

Title: Historical and contemporary signatures of selection in response to transmissible cancer in the Tasmanian Devil (*Sarcophilus harrisii*)

Authors: Amanda R. Stahlke<sup>1</sup>, Brendan Epstein<sup>2</sup>, Soraia Barbosa<sup>1</sup>, Austin Patton<sup>2</sup>, Sarah A. Hendricks<sup>1</sup>, Anne Veillet<sup>1</sup>, Alexandra K Fraik<sup>2</sup>, Barbara Schönfeld<sup>3</sup>, Hamish I. McCallum<sup>4</sup>, Rodrigo Hamede<sup>3</sup>, Menna E. Jones<sup>3</sup>, Andrew Storfer<sup>2</sup>, Paul A. Hohenlohe<sup>1</sup>

Affiliations:

1. Institute for Bioinformatics and Evolutionary Studies (IBEST), University of Idaho, Moscow, ID 83844, USA
2. School of Biological Sciences, Washington State University, Pullman, WA 99164, USA
3. School of Natural Sciences, University of Tasmania, Hobart, TAS 7005, Australia
4. Environmental Futures Research Institute, Griffith University, Nathan, QLD 4111, Australia

Abstract

Tasmanian devils (*Sarcophilus harrisii*) are evolving in response to a unique transmissible cancer, devil facial tumour disease (DFTD), first described in 1996. Persistence of wild populations and the recent emergence of a second independently evolved transmissible cancer suggest that transmissible cancers may be a recurrent feature in devils. We used a targeted sequencing approach, RAD-capture, to identify genomic regions subject to rapid evolution in approximately 2,500 devils as DFTD spread across the species range. We found evidence for genome-wide contemporary evolution, including 186 candidate genes related to cell cycling and immune response. We then searched for signatures of recurrent selection with a molecular evolution approach and found widespread evidence of historical positive selection in devils relative to other marsupials. We identified both contemporary and historical selection in 19 genes and enrichment for contemporary and historical selection independently in 22 gene sets. Nonetheless, the overlap between candidates for historical selection and for contemporary response to DFTD was lower than expected, supporting novelty in the evolutionary response of devils to DFTD. Our results can inform management actions to conserve adaptive capacity of devils by identifying high priority targets for genetic monitoring and maintenance of functional diversity in managed populations.

Keywords: Rapid evolution, molecular evolution, gene set overlap, wildlife disease, conservation genomics, RAD-capture

## Introduction

Species are subject to selection by pathogens throughout their evolutionary history, shaping lineage diversification and leading to complex cellular and molecular defensive mechanisms (1). Still, emerging infectious diseases (EIDs) can cause mass mortality and, given sufficient survival and genetic variation, initiate rapid adaptive evolution in a naïve host population (2). Although the prevalence and severity of EIDs in wildlife populations is now well-recognized (3-6), we are just beginning to understand the evolutionary impacts of disease in wildlife. We have a relatively short recorded history of infectious disease in wildlife, and therefore a limited ability to predict outcomes or intervene when warranted (7, 8).

High-throughput DNA-sequencing techniques and the generation of high-quality annotated reference genomes have revolutionized our ability to monitor and identify mechanisms of evolutionary responses to pathogens (8-10). Inter-specific comparisons of non-synonymous and synonymous variation (dN/dS) within protein-coding regions have long been used to identify positive selection at immune-related loci (11, 12). In the genomic era, these analyses have been extended beyond a few well-characterized genes and a few well-studied species; for example, to reveal shared positive selection hotspots among viral immune pathways in both birds and mammals (13). At the population level, rapid evolution in response to disease can be detected by tracking changes in allele frequency before, during, and after the outbreak of disease (14, 15). However, such analyses are resource-intensive and therefore still challenging in wildlife systems (but see 16, 17, 18). Reduced representation techniques such as restriction-site associated DNA-sequencing (RADseq) (19) have made the acquisition of genome-wide, time-series genetic data more accessible in non-model systems (20). By integrating these resources and tests of selection at differing temporal scales, we can assess whether species that show rapid evolution in response to contemporary pathogens also show evidence of historical selection to similar pathogens.

A striking example of an EID acting as an extreme selective force in wildlife is devil facial tumour disease (DFTD), a transmissible cancer first described in 1996 afflicting wild Tasmanian devils (*Sarcophilus harrisii*) (21). Tasmanian devils are the largest extant carnivorous marsupial, with contemporary wild populations restricted to the Australian island of Tasmania. As a transmissible cancer, DFTD tumour cells are transmitted between hosts, behaving as a pathogen (22). Transmission typically occurs as devils bite each other during the mating season after devils have reached sexual maturity (23, 24). With few notable exceptions documenting regression (25), DFTD tumors escape recognition, become malignant, and can kill their hosts within six months (26). Starting from a single Schwann cell origin (27), DFTD has now swept across nearly the entire species range (Figure 1A). Devil populations have declined species-wide ~80% (28) with local declines in excess of 90%. Most populations will reach 90% decline after about 6 years, with just a few exceptions (29). This is particularly alarming because devils have notoriously low genome-wide diversity, attributed to climate- and anthropogenic-induced bottlenecks (30-32). Depleted genetic diversity at immune-related loci has likely further contributed to DFTD vulnerability (33).

Remarkably, a second independent transmissible cancer, DFT2, was described 20 years later in 2014 (34, 35). Comparative and functional analyses of DFTD and DFT2 showed similar drivers of cancerous mutations and tissue type of origin (36). Low genetic diversity, chromosomal fragility (37), a reportedly high incidence of non-transmissible neoplasms (38), and injury-prone biting behaviour (39) may contribute to a predisposition to transmissible cancers in devils (40). Devils have shown signs of resilience to DFTD. Females at five sites were found to exhibit precocial breeding after the arrival of DFTD, potentially increasing their lifetime productivity despite high mortality due to DFTD (41). DFTD has been spontaneously cleared (i.e., regressed) in some individuals (25). Long-term, field-based pattern matching and individual-based simulations predicted that cyclical co-existence or DFTD extirpation are more likely scenarios than devil extinction (42). Incorporating evolutionary responses by devils into such predictive models could also increase the likelihood of devil persistence. Taken together, these findings suggest that transmissible cancers may be a recurring selective force in the Tasmanian devil lineage.

Because of the threat of DFTD and DFT2 to devil populations, there are ongoing conservation efforts, including the establishment of a captive devil insurance meta-population. The insurance population is managed to maintain genome-wide genetic diversity and serve as a source for re-introductions in an effort to increase genetic diversity and size of wild populations (43). To inform conservation efforts, it is important to understand what types of genetic variation in natural populations may allow for evolutionary rescue from disease and maintain adaptive potential for future threats (44). Given evidence for rapid evolution in response to DFTD, monitoring of genetic variation at candidate adaptive loci could help evaluate the adaptive potential of wild populations (44, 45). In heavily managed (e.g. captive) populations, loci associated with an adaptive response to disease could be included in genotyping panels for maintaining genetic diversity (46).

Ongoing long-term field and genomic studies have provided a wealth of resources to examine responses to DFTD. Since the first disease detection in 1996, field sites have been established across the island of Tasmania to consistently monitor the spread and effects of the disease and wild devil populations (41). A high-quality reference genome for the devil yields deep evolutionary history insight and fine-scale mapping for functional inference of individual genotypes (27, 32). Population genomic studies have already shown that devils are rapidly evolving in response to DFTD (2, 47, 48). From these data, a RAD-capture (Rapture; 49) panel has been developed to target 16,000 putative candidates and demographically-informative loci, facilitating the collection of data across hundreds of individuals in a single lane of sequencing. This panel was used to identify genes associated with DFTD-related phenotypes, including infection and survival in females (50). We also now have access to the shared and divergent evolutionary histories owing to a robust, dated phylogenetic reconstruction (51), and high quality, readily accessible, annotated reference genomes of the South American grey-tailed opossum (*Monodelphis domestica*) (52) the tammar wallaby (*Notamacropus eugenii*) (53), and the koala (*Phascolarctos cinereus*) (54).

Here we identify targets of selection and signatures of adaptation at both contemporary and historical time scales in Tasmanian devils. We expand upon previous studies of contemporary selection with a much larger sample size including several time points for sites across the entire island, using a RAD-capture panel for targeted sequencing of almost 16,000 loci. Then, we test for evidence of recurrent selection by examining shared contemporary and historical signatures of selection, in terms of either specific genes or functional genetic pathways that could relate to transmissible cancer. We propose three hypotheses: 1) If transmissible cancer is a novel selective force acting on Tasmanian devils, genes under contemporary selection by DFTD will be different from genes with a signature of historical positive selection. 2) If transmissible cancer is a recurrent selective force in the devil lineage that targets the same set of genes repeatedly, we may expect an overrepresentation of the same genes under both contemporary and historical selection. Alternatively, 3) recurrent transmissible cancers may impose selection on different genes within a common set of pathways, or even on genes in different functional groups. These alternatives can inform conservation efforts to manage genetic diversity for resilience in natural devil populations, and any genes or functional pathways that show both contemporary and historical selection may be relevant to cancer resistance more broadly.

## Materials and Methods

### *Rapture Sequencing for Contemporary Selection*

We used the Rapture method (combining RAD and sequence capture; 49) to conduct targeted genotyping of single-nucleotide polymorphisms (SNPs) across 2,562 individuals from multiple Tasmanian devil populations, sampled both before and after DFTD appeared in each population (Figure 1A, Supplemental Table S1). For SNP genotyping, we constructed Rapture libraries following Ali et al (2016), using the restriction enzyme *Pst*I and a capture array targeting 15,898 RAD loci selected for membership in one of three functional categories: 1) those showing signatures of DFTD-related selection from previous work (Epstein et al. 2016), 2) loci close to genes with known cancer or immune function, and 3) loci widely distributed across the genome (See 48, 50 for more details on the development of this array.). See Supplemental Materials S1 for multiplexing, read processing, and genotyping details.

### *Spatial and temporal analysis of contemporary selection*

We identified candidate SNPs by combining the results of three analyses: raw change in allele frequency in each population after DFTD (hereafter  $\Delta af$ ), and two methods that estimate strength of selection from allele frequencies at multiple time points in multiple populations, the method of Mathieson & McVean (14), (hereafter *mm*), which allows the estimated selection coefficient to vary over space; and *spatpg* (15), which allows the selection coefficient to vary over time as well as space. To reduce the signal-to-noise ratio, we then combined three tests for contemporary selection with a de-correlated composite of multiple signals (*DCMS*) (55). See Supplemental Materials S1 for details of each analysis.

### *Historical Selection*

Today, devils are the only species of marsupials with any known history of transmissible cancer. We combined existing genomic resources for the South American grey-tailed opossum and tammar wallaby from the Ensembl database (56) and the recently published transcriptome assembly of the koala (54) to identify signatures of positive selection in devils, relative to the three other annotated marsupial genomes using the branch-site test of PAML (Phylogenetic Analysis by Maximum Likelihood (57, 58). The branch-site test compares the likelihood scores for two models which estimate dN/dS among site classes of the multi-sequence alignment and branches. In the neutral model, all site classes and branches are constrained to  $dN/dS \leq 1$ . In the alternative model, dN/dS of site class 2 is allowed to exceed 1 for only the foreground branch, while constraining the background branches to  $dN/dS \leq 1$ . See Supplemental Materials S1 for details regarding orthology identification and PAML implementation.

### *Comparison of Contemporary and Historical Signatures*

To test whether the number of genes under contemporary selection differed from the genome-wide background, we compared the number of overlapping candidate and non-candidate genes with Fisher's one-tailed test. To test for differences in the strength of selection, we compared the distributions of dN/dS magnitude  $> 1$  and the proportion of sites per gene with  $dN/dS > 1$  found in Site Class 2 of the branch-site test. We used two nonparametric tests of equality to compare the distributions; our null hypothesis was that the two samples came from the same population (59) (60). To identify key mechanisms of adaptation among candidate genes, we used gene ontology (GO) term enrichment analysis using the SNP2GO package (61) and the PANTHER web-interface (62). We capitalized on the wealth of ongoing research in devils and DFTD by comparing our contemporary and historical candidates to those previously identified using different datasets and analytical approaches (2, 47, 48, 50, 63). We then tested for overrepresentation of contemporary and historical candidates in gene sets of the molecular signatures database (MsigDB) (64). See Supplemental Materials S1 for details of these comparisons.

## **Results**

### *Genomic Data Processing*

To test for contemporary selection, we sampled a total of 2,562 individuals across six localities of Tasmania before and after DFTD prevalence (Table 1; Supplemental Table S1; Figure 1A), with a RAD-capture array (50). After filtering, we retained 14,585 – 22,878 SNPs variants for downstream analysis, depending on the sampled time point and population.

### *Evidence for contemporary selection*

In the top 1% of DCMS scores, we identified 144 candidate SNPs for contemporary selection by DFTD; of these, 79 had annotated genes (186 total) within 100 kb (Figure 2; Supplemental Table S4). In our comparison of previously identified candidate genes in devils (47, 48, 50, 63), we found many overlapping candidates (See *Discussion*), but only significant enrichment among our contemporary candidates for those associated with disease-related phenotypes (50), with the largest odds ratio for genes associated with phenotypes in females (14 genes,  $p$ -value=5.1e-08, Odds ratio=7.1). Gene ontology enrichment analysis found middle ear morphogenesis

(GO:0042474) significantly enriched among contemporary candidate SNPs ( $FDR < 0.05$ ). Five candidate SNPs were within the 100 kb window of two genes associated with this term: EYA1 and PRKRA. Both EYA1 and PRKRA are involved in cell proliferation and migration and implicated in tumour suppression and angiogenesis (65-67).

### *Evidence for historical selection*

Using the branch-site test for positive selection in PAML, we found 1,982 genes with a signal of historical positive selection (Figure 3, red); Supplementary Table S5). The majority of genes were classified as having a molecular function of binding or catalytic activity; involved in metabolic and cellular processes, or biological regulation; and a plurality as participating in the Wnt signalling pathway, which is keystone regulator of stem cell proliferation, including in cancer (68).

### *Comparing historical and contemporary selection*

Twenty-one contemporary candidate genes were also candidates for historical selection (Supplemental Table S6). The distributions of dN/dS estimates (Figure 3a) and proportion of sites (Figure 3b) in genes under historical positive selection from the genome-wide background and contemporary candidates similarly had an overall low magnitude of dN/dS and a low proportion of sites under historical positive selection. The contemporary candidates did not show evidence of more widespread historical selection compared to the genome-wide background according to tests of equality (59, 60).

We identified 22 gene sets from the Molecular Signatures Database (64) independently enriched within the contemporary candidate set and the historical candidate sets (Supplemental Table S8). Note that no single gene was under both historical and contemporary selection within these sets. The cancer gene neighbourhoods of correlated expression (CGN of MSigDb) from the MORF compendium (64), shared among the highest rates of overlap (approx. 30-40%) for both contemporary and historical candidate sets (Supplemental Figure S4). After correcting for multiple testing in each analysis, only the gene set for the gene expression neighbourhood of RBBP8 was still significantly enriched. Importantly, the permutation test of shared gene sets found *fewer* shared overlaps between historical and contemporary selection than expected by chance ( $p = 0.012$ , Supplemental Figure S4).

## **Discussion**

Here we used targeted sequencing and a molecular evolutionary analytical framework to show widespread positive selection in the Tasmanian devil across the genome at both historical and contemporary timescales. We identified 186 candidate genes for rapid allelic shifts following the arrival of DFTD at the contemporary scale in wild devil populations and 1,982 candidate genes for historical positive selection within devils relative to other marsupials. Statistical tests comparing degree of overlap and distributions among these candidates did not support recurrent selection on a common set of genes in response to transmissible cancer. Our results suggest that the specific genes implicated in the contemporary evolutionary response to DFTD are mostly novel targets compared to the genome-wide signature of historical selection. That is,

most of the genes under selection in devils in response to DFTD have not been under recurrent selection over macroevolutionary timescales. However, we also found support for overlapping contemporary and historical selection on some gene networks. Our work contributes to mounting evidence identifying possible mechanisms by which devil populations are persisting and rapidly evolving in the face of DFTD despite overall low genetic diversity and population bottlenecks (2, 25, 50, 69, 70). Broadly, this type of approach can be applied to selection analyses of novel threats in wildlife populations in the current era of anthropogenic global change and guide monitoring and management actions focused on genetic adaptive potential.

### *Contemporary Responses to DFTD*

Across six study sites that span the geographic range of Tasmanian devils, and genomic samples of over 2500 devils collected over 15 years, we found widespread signatures of rapid evolution in populations impacted by DFTD. Despite concerns that devils may not harbour sufficient genetic diversity to sufficiently respond to DFTD, we identified 186 candidate genes (Supplemental Table S4) with robust signatures of positive selection, reaffirming the adaptive potential of wild devils described in a growing body of work (2, 47, 48, 70). Our targeted RAD-capture sampling approach allowed us to sequence many more individual devils than previous studies and examine specific loci and regions of interest. These samples spanned six genetic populations shown to be locally adapted to differing habitats (48, 71). By incorporating thorough time-series sampling at each site throughout the course of the disease and bioinformatic methods that explicitly allow selection to vary over time and populations (*spatpg* and *mm*), our results provide robust evidence of rapid evolution across the island in response to DFTD.

In line with previous population genomic studies (2, 47, 48, 50, 63), our analysis of contemporary evolution detected a putatively adaptive response related to the immune system, cell adhesion, and cell-cycle regulation (Supplemental Table S4). Our GO enrichment result for middle ear development (GO:0042474) among contemporary candidates may highlight selection for interactions with the peripheral nervous system and cell proliferation. Whereas Hubert and colleagues (47) suggested genes with nervous system annotations may be indicative of selection for behavioural changes, others have pointed out the importance and vulnerability of peripheral nerve repair by Schwann cells in devils, given the prevalence of biting and Schwann cell origin of DFT (40). Regardless, genes and their products at the interface of the nervous system and proliferation could prove to be important candidates. Significant enrichment for genes associated with devil case-control, age, and survival (50) among our contemporary candidates is a strong indicator that these contemporary candidates likely confer relevant phenotypic change. We also confirmed five of the seven candidates identified previously by Epstein and colleagues (2) with RADseq (approximately 90,000 SNPs) in just three populations, specifically CRBN, ENSSHAG00000007088 (uncharacterized), and the previously identified chromosome 3 region including THY1, USP2, C1QTNF5. In contrast, we identified only two (TRNT1 and FSHB) of the 148 candidates from a re-analysis of that same dataset which identified population-specific responses (47). Candidates associated with disease prevalence identified with a genotype-environment-association approach (48) were also functionally

associated with transcription and cellular regulation, but relatively few of the same specific genes were identified (GRIP2, KYNU, ADAMST9, NDUFS3, PLEKHA3).

Among our contemporary candidates in the top 1% of DCMS scores across all tests, we did not identify any of the genes or regions that had been associated with host variation responsible for tumour regression (63). However, we found PAX3, which encodes a transcription factor associated with angiogenesis (72), as a top candidate for DCMS of allele frequency change ( $\Delta af$ ), possibly reflecting differences in test sensitivities. Among tumour genomic variants associated with regression (73), only the gene encoding Janus kinase and microtubule interacting protein 3 (JAKIMP3) was also found among our contemporary candidates. In general, the paucity of candidates shared between our contemporary analysis and regression studies suggests that regression is not the dominant form of phenotypic response to DFTD.

Our results affirm repeated, concordant responses across the island as described previously (2), with greater similarity between analytical approaches than within populations. This result is consistent with rapid evolution facilitated by selection for standing variation that was largely shared across populations prior to disease arrival. This timescale would likely be too short for new mutations to play a substantial role in DFTD response, and genetic variation is shared across the species range, despite some geographic population structure (2, 71).

#### *Historical selection in devils*

With our genome-wide molecular evolution approach (57), we found similarly widespread historical positive selection across the devil genome, in 1,977 genes, or about 31% of all 6,193 orthologs tested (Supplemental Table S5). The branch-site test is known to be less conservative than related models, particularly when divergence among species is large (74), but the rates of genome-wide selection we found in devils are similar to those described in other taxa; e.g. 17-73% of avian orthologs (13). To our knowledge, this is the first genome-wide survey of positive selection specifically among the publicly available annotated marsupials, spanning divergence with origins in the Cretaceous (approx. 80 my) (51).

We did not find preferential positive selection for immunity-related genes, as has been shown in primates (1), eutherian mammals more generally (75), and birds (13). Instead, we found the highest proportion of pathways under historical selection to be functionally classified within the Wnt pathway, a key signalling cascade regulating cell adhesion and implicated in carcinogenesis (76). As genomic resources grow and improve in marsupials (10), interspecific analyses for positive selection at finer scales may reveal more recent and specific responses in Tasmanian devils. Analytical resolution could also be improved by finer-scale annotation and curated databases of immune-system processes in marsupials to complement genome assembly efforts. Interspecific analyses of more recently diverged species could also have greater power to inform immune-system and cancer vulnerabilities in humans (77).

#### *Comparing Contemporary and Historical Timescales*

Remarkably few transmissible cancers have been discovered in nature (three in mammals, and a small handful in bivalve molluscs) (78, 79), and yet two of those independent clonally-transmitted cancers have been discovered in Tasmanian devils in less than 20 years. This, and the observed rapid evolutionary response to disease would suggest that transmissible cancers may be a recurrent event in devils. We tested for recurrent adaptation to transmissible cancers in devils by examining whether the contemporary signature of selection in response to DFTD (Figure 2) was reflected by positive selection at a deeper evolutionary time scale. We compared the distributions of historical selection in contemporary candidates to those of the genome-wide background (Figure 3) and by evaluating contemporary and historical gene set overlaps (Supplemental Tables S7 and S8). Our results did not conclusively support or reject the hypothesis of recurrent selection by repeated transmissible cancers. We found no significant overlap between contemporary candidates with signatures of historical selection and there was no significant difference in the proportion of sites or magnitude of historical selection between contemporary and the genome-wide background selection (Figure 3). We broadened the gene-by-gene comparison using the MSigDb gene sets to infer functional sets related to disease responses and found that shared gene set enrichment occurred less than expected by chance (Supplemental Figure S5). This suggests that these 22 gene sets could be specific targets warranting further investigation, instead of a proportional representation of the broad genome-wide background of historical selection. The 19 candidate genes (Supplemental Table S6) and 22 shared gene sets (Supplemental Table S8, Supplemental Figure S4) raise interesting targets for understanding functionally important variation in devils. Our results suggest a largely novel response to DFTD, but still highlight potential mechanisms of species persistence.

The nineteen candidates for recurrent selection (Supplemental Table S6) are generally related to three main themes: transcription regulation, the nervous system, and the centrosome. Four of these candidates for recurrent selection were previously associated with disease-related phenotypes by Margres and colleagues (50): SRP72, ARL9, ENSSHAG00000004590 (Novel, Set1DB), ENSSHAG00000011607 (Novel, MAPK). We additionally found 82 historical candidates previously identified in the top 1% of SNPs associated with disease-related phenotypes and three genes in the top 0.1% associated with large-effect sizes for female case-control and survival (50): PRKA10, ST8SIA2, and SLC12A8. PRKA10 binds to regulatory subunits of proteinase K (80). These results lend support to the hypothesis of recurrent selection by transmissible cancers, but the lack of significant overlap suggest that this could occur as a random sampling from two large lists of candidates. These three genes were not identified in our tests for contemporary selection. Both our contemporary selection analysis and the genome-wide association study (GWAS) approach used by Margres and colleagues (50) are statistically limited by small populations, sample size, and the time scale over which DFTD-related selection has occurred. By considering the complement of these results together, the overlapping historical, GWAS, and contemporary candidates may still be promising targets of recurrent selection along similar functional axes.

Gene overlap analysis using the immunologic, curated, and cancer gene network sets of the Molecular Signatures Database highlighted gene expression in cancer-related genes as a shared

functional feature of both historical and contemporary selection in devils. We focus here on the 22 significant overlaps found with both contemporary and historical candidates (Supplemental Table S8). The computationally mined co-expression neighbourhoods of cancer-related genes from the MORF compendium (64) had the highest rates of overlap for both contemporary and historical sets (Supplemental Figure S4). Among all gene sets, the strongest evidence for recurrent selection in functional pathways was for the CGN of the retinoblastoma binding protein 8 (MORF\_RBBP8), which was the only significant set after correcting for multiple testing. RBBP8 regulates cell proliferation and has been implicated as a tumour suppressor as a modulator of BRCA1 (81). High rates of overlap among the MORF sets could be because co-expressed genes are often subject to similar selective force at similar rates (75); or because co-expression and signatures of selection are more likely to occur within close physical proximity (82), although these genes are all on different super-contigs of the devil genome. Although variation in host gene expression has been shown to confer rapid evolution of disease resistance (83), differential expression among devils with differing disease status has not yet been investigated.

#### *Transmissible Cancer as a Recurrent Selective Force?*

The low prevalence of genes with both contemporary and historical signatures of selection suggest a novel response to DFTD compared to historic selection in the devil lineage. However, our results do not conclusively eliminate or support the possibility that devils have previously been impacted by transmissible cancers. The low genetic diversity observed in devils could be the result of the widespread historical positive selection resulting from transmissible cancers or other diseases (84), or historical bottlenecks due to climate change and habitat loss (30-32). The genes (Supplemental Table S6) and functional gene sets (Supplemental Table 8; Supplemental Figure S4) under selection at both timescales may reflect historical transmissible cancers, or indicate ongoing selection by a similar selective force. A hypothesis for the lack of greater overlap than we observed between historical and contemporary candidates is that there is redundancy in the genetic mechanisms underlying resistance to transmissible cancers, potentially as a result of repeated selection for resistance (85). It could be that historical purifying selection already eliminated variation at genes associated with cancer resistance.

The widespread contemporary evolution we found in devils reflects the recent prediction (86) that response to an emergent disease is most likely controlled by many genes conferring quantitative resistance (87), for example by reducing the within-host growth rate of tumors. By extension, DFTD is predicted to become less virulent in the short-term. If DFTD persists long-term in the devil population with ongoing coevolution, it may lead to diversifying selection for specific, qualitative gene-for-gene host resistance mechanisms (86). Phylodynamic study of DFTD could help further test these hypotheses by testing for relationships between genomic variation and transmission rates (88). Indeed, phylodynamic analysis of DFTD as it spread across the island supports the hypothesis that devils may be mounting a response; transmission rates have decayed such that DFTD appears to be shifting from emergence to endemism (Patton et al. 2020, In Review). Although host-genomic variation was not jointly considered in this analysis,

the combined evidence of multiple studies demonstrating rapid evolutionary response of devils to DFTD, including this one, support these interpretations.

### *Conservation Implications*

Calls have been made to consider the historical context of adaptation when proposing conservation management solutions based on genomic results (89). Our analysis of historical selection largely supports the hypothesis that DFTD is a newly emerging and novel selective force, distinctly shaping today's remaining wild devils. The targets of novel selection that we identified (Figure 2, Supp Table S4) and their functional roles should be considered for prioritization of monitoring and conservation in light of DFTD. At the same time, the wide distribution of contemporary candidates across the genome also highlights the importance of standing genetic variation to continue to respond to unique selective forces, including their local environments (48). The low genetic diversity observed in devils may be the result of historical population declines (30-32) and/or similarly diffuse selection operating historically in the devil lineage. Genomic monitoring could be useful for maintaining both functional diversity at candidate loci and genome-wide variation in breeding captive populations (46, 90, 91) and in the wild. Multiple genomic tools are available for targeted monitoring of large sets of loci (e.g. 92, 93) and could be used to track adaptive evolution and potential in the form of genetic diversity (44). However, before management decisions are made for specific genes further work should first identify favoured alleles and functional fitness effects for the genes we found in linkage with our genotyped SNP markers (Supplemental Table S4) .

At the time of writing, DFTD still has yet to reach all of the populations of devils in the far west (Fig. 1a), and although wild populations have persisted, DFTD continues to circulate throughout the island. We still do not know whether and how devils may be particularly susceptible to transmissible cancers, and our work cannot shed light on possible mechanisms for origin of these diseases. Instead our results inform mechanisms of tolerance/resistance to DFTD after it has emerged. To maintain adaptive capacity over the long term in the face of similar recurrent selective forces (e.g. transmissible cancers), our results warrant (1) the monitoring of genetic variation in broad functional groups, for example in genes related to transcription and cell proliferation; and (2) management strategies to maintain genetic diversity across those broad groups. This study could provide a list of candidate loci for development of a genotyping panel for either purpose, with flexibility to target many or fewer loci. At the same time, given urgent and unpredictable present-day threats including not just emerging diseases but environmental change and population fragmentation, it is important also for monitoring and population management to account for genetic variation across the genome.

### *Data and Script Accessibility*

Demultiplexed sequence data has been deposited at NCBI under Bio-Project PRJNA306495 (<http://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA306495>) and BioProject PRJNA634071 (<http://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA634071>). Code and tabular results are available at [https://github.com/Astahlke/contemporary\\_historical\\_sel\\_devils](https://github.com/Astahlke/contemporary_historical_sel_devils).

### *Acknowledgements*

The authors would like to acknowledge Michael R. Miller, Sean M. O'Rourke, Cody G. Wiench, for their contributions to the Rapture data; and Rebecca Johnson and Denis O'Meally for providing the koala transcriptome data. This work was funded by NSF grant DEB-1316549 and NIH grant R01-GM126563 to A.S., P.A.H., M.E.J., and H.I.M. as part of the joint NSF-NIH-USDA Ecology and Evolution of Infectious Diseases program. Genomics and bioinformatics were supported by an Institutional Development Award (IDeA) from the National Institute of General Medical Sciences of the NIH under grant number P30 GM103324. Sample collection was funded under Australian Research Council grants DE170101116, DP110102656, LP0989613 and LP0561120 to R.H., M.E.J., H.I.M., and A.S., an ARC Future Fellowship FT100100031 to M.E.J., and multiple awards from the University of Tasmania Foundation Eric Guiler Research Grant.

#### *Authors' contributions*

ARS, HIM, MEJ, AS, and PAH conceived and designed the study. RH and MEJ conducted fieldwork and sampling. ARS, BE, SB, SAH, AV, BS conducted genomics labwork. ARS, BE, SB, AP, SAH, AKF, and PAH conducted bioinformatic analyses. ARS wrote the manuscript with contributions from all authors.

*Tables and Figures*

Table 1. Number of adults sampled before and after the year of first detection of DFTD at each site. See Supplemental Table S1 for sample size for each year at each locality.

<b>Location</b>	<b>Year of First Detection</b>	<b>Samples Before</b>	<b>Samples After</b>	<b>Total</b>
Mt. William	1996	0	155	155
Freycinet	2001	300	382	682
Forestier	2004	131	552	683
Fentonbury	2005	99	169	268
West Pencil Pine	2006	52	348	400
Narawntapu	2007	224	150	374
<b>Total</b>		806	1756	2562

Figure 1. A) Map of the six contemporary sampling locations relative to disease prevalence over time (red lines) with the year of first detection labeled at each site. B) Pruned, unrooted phylogenetic tree of the marsupials (51) used to estimate genome-wide historical selection on the devil lineage. Devil cartoon by David Hamilton. From top to bottom: The tammar wallaby (*Notamacropus eugenii*), koala (*Phascolarctos cinereus*), the Tasmanian devil (*Sarcophilus harrisii*), and the South American grey-tailed opossum (*Monodelphis domestica*).

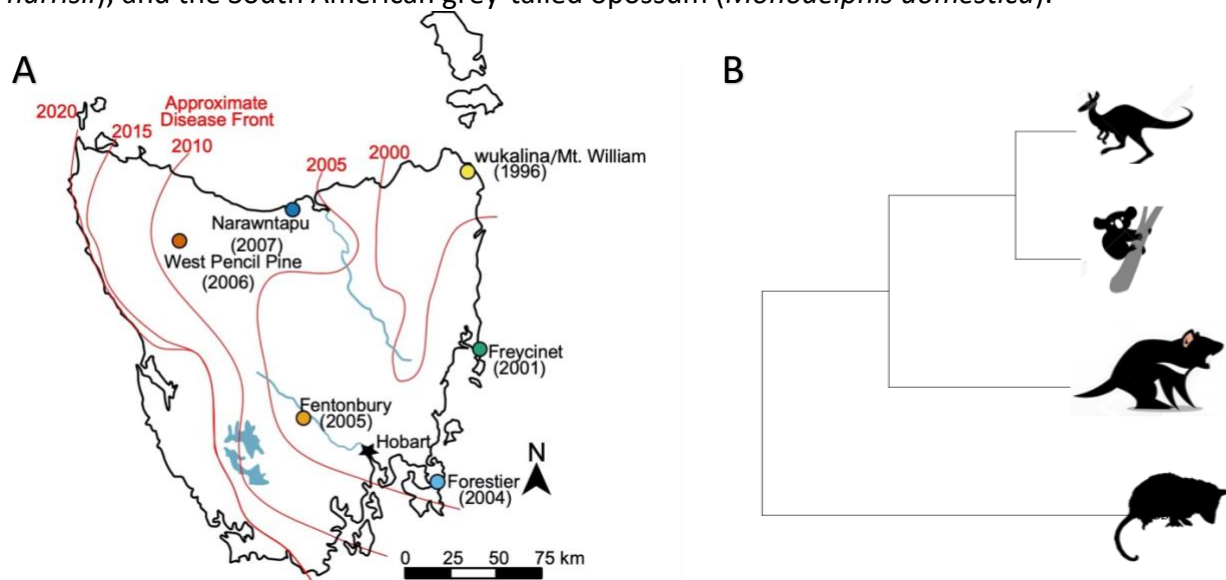


Figure 2. Results of each elementary test of contemporary selection across sites and composite score for candidate (in filled points) and noncandidate (in open squares) SNPs, ordered by chromosome. From top to bottom: P-values for change in allele frequency ( $\Delta af$ ), Mathieson and McVean ( $mm$ ) (14),  $spatpg$  (15), and  $DCMS$  (55).

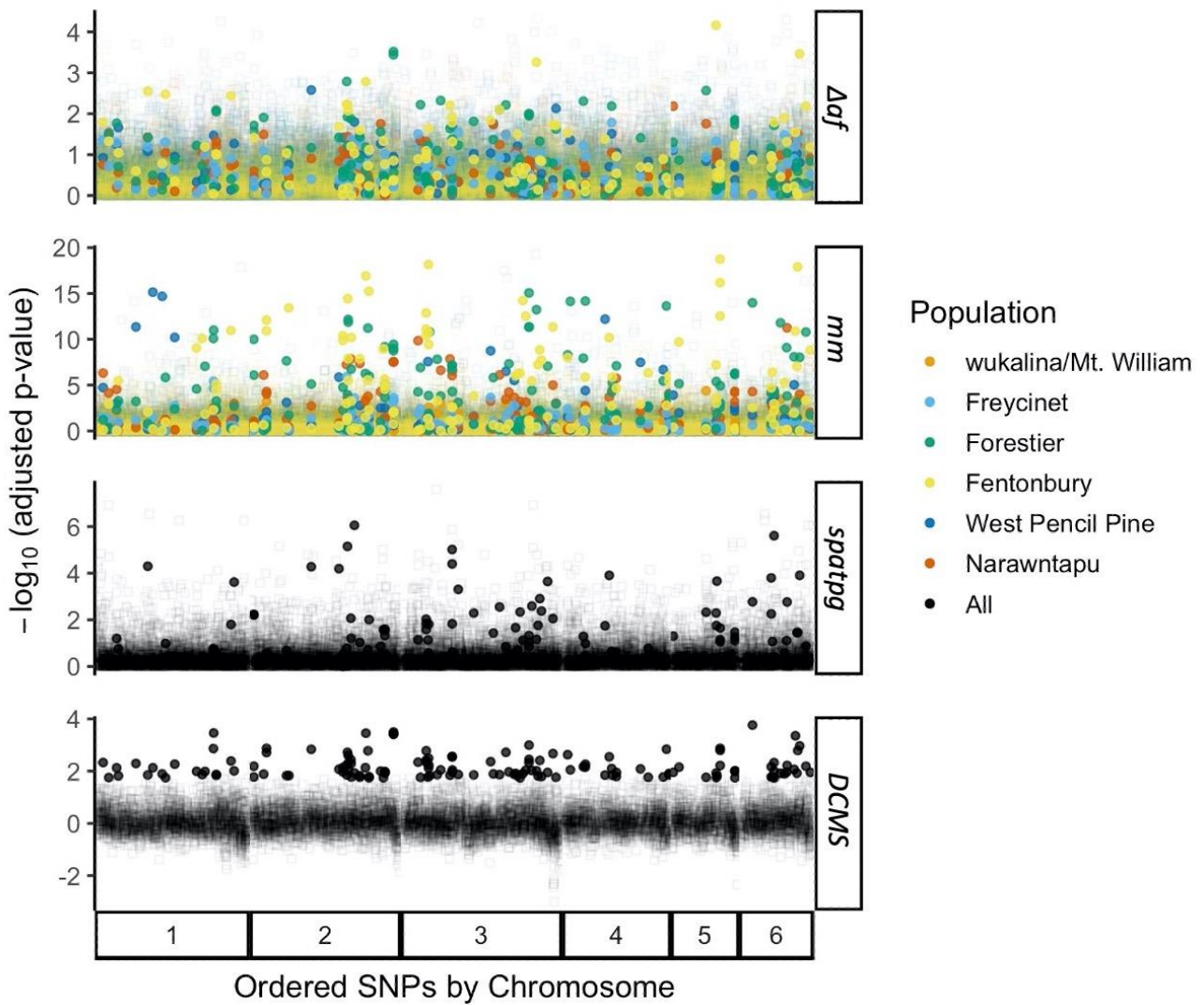
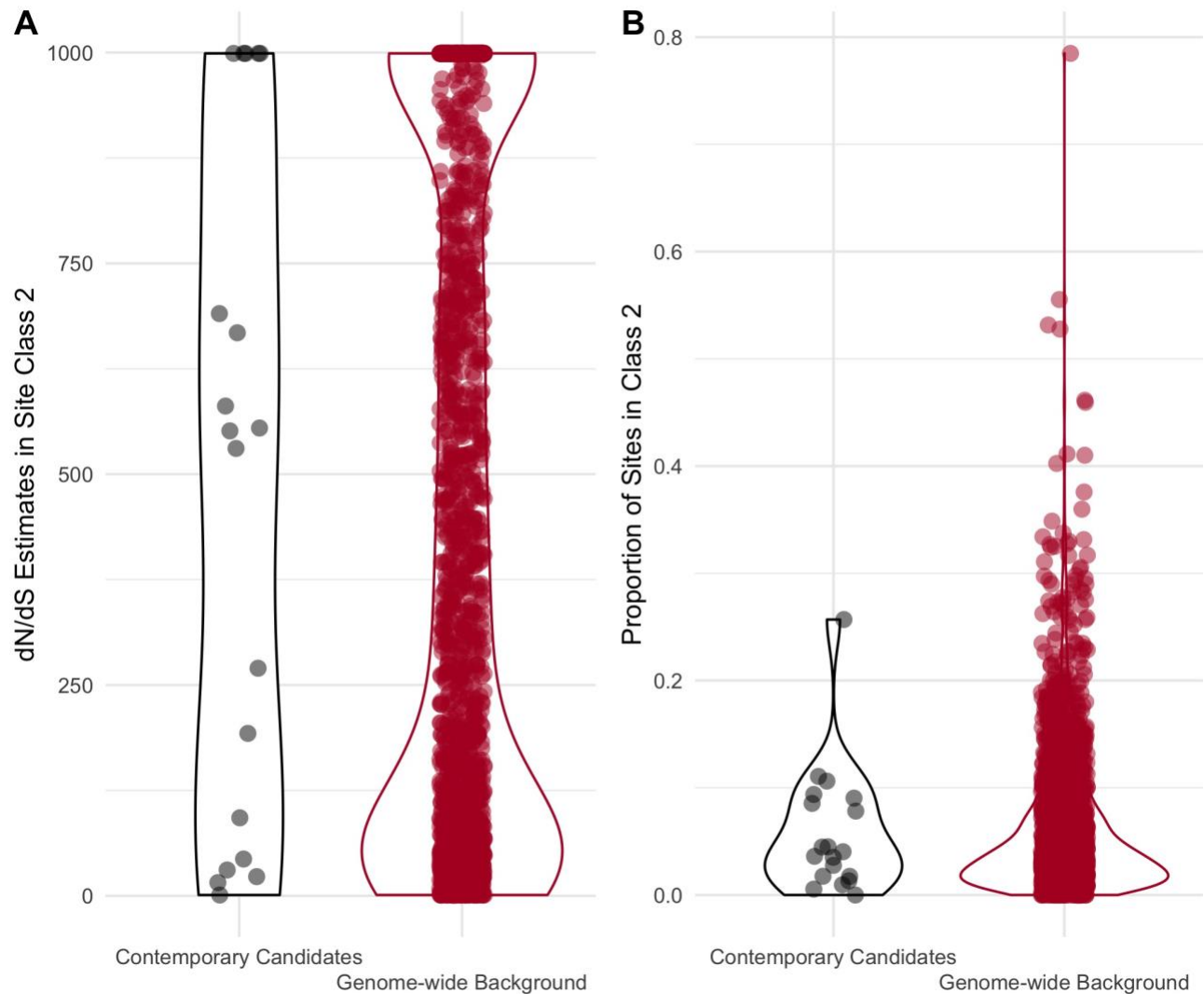


Figure 3. Distributions of statistically significant historical positive selection (57) for contemporary candidates (N=19) and the genome-wide background (N=1,982). Each point represents the results for a single gene. A) Estimates of dN/dS and B) proportion of sites classified as positive selection in the devil branch.



## References

1. Barreiro LB, Quintana-Murci L. From evolutionary genetics to human immunology: how selection shapes host defence genes. *Nat Rev Genet.* 2010;11(1):17-30.
2. Epstein B, Jones M, Hamede R, Hendricks S, McCallum H, Murchison EP, et al. Rapid evolutionary response to a transmissible cancer in Tasmanian devils. *Nature communications.* 2016;7:12684.
3. De Castro F, Bolker B. Mechanisms of disease-induced extinction. *Ecology Letters.* 2005;8(1):117-26.
4. Smith KF, Sax DF, Lafferty KD. Evidence for the role of infectious disease in species extinction and endangerment. *Conservation biology.* 2006;20(5):1349-57.
5. Jones KE, Patel NG, Levy MA, Storeygard A, Balk D, Gittleman JL, et al. Global trends in emerging infectious diseases. *Nature.* 2008;451(7181):990-3.
6. Lafferty KD. The ecology of climate change and infectious diseases. *Ecology.* 2009;90(4):888-900.
7. McCallum H, Dobson A. Detecting disease and parasite threats to endangered species and ecosystems. *Trends in ecology & evolution.* 1995;10(5):190-4.
8. Storfer A, Kozakiewicz CP, Beer MA, Savage AE. Applications of Population Genomics for Understanding and Mitigating Wildlife Disease. 2020.
9. McKnight DT, Schwarzkopf L, Alford RA, Bower DS, Zenger KR. Effects of emerging infectious diseases on host population genetics: a review. *Conservation genetics.* 2017;18(6):1235-45.
10. Brandies P, Peel E, Hogg CJ, Belov K. The Value of Reference Genomes in the Conservation of Threatened Species. *Genes.* 2019;10(11):846.
11. Li W-H. Unbiased estimation of the rates of synonymous and nonsynonymous substitution. *Journal of molecular evolution.* 1993;36(1):96-9.
12. Yang Z. Likelihood ratio tests for detecting positive selection and application to primate lysozyme evolution. *Molecular biology and evolution.* 1998;15(5):568-73.
13. Shultz AJ, Sackton TB. Immune genes are hotspots of shared positive selection across birds and mammals. *Elife.* 2019;8:e41815.
14. Mathieson I, McVean G. Estimating selection coefficients in spatially structured populations from time series data of allele frequencies. *Genetics.* 2013;193(3):973-84.
15. Gompert Z. Bayesian inference of selection in a heterogeneous environment from genetic time-series data. *Mol Ecol.* 2016;25(1):121-34.
16. Alves JM, Carneiro M, Cheng JY, de Matos AL, Rahman MM, Loog L, et al. Parallel adaptation of rabbit populations to myxoma virus. *Science.* 2019;363(6433):1319-26.
17. Monello RJ, Galloway NL, Powers JG, Madsen-Bouterse SA, Edwards WH, Wood ME, et al. Pathogen-mediated selection in free-ranging elk populations infected by chronic wasting disease. *Proceedings of the National Academy of Sciences.* 2017;114(46):12208-12.
18. Lilley TM, Wilson IW, Field KA, Reeder DM, Vodzak ME, Turner GG, et al. Genome-wide changes in genetic diversity in a population of *Myotis lucifugus* affected by white-nose syndrome. *G3: Genes, Genomes, Genetics.* 2020;10(6):2007-20.

19. Baird NA, Etter PD, Atwood TS, Currey MC, Shiver AL, Lewis ZA, et al. Rapid SNP discovery and genetic mapping using sequenced RAD markers. *PloS one*. 2008;3(10):e3376.
20. Andrews KR, Good JM, Miller MR, Luikart G, Hohenlohe PA. Harnessing the power of RADseq for ecological and evolutionary genomics. *Nature Reviews Genetics*. 2016;17(2):81.
21. Hawkins CE, Baars C, Hesterman H, Hocking GJ, Jones ME, Lazenby B, et al. Emerging disease and population decline of an island endemic, the Tasmanian devil *Sarcophilus harrisii*. *Biological Conservation*. 2006;131(2):307-24.
22. Pearse AM, Swift K. Allograft theory: transmission of devil facial-tumour disease. *Nature*. 2006;439(7076):549.
23. Hamilton DG, Jones ME, Cameron EZ, McCallum H, Storfer A, Hohenlohe PA, et al. Rate of intersexual interactions affects injury likelihood in Tasmanian devil contact networks. *Behavioral Ecology*. 2019;30(4):1087-95.
24. Hamede RK, McCallum H, Jones M. Seasonal, demographic and density-related patterns of contact between Tasmanian devils (*Sarcophilus harrisii*): Implications for transmission of devil facial tumour disease. *Austral Ecology*. 2008;33(5):614-22.
25. Pye R, Hamede R, Siddle HV, Caldwell A, Knowles GW, Swift K, et al. Demonstration of immune responses against devil facial tumour disease in wild Tasmanian devils. *Biol Lett*. 2016;12(10).
26. Siddle HV, Kreiss A, Tovar C, Yuen CK, Cheng Y, Belov K, et al. Reversible epigenetic down-regulation of MHC molecules by devil facial tumour disease illustrates immune escape by a contagious cancer. *Proceedings of the National Academy of Sciences*. 2013;110(13):5103-8.
27. Murchison EP, Tovar C, Hsu A, Bender HS, Kheradpour P, Rebbeck CA, et al. The Tasmanian devil transcriptome reveals Schwann cell origins of a clonally transmissible cancer. *Science*. 2010;327(5961):84-7.
28. Lazenby BT, Tobler MW, Brown WE, Hawkins CE, Hocking GJ, Hume F, et al. Density trends and demographic signals uncover the long-term impact of transmissible cancer in Tasmanian devils. *Journal of Applied Ecology*. 2018;55(3):1368-79.
29. McCallum H, Jones M, Hawkins C, Hamede R, Lachish S, Sinn DL, et al. Transmission dynamics of Tasmanian devil facial tumor disease may lead to disease-induced extinction. *Ecology*. 2009;90(12):3379-92.
30. Brüniche-Olsen A, Jones ME, Burrridge CP, Murchison EP, Holland BR, Austin JJ. Ancient DNA tracks the mainland extinction and island survival of the Tasmanian devil. *Journal of biogeography*. 2018;45(5):963-76.
31. Brüniche-Olsen A, Jones ME, Austin JJ, Burrridge CP, Holland BR. Extensive population decline in the Tasmanian devil predates European settlement and devil facial tumour disease. *Biology letters*. 2014;10(11):20140619.
32. Patton AH, Margres MJ, Stahlke AR, Hendricks S, Lewallen K, Hamede RK, et al. Contemporary demographic reconstruction methods are robust to genome assembly quality: A case study in Tasmanian Devils. *Molecular biology and evolution*. 2019;36(12):2906-21.
33. Siddle HV, Kreiss A, Eldridge MDB, Noonan E, Clarke CJ, Pyecroft S, et al. Transmission of a fatal clonal tumor by biting occurs due to depleted MHC diversity in a threatened carnivorous marsupial. *Proceedings of the National Academy of Sciences*. 2007;104(41):16221-6.
34. Pye RJ, Pemberton D, Tovar C, Tubio JMC, Dun KA, Fox S, et al. A second transmissible cancer in Tasmanian devils. *Proceedings of the National Academy of Sciences*. 2016;113(2):374-9.

35. James S, Jennings G, Kwon YM, Stammnitz M, Fraik A, Storfer A, et al. Tracing the rise of malignant cell lines: Distribution, epidemiology and evolutionary interactions of two transmissible cancers in Tasmanian devils. *Evolutionary applications*. 2019;12(9):1772-80.
36. Stammnitz MR, Coorens THH, Gori KC, Hayes D, Fu B, Wang J, et al. The origins and vulnerabilities of two transmissible cancers in Tasmanian devils. *Cancer Cell*. 2018;33(4):607-19.
37. Deakin JE, Bender HS, Pearse A-M, Rens W, O'Brien PCM, Ferguson-Smith MA, et al. Genomic restructuring in the Tasmanian devil facial tumour: chromosome painting and gene mapping provide clues to evolution of a transmissible tumour. *PLoS genetics*. 2012;8(2):e1002483.
38. Peck SJ, Michael SA, Knowles G, Davis A, Pemberton D. Causes of mortality and severe morbidity requiring euthanasia in captive Tasmanian devils (*Sarcophilus harrisii*) in Tasmania. *Australian veterinary journal*. 2019;97(4):89-92.
39. Hamede RK, McCallum H, Jones M. Biting injuries and transmission of Tasmanian devil facial tumour disease. *Journal of Animal Ecology*. 2013;82(1):182-90.
40. Patchett AL, Flies AS, Lyons AB, Woods GM. Curse of the devil: molecular insights into the emergence of transmissible cancers in the Tasmanian devil (*Sarcophilus harrisii*). *Cellular and Molecular Life Sciences*. 2020.
41. Jones ME, Cockburn A, Hamede R, Hawkins C, Hesterman H, Lachish S, et al. Life-history change in disease-ravaged Tasmanian devil populations. *Proceedings of the National Academy of Sciences*. 2008;105(29):10023-7.
42. Wells K, Hamede RK, Jones ME, Hohenlohe PA, Storfer A, McCallum HI. Individual and temporal variation in pathogen load predicts long-term impacts of an emerging infectious disease. *Ecology*. 2019;100(3):e02613.
43. Hogg CJ, Lee AV, Srb C, Hibbard C. Metapopulation management of an Endangered species with limited genetic diversity in the presence of disease: the Tasmanian devil *Sarcophilus harrisii*. *International Zoo Yearbook*. 2017;51(1):137-53.
44. Hohenlohe PA, McCallum HI, Jones ME, Lawrance MF, Hamede RK, Storfer A. Conserving adaptive potential: lessons from Tasmanian devils and their transmissible cancer. *Conservation Genetics*. 2019;20(1):81-7.
45. Wright BR, Farquharson KA, McLennan EA, Belov K, Hogg CJ, Grueber CE. A demonstration of conservation genomics for threatened species management. *Molecular Ecology Resources*. 2020.
46. Grueber CE, Peel E, Wright B, Hogg CJ, Belov K. A Tasmanian devil breeding program to support wild recovery. *Reproduction, Fertility and Development*. 2019;31(7):1296-304.
47. Hubert J-N, Zerjal T, Hospital F. Cancer-and behavior-related genes are targeted by selection in the Tasmanian devil (*Sarcophilus harrisii*). *PloS one*. 2018;13(8):e0201838.
48. Fraik AK, Margres MJ, Epstein B, Barbosa S, Jones M, Hendricks S, et al. Disease swamps molecular signatures of genetic-environmental associations to abiotic factors in Tasmanian devil (*Sarcophilus harrisii*) populations. *Evolution*. 2020;n/a(n/a).
49. Ali OA, O'Rourke SM, Amish SJ, Meek MH, Luikart G, Jeffres C, et al. RAD Capture (Rapture): Flexible and efficient sequence-based genotyping. *Genetics*. 2016;202(2):389-400.
50. Margres MJ, Jones ME, Epstein B, Kerlin DH, Comte S, Fox S, et al. Large-effect loci affect survival in Tasmanian devils (*Sarcophilus harrisii*) infected with a transmissible cancer. *Molecular ecology*. 2018;27(21):4189-99.

51. Mitchell KJ, Pratt RC, Watson LN, Gibb GC, Llamas B, Kasper M, et al. Molecular phylogeny, biogeography, and habitat preference evolution of marsupials. *Molecular biology and evolution*. 2014;31(9):2322-30.
52. Mikkelsen TS, Wakefield MJ, Aken B, Amemiya CT, Chang JL, Duke S, et al. Genome of the marsupial *Monodelphis domestica* reveals innovation in non-coding sequences. *Nature*. 2007;447(7141):167-77.
53. Renfree MB, Papenfuss AT, Deakin JE, Lindsay J, Heider T, Belov K, et al. Genome sequence of an Australian kangaroo, *Macropus eugenii*, provides insight into the evolution of mammalian reproduction and development. *Genome biology*. 2011;12(8):1-26.
54. Johnson RN, O'Meally D, Chen Z, Etherington GJ, Ho SYW, Nash WJ, et al. Adaptation and conservation insights from the koala genome. *Nature genetics*. 2018;50(8):1102.
55. Ma Y, Ding X, Qanbari S, Weigend S, Zhang Q, Simianer H. Properties of different selection signature statistics and a new strategy for combining them. *Heredity*. 2015;115(5):426.
56. Aken BL, Achuthan P, Akanni W, Amode MR, Bernsdorff F, Bhai J, et al. Ensembl 2017. *Nucleic acids research*. 2016;45(D1):D635-D42.
57. Yang Z. PAML 4: phylogenetic analysis by maximum likelihood. *Molecular biology and evolution*. 2007;24(8):1586-91.
58. Yang Z. PAML: a program package for phylogenetic analysis by maximum likelihood. *Bioinformatics*. 1997;13(5):555-6.
59. Massey Jr FJ. The Kolmogorov-Smirnov test for goodness of fit. *Journal of the American statistical Association*. 1951;46(253):68-78.
60. Scholz FW, Stephens MA. K-sample Anderson-Darling tests. *Journal of the American Statistical Association*. 1987;82(399):918-24.
61. Szkiba D, Kapun M, von Haeseler A, Gallach M. SNP2GO: functional analysis of genome-wide association studies. *Genetics*. 2014;197(1):285-9.
62. Mi H, Muruganujan A, Ebert D, Huang X, Thomas PD. PANTHER version 14: more genomes, a new PANTHER GO-slim and improvements in enrichment analysis tools. *Nucleic acids research*. 2018;47(D1):D419-D26.
63. Wright B, Willet CE, Hamede R, Jones M, Belov K, Wade CM. Variants in the host genome may inhibit tumour growth in devil facial tumours: evidence from genome-wide association. *Sci Rep*. 2017;7(1):423.
64. Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proceedings of the National Academy of Sciences*. 2005;102(43):15545-50.
65. Wu K, Li Z, Cai S, Tian L, Chen K, Wang J, et al. EYA1 phosphatase function is essential to drive breast cancer cell proliferation through cyclin D1. *Cancer research*. 2013;73(14):4488-99.
66. Cai S, Cheng X, Liu Y, Lin Z, Zeng W, Yang C, et al. EYA1 promotes tumor angiogenesis by activating the PI3K pathway in colorectal cancer. *Experimental cell research*. 2018;367(1):37-46.
67. Meurs EF, Galabru J, Barber GN, Katze MG, Hovanessian AG. Tumor suppressor function of the interferon-induced double-stranded RNA-activated protein kinase. *Proceedings of the National Academy of Sciences*. 1993;90(1):232-6.
68. Reya T, Clevers H. Wnt signalling in stem cells and cancer. *Nature*. 2005;434(7035):843-50.

69. Margres MJ, Ruiz-Aravena M, Hamede R, Chawla K, Patton AH, Lawrance MF, et al. Spontaneous tumor regression in Tasmanian devils associated with RASL11A activation. *Genetics*. 2020.
70. Brüniche-Olsen A, Austin JJ, Jones ME, Holland BR, Burr ridge CP. Detecting selection on temporal and spatial scales: a genomic time-series assessment of selective responses to devil facial tumor disease. *PloS one*. 2016;11(3).
71. Hendricks S, Epstein B, Schönfeld B, Wiench C, Hamede R, Jones M, et al. Conservation implications of limited genetic diversity and population structure in Tasmanian devils (*Sarcophilus harrisii*). *Conservation Genetics*. 2017;18(4):977-82.
72. Fang WH, Wang Q, Li HM, Ahmed M, Kumar P, Kumar S. PAX 3 in neuroblastoma: oncogenic potential, chemosensitivity and signalling pathways. *Journal of cellular and molecular medicine*. 2014;18(1):38-48.
73. Margres MJ, Ruiz-Aravena M, Hamede R, Jones ME, Lawrance MF, Hendricks SA, et al. The genomic basis of tumor regression in Tasmanian devils (*Sarcophilus harrisii*). *Genome biology and evolution*. 2018;10(11):3012-25.
74. Zhang J. Frequent false detection of positive selection by the likelihood method with branch-site models. *Molecular Biology and Evolution*. 2004;21(7):1332-9.
75. Kosiol C, Vinař T, da Fonseca RR, Hubisz MJ, Bustamante CD, Nielsen R, et al. Patterns of positive selection in six mammalian genomes. *PLoS genetics*. 2008;4(8).
76. Nelson WJ, Nusse R. Convergence of Wnt,  $\beta$ -catenin, and cadherin pathways. *Science*. 2004;303(5663):1483-7.
77. Wakefield MJ, Graves JAM. Marsupials and monotremes sort genome treasures from junk. *Genome biology*. 2005;6(5):218.
78. Metzger MJ, Reinisch C, Sherry J, Goff SP. Horizontal transmission of clonal cancer cells causes leukemia in soft-shell clams. *Cell*. 2015;161(2):255-63.
79. Strakova A, Murchison EP. The cancer which survived: insights from the genome of an 11000 year-old cancer. *Curr Opin Genet Dev*. 2015;30:49-55.
80. Beene DL, Scott JD. A-kinase anchoring proteins take shape. *Current opinion in cell biology*. 2007;19(2):192-8.
81. Li S, Chen P-L, Subramanian T, Chinnadurai G, Tomlinson G, Osborne CK, et al. Binding of CtIP to the BRCT repeats of BRCA1 involved in the transcription regulation of p21 is disrupted upon DNA damage. *Journal of Biological Chemistry*. 1999;274(16):11334-8.
82. Oliver B, Parisi M, Clark D. Gene expression neighborhoods. *Journal of biology*. 2002;1(1):4.
83. Bonneaud C, Balenger SL, Russell AF, Zhang J, Hill GE, Edwards SV. Rapid evolution of disease resistance is accompanied by functional changes in gene expression in a wild bird. *Proceedings of the National Academy of Sciences*. 2011;108(19):7866-71.
84. Guiler E. *The Tasmanian devil*: St. David's Park Pub.; 1992.
85. Casanova J-L, Abel L, editors. *Human genetics of infectious diseases: Unique insights into immunological redundancy* 2018 2018: Elsevier.
86. Miller IF, Metcalf CJE. Evolving resistance to pathogens. *Science*. 2019;363(6433):1277-8.
87. Gandon S, Michalakis Y. Evolution of parasite virulence against qualitative or quantitative host resistance. *Proceedings of the Royal Society of London Series B: Biological Sciences*. 2000;267(1447):985-90.

88. Pybus OG, Rambaut A. Evolutionary analysis of the dynamics of viral infectious disease. *Nature Reviews Genetics*. 2009;10(8):540-50.
89. Kardos M, Shafer ABA. The Peril of Gene-Targeted Conservation. *Trends Ecol Evol*. 2018;33(11):827-39.
90. McLennan EA, Grueber CE, Wise P, Belov K, Hogg CJ. Mixing genetically differentiated populations successfully boosts diversity of an endangered carnivore. *Animal Conservation*. 2020.
91. Wright B, Morris K, Grueber CE, Willet CE, Gooley R, Hogg CJ, et al. Development of a SNP-based assay for measuring genetic diversity in the Tasmanian devil insurance population. *BMC Genomics*. 2015;16:791.
92. Campbell NR, Harmon SA, Narum SR. Genotyping-in-Thousands by sequencing (GT-seq): A cost effective SNP genotyping method based on custom amplicon sequencing. *Mol Ecol Resour*. 2015;15(4):855-67.
93. von Thaden A, Nowak C, Tiesmeyer A, Reiners TE, Alves PC, Lyons LA, et al. Applying genomic data in wildlife monitoring: Development guidelines for genotyping degraded samples with reduced single nucleotide polymorphism panels. *Molecular Ecology Resources*. 2020;20(3).