

1 **Assessing Confidence in Root Placement on Phylogenies:** 2 **An Empirical Study Using Non-Reversible Models**

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10 ABSTRACT

11 Using time-reversible Markov models is a very common practice in phylogenetic
12 analysis, because although we expect many of their assumptions to be violated by empirical
13 data, they provide high computational efficiency. However, these models lack the ability to
14 infer the root placement of the estimated phylogeny. In order to compensate for the inability of
15 these models to root the tree, many researchers use external information such as using outgroup
16 taxa or additional assumptions such as molecular-clocks. In this study, we investigate the utility
17 of non-reversible models to root empirical phylogenies and introduce a new bootstrap measure,
18 the *rootstrap*, which provides information on the statistical support for any given root position.

19 Availability and implementation: A python script for calculating rootstrap support values is
20 available at <https://github.com/suhanaser/Rootstrap>.

21 [phylogenetic inference, root estimation, bootstrap, non-reversible models]

22

23 MAIN TEXT

24 The most widely used method for rooting trees in phylogenetics is the outgroup method.
25 Although the use of an outgroup to root an unrooted phylogeny usually outperforms other
26 rooting methods (Huelsenbeck, et al. 2002), the main challenge with this method is to find an
27 appropriate outgroup (Watrous and Wheeler 1981; Maddison, et al. 1984; Smith 1994;
28 Swofford, et al. 1996; Lyons-Weiler, et al. 1998; Milinkovitch and Lyons-Weiler 1998).
29 Outgroups that are too distantly-related to the ingroup may have substantially different
30 molecular evolution than the ingroup, which can compromise accuracy. And outgroups that are
31 too closely related to the ingroup may not be valid outgroups at all.

32 It is possible to infer the root of a tree without an outgroup using molecular clocks
33 (Huelsenbeck, et al. 2002; Drummond, et al. 2006). A strict molecular-clock assumes that the
34 substitution rate is constant along all lineages, a problematic assumption especially when the
35 ingroup taxa are distantly related such that their rates of molecular evolution may vary.
36 Relaxed molecular-clocks are more robust to deviations from the clock-like behaviour
37 (Drummond, et al. 2006), although previous studies have shown that they can perform poorly
38 in estimating the root of a phylogeny when those deviations are considerable (Tria, et al. 2017).

39 Other rooting methods rely on the distribution of branch lengths, including Midpoint
40 Rooting (MPR) (Farris 1972), Minimal Ancestor Deviation (MAD) (Tria, et al. 2017), and
41 Minimum Variance Rooting (MVR) (Mai, et al. 2017). Such methods also assume a clock-like
42 behaviour; however, they are less dependent on this assumption as the unrooted tree is
43 estimated without it. Similar to inferring a root directly from molecular-clock methods, the
44 accuracy of those rooting methods decreases with higher deviations from the molecular-clock
45 assumption (Mai, et al. 2017).

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46 Other less common rooting methods that can be used in the absence of outgroup are:
47 rooting by gene duplication (Dayhoff and Schwartz 1980; Gogarten, et al. 1989; Iwabe, et al.
48 1989), indel-based rooting (Rivera and Lake 1992; Baldauf and Palmer 1993; Lake, et al.
49 2007), rooting species tree from the distribution of unrooted gene trees (Allman, et al. 2011;
50 Yu, et al. 2011), and probabilistic co-estimation of gene trees and species tree (Boussau, et al.
51 2013).

52 All the methods mentioned above, apart from the molecular-clock, infer the root
53 position independently of the ML tree inference. The only existing approach to include root
54 placement in the ML inference is the application of non-reversible models. Using non-
55 reversible substitution models relaxes the fundamental assumption of time-reversibility that
56 exists in the most widely used models in phylogenetic inference (Jukes and Cantor 1969;
57 Kimura 1980; Hasegawa, et al. 1985; Tavaré 1986; Dayhoff 1987; Jones, et al. 1992; Tamura
58 and Nei 1993; Whelan and Goldman 2001; Le and Gascuel 2008). This in itself is a potentially
59 useful improvement in the fit between models of sequence evolution and empirical data. In
60 addition, since non-reversible models naturally incorporate a notion of time, the position of the
61 root on the tree is a parameter that is estimated as part of the ML tree inference. Since the
62 incorporation of non-reversible models in efficient ML tree inference software is relatively new
63 (Minh, et al. 2020), we still understand relatively little about the ability of non-reversible
64 models to infer the root of a phylogenetic tree, although a recent simulation study has shown
65 some encouraging results (Bettisworth and Stamatakis 2020).

66 Regardless of the rooting method and the underlying assumptions, it is crucial that we
67 are able to estimate the statistical confidence we have in any particular placement of the root
68 on a phylogeny. A number of previous studies have sensibly use ratio likelihood tests such as
69 the Shimodaira-Hasegawa (SH) test (Shimodaira and Hasegawa 1999) and the Approximately
70 Unbiased (AU) test (Shimodaira 2002) to compare a small set of potential root placements,

71 rejecting some alternative root placements in favour of the ML root placement e.g.(Nardi, et
72 al. 2003; Steenkamp, et al. 2006; Jansen, et al. 2007; Moore, et al. 2007; Williams, et al. 2010;
73 Kocot, et al. 2011; Zhou, et al. 2011; Whelan, et al. 2015; Zhang, et al. 2018), these tests are
74 still somewhat limited in that they do not provide the level of support the data have for a certain
75 root position.

76 There is strong empirical evidence that molecular evolutionary processes are rarely
77 reversible (Squartini and Arndt 2008; Naser-Khdour, et al. 2019), but few studies have
78 explored the accuracy of non-reversible substitution models to root phylogenetic trees
79 (Huelsenbeck, et al. 2002; Yap and Speed 2005; Williams, et al. 2015; Cherlin, et al. 2018;
80 Bettisworth and Stamatakis 2020). Most studies that have looked at this question in the past
81 have focused on either simulated datasets (Huelsenbeck, et al. 2002; Jayaswal, et al. 2011;
82 Cherlin, et al. 2018; Bettisworth and Stamatakis 2020) or relatively small empirical datasets
83 (Yang and Roberts 1995; Yap and Speed 2005; Jayaswal, et al. 2011; Heaps, et al. 2014;
84 Williams, et al. 2015; Cherlin, et al. 2018). In both cases, the addressed substitution models
85 were nucleotide models, and to our knowledge, no study has yet investigated the potential of
86 amino acid substitution models in inferring the root placement of phylogenetic trees.

87 In this paper, we focus on evaluating the utility of non-reversible amino acid and
88 nucleotide substitution models to root the trees, and we introduce a new metric, the *rootstrap*
89 *support value*, which estimates the extent to which the data support every possible branch as
90 the placement of a root in a phylogenetic tree. Unlike previous studies that used Bayesian
91 methods with non-reversible substitution models to infer rooted ML trees (Heaps, et al. 2014;
92 Cherlin, et al. 2018), we will conduct our study in a Maximum likelihood framework using IQ-
93 TREE (Minh, et al. 2020). A clear advantage of the Maximum likelihood over the Bayesian
94 analysis is that there is no need for a prior on the parameter distributions, which sometimes can
95 affect tree inference (Huelsenbeck, et al. 2002; Cherlin, et al. 2018). Even though estimating

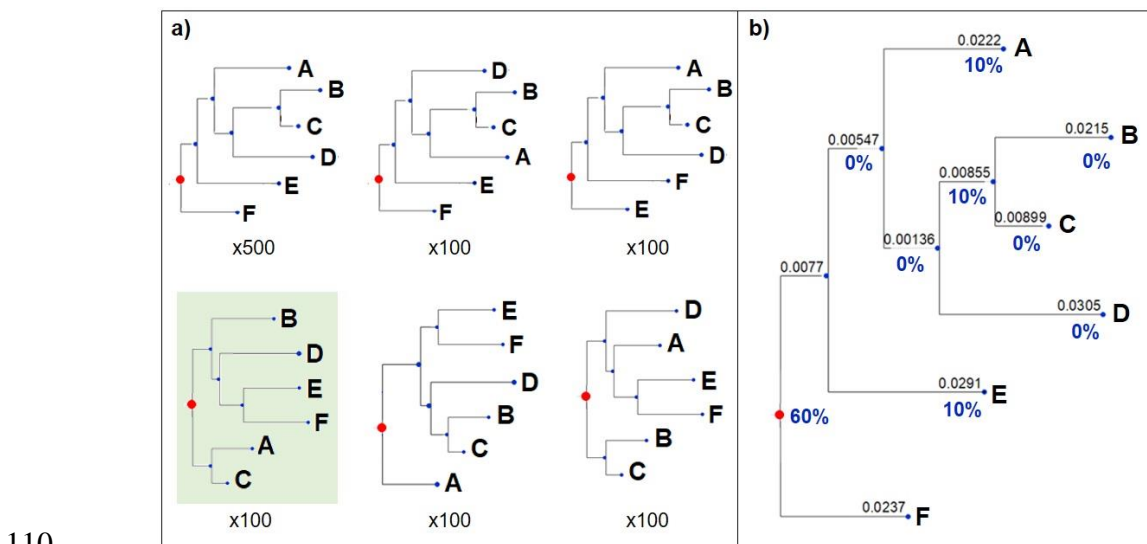
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96 the non-reversible model's parameters by maximizing the likelihood function seems more
97 computationally intensive than calculating posterior probabilities (Huelsenbeck, et al. 2002),
98 the IQ-TREE algorithm sufficiently fast to allow us to estimate root placements, with *rootstrap*
99 *support* for very large datasets.

100 MATERIAL AND METHODS

101 *The "Rootstrap" Support, and measurements of error in root placement*

102 To compute rootstrap supports, we conduct a bootstrap analysis, i.e., resampling alignment
103 sites with replacement, to obtain a number of bootstrap trees. We define the *rootstrap* support
104 for each branch in the ML tree, as the proportion of bootstrap trees that have the root on that
105 branch. Since the root can be on any branch in a rooted tree, the rootstrap support values are
106 computed for all the branches including external branches. The sum of the rootstrap support
107 values along the tree are always smaller than or equal to one. A sum that is smaller than one
108 can occur when one or more bootstrap replicates are rooted on a branch that does not occur in
109 the ML tree (Fig. 1).

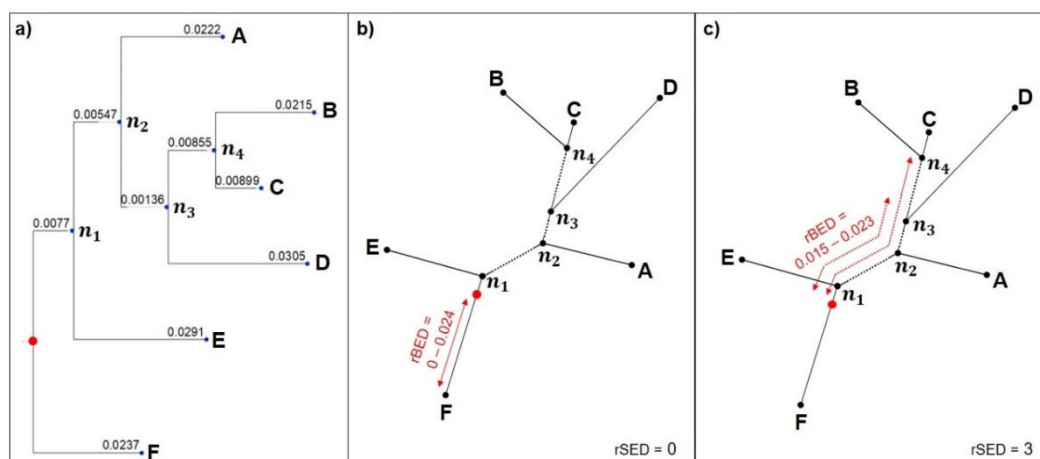


110
111 **FIGURE 1. Illustration of the rootstrap concept.** (a) The bootstrap replicates trees. (b)
112 The ML tree with the rootstrap support values for each branch. Note that the sum of the
113 rootstrap support values is less than 100% due to 100 bootstrap replicates trees (green)
114 have their root at a branch that does not exist in the ML tree.

115

116 If the true position of the root is known (e.g. in simulation studies) or assumed (e.g. in
117 the empirical cases we present below), we can calculate additional measurements of the error
118 of the root placement. We introduce two such measurements here: *root branchlength error*
119 *distance* (rBED) and *root split error distance* (rSED). Since the non-reversible model infers
120 the exact position of the root on a branch, we define the *root branchlength error distance*
121 (rBED) as the range between the minimum and maximum distance between the inferred root
122 position and the “true root” branch. If the true root is on the same branch as the ML tree root,
123 then rBED will be between 0 and the distance between the ML tree root and the farthest point
124 on that branch (Fig. 2). Since rBED is based on branch lengths only, it ignores the absolute
125 number of splits between the ML tree root and the true root; and therefore, the rBED for the
126 true root being on the same ML root branch can be bigger than the rBED for the true root
127 being on a different branch (e.g. Fig. 2). In order to account for the number of splits (nodes)
128 between the ML tree root and the true root, we define *root split error distance* (rSED) as the
129 number of splits between the ML root branch and the branch that is believed to contain the
130 true root (Fig. 2).

131



132 FIGURE 2. An example to illustrate the root error distance. (a) the ML rooted tree, (b) the
133 root branch-length error distance (rBED) if the true root is believed to be on the same ML

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134 root branch (rSED = 0), (c) the rBED if the true root is believed to be on the branch between
135 D and the clade of C + B (rSED = 3).

136

137 The rootstrap, rBED, and rSED assess different aspects of the root placement. While the
138 rootstrap offers an indication of the support that the data have for a certain branch to be the
139 root branch, rBED and rSED provide an estimation to the accuracy of the method in
140 estimating the exact root position if the root position is known or assumed in advance. In
141 other words, the rootstrap value is a measure for the adequacy of the data to validate a root
142 placement given the model, while rBED, and rSED are measures of the accuracy of the non-
143 reversible model to find the root placement given the data.

144 *Empirical Datasets*

145 Because non-reversible amino acid models require the estimation of a large number of
146 parameters, and because we suspected that the information in any such analysis on the
147 placement of the root branch of a tree might be rather limited, we searched for empirical
148 datasets that met a number of stringent criteria:

149 (1) Existence of both DNA and amino acid multiple sequences alignments (MSA) for the
150 same loci.

151 (2) Genome-scale MSAs to ensure that the MSAs have as much information as possible with
152 which to estimate the non-reversible models' free parameters and the root position. Since
153 we do not know the number of sites required to correctly infer the rooted ML tree, we
154 define 100,000 sites as the minimum number of required sites. This also allows us to
155 subsample the dataset to explore the ability of smaller datasets to infer root positions.

156 (3) Highly-curated alignments: since the quality of the inferred phylogeny is highly
157 dependent on the quality of the MSA (Philippe, et al. 2011), we focussed on datasets that
158 were highly-curated for misalignment, contamination, and paralogy.

159 (4) Existence of several clades for which there is a very strong consensus regarding their root
160 placement. Since we are interested in evaluating the performance of non-reversible
161 models to infer root placements in an empirical rather than a simulation context, we need
162 to identify monophyletic sub-clades for which we can be almost certain about their root
163 position. This enables us to divide the dataset into non-overlapping sub-clades for which
164 we are willing to assume we know the root positions. Furthermore, we define the
165 minimum number of taxa in each sub-dataset as five.

166 We initially identified a number of genome-scale datasets that contained large numbers of
167 nucleotide and amino acid MSAs. In many cases, it was difficult to determine whether these
168 alignments had been rigorously curated, and even more challenging to find datasets for which
169 the root position of a number of subclades could be assumed with confidence. The only dataset
170 that met all of our criteria was a dataset of placental mammals with 78 ingroup taxa and
171 3,050,199 amino acids (Wu, et al. 2019). This dataset was originally published as an MSA
172 (Liu, et al. 2017) based on very high-quality sequences from Ensembl, NCBI, and GenBank
173 databases. After receiving detailed critiques for potential alignment errors (Gatesy and Springer
174 2017), the dataset was further processed to remove potential sources of bias and error, and an
175 updated version of the dataset was recently published (Wu, et al. 2018). The fact that this
176 alignment comes from one of the most well-studied clades on the planet, has been
177 independently curated and critiqued by multiple groups of researchers and includes truly
178 genome-scale data, makes it ideally suited for our study.

179

180 *Selecting Clades with a Well-Defined Root*

181 Since our main objective in this study is to evaluate the effectiveness of non-reversible
182 models and the rootstrap value in estimating and measuring the support for a given root
183 placement on empirical datasets, we must identify a collection of sub-clades of the larger

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184 mammal dataset for which it is reasonable to assume a root position. We acknowledge, of
185 course, that outside a simulation framework it is not possible to be certain of the position of the
186 root position of a clade. Nevertheless, it is possible to identify clades for which the position of
187 the root is well supported and non-controversial, thus minimising the chances that the
188 assumption of particular root position is incorrect. To achieve this, we analysed the root
189 position of each order and superorder in the dataset, and defined “*well-defined clades*” that
190 fulfilled **all** of the following criteria:

- 191 (1) It contains at least five taxa. This ensures that the probability of obtaining a random ML
192 rooted tree to be at most 0.95%. For clades with four taxa, there are 15 different rooted
193 topologies, and therefore a 6.7% probability to get any of these topologies by chance. On
194 the other hand, for clades with at least five taxa, there are at least 105 different rooted
195 topologies and maximum probability of 0.95% to randomly get one of them as the ML
196 tree.
- 197 (2) The bootstrap support for the deepest two levels of branches leading to that clade in the
198 phylogenetic tree calculated from the whole dataset is 100%: since the bootstrap value
199 indicates the support the data have for a certain branch, we require 100% support for the
200 deepest two levels of branches leading to a certain clade in the whole tree (Appendix Fig.
201 A.1). This requirement ensures that there is strong support in the dataset for the root
202 position of the clade when the entire dataset is rooted with an outgroup.
- 203 (3) The gene concordance factor (gCF) and the site concordance factor (sCF) for the deepest
204 two levels of branches leading to the clade are significantly greater than 33%. The site
205 Concordance Factor (sCF) is calculated by comparing the support of each site in the
206 alignment for the different arrangements of quartet around a certain branch. In other
207 words, an sCF of 33% means equal support for any of the possible arrangements.

208 Therefore, we require that the sCF of the deepest two levels of branches leading to that
209 clade to be significantly greater than 33%. The gene Concordance Factor (gCF) of a
210 branch is calculated as the proportion of gene trees contain that branch. Although there is
211 no threshold regarding the required proportion of genes concordant with a certain branch,
212 for convenience, we define branches with gCF significantly greater than 33% as
213 branches that are concordant with enough genes in the alignment (Minh, et al. 2020). To
214 test whether the sCF and the gCF are significantly greater than 33%, we use a simple
215 binomial test with a success probability of 0.33.

216 (4) At least 95% of the studies that have been published in the last decade support this clade:
217 we searched google scholar for all published papers since 2009 that determine the root of
218 the addressed clade. We then checked if at least 95% of those papers agree that the root
219 position of the clade matches that in the ML tree we estimate from the whole dataset (see
220 supplementary material).

221 *Estimating unrooted Phylogenies*

222 For the whole nucleotide and amino-acid datasets with ingroup and outgroup taxa, we
223 inferred the unrooted phylogeny using IQ-TREE (Nguyen, et al. 2015) with the best-fit fully
224 partitioned model (Chernomor, et al. 2016) and edge-linked substitution rates (Duchene, et al.
225 2020). We then determined the best-fit reversible model for each partition using ModelFinder
226 (Kalyaanamoorthy, et al. 2017). See the algorithm for finding well-defined clades in
227 Appendix Algorithm A.1.

228 *Estimating Rooted phylogenies*

229 For each well-defined clade, we first removed all other taxa from the tree and then
230 sought to infer the root of the well-defined clade using non-reversible models without
231 outgroups. Using the best partitioning scheme from the reversible analysis, we inferred the

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232 rooted tree for each well-defined clade with the non-reversible models for amino acid (NR-
233 AA) and nucleotide (NR-DNA) sequences (Minh, et al. 2020). This approach fits a 12-
234 parameter non-reversible model for DNA sequences, and a 380-parameter non-reversible
235 model for amino acids. Details of the command lines used are provided in the supplementary
236 material section “Algorithm A.2”. Each analysis returns a rooted tree. We performed 1000
237 non-parametric bootstraps of every analysis to measure the rootstrap support.

238 To assess the performance of the rootstrap and the ability of non-reversible models to
239 estimate the root of the trees on smaller datasets, we also repeated every analysis on
240 subsamples of the complete dataset. For each well-defined clade, we performed analysis on
241 the complete dataset (100%) as well as datasets with 10%, 1% and 0.1% of randomly-
242 selected loci from the original alignment.

243 *The confidence set of root branches using the Approximately Unbiased test*

244 In addition to the rootstrap support, we calculate the confidence set of all the branches
245 that may contain the root of the ML tree using the Approximately Unbiased (AU) test
246 (Shimodaira 2002). To do this, we re-root the ML tree with all possible placements of the root
247 (one placement for each branch) and calculate the likelihood of each tree. Using the AU test,
248 we then ask which root placements can be rejected in favour of the ML root, using an alpha
249 value of 5%. We define the *root branches confidence set* as the set of root branches that are not
250 rejected in favour of the ML root placement.

251 *Reducing systematic bias by removing third codon positions and loci that fail the MaxSym* 252 *test*

253 As it is common in many phylogenetic analyses to remove third codon positions from
254 the alignment (Swofford, et al. 1996), we wanted to assess the effect of removing third codon
255 positions on the root inference and the rootstrap values in nucleotide datasets. For that

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256 purpose, we remove all the third codon positions from the nucleotide alignments and re-ran
257 the analysis using the NR-DNA model.

258 Moreover, although the NR-AA and NR-DNA models relax the reversibility assumption,
259 they still assume stationarity and homogeneity. To reduce the systematic bias produced by
260 violating these assumptions, we used the MaxSym test (Naser-Khdour, et al. 2019) to remove
261 loci that violate those assumptions in the nucleotide and amino acid datasets, and then re-ran
262 all analyses as above.

263 *Applying the methods to two clades whose root position is uncertain*

264 In addition to the well-defined clades, we used the methods we propose here to infer
265 the root of two clades of mammals whose root position is controversial; Chiroptera and the
266 Cetartiodactyla.

267 There is a controversy around the root of the Chiroptera (bats) in literature. The two
268 most popular hypotheses are: 1) the Microchiroptera-Megachiroptera hypothesis; where the
269 root is placed between the Megachiroptera, which contains the family Pteropodidae, and the
270 Microchiroptera, which contains all the remaining Chiroptera families. This hypothesis is
271 well supported in the literature (Agnarsson, et al. 2011; Meredith, et al. 2011). However,
272 more recent studies seem to provide less support for this hypothesis; 2) the
273 Yinpterochiroptera-Yangochiroptera hypothesis, in which the Yangochiroptera clade includes
274 most of Microchiroptera and the Yinpterochiroptera clade includes the rest of
275 Microchiroptera and all of Megachiroptera. There is growing support for this hypothesis in
276 the literature (Meganathan, et al. 2012; Tsagkogeorga, et al. 2013; Ren, et al. 2018; Reyes-
277 Amaya and Flores 2019).

278 Similar to Chiroptera, the root of Cetartiodactyla remains contentious in the literature.
279 The three main hypotheses regarding the root of Cetartiodactyla are: 1) Tylopoda as the sister

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280 group for all other cetartiodactylans; 2) Suina as the sister group for all other
281 cetartiodactylans; 3) the monophyletic clade containing Tylopoda and Suina as the sister
282 group for all other cetartiodactylans.

283 To ascertain whether certain sites or loci had very strong effects on the placement of
284 the root we follow the approach of Shen et. al. (Shen, et al. 2017) and calculate the difference
285 in site-wise log-likelihood scores (Δ SLS) and gene-wise log-likelihood scores (Δ GLS) between
286 the supported root positions for each clade.

287

288 RESULTS

289 *Inference of the mammal tree and selection of well-defined clades*

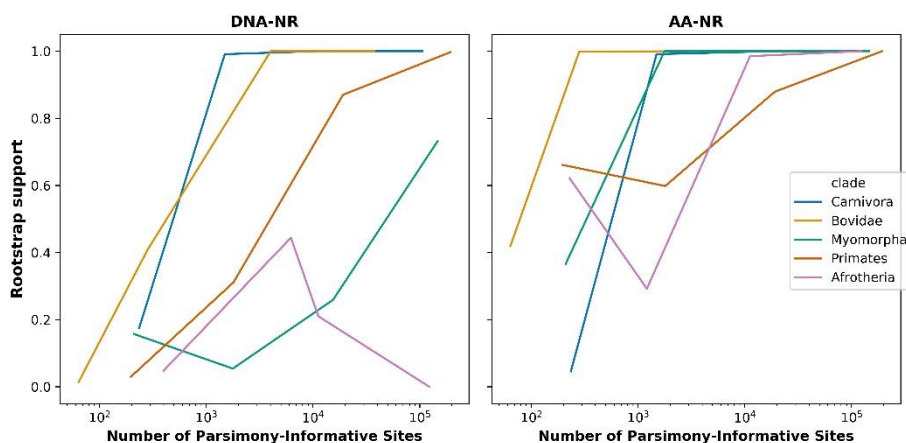
290 The trees inferred from the whole datasets with the nucleotide-reversible model and
291 the amino-acid-reversible model (Appendix Fig. A.2, Appendix Fig. A.3, Appendix Table
292 A.2) are consistent with the published tree (Liu, et al. 2017). Five clades met all the criteria of
293 well-defined clades, namely, Afrotheria, Bovidae, Carnivora, Myomorpha, and Primates in
294 both amino acid and nucleotide datasets (see Appendix Table A.1 and Appendix Table A.2).

295 *High accuracy of the AA non-reversible model in inferring the root*

296 Using NR-AA, we inferred the correct root with very high rootstrap support for all
297 five well-defined clades (Appendix Table A.3). Moreover, for all the five clades, the true root
298 was the only root placement in the confidence set of the AU test.

299 Our results show that using only 10% of the sites in the amino acid alignments gave
300 very high rootstrap support values (> 98%) for most of these clades (Fig. 3). Moreover, in
301 some datasets, 1% of the sites were enough to give a very high rootstrap support value. Yet,
302 using only 0.1% decreased the rootstrap support value noticeably in all datasets (Appendix

303 Table A.3). These values are shown for each dataset in Figure 3, where the X-axis is plotted
 304 in terms of parsimony-informative sites to allow for a more direct comparison between
 305 datasets. Although the rootstrap support for the true root improves as the number of
 306 parsimony-informative sites increase, in some datasets (e.g. Afrotheria nucleotide dataset)
 307 this is not the case (Fig. 3).



308
 309 FIGURE 3. The rootstrap support value for each clade as a function of the number of
 310 parsimony-informative sites.

311

312

313 *Poor performance of the DNA non-reversible model in inferring the root*

314 We correctly inferred the root for four out of the five nucleotide datasets with the NR-
 315 DNA model. However, the rootstrap support was generally lower than in the amino-acid
 316 datasets (Fig. 3, Appendix Tables A.3 and A.4). In addition, our results show that removing
 317 the third codon positions does not improve the rootstrap support value. In contrast, in some
 318 datasets removing third codon positions decreased the rootstrap support value and increased
 319 the rSED (Table 1).

320 TABLE 1. Rootstrap support and rSED values in whole nucleotide datasets and
 321 nucleotide datasets without third codon positions.

Clades	All loci		Without 3rd	
	rootstrap	rSED	rootstrap	rSED
Afrotheria	0.0%	2	0.0%	2
Primates	90.1%	0	90.1%	0

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Myomorpha	15.8%	0	15.8%	1
Carnivora	100.0%	0	100.0%	0
Bovidae	82.5%	0	82.5%	0

322

323 *Removing loci that violate the stationarity and homogeneity assumptions improves the*
 324 *rootstrap support*

325 As expected, our results show that removing loci that fail the MaxSym test improves
 326 the root placement inference and the rootstrap support values when the rootstrap support
 327 value was less than 100% and/or the root placement was inferred incorrectly, as the case in
 328 some nucleotide datasets (TABLE 2).

329 TABLE 2. Rootstrap support values in whole datasets and datasets with loci that passed
 330 the MaxSym test only.

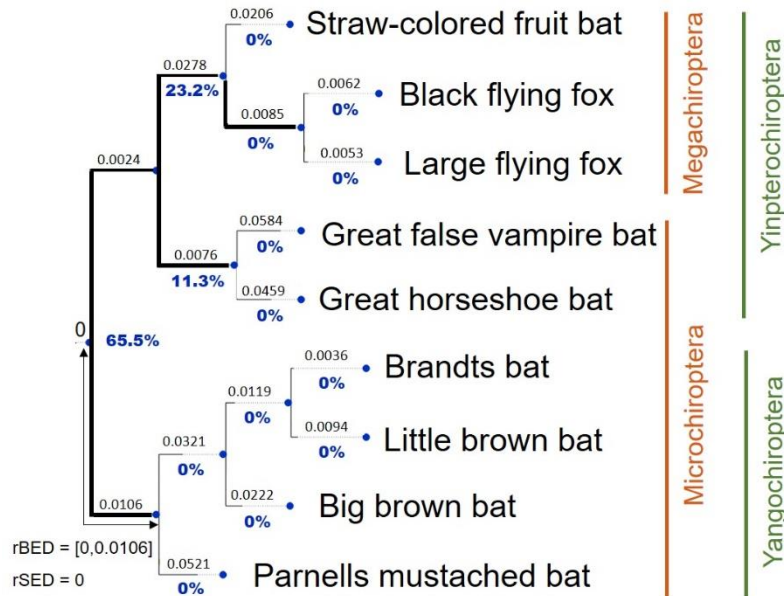
Clade	Amino Acid		Nucleotide	
	all loci	Passed MaxSym	all loci	Passed MaxSym
Afrotheria	100.0%	100.0%	0.0%	8.4%
Primates	100.0%	100.0%	99.7%	99.9%
Myomorpha	100.0%	100.0%	73.2%	88.3%
Carnivora	100.0%	100.0%	100.0%	100.0%
Bovidae	100.0%	100.0%	100.0%	100.0%

331

332 *Microchiroptera-Megachiroptera or Yinpterochiroptera-Yangochiroptera?*

333 Using the whole amino acid dataset, our results show 65.5% rootstrap support for the
 334 Yinpterochiroptera-Yangochiroptera hypothesis and 23.2% for the Microchiroptera -
 335 Megachiroptera hypothesis. The remaining 11.3% of the rootstrap support goes to supporting
 336 the branch leading to Rhinolophoidea as root branch of the bats (Fig. 4). Removing amino
 337 acid loci that fail the MaxSym test (110 loci) gives similar results, with 65.9% rootstrap
 338 support for the Yinptero-Yango hypothesis and 25.6% rootstrap support for the Micro-Mega
 339 hypothesis. In both cases, the AU test could not reject any of the three root positions that
 340 received non-zero rootstrap support (Appendix Table A.5).

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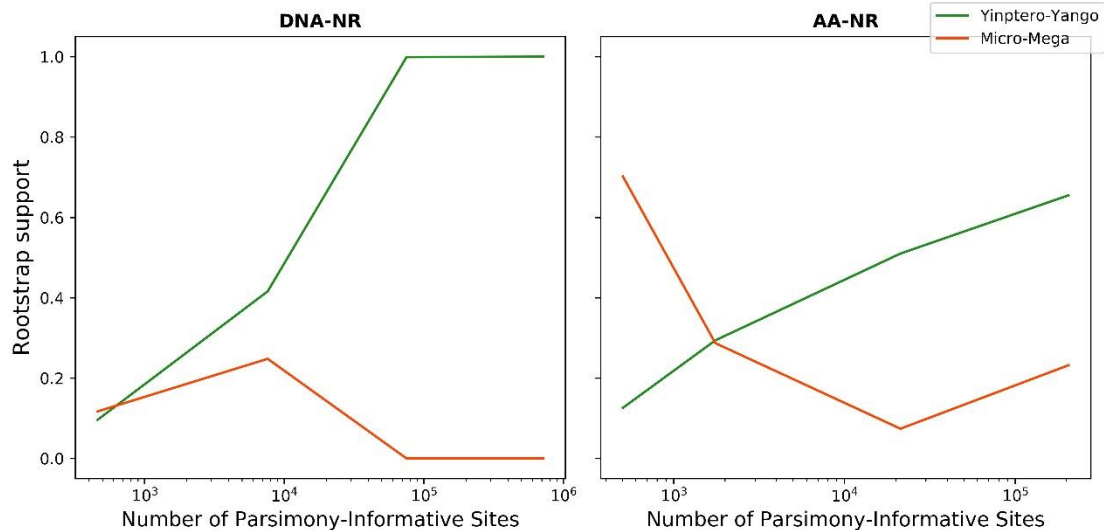
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342 FIGURE 4. The ML rooted tree as inferred from the whole Chiroptera amino acid dataset.
343 Bold branches are branches in the AU confidence set. Blue values under each branch are the
344 rootstrap support values.

345 Using the NR-DNA model gives 100% rootstrap support for the Yinptero-Yango
346 hypothesis, and we can confidently reject the Micro-Mega hypothesis in favour of the
347 Yinptero-Yango hypothesis using the AU test (Appendix Fig. A.4). Yet, removing nucleotide
348 loci that fail the MaxSym test (~25% of the loci) decreases the support for the Yinptero-
349 Yango hypothesis to 90.1%, although we can still confidently reject the Micro-Mega
350 hypothesis using the AU test (Appendix Table A.5).

351 Interestingly, when we randomly subsample 10%, 1%, and 0.1% of the loci in the
352 nucleotide dataset, we consistently get the Yinptero-Yango hypothesis as the ML tree and the
353 solely rooted topology in the AU confidence set (Appendix Table A.5). Moreover, the
354 rootstrap support value for the Yinptero-Yango hypothesis increases and the rootstrap support
355 value for the Micro-Mega hypothesis decreases as more parsimony-informative sites are
356 added to the alignment, for both nucleotide and amino acid datasets (Fig. 5, Appendix Table
357 A.5).

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358

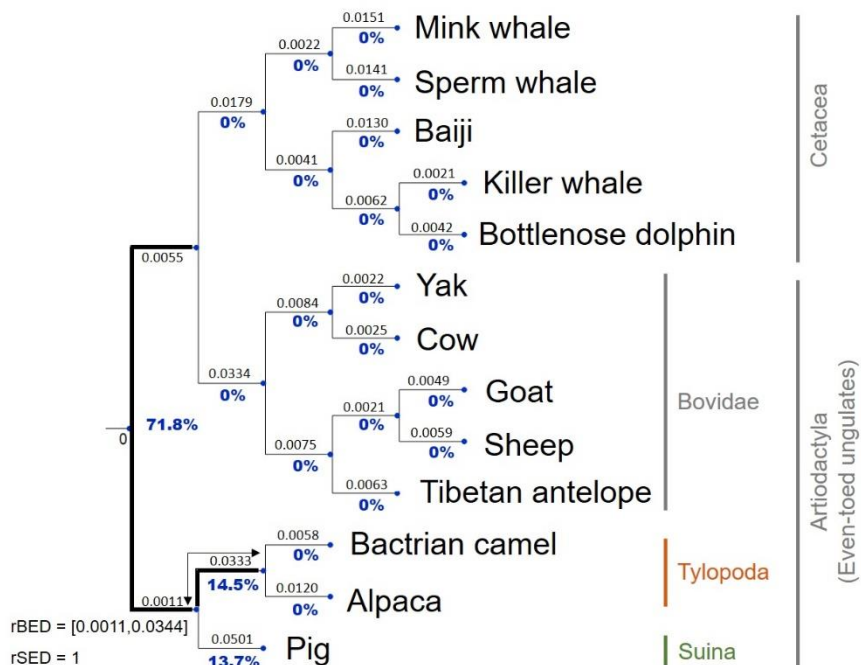
359 **FIGURE 5.** Rootstrap support value as a function of the number of parsimony-informative
360 characters in the Chiroptera nucleotide and amino acid datasets.

361 The Δ GLS and Δ SLS values (Shen, et al. 2017) reveal that approximately half of the
362 nucleotide and amino acid loci prefer the Yinptero-Yango hypothesis while the other half
363 prefers Micro-Mega hypothesis. Furthermore, slightly less than half of the nucleotide sites
364 prefer the Yinptero-Yango hypothesis. However, more than two-thirds of the amino acid sites
365 prefer the Yinptero-Yango hypothesis (Appendix Fig. A.5). The distributions of Δ GLS and
366 Δ SLS (Appendix Fig. A.6) show that a small proportion of the amino acid loci (~1%) have
367 very strong support for the Micro-Mega hypothesis, and removing those loci from the
368 alignment increased the rootstrap support for the Yinptero-Yango hypothesis to 76.6%.
369 Nonetheless, both root placements are still in the confidence set of the AU test (Appendix
370 Table A.5) with the amino acid dataset. On the other hand, removing nucleotide loci with the
371 highest absolute Δ GLS value still gives the Yinptero-Yango hypothesis as the ML tree and
372 the sole topology in the AU confidence set. We conclude that while the nucleotide data show
373 a clear preference to the Yinptero-Yango hypothesis, the amino acid data do not allow us to
374 distinguish between the two leading hypotheses for the placement of the root of the
375 Chiroptera based on rooting with non-reversible models.

376 *The ambiguous root of Cetartiodactyla*

377 The ML tree inferred with the whole amino acid dataset places the clade containing
378 Tylopoda (represented by its only extant family; Camelidae) and Suina as the most basal
379 cetartiodactylan clade with 71.8% rootstrap support (Fig. 6). Yet, The AU test did not reject
380 Tylopoda as the most basal clade. On the other hand, the ML tree inferred with the whole
381 nucleotide dataset places Tylopoda as the most basal clade with 71.0% rootstrap support, and
382 we can confidently reject the Tylopoda + Suina hypothesis using the AU test (Appendix Fig.
383 A.7).

384 Removing the amino acid loci that failed the MaxSym test (~1%) still places Tylopoda +
385 Suina as the basal-most clade, yet, it decreases the rootstrap support for the Tylopoda + Suina
386 hypothesis to 63.3% and increases the rootstrap support for the Tylopoda hypothesis to
387 28.5%. However, we still cannot reject either of the hypotheses using the AU test (Appendix
388 Table A.6).

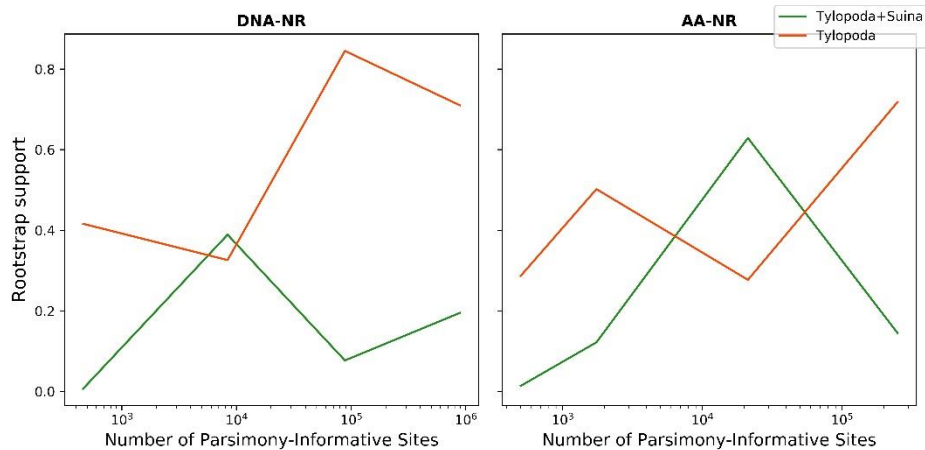


389

390 FIGURE 6. The ML rooted tree of as inferred from the whole Cetartiodactyla amino acid
391 dataset. Bold branches are branches in the AU confidence set. Blue values under each branch
392 are the rootstrap support values.

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393 Removing the nucleotide loci that failed the MaxSym test (~1%) still places Tylopoda
394 as the basal-most clade and the only rooted topology in the AU confidence set. However, it
395 decreases the rootstrap support for the Tylopoda hypothesis to 68.7% and increases the
396 rootstrap support for the Tylopoda + Suina hypothesis to 20.1% (Appendix Table A.6).
397 The results from the subsample datasets are mixed (Fig. 7). Analyses on smaller datasets
398 show no clear pattern in the placement of the root (Appendix Table A.6), leading us to
399 conclude only that the analyses of the whole dataset is likely to provide the most accurate
400 result, but that it is plausible that adding more data may lead to different conclusions in the
401 future.



402

403 FIGURE 7. rootstrap support value as a function of the number of parsimony-informative
404 characters in the Cetartiodactyla nucleotide and amino acid datasets.

405 Δ GLS analyses reveal that approximately, half of the amino acid and nucleotide loci
406 favour the Tylopoda+Suina hypothesis, while the other half of loci favour the Tylopoda
407 hypothesis (Appendix Figs. A.8-9). On the other hand, two-thirds of the amino acid sites and
408 more than 80% of the nucleotide sites favour the Tylopoda+Suina hypothesis. Removing 1%
409 of the amino acid loci with the highest absolute Δ GLS values still places Tylopoda + Suina as
410 the most basal clade. However, the rootstrap support of the Tylopoda + Suina decreased to
411 63.2% and the rootstrap support for the Tylopoda hypothesis remains approximately the same

412 (~14.5%), while the rootstrap support for the Suina hypothesis increases from 13.7% to
413 22.4%. Yet, both the Tylopoda + Suina hypothesis and the Tylopoda hypothesis are in the
414 confidence set of the AU test, while the Suina hypothesis is rejected by the AU test
415 (Appendix Table A.6).

416 Removing 1% of the nucleotide loci with the highest absolute Δ GLS values gives the
417 Tylopoda+Suina as the most basal clade of Cetartiodactyla with 39.7% rootstrap support.
418 However, the solely rooted topology in the AU confidence set is the topology in which the
419 root is placed on the branch leading to Suina (Appendix Table A.6). We conclude that neither
420 the nucleotide nor the amino acid data are adequate to infer the root placement of
421 Cetartiodactyla with non-reversible models.

422 DISCUSSION

423 In this paper, we introduced a new measure of support for the placement of the root in
424 a phylogenetic tree, the rootstrap support value, and applied it to empirical amino acid and
425 nucleotide datasets inferred using non-reversible models implemented in IQ-TREE (Minh, et
426 al. 2020). The rootstrap is a useful measure because it can be used to assess the statistical
427 support for the placement of the root in any rooted tree, regardless of the rooting method. In a
428 Maximum Likelihood setting, interpretation of the rootstrap support is similar to the
429 interpretation of the classic nonparametric bootstrap. In a Bayesian setting, the same
430 procedure could be used to calculate the posterior probability of the root placement given a
431 posterior distribution of trees. It is noteworthy that the rootstrap support value is not a
432 measure of the accuracy of the root placement and therefore should not be interpreted as
433 such. However, it provides information about the robustness of the root inference with regard
434 to resampling the data. This interpretation is consistent with the interpretation of the

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435 nonparametric bootstrap (Holmes 2003) but with regard to the root placement instead of the
436 whole tree topology.

437 In addition to the rootstrap support value, we introduced another two metrics; the root
438 branch-length error distance (rBED), and the root split error distance rSED. Similar to the
439 rootstrap metric, these additional metrics can be used in with any approach that generates
440 rooted phylogenetic trees. We note that both metrics require the true position of the root to be
441 known (or assumed) and that the rBED requires the rooting method to be able to accurately
442 place the root in a specific position of the root branch.

443 In this study, we used these and other methods to assess the utility of non-reversible
444 models to root phylogenetic trees in a Maximum Likelihood framework. We focussed on
445 applying these methods to a large and very well curated phylogenomic dataset of mammals,
446 as the mammal phylogeny provides perhaps the best opportunity to find clades for which the
447 root position is known with some confidence. As expected, our results show an exponential
448 increase in the rootstrap support for the true root as we add more information to the MSA.
449 Our results suggest that non-reversible amino-acid models are more useful for inferring root
450 positions than non-reversible DNA models, which is consistent with results from previous
451 simulations using the NR-DNA model (Bettisworth and Stamatakis 2020). One explanation
452 for this difference between the NR-DNA and the NR-AA models is the bigger character-state
453 space of the NR-AA models. These models have 400 parameters (380 rate parameters and 20
454 amino acid frequencies) whereas NR-DNA models have only 16 parameters (12 rate
455 parameters and 4 nucleotide frequencies). This could allow the NR-AA model to capture the
456 evolutionary process better than the NR-DNA model, potentially providing more information
457 on the root position of the phylogeny. This hypothesis requires some further exploration
458 though, and we note that the actual character-space of amino acids is much smaller than

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459 accommodated in NR-DNA models due to functional constraints on protein structure
460 (Dayhoff, et al. 1978).

461 Another explanation for the difference in performance between the NR-AA and NR-
462 DNA models is that higher compositional heterogeneity in nucleotide datasets may bias tree
463 inference. In principle, this bias can be alleviated by removing loci that violate the
464 stationarity and homogeneity assumptions (Naser-Khdour, et al. 2019). Our results suggest
465 that this may be the case for the datasets we analysed: we show that removing loci that
466 violate the stationarity and homogeneity assumptions improves the accuracy and statistical
467 support for the placement of the root. This is not surprising since the robustness of the
468 rootstrap, similar to the bootstrap, relies on the consistency of the inference method, so
469 removing systematic bias should improve its performance.

470 We used the non-reversible approach to rooting trees along with the rootstrap support
471 to assess the evidence for different root placements in the Chiroptera and Cetartiodactyla.
472 Using the amino acid datasets we found that in both cases, although there tended to be higher
473 rootstrap support for one hypothesis, neither of the current hypotheses for either dataset could
474 be rejected. These results emphasize the importance of the rootstrap support value as a
475 measure of the robustness of the root estimate given the data. In both the Chiroptera and
476 Cetartiodactyla datasets the root placement varied among subsamples of the dataset, and the
477 rootstrap support reflects this uncertainty. In both cases, the amino acid data is inadequate to
478 distinguish between certain root placements. On the other hand, in both the Chiroptera and
479 Cetartiodactyla, the nucleotide datasets appear to show stronger support for a single root
480 placement. This difference between the amino acid and the nucleotide datasets results may be
481 due to greater phylogenetic signal contained in the nucleotide characters.

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482 Our results demonstrate that both non-reversible models can be surprisingly useful for
483 inferring the root placement of phylogenies in the absence of additional information (such as
484 outgroups) or assumptions (such as molecular clocks). Indeed, we show that root placements
485 appear to be accurate even with fairly datasets as small as 50 well-curated loci between fairly
486 closely-related taxa such as orders of mammals. We hope that the combination of non-
487 reversible models and rootstrap support will add another tool to the phylogeneticist's arsenal
488 when it comes to inferring rooted phylogenies.

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492 REFERENCES

- 493 Agnarsson I, Zambrana-Torrel CM, Flores-Saldana NP, May-Collado LJ. 2011. A
494 time-calibrated species-level phylogeny of bats (Chiroptera, Mammalia). *PLoS*
495 *Curr* 3:RRN1212.
- 496 Allman ES, Degnan JH, Rhodes JA. 2011. Identifying the rooted species tree from the
497 distribution of unrooted gene trees under the coalescent. *J. Math. Biol.* 62:833-
498 862.
- 499 Baldauf SL, Palmer JD. 1993. Animals and fungi are each other's closest relatives:
500 congruent evidence from multiple proteins. *Proceedings of the National*
501 *Academy of Sciences* 90:11558-11562.
- 502 Bettisworth B, Stamatakis A. 2020. RootDigger: a root placement program for
503 phylogenetic trees. *bioRxiv*:2020.2002.2013.935304.
- 504 Boussau B, Szollosi GJ, Duret L, Gouy M, Tannier E, Daubin V. 2013. Genome-scale
505 coestimation of species and gene trees. *Genome Res.* 23:323-330.

NASER-KHDOUR ET AL.

- 506 Cherlin S, Heaps SE, Nye TMW, Boys RJ, Williams TA, Embley TM. 2018. The Effect
507 of Nonreversibility on Inferring Rooted Phylogenies. *Mol. Biol. Evol.* 35:984-
508 1002.
- 509 Chernomor O, von Haeseler A, Minh BQ. 2016. Terrace Aware Data Structure for
510 Phylogenomic Inference from Supermatrices. *Syst. Biol.* 65:997-1008.
- 511 Dayhoff M. 1987. A model of evolutionary change in proteins. *Atlas of protein*
512 *sequence and structure* 5:suppl. 3.
- 513 Dayhoff M, Schwartz R. 1980. Prokaryote evolution and the symbiotic origin of
514 eukaryotes. *Endocytobiology: endosymbiosis and cell biology: a synthesis of*
515 *recent research* 1:63-84.
- 516 Dayhoff M, Schwartz R, Orcutt B. 1978. A model of evolutionary change in proteins.
517 *Atlas of protein sequence and structure* 5:345-352.
- 518 Drummond AJ, Ho SY, Phillips MJ, Rambaut A. 2006. Relaxed phylogenetics and
519 dating with confidence. *PLoS Biol.* 4:e88.
- 520 Duchene DA, Tong KJ, Foster CSP, Duchene S, Lanfear R, Ho SYW. 2020. Linking
521 Branch Lengths across Sets of Loci Provides the Highest Statistical Support for
522 Phylogenetic Inference. *Mol. Biol. Evol.* 37:1202-1210.
- 523 Farris JS. 1972. Estimating Phylogenetic Trees from Distance Matrices. *Am. Nat.*
524 106:645-&.
- 525 Gatesy J, Springer MS. 2017. Phylogenomic red flags: Homology errors and zombie
526 lineages in the evolutionary diversification of placental mammals. *Proc Natl*
527 *Acad Sci U S A* 114:E9431-E9432.
- 528 Gogarten JP, Kibak H, Dittrich P, Taiz L, Bowman EJ, Bowman BJ, Manolson MF,
529 Poole RJ, Date T, Oshima T, et al. 1989. Evolution of the vacuolar H⁺-ATPase:
530 implications for the origin of eukaryotes. *Proc Natl Acad Sci U S A* 86:6661-
531 6665.
- 532 Hasegawa M, Kishino H, Yano T. 1985. Dating of the human-ape splitting by a
533 molecular clock of mitochondrial DNA. *J. Mol. Evol.* 22:160-174.

ROOTSTRAP SUPPORT

- 534 Heaps SE, Nye TM, Boys RJ, Williams TA, Embley TM. 2014. Bayesian modelling of
535 compositional heterogeneity in molecular phylogenetics. *Stat Appl Genet Mol*
536 *Biol* 13:589-609.
- 537 Holmes S. 2003. Bootstrapping phylogenetic trees: Theory and methods. *Statistical*
538 *Science* 18:241-255.
- 539 Huelsenbeck JP, Bollback JP, Levine AM. 2002. Inferring the Root of a Phylogenetic
540 Tree. *Syst. Biol.* 51:32-43.
- 541 Iwabe N, Kuma K, Hasegawa M, Osawa S, Miyata T. 1989. Evolutionary relationship of
542 archaeobacteria, eubacteria, and eukaryotes inferred from phylogenetic trees of
543 duplicated genes. *Proc Natl Acad Sci U S A* 86:9355-9359.
- 544 Jansen RK, Cai Z, Raubeson LA, Daniell H, Depamphilis CW, Leebens-Mack J, Muller
545 KF, Guisinger-Bellian M, Haberle RC, Hansen AK, et al. 2007. Analysis of 81
546 genes from 64 plastid genomes resolves relationships in angiosperms and
547 identifies genome-scale evolutionary patterns. *Proc Natl Acad Sci U S A*
548 104:19369-19374.
- 549 Jayaswal V, Ababneh F, Jermin LS, Robinson J. 2011. Reducing model complexity of
550 the general Markov model of evolution. *Mol. Biol. Evol.* 28:3045-3059.
- 551 Jones DT, Taylor WR, Thornton JM. 1992. The rapid generation of mutation data
552 matrices from protein sequences. *Comput. Appl. Biosci.* 8:275-282.
- 553 Jukes TH, Cantor C. 1969. Evolution of protein molecules. In: Munro HN, editor. In
554 *Mammalian Protein Metabolism*.
- 555 Kalyanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermin LS. 2017.
556 ModelFinder: fast model selection for accurate phylogenetic estimates. *Nat.*
557 *Methods* 14:587-589.
- 558 Kimura M. 1980. A Simple Method for Estimating Evolutionary Rates of Base
559 Substitutions through Comparative Studies of Nucleotide-Sequences. *J. Mol.*
560 *Evol.* 16:111-120.

NASER-KHDOUR ET AL.

- 561 Kocot KM, Cannon JT, Todt C, Citarella MR, Kohn AB, Meyer A, Santos SR, Schander
562 C, Moroz LL, Lieb B, et al. 2011. Phylogenomics reveals deep molluscan
563 relationships. *Nature* 477:452-456.
- 564 Lake JA, Herbold CW, Rivera MC, Servin JA, Skophammer RG. 2007. Rooting the tree
565 of life using nonubiquitous genes. *Mol. Biol. Evol.* 24:130-136.
- 566 Le SQ, Gascuel O. 2008. An improved general amino acid replacement matrix. *Mol.*
567 *Biol. Evol.* 25:1307-1320.
- 568 Liu L, Zhang J, Rheindt FE, Lei F, Qu Y, Wang Y, Zhang Y, Sullivan C, Nie W, Wang
569 J, et al. 2017. Genomic evidence reveals a radiation of placental mammals
570 uninterrupted by the KPg boundary. *Proc Natl Acad Sci U S A* 114:E7282-
571 E7290.
- 572 Lyons-Weiler J, Hoelzer GA, Tausch RJ. 1998. Optimal outgroup analysis. *Biol. J.*
573 *Linn. Soc.* 64:493-511.
- 574 Maddison WP, Donoghue MJ, Maddison DR. 1984. Outgroup Analysis and Parsimony.
575 *Systematic Zoology* 33:83-103.
- 576 Mai U, Sayyari E, Mirarab S. 2017. Minimum variance rooting of phylogenetic trees
577 and implications for species tree reconstruction. *PLoS One* 12:e0182238.
- 578 Meganathan PR, Pagan HJ, McCulloch ES, Stevens RD, Ray DA. 2012. Complete
579 mitochondrial genome sequences of three bats species and whole genome
580 mitochondrial analyses reveal patterns of codon bias and lend support to a basal
581 split in Chiroptera. *Gene* 492:121-129.
- 582 Meredith RW, Janecka JE, Gatesy J, Ryder OA, Fisher CA, Teeling EC, Goodbla A,
583 Eizirik E, Simao TL, Stadler T, et al. 2011. Impacts of the Cretaceous Terrestrial
584 Revolution and KPg extinction on mammal diversification. *Science* 334:521-524.
- 585 Milinkovitch MC, Lyons-Weiler J. 1998. Finding optimal ingroup topologies and
586 convexities when the choice of outgroups is not obvious. *Mol. Phylogen. Evol.*
587 9:348-357.

ROOTSTRAP SUPPORT

- 588 Minh BQ, Hahn MW, Lanfear R. 2020. New methods to calculate concordance factors
589 for phylogenomic datasets. *Mol. Biol. Evol.*
- 590 Minh BQ, Schmidt HA, Chernomor O, Schrempf D, Woodhams MD, von Haeseler A,
591 Lanfear R. 2020. IQ-TREE 2: New Models and Efficient Methods for
592 Phylogenetic Inference in the Genomic Era. *Mol. Biol. Evol.* 37:1530-1534.
- 593 Moore MJ, Bell CD, Soltis PS, Soltis DE. 2007. Using plastid genome-scale data to
594 resolve enigmatic relationships among basal angiosperms. *Proc Natl Acad Sci U*
595 *S A* 104:19363-19368.
- 596 Nardi F, Spinsanti G, Boore JL, Carapelli A, Dallai R, Frati F. 2003. Hexapod origins:
597 monophyletic or paraphyletic? *Science* 299:1887-1889.
- 598 Naser-Khdour S, Minh BQ, Zhang W, Stone EA, Lanfear R. 2019. The Prevalence and
599 Impact of Model Violations in Phylogenetic Analysis. *Genome Biol Evol.*
- 600 Nguyen LT, Schmidt HA, von Haeseler A, Minh BQ. 2015. IQ-TREE: a fast and
601 effective stochastic algorithm for estimating maximum-likelihood phylogenies.
602 *Mol. Biol. Evol.* 32:268-274.
- 603 Philippe H, Brinkmann H, Lavrov DV, Littlewood DT, Manuel M, Worheide G, Baurain
604 D. 2011. Resolving difficult phylogenetic questions: why more sequences are not
605 enough. *PLoS Biol.* 9:e1000602.
- 606 Ren M, Sun HJ, Bo SQ, Zhang SY, Hua PY. 2018. Parallel amino acid deletions of
607 prestin protein in two dramatically divergent bat lineages suggest the complexity
608 of the evolution of echolocation in bats. *Acta Chiropterologica* 20:311-317.
- 609 Reyes-Amaya N, Flores D. 2019. Hypophysis size evolution in Chiroptera. *Acta*
610 *Chiropterologica* 21:65-74.
- 611 Rivera MC, Lake JA. 1992. Evidence That Eukaryotes and Eocyte Prokaryotes Are
612 Immediate Relatives. *Science* 257:74-76.
- 613 Shen XX, Hittinger CT, Rokas A. 2017. Contentious relationships in phylogenomic
614 studies can be driven by a handful of genes. *Nat Ecol Evol* 1:126.

NASER-KHDOUR ET AL.

- 615 Shimodaira H. 2002. An approximately unbiased test of phylogenetic tree selection.
616 *Syst. Biol.* 51:492-508.
- 617 Shimodaira H, Hasegawa M. 1999. Multiple comparisons of log-likelihoods with
618 applications to phylogenetic inference. *Mol. Biol. Evol.* 16:1114-1116.
- 619 Smith AB. 1994. Rooting Molecular Trees - Problems and Strategies. *Biol. J. Linn. Soc.*
620 51:279-292.
- 621 Squartini F, Arndt PF. 2008. Quantifying the Stationarity and Time Reversibility of the
622 Nucleotide Substitution Process. *Molecular Biology and Evolution* 25:2525-
623 2535.
- 624 Steenkamp ET, Wright J, Baldauf SL. 2006. The protistan origins of animals and fungi.
625 *Mol. Biol. Evol.* 23:93-106.
- 626 Swofford D, Olsen G, Waddell P. 1996. Phylogenetic inference. In: David M. Hillis
627 CM, Barbara K. Mable, editor. *Molecular Systematics*, 2nd edn: Sunderland,
628 Mass. : Sinauer Associates. p. 407-513.
- 629 Tamura K, Nei M. 1993. Estimation of the number of nucleotide substitutions in the
630 control region of mitochondrial DNA in humans and chimpanzees. *Mol. Biol.*
631 *Evol.* 10:512-526.
- 632 Tavaré S. 1986. Some probabilistic and statistical problems in the analysis of DNA
633 sequences. *Lectures on Mathematics in the Life Sciences* 17.
- 634 Tria FDK, Landan G, Dagan T. 2017. Phylogenetic rooting using minimal ancestor
635 deviation. *Nat Ecol Evol* 1:193.
- 636 Tsagkogeorga G, Parker J, Stupka E, Cotton JA, Rossiter SJ. 2013. Phylogenomic
637 analyses elucidate the evolutionary relationships of bats. *Curr. Biol.* 23:2262-
638 2267.
- 639 Watrous LE, Wheeler QD. 1981. The out-Group Comparison Method of Character
640 Analysis. *Systematic Zoology* 30:1-11.

ROOTSTRAP SUPPORT

- 641 Whelan NV, Kocot KM, Moroz LL, Halanych KM. 2015. Error, signal, and the
642 placement of Ctenophora sister to all other animals. *Proc Natl Acad Sci U S A*
643 112:5773-5778.
- 644 Whelan S, Goldman N. 2001. A general empirical model of protein evolution derived
645 from multiple protein families using a maximum-likelihood approach. *Mol. Biol.*
646 *Evol.* 18:691-699.
- 647 Williams KP, Gillespie JJ, Sobral BW, Nordberg EK, Snyder EE, Shallom JM,
648 Dickerman AW. 2010. Phylogeny of gammaproteobacteria. *J. Bacteriol.*
649 192:2305-2314.
- 650 Williams TA, Heaps SE, Cherlin S, Nye TM, Boys RJ, Embley TM. 2015. New
651 substitution models for rooting phylogenetic trees. *Philos Trans R Soc Lond B*
652 *Biol Sci* 370:20140336.
- 653 Wu S, Edwards S, Liu L. 2019. Data from: Genome-scale DNA sequence data and the
654 evolutionary history of placental mammals. In: Figshare.
- 655 Wu S, Edwards S, Liu L. 2018. Genome-scale DNA sequence data and the evolutionary
656 history of placental mammals. *Data Brief* 18:1972-1975.
- 657 Yang ZH, Roberts D. 1995. On the Use of Nucleic-Acid Sequences to Infer Early
658 Branchings in the Tree of Life. *Mol. Biol. Evol.* 12:451-458.
- 659 Yap VB, Speed T. 2005. Rooting a phylogenetic tree with nonreversible substitution
660 models. *BMC Evol. Biol.* 5:2.
- 661 Yu Y, Warnow T, Nakhleh L. 2011. Algorithms for MDC-based multi-locus phylogeny
662 inference: beyond rooted binary gene trees on single alleles. *J. Comput. Biol.*
663 18:1543-1559.
- 664 Zhang SQ, Che LH, Li Y, Dan L, Pang H, Slipinski A, Zhang P. 2018. Evolutionary
665 history of Coleoptera revealed by extensive sampling of genes and species. *Nat*
666 *Commun* 9:205.
- 667 Zhou X, Xu S, Yang Y, Zhou K, Yang G. 2011. Phylogenomic analyses and improved
668 resolution of Cetartiodactyla. *Mol Phylogenet Evol* 61:255-264.