targeting a single gene with one (single) or two
918 (double) gRNAs. The dotted diagonal line represents equal phenotypic strength, based
919 on FACS enrichment. **B)** Arrayed analysis of phenotypic strength of single essential

920 genes for autophagy when targeted with one (yellow) or two (blue) gRNAs. Error bars
921 represent standard error of mean (SEM) over three biological replicates (n=3). **C**) Density
922 plots of delta log₂FC ($\Delta\log_2\text{FC}$) value analyses of single essential genes for autophagy,
923 computed by MAX, SUM, MIN, MULT, and LOG models.

924

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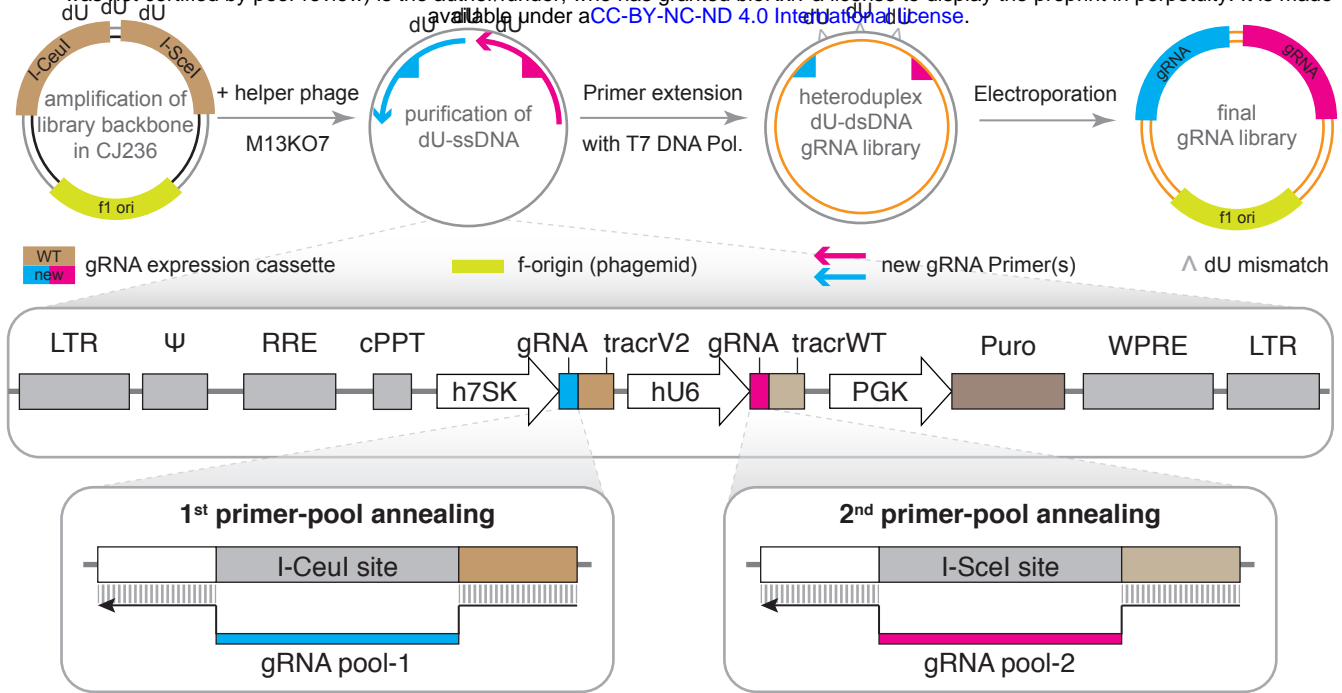
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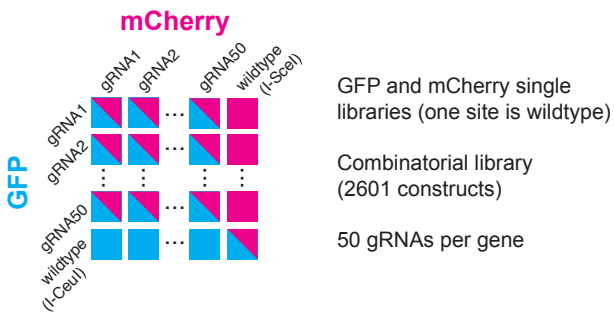
Figure 1

A

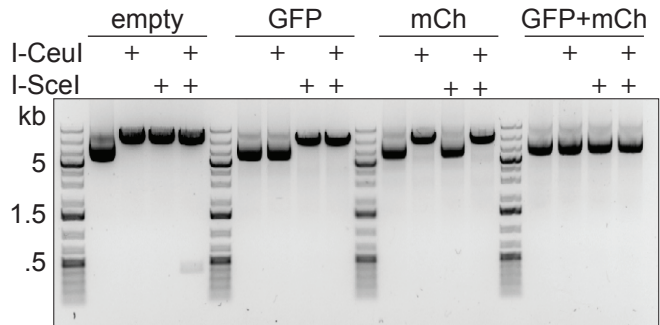
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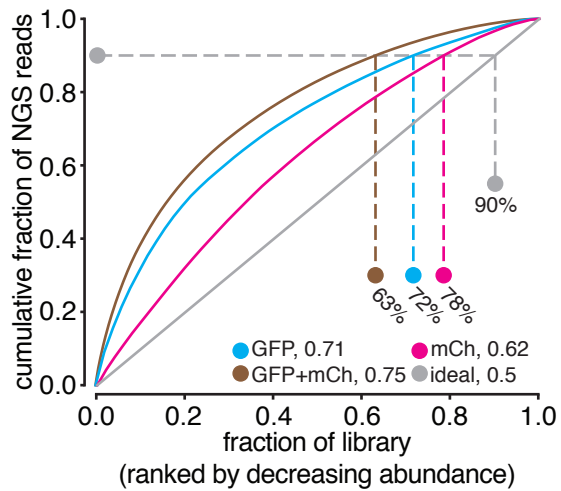
B



C



D



E

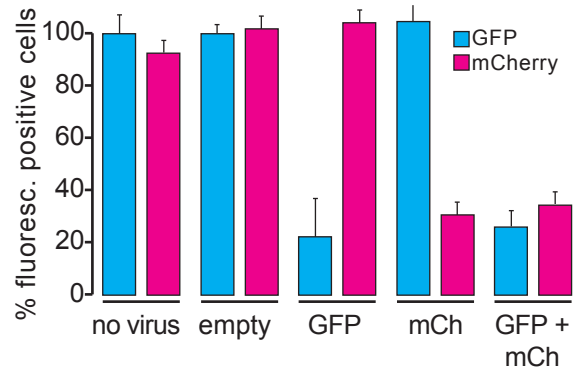


Figure 2

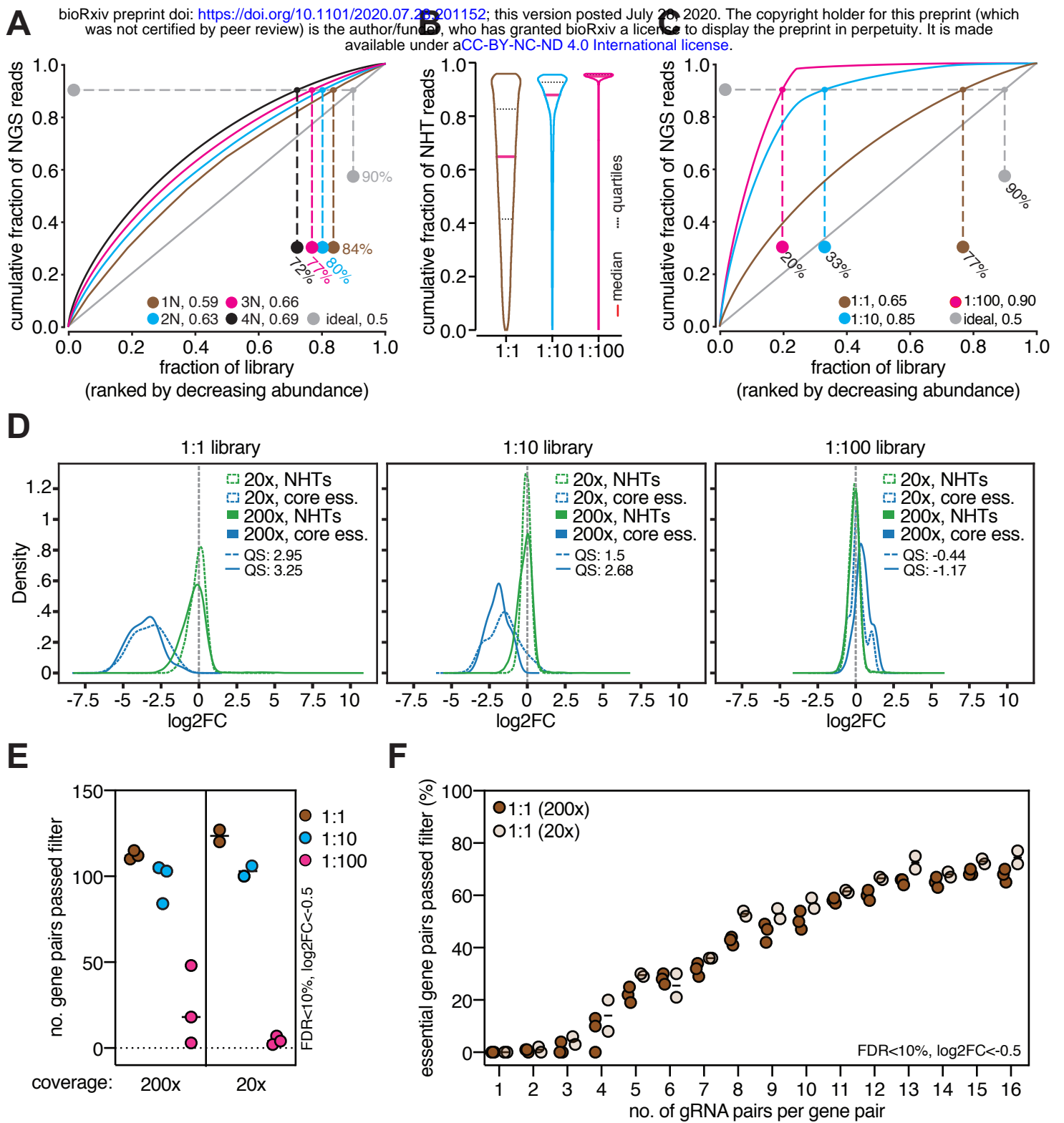


Figure 3

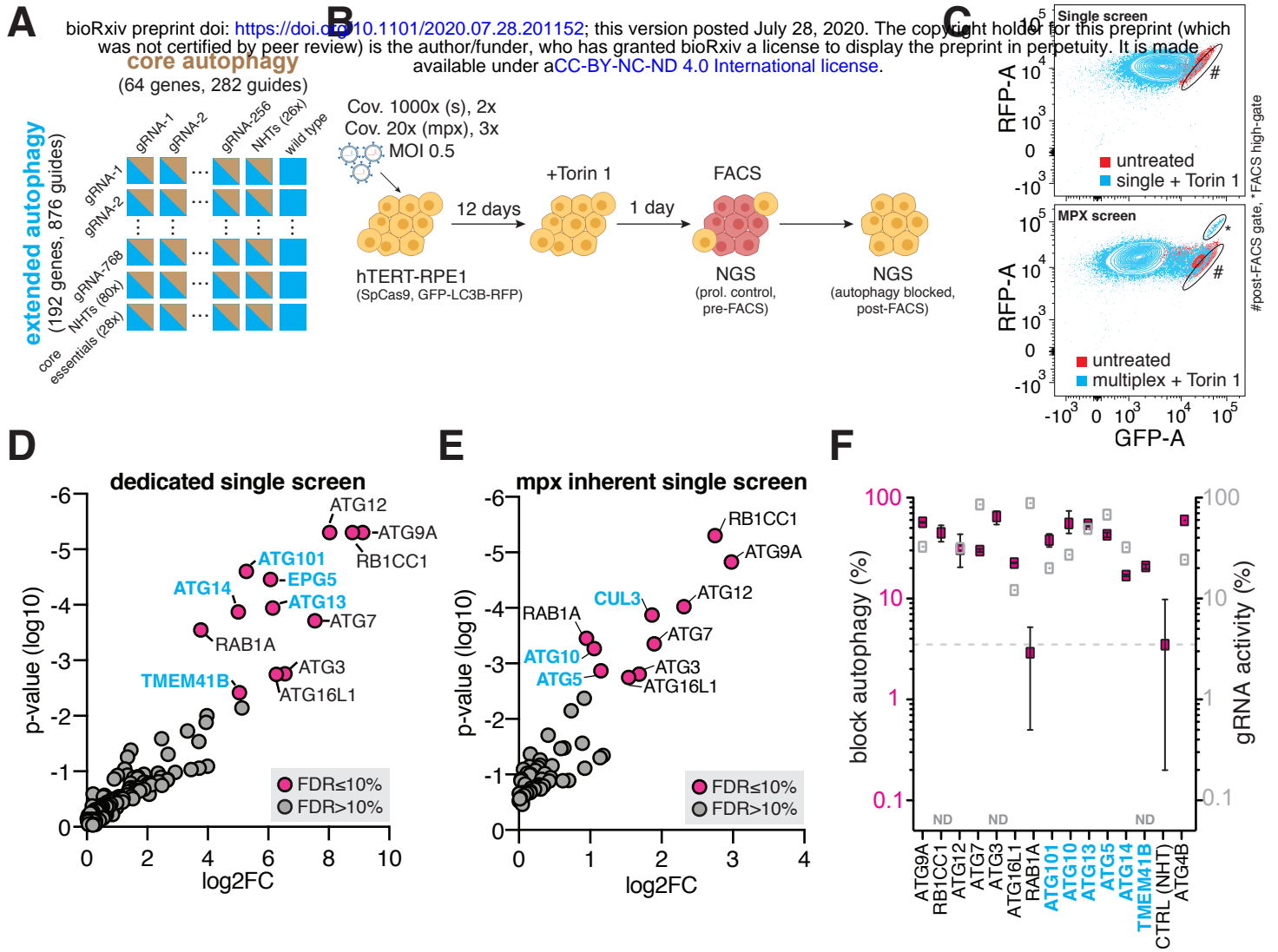
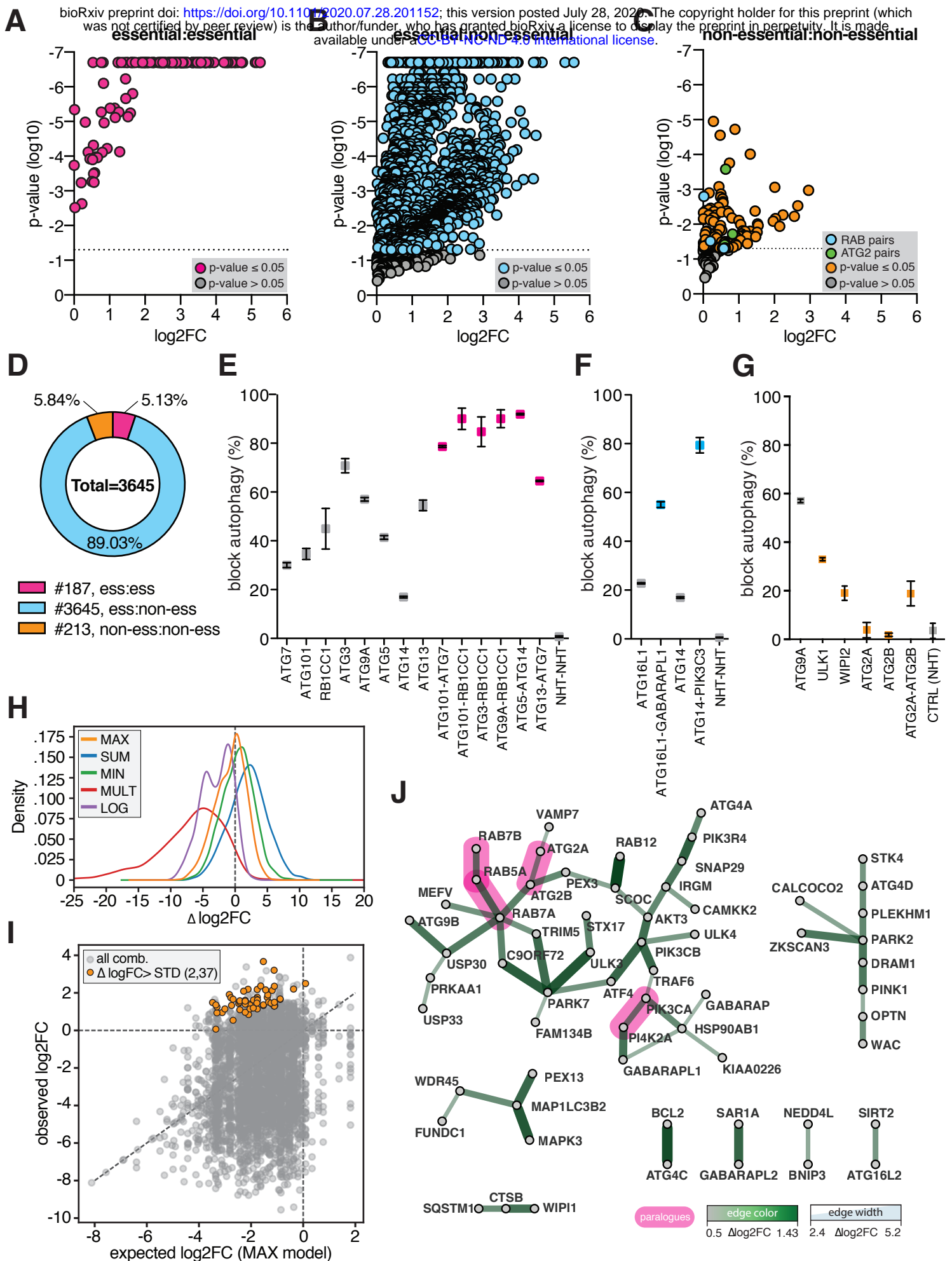
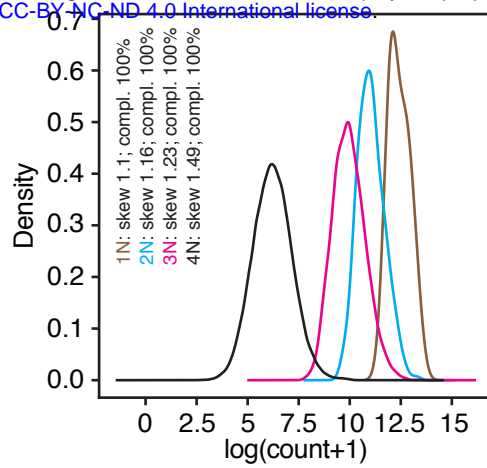
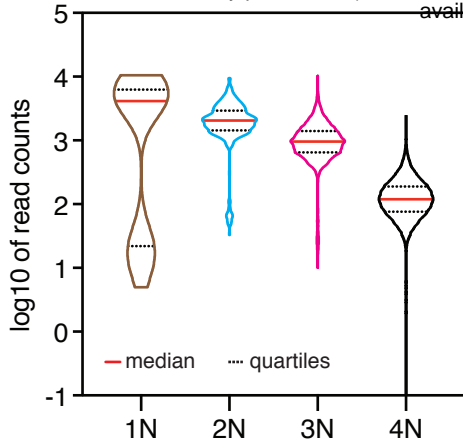


Figure 4

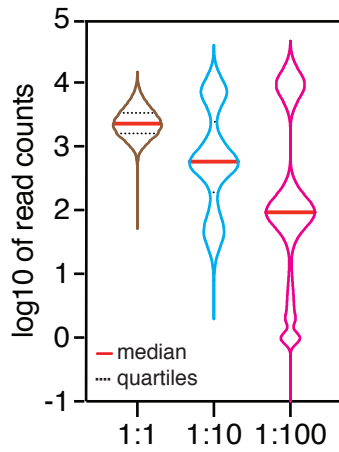


Supp. Figure 2

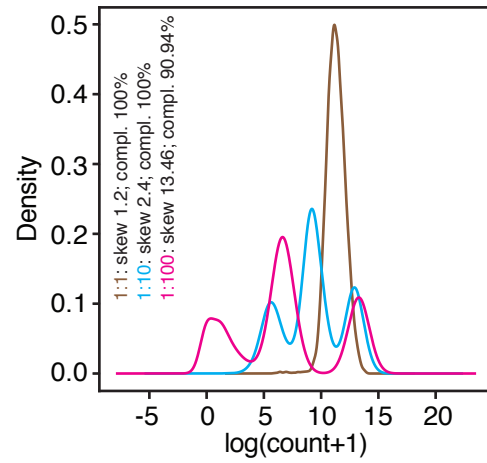
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C

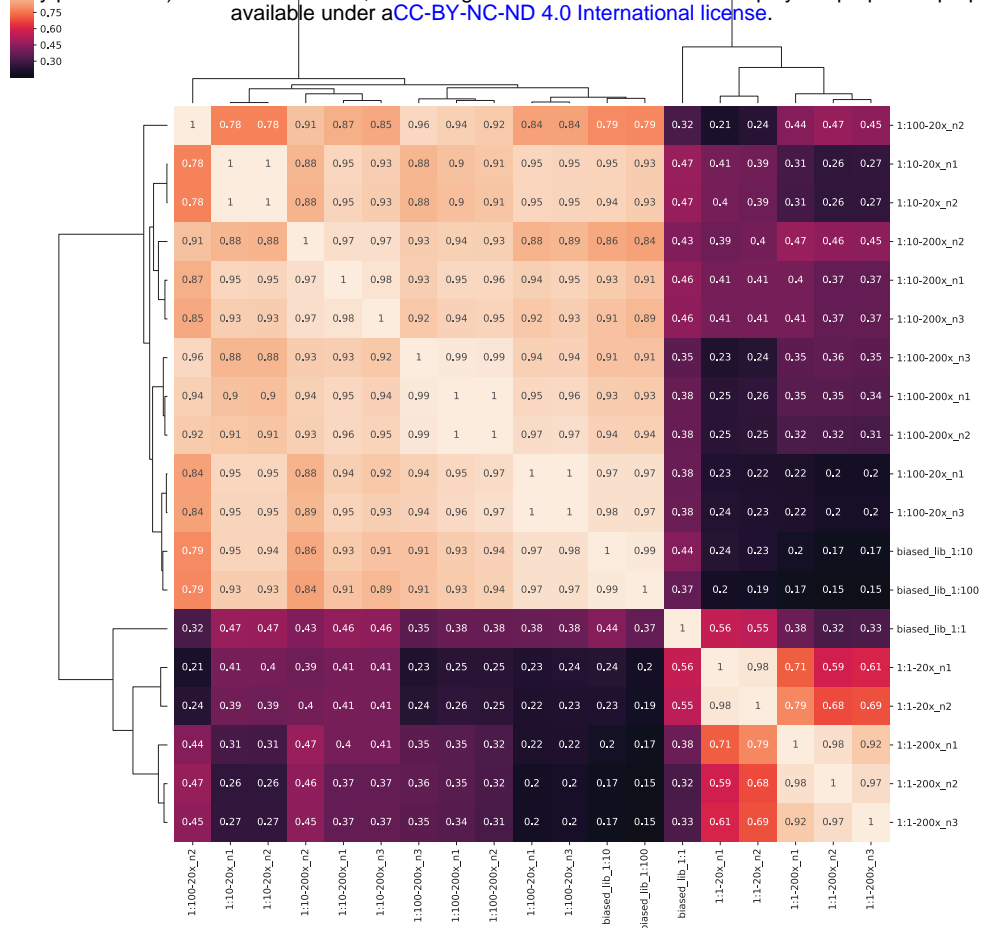


D

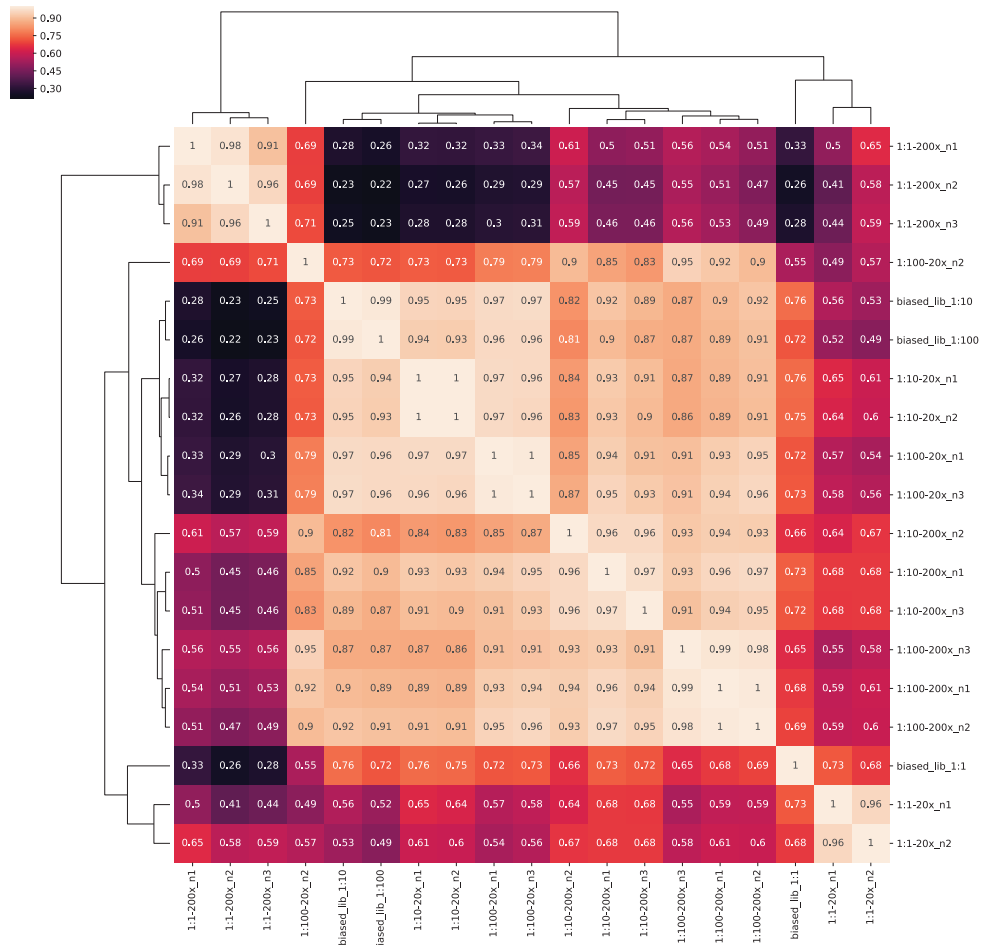


Supp. Figure 3

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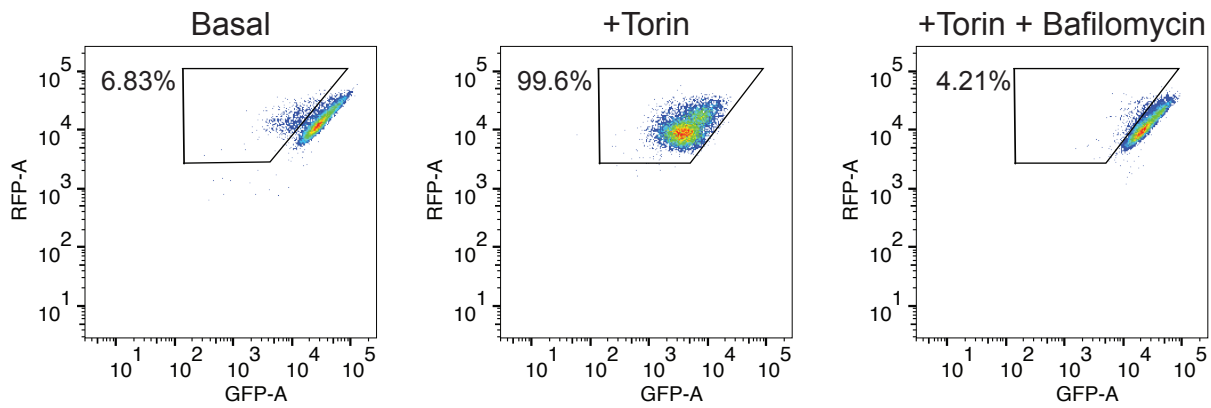
B



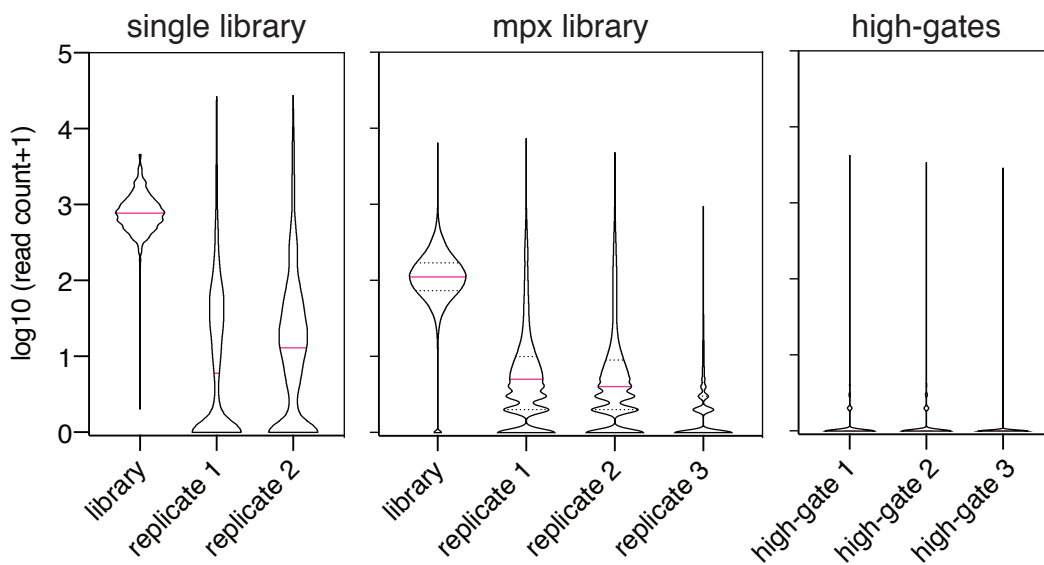
Supp. Figure 4

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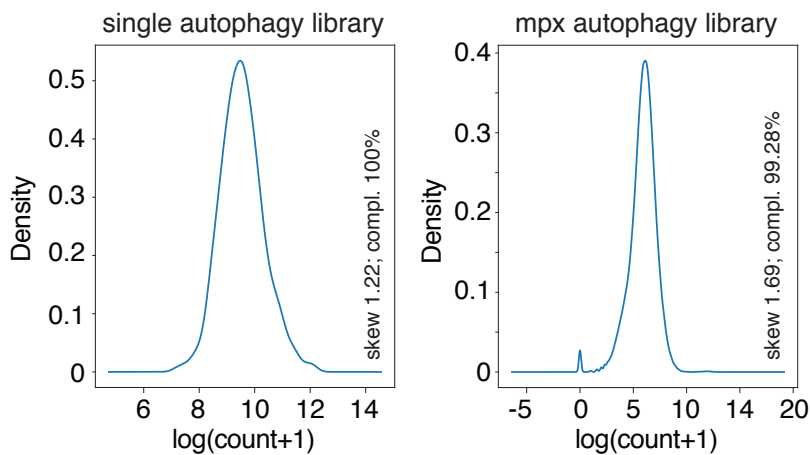
A



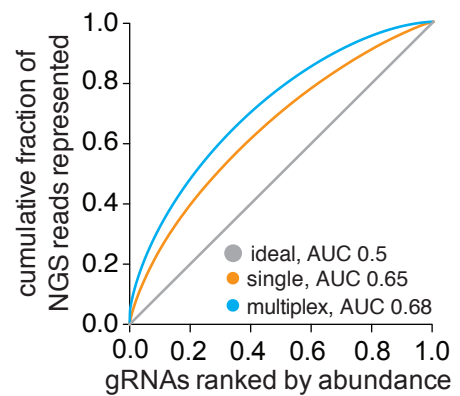
B



C

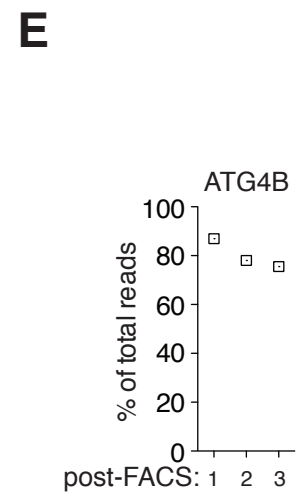
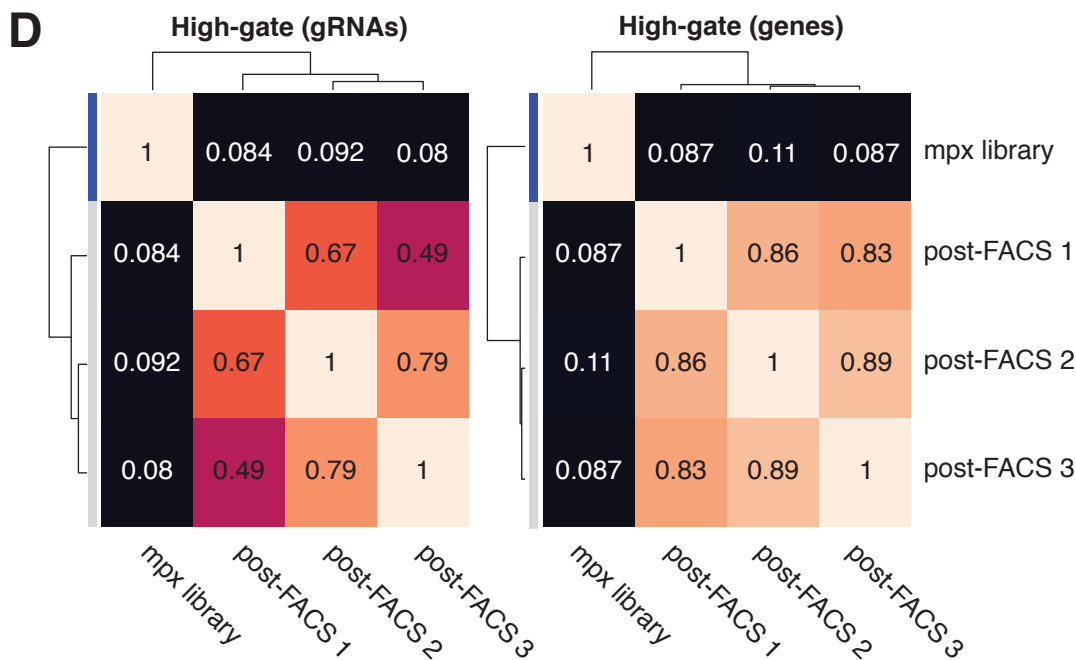
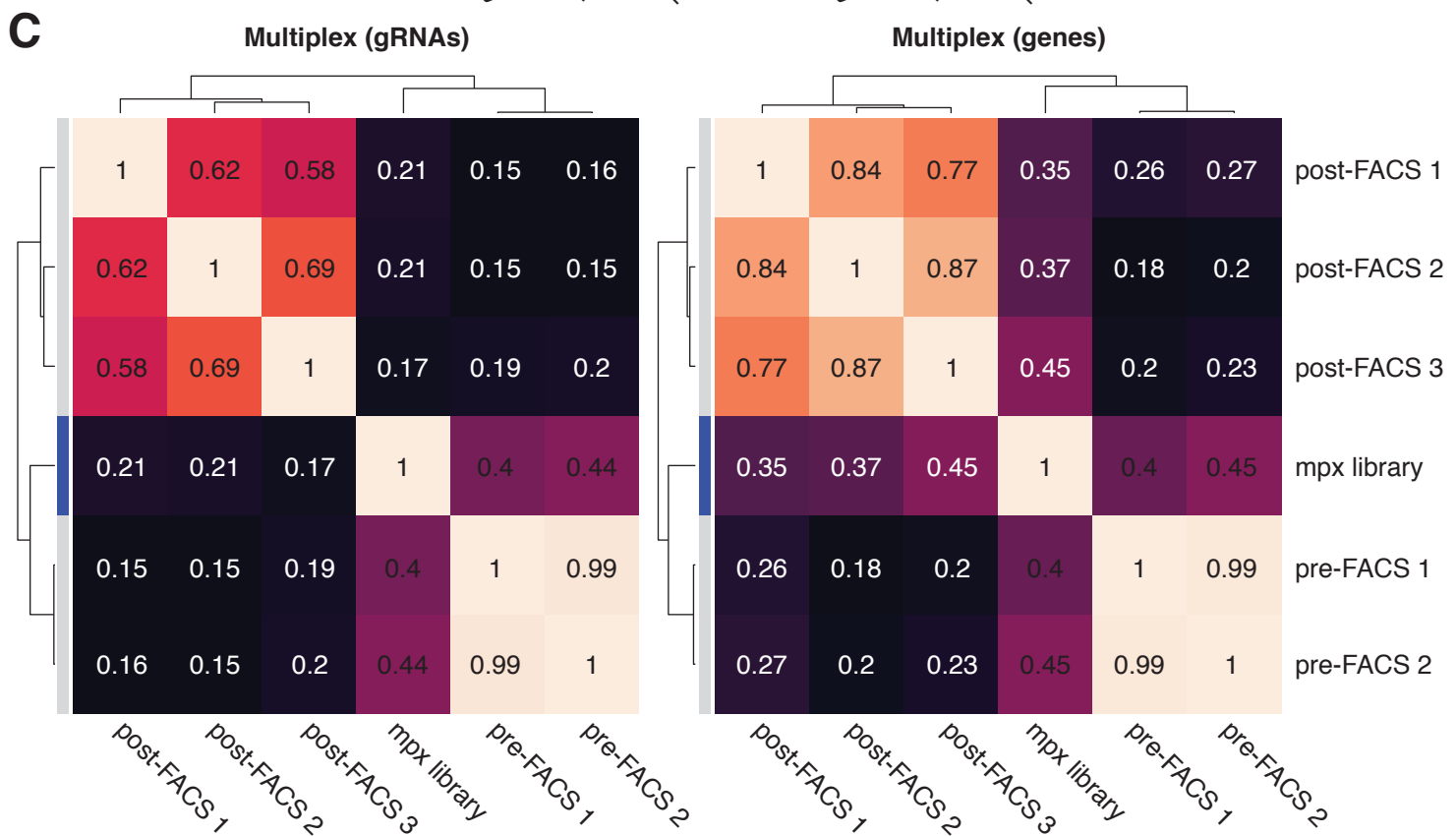
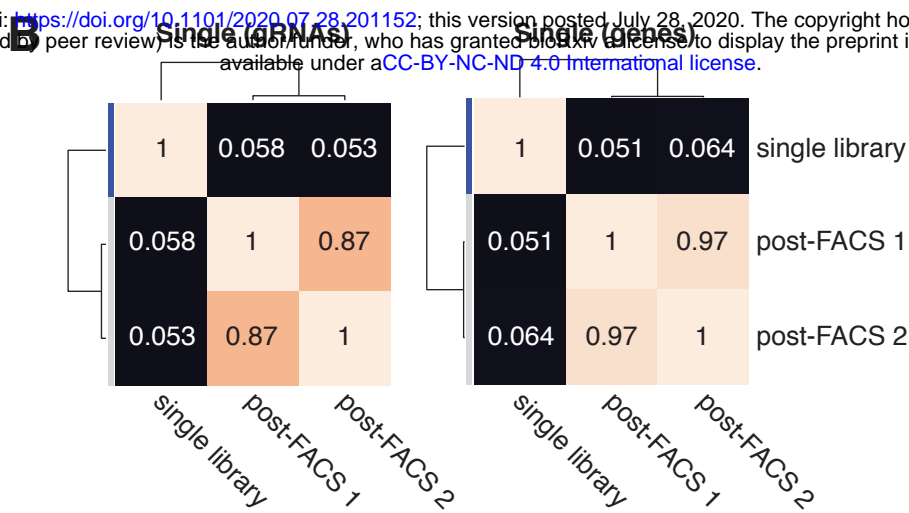
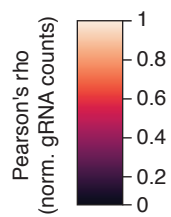


D



Supp. Figure 5

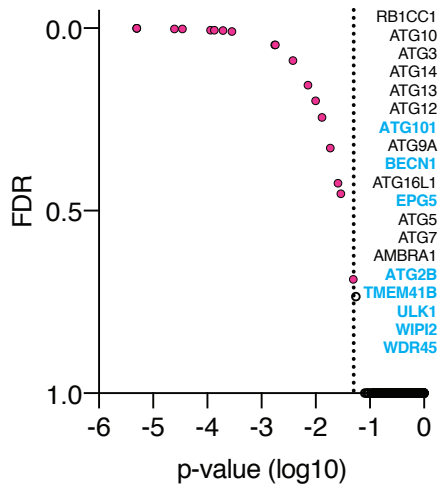
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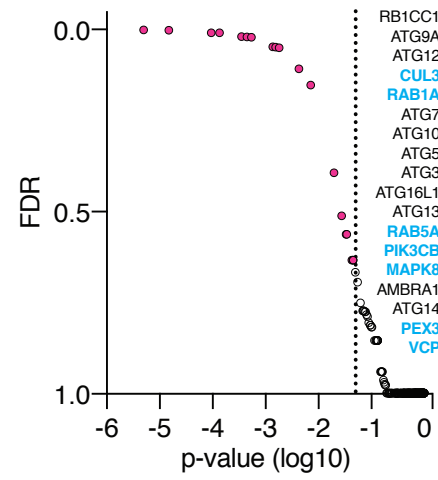
Supp. Figure 6

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A dedicated single screen



B mpX inherent single screen



Supp. Figure 7

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