Deeply sequenced infectious clones of key cassava begomovirus

isolates from Cameroon

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Running head: Full sequences for key clones of ACMV and EACMCV

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Abstract

We Illumina-sequenced two pairs of widely used infectious clones (4

plasmids) of the bipartite begomoviruses African cassava mosaic virus

(ACMV) and East African cassava mosaic Cameroon virus. The

ACMV clones were quite divergent from published expectations. Raw

reads, consensus plasmid sequences, and the infectious clones

themselves are all publicly available.

Main text

Infectious clones are the central tool of molecular virology. Cloned

isolates of African cassava mosaic virus (ACMV) and East African

cassava mosaic Cameroon virus (EACMCV) provided conclusive

proof of synergy between two major clades of cassava begomoviruses

(1), which is a defining feature of the epidemic of mosaic disease that

has devastated cassava production in sub-Saharan Africa (2, 3). Herein

we announce new sequence resources for these frequently used clones,

which confirm the EACMCV clones and clarify the identity of the

ACMV clones.

Complete and accurate plasmid sequences considerably simplify

molecular analysis of infectious clones and design of new constructs.

To confirm the sequences of the four plasmids of interest, libraries

were prepared from Covaris-sheared plasmid DNA in triplicate with

the NEBNext Ultra II kit and sequenced on the Illumina NextSeq 500

platform in the 150 bp paired-end read configuration.

These plasmids (1, 4, 5) had not been fully sequenced, but deduced

sequence maps (including partial-tandem-dimer virus segment inserts)

were available. Reads were trimmed with CutAdapt 1.16 (6) and

aligned to these sequences with the Burrows-Wheeler Aligner (BWA-

MEM v0.7.13 (7)). Variants relative to the reference sequence were

identified with samtools v1.8 (8) and VarScan v2.4.4 (9).

The EACMCV DNA-A and DNA-B clones had four and two small

differences relative to their corresponding GenBank accessions

(AF112354.1 and FJ826890.1). The consensus sequences of the

ACMV clones, however, were 3.1% and 5.8% divergent from the

sequences (AF112352.1 and AF112353.1) originally reported by

Fondong et al. (1), as calculated with Sequence Demarcation Tool v1.2

(10). This difference was not entirely unexpected, because of the

parallel history of two sets of ACMV clones: infectious partial-

tandem-dimer clones were made via restriction-digestion/ligation from

sap-inoculated *Nicotiana benthamiana* plants whereas the monomer-

segment-unit clones were cloned with PCR from the same original

cassava field sample (1). We expect these complete infectious clone

sequences will be of great utility to the community, given that many

follow-up publications (11–22) specifically referenced the related but

non-identical monomer sequences (AF112352.1 and AF112353.1).

We have deposited consensus sequences for the four plasmids and the

relevant ACMV segments and have additionally deposited the

plasmids with Addgene to simplify sharing and ensure their continued

availability. The deep coverage (over 20,000-fold; Figure 1), which

was consistent across three technical sequencing replicates, assures the

correctness of the partial-tandem-dimer sequences.

Our results underscore the value of confirming the sequence of

molecular clones. An additional example from the geminivirus

literature is the Nigerian Ogoroco clone of ACMV (23). Kittelmann et

al. (24) thoroughly documented that the infectious DNA-A clone is not

100% identical to the related GenBank accession (AJ427910.1). We

do not believe the large literature on the Cameroonian ACMV isolates

requires dramatic reinterpretation, but we recommend using the

revised sequences described here for future analyses.

Data availability: Plasmids will be made publicly available via

Addgene (plasmid # 159134 to 159137). Sequences for full plasmids

(MT856192 to MT856195) and ACMV segments (MT858793 and

MT858794) are available in GenBank. Raw Illumina data are available

from the NCBI Sequence Read Archive (PRJNA649777).

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Figure 1. Plots of Illumina read depth across the length of the four infectious clone plasmids. The region in each plasmid corresponding each virus segment partial-tandem-dimer unit is indicated with a black line under each graph. White lines demarcate the boundaries of the unique and duplicated regions of the concatamer. Each virus segment monomer unit (in between two replication-origin nick sites) is shown in blue. Canonical virus genes are indicated with gray arrows, left to right for virus sense (AV1, AV2, BV1) and right to left for complementary sense (AC1 to AC4, BC1). Uneven read depth for the ACMV DNA-B plasmid is due to instability (truncation) in *E. coli*, which is evident in single-cut restriction digests.

References

References

1.   Fondong VN, Pita JS, Rey MEC, de Kochko A, Beachy RN, Fauquet CM. 2000. Evidence
of synergism between African cassava mosaic virus and a new double-recombinant
geminivirus infecting cassava in Cameroon. Journal of General Virology 81:287–297.

2.   Pita JS, Fondong VN, Sangaré A, Otim-Nape GW, Ogwal S, Fauquet CM. 2001.
Recombination, pseudorecombination and synergism of geminiviruses are determinant keys
to the epidemic of severe cassava mosaic disease in Uganda. Journal of General Virology
82:655–665.

3.   Mbewe W, Hanley-Bowdoin L, Ndunguru J, Duffy S. 2020. Cassava viruses:
Epidemiology, evolution, and management. *In* Ristaino, JB, Records, A (eds.), Emerging
plant diseases and global food security. The American Phytopathological Society, St. Paul.

4.   Fondong VN, Chen K. 2011. Genetic variability of East African cassava mosaic Cameroon
virus under field and controlled environment conditions. Virology 413:275–282.

5.   Chowda Reddy RV, Dong W, Njock T, Rey MEC, Fondong VN. 2012. Molecular
interaction between two cassava geminiviruses exhibiting cross-protection. Virus Research
163:169–177.

6.   Martin M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing
reads. EMBnet.journal 17:10–12.

7.   Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-
MEM. arXiv:13033997 [q-bio].

8.   Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin

R. 2009. The Sequence Alignment/Map format and SAMtools. Bioinformatics 25:2078–

2079.

9.   Koboldt DC, Zhang Q, Larson DE, Shen D, McLellan MD, Lin L, Miller CA, Mardis ER,

Ding L, Wilson RK. 2012. VarScan 2: Somatic mutation and copy number alteration

discovery in cancer by exome sequencing. Genome Res 22:568–576.

10.   Muhire BM, Varsani A, Martin DP. 2014. SDT: A virus classification tool based on

pairwise sequence alignment and identity calculation. PLOS ONE 9:e108277.

11.   Vanitharani R, Chellappan P, Fauquet CM. 2003. Short interfering RNA-mediated

interference of gene expression and viral DNA accumulation in cultured plant cells. PNAS

100:9632–9636.

12.   Vanitharani R, Chellappan P, Pita JS, Fauquet CM. 2004. Differential roles of AC2 and

AC4 of cassava geminiviruses in mediating synergism and suppression of

posttranscriptional gene silencing. J Virol 78:9487–9498.

13.   Chellappan P, Vanitharani R, Fauquet CM. 2004. Short interfering RNA accumulation

correlates with host recovery in DNA virus-infected hosts, and gene silencing targets

specific viral sequences. J Virol 78:7465–7477.

14.   Chellappan P, Vanitharani R, Fauquet CM. 2005. MicroRNA-binding viral protein

interferes with Arabidopsis development. PNAS 102:10381–10386.

15.   Chellappan P, Vanitharani R, Ogbe F, Fauquet CM. 2005. Effect of temperature on

geminivirus-induced RNA silencing in plants. Plant Physiol 138:1828–1841.

16.   Amin I, Patil BL, Briddon RW, Mansoor S, Fauquet CM. 2011. A common set of

developmental miRNAs are upregulated in *Nicotiana benthamiana* by diverse

begomoviruses. Virology Journal 8:143.

17.   Reyes MI, Nash TE, Dallas MM, Ascencio-Ibáñez JT, Hanley-Bowdoin L. 2013. Peptide

aptamers that bind to geminivirus replication proteins confer a resistance phenotype to

tomato yellow leaf curl virus and tomato mottle virus infection in tomato. Journal of

Virology 87:9691–9706.

18.   Ndunguru J, León LD, Doyle CD, Sseruwagi P, Plata G, Legg JP, Thompson G, Tohme J,

Aveling T, Ascencio-Ibáñez JT, Hanley-Bowdoin L. 2016. Two novel DNAs that enhance

symptoms and overcome CMD2 resistance to cassava mosaic disease. Journal of Virology

90:4160–4173.

19.   Patil BL, Bagewadi B, Yadav JS, Fauquet CM. 2016. Mapping and identification of cassava

mosaic geminivirus DNA-A and DNA-B genome sequences for efficient siRNA expression

and RNAi based virus resistance by transient agro-infiltration studies. Virus Research

213:109–115.

20.   Beyene G, Chauhan RD, Wagaba H, Moll T, Alicai T, Miano D, Carrington JC, Taylor NJ.

2016. Loss of CMD2-mediated resistance to cassava mosaic disease in plants regenerated

through somatic embryogenesis. Molecular Plant Pathology 17:1095–1110.

21. Kuria P, Ilyas M, Ateka E, Miano D, Onguso J, Carrington JC, Taylor NJ. 2017.

Differential response of cassava genotypes to infection by cassava mosaic geminiviruses.

Virus Research 227:69–81.

22. Chauhan RD, Beyene G, Taylor NJ. 2018. Multiple morphogenic culture systems cause loss

of resistance to cassava mosaic disease. BMC Plant Biology 18:132.

23. Briddon RW, Liu S, Pinner MS, Markham PG. 1998. Infectivity of African cassava mosaic

virus clones to cassava by biolistic inoculation. Arch Virol 143:2487–2492.

24. Kittelmann K, Rau P, Gronenborn B, Jeske H. 2009. Plant geminivirus Rep protein induces

rereplication in fission yeast. Journal of Virology 83:6769–6778.