

1 **A new molecular method for the exploration of hybrid**
2 **zones between two toad species of conservation interest**

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26

27 **Abstract**

28 Analyzing hybrid zones between previously isolated lineages allows insight into
29 processes determining the fate of re-encounter of these taxa. The distributions of Fire-
30 bellied (*Bombina bombina*) and Yellow-bellied toads (*B. variegata*) meet in the
31 Carpathian Basin resulting in a narrow contact zone in the foothill regions, where hybrids
32 often appear. Our aim was to explore a transient zone between *B. bombina* and *B.*
33 *variegata* within the Carpathian Basin along a transect including the Börzsöny Hills in
34 Hungary and Krupinská Planina in Slovakia. We visited 28 locations in these areas and
35 collected altogether 230 specimens, photographed and sampled them using buccal swabs.
36 In order to distinguish between the two species and determine hybrid individuals, we used
37 mitochondrial markers and designed a novel technique based on the restriction of the
38 *Ncx-1* gene. The geographical distribution pattern of these two species delivered
39 unexpected results, as Börzsöny Hills was completely colonised by *B. bombina* including
40 locations which can be classified as typical habitats of *B. variegata*. Moreover, in
41 Krupinská Planina many locations were also colonised by *B. bombina*, including high
42 altitude ponds. The most remote sites still harbour *B. variegata* populations, but seven of
43 these were found with hybrid individuals. This pattern may indicate the northward and
44 altitudinal range expansion of *B. bombina* and the colonisation of habitats of its sister
45 species in these areas. Our results warrant enhanced attention to hybrid zones, where
46 introgression and changes in population composition may reflect recent rapid
47 environmental alterations and redirect conservation focus.

48 **Keywords**

49 *Bombina bombina*, *Bombina variegata*, Range expansion, Climate change,
50 Contact zone, Distribution mapping, Introgression

51 **Introduction**

52 One of the main processes driving speciation is geographic isolation due to one or
53 more vicariant events leading to divergent lineages [1]. On one hand, different ecological
54 constraints distinctly shape the two lineages, facilitating ecological differentiation (i.e.
55 adaptation to different habitats). On the other hand, allopatric distribution may lead to the
56 accumulation of random genetic differences between the isolated lineages. Both types of
57 genetic divergence may potentiate reproductive isolation. However, if there is insufficient
58 time available for speciation, incomplete reproductive isolation may result in
59 hybridization between the sister-lineages upon re-encounter [2,3]. In Europe, several
60 cases of hybridization are considered to be a consequence of warming periods after glacial
61 ages, when previously allopatric populations of sister lineages became parapatric due to
62 area-expansions and came into secondary contact [4]. Analyzing hybrid zones of
63 genetically and ecologically differentiated, but not reproductively isolated taxa allows
64 insight into admixture processes determining the fate of the closely related lineages (see
65 e.g. [5]).

66 The hybrid zone between the European fire-bellied and yellow-bellied toads
67 (*Bombina bombina* and *B. variegata*; Bombinatoridae) in Central Europe has been
68 identified by morphologic, electrophoretic and genetic analyses [6–9]. The two species
69 are similar with a warty, cryptically coloured dorsal and aposematic, brightly coloured
70 ventral pattern [10,11]. The first attempts to distinguish between sister species and
71 hybrids were based on morphological characteristics. The most commonly used trait was
72 the extension and connectivity of the ventral patches at particular body parts,
73 supplemented with morphometric measurements, including snout-vent length, head
74 length, head width, distance between eyes, forelimb length, femur length, tibia length,

75 and foot length [8,12]. However, transitional morphological characters between *B.*
76 *bombina* and *B. variegata* had been observed as early as the 1890s [13], due to
77 hybridization in the contact zones [9,14]. Therefore, more reliable identification based on
78 molecular methods were designated including protein electrophoresis [7,9,15,16] and
79 DNA techniques using mitochondrial [8,17] and nuclear gene markers [18] or
80 microsatellites [19,20]. Molecular analyses revealed that introgression has not been
81 detected outside the hybrid zones and the genetic structures vary in pure and hybrid
82 populations [10,17–19,21,22]. The hybrid zones cover different genetic structures
83 depending on their location [18].

84 The location of the hybrid zone is determined by environmental factors due to the
85 diverse adaptations to different habitats of the sister species. The distribution of *B.*
86 *bombina* covers the lowlands of Europe from Germany to Russia and probably extends
87 even beyond the Ural Mountains, while *B. variegata* occurs mainly in the highlands of
88 the Balkan Peninsula, Central and Western Europe and is absent east of the Carpathian
89 Ridges [23]. Thus, the two species have parapatric distribution with a narrow but extended
90 contact zone stretching from Germany southward to Bulgaria [24]. Based on mtDNA
91 analyses, *B. bombina* is composed of two clades confined to Northern and Southern
92 Europe, with low mtDNA divergence between and within clades [17,18]. In contrast,
93 mtDNA variation of *B. variegata* in its range is extensive, and four distinct haplotypes
94 are identified. Hence the overall pattern of their recent distribution reflects a post-glacial
95 radiation from Balkan, Mediterranean and Carpathian refugia, whereas recolonization of
96 European habitats by *B. bombina* initiated from one refugium near the Black Sea coast
97 [17,18,25].

98 The Carpathians and their surrounded basin with complex topography created a
99 complicated and diverse contact zone between the two species. The inner part of the
100 Carpathian Basin is divided by mid-sized ridges with the highest elevation of Kékes
101 (Hungary) up to 1015 m above sea level (asl), where *B. variegata* occurs in isolated
102 populations inhabiting small temporary ponds and puddles at these mountainous
103 enclaves. In contrast, previous studies indicate that *B. bombina* prefers larger, more
104 permanent water bodies at lowland, open areas with more connected populations [21,26].
105 Due to the different habitat preferences, the two species contact mostly along the foothill
106 regions, forming an extended contact zone in the Carpathian Basin with resulting hybrid
107 individuals in many populations [8,16]. Gollmann and colleagues [7] studied a shorter
108 section of the ridges on the border of northern Hungary and southern Slovakia in the
109 Aggtelek Karst region and revealed a hybrid zone between the two species with highly
110 variable population structures. Sas and colleagues [11] reported a pure hybrid population
111 around Oradea, Romania. Evidence of hybridizing populations from other sections of the
112 Carpathians were also shown [17,19,22].

113 Our aim was to explore a transient zone between *B. bombina* and *B. variegata* and
114 assess the distribution of the two species and their hybrids from the inner part of the
115 Carpathian Basin to the southern edge of the Carpathian Ridges along an approximately
116 50 km long north-south transect (Fig 1). We also aimed to test a novel molecular approach
117 to distinguish between the two species and determine hybrid individuals.

118 **Fig 1. Map of the sampling area.** Circles indicate the assessed locations (L1-21) and
119 empty circles represent locations where individuals were not observed (L22-L28).
120 Squares represent settlements. White, light grey and dark grey areas represent the relief
121 under 200 m, between 200-400 m and above 400 m, respectively. Locations containing

122 hybrid individuals are underlined.

123 **Materials and Methods**

124 **Study area**

125 Börzsöny Hills are located in northern Hungary. In the north they are separated
126 from Krupinská Planina in Slovakia by Ipeľ River's valley, which also demarcates the
127 border between the two countries (Fig 1.). The highest elevation in Börzsöny is 938 m asl
128 and, due to its rugged terrain, most of the valleys are strongly shaded with cold
129 microclimates. Krupinská Planina is a sibling mountain of Börzsöny in Slovakia having
130 identical rock material; a Miocene andesite. It is lower (535 m asl), but its terrain is
131 heavily rugged as well. Published herpetofaunistic data from Börzsöny is old [27] and
132 while plenty of data were collected in an online database (<https://herpterkep.mme.hu>), no
133 systematic assessment was made for *Bombina* species prior to this study. The
134 herpetofauna of Krupinská Planina had hardly been explored before we started our
135 investigation. The only record was available from Slovakian sources is a summary of a
136 2-year (2008-2009) data collection. Both *Bombina* species had been found in the survey,
137 but the author did not analyse their presence deeper [28].

138 **Sample collection**

139 We searched for *Bombina* habitats based on literature sources and an online data
140 base (herpterkep.mme.hu). Potential habitats were marked on satellite maps
141 (GoogleEarth). In total, we visited 28 locations (S1 Table) and sampled 230 toads
142 between 2014 and 2018. Individuals were caught by hand and DNA samples were
143 collected using a non-invasive buccal swabbing. Each individual was placed into a
144 transparent plastic box and fixed by gently pressing a plastic sponge against their back.

145 The ventral pattern of each toad was photographed for further analyses (not presented
146 here, S1 Fig and they were then released at the capture site).

147 **Amplification and analysis of the nuclear *Ncx-1* gene**

148 The collected swabs were stored in 70% ethanol until the DNA extraction by
149 phenol-chloroform method [29]. The following primers were used to amplify a 846 base
150 pair (bp) long region of the *Ncx-1* gene: NcxF (5'- TCATCCGCTCCTGAAATTCT -3')
151 and NcxR (5'- CACAGTCCCACAGTTTTCCA -3') [18]. All PCR reactions were
152 implemented with MyTaq Ready Mix (Bioline) according to the manufacturer's
153 instructions. The reactions contained 80 ng DNA and 8 μ M of each primer. The PCR
154 conditions were the following: an initial denaturation step at 95 °C for 5 min, 30 cycles
155 with denaturation at 95 °C for 20 sec, an annealing temperature of 62 °C for 20 sec, and
156 elongation at 72 °C for 20 sec and a final elongation at 72 °C for 2 min. The 846 bp long
157 PCR product was analysed in 1% agarose gel.

158 To find a restriction fragment length polymorphism (RFLP) loci *Ncx-1* gene,
159 haplotypes were downloaded from the National Center for Biotechnology Information
160 (NCBI) and aligned to a consensus sequence both in *B. bombina* and *B. variegata* with
161 ClustalW2 (EMBL-EBI). Three haplotypes were obtained from *B. bombina* and 16
162 sequences originated from *B. variegata* according to Fijarczyk et al. [18]. The GenBank
163 accession numbers can be found in S2 Table. The created *B. bombina* and *B. variegata*
164 *Ncx-1* consensus sequences were compared pairwise with MultAlign [30]. With
165 NEBCutter [31] and dCAPS Finder [32] four HpyCH4V restriction enzyme sites
166 (TG^CA) were detected in *B. bombina*, with one out of four missing from the *B.*
167 *variegata* sequences (S2 Fig). The amplified PCR fragments were digested with
168 HpyCH4V enzyme (New England Biolab) at 37 °C for 3 hours. 10 μ l of each digestion

169 reaction were loaded into 2.5 % TAE agarose gel and GeneRuler™ 1 kb Plus DNA
170 Ladder (Fermentas) was used as a molecular weight marker. The digestion results in a
171 694 bp long fragment in case of *B. variegata*. *B. bombina* samples cleave to a 369 bp and
172 325 bp long fragments. The samples of hybrid individuals carry both the 694 bp, 369 bp
173 and the 325 bp long fragments. The remaining 152 bp split into 96 bp, 32 bp and 24 bp
174 long fragments in both genotypes (Fig 2).

175 **Fig 2. Electrophoretic differentiation of the digested PCR fragments specific to Ncx-**
176 **1 gene.** The amplified PCR product from Ncx-1 gene before enzymatic digestion: *B.*
177 *variegata* (Bv; identifier of the individual: 101), *B. bombina* (Bb; identifier of the
178 individual: 189) and hybrid (Bv × Bb; identifier of the individual: 169) Fragments
179 resulted by the digestion with HpyCH4V restriction enzyme: Bv* (101), Bv × Bb* (169),
180 Bb* (189). MM - molecular weight marker.

181 **Amplification of the mitochondrial fragments**

182 Six samples were processed at each location where one species emerged. All the
183 samples were analysed at the hybrid locations to detect the types of the mitochondria that
184 occur. The primers were based on the total *B. bombina* and *B. variegata* mitochondrial
185 genome at NCBI (EU115993.1, NC_009258.1). Two mitochondrial primer pairs were
186 used to distinguish the two species: RadF (5'- CAGCTAGTATCAACCCACCAGAT -
187 3') and RadR (5'- TTGATCTGTTGCTGGGTACGTCTTG -3') primers are specific for
188 *B. bombina*, TynF (5'- CAATAAAATTCAACCGCCAACAAT -3') and TynR (5'-
189 AAGTTGATCTGTTGCTGGGTATGTTCTA -3') primers are specific for *B. variegata*
190 mitochondria [33]. The PCR reactions were completed with Mytaq Read Mix (Bioline)
191 according to the manufacturer's instruction. The PCR cycles were the following in case
192 of the Rad primer pair: an initial denaturation step at 95 °C for 5 min, 35 cycles with

193 denaturation at 95 °C for 20 sec, an annealing temperature of 63 °C for 20 sec, and
194 elongation at 72 °C for 20 sec and a final elongation at 72 °C for 2 min. With the Tyn
195 primer pair the same PCR protocol was used except that the annealing temperature was
196 60 °C. The resulting DNA fragments were analyzed at 2.5 % agarose gel containing
197 ethidium-bromide with GeneRuler™ 1 kb Plus DNA Ladder (Fermentas). The RadF-R
198 primers amplify a 173 bp long fragment, the amplicon with the TynF-R primers is 196 bp
199 long (Fig 3).

200 **Fig 3. The amplification of mitochondrial regions with Tyn and Rad primer pairs**
201 **specific for *B. bombina* (Bb) and *B. variegata* (Bv) respectively.** According to the
202 mitochondrial marker the individual 189 carries Bb genotype, while the individual 101
203 carries Bv genotype. Individual 169 and 175 are hybrids. MM - molecular weight marker.

204 **Sequencing**

205 The PCR amplified *Ncx-1* gene fragments were sequenced by Sanger sequencing
206 (Eurofins Genomics, Ebersberg, Germany), and resulting chromatograms analysed with
207 Chromas 2.6.5 software (Technelyium).

208 **Results**

209 **Identifying a new nuclear marker in *Ncx-1* gene**

210 Altogether, we analysed 230 samples from 21 sampling sites located in Hungary
211 and Slovakia (Table 1). We could successfully amplify an 846 bp long fragment of the
212 *Ncx-1* gene in all 230 cases. The designed restriction enzyme digestion with HpyCH4V
213 could distinguish between the consensus sequences assigned to the two species and the
214 hybrid individuals. To validate our results, we Sanger sequenced 4 *B. variegata*, 7 *B.*
215 *bombina* and all the 26 hybrid individuals. The sequencing confirmed the result of the

216 *Ncx-1* digestion in all cases as the heterozygous locus was specific to the individuals
217 which were considered hybrids based on the digestion fragment composition (S2 Fig).

218 **Table 1. Species identification results according to nuclear marker *Ncx-1* at each**
219 **location.**

Location	Sample size	Nuclear marker		
		Bb	Bv	H
L1	29	29	-	-
L2	1	1	-	-
L3	2	2	-	-
L4	11	11	-	-
L5	15	15	-	-
L6	5	5	-	-
L7	10	10	-	-
L8	10	10	-	-
L9	15	15	-	-
L10	5	5	-	-
L11	6	6	-	-
L12	19	14	-	5
L13	20	20	-	-
L14	15	15	-	-
L15	3	2	-	1
L16	9	-	6	3
L17	34	6	14	14
L18	7	-	6	1
L19	2	-	2	-
L20	10	5	4	1
L21	2	-	1	1
Sum	230	171	33	26

220 Abbreviations: Bb: *Bombina bombina*, Bv: *Bombina variegata*, H: hybrid individual

221 mtDNA markers

222 One hundred and nineteen samples were analysed with two mitochondrial markers
223 from 14 locations (including all 7 locations with hybrids and half of the locations where
224 only *B. bombina* was found) with the two primer pairs that can distinguish between the
225 *B. bombina* and *B. variegata* mitochondria. The analysed 119 samples showed that the

226 phenotype and the *Ncx-1* gene results agreed with the mitochondrial markers at all 7 tested
227 locations where we found only *B. bombina* individuals (Table 1). Among the 7 sites with
228 hybrids, at one location the mitochondrial markers revealed *B. bombina* genotype, at five
229 locations the results showed *B. variegata* mitochondria. There was one location at the
230 hybrid zone - Plášťovce (L12) - where we detected both *B. bombina* and *B. variegata*
231 mitochondrion. Table 2 summarizes the result of the analyses based on the two
232 mitochondrial markers in 22 hybrid individuals (in 4 individuals we failed to amplify
233 mtDNA; see S3 Table for more details).

234 **Table 2. Combined mitochondrial and nuclear genotypes of individuals from**
235 **locations where hybrids occurred.**

Nuclear marker	Mitochondrial marker	Sample size	Location ID
Bb	Bb	14	L12, L15, L17
Bb	Bv	7	L17, L20
Bv	Bv	27	L16, L17, L18, L20, L21
H	Bb	3	L12, L15
H	Bv	19	L12, L16, L17, L18, L20

236 Abbreviations: Bb: *Bombina bombina*, Bv: *Bombina variegata*, H: hybrid individual

237 **Distribution of the two species in the study area**

238 Based on our species identification method, *Bombina bombina* occurred at 17
239 locations (including all locations in Börzsöny Hills), while *B. variegata* occurred at six
240 locations. In 13 locations all individuals proved to be *B. bombina*. At one location
241 (Čekovce) we found only *B. variegata*. We identified 7 locations where hybrid
242 individuals occurred. In total, we caught 26 hybrid individuals identified with the *Ncx-1*
243 nuclear gene marker at seven locations: Plášťovce, Brezovo, Čabrad, Litava, Medovarce,

244 Pribelce, Chrťany. At Litava and Pribelce we found both *B. bombina*, *B. variegata* and
245 also hybrid toads.

246 The examined toad populations in the Börzsöny Hills and the Ipel' Valley consist
247 of *B. bombina* specimens, and both species were found in the Krupinská Planina area. *B.*
248 *variegata* appears exclusively in the coolest, remote mountainous areas of the region,
249 while the edges of the plateau at even higher elevations were occupied by *B. bombina*.
250 On average, *B. variegata* occupied higher elevations than *B. bombina*, but the two species
251 reached the same maximum altitude. Hybrids occurred at intermediate altitude between
252 the two species median elevations (Fig 4). However, there was no significant difference
253 between the altitudinal distributions of the two species and their hybrids (one-way
254 ANOVA, for all location: $F_{2,27} = 0.473$; $P = 0.63$; only for locations in Krupinská Planina,
255 excluding *B. bombina* sites in Börzsöny and Ipel' Valley: $F_{2,18} = 1.064$; $P = 0.37$). In the
256 northern half of the Krupinská Planina we investigated 7 more locations (all artificial
257 ponds), where we did not find any *Bombina* specimens (Fig 1).

258 **Fig 4. The altitudinal distribution of the two species and their hybrids in the two**
259 **areas.** Horizontal line is the median, whiskers represent range, boxes represent
260 interquartiles and the circle indicates an outlier (deviating from the boundary of the
261 interquartile range (IQR) by more than $1.5 \times \text{IQR}$).

262 Discussion

263 Our study has four main achievements. First, using restricted digestion of a
264 nuclear gene we invented a new genetic test for the identification of *B. bombina* and *B.*
265 *variegata* and their hybrids. Second, we successfully applied this method on *Bombina*
266 samples from a putative hybrid zone and identified the two homozygous and the
267 heterozygous genotypes. Third, we compared the nuclear genotypes of the sampled

268 individuals with their haplotypes to a mitochondrial marker and found various genomic
269 combinations in the actual hybrid zone. Fourth, we identified that, in contrast to Börzsöny
270 Hills, Krupinská Planina is inhabited by both *B. bombina* and *B. variegata* populations
271 which produce hybrids in their contact zones.

272 **Evaluation of molecular identification**

273 The distribution patterns we explored are characteristically different at the two
274 sides of the Ipeľ Valley. While in Börzsöny Hills (Hungary) we found only pure *B.*
275 *bombina* populations even at the highest locations and in the smallest wheel track water
276 bodies, in Krupinská Planina (Slovakia) the distribution of the two species and hybrids
277 show a clear geographic and altitudinal pattern. *B. bombina* occupies the lowest and more
278 southern locations, while *B. variegata* occurs at the most northern location and highest
279 altitude, while intermediate locations (both in altitudinal and geographic terms) are
280 inhabited by hybrid populations, where *B. variegata* appear to be the more abundant
281 parent species. It is an interesting question if Börzsöny Hills harboured *B. variegata*
282 populations in the past. Today's remaining *B. variegata* populations in northern Hungary
283 are mostly found in ranges including Zemplén or Aggtelek which are continuous with the
284 Slovakian Carpathians. However, there is still *B. variegata* in Mátra Hills, a range very
285 similar to Börzsöny in its more isolated location and geography. If *B. variegata* occurred
286 in Börzsöny Hills in the past, it has been most likely outcompeted by its congener, which
287 may be a potential genetic outcome of the introgression if environmental conditions
288 favour one of the species. However, we did not detect hybridization introgression, as we
289 only found pure *B. bombina* specimens in Börzsöny, based on both the nuclear and
290 mitochondrial genomes.

291 In contrast, in the Krupinská Planina the greatest frequency of hybrid individuals
292 has *B. variegata* mitochondrial haplotype. This suggests that most hybrids have *B.*
293 *variegata* maternal origins, implying that hybridization arose from matings between *B.*
294 *variegata* females and *B. bombina* males. This is consistent with the pattern in other frog
295 species, where males disperse further than females [34], and that indiscriminate, coercive
296 male frogs readily mate with heterospecific females both under experimental and natural
297 conditions [35,36]. As hybrids occurred mostly alongside *B. variegata* individuals, it
298 seems likely that *B. bombina* genes introgressed into these populations via occasional
299 dispersal of *B. bombina* individuals.

300 Climate change will alter (or most probably already is altering) the recent pattern
301 of habitats and biotopes of Europe and the entire globe as it has become clear by the end
302 of the 20th century [37,38]. As a response, several species will certainly shift their range
303 adapting to the new circumstances [39,40]. Although in many cases studies revealed rapid
304 and sometimes irreversible decline in amphibian populations as a consequence of global
305 warming [41–43], some species will respond by dispersing and expanding their ranges
306 [44]. The current isolated distribution patches of *B. variegata* probably resulted from the
307 recent spreading of its congener, which outcompeted it from lower elevations of its former
308 distribution area [8]. This scenario may also have occurred in the Börzsöny Hills,
309 according to our results. In our case, we posit that if characteristically lowland species (*B.*
310 *bombina*) colonize habitats where the ecological conditions would be more favourable
311 for highland species (*B. variegata*), it reflects a current area expansion of the first species.
312 Possible area expansion of *B. bombina* has been documented during the last decade at the
313 northern edge of its range [45]. In Brandenburg province of Eastern Germany a detailed
314 analysis suggested a possible area expansion of *B. bombina*, as further habitats are

315 becoming suitable for the species due to predicted changes in climate and associated
316 future environmental conditions [46].

317 As we did not investigate *Bombina* dispersal processes between ponds in our study
318 area, nor their individual or environmental factors, we can only speculate that absence of
319 either species in the artificial ponds in the northernmost portion of our study site is a result
320 of (i) the relatively short time since these ponds have been established coupled with
321 relatively short dispersal distances of *Bombina* species [47], (ii) barriers to dispersal
322 within the landscape matrix including unsuitable microclimates and habitat for facilitating
323 movement [48], and/or (iii) the fact that these permanent ponds were artificial, deeper and
324 moderate in size which are less likely to be utilized by *B. variegata* in such contexts [49].
325 Nevertheless, we feel that extended investigation of ongoing patterns of dispersal,
326 occupancy and interspecific dynamics in such hybrid zones is warranted.

327 Both *Bombina* species are considered reliable indicators of habitat quality and
328 listed under the European Union's Council Directive 92/43/EEC on the conservation of
329 natural habitats and of wild fauna and flora (<https://eur-lex.europa.eu>). Being species of
330 community interest, their population sizes should be regularly monitored. Our research
331 highlights that special attention should be paid to hybrid zones, where changes of
332 population composition may reflect effects of recent rapid environmental alterations,
333 most notably climatic changes. As morphological identification of individuals could be
334 problematic due to the large overlap between phenotypes, our new and simple genetic
335 method provides a useful tool to track the hybrid populations genetic composition.

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493

494 **Supporting information**

495 **S1 Fig. Photos of three individuals (Bv, Bv x Bb, Bb).** Sampling locations and
496 individual IDs are indicated at the photos.

497 **S2 Fig. Side-by-side alignment of Bv consensus and Bb consensus sequence of the**
498 ***Ncx-1* gene.** Stars represent the distinct nucleotides. Underlined regions mark the
499 recognition site of the HpyCH4V restriction enzyme.

500 **S1 Table. The name and geocoordinates of sampling locations.**

501 **S2 Table. GenBank accession numbers of the sequences used to align the consensus**
502 **sequence of the *Ncx-1* gene.**

503 **S3 Table. Combined mitochondrial and nuclear genotypes of individuals at each**
504 **locations where hybrids were found.** In case of 2 individual in L12, 6 individuals in
505 L20 and 1 individual in L17, L18 and L21 we failed to amplify any mtDNA. In L12,
506 L17 and L18 the individuals was 1hybrids according to the nuclear marker.

507 **S4 Table. Sample size of individuals of which mtDNA amplification was failed at**
508 **each location of the hybrid zone**

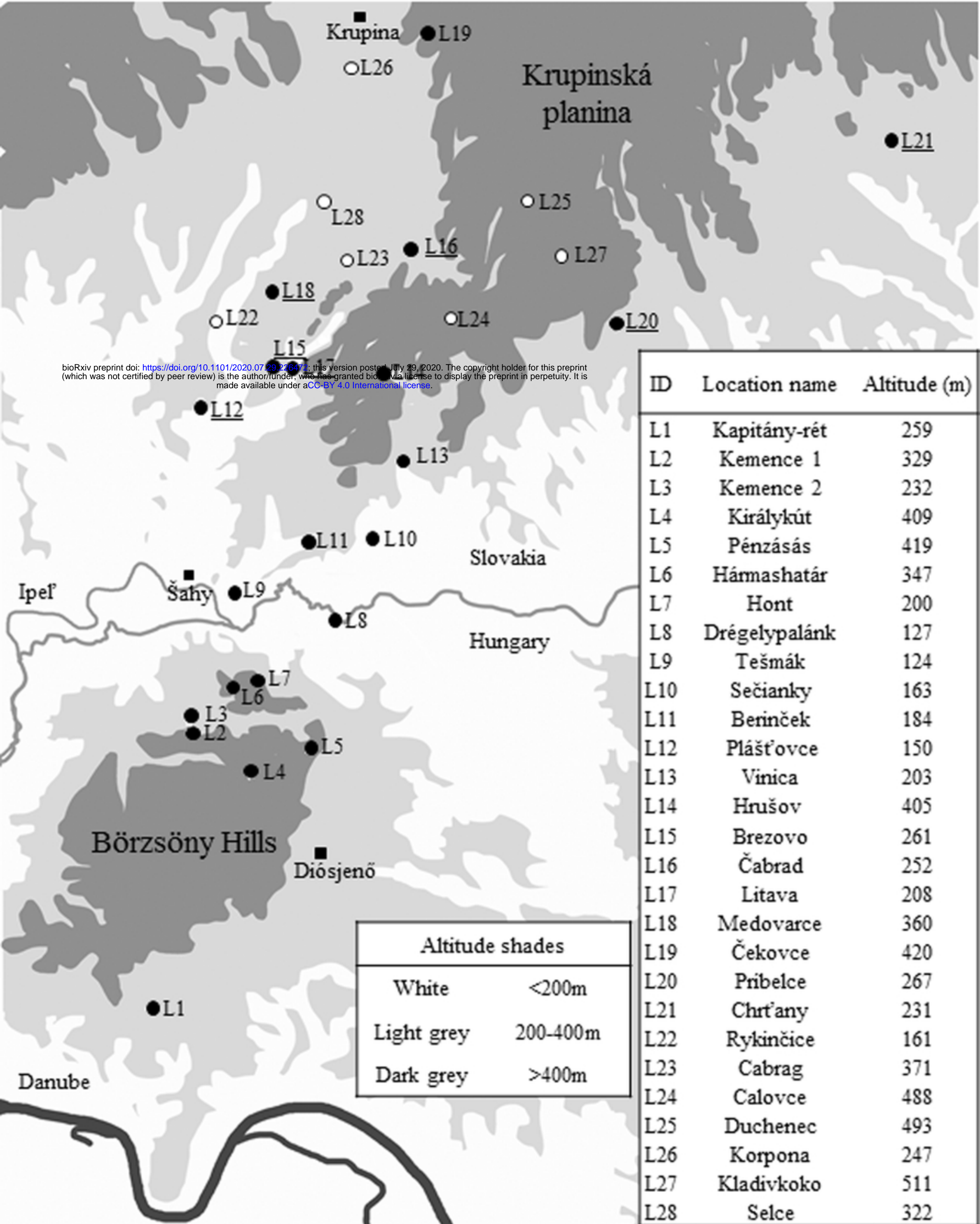


Fig1

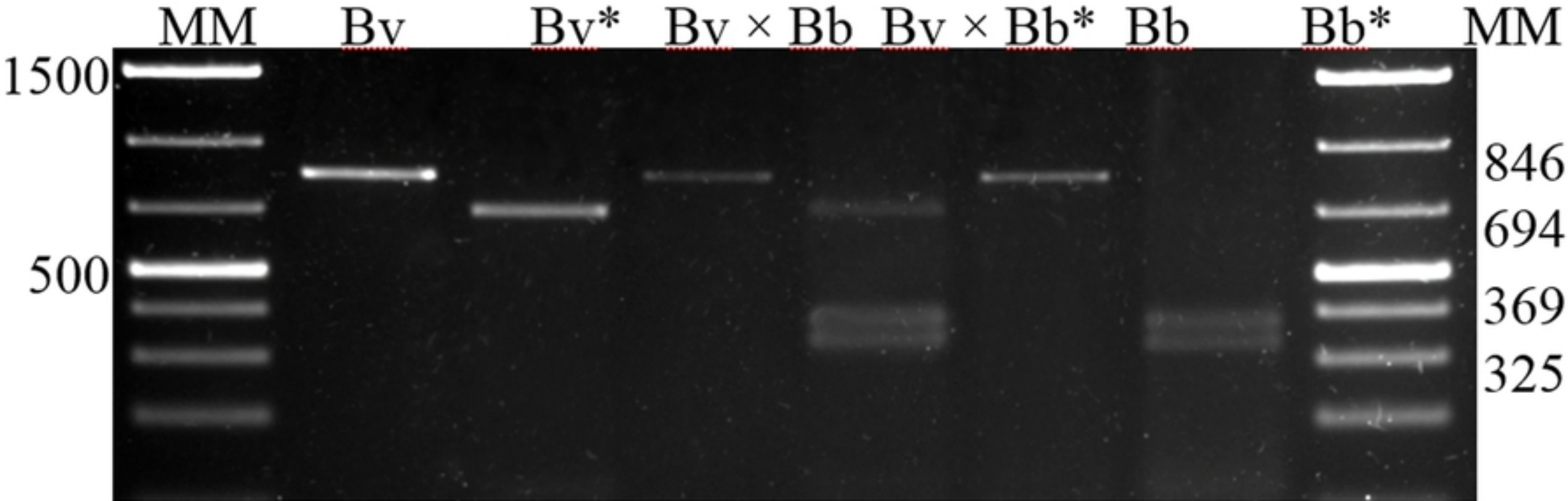


Fig2

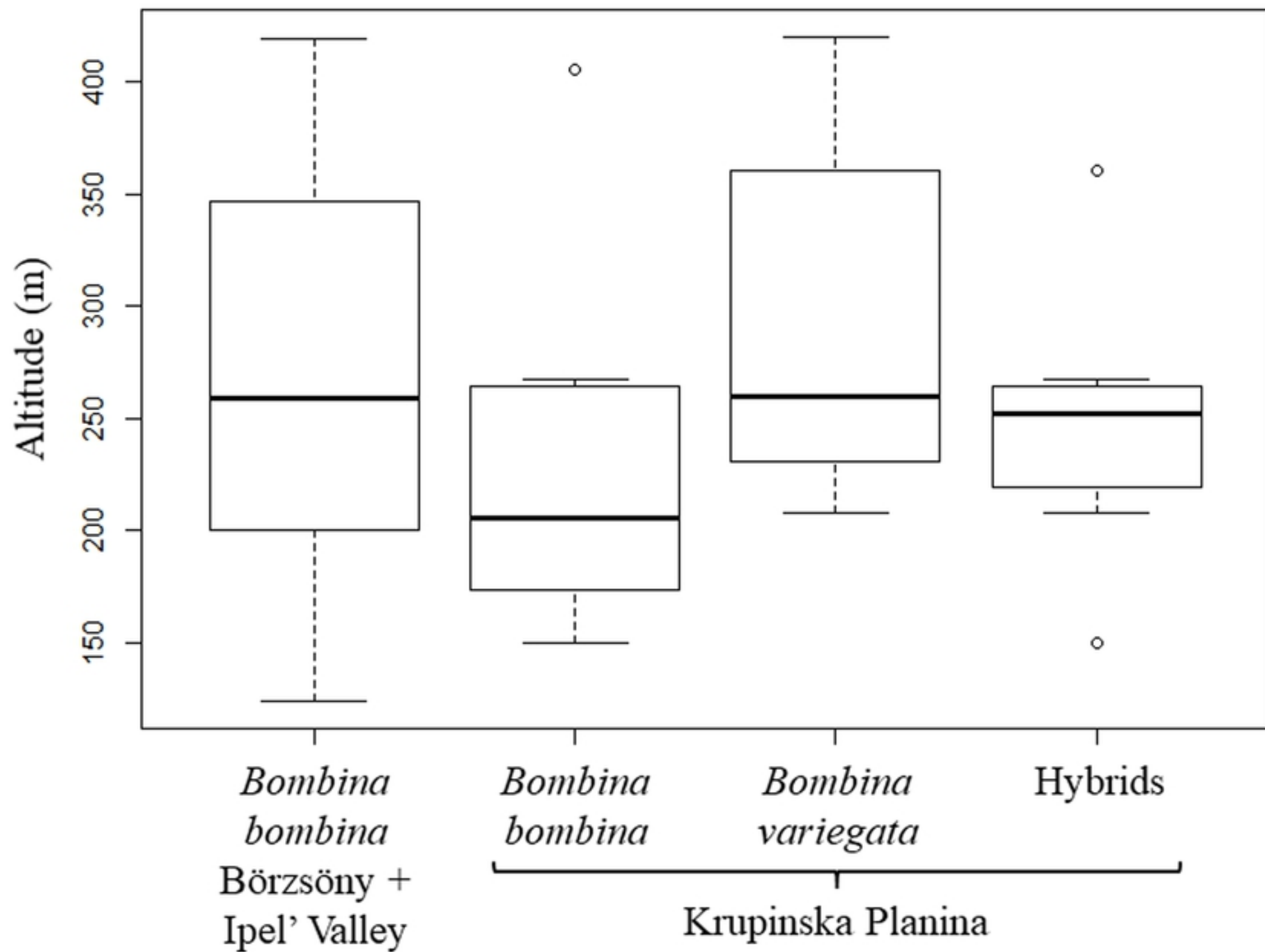


Fig4