Seasonal climate variations promote bacterial α-diversity in soil

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Abstract

Ecological theory suggests that temporal environmental fluctuations can contribute

greatly to diversity maintenance. Given bacteria's short generation time and rapid

responses to the environmental change, seasonal climate fluctuations are very likely to

play an important role in maintaining the extremely high α-diversity of soil bacterial

community, which has been unfortunately neglected in previous studies. Here, with

in-depth analyses of two previously published high-quality soil bacterial datasets at

global scale, we found that soil bacterial α-diversity was positively correlated with

both seasonal variations of temperature and precipitation. Furthermore, piecewise

structural equation models showed that seasonal variations of temperature or

precipitation directly promoted soil bacterial α-diversity in each dataset. Our findings

implied that fluctuation-dependent mechanisms of diversity maintenance presumably

operate in soil bacterial communities and highlighted that both the average values and

temporal variations of abiotic environmental factors should be considered when

investigating soil bacterial community composition in the future.

The extraordinarily high α-diversity of soil bacterial communities has fascinated and

puzzled microbial ecologists [1, 2]. In contrast to the considerable advances in

characterizing soil bacterial biogeographic patterns across spatial and environmental

gradients (e.g. [3, 4]), little attempt has been made to infer the underlying mechanisms

of diversity maintenance. The high spatial heterogeneity of soil was generally thought

as an important factor in maintaining bacterial α-diversity [5]; surprisingly, temporal

environmental fluctuations promoting diversity maintenance firstly suggested by

Hutchinson was rarely mentioned, which is one of the most influential ideas in

community ecology [6, 7]. The fluctuation-dependent coexistence mechanisms are

particularly important for micro-organisms due to their short generation time and

rapid responses to environmental changes [8, 9]. Temporal changes in temperature

and precipitation can significantly alter soil physiochemical and nutritional status,

which jointly enhance temporal environmental fluctuations in soil, and drive the

seasonal species turnover in bacterial community [10]. However, in nearly all of the

bacterial biogeographic studies, only the mean annual temperature (MAT) and annual

precipitation (AP) were chosen as climatic predictors of soil bacterial community

diversity and composition, whereas seasonal climate variations were unfortunately

neglected. Here we tested the prediction that seasonal climate variations promote soil

bacterial α-diversity derived from the fluctuation-dependent coexistence theory, with

two previously published surveys covering wide geographic and climatic gradients:

global topsoil microbiome (hereafter "topsoil data") [4] and global soil atlas (hereafter

"atlas data") [11]. Details of data acquisition, processing, and statistical analyses were

provided in supplementary methods.

The climate data were obtained from the WorldClim database

(http://www.worldclim.org), whereby the standard deviation of monthly mean

temperature and the coefficient of variation of monthly precipitation were used to

represent temperature seasonality (TS) and precipitation seasonality (PS), respectively.

Soil bacterial α-diversity was measured as the number of observed phylotypes

(hereafter richness). As predicted, richness of soil bacteria significantly increased with

TS and PS in both datasets (Fig 1a-d; linear regression, $p < 0.05$). Piecewise structural

equation model was then fitted to infer how geographic (absolute attitude), climatic

(MAT, AP, TS, PS) variables and soil physiochemical properties (pH and C/N ratio,

the most important two soil physiochemical factors in determining bacterial diversity

and composition) interact to determine richness of soil bacteria. The modeling results

demonstrated significant direct effect of PS (Fig 2a; standardized coefficient $\beta =$

0.206, $p < 0.001$) in promoting richness of soil bacteria in topsoil data, and TS (Fig 2b;

standardized coefficient $\beta = 0.188$, $p < 0.001$) in atlas data. Note that the average of

TS was significantly larger in atlas data, whereas the average of PS was significantly

larger in topsoil data (Fig 1e, f; Mann-Whitney test, $p < 0.001$). It is highly possible

that TS and PS simultaneously promote soil bacterial α-diversity, but their effects are

more likely to be detected when samples with high levels of seasonal variations are

included.

Theoretically, there are two classes of fluctuation-dependent coexistence

mechanisms: the storage effect and relative nonlinearity of competition [7, 9]. The

storage effect mediates coexistence via temporal niche partitioning [7], and its

operation requires that (1) species differ in their responses to environments, which

results in (2) the relative strength of interspecific and intraspecific competition

varying with fluctuating environments; and that (3) there are mechanisms buffering

population from extinction under unfavorable conditions [7], such as a variety of

physiological stress-resistance and dormancy mechanisms of soil bacteria [12]. There

has been much experimental evidence that soil bacteria show diverse responses to

moisture and temperature gradients. For example, a collection of soil bacterial isolates

exhibited a wide range of responses to moisture gradient, and the derived niche

parameters suggested a potential of coexistence via partitioning the moisture niche

axis [13]. In addition to dry- and wet-adapted bacteria, stress-tolerant and

opportunistic strategies of soil bacteria in response to drought or raining events were

also discovered, as well as sensitive taxa [12, 14]. Likewise, both warm- and

cold-responsive taxa were identified [15], and soil bacterial communities can rapidly

diverge under contrasting experimental temperature treatments [16]. Besides evidence

strongly supporting differential responses to moisture and temperature (requirement 1)

discussed above, repeated reports of seasonal turnover in soil bacterial community

structure (e.g. [10]) implied altered competitive intensities among soil bacteria under

fluctuating environments (requirement 2); although distinguishing whether it is due to

the fluctuating temperature and moisture *per se*, and/or the accompanying seasonal

changes in availability of various resources was difficult. So far, rigorously testing of

the storage effect has been restricted to simplistic microcosms [17, 18]; although

being presumably important, its role in maintaining soil bacterial α-diversity in

natural system remains unexamined yet.

By contrast, relative nonlinearity of competition might not be the major

mechanism responsible for the promoting effect of seasonal climate variations on the

soil bacterial α-diversity. Coexistence via relative nonlinearity of competition is based

on that the per capita growth rates of species are different nonlinear functions of

limiting resource [7, 8, 19]. Such nonlinearity can result in the fluctuation of limiting

resource, and the stable coexistence can achieve when each species is disadvantaged

relative to the others in the fluctuation environment caused by itself, but not

necessarily requires fluctuation of external conditions [7, 8, 19]. On the other hand,

previous modeling investigations emphasizing the importance of relative nonlinearity

of competition merely focused on microorganisms in homogenous aquatic systems [8,

9], and their implications cannot be directly generalized to the soil environment with

high spatial heterogeneity, where the resources are not simultaneously accessible to

all species.

In conclusion, our findings supported the idea that seasonal temperature and

precipitation variations promote soil bacterial α-diversity via fluctuation-dependent

mechanisms of diversity maintenance. Our study benefited from applying classic

ecological theories to interpret pattern and get mechanistic insight [20], and

highlighted the importance of considering both the average values and temporal

variations of abiotic environmental factors to understand the soil bacterial community

composition. Further experimental studies designed to investigate the specific

coexistence mechanisms will shed new light on the diversity maintenance of soil

bacterial communities in the future.

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Figure legends

Fig 1 The relationships between soil bacterial richness (the number of observed phylotypes) and temperature (a, c) or precipitation seasonality (b, d), with the extent of climatic seasonality captured (e, f) in topsoil and atlas data. The colors represent different datasets (topsoil data: red, atlas data: blue). In panels (e, f), the lower and upper ends of the boxes represent 25% and 75% of the range, respectively; lines in the boxes indicate medians; triangles indicate maximums and minimums; squares indicate mean values; and whiskers represent $\pm 1.5 \times$ the interquartile range (IQR, defined as the upper quartile minus the lower quartile); different letters denote significant differences in climatic seasonality between two datasets (Mann-Whitney test, $p < 0.001$).

Fig 2 Structural equation models (piecewise SEM) of geographic (absolute latitude), climatic (mean annual temperature, annual precipitation, temperature seasonality, precipitation seasonality) variables, and soil physiochemical properties (pH, C/N ratio) as predictors of soil bacterial richness for topsoil data (a) and atlas data (b). Red arrows represent positive paths, and black arrows represent negative paths; only significant paths were shown ($p < 0.05$). Standardized effect sizes of path coefficients were reported and indicated by path thickness. Shipley's test of directed separation: Fisher's C statistic (if $p > 0.05$, then there are no missing associations and the model reproduces the data well) and AIC were used to evaluate the overall fit of the model. Details of model evaluation and coefficients estimation were provided in supplementary methods and results.

(a) Topsoil data

Fisher's C = 2.98, p = 0.936

AIC = 78.98

Annual precipitation
R^2 = 0.528

Mean annual temperature
R^2 = 0.757

Temperature seasonality
R^2 = 0.63

Distance to equator (absolute latitude)

Soil pH
R^2 = 0.424

C / N ratio
R^2 = 0.11

Precipitation seasonality
R^2 = 0.301

Soil bacterial α-diversity (richness)
R^2 = 0.546

0.348
-0.581
-0.367
-0.388
0.59
0.464
-0.87
-0.321
0.499
0.569
-0.392
-0.336
0.698
-0.351
0.206
0.371

(b) Atlas data

Fisher's C = 9.74, p = 0.94

AIC = 75.74

Annual precipitation
R^2 = 0.262

Mean annual temperature
R^2 = 0.722

Temperature seasonality
R^2 = 0.572

Distance to equator (absolute latitude)

Soil pH
R^2 = 0.504

C / N ratio
R^2 = 0.114

Precipitation seasonality
R^2 = 0.279

Soil bacterial α-diversity (richness)
R^2 = 0.443

-0.71
-0.646
-0.589
-0.339
-0.25
0.476
-0.85
-0.583
0.195
0.17
-0.21
0.572
-0.149
-0.819
0.216
-0.337
0.188
-0.987