Genetic diversity analysis of Chinese plum (*Prunus salicina* L.) based on whole-genome resequencing

Xiao Wei[1,#], Fei Shen[2,#], Qiuping Zhang[1,*], Ning Liu[1], Yuping Zhang[1], Ming Xu[1], Shuo Liu[1], Yujun Zhang[1], Xiaoxue Ma[1], Weisheng Liu[1,*]

1 *Liaoning Institute of Pomology, Yingkou 115009*

2 *Beijing Agro-biotechnology Research Center, Beijing Academy of Agriculture and Forestry Sciences, Beijing 100193*

[#] Authors that contributed equally to this work.

[*] Corresponding author: Qiuping Zhang (e-mail: lbzhangqiuping2@163.com) and Weisheng Liu (e-mail: wsliulaas@163.com)

Abstract

Chinese plum (*Prunus salicina* L.), also known as Japanese plum, is gaining importance due to their extensive genetic diversity and nutritional attributes beneficial for human health. Single-nucleotide polymorphisms (SNPs) are the most abundant form of genomic polymorphisms and are widely used in population genetics research. Here, we construed high-density haplotype map by whole-genome resequencing of 67 *Prunus* accessions with a depth of ~20× to evaluate the genome-level diversity and population structure. The phylogenetic analysis, the principal component analysis, and the population structure profiling, indicated that the 67 plum accessions could be classified into four groups corresponding to their origin location, the southern cultivar group (SCG), the northern cultivar group (NCG), the foreign cultivar group (FG), and the mixed cultivar group (MG). Some cultivars from South China were clustered with the other three groups. The genetic diversity indices including the private allele number, the observed heterozygosity, the expected heterozygosity, and the nucleotide diversity of the SCG were higher than those of the NCG. The gene flow from the SCG to the FG was detected as well. We concluded that the origin center of Chinese plum was at the Yangtze River Basin in South China. This study provided genetic variation features and population structure of Chinese plum cultivars, laying a foundation for breeders to use diverse germplasm and allelic variants for improving Chinese plum varieties.

Keywords: *Prunus salicina* Lindl.; whole-genome resequencing; population structure; genetic introgression

1. Introduction

Plum is one of the most important fruit crops in the world, possessing extensive genetic diversity and high economic value. The largest plum producer is China, with an annual production of 6,801,187 metric tons in 2018, accounting for 53.9% of the world's total [1]. Within the genus, *Prunus*, Chinese plum (*Prunus salicina* L., 2n = 2x = 16), also known as Japanese plum, is widely grown for fresh market consumption and the canning industry, which includes both pure Chinese plum and its hybrids with other diploid plum species, such as *Prunus simonii* Carr., *Prunus cerasifera* Ehrh., *Prunus americana* Marsh., and others [2].

Chinese plum possible originates from the Yangtze River Basin, and now there are abundant plum germplasm resources in China [3]. With its long growing history and extensive geographical distribution, there are more than 800 indigenous plum cultivars in China derived from *Prunus salicina* L. alone. Over 600 of them are now preserved at National Germplasm Repository for Plums and Apricots (NGRPA) located in Xiongyue, Liaoning Province, China [4,5]. Yu et al. [6] carried out comprehensive phenotyping of 405 Chinese plum cultivars and its hybrids from NGRPA, totally 32 morphological and agronomic characters were investigated, of which their coefficients of variations (CVs) were calculated ranging from 14.85% to 47.09%, suggesting that the genetic variability of Chinese plums distributed widely.

Genetic variability is the prerequisite for any plant breeding program. Learning the extent and structure of genetic variation in germplasm collections is a crucial step for the efficient conservation and utilization of biodiversity in cultivated crops, and using

diverse plum resources to broaden the genetic base of worldwide plum cultivars is the critical objective for plum breeders [2,7]. Previous efforts have been made to understand the genetic diversity of plum better. Zhang and Zhang [8] took the lead in collecting plum germplasm resources and evaluating their genetic diversity on the basis of morphological traits and isozyme polymorphisms, whereas, morphological traits were highly susceptible to environmental factors, so the estimates of genetic diversity were not precise; while isozymes had a low degree of polymorphism and hence were not efficient enough for the characterization of germplasm genetic diversity [9]. Overcoming the above-mentioned problems, DNA-based markers became widely applied in plum genetic diversity analysis, including random amplified polymorphic DNAs (RAPDs) [5,10], simple sequence repeats (SSRs) [11-13], inter-simple sequence repeats (ISSRs) [2,14]. In addition, benefiting from the advances in next-generation sequencing (NGS) technologies, genotyping by sequencing (GBS), provides a great wealth of information that makes it possible to identify thousands of single nucleotide polymorphisms (SNPs), which, after adequate filtering, allow us to carry out detailed genetic diversity studies [7,15-17].

By conducting DNA-based marker assessments, Chinese plums were generally classified into two major groups, the southern cultivar group (SCG) and the northern cultivar group (NCG) [18-20]. The *P. salicina* system in South China was rather primitive than that in North China basing on a palynology study, and the spread of *P. salicina* was from the south to the north [21]. However, simply cultivar clustering works are inadequate for the integrated study of genetic diversity of Chinese plum. As

80 described in peach, large scale SNP data-based diversity analysis boosted the
81 deciphering of the evolution and domestication of peach germplasm resources
82 [22,23].
83 In the present study, we used SNP markers generated from high depth whole-genome
84 resequencing data (an average depth of ~20×) for the first time to elucidate the
85 pattern of genetic diversity, population structure, and domestication of a diverse *P.*
86 *salicina* collection. An in-depth understanding of their genetic relationships could
87 benefit plum germplasm conservation and utilization, and lead to new cultivar
88 improvement.
89

90 **2. Results**

91 *2.1 Markers development based on whole-genome resequencing*

92 We collected 67 plum accessions representing different geographic and morphological
93 characteristics (Table S1). The 67 plum accessions were sequenced using the Illumina
94 HiSeq 2500 platform with a sequence depth higher than 20×; a totally 462.6 Gb clean
95 data were retained with an average of 6.9 Gb for each accession after filtering out
96 low-quality reads. The quality of the sequencing data was high with a Q30 > 85%,
97 and a GC content slightly fluctuated around 38.0%. The mapping rate to the peach
98 genome was range from 85.73% to 88.57%, with a average of 87.49% (Table S1). We
99 called SNPs with the unique mapped reads, and a total of 16,600,033 SNPs and
100 2,107,664 INDELs were identified across the 67 accessions using SAMtools v1.4
101 software. After filtering out these in low quality, we obtained 14,549,234 SNPs which

could ensure the accuracy and reliability of subsequent genetic diversity and population structure analyses (Figure 1). About 43.81% of the SNPs were located in intergenic regions, and 11.88% were in coding regions. The non-synonymous to synonymous substitution ratio (dN/dS) for the SNPs in the coding regions was 15.92. Transitions were found in a proportion of 59.58% (10315764/16600033), with a transition/transversion ratio (Ts/Tv ratio) of 1.47 (Figure 1, Table 1).

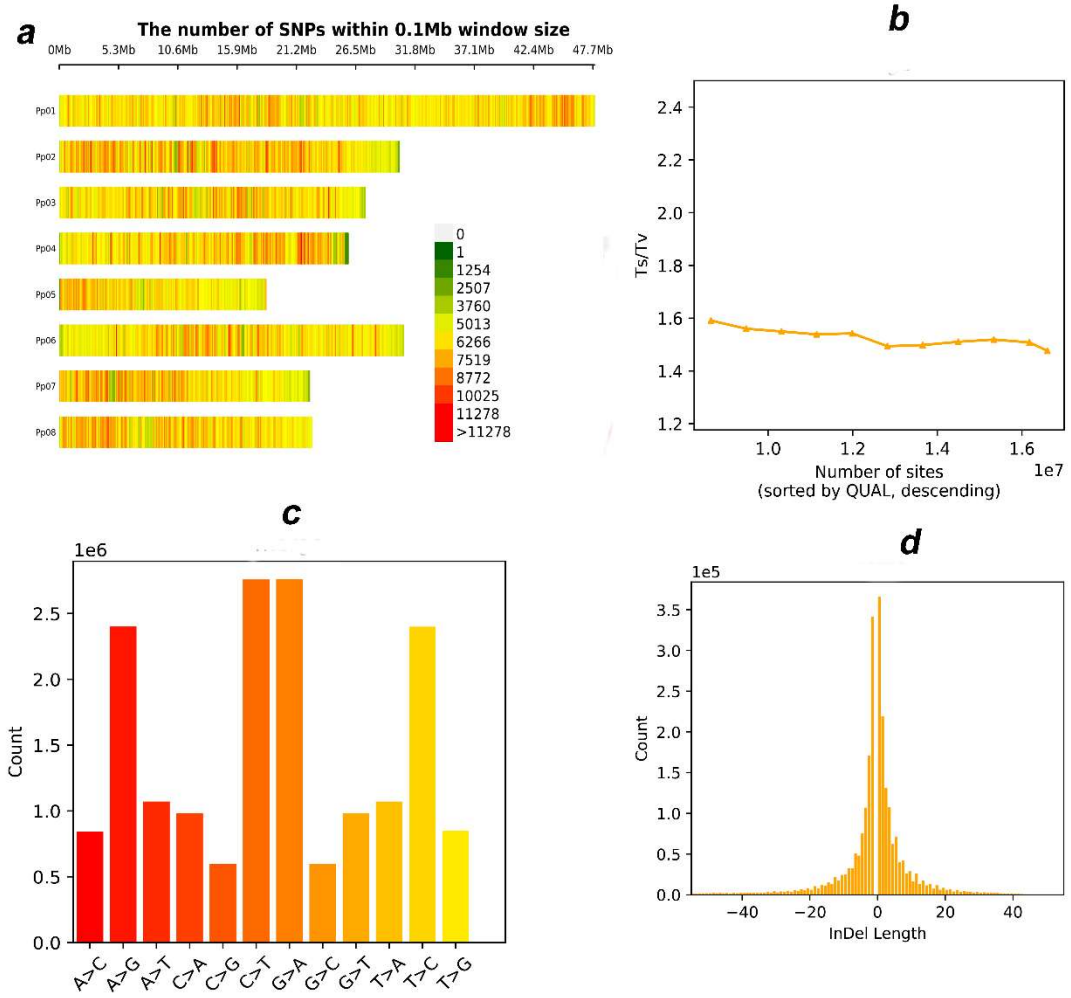


Figure 1. The statistics of markers generated from the whole-genome resequencing of the 67 plum accessions. a) The chromosome-scale SNP distribution; b) The distribution of transition/transversion ratio (Ts/Tv ratio); c) The counts of different types of transitions and transversions; d) The counts of genome-wide insertions and

113 deletions.

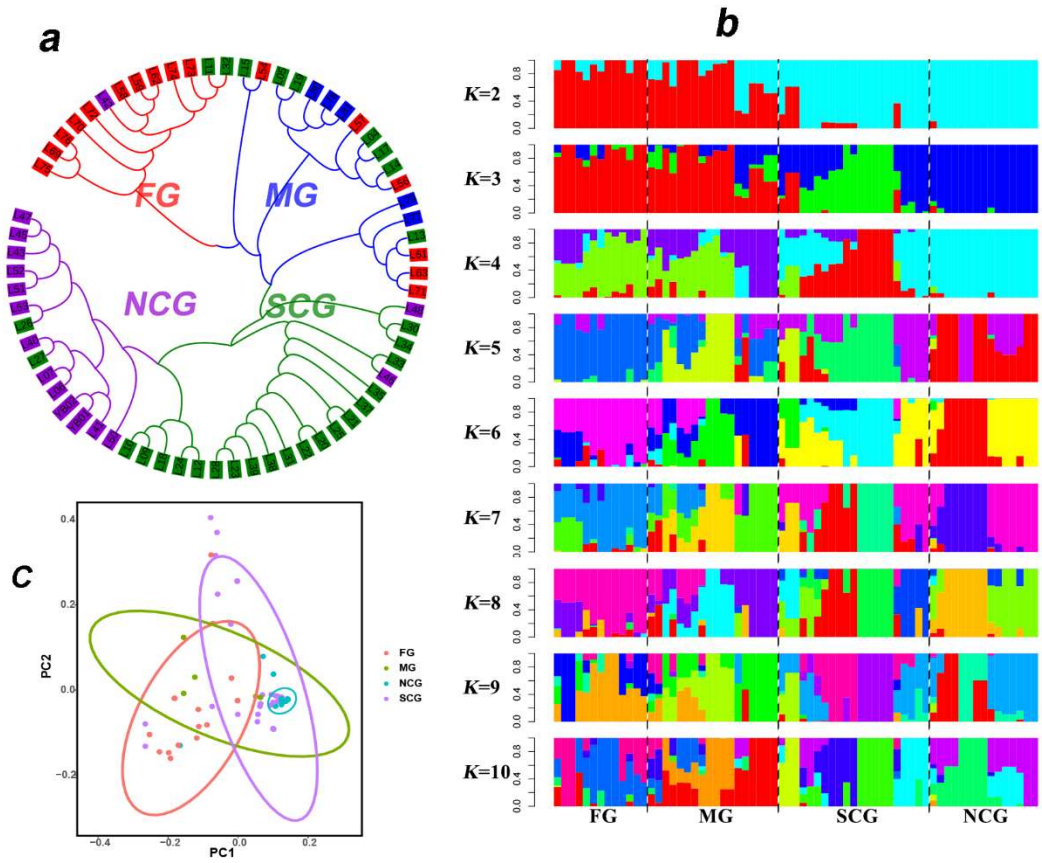
114 **Table 1**. Statistics of variations across different chromosomes

Categories	Numbers	Ratio
Raw SNPs	16,600,033	88.73%
InDels	2,107,664	11.27%
Filtered SNPs	14,549,234	87.65%
Intergenic regions SNPs	7,272,007	43.81%
Coding regions SNPs	1,971,748	11.88%
Transition	10,315,764	59.58%
Transversion	6,999,126	40.42%
Transition/Transversion	--	147.00%
Non-synonymous SNPs	1,855,201	94.09%
Synonymous SNPs	116,547	5.91%

115 *2.2 Phylogenetic analysis*

116 To exploit the relationship among the Chinese plum, a Neighboring-joining
117 phylogenetic tree of the 67 accessions was constructed with all SNPs (Figure 2a). The
118 phylogenetic tree classified the accessions into four main groups which are
119 corresponding to their origin location: (1) the southern cultivar group (SCG) was
120 comprised of the plum cultivars from Sichuan, Guizhou, Yunnan, Guangdong,
121 Guangxi, Zhejiang, and Fujian Province; (2) the northern cultivar group (NCG)
122 comprised of plum cultivars mainly from Hebei, Henan, Shandong, Shaanxi Province,
123 and the south part of Liaoning Province; (3) the foreign cultivar group (FG) included
124 plum cultivars from the USA and Japan; and (4) the mixed cultivar group (MG)
125 consisted of newly bred cultivars from the NGRPA and several cultivars originated
126 SCG and FG. Notably, we fould 'Saozouli' and 'Shuili' cultivars collected from south
127 China were clustered with NCG, 'Zaohuangli' and 'Jinshali' with FG, and 'Wanshu
128 huanai', 'Huahongli' and 'Abazhoumeiguili' with MG, suggesting that SCG is more

129 diverse than any other group.



130

131 **Figure 2**. The population structure of the 67 plum accessions. a) Neighboring-joining

132 phylogenetic tree constructed using SNPs at fourfold degenerate site. Each group was

133 color coded; b) Bayesian model-based clustering of the 67 plum accessions with the

134 number of ancestry kinship (*K*) from 2 to 10; c) Principle Component Analysis (PCA)

135 of the 67 plum accessions. FG, the foreign cultivar group; MG, the mixed cultivar

136 group; SCG, the southern cultivar group; NCG, the northern cultivar group.

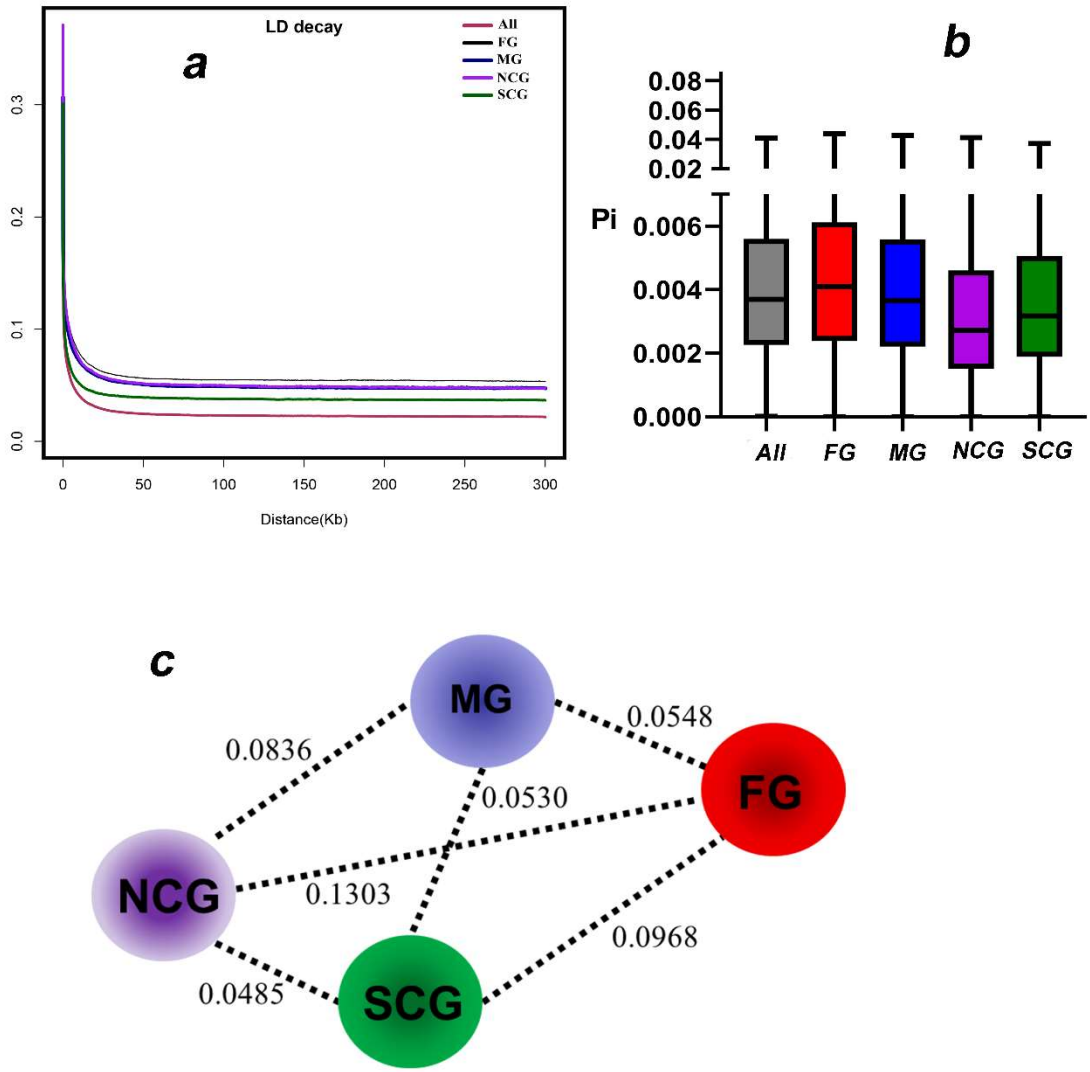
137 *2.3 Population structure analysis*

138 Population genetic structure analysis was performed based on high-quality SNPs.

139 We employed 5-fold cross validation to inference the number of ancestor population,

140 K (Figure 2b). The NCG was divergent from SCG when the K >2, suggesting the

141 NCG was derived from SCG. We observed that SCG includes individuals with similar
142 or different genetic structure to these from other groups like NCG and FG, further
143 indicating that it has higher diversity than others. The observation was further
144 supported by genetic diversity analysis with π (Figure 3b). Thus, other groups
145 including FG and NCG potentially associated with SCG during plum evolution.



146

147 **Figure 3**. The genetic diversity of different groups. a) The decay of linkage
148 disequilibrium (LD) measured as the squared correlation coefficient (r^2) by pairwise
149 physical distance; b) The nucleotide diversity (π) of different groups; c) The genetic
150 differentiation analysis between groups. The values between pairs indicate population

divergence (F_{ST}). FG, the foreign cultivar group; MG, the mixed cultivar group; SCG, the southern cultivar group; NCG, the northern cultivar group.

2.4 Principal component analysis

To further confirm the relationship among the cultivars, we performed principal component analysis (PCA) toward the 67 plum accessions (Figure 2c). Based on the principal component plot with first two principal components, the NCG exhibited a relatively close relationship with SCG, which was consistent with the phylogenetic tree and population structure analysis. We deduced that the NCG could be considered as a more independent subgroup of SCG and may be derived from a specific ecotype. Furthermore, samples from SCG showed some degree of correlation, suggesting the close relationship between SCG and others. The results further confirm that other groups potentially originate from SCG.

2.5 Genetic diversity analysis

As shown in Table 2, the expected heterozygosity (He) of the *Prunus* populations varied between 0.234 and 0.304; the observed heterozygosity (Ho) of the *Prunus* populations ranged between 0.328 and 0.429; Wright's F-statistic (F_{IS}) of the *Prunus* populations varied between –0.241 and –0.146; the number of private alleles (A_P) in the *Prunus* populations varied between 546 and 31,285; the nucleotide diversity (π) ranged between 0.00358 and 0.00467. The mean nucleotide variation of *P. salicina* was higher than that of other perennial crops, such as peach (π = 0.0015) [24], cassava (π = 0.0026) [25], and apricot (π = 0.0027) [26], but lower than that of date palm (π = 0.0092) [27]. The Tajima's D values of the four groups were all tested positive (1.002

173 – 1.497) and significantly different from zero. Thus, the null hypothesis of neutral
174 evolution could be rejected. As shown in Figure 3a, the LD decay rate was fastest in
175 the SCG and was lowest in the FG, suggesting a lower frequency of genetic
176 recombination in the FG, as they included artificially hybridized cultivars.

177 **Table 2**.The statistical values of genetic diversity within different populations.

Groups	Private allele number (Ap)	Observed heterozygosity (Ho)	Expected heterozygosity (He)	Nucleotide diversity (π)	Wright's F-statistic (F_{IS})	Tajima's D
FG	31,285	0.4290	0.3036	0.00467	-0.2411	1.4974
MG	30,112	0.3658	0.2846	0.00433	-0.1560	1.0094
SCG	27,287	0.3365	0.2606	0.00395	-0.1460	1.0022
NCG	546	0.3280	0.2336	0.00358	-0.1829	1.3211

178 *2.6 Genetic differentiation analysis*

179 The F_{ST} between the four groups varied between 0.0485 and 0.1303 (Fig. 3). The F_{ST}
180 value of MG-FG, MG-SCG, and SCG-NCG pairs was relatively lower among all the
181 group-pairs analyzed, indicating that the genetic differences within populations were
182 higher than those between populations and the possible genetic exchange between
183 populations. The F_{ST} value between the NCG and FG groups were the largest among
184 all the groups analyzed, which may have been due to geographical isolation, low gene
185 flow between populations, and significant genetic differences.

186 *2.7 Gene flow analysis and inferred evolutionary path*

187 We analyzed the gene flow between the four geographic groups (Figure 4b, 4c).
188 Allowing one or two migration events (m = 1 or 2), we observed that gene flow
189 occurred between the SCG and FG accessions, which likely reflected their many
190 shared genomic components due to hybridization in their domestication and breeding
191 histories. Considering the geographical locations of SCG and NCG, though these two

ecological groups were close to each other, we found no gene flow between them, indicating a relatively independent domestication process.

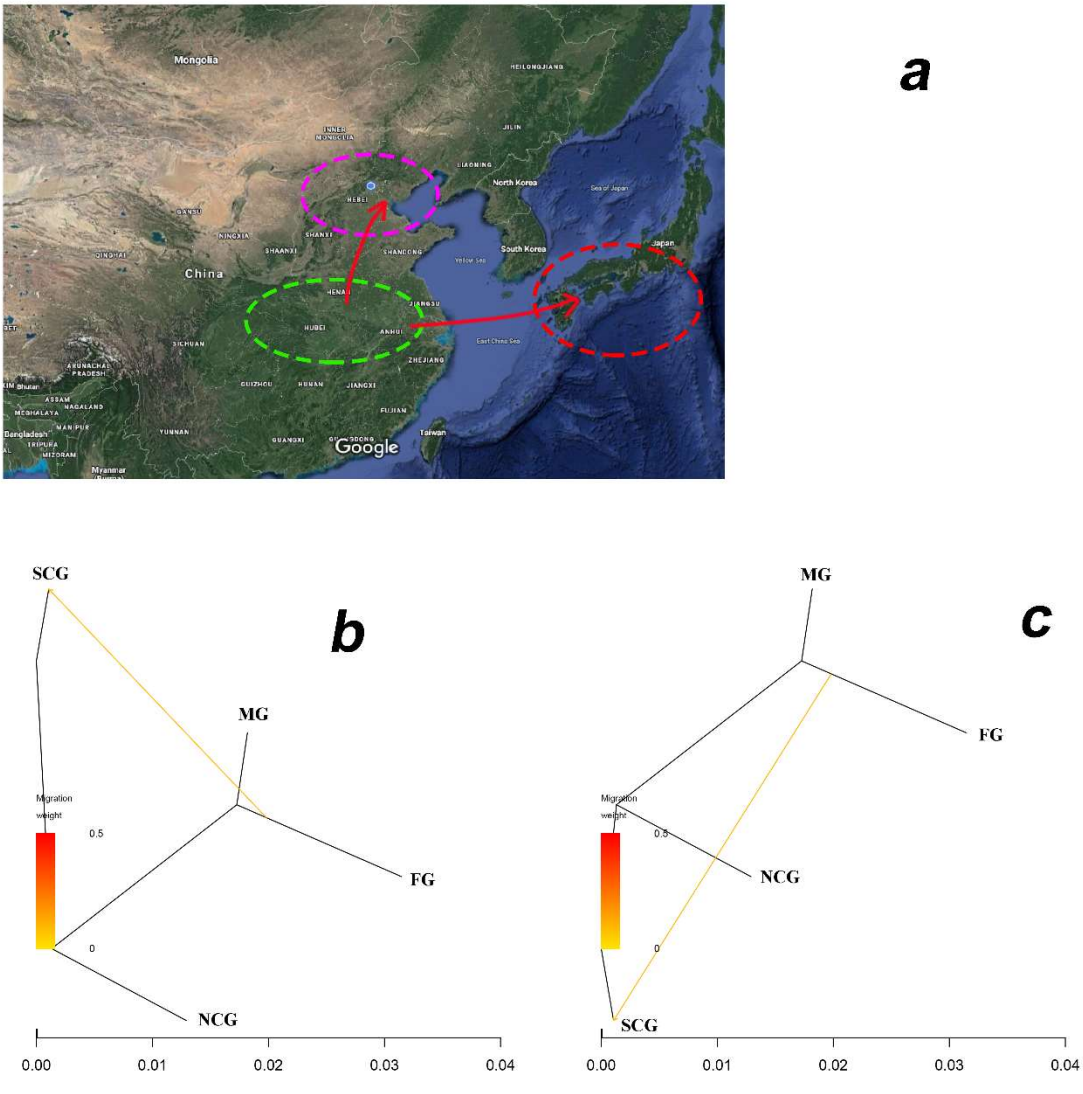


Figure 4. The inferred evolutionary path (a) and gene flow analysis between populations as inferred by TreeMix using a model with one (b) and two (c) admixture events. Admixtures are colored according to their weight. FG, the foreign cultivar group; MG, the mixed cultivar group; SCG, the southern cultivar group; NCG, the northern cultivar group.

3. Discussion

202 SNPs have become tremendously important as markers for genetics research in plants
203 because they have been found to be in high frequency, display a lower mutation rate
204 compared to SSR-based markers, and they are uniformly distributed across the
205 genome [28]. NGS is a kind of low-cost technologies to discover large numbers of
206 SNPs for extensive genetic studies, including within-species diversity analysis,
207 linkage map construction, and genome-wide association studies (GWAS), which is
208 allowing significant advances in plant genetics and breeding [6,22,23,29]. The genus
209 *Prunus* has shown conserved intraspecific and intragenic collinearity in the *Rosaceae*
210 family, and peach has been considered as a model species for the genus *Prunus*
211 regarding multiple genetic types of research [30-33]. On these bases, for the first time,
212 we conducted high-depth (~20×) whole-genome resequencing of 67 plum accessions
213 using the peach genome sequence as a reference, obtaining an average of 6.9 Gb
214 high-quality data for each acquisition, which ensured the accurate and comprehensive
215 population genetic analysis.
216 Based on the high-quality SNPs, the 67 plum accessions in this study were divided
217 into four groups, the SCG, NCG, FG, and MG, supported by the phylogenetic tree, the
218 population structure analysis, and the PCA results, which were partly in accordance
219 with the previously obtained results through RAPD, ISSR, and SSR marker assessing
220 [18-20]. We noticed that several cultivars from the south part of China distributed in
221 other three groups (Figure 2a), combining with the fact that the *P. salicina* system in
222 South China was rather primitive than that in North China basing on a palynology
223 study, we speculated that, for SCG, the genetic background was relatively broader, the

224 gene exchange was more frequent with other groups, and the possible origin center of
225 Chinese plum was at the Yangtze River Basin in South China [3,20,21,34]. For the
226 five newly bred cultivars from the NGRPA in MG, one of their parents was from the
227 USA (‘Blackamber’, ‘Friar’) or Japan (‘Akihime’) and the other parent was native to
228 China, and the gene flow between SCG and FG was detected (Figure 4). Thus, we
229 assumed that those cultivars in MG were genetically related. As mentioned in other
230 studies, most of the Chinese plum cultivars from Japan and the improved Chinese
231 plum hybrids from the USA were found distributed across Chinese indigenous plum
232 group through dendrogram analysis [2,5,20], which was according with the *P.*
233 *salicina*-predominant genetic components of improved Chinese plum hybrids [35],
234 and in agreement with the fact that China-originated plum cultivars were initially
235 introduced to Japan, and then imported to the USA in 1870 from Japan by Luther
236 Burbank [3].
237 Southwest China was speculated as a primitive domestication center of plum as there
238 were wild plum populations discovered in Yunnan, Sichuan, and Guizhou Province
239 [20]. Suggesting by the high Shannon index of diversity, the high value of average
240 sufficient allele, and the high expected heterozygosity, the Chinese plums cultivated in
241 southwest China were tested with high genetic diversity using SSR markers [20].
242 Additionally, by analyzing the phenotyping variation of 405 plum accessions
243 distributed in different geographical regions in China, the fruit weight of cultivars in
244 South China exhibited the most significant coefficient of variation [6]. The π, Ho, and
245 He of the SCG calculated with SNPs in our study were higher than those of the NCG,

corresponding with the higher genetic diversity of the SCG; these genetic diversity indices of the SCG were lower than that of the FG and MG, presumably due to the artificial hybridized cultivars in the FG and MG.

Generally, the genetic differentiation between populations is considered moderate when the F_{ST} value is higher than 0.05 and is considered highly differentiated when the F_{ST} value is higher than 0.15 [36]. In our study, the NCG-FG pair possessed the highest F_{ST} value, following by the SCG-FG couple. In contrast, the F_{ST} value of the MG-FG was significantly lower as there were five newly selected plum cultivars by NGRPA in the MG with foreign lineage from the FG. Human intervention via the artificial selection of favorable phenotypic traits to enhance production and improve desirable agronomic traits can both reduce the levels of genetic variability and skew allele frequencies [37].

4. Materials and Methods

4.1 Plant materials

A diverse collection consisting of 67 *Prunus* spp. accessions, including 65 *P. salicina* accessions and two *P. simonii* accessions, were selected for the whole-genome resequencing study (Table S1). All these trees were held at NGRPA located in Xiongyue county, Liaoning Province, China (40°18′ N, 122°16′ E) in a planting density of 3.0 × 4.0 m, trained to the open vase system with three or four main branches, and maintained under conventional management and pest control operations.

268 *4.2 DNA extraction, library preparation, and sequencing*

269 Genomic DNAs were isolated from the young and healthy leaf samples of the 67
270 accessions using a modified CTAB protocol [38]. The quality and integrity of the
271 DNAs were examined using a NanoDrop® spectrophotometer (ND-1000, Thermo
272 Fisher Scientific Inc., USA) followed by electrophoresis on 1% agarose gels.
273 Quantification of the DNA samples was assessed with Qubit™ (2.0 Fluorometer,
274 Invitrogen, Carlsbad, CA, USA).

275 High molecular weight DNA aliquots with 230/260 and 260/280 ratios ranging
276 between 1.8 – 2.0 and 1.8 – 2.2, respectively, were then sent to BGI (Shenzhen, China)
277 for library construction and sequencing. The insert-size of the libraries was 500 bp,
278 and the length of pair-end reads was 150 bp. All the libraries were sequenced using
279 the Illumina HiSeq 2500 platform (Illumina, San Diego, CA, USA).

280 *4.3 Reads mapping, SNP calling, and SNP annotation*

281 The qualified paired-end reads of each accession were aligned against the peach
282 reference genome v2.0 [24] using BWA v0.7.12-r1039 with the parameters mem -t 4
283 -k 32 –M [39], and SNPs were identified using SAMtools v1.4 [40]. Low-quality
284 SNPs were filtered out by minimum minor allele frequency (mnMAF < 0.01) and
285 missing data per site (MDpS > 10%), and finally converted into Variant Call Format
286 file (VCF). Gene-based SNP annotation was performed using the ANNOVAR
287 package v2018-04-16 [41]. Based on the reference genome annotation, SNPs were
288 categorized as occurring in exonic regions (overlapping with a coding exon), intronic
289 regions (overlapping with an intron), upstream and downstream regions (within a 1 kb

290 region upstream or downstream from the transcription start site), or intergenic regions.
291 Additionally, the heterozygosity was calculated using VCFtools v0.1.14 [42].

292 *4.4 Population structure analysis*

293 TreeBeST v1.9.2 software was used to calculate the distance matrix [43]. RAxML
294 v8.2.12 was used to construct the ML phylogenetic tree, and 1000 bootstrap replicates
295 were used [44]. The obtained phylogenetic tree was visualized using MEGA v5.05
296 [45].

297 The principal component analysis was performed using the PLINK v1.07 software
298 with default parameters [46].

299 We used ADMIXTURE v1.23 to infer the population structure [47]. To identify the
300 best genetic clusters *K*, cross-validation error was tested for each *K* value from 2 to 10.
301 The termination criterion was 1e-6 (stopping when the log-likelihood increased by
302 less than 1e-6 between iterations).

303 *4.5 Linkage disequilibrium (LD) analysis*

304 Linkage disequilibrium (LD) was calculated using SNPs with MAF greater than 0.05
305 through PLINK v1.07 software with the following setting: --file --r2 --ld-window
306 99999 --ld-window-kb 200 --out. LD decay was calculated based on the squared
307 correlation coefficient (r^2) values between two SNPs and the physical distance
308 between the two SNPs [46].

309 *4.6 Gene flow analysis*

310 The TreeMix v1.13 software was used to evaluate the gene flow among different
311 groups with the parameters -se-bootstrap-k 1000 -m, where the number (-m) varied

312 from one to three [48].

313 *4.7 Genetic diversity and differentiation analysis*

314 To evaluate the genetic diversity and differentiation, we used a 100-kb sliding window
315 with a step size of 10 kb to calculated the number of private alleles (A_P), the observed
316 heterozygosity (Ho), the expected heterozygosity (He), the nucleotide diversity (π),
317 the Tajima's D value, Wright's F-statistic (F_{IS}), and the population-differentiation
318 statistics (Fixation index, F_{ST}) using VCFtools v0.1.14 [42].

319

320 **5. Conclusions**

321 In the current study, using high-quality SNPs generated from high-depth
322 whole-genome resequencing of 67 *Prunus* accessions, we concluded that the origin
323 center of Chinese plum was at the Yangtze River Basin in South China. This study
324 also provided genetic variation features of plum cultivars and dissected their genetic
325 diversity and population structure, laying a foundation for breeders to use diverse
326 germplasm and allelic variants towards developing improved Chinese plum varieties.

327

328 **Data availability:** All of the raw reads of the plum accessions generated in this study
329 have been deposited in the public database of National Center of Biotechnology
330 Information under PRJNA659814.

331 **Supplementary Materials**: Supplementary material can be found at http://www.

332 **Authors' contributions:** ZQP and LWS conceived and designed the experiments.
333 WX and SF performed the experiments. WX, SF and ZQP analyzed the data and

wrote the paper. LN, ZYP, XM, LS, MXX and ZYJ collected samples and managed the materials. LWS revised the manuscript. All authors have read and approved the final manuscript.

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Conflict of interest: The authors declare no conflict of interest.

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