Investigation of Capsule-Inspired Neural Network Approaches for DNA Methylation

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Abstract

DNA methylation (DNAm) alterations are implicated with aging and diseases by regulating gene expression. DNAm deep-learning approaches can capture features associated with aging, cell type, and disease progression, but lack incorporation of prior biological knowledge. We present deep-learning software, *MethylCapsNet* and *MethylSPWNet*, that group CpGs into user-specified or predefined biologically relevant groupings (eg. gene promoter or CpG island context) related to diagnostic and prognostic outcomes. We train our models on a cohort (n=3,897) to classify central nervous system tumors and compare to existing approaches. Our methodology presents opportunities to increase interpretability of disease mechanisms through utilization of biologically relevant annotations.

Background

DNA methylation (DNAm) is a key epigenetic regulator of gene expression in health and disease states, processes of aging and cellular differentiation/stemness, and response to environmental exposures [1–3]. DNAm of cytosine in the context of cytosine-guanine dinucleotides (CpG) sites can be measured with standardized genome-scale oligonucleotide bead-arrays at hundreds of thousands of sites [4,5]. Though a CpG is either unmethylated or methylated, fluorescence signal intensities from array measures of bulk biospecimen DNA are used to derive a beta value measure which approximates the proportion of methylated DNA copies. Gene promoter CpG island methylation is associated with repression of transcription, whereas unmethylated CpG islands are permissive to gene transcription. Alterations to DNAm have a well-established role in carcinogenesis and tumor progression, including inactivation of tumor suppressor genes, aberrant

oncogene expression, and loss of repression of repetitive element sequences that contribute to genomic instability [6,7].

According to the current World Health Organization classification scheme, there are over 38 known Central Nervous System (CNS) tumor subtypes defined by histopathological features that are mostly representative of glioma, ependymoma, and embryonal subtypes.[8] Within those categorizations, over 80 further delineations are specified by molecular subtyping. DNAm alterations have been heavily implicated in the development and prognosis of CNS tumors. For instance, epigenetic silencing of *MGMT* is associated with an improved response to chemotherapy in glioblastoma patients through the deactivation of crucial DNA repair mechanisms[9]. *IDH* mutations are associated with improved survival in glioma patients through subsequent global hypermethylation of CpG island promoters, known as induction of the CpG Island Methylator Phenotype (CIMP)[10–13]. Other examples include hypermethylation of WNT and SHH pathways in medulloblastoma patients.[14] The success of differential methylation analyses in characterizing CNS tumors has recently led to the development of DNAm classifiers of brain tumors as companion diagnostic tools to understand and correctly diagnose challenging histologic cases and for selection of targeted therapies[8].

While the development of this methylation-based brain-tumor machine learning classifier has been heralded as an improvement, existing diagnostic framework clinically applicable classifiers use only a small subset of measured CpGs, in this case, 10,000 CpGs [15]. While the scope of features used provides for an accurate DNAm prediction model, incorporating additional CpG predictors, of which may be better suited for machine learning analyses by merit of their

prohibitive dimensionality, may allow the resolution of tumor classes otherwise not identified, and aid in understanding relationships with outcomes [16]. Deep learning algorithms are a subclass of machine learning approaches that are based on the use of artificial neural networks (ANN) [17–19]. Multi-layer perceptrons (MLP) represent a subclass of neural networks that treat the input data as a one-dimensional vector and then pass the information from one set of nodes to a subsequent set of nodes through fully connected layers of weights/parameters. The information at the subsequent layer of nodes is transformed using non-linear transforms/activations/link functions such as the sigmoid (logistic), hyperbolic tangent (tanh), or rectified linear unit (ReLU) transforms. These types of analyses are common for deep learning analyses of DNAm data, where the input data is a list of beta values for each subject[20].

DNAm deep learning frameworks, such as MethylNet, can accurately characterize tissue, disease states, and infer subject age and cell type proportions through unsupervised embedding, generation, classification, and regression tasks,[20–24] while attempting to ascribe important methylated loci through the use of model interpretability frameworks such as SHAP [25] or LIME[26]. While the inclusion of more CpGs presents an opportunity to expand the space of biologically testable hypotheses [20], the CpGs included in these classifiers have been poorly characterized; statistical challenges (e.g., multicollinearity) with interpretations and generation of associations with pathways remain understudied.[27]

Multicollinearity, the unusually high correlation between features, can be addressed with careful feature-selection or grouping, such as with elastic net penalization[1,28]. Feature selection methods and statistical learning methods such as sparse Group LASSO and network

regularization have identified important CpGs in highly complex data.[29–33] More recent work has called for a greater understanding of the implications of DNAm-DNAm interactions through the incorporation of Gaussian Graphical Models, Canonical Correlation Analysis, and module discovery through weighted gene co-methylation networks. [34–50] There is growing support for the use of novel deep learning methods to aggregate, group, and select CpGs by their local context (e.g., genes) in an effort to connect and interpret the data with clinical outcomes.[51–53] Incorporation of prior biological knowledge not only improves the transparency and interpretability of the modeling approach but also reduces noise while increasing signal by meaningfully pruning redundant relationships between predictors. [54]

Capsule Networks are a promising approach for grouping CpGs that harness their statistical interactions and relate the groupings of predictors to clinical and biological outcomes.[27] Capsule networks explicitly model the relationship between images or genomic information by its constituent parts, or capsules, by parameterizing pose matrices (unitary transformations), and then hierarchically associate each of these parts independently to higher-order targets of interest. In addition, recent deep neural network modeling approaches have utilized Group LASSO penalties for deep neural networks. [55]

Here we provide a framework for Capsule Network inspired deep learning with methylation data. We investigated the organization of CpG features into DNAm capsules, which represent local contexts that can be related to one another. *MethylSPWNet* and *MethylCapsNet* organize sets of CpGs into a series of capsules defined by higher-order genomic contexts and performs classification tasks. To bring additional interpretability to existing deep learning approaches

while capturing hierarchical association networks, we propose and explore *MethylSPWNet* (Sparse Pathway Network) and *MethylCapsNet* as deep learning analogs of these enrichment tests. We provide recommendations for the growth of these capsule and network-deriving models and provide open-source software for training these models. There are a growing number of pertinent use cases for deep learning DNAm analyses (e.g., aging, cell-type estimation, disease diagnosis and prognosis); the *MethylCapsNet* framework proposes to expand the broad utility of these tools by allowing end-users to construct their unique capsules that represent an array of biologically plausible contexts that further explain their target of interest.

Results

To illustrate the potential utility of capsule-inspired neural network approaches, we revisited the Capper et al. [8] dataset used to train a model that differentiates CNS tumors [56]. CNS tumor histology is largely characterized by the presence or absence of morphologically distinct cells of origin, including neuronal, astrocytic, microglial, oligodendrocytic, and Schwann cells. We aimed to predict the 38 histological subtypes of CNS tumors (39 classes, including controls) as a test case for the capsule-inspired neural network approaches. We compare the *MethylCapsNet* and *MethylSPWNet* frameworks for capsule organization with the existing *MethylNet* framework (which does not account for Capsule organized information [20]), and with Group LASSO Logistic Regression Classifier organized by capsules. Details of modeling approaches, fitting procedures, and capsule selection are in the Methods section.

Classification Study

Table 1: Classification Results for Group LASSO, MethylNet, MethylSPWNet, and MethylCapsNet*

Approach	Accuracy±SE	Recall±SE	Precision±SE	F1-Score±SE
Group LASSO	0.077±0.0099	0.23±0.016	0.28±0.032	0.17±0.016
MethylNet	0.97±0.0061	0.99±0.0099	0.96±0.011	0.97±0.011
MethylSPWNet	0.98±0.0049	0.96±0.0088	0.96±0.0085	0.96±0.0089
MethylCapsNet	0.98±0.0044	0.98±0.012	0.98±0.01	0.98±0.011

*All scores are reported as macro-measures across all subtypes; 95% confidence intervals estimated using 1000-sample non-parametric bootstrap

We trained each of the modeling approaches to differentiate 38 histological subtypes of CNS tumors and compared their classification performance via a 1000-iteration non-parametric bootstrap of F1-scores over the test set, which balances sensitivity and specificity and reduces the bias in output. Our results indicate that *MethylNet*, *MethylSPWNet*, and *MethylCapsNet* can all achieve very similar high performance on a common data set (Table 1). Besides, the neural network approaches achieved considerably higher performance than the Group Logistic Regression approach. A breakdown of classification scores has been included in the supplementary material. Since all three approaches offer similar performance on classifying brain tumors, we next sought to uncover overlap or complementary insights provided by each modeling approach based on their organization of data. The high predictive accuracy of both capsule approaches provided grounds for exploring the factors related to its decision-making process for increased transparency and validation of our approach.

Clustering Gene-Level Brain Cancer Embeddings

Until this point, our unit of analysis has been individual CpGs. Summarizing gene-level methylation using median or mean methylation is generally not appropriate because the

relationship between methylation state and gene expression can vary depending on the genomic context (e.g., promoter and gene body). However, while training *MethylSPWNet* to predict tumor histological subtype, the model learns to generate gene-level summaries of methylation by updating the weight of each CpG when aggregating beta values of CpGs on the gene-level (See Methods "Description of MethylSPWNet"). This gene-level aggregation has the effect of transforming a design matrix of samples by CpGs into samples by genes. Gene-level embeddings correlated with outcome (here tumor histological subtype), can then be interrogated for their relationship with known pathways and gene networks. To visualize gene-level embeddings, we generated cluster heatmaps, where rows constitute observations and columns constitute genes, showing plots of the top 2000 variable genes from neural network gene-level embeddings (Supplementary Figures 1-2).

To assess the representation capacity of MethylNet, MethylCapsNet, and MethylSPWNet embeddings, we clustered embeddings with histologic tumor subtype, cell of origin, and histological subtype with the molecular subclass (Supplementary Table 1). Preliminary clustering of the observations demonstrates, for instance, the inability to differentiate *IDH* mutant subtypes of glioma when defined by median methylation versus the neural network parameterization. There exists observed concordance between hierarchical clustering this embedding space and the brain cancer subtypes, as either defined by their molecular subtypes, the original histological subtypes that the model was trained on, or higher-order cells of origin (e.g., mesenchymal, ependymal, neuro-glial origin). *MethylSPWNet* had the highest degree of concordance with histological and molecular subtypes within the gene-level embedding space [57] (V-Measure 0.72±0.0059; Supplementary Table 1). The ability to recapitulate relevant

histological subtypes of CNS tumors through the embeddings alone is further corroborated by embedding plots [57], where MethylCapsNet appears to generate the best separation and differentiation of subtypes (Silhouette Score: 0.52±0.0048), followed by MethylNet (Silhouette Score: 0.25±0.01), and MethylSPWNet (Silhouette Score: 0.1±0.0087), estimated using a 1000-sample non-parametric bootstrap. Since these three approaches were trained to recognize histological subtype, the signal of the cell of cancer origin and molecular subtypes were less well captured.

Gene-Level and Modularity Enrichment Analyses

Next, we aimed to evaluate the utility of the group-regularized deep learning approach for capsule-organized summaries of DNAm on the gene-level for pathways and gene network analyses. We performed a preliminary analysis of pathway and module detection based on the extraction of hypervariable genes across the neural network embeddings, and Louvain clustering of networks of genes based on the pairwise correlation between the genes. Further description of the methods and results is provided in the supplementary material (Supplementary Table 2; Supplementary Figure 3-4).

A description and flow-chart showing an overview of methods for pathways and gene network analysis downstream from *MethylSPWNet* can be found in the Methods section "Description of Potential Downstream Analyses". We focused our presentation of results on three specific CNS tumor subtypes: glioblastoma (GBM), low-grade glioma (LGG), and medulloblastoma (MB). Gene-level embeddings (gene by sample), and derived from those embeddings, pathways and gene network analysis results are shown in Figures 1, 2 and 3 respectively. Gene-level

embeddings, as well as pathways and gene network analysis results for all 38 CNS tumor

subtypes, are provided in the supplementary materials and additional files.

Figure 1: Clustermap of gene-level embeddings for 2000 most variable genes in a) GBM; b) LGG; c)
MB. Presence of the subtype is indicated by the left color track; yellow indicates the presence of controls;
green, purple, and pink indicates the presence of the GBM, LGG, and MB subtypes respectively; columns
have been standardized to highlight trends; in the heatmap, red indicates high *MethylSPWNet* embedding
values, while blue indicates low embedding values

Subtype-Specific Pathways Discovery

Using the gene-level embeddings acquired from fitting *MethylSPWNet*, we sought to calculate

genes that were differentially embedded (as defined by differences in their gene-level embedding

values) between disease and controls (empirical Bayes) and determine the associations of these

genes with some of their correspondent pathways (see Methods "Description of Potential

Downstream Analyses"). A visualization of pathway networks and output of gene-subtype

associations and pathway enrichment statistics are provided in Figure 2. The results of empirical

Bayes outputs of the differential embedding analyses for each subtype are provided as Additional

Files.

From our analysis of the selected three CNS tumor subtypes (GBM, LGG, and MB), we found many top differential genes have been implicated for these subtypes in prior literature. For instance, *TACC* and *FGFBP2* (Figure 2b) highly differentiated GBM from normal and have been previously implicated with a tumorigenic gene fusion event (*TACC-FGFR3* [58–60]); *RASGRF2* (Figure 2b) has been linked to congenital GBM[61], *LGI2* and *NPY5R* (Figure 2b) have both been related to changes in Sox2 expression[62], which promotes cellular plasticity in GBM. An interesting associated pathway for GBM was Type I diabetes mellitus (Figure 2a-b), which is an autoimmune disease, of which its spurious association could be related to found associations with several immune evading markers. For LGG, *TLK1* (Figure 2d), a serine-threonine kinase associated with replication, focal adhesion, and cell cycle, has previously been implicated in gliomas[63]; as have *ELL2* (Figure 2d), which was known to be regulated by microRNA (miRNA)-mediated gene silencing, and low *ELL2* expression was reported to be a marker for poorer survival in GBM patients[64]. In addition, *GNL1* (Figure 2d) was found in our analysis to be associated with LGG and has been identified as being related to cell proliferation, given its role in the phosphorylation of Rb[65]. We also found associations with the Opioid-signaling pathway and G alpha (i) signaling events [66], and the tyrosine kinase receptor pathway VEGFR (Vascular Endothelial Growth Factor Receptor) and downstream signaling pathway ERK (Figure 2c-d), largely involved with proliferation and angiogenesis[67]. Regarding MB, as examples, we uncovered *NRBP2* (Nuclear Receptor Binding Protein 2; Figure 2f), which had been shown to be downregulated in MB[68], and *SOX14* (Figure 2f), part of the *SOX* family which largely determines cell fate and thus heavily implicated across many CNS tumors[69]. Additionally, pathways such as Muscle Contraction (Figure 2e-f) have been associated with specific molecular subtypes of MB[14,70].

A

transmembrane transporter transport

synaptic trans signaling

distal axon growth

transmitter gated channel

regulation organismal process

part morphogenesis differentiation

collagen trimer matrix

phosphotransferase alcohol binding

extracellular space exosome

vesicle secretory cytoplasmic

B

Differential Embedding

gene	logFC	AveExpr	t	adj.P.Val	B
TACC3	-1.71	0.394	-23.0	1.30e-47	107.0
LGI2	-1.76	0.368	-20.6	3.32e-42	93.6
RASGRF2	-1.94	0.464	-20.0	5.56e-41	90.4
FGFBP2	-1.62	0.374	-19.8	1.44e-40	89.2
NPY5R	-1.81	0.548	-19.4	7.06e-40	87.3

Pathway Enrichment

Pathway	Description	FDR
REAC:R-HSA-112316	Neuronal System	0.00037
REAC:R-HSA-112315	Transmission across Chemical Synapses	0.00980
KEGG:05033	Nicotine addiction	0.00078
KEGG:04080	Neuroactive ligand-receptor interaction	0.00100
KEGG:04940	Type I diabetes mellitus	0.02200

C

opioid signalling events

cell junction anchoring

postsynaptic transmission

projection boundary body

extracellular space endomembrane

vegfr2 erk1 2

ion binding anion

component plasma membrane

embryonic organ embryo

organismal process multicellular

synaptic trans signaling

response regulation communication

neuron development morphogenesis

regulation biological quality

biological adhesion cell

adhesion molecules homophilic

secretion regulation transport

movement cell subcellular

inorganic transporter activity

actin filament process

junction organization assembly

D

Differential Embedding

gene	logFC	AveExpr	t	adj.P.Val	B
TLK1	1.180	-0.0191	12.5	2.63e-18	38.9
LIMS1	0.991	-0.3830	11.4	2.92e-16	33.5
ELL2	1.340	-0.9720	11.1	9.77e-16	31.9
TSR2	1.160	0.1090	10.9	2.03e-15	30.9
GNL1	1.220	-0.0613	10.7	5.22e-15	29.7

Pathway Enrichment

Pathway	Description	FDR
REAC:R-HSA-112314	Neurotransmitter receptors and postsynaptic signal transmission	3.5e-05
REAC:R-HSA-112316	Neuronal System	1.2e-04
REAC:R-HSA-112315	Transmission across Chemical Synapses	2.5e-03
REAC:R-HSA-111885	Opioid Signalling	1.2e-02
REAC:R-HSA-418594	G alpha (i) signalling events	2.0e-02

E

transmembrane ion activity

muscle contraction heart

cell body projection

hormone levels secretion

regulation localization transport

metabolic process phosphorus

establishment localization transport

component plasma membrane

signaling synaptic trans

cellular response external

intracellular vesicle cytoplasmi

regulatory binding dna

anatomical structure developmental

cell morphogenesis neuron

epithelial tube morphogenesis

F

Differential Embedding

gene	logFC	AveExpr	t	adj.P.Val	B
VPS13B	1.99	-0.289	26.4	1.05e-48	109.0
NRBP2	2.55	-1.200	26.3	1.05e-48	109.0
PROP1	2.03	-0.585	25.8	4.32e-48	107.0
SOX14	2.64	-1.510	24.1	4.03e-45	100.0
TFCP2	2.21	-1.350	23.9	8.13e-45	99.1

Pathway Enrichment

Pathway	Description	FDR
REAC:R-HSA-112316	Neuronal System	9.4e-08
REAC:R-HSA-397014	Muscle contraction	4.4e-05
REAC:R-HSA-5576891	Cardiac conduction	2.4e-04
REAC:R-HSA-112315	Transmission across Chemical Synapses	1.3e-03
REAC:R-HSA-1296071	Potassium Channels	1.1e-02

Figure 2: Example Output from Pathway Enrichment Analyses for a-b) GBM-Control embeddings; c-d) LGG-Control embeddings; e-f) MB-Control embeddings; left-side plots: corresponds to summaries of GO/KEGG/Reactome enrichments using EnrichmentMap in Cytoscape, plotted via RCy3; each small node corresponds to a pathway; node and text size proportional to the number of overlapping genes after differential gene-level embedding analysis; edge between nodes corresponds to shared genes; large ellipses correspond to found clusters using Markov Chain clustering, three words extracted to annotate cluster using a word cloud algorithm; right-side plots: example output of differential embedding (limma) and pathway enrichment analyses (g:Profiler) on embeddings; top 5 genes and select pathways listed of the many uncovered in each analysis.

Derivation of Weighted Gene Co-Methylation Networks

To investigate the gene-level embeddings, we derived disease-specific modules of genes using

Weighted Gene Correlation Network Analysis (WGCNA) [48] for each of the 38 brain cancer

subtypes (each paired to normal controls) using the WGCNA R package (see Methods

"Description of Potential Downstream Analyses").

We derived 606 modules of genes across the 38 subtypes (37 networks were derived, one

subtype was omitted for low sample count), 297 of which were significantly associated with

subtype (all P-values < 0.05). We have included as Additional Files the module membership of

each of the genes, module expression across the samples for each subtype, hub genes for each

module (genes that are located most centrally in each subnetwork), and statistics that relate each

module to the subtype. Connectivity of individual genes from the generated WGCNA modules

for GBM, LGG, and MB subtypes are shown in Figure 3. Tables of top hub genes from selected

modules strongly associated with each subtype are shown.

Some of the hub genes of the WGCNA modules were found to be correspondent with prior

knowledge about their respective subtypes. For instance, in GBM, *RASGRF4* (blue module;

Figure 3a-b) had been previously described, silencing of *MGMT* (green module; Figure 3a-b)

plays a significant role in the progression of GBM through inactivation of its DNA repair

mechanisms[9]; *MIR33B* (brown module; Figure 3a-b) is also related to GBM progression by

regulating cell proliferation, invasion, migration, and MYC signaling[71,72]; finally, the role of

platelet factors (*PF4*; blue module) and CpG island hypermethylation (*BARHL2*; turquoise

module) is not inadmissible for GBM[73,74]. Examples of hub genes in LGG include *NRP1*

(black module; Figure 3d; which has been shown to be related to poor prognosis in gliomas and signals through microglia/macrophages)[75], *PTPRZ1* (pink module; Figure 3d; promotes malignant growth in GBM)[76], and *COL6A3* (green module; Figure 3d; member of genes serving to form the tumor vasculature). Finally, for MB, *SOX 14* and *SOX17* were found in the green module (Figure 3f; cell fate)[69], while *CD4*, the gene that codes for the membrane glycoprotein of the CD4 T-cell, was found in the green-yellow module (Figure 3f; corroborated by the immune infiltration patterns of the stroma for MB), and *SLIT3*, a gene characterized by axon guidance and consequently tumor growth, migration, and angiogenesis, was found to be central to the black module (Figure 3f). Interestingly, *GFPT2* (amino acid metabolism, implicated with higher expression and lower GBM survival [77]) and *SYNPO*, central to the black module of MB, were also found to be central to the blue module of GBM.

A

B

GBM Significant Module Hub Genes

Description	blue	green	brown	turquoise
p-value	9.5e-37	6.5e-18	2.3e-11	3.5e-12
Correlation with GBM	0.81	0.62	-0.5	-0.52
Hub Gene 1	GFPT2	TNFRSF6B	MIR33B	LYRM4
Hub Gene 2	SYNPO	C3orf20	TSR3	PABPC3
Hub Gene 3	ENPEP	TSLP	LOC285796	LOC402778
Hub Gene 4	PF4	MYOM2	SERF2	BARHL2
Hub Gene 5	LOC285501	LOC220930	ADPRHL1	FAM71F1
Hub Gene 6	LRRC14B	PLA2G2D	ADCK2	DNAH1
Hub Gene 7	TRAM1L1	CDH23	NCS1	GALR2
Hub Gene 8	RASGRF2	MGMT	IL31	KLK6

C

D

LGG Significant Module Hub Genes

Description	pink	green	blue	black
p-value	1.7e-09	1.4e-06	4.8e-08	9.7e-12
Correlation with LGG	0.55	0.45	-0.51	-0.61
Hub Gene 1	PTPRZ1	COL6A3	PABPC3	NRP1
Hub Gene 2	MLST8	FLJ36000	PC	TMPRSS4
Hub Gene 3	MAP10	ATP6V0A4	PALMD	IGDCC4
Hub Gene 4	FCGR2A	METTL9	CTBP1	WFIKKN2
Hub Gene 5	C17orf99	C10orf71	SLC25A18	COL17A1
Hub Gene 6	FOXI2	GABBR2	GFRA4	ADPRHL1
Hub Gene 7	DNAJB8	CTSF	MIR486-1	MELTF
Hub Gene 8	SULT1A2	ERBB4	ASXL2	MIR134

E

F

MB Significant Module Hub Genes

Description	black	midnightblue	greenyellow	green
p-value	7.5e-47	1.9e-23	6.1e-29	2.9e-48
Correlation with MB	0.9	0.75	-0.8	-0.91
Hub Gene 1	GFPT2	GGACT	CD4	SOX17
Hub Gene 2	SLIT3	FOLR1	ST6GALNAC5	R3HDM2
Hub Gene 3	PDCD6	SCGB1D4	HSPB8	LHX1
Hub Gene 4	PCDHGA9	ADGRF2	RASSF9	AIFM2
Hub Gene 5	ENPEP	HEATR4	LIMS2	PPP4R3C
Hub Gene 6	ZAR1	COL7A1	MYPN	MIR196A2
Hub Gene 7	SYNPO	CPED1	LOC399815	SOX14
Hub Gene 8	C5orf52	CHD8	PANX2	ABCA2

Figure 3: Example Output from WGCNA Analyses for a-b) GBM-Control embeddings; lack of connectivity between the blue/black modules and the other models suggests lack of functional relationship; c-d) LGG-Control embeddings; e-f) MB-Control embeddings; left-side plots: summarized gene-gene networks using the MAPPER algorithm; nodes indicate genes or dense grouping of genes; colored by discovered module via WGCNA; edges indicate co-methylation; right-side plots: example output from WGCNA analysis; the subset of gene-gene modules; p-value indicates whether significantly associated with tumor subtype; correlation value indicates directionality and strength of this relationship; select hub genes indicate genes with the highest centrality in each subnetwork/module.

Enrichment of Neural Network CpGs for Gene and Island Contexts

As Weighted Gene Correlation Network Analysis (WGCNA) identifies significant associations with known pathways and novel gene-gene co-methylation networks, we sought to investigate the CpG-specific parameters that corresponded to producing the embeddings to understand better why the neural network decided to upweight some CpGs, but not others. To better elucidate the genomic contexts that *MethylSPWNet* found to be important, we explored the CpG island context and spatial relationship to the transcriptional start site (TSS) (Methods section "CpG Island/Gene Context Analysis"). CpG islands (CGI) are CpG dense regions; approximately 60% of gene promoters contain CpG islands[78]. CpG shores immediately flank the CGIs by up to 2kb, shelves flank the shores by an additional 2kb, as regional CpG density decreases. Variables for the spatial relationship to the TSS include TSS1500 and TSS200, within 1500 bp and 200 bp of the TSS, respectively. Additional TSS variables are the 5'UTR immediately downstream of the TSS, the first exon, gene body, and 3'UTR.

Of note, we found that CpGs with positive weights (rank-ordered) were depleted for promoter island regions (defined as having TSS1500/TSS200 annotation and not open sea) (OR=0.69; p=0.04) as compared to sites not included in promoter-island regions (OR=1.45; p=0.04). However, when limiting the set of CpGs to only promoter regions (i.e. TSS1500/TSS200), we noted that positive weight CpGs were enriched for island context (i.e. not in an open sea region) (OR=1.21; p=0.03), while negative weight CpGs were depleted for the CGI context (OR=0.89; p=0.02). Furthermore, both the positive and negative weights of intragenic CpGs were depleted for association with the correspondent methylated promoters (as compared to unmethylated promoters) for their respective genes (positive weight OR=0.54, p<0.01; negative weight

OR=0.44, p<0.01). We have included the set of tables for the relationship between CpG weight and independently considered contexts in the Supplementary Materials (Supplementary Table 3; Supplementary Figures 4).

MethylCapsNet Module Enrichment

From the embedding module discovery analysis and further contextualization of the neural network CpG-weights, we observed that information encoded in *MethylSPWNet* corresponds to key pathways associated with various CNS tumors and important genomic contexts. *MethylCapsNet* offers the ability to infer more granular relationships between capsules on the individual sample level when we are able to reduce the number of parameters specified. The primary capability and emphasis of the capsule network approach is to compare capsules to each other as well as directly relate them to particular outcomes of interest via the dynamic construction of a bipartite network (gene-subtype relationships) as part of the training and prediction process. For the CapsNet analysis, we pre-selected a subset of genes previously shown to be implicated in various types of brain cancer (see "Selection of Capsules for *MethylCapsNet* and *MethylSPWNet*" in Methods). As such, we believe it would not be appropriate to test for enrichment of these genes due to the pre-selection procedure that introduces a bias. Instead, we derived modules of genes that the neural network deemed to have a coordinated DNAm response in elucidating particular subtypes. Our modularity analysis projects the estimated bipartite graph (gene-subtype) across samples into a univariate graph (gene-gene), then clusters the graph using Louvain modularity to yield four modules of genes (green, red, blue, yellow) (Figure 7).

Here, we offered an example of the kinds of inferences that can be made from the resultant unipartite network and subsequent clustering. For instance, the yellow module implicates relationships between *WNT3A* and *EGFR*, heavily implicated in IGF and WNT signaling. The red module features the relationship between *FRZ* and *APC*, both of which are heavily involved in WNT signaling (*APC* forms the complex to inhibit the accumulation of β-*catenin*, while the binding of WNT to frizzled family receptors may degrade this inhibition and permit cell proliferation[79]). Of the green module, *IDH3G* and *NPR3* (linked to energy metabolism, gene fusion, and chromatin remodeling) were related to both *LRDD* (pro-apoptotic *MAPK* pathway) and *WIF1* (both previously implicated WNT signaling suppressors) from the yellow module. *KIAA1549*, related to astrocytomas and fused to *BRAF* for its progression to oncogenesis, was implicated with *WNT1* in the blue module[80]. Insulin-like Growth Factor Binding Protein 2 (*IGFB2*; glioma oncogene) of the red module appeared to be negatively correlated with many of the genes across the yellow and green modules[81]. *TP53* (which lacked consequential methylation patterns) and *MYC* [82], *MGMT,* and *TERT* share relationships with each other, but not with the other modules, perhaps highlighting how ubiquitous these somatic alterations are for oncogenesis.

Figure 7: Clustermap of weighted bipartite network projection of select gene capsules' relationship to various CNS tumors using the *MethylCapsNet* modeling approach; rows and columns are genes, row colors denote modules (green, red, blue, yellow) of genes found using Louvain modularity; values in matrix denote Pearson's correlation between genes

Despite having highlighted potentially interesting relationships, we acknowledge that there will

be future opportunity to increase the search space of possible relationships between genes and

their coordinated response for bringing about subtypes of brain cancer. In the Supplementary

Material, we provide the routing matrix that was used to align each gene to a correspondent CNS subtype, which is the predicted gene-subtype bipartite network averaged across individuals (Supplementary Figure 5). In addition to this, we have provided a locally-deployable web application that allows the user to interrogate their uncovered capsules and form networks on the individual-level and aggregated across patient subgroups or disease subtypes using gene-capsule specific embeddings (Supplementary Figure 6). For instance, in our supplemental material, we demonstrate how the *MethylCaps* web application can be used to derive individual networks for LGG and GBM, identifying *ANO9* and *EGFR,* respectively, as implicated in these conditions (Supplementary Figure 7-8). These genes and their associated pathways have been heavily implicated in tumorigenesis and gliomas [83,84].

Discussion

Recent reviews and initial explorations discussed the potential utility of capsule networks to relate biologically-organized capsules to each other and known disease outcomes. [27,85–87] In this work, we set out to perform a preliminary evaluation that shows the feasibility and suitability of DNA methylation capsules for deep learning analyses as a means to organize CpG information to higher-order contexts to improve prediction and transparency while uncovering instances of coordinated gene-level methylation patterns. In our analyses, we compared several state-of-the-art predictive modeling methods for DNA methylation classification of brain tumors. We demonstrated that capsule-based deep learning approaches could achieve performance on par with existing deep learning models and proving better than existing traditional statistical frameworks for analyzing DNA methylation data. Our work demonstrated new insights when compared to other existing methylation-based tumor classification schemes currently used,

which are often based on a small subset of CpGs, and lack interpretation of the loci selected[8,88]. We demonstrated the efficacy of increasing the use of the available CpGs on the Illumina 450k Array, ultimately using 200,000 loci before sub-setting by context.

DNA methylation capsules focused at the gene level can disentangle important CpGs that might otherwise be down-weighted in a feature-by-feature deep learning unsupervised or supervised learning approach. These CpGs demonstrated substantial overlap with genes known to be related to tumorigenesis in the brain, such as *NOTCH1*, *PTEN*, and *GNAS*. This is consistent with previous studies that demonstrated mutations common in brain tumors, such as *IDH1*, are correlated with disruptions in methylation.[10,89–94]

The context-specific CpG weight enrichment analyses suggest that within promoter regions, island context is important for differentiating different CNS tumor subtypes, but taken as a whole, regions outside of CpG promoter islands are important for capturing this heterogeneity. Furthermore, outside of the promoter context (supposedly regions that better capture tumor heterogeneity), the ability of intragenic CpGs to distinguish tumor subtypes is still dependent on the promoter methylation status of the respective gene. Clustering of CpGs with the highest weights at CpG islands, shores, gene bodies, and transcription start sites will help us understand where the most diagnostically relevant sites are in the genome, but demands additional investigation.

We also presented a few examples of the potential downstream applications of capsule-based approaches. In particular, our framework demonstrated the ability to relate derived gene-level

measures of *MethylSPWNet* to known disease pathways via differential methylation analysis of the gene-level embeddings and gene-gene co-methylation networks via WGCNA. Finally, we provided a preliminary interpretation of bipartite (gene-subtype) and unipartite (gene-gene) networks, which can be derived by *MethylCapsNet* web framework.

There are a few limitations to this work, presenting room for future improvements in the analytical method. First, the selection of included CpG loci is biased on limited sites available from the Illumina Array platform. At present, the utilization of an Illumina methylation array platform is more tractable due to lower technical costs and expertise required[95] than Whole Genome Bisulfite Sequencing (WGBS) that requires substantial sequencing depth on the order of 100x for comparable precision[96].

Second, a reduced set of genes were fit using capsule network. In its current formulation, it remains challenging to run *MethylCapsNet* at scale due to the heavy computational demand and the large number of free parameters. Since *MethylCapsNet* can only analyze approximately one thousand capsules at a time, the capsule selection step is critical to the successful application of the method. This parameter space should be reduced by finding some marriage between the scalability enabled by MethylSPWNet and perhaps greater transparency offered by MethylCapsNet. Presently, we advise end users to utilize *MethylSPWNet* when the number of contexts under evaluation is large (i.e. on the order of at least one thousand capsules), or if the number of CpGs per gene is small, and to utilize *MethylCapsNet* when the number of contexts under consideration is smaller (i.e. under the order of one thousand capsules). For instance, relating all genes without manual curation of gene sets can be done using *MethylSPWNet*, while

relating curated gene sets, perhaps regions of the genome fragmented by consistent windows, or larger DNAm CpG modules that have been uncovered through methods such as WGCNA. In addition, the adoption of capsule-inspired approaches that explicitly form networks via their routing mechanisms presents a future area of research [86]. It is also assumed that *MethylCapsNet* capsules that are more closely embedded are interacting, but it is not entirely clear what is the nature of these interactions without incorporating gene expression data and mQTL analyses (methylated Quantitative Trait Loci). Here, there is an opportunity to further apply concepts from topological data analysis (TDA), such as Mapper[97–101], to distill the key functional relationships from high dimensional, complex data.

Existing classification frameworks currently used in the clinical setting for aiding brain cancer diagnosis only utilize a small subset of the total possible set of CpGs that can be measured. Current modeling approaches can be difficult to trust or use to study new network biology until they can take into account a larger, more complete set of predictors. However, it is also important to note that doing so would introduce additional noise into the modeling approach, but incorporation of prior biological knowledge can help reduce noise while improving the detection of biologically relevant signals. By demonstrating that the organization of CpGs into their respective genomic contexts, we present further opportunity to reduce the feature space and disentangle correlation and collinearity between CpG sites for the creation of a new class of transparent, clinically tractable models. Despite having demonstrated the promising downstream analyses which users may readily adopt through our framework, we acknowledge that there is ample opportunity for further development of related methods and their use cases.

Conclusions

Here, we have demonstrated the feasibility and utility of DNAm based capsules for performing disease classification and potentially determining dysregulated genes for these diseases. We found that DNA methylation capsule methods are able to predict brain cancer subtypes with high accuracy and present convenient means for organizing data over traditional techniques for studying DNA methylation data. As such, we advocate for the organization of well-defined DNAm capsules as a means to improve the accuracy, transparency, and broad applicability of DNAm deep learning models. Future deep learning prognostic models that reimagine the formation and incorporation of DNA methylation capsules, paired with cell type inference, gene expression, and/or corroborating chromatin capture, may serve as grounds for the derivation of unknown heterogeneity.

Methods

Overview of Framework

The *MethylCapsNet* methodology presents an extension of the *MethylNet* framework [20] and is implemented as a command-line interface that allows the user to group CpGs into capsules, and then dynamically route the capsules to make a prediction and interpret the results.

MethylCapsNet utilizes separate MLPs for every set of CpGs (one set per context) to derive context-specific embeddings (separate context embeddings per each individual), and dynamic routing processes force information from child capsules into disease/categorical outcomes (Figure 4a). The information is hierarchical because each child capsule may only align with one parent capsule. Once the capsule network is fit, graph structures that describe the relationships

between the contexts for each individual can be derived by thresholding the correlation between

pairwise n-dimensional context embeddings. Highly conserved biological networks can be

derived by thresholding the number of individuals that share the same edge between the contexts.

The simplest genomic context considered are genes that the CpGs annotate to; other capsules can

be defined by, for instance, genomic region, or pathway / biological process annotation.

MethylSPWNet is a specialized neural network architecture that routes beta values from the

CpGs in each context into a single node representing the context (Figure 4b); each CpG is given

a weight based on the importance of its contribution, both on the gene level and towards the

classification task as a whole. This information passes through additional neural network layers

that dynamically relate latent sets of predictors to outcomes of interest, whether they be

prognostic or diagnostic in nature [53,102]. Much like Group LASSO approaches (Figure 4c),

group L1-penalization can be utilized on the CpG weights routed to each gene to select relevant

genes of interest.

Figure 4: Description of Modeling Approaches: a) *MethylCapsNet* separate MLPs are utilized to form
capsule level embeddings from beta values of CpG groupings, which are then associated with outcome
targets of interest through dynamic routing of information; these embeddings may be studied to propose
networks based on co-methylation on the individual level; b) *MethylCapsNet* aggregates the beta values
of groups of CpGs through one locally connected layer; nodes of the resulting layer represent biologically
meaningful units that are passed through an MLP for final prediction; group L1 penalties are applied to

prune genes/capsules unrelated to the outcome; c) Group LASSO models are of the generalized linear model specification and penalize sets of parameters corresponding to each capsule/gene

Figure 5: Description of Framework: a) User selects capsules to group sets of CpGs from their own supplied list or a prespecified annotation set; b) User selects modeling approach, *MethylCapsNet*, *MethylSPWNet*, *Group LASSO*; c) Hyperparameter search conducted to reveal ideal model specification; d) Final models are fit, and capsules are interrogated for relationships with one another, relationships to the outcome, and highly weighted CpGs; red indicates an important capsule; *MethylCapsNet* relates the Capsule to each other per individual; *MethylSPWNet* locates important CpGs within a capsule and contextualizes its location

The software implementation (Figure 5) is comprised of modules pertaining to prediction and interpretation tasks, which take into account the relationships and embeddings derived through the training process.

Data Preparation

DNAm data from CNS tumors (n=3,897) were accessed from the GEO archive (GSE109381), preprocessed using *PyMethylProcess* [21], and divided into 70%/10%/20% training, validation, and testing sets (*MethylationArray* objects) via *PyMethylProcess*. The 200,000 most variable CpG loci across the training samples were retained for analysis. Sets of CpGs tracked to genes, which were then selected to form capsules. The original set of 200,000 CpGs was used as features for the *MethylNet* approach, the complete set of intersecting gene capsules with more than five associated CpGs were used for the *MethylSPWNet* (n=10,341; 139,028 CpGs), and Group LASSO approaches, and a reduced set of capsules (n=55), were utilized for *MethylCapsNet* after manual curation and a hyperparameter search (see "Selection of Capsules for *MethylCapsNet* and *MethylSPWNet*").

Description of Potential Downstream Analyses

After fitting a *MethylSPWNet* model (and *MethylCapsNet*), the user may further interrogate the gene-level embeddings depending on the research question being addressed. The user may explore how each gene relates to each outcome or how they relate to one another, the details of which have been included in an informative flow diagram (Figure 6) (separate text describing information, that can be extracted after fitting *MethylCapsNet* can be found in the section "Description of Capsule Network").

Differentially methylated genes from gene-level values derived by the neural network embeddings in each of GBM, LGG, and MB, were identified with *limma* [103]comparing to nontumor control tissue. Least-squares regression and empirical Bayes moderated F-test of the fit model produced FDR-adjusted p-values and log-odds ratios for the degree of differential embedding (the extent to which gene-level embeddings vary between subtypes). We profiled functionally enriched pathways using the g:Profiler package [104] after selection of genes below an FDR-adjusted significance threshold and visualized the results (relating pathways by the number of shared genes, clustering into higher-order pathways via Markov clustering) using EnrichmentMap, as part of the Cytoscape network visualization framework [105,106].

The pairwise correlation between *SPWNet*-derived gene methylation was calculated using Pearson's correlation coefficient. Weighted adjacency matrices were calculated from the pairwise correlation matrices for each of the subtypes using the power adjacency function, which takes the co-methylation to a power specified separately for each subtype. To further cluster the genes, the weighted adjacency is transformed into a topological overlap matrix (*TOM*), which is defined by the extent to which two genes share a third common gene. Finally, hierarchical clustering is applied to derive the final modules of genes. Finally, to relate each module with the disease subtype via the aforementioned least squares and empirical Bayes differential analyses methods, we calculated eigengenes (1st principal component) for the genes in each module to further reduce the design matrix (samples by modules) [103]. A large number of genes (on the order of five thousand) for such a summary gene network plot may make plotting the individual genes cumbersome and hard to understand, so we utilized Mapper[97,99,100], a tool from

Topological Data Analysis, to further summarize and portray the relationships between the genes in the network summary plot.

Figure 6: Flow Diagram for Possible Downstream Applications of *MethylSPWNet*: a) User fits *MethylSPWNet* model to predict brain cancer subtypes; b) gene embedding matrix (samples by genes; in this example GBM vs Control) is extracted from the model from the test samples; user decides whether

they want to **(c-e)** associate gene-level embeddings with disease outcome, or **(f-h)** relate genes to one another to imply functional relationships; c) user opts for pathways analysis; *limma* is used to identify genes with differential embeddings between GBM and controls using an empirical bayes moderated linear model; d) significant genes may be passed into g:Profiler or *limma*'s internal functions *camera*, *goana*, and *kegga*, for pathways enrichment over GO, KEGG, Reactome, etc. databases; e) pathways are summarized and visualized using EnrichmentMap, accessed via RCy3; f) user opts for gene correlation analysis (WGCNA), first calculating correlation between genes, then calculating topological overlap over correlation transformed by power, hierarchical clustering is applied to deduce modules; g) gene-gene networks and module membership is visualized using Mapper; h) modules related to GBM are extracted using *limma* by comparing projections into first eigengene (samples by modules matrix) for each module for GBM vs controls

CpG Island/Gene Context Analysis

Using the MethylSPWNet, each CpG is assigned a weight that relates the CpG to its associated

gene or genomic context. These CpG weights are learned by the neural network and can be used

to rank genes based on their relative importance (rank assigned by maximum absolute CpG

weight), an alternative measurement to the modularity analysis. Inspection of the weights of

CpGs within each gene can provide insight as to sites and contexts that are important for the

prediction of brain cancer subtypes. Further, investigation of weights spatially across the genome

may give rise to important patterns and motifs that could warrant future investigation. In the

supplementary material, we first considered the aforementioned contexts independently and did

not consider the joint impact of context (e.g., did not associate with island-promoter regions,

which are generally considered to be regions more causally related to changes in their

expression). We then considered sites that were associated with island promoter regions

(including shore and shelf context; more causally associated with gene expression) and

separately compared the overlap of the CpGs correspondent to top positive and negative weights

to the CpGs that were unassociated with this context (open sea and not TSS200/1500). We

separately considered CpGs within the promoter regions. Finally, we considered intragenic CpGs

and whether or not their corresponding gene's promoter was methylated or unmethylated (as

operationalized by calculating a beta-value methylation cutoff via local minima in the distribution of beta-values; any CpG with a beta value above the threshold was methylated; CpG methylation was averaged across each promoter and subject to the threshold to determine methylation status). We calculated odds-ratios for enrichment/depletion in these contexts using Fischer's exact tests. Without matched gene expression information, we were unable to make any causal claims/inferences about how these contexts modify gene expression to bring about these disease states.

Description of Capsule Network

The capsule network featured in this work operates by first finding representations of the given CpG sets as denoted by the primary capsule formation. The features, CpGs, of the CpG sets are fed into parallel implementations of a multi-layer perceptron, f_j, where the output dimensions of each of the neural networks are the same. Thus, the dimensionality of the primary capsules reflects the number of output neurons, which is a latent representation of each CpG set, times the number of capsules, per individual. The mathematical formulation of this transformation is presented below:

$$\overrightarrow{z_{gene_j}} = f_j\left(\overrightarrow{x_j}\right)$$

For a single individual, the capsules, represented by row vectors, are stacked to form a capsule matrix:

$$\overleftrightarrow{Z} = \begin{bmatrix} \overrightarrow{z_{gene_1}} \\ \vdots \\ \overrightarrow{z_{gene_n}} \end{bmatrix}$$

The primary capsules are transformed by an affine transformation, which is a set of learnable parameters that seek to rotate, scale, and shift the data, and may encode information pertaining to the interactions between capsules.

$$\overrightarrow{Z_*} = \overleftrightarrow{W}\, \overrightarrow{Z}$$

Each primary child capsule's information is then dynamically routed to parent hidden or output capsules.

$$\overrightarrow{Y} = \sigma(\overleftrightarrow{C}\overrightarrow{Z_*}) = sq\left(\begin{bmatrix} \overrightarrow{y_{class\ 1}} \\ \vdots \\ \overrightarrow{y_{class\ m}} \end{bmatrix}\right)$$

Where:

$$\overrightarrow{y_{class\ j}} = sq\left(\sum_i C_{gene i, class j}\, \overrightarrow{z_{gene\ \iota}}\right)$$

The aim of dynamic routing is to force the information encoded into each child to align with one parent capsule, thus utilized to calculate $\overleftrightarrow{C} = \{C_{ij}\}$, a bipartite network relating the child-parent-capsule. Each child is represented by a vector of the same length as the output of the parent capsule. Analogous to the non-linear transformation of the sum of the information output from the previous layer of neurons for traditional neural networks, for each parent capsule, the child-capsule values are summed, and then a non-linear transform called a squash function, $sq(\vec{x}) = \frac{\|\vec{x}\|^2}{1+\|\vec{x}\|^2}\frac{\vec{x}}{\|\vec{x}\|}$, is applied to effectively zero-out, or *squash*, child capsules that do not agree with parent capsules.

The contributions of each child to a parent are weighted, but two constraints are imposed: First, the weights from each child to its parents must sum to 1. Second, a reward for the alignment of a

child to exactly one parent, a dynamic routing by agreement mechanism, where an iterative

process updates the weights between the child and parent by adding their dot product. The update

mechanism for calculating $\vec{C}$ is recapitulated below; after initializing $\vec{C_i} = softmax(\vec{\beta_i} = \vec{0})$ for

$r \in \{1, 2, 3, ...\}$ iterations:

$$\vec{C_i} = softmax(\vec{\beta_i})$$

$$\vec{Y_j} = sq\left(\sum_i C_{genei,classj} \overrightarrow{z_{gene\,i}}\right)$$

$$\beta_{ij} = \beta_{ij} + \overrightarrow{Y_{classj}} \cdot \overrightarrow{z_{gene\,i}}$$

This formula is simplified from its original derivation and utilizes a few notational shortcuts.

Applying this operation for r iterations per batch per training epoch effectively prunes the other

connections between the child and its parents as it converges on a single parent from which to

send its information. Each output capsule per individual, $\vec{Y_j}$, is represented by a vector in some n-

dimensional space. The output capsule with the highest L2-norm, $\left\|\vec{Y_j}\right\|_2$, is selected as the

predicted class, and a margin loss is applied to penalize the model when it fails to either

concretely have a very high ($m^+ = 0.9$) or very low probability ($m^- = 0.1$) of prediction.

$$L_{margin:\ classj} = \delta_{y_i,j}\, max\left(0, m^+ - \left\|\vec{Y_j}\right\|\right)^2 + \lambda\left(1 - \delta_{y_i,j}\right) max\left(0, \left\|\vec{Y_j}\right\| - m^-\right)^2$$

The Kronecker's delta $\delta_{y_i,j}$ is equal to one when the outcome for individual i is equal to the j-th

class, thereby activating the left-hand margin loss that penalizes the model if the probability is

below $m^+ = 0.9$. On the right-hand of the equation, $\delta_{y_i,j}$ is equal to zero when the outcome for

individual i is not equal to the j-th class, thereby activating the right-hand margin loss that

penalizes the model if the probability is above $m^- = 0.1$.

The model is also penalized based on how much the original methylation array could be constructed from the true class's output capsule via a decoder neural network, $\widehat{X} \sim p_\phi(\vec{X}|\overrightarrow{Y_J})$:

$$L_{reconstruct} \propto \left(\widehat{X} - \vec{X}\right)^2$$

Of the most interest to a biologist may be the primary capsule embeddings per individual, $\overrightarrow{Z_*}$, which demonstrate interactions between these biological hypotheses and how the outcome of interest is separable within certain genomic context, and the weights between the primary and output capsules, $\overleftrightarrow{C}$, a bipartite graph, which demonstrates how these genomic regions are related hierarchically and have implications for parent processes. The coordinated response of capsules can also be derived through a bipartite projection of $\overleftrightarrow{C}$ into a unipartite network of capsules. Secondarily, of importance are the concatenation of the primary capsules, which demonstrate overall class separation, and the decoded output. Tweaking the embeddings or L2 norm of the output capsules and decoding can potentially effectively generate methylation data conditionally on outcomes of interest and interpolate between purified states, though this aspect was unexplored due to prohibitive dimensionality.

Description of MethylSPWNet

MethylSPWNet is the deep learning analog of a Group LASSO Regression model. The beta values for the CpGs for each gene, $\overrightarrow{x_j}$, is transformed into a single value, z_{genej}, through the multiplication of a set of gene-specific CpG weight matrices, $\overrightarrow{w_j}$. These weights are updated throughout the training process to minimize the divergence between observed and expected

outcomes; the magnitude of the weights dictates how much information from each CpG should be considered. The final gene-level summary value is given by:

$$z_{genej} = \sigma\left(\overrightarrow{w_j} \cdot \overrightarrow{x_j}\right)$$

The gene-level summary values are concatenated to form an array of gene-level summaries ($\vec{z} \in \mathbb{R}^n$):

$$\vec{z} = \left[z_{gene1}\ z_{gene2}\ z_{gene3} \cdots z_{genen}\right]$$

The final prediction for the network can be obtained using the following transformation via an MLP, f:

$$\hat{y} = f(\vec{z})$$

$$\hat{p} = softmax(\hat{y})$$

In the classification case, this predicted outcome is compared to the expected outcome via:

$$L_{CE} = \frac{-\sum_i \sum_c y_{i,c} log\left(\widehat{p_{i,c}}\right)}{N}$$

We applied Group L1 regularization to these weights to cause certain genes to drop out, returning genes important for the prediction of the cancer subtypes. The final LASSO penalty is given by:

$$L_{L1} = \sum_j \sqrt{d_j}\ \left\|\overrightarrow{w_{gene\ j}}\right\|_2$$

Where d_j is the number of CpGs assigned to that gene. An intermediate layer of the neural network, $\vec{z}$, stores gene-level summaries of DNAm information, and $\overrightarrow{w_{gene\ j}}$ contains the importance of each CpG for a particular gene.

Here, we contrast this summary measure, $z_{genej} = \sigma\left(\overrightarrow{w_j} \cdot \overrightarrow{x_j}\right)$ to a more traditional summary

measure, such as the median or mean methylation (mean displayed on the right): $\beta_{genej} = \frac{\sum_k \beta_{jk}}{d_j}$.

Assuming the mean as our measurement, for simplification, it can be seen that each CpG is given

equal weight $\frac{1}{d_j}$, while for *MethylSPWNet*, each CpG is given weight w_{jk}, which is learnable and

reflective of the relative contribution of the given methylation beta value to the aggregate

measure. A comparison between z_{genej} and β_{genej} can be found in the supplementary materials.

Hyperparameter Scans

MethylCapsNet includes the use of a hyperparameter optimization scheme, accessible

through the *methylcaps-hypscan* module. Currently offered by the package is the availability to

scan a number of hyperparameters, including the number of training epochs, length of the

genomic region, the minimum number of CpGs to constitute a capsule, weighting schemes for

reconstruction loss and survival loss, learning rate, in addition to other capsule network focused

hyperparameters. Additionally, the search of *MethylNet* model architecture, randomized neural

network topologies, was replaced by a framework that searches for the ideal number of neurons

per neural network layer, conditional upon the choice of the number of layers. There are three

search strategies for optimization, including randomized searches and Bayesian optimization

techniques. This scheme differs from *MethylNet*, as both the neural network topology and set of

hyperparameters can be optimized through the application of successive Gaussian processes to

update some prior of losses over the set of hyperparameters. However, the results presented in

this manuscript utilized the randomized search design. The jobs can be launched in parallel and

scaled to meet the demands of a larger compute cluster.

Capsule Generation

Capsules specify the groupings of CpGs of the *MethylationArray* object. Capsule selection has been incorporated into the *hyperparameter_scan* and the *methylcaps-model* subcommands. Application Programming Interface (API) access to capsule selection and the building may be accessed through the *build_capsules* script. Prespecified capsules include annotations from gene sets from MSigDB and GSEA: 1) genes, 2) gene upstream/downstream, 3) the following Illumina methylation array annotations – UCSC_RefGene_Name, UCSC_RefGene_Accession, UCSC_RefGene_Group, UCSC_CpG_Islands_Name, Relation_to_UCSC_CpG_Island, Phantom, DMR, Enhancer, HMM_Island, Regulatory_Feature_Name, Regulatory_Feature_Group, and DHS – and 4) the following GSEA gene sets: C5.BP, C6, C1, H, C3.MIR, C2.CGP, C4.CM, C5.CC, C3.TFT, C5.MF, C7, C2.CP, C4.CGN. Users can also specify their own capsules through the presentation of a pickled dictionary containing a DataFrame that maps each CpG to a context name of choice. In addition, capsule generation has been integrated with BedTools [107] (*genomic_binned* selection), which can break up the entire hg19 genome into overlapping windows of fixed width, CpGs in these windows will belong to these capsules. We recommend the utilization of the Circos tool [108] for visualization of derived capsule relationships using the *genomic_binned* option.

Selection of Capsules for *MethylCapsNet* and *MethylSPWNet*

For the training of *MethylSPWNet*, we utilized all genes that overlapped with the 200,000 most variable CpGs across the CNS tumors. For *MethylCapsNet*, we could not utilize the complete set of genes due to the number of free parameters, a gene list of 650 genes was manually curated for

MethylCapsNet that included genes related to WNT, SHH, DKK1, beta-catenin, SFRP, NPR3,

amongst others; this list was reduced to 55 genes via recognition of genes by domain experts and

thresholding of the minimum number of CpGs. As a further description, for both approaches,

hyperparameter scans were utilized for pruning genes that did not contain a minimum number of

CpGs (this threshold was varied via the hyperparameter scan), resulting in a lower number of

genes than originally specified (n=10,341). In future iterations of the Capsule Network-based

approach, the constraints on gene selection will be lifted via reduction of free parameters and

adoption of explicit network building approaches.

Co-Methylation Embedding Modules

MethylSPWNet derived gene-level methylation summaries/embeddings were correlated to each

other and within their own set of top genes; Louvain modularity was performed on a k-nearest

neighbors graph of *MethylSPWNet* gene-level embeddings to establish preliminary modules. For

MethylSPWNet, final gene co-methylation/embedding analysis was carried out on a subtype-

specific basis, done so through the use of WGCNA.

For *MethylCapsNet*, capsule level embeddings were averaged across all individuals to form

overall embeddings, though just as relevant, these approaches can be extended to capsule-level

embeddings on the individual level or aggregated across meaningful subgroups. To derive final

measures of coordinated response between capsules, we averaged the coefficients of the routing

matrix across the individuals to form a weighted bipartite graph, and calculated a bipartite

projection of the graph to form a unipartite graph of capsules. We utilized the Louvain

modularity algorithm to discover hubs in this network and performed enrichment analyses on the pathway level using enrichr [109] to describe these hubs.

Analysis of CpG Weights derived from *MethylSPWNet*

MethylSPWNet derives CpG-specific weights, $\vec{w}$, that relates each CpG to its respective gene. We rank-ordered, reverse rank-ordered, and absolute-value reverse rank-ordered these lists to yield CpGs that were important to the differentiation of the tumor types. We subset the first 1000 CpGs and marked which genes they corresponded to and tested for enrichment using enrichr in our preliminary weight analysis. Finally, weights were also rank-ordered and reverse-rank ordered to yield the set of top negative and positive weights, respectively, the CpGs correspondent to the top number of CpGs (selected to highlight tendencies of enrichment and depletion) were related to the various island and gene context.

Method to Cluster Gene-Level Brain Cancer Embeddings by Samples

Recall that embeddings for individuals for *MethylSPWNet* were given in the following form (the design matrix is of dimensionality samples by genes):

$$\vec{z} = \begin{bmatrix} z_{gene1} \ z_{gene2} \ z_{gene3} \cdots z_{genen} \end{bmatrix}$$

For *MethylNet*, the embeddings are derived using the encoder:

$$\vec{z} = f(\vec{x})$$

Embeddings for individuals using the *MethylCapsNet* approach (of dimensionality samples by genes by latent dimensions) can be obtained by either averaging or concatenating (an aggregation, or *AGG* operator) the gene-level embeddings:

$$\overleftrightarrow{z} = AGG\left(\begin{bmatrix} \overrightarrow{z_{gene1}} \\ \vdots \\ \overrightarrow{z_{genen}} \end{bmatrix}\right)$$

Stacking these vectors for individuals would yield a design matrix that can be clustered using methods such as hierarchical clustering. We implemented hierarchical clustering using *scikit-learn* (>0.22) and found 14 clusters to compare against true labels of cell-of-origin, histological subtype, and histological and molecular subtype using the v-measure statistic and cluster separation using the Silhouette coefficient.

Software Availability

The software (*MethylCapsNet*, *MethylSPWNet*, and Group LASSO logistic regression) is open source and can be found on GitHub at https://github.com/Christensen-Lab-Dartmouth/MethylCapsNet, on PyPI under the tag *methylcapsnet*, and on Docker at *joshualevy44/methylcapsnet*. While new features may be developed for the *MethylCapsNet* framework, community contributions are welcome in the form of GitHub pull requests and issues. A test pipeline is available in the software implementation, a Wiki, help documentation, and example R scripts for possible downstream analyses can be found in the GitHub repository.

Web Application

We have developed a web application for the submission and investigation of *MethylCapsNet* outputs. The web application features three modules. The first is the network projection model, where capsules are related two to each other across subtypes, and network configurations can be changed by having some user tweak the relationships between the capsules and conservation.

The second module displays routing information, and the third module displays embedding information. Usage is detailed in the wiki.

Analysis Hardware and Software

The analyses run for this work were optimized utilizing K80 GPUs at the Dartmouth Research Computing Cluster. The algorithms were designed using Python 3.7, PyTorch version 1.1, and CUDA 9.0.

Dataset Availability

We acquired data from GEO accession GSE109381 preprocessed data using *PyMethylProcess* and the subselected 200K of the most hypervariable CpGs (to focus on CpGs that may better differentiate CNS tumor subtypes) after functional normalization was applied to the data. SNPs and autosomal probes were omitted. Preprocessing steps have been detailed in using the pipeline of *PyMethylProcess* [21].

Abbreviations

ANN – Artificial Neural Network
CGI – CpG Island
CIMP – CpG Island Methylator Phenotype
CNS – Central Nervous System
CpG – Cytosine Guanine Dinucleotide
DNAm – DNA Methylation
GBM – Glioblastoma Multiforme
LGG – Low-Grade Glioma
MB – Medulloblastoma
MLP – Multi-Layer Perceptron
mQTL – Methylated Quantitative Trait Loci
ReLU – Rectified Linear Unit

Tanh – Hyperbolic Tangent
TDA – Topological Data Analysis
TOM – Topological Overlap Matrix
WGCNA – Weighted Gene Correlation Network Analysis

Declarations

Ethics Approval and Consent to Participate

Not applicable.

Consent for Publication

Not applicable.

Availability of Data and Materials

Data used in this study was acquired from GEO accession GSE109381. Test data is available in our GitHub repository.

Correspondence Statement

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Competing Interests

Not applicable.

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Authors' Contributions

The conception and design of the study were contributed by JJL and BCC. Implementation, programming, data acquisition, and analyses were by JJL. All authors contributed towards refining the analytic plan and direction. All authors contributed to the writing and editing of the manuscript. CLP and JJL tested the pipeline.

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References

1. Bell CG, Lowe R, Adams PD, Baccarelli AA, Beck S, Bell JT, et al. DNA methylation aging clocks: challenges and recommendations. Genome Biol. 2019;20:249.

2. Khavari DA, Sen GL, Rinn JL. DNA methylation and epigenetic control of cellular differentiation. Cell Cycle. 2010;9:3880–3.

3. Christensen BC, Houseman EA, Marsit CJ, Zheng S, Wrensch MR, Wiemels JL, et al. Aging and environmental exposures alter tissue-specific DNA methylation dependent upon CpG island context. PLoS Genet. 2009;5:e1000602.

4. Dedeurwaerder S, Defrance M, Calonne E, Denis H, Sotiriou C, Fuks F. Evaluation of the Infinium Methylation 450K technology. Epigenomics. 2011;3:771–84.

5. Moran S, Arribas C, Esteller M. Validation of a DNA methylation microarray for 850,000 CpG sites of the human genome enriched in enhancer sequences. Epigenomics. 2016;8:389–99.

6. Heyn H, Esteller M. DNA methylation profiling in the clinic: applications and challenges. Nature Reviews Genetics. Nature Publishing Group; 2012;13:679–92.

7. Dor Y, Cedar H. Principles of DNA methylation and their implications for biology and medicine. The Lancet. Elsevier; 2018;392:777–86.

8. Capper D, Jones DTW, Sill M, Hovestadt V, Schrimpf D, Sturm D, et al. DNA methylation-based classification of central nervous system tumours. Nature. 2018;555:469–74.

9. Hegi ME, Diserens A-C, Gorlia T, Hamou M-F, de Tribolet N, Weller M, et al. MGMT Gene Silencing and Benefit from Temozolomide in Glioblastoma. New England Journal of Medicine. Massachusetts Medical Society; 2005;352:997–1003.

10. Turcan S, Rohle D, Goenka A, Walsh LA, Fang F, Yilmaz E, et al. IDH1 mutation is sufficient to establish the glioma hypermethylator phenotype. Nature. 2012;483:479–83.

11. Noushmehr H, Weisenberger DJ, Diefes K, Phillips HS, Pujara K, Berman BP, et al. Identification of a CpG Island Methylator Phenotype that Defines a Distinct Subgroup of Glioma. Cancer Cell. 2010;17:510–22.

12. Christensen BC, Smith AA, Zheng S, Koestler DC, Houseman EA, Marsit CJ, et al. DNA Methylation, Isocitrate Dehydrogenase Mutation, and Survival in Glioma. J Natl Cancer Inst. 2011;103:143–53.

13. J. Dabrowski M, Wojtas B. Global DNA Methylation Patterns in Human Gliomas and Their Interplay with Other Epigenetic Modifications. Int J Mol Sci [Internet]. 2019 [cited 2020 Jul 21];20. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6678179/

14. Cavalli FMG, Remke M, Rampasek L, Peacock J, Shih DJH, Luu B, et al. Intertumoral Heterogeneity within Medulloblastoma Subgroups. Cancer Cell. 2017;31:737-754.e6.

15. Maros ME, Capper D, Jones DTW, Hovestadt V, von Deimling A, Pfister SM, et al. Machine learning workflows to estimate class probabilities for precision cancer diagnostics on DNA methylation microarray data. Nature Protocols. Nature Publishing Group; 2020;15:479–512.

16. Rauschert S, Raubenheimer K, Melton PE, Huang RC. Machine learning and clinical epigenetics: a review of challenges for diagnosis and classification. Clinical Epigenetics. 2020;12:51.

17. LeCun Y, Bengio Y, Hinton G. Deep learning. Nature. 2015;521:436–44.

18. Hinton GE. Connectionist learning procedures. Artificial Intelligence. 1989;40:185–234.

19. Krizhevsky A, Sutskever I, Hinton GE. ImageNet Classification with Deep Convolutional Neural Networks. In: Pereira F, Burges CJC, Bottou L, Weinberger KQ, editors. Advances in Neural Information Processing Systems 25 [Internet]. Curran Associates, Inc.; 2012 [cited 2019 Jun 10]. p. 1097–1105. Available from: http://papers.nips.cc/paper/4824-imagenet-classification-with-deep-convolutional-neural-networks.pdf

20. Levy JJ, Titus AJ, Petersen CL, Chen Y, Salas LA, Christensen BC. MethylNet: an automated and modular deep learning approach for DNA methylation analysis. BMC Bioinformatics. 2020;21:108.

21. Levy JJ, Titus AJ, Salas LA, Christensen BC. PyMethylProcess - convenient high-throughput preprocessing workflow for DNA methylation data. Bioinformatics. 2019;

22. Titus AJ, Wilkins OM, Bobak CA, Christensen BC. Unsupervised deep learning with variational autoencoders applied to breast tumor genome-wide DNA methylation data with biologic feature extraction. bioRxiv. 2018;433763.

23. Titus AJ, Bobak CA, Christensen BC. A New Dimension of Breast Cancer Epigenetics - Applications of Variational Autoencoders with DNA Methylation. 2018 [cited 2019 Jun 15]. p. 140–5. Available from: https://www.scitepress.org/PublicationsDetail.aspx?ID=kxmWGvGaZh0=&t=1

24. Angermueller C, Lee HJ, Reik W, Stegle O. DeepCpG: accurate prediction of single-cell DNA methylation states using deep learning. Genome Biology. 2017;18:67.

25. Lundberg SM, Lee S-I. A Unified Approach to Interpreting Model Predictions. In: Guyon I, Luxburg UV, Bengio S, Wallach H, Fergus R, Vishwanathan S, et al., editors. Advances in Neural Information Processing Systems 30 [Internet]. Curran Associates, Inc.; 2017 [cited 2019 Jun 10]. p. 4765–4774. Available from: http://papers.nips.cc/paper/7062-a-unified-approach-to-interpreting-model-predictions.pdf

26. Ribeiro MT, Singh S, Guestrin C. "Why Should I Trust You?": Explaining the Predictions of Any Classifier. arXiv:160204938 [cs, stat] [Internet]. 2016 [cited 2020 Feb 9]; Available from: http://arxiv.org/abs/1602.04938

27. Camacho DM, Collins KM, Powers RK, Costello JC, Collins JJ. Next-Generation Machine Learning for Biological Networks. Cell. 2018;173:1581–92.

28. Horvath S. DNA methylation age of human tissues and cell types. Genome Biol. 2013;14:R115.

29. Handl L, Jalali A, Scherer M, Eggeling R, Pfeifer N. Weighted elastic net for unsupervised domain adaptation with application to age prediction from DNA methylation data. Bioinformatics. 2019;35:i154–63.

30. Sun H, Wang S. Penalized logistic regression for high-dimensional DNA methylation data with case-control studies. Bioinformatics. 2012;28:1368–75.

31. Zhou W, Lo S-H. Analysis of genotype by methylation interactions through sparsity-inducing regularized regression. BMC Proceedings. 2018;12:40.

32. Choi J, Kim K, Sun H. New variable selection strategy for analysis of high-dimensional DNA methylation data. J Bioinform Comput Biol. 2018;16:1850010.

33. Dong NT, Khosla M. Revisiting Feature Selection with Data Complexity. bioRxiv. 2019;754630.

34. Sun L, Namboodiri S, Chen E, Sun S. Preliminary Analysis of Within-Sample Co-methylation Patterns in Normal and Cancerous Breast Samples. Cancer Inform. 2019;18:1176935119880516.

35. Rickabaugh TM, Baxter RM, Sehl M, Sinsheimer JS, Hultin PM, Hultin LE, et al. Acceleration of age-associated methylation patterns in HIV-1-infected adults. PLoS ONE. 2015;10:e0119201.

36. Zhang J, Huang K. Pan-cancer analysis of frequent DNA co-methylation patterns reveals consistent epigenetic landscape changes in multiple cancers. BMC Genomics. 2017;18:1045.

37. Gomez L, Odom GJ, Young JI, Martin ER, Liu L, Chen X, et al. coMethDMR: accurate identification of co-methylated and differentially methylated regions in epigenome-wide association studies with continuous phenotypes. Nucleic Acids Res. 2019;47:e98–e98.

38. Lien TG, Borgan Ø, Reppe S, Gautvik K, Glad IK. Integrated analysis of DNA-methylation and gene expression using high-dimensional penalized regression: a cohort study on bone mineral density in postmenopausal women. BMC Medical Genomics. 2018;11:24.

39. Ng B, Jafarzadeh S, Cole D, Goldenberg A, Mostafavi S. DNA Methylation Network Estimation with Sparse Latent Gaussian Graphical Model. 2018.

40. Davies M, Volta M, Pidsley R, Lunnon K, Dixit A, Lovestone S, et al. Functional annotation of the human brain methylome identifies tissue-specific epigenetic variation across brain and blood. Genome biology. 2012;13:R43.

41. Cui Z-J, Zhou X-H, Zhang H-Y. DNA Methylation Module Network-Based Prognosis and Molecular Typing of Cancer. Genes (Basel) [Internet]. 2019 [cited 2020 Feb 9];10. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6722866/

42. Mallona I, Aussó S, Díez-Villanueva A, Moreno V, Peinado MA. Modular dynamics of DNA co-methylation networks exposes the functional organization of colon cancer cells' genome. bioRxiv. 2018;428730.

43. Tremblay BL, Guénard F, Lamarche B, Pérusse L, Vohl M-C. Network Analysis of the Potential Role of DNA Methylation in the Relationship between Plasma Carotenoids and Lipid Profile. Nutrients. 2019;11:1265.

44. Mallik S, Bandyopadhyay S. WeCoMXP: Weighted Connectivity Measure Integrating Co-Methylation, Co-Expression and Protein-Protein Interactions for Gene-Module Detection. IEEE/ACM Trans Comput Biol Bioinform. 2018;

45. Wang F, Xu H, Zhao H, Gelernter J, Zhang H. DNA co-methylation modules in postmortem prefrontal cortex tissues of European Australians with alcohol use disorders. Scientific Reports. 2016;6:1–11.

46. Bartlett TE, Olhede SC, Zaikin A. A DNA Methylation Network Interaction Measure, and Detection of Network Oncomarkers. PLoS One [Internet]. 2014 [cited 2020 Feb 10];9. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3882261/

47. Horvath S, Zhang Y, Langfelder P, Kahn RS, Boks MPM, van Eijk K, et al. Aging effects on DNA methylation modules in human brain and blood tissue. Genome Biol. 2012;13:R97.

48. Langfelder P, Horvath S. WGCNA: an R package for weighted correlation network analysis. BMC Bioinformatics. 2008;9:559.

49. Akulenko R, Helms V. DNA co-methylation analysis suggests novel functional associations between gene pairs in breast cancer samples. Hum Mol Genet. 2013;22:3016–22.

50. Affinito O, Palumbo D, Fierro A, Cuomo M, De Riso G, Monticelli A, et al. Nucleotide distance influences co-methylation between nearby CpG sites. Genomics. 2020;112:144–50.

51. Hao J, Kim Y, Kim T-K, Kang M. PASNet: pathway-associated sparse deep neural network for prognosis prediction from high-throughput data. BMC Bioinformatics. 2018;19:510.

52. Hao J, Masum M, Oh JH, Kang M. Gene- and Pathway-Based Deep Neural Network for Multi-omics Data Integration to Predict Cancer Survival Outcomes. In: Cai Z, Skums P, Li M, editors.

Bioinformatics Research and Applications. Cham: Springer International Publishing; 2019. p. 113–24.

53. Borisov V, Haug J, Kasneci G. CancelOut: A Layer for Feature Selection in Deep Neural Networks. In: Tetko IV, Kůrková V, Karpov P, Theis F, editors. Artificial Neural Networks and Machine Learning – ICANN 2019: Deep Learning. Cham: Springer International Publishing; 2019. p. 72–83.

54. Crawford J, Greene CS. Incorporating biological structure into machine learning models in biomedicine. Current Opinion in Biotechnology. 2020;63:126–34.

55. Xie G, Dong C, Kong Y, Zhong JF, Li M, Wang K. Group Lasso Regularized Deep Learning for Cancer Prognosis from Multi-Omics and Clinical Features. Genes (Basel) [Internet]. 2019 [cited 2020 Mar 29];10. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6471789/

56. Barthel FP, Johnson KC, Wesseling P, Verhaak RGW. Evolving Insights into the Molecular Neuropathology of Diffuse Gliomas in Adults. Neurol Clin. 2018;36:421–37.

57. Artemenkov A, Panov M. NCVis: Noise Contrastive Approach for Scalable Visualization. 2020 [cited 2020 May 6]; Available from: https://arxiv.org/abs/2001.11411v1

58. Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, et al. STRING v11: protein–protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. Nucleic Acids Res. 2019;47:D607–13.

59. Babic I, Mischel PS. Multiple functions of a glioblastoma fusion oncogene. J Clin Invest. 2013;123:548–51.

60. Parker BC, Annala MJ, Cogdell DE, Granberg KJ, Sun Y, Ji P, et al. The tumorigenic FGFR3-TACC3 gene fusion escapes miR-99a regulation in glioblastoma. J Clin Invest. 2013;123:855–65.

61. Macy ME, Birks DK, Barton VN, Chan MH, Donson AM, Kleinschmidt-DeMasters BK, et al. Clinical and molecular characteristics of congenital glioblastoma. Neuro Oncol. Oxford Academic; 2012;14:931–41.

62. Berezovsky AD, Poisson LM, Cherba D, Webb CP, Transou AD, Lemke NW, et al. Sox2 Promotes Malignancy in Glioblastoma by Regulating Plasticity and Astrocytic Differentiation. Neoplasia. 2014;16:193-206.e25.

63. Ibrahim K, Abdul Murad NA, Harun R, Jamal R. Knockdown of Tousled-like kinase 1 inhibits survival of glioblastoma multiforme cells. International Journal of Molecular Medicine. Spandidos Publications; 2020;46:685–99.

64. Huang Q, Zhang X-W, Ma Y-S, Lu G-X, Xie R-T, Yang H-Q, et al. Up-regulated microRNA-299 corrected with poor prognosis of glioblastoma multiforme patients by targeting ELL2. Jpn J Clin Oncol. Oxford Academic; 2017;47:590–6.

65. Krishnan R, Boddapati N, Mahalingam S. Interplay between human nucleolar GNL1 and RPS20 is critical to modulate cell proliferation. Scientific Reports. Nature Publishing Group; 2018;8:11421.

66. Friesen C, Hormann I, Roscher M, Fichtner I, Alt A, Hilger R, et al. Opioid receptor activation triggering downregulation of cAMP improves effectiveness of anti-cancer drugs in treatment of glioblastoma. Cell Cycle. 2014;13:1560–70.

67. Pearson JRD, Regad T. Targeting cellular pathways in glioblastoma multiforme. Signal Transduct Target Ther. 2017;2:17040.

68. Xiong A, Roy A, Spyrou A, Weishaupt H, Marinescu VD, Olofsson T, et al. Nuclear Receptor Binding Protein 2 Is Downregulated in Medulloblastoma, and Reduces Tumor Cell Survival upon Overexpression. Cancers (Basel) [Internet]. 2020 [cited 2020 Aug 9];12. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7352854/

69. de la Rocha AMA, Sampron N, Alonso MM, Matheu A. Role of SOX family of transcription factors in central nervous system tumors. Am J Cancer Res. 2014;4:312–24.

70. Rivero-Hinojosa S, Lau LS, Stampar M, Staal J, Zhang H, Gordish-Dressman H, et al. Proteomic analysis of Medulloblastoma reveals functional biology with translational potential. acta neuropathol commun. 2018;6:48.

71. Qi Y, Gao Y. Clinical significance of miR-33b in glioma and its regulatory role in tumor cell proliferation, invasion and migration. Biomarkers in Medicine. Future Medicine; 2020;14:539–48.

72. Wang X, Huang Z, Wu Q, Prager BC, Mack SC, Yang K, et al. MYC-regulated Mevalonate Metabolism Maintains Brain Tumor Initiating Cells. Cancer Res. 2017;77:4947–60.

73. Marx S, Xiao Y, Baschin M, Splittstöhser M, Altmann R, Moritz E, et al. The Role of Platelets in Cancer Pathophysiology: Focus on Malignant Glioma. Cancers (Basel) [Internet]. 2019 [cited 2020 Aug 9];11. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6521321/

74. Wu X, Rauch TA, Zhong X, Bennett WP, Latif F, Krex D, et al. CpG island hypermethylation in human astrocytomas. Cancer Res. 2010;70:2718–27.

75. Caponegro MD, Moffitt RA, Tsirka SE. Expression of neuropilin-1 is linked to glioma associated microglia and macrophages and correlates with unfavorable prognosis in high grade gliomas. Oncotarget. 2018;9:35655–65.

76. Xia Z, Ouyang D, Li Q, Li M, Zou Q, Li L, et al. The Expression, Functions, Interactions and Prognostic Values of PTPRZ1: A Review and Bioinformatic Analysis. J Cancer. 2019;10:1663–74.

77. Panosyan EH, Lin HJ, Koster J, Lasky JL. In search of druggable targets for GBM amino acid metabolism. BMC Cancer [Internet]. 2017 [cited 2020 Aug 9];17. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5331648/

78. Rodríguez-Paredes M, Esteller M. Cancer epigenetics reaches mainstream oncology. Nat Med. 2011;17:330–9.

79. Chenn A. Wnt/β-catenin signaling in cerebral cortical development. Organogenesis. 2008;4:76–80.

80. Testa U, Castelli G, Pelosi E. Genetic Abnormalities, Clonal Evolution, and Cancer Stem Cells of Brain Tumors. Med Sci (Basel) [Internet]. 2018 [cited 2020 Jul 1];6. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6313628/

81. Li T, Forbes ME, Fuller GN, Li J, Yang X, Zhang W. IGFBP2: integrative hub of developmental and oncogenic signaling network. Oncogene. Nature Publishing Group; 2020;39:2243–57.

82. Ishak G, Leal MF, dos Santos NPC, Demachki S, Nunes CAM, do Nascimento Borges B, et al. Deregulation of MYC and TP53 through genetic and epigenetic alterations in gallbladder carcinomas. Clin Exp Med. 2015;15:421–6.

83. Jun I, Park HS, Piao H, Han JW, An MJ, Yun BG, et al. ANO9/TMEM16J promotes tumourigenesis via EGFR and is a novel therapeutic target for pancreatic cancer. Br J Cancer. 2017;117:1798–809.

84. Hatanpaa KJ, Burma S, Zhao D, Habib AA. Epidermal Growth Factor Receptor in Glioma: Signal Transduction, Neuropathology, Imaging, and Radioresistance. Neoplasia. 2010;12:675–84.

85. Sabour S, Frosst N, Hinton GE. Dynamic Routing Between Capsules. arXiv:171009829 [cs] [Internet]. 2017 [cited 2020 Jan 18]; Available from: http://arxiv.org/abs/1710.09829

86. Venkatraman S, S B, Sarma R. Building Deep, Equivariant Capsule Networks. 2019.

87. Wang L, Miao X, Zhang J, Cai J. MultiCapsNet: a interpretable deep learning classifier integrate data from multiple sources. bioRxiv. 2019;570507.

88. Danielsson A, Nemes S, Tisell M, Lannering B, Nordborg C, Sabel M, et al. MethPed: a DNA methylation classifier tool for the identification of pediatric brain tumor subtypes. Clin Epigenetics. 2015;7:62.

89. Hovestadt V, Jones DTW, Picelli S, Wang W, Kool M, Northcott PA, et al. Decoding the regulatory landscape of medulloblastoma using DNA methylation sequencing. Nature. 2014;510:537–41.

90. Baeza N, Weller M, Yonekawa Y, Kleihues P, Ohgaki H. PTEN methylation and expression in glioblastomas. Acta Neuropathol. 2003;106:479–85.

91. Capaccione KM, Pine SR. The Notch signaling pathway as a mediator of tumor survival. Carcinogenesis. 2013;34:1420–30.

92. Fan X, Matsui W, Khaki L, Stearns D, Chun J, Li Y-M, et al. Notch pathway inhibition depletes stem-like cells and blocks engraftment in embryonal brain tumors. Cancer Res. 2006;66:7445–52.

93. Li J, Yen C, Liaw D, Podsypanina K, Bose S, Wang SI, et al. PTEN, a putative protein tyrosine phosphatase gene mutated in human brain, breast, and prostate cancer. Science. 1997;275:1943–7.

94. He X, Zhang L, Chen Y, Remke M, Shih D, Lu F, et al. The G protein α subunit Gαs is a tumor suppressor in Sonic hedgehog-driven medulloblastoma. Nat Med. 2014;20:1035–42.

95. Pidsley R, Zotenko E, Peters TJ, Lawrence MG, Risbridger GP, Molloy P, et al. Critical evaluation of the Illumina MethylationEPIC BeadChip microarray for whole-genome DNA methylation profiling. Genome Biology. 2016;17:208.

96. Zhou L, Ng HK, Drautz-Moses DI, Schuster SC, Beck S, Kim C, et al. Systematic evaluation of library preparation methods and sequencing platforms for high-throughput whole genome bisulfite sequencing. Scientific Reports. Nature Publishing Group; 2019;9:10383.

97. Bodnar C, Cangea C, Liò P. Deep Graph Mapper: Seeing Graphs through the Neural Lens. arXiv:200203864 [cs, stat] [Internet]. 2020 [cited 2020 Feb 20]; Available from: http://arxiv.org/abs/2002.03864

98. Veen HJ van, Saul N, Eargle D, Mangham SW. Kepler Mapper: A flexible Python implementation of the Mapper algorithm. Journal of Open Source Software. 2019;4:1315.

99. Wang T, Johnson T, Jie Z, Huang K. Topological Methods for Visualization and Analysis of High Dimensional Single-Cell RNA Sequencing Data. Pacific Symposium on Biocomputing. 2019;24:350–61.

100. Rizvi AH, Camara PG, Kandror EK, Roberts TJ, Schieren I, Maniatis T, et al. Single-cell topological RNA-Seq analysis reveals insights into cellular differentiation and development. Nat Biotechnol. 2017;35:551–60.

101. Lum PY, Singh G, Lehman A, Ishkanov T, Vejdemo-Johansson M, Alagappan M, et al. Extracting insights from the shape of complex data using topology. Scientific Reports. Nature Publishing Group; 2013;3:1236.

102. Singh D, Yamada M. FsNet: Feature Selection Network on High-dimensional Biological Data. arXiv:200108322 [cs, stat] [Internet]. 2020 [cited 2020 Feb 22]; Available from: http://arxiv.org/abs/2001.08322

103. Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, et al. limma powers differential expression analyses for RNA-sequencing and microarray studies. Nucleic Acids Res. Oxford Academic; 2015;43:e47–e47.

104. Raudvere U, Kolberg L, Kuzmin I, Arak T, Adler P, Peterson H, et al. g:Profiler: a web server for functional enrichment analysis and conversions of gene lists (2019 update). Nucleic Acids Res. Oxford Academic; 2019;47:W191–8.

105. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: A Software Environment for Integrated Models of Biomolecular Interaction Networks. Genome Res. 2003;13:2498–504.

106. Gustavsen JA, Pai S, Isserlin R, Demchak B, Pico AR. RCy3: Network biology using Cytoscape from within R. F1000Res [Internet]. 2019 [cited 2020 Jul 2];8. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6880260/

107. Quinlan AR, Hall IM. BEDTools: a flexible suite of utilities for comparing genomic features. Bioinformatics. Oxford Academic; 2010;26:841–2.

108. Krzywinski M, Schein J, Birol İ, Connors J, Gascoyne R, Horsman D, et al. Circos: An information aesthetic for comparative genomics. Genome Res. 2009;19:1639–45.

109. Chen EY, Tan CM, Kou Y, Duan Q, Wang Z, Meirelles GV, et al. Enrichr: interactive and collaborative HTML5 gene list enrichment analysis tool. BMC Bioinformatics. 2013;14:128.