

A Novel Factor in Olfactory Ensheathing Cell-Astrocyte Crosstalk: Anti-Inflammatory Protein α -Crystallin B

Aybike Saglam^{1,2*}, Anne L. Calof^{3#} and Susan Wray^{1†‡}

¹Cellular & Developmental Neurobiology Section, NINDS, NIH, Bethesda, MD

²Program in Neuroscience & Cognitive Science, Univ. of Maryland, College Park

³Dept of Anatomy & Neurobiology and the Center for Complex Biological Systems, Univ. of California, Irvine, California

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*E-mail: asaglam@umd.edu

#Email: alcalof@uci.edu

†Corresponding author.

‡E-mail: wrays@ninds.nih.gov

Main Points:

- Astrocytes uptake OEC-secreted exosomes.
- WT OEC-exosomes, but not CryAB-null OEC-exosomes, block nuclear NF κ B translocation in astrocytes.
- CryAB, and other factors secreted by OECs, suppresses multiple neurotoxicity-associated astrocyte transcripts.

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Abstract

Astrocytes are key players in CNS neuroinflammation and neuroregeneration that may help or hinder recovery, depending on the context of the injury. Although pro-inflammatory factors that promote astrocyte-mediated neurotoxicity have been shown to be secreted by reactive microglia, anti-inflammatory factors that suppress astrocyte activation are not well-characterized. Olfactory ensheathing cells (OECs), glial cells that wrap axons of olfactory sensory neurons, have been shown to moderate astrocyte reactivity, creating an environment conducive to regeneration. Similarly, astrocytes cultured in medium conditioned by cultured OECs (OEC-CM) show reduced nuclear translocation of Nuclear Factor kappa-B (NF κ B), a pro-inflammatory protein that induces neurotoxic reactivity in astrocytes. In this study, we screened primary and immortalized OEC lines to identify these factors and discovered that Alpha B-crystallin (CryAB), an anti-inflammatory protein, is secreted by OECs via exosomes, coordinating an intercellular immune response. Our results showed: 1) OEC exosomes block nuclear NF κ B translocation in astrocytes while exosomes from *CryAB*-null OECs could not; 2) OEC exosomes could be taken up by astrocytes and 3) CryAB treatment suppressed multiple neurotoxicity-associated astrocyte transcripts. Our results indicate that OEC-secreted factors are potential agents that can ameliorate, or even reverse, the growth-inhibitory environment created by neurotoxic reactive astrocytes following CNS injuries.

1 Introduction

Damage to the central nervous system (CNS) provokes morphological and molecular changes in astrocytes, causing them to become ‘reactive astrocytes’ (Liddelow & Barres, 2017). These reactive cells play positive roles during CNS injury, such as confining inflammation by surrounding the damaged tissue and creating a barrier between it and uninjured tissues (Silver, *et al.*, 2015). Reactive astrocytes have been traditionally characterized by increased expression of intermediate filament proteins such as GFAP (Glial Fibrillary Acidic Protein), vimentin, and nestin (summarized in Liddelow & Barres, 2017). Excessive or sustained astrocyte reactivity is characterized by activation of pro-inflammatory pathways such as the NF κ B pathway (Liddelow & Barres., 2017; Wheeler 2020). This activity can be deleterious to functional recovery, since it can lead to chronic inflammation and neurotoxicity (Sofroniew, *et al.*, 2010). A better understanding of the molecular mechanisms that govern astrocyte reactivity would therefore be helpful to create environments conducive to regeneration following CNS injury.

The mammalian olfactory system shows robust neurogenesis throughout life. Data suggest that both neural niche signals and the surrounding glia, including olfactory ensheathing cells (OECs), give the olfactory mucosa this unique capability (Li, *et al.*, 2005; Roet & Verhaagen, 2014). Several groups have transplanted OECs into CNS injury sites, and observed improved axonal regeneration (Li *et al.*, 1997; Imaizumi *et al.*, 2000), functional recovery (Johansson *et al.*, 2005), reduced astrocytic scar tissue (Ramer *et al.*, 2004), and an attenuated hostile astrocyte response (Lakatos *et al.*, 2003; summarized in Roet & Verhaagen, 2014). Moreover, factors secreted by OECs have been shown to moderate astrocyte reactivity, at least insofar as their presence results in reduction of GFAP expression and nuclear translocation of NF κ B (Chuah *et al.*, 2011; O'Toole *et al.*, 2007). Identification of molecules secreted by OECs, which specifically affect astrocyte reactivity, should lead not only to a better understanding of the crosstalk between astrocytes and OECs; it may also reveal mechanisms that can block the metamorphosis of astrocytes into neurotoxic cells.

To identify such molecules, this study used lipopolysaccharide (LPS)-treated astrocytes, a model for neurotoxic reactive astrocytes, and assessed conditioned medium (CM) from immortalized clonal mouse OEC cell lines (Calof & Guevara, 1993). Nuclear NF κ B translocation in astrocytes was measured to determine if CM from these cell lines could mimic primary OEC-CM, which blocked the LPS pro-inflammatory response in astrocytes. Two immortalized cell lines were chosen for further study: one whose CM mimicked the effect of primary OECs (positive control); and a second, whose CM did not block the LPS response in astrocytes (negative control). These two cell lines and primary OECs were challenged with LPS, and the conditioned media screened by mass spectrometry. Using this strategy, the heat-shock protein

CryAB (D'Agostino *et al.*, 2013) was identified. Subsequent experiments showed that: 1) CryAB is secreted by OECs via exosomes; 2) exogenous CryAB suppressed LPS-induced astrocyte reactivity; 3) exosomes containing CryAB are taken up by astrocytes; and 4) unlike wildtype (WT) OEC-exosomes, *CryAB*-null (*CryAB*^{-/-}) OEC exosomes fail to suppress LPS-induced astrocyte reactivity measured by nuclear NFκB translocation. Finally, examination of transcripts that are associated with neurotoxic-reactive astrocytes (Liddelow *et al.*, 2017) revealed that either exogenous CryAB or OEC-CM can suppress expression of several of these transcripts. Taken together, the data indicate that CryAB secreted by OECs, via exosomes, is an important factor for OEC-astrocyte crosstalk that can block astrocytes from becoming neurotoxic cells. Ultimately, mimicking appropriate astrocyte-OEC crosstalk *in vivo* may contribute to an environment conducive to regeneration following a broad range of CNS injuries.

2 Materials & Methods

2.1 Mice

All mice were maintained, and all animal handling procedures were performed according to protocols approved by the National Institutes of Health NINDS Institutional Animal Care and Use Committee. *CryAB*-null (*CryAB*^{Del(9Hspb2-Cryab)1Wavr}, henceforth referred to as *CryAB*^{-/-}) mice (Brady *et al.*, 2001) were obtained as homozygous sperm, revived by IVF using eggs from C57bl6/N mice (Jackson Laboratory), and resulting heterozygotes intercrossed to obtain *CryAB*^{-/-} and *CryAB*^{+/+} (wildtype, WT) lines, which were used as the source for OEC primary cultures (see below). Mice were genotyped using a three-primer PCR protocol: 5'-TAGCTTAATAATCTGGGCCA-3', 5'-GGAGTTCACAGGAAGT-ACC-3', and 5'-TGGAAGGATTGGAG-CTACGG-3' primers were used in 4:1:1 molar ratio. Amplification was performed for 40 cycles at 94°C for 15 sec, 62°C for 30 sec and 72°C at 1 min. PCR produced a 310-bp product for the WT allele and a 600-bp product for the null allele.

2.2 Cell culture and reagents

Primary cultures of OECs were generated as described previously (Dairaghi *et al.*, 2018). Briefly, olfactory bulbs of postnatal (PN) day 0-7 mice were collected and placed in an enzyme mix: 300µg/ml hyaluronidase (Sigma, Cat# H3631, St. Louis, MO), 30U/ml dispase I (Sigma, Cat# D4818), 1.2 mg/ml collagenase type 4 (Worthington, Cat# 43E14231, Lakewood, NJ), 10U/ml DNase I (Worthington, Cat# 54E7315); for 35 min at 37°C with constant agitation (Au & Roskams, 2003). Cells were run through a 40µm cell strainer to remove non-dissociated tissue pieces and then washed with DMEM-F12 medium. Subsequently, cells were purified by the differential cell adhesion method (Nash *et al.*, 2001), which consists of three steps: 1) Cells were seeded into uncoated T75 flasks (4x10⁶ viable cells/flask, VWR, Cat# 734-2788, Radnor, PA) for 18 hrs to remove fibroblasts; 2) the supernatant of the first step was seeded into another uncoated flask for up to 36 hrs to remove astrocytes; and finally 3) the supernatant of the second flask was seeded onto poly-L-lysine (Sigma, Cat# P4707)-coated flasks to grow primary OECs. Cells were cultured for up to 2 weeks and medium was changed every 2–3 days. OECs constituted more than 90% of the cells in the culture based on p75 and S100B immunostaining (data not shown). For OECs to be co-cultured with primary astrocytes, the medium was gradually changed to serum-free medium (Klenke & Taylor-Burds, 2012) supplemented with 5ng/ml HB-EGF (PeproTech, Cat# 100-47, Rocky Hill, NJ), and B27 (Thermo Fisher Scientific, Cat# A3582901) to provide a medium compatible with astrocyte culture, since serum has been shown to induce astrocyte reactivity (Foo *et al.*, 2011).

Primary astrocyte cultures were obtained by magnetic sorting as previously described (Holt *et al.*, 2019), with some modifications. Briefly, 10-20 cortices of PN day 2-4 pups were dissociated using the MACS Neural

Tissue Dissociation Kit-T (Miltenyi Biotec, Cat# 130-093-231, Auburn, CA) at 37°C (5% CO₂, 30 min). Non-dissociated tissue was removed using a 40µm cell strainer (Fisher Scientific, Cat# 22-363-547), and the remaining cell solution was centrifuged (300g, 5 min). Next, a discontinuous density gradient, prepared using 1:1 albumin-ovomuroid solution (10mg/ml of each) (Worthington, Cat# OI; GeminiBio, Cat# 700-102P, West Sacramento, CA), was used to remove cell debris and inhibit enzyme activity. The cell pellet was resuspended in 80µl Hank's Balanced Salt Solution (HBSS) (Gibco, Cat# 14025-092) plus 20µl anti-GLAST (ACSA-1) MicroBeads (Miltenyi Biotec, Cat# 130-095-825, Auburn, CA) for up to 10⁷ cells, and incubated for 10 min (4°C). Cells were washed and incubated in 90µl HBSS plus 10µl anti-Biotin MicroBeads for another 15 min (4°C) before running through MACS column for positive selection of astrocytes. Cells were cultured for one week and then the same procedure was followed with anti-Prominin-1 MicroBeads (Miltenyi Biotec, Cat# 130-092-564) for the negative selection of radial glia, followed by another positive selection with anti-GLAST antibody the same day, to increase the purity of astrocyte cultures. Sorted cells were cultured in T25 flasks coated with poly-L-lysine, in 5ml serum-free astrocyte culture medium (ACM, described above). In our hands, astrocytes isolated by this method and cultured in ACM were not reactive when stained with NFκB (not shown). The same method was adjusted to obtain oligodendrocyte cultures using anti-O4-MicroBeads (Miltenyi Biotec, Cat# 130-096-670). The immortalized mouse astrocyte line C8D30 (ATCC, VA, USA) was cultured in DMEM-F12 (Gibco, Cat# 10313-02, 11765-054, Long Island, NY) containing 10% Fetal Bovine Serum (FBS) (Gibco, Cat# 10438-026), plus 0.5% antibiotic-antimycotic (Gibco, Cat# 15240-062) at 37°C in 5%CO₂.

2.3 Immortalized OEmyc790 Cell Lines

Six immortalized OEC lines (OEmyc790-C7s.D, D6s.AB8, C6s.BG9, D10 and D4), derived from retrovirus-mediated transformation of primary embryonic mouse olfactory epithelium cultures derived from E15 mouse embryos (Calof & Guevara, 1993), were analyzed; two lines, OEmyc790-C7s.D (C7) and OEmyc790-C4 (C4), were used for the studies described below. Cells were plated on cell culture plates (Fisher Scientific, Cat# 130190, Waltham, MA) and cultured in DMEM-F12 as described above. Medium was changed every 3-5 days. When 60% confluent, a 1:4 dilution of trypsin was used (Gibco, Cat# 15400054) to split the cells into thirds.

2.4 Primary antibodies and recombinant proteins

The following antibodies were used: CryAB rabbit polyclonal antibody (Millipore, Cat# ABN185, Darmstadt, Germany, 1:1K for WB, 1:4K for immunofluorescence (IF); Histone mouse monoclonal antibody (Fisher Scientific, Cat# AHO1432, Waltham, MA, 1:200 for WB); NFκB rabbit polyclonal antibody (C-20, Santa Cruz, Cat# sc-372, Santa Cruz, CA, 1:650 for WB, 1:750 for IF); Sox10 goat polyclonal antibody

(N-20, Santa Cruz, Cat# sc-17342, 1:300 for IF); Alix mouse monoclonal antibody (3A9, Cell Signaling, Cat# 2171T, 1:1K for WB); GFAP chicken polyclonal antibody (Aves, 1:4K for IF); Flotillin-1 rabbit polyclonal antibody (D2V7J), Cell Signaling, Cat# 18634, 1:1K for WB); β -actin mouse monoclonal antibody (AC-74, Millipore, Cat# A2228, 1:1K for WB); Tomm20 rabbit polyclonal antibody (FL-145, Santa Cruz, Cat# sc-11415, 1:1K for WB); CD63 biotinylated antibody (Miltenyi Biotec, Cat# 130-108-922, Auburn, CA, 1:15 for IF); BLBP mouse monoclonal antibody (Abcam, Cat# ab131137, 1:2K for IF) and p75-NGFR rabbit polyclonal antibody (Millipore, Cat# AB1554, 1:5K). Recombinant chicken Anosmin1 (MyBioSource, Cat# MBS963562-COA, San Diego, CA) was used at 5nM while recombinant CryAB protein (MyBioSource, Cat# MBS964495) and recombinant myoglobin (MyBioSource, Cat# MBS142891) were used at 50ng/ml unless stated otherwise.

2.5 Mass spectrometry

Primary OECs, and immortalized C7 and C4 OEC cell lines, were established by seeding them in T75 flasks at a concentration of 8×10^5 cells/flask in regular growth medium. To concentrate secreted proteins, the cells in each flask were rinsed and media replaced with 10ml/flask of Earle's Balanced Salt Solution (EBSS) with 5.5mM D-Glucose (Gibco, Cat# 14155-063); conditioned medium (CM) was collected after 48 hrs total of incubation. For the last 2 hrs of the 48-hr collection period, either $1 \mu\text{l/ml}$ LPS (Sigma, Cat# L6529) or 5nM recombinant chicken Anosmin1 was added. CM was then collected, centrifuged to remove debris, and frozen at -80°C . Frozen samples were freeze-dried using a lyophilizer (Novalyph-NL150, Savant Instruments, Holbrook, NY). The pellets were reconstituted in water and bicinchoninic acid (BCA) protein assay was performed. $200 \mu\text{g}/60 \mu\text{l}$ protein per group was submitted for mass spectrometry analysis (NINDS Protein Facility, NIH). Each sample was digested with trypsin. Tandem Mass Tag (TMT) labeled samples were mixed together (TMT 126-131). The mixture was separated using hydrophilic interaction liquid chromatography (HILIC) high performance liquid chromatography (HPLC) system. Five HILIC fractions were collected from the mixed sample. One liquid chromatography-tandem mass spectrometry (LC/MS/MS) experiment was performed for each HILIC fraction, using an Orbitrap Fusion Lumos Mass Spectrometer (Thermo Fisher Scientific, Waltham, MA) coupled to a 3000 Ultimate high-pressure liquid chromatography instrument (Thermo Fisher Scientific). Proteome Discoverer 2.2 software used for database search and TMT quantification, and data were mapped against the Sprout mouse database. "Primary OEC+LPS-CM" was used as reference to calculate the ratio for LPS treated samples; "Primary OEC+5nMA1-CM" was used as reference to calculate the ratio for 5nMA1-treated samples. No normalization was performed. See Supplemental Data 1 and 2 for obtained values.

2.6 Immunoblot analysis

As a readout of reactivity, quantitative immunoblot analysis was performed on nuclear fractions of immortalized C8D30 astrocytes treated with 1 μ l/ml LPS or vehicle control for 2 hrs. For co-culture groups, OECs seeded on porous inserts (0.4 μ m Millicell Cell Culture Insert, Millipore, Cat# PICM0RG50) were placed on top of astrocytes for 24 hrs and were discarded at the end of the incubation period, so that only astrocytes were collected for subsequent protein analysis. For the CM treated groups, CM from each line was collected (after 24 hr incubation) and then added to astrocytes for 22 hrs, followed by a 2-hour LPS treatment. Astrocytes were then collected by scraping and the CNMCS Compartmental Protein Extraction Kit (BioChain Cat# K3013010 Hayward, CA) with protease/phosphatase inhibitors (PI, Cell Signaling, Cat# 5872S, Danvers, MA) was used and the nuclear fractions were isolated for each treatment condition. The fractions were run on BioRad Mini-Protean TGX Stain-Free Gels (Cat#4568084), transferred to PVDF stain-free blot (Trans-Blot Turbo Transfer Pack, Cat#1704156) via the Trans-Blot Turbo transfer system (BioRad), and blocked with 5% dry milk (BioRad, Cat #170-6404) prior to staining with NF κ B antibody. Membranes were exposed to Clarity enhanced chemiluminescence (ECL) reagent (Cat. # 170-5061, Bio-Rad) for 5 min and the signal was detected using ChemiDoc MP (Cat. # 170-8280, Bio-Rad). Quantification of band intensities was calculated using Image Lab 5.0 software (Bio-Rad) and normalized by the loading control immunostained for Histone on the same sample and the same blot. Three biological replicates were used for statistical analysis.

2.7 Quantitative immunofluorescence

Fluorescent immunostaining for nuclear NF κ B and cytoplasmic NF κ B was quantified in immortalized C8D30 astrocytes following 2-hr treatment with 1 μ g/ml LPS or a cocktail of 3 cytokines: IL-1 α (3ng/ml, Sigma, Cat# I3901), TNF α (30ng/ml, Cell Signaling, Cat# 8902SF) and C1q (400ng/ml, MyBioSource, Cat# MBS143105, San Diego, CA), as follows: After immunofluorescence staining for NF κ B, confocal images were taken on a Zeiss LSM 800 Confocal Microscope (Carl Zeiss, Thornwood, NY). A defined area was measured in both nuclear and cytoplasmic compartments for each astrocyte, and the fluorescence intensity was quantified for each area in each cell using Imaris software. The ratio of nuclear to cytoplasmic fluorescence intensity was used as a quantitative readout of astrocyte reactivity. Median values were calculated for each biological replicate (N=3) obtained from multiple images (2-3/well) containing a total of ~100 cells/treatment (Figure 1C, cell numbers in Supplemental Table 1) or ~50 cells/treatment (Figure 2B, cell numbers in Supplemental Table 2). Values greater than one indicate that the NF κ B value was higher in the nucleus compared to cytoplasm, and cells were reactive. Statistics (ANOVA) were performed and the average median value \pm standard deviation (SD) per treatment plotted.

2.8 Isolation of exosomes and exosome uptake experiments

For isolation of exosomes, the protocol of Adolf and colleagues (2018) was used with slight modifications. Briefly, 24 hr prior to exosome collection, cells were washed and medium changed to exosome depleted medium (EDM) containing 10% exosome-depleted FBS (Gibco, Cat# A27208-03). CM was collected, (protease inhibitor (PI) was added immediately for immunoblotting) and samples kept at 4°C until exosome isolation. Exosomes were isolated through three centrifugation steps: 1) CM was spun for 10 min at 2,000g to remove debris; 2) the resulting supernatant was centrifuged for 30 min at 10,000g to pellet microvesicles; and 3) this second supernatant was centrifuged for 4 hrs at 100,000g (Optima MAX-XP ultracentrifuge, TLA-100.3 rotor, Beckman Coulter). Following ultracentrifugation, pelleted exosomes were re-suspended in buffer (for ELISA and immunoblotting) or cell culture medium, as required. For astrocyte uptake experiments, isolated OEC-exosomes were resuspended by pipetting and added directly to the culture medium of *CryAB*^{-/-} astrocytes for 4 hrs. Cultures were then fixed with 4% paraformaldehyde and stained for markers of interest. Images were taken on a stimulated emission depletion (STED) confocal microscope (Leica, Wetzlar, Germany) for the visualization of internalized exosomes in astrocytes.

2.9 CryAB Immunoprecipitation

CryAB was immunoprecipitated (IP-CryAB) from isolated OEC-exosome fractions that were lysed in RIPA buffer. Briefly, 200µl Protein A Dynabeads (30mg/ml, Invitrogen, Cat# 10001D, Carlsbad, CA) were washed 3 times in PBS+ 0.1% Tween (PBST) using a magnetic stand (Millipore, PureProteome Cat# LSKMAGS08), CryAB antibody (400µl, 1:50 (10 µg/mL) in PBST) was added to the beads, and the mixture was incubated (30 min, RT) with constant agitation. The antibody solution was removed, beads washed (3x), exosome fractions resuspended in PBS were added, and the mixture was incubated overnight (4°C). Beads were then washed (4x) and CryAB protein eluted by addition of 60µl of 0.2M Glycine (pH 2.5); the pH of the eluate was neutralized by addition of 5µl of 1M Tris (pH 8.5). Cell culture, immunoblotting or ELISA was performed.

2.10 ELISA

CryAB concentration was measured in exosome fractions using a competitive ELISA kit (MyBioSource, Cat# MBS7239470, San Diego, CA) according to manufacturer's instructions. Isolated exosomes were sonicated and lysed in Buffer M (containing NP40) plus PI from Protein Extraction Kit (BioChain). Equivalent quantities of total exosomal protein or supernatant CM protein, determined by BCA protein assay, were brought to equivalent volumes in EDM. 100µl of samples were added to each well and measured with a microplate reader (FlexStation 3; Molecular Devices, Sunnyvale, CA). Results were analyzed with SoftMax Pro Software (Molecular Devices).

2.11 Quantitative RT-PCR (q-RT-PCR)

cDNA synthesis was performed using SuperscriptTM III reverse transcriptase (Invitrogen), and PCR carried out using the ViiA7 Real-Time PCR System (Applied Biosystems, Waltham, MA) in 20 μ l final volume, containing 10 μ l of SsoAdvanced Universal SYBR Green Supermix (BioRad Cat#1725271), 2 μ l of a primer mix with a concentration of 1 μ M of each primer and 1 μ l of cDNA and 7 μ l water. Samples were run in triplicate. The expression levels of genes of interest were normalized using the primers (forward; reverse) (AGTGCCAGCCTCGTCCCGTA; TGAGCCCTTCCACAATGCCA), for expression of GAPDH (see Supplemental Data 3 for obtained values). All other primer sequences are detailed in (Liddelow *et al.*, 2017; Clarke *et al.*, 2018). Data were analyzed by one-way ANOVA followed by Dunnett's multiple post hoc test.

2.12 Statistical analysis and cell counting

All statistical analyses were done using GraphPad Prism 8.00 software. The results are shown as mean \pm SD. Statistical analysis was performed using one-way or two-way ANOVA, unless otherwise stated. Probability values of 0.05 ($p \leq 0.05$) were considered to indicate statistical significance. N=biological replicates, n=technical replicates.

3 Results

3.1 OECs secrete anti-inflammatory factor(s) that reduce astrocyte reactivity

Nuclear translocation of the pro-inflammatory protein NF κ B was used as a readout of astrocyte reactivity evoked by bacterial endotoxin LPS (Rothhammer *et al.*, 2016), as measured by immunoblot analysis of the nuclear fraction of astrocyte lysates (Figure 1A, inset). As expected, NF κ B increased in the nuclear fraction of immortalized C8D30 astrocytes treated with LPS (Figure 1A, gray bars, $p \leq 0.05$). Co-culture of astrocytes with OECs blocked nuclear translocation of NF κ B in response to LPS, as previously reported (Hale *et al.*, 2011; Figure 1A, purple bars). Adding CM from untreated OEC monocultures, (OEC-CM), also decreased NF κ B translocation into nuclei of astrocytes exposed to LPS (Figure 1A, red bars), indicating that anti-inflammatory factor(s) are secreted by OECs even in the absence of a stress signal.

To facilitate identification of OEC factors of interest, the anti-inflammatory capacity of six immortalized OEC lines (Calof & Guevara, 1993) were screened. Immortalized astrocytes (C8D30) were treated with CM from the different OEC lines, treated with LPS, immunostained for NF κ B (Figure 1B), and the nuclear/cytoplasmic ratio of NF κ B immunostaining was determined. As shown in Figure 1C, CM from two of the immortalized OEC lines, C7 and D6, significantly reduced nuclear NF κ B translocation in C8D30 astrocytes compared to LPS treatment alone (Figure 1C, red bars, $p \leq 0.05$), while D4 and C4 CM were similar to LPS alone. Original characterization of these immortalized OEC lines had been based on morphology and immunostaining with markers expressed by primary OECs (Calof & Guevara, 1993). Characterization of the re-grown lines was consistent with earlier reports, with C7 cells, for example, showing heterogeneous morphologies depending on culture conditions and density (Figure 1D): these included Schwann Cell spindle-like (majority; Figure 1Da), astrocyte-like type1 (Figure 1Db), and astrocyte-like type2 (Figure 1Dc) morphologies (Huang *et al.*, 2008). Cell lines were re-examined by immunofluorescence for expression of OEC-specific markers, such as p75, Sox10, and brain lipid-binding protein (BLBP). Both C7 and C4 cell lines were positive for these OEC markers (Figure 1E). Since C7-CM significantly suppressed nuclear NF κ B translocation in astrocytes, whereas C4-CM did not (Figure 1B, C), and both lines expressed the OEC markers tested, C7-CM was used as a positive control, and C4-CM as a negative control in further experiments.

3.2 OEC-secreted CryAB suppresses LPS-induced astrocyte reactivity

To identify OEC-derived molecules potentially involved in crosstalk between OECs and astrocytes, secreted proteins from primary OEC-CM, C7-CM and C4-CM were compared by mass spectrometry. Before collection of CM, LPS was added to cultures as a stress signal. Secreted proteins from LPS-treated cells were ranked based on 1) their abundance in C7-CM compared to C4-CM; 2) their abundance in C7-CM compared to primary OEC-CM; and 3) absence from C4-CM (Figure 2A). Proteins that were secreted at similar levels

by C7 cells and primary OECs (Figure 2A, horizontal dashed line), but are not likely to be present in C4-CM (Figure 2A, X axis) were determined. Based on these criteria, we identified two proteins of particular interest: the heat shock protein alpha crystallin B chain (CryAB), and the cell surface glycoprotein MUC18 (MCAM). To identify OEC secreted molecules in response to an endogenous signal from astrocytes, similar experiments were performed after treatment with Anosmin 1, an extracellular binding protein secreted by mature astrocytes (Gianola *et al.*, 2009) and shown to act on OECs (Hu *et al.*, 2019). Even though the ortholog is yet to be identified for this protein in mice, we observed a robust migration of primary mouse OECs towards recombinant Anosmin-1 (personal observation). Notably, both CryAB and MCAM were identified as major secreted proteins in this screen as well (Figure 2B). CryAB was selected for further study because of its known role as an anti-inflammatory protein involved in stress responses by CNS glia (e.g., Ousman *et al.*, 2007; Kuipers *et al.*, 2017), and because it was the most abundant protein fitting our criteria in screens of both LPS (Fig. 2A) and Anosmin-1 (Fig. 2B) treated samples. Recombinant CryAB protein mimicked the effect of OEC-CM or C7-CM on astrocyte reactivity, as measured by suppressed nuclear translocation of NF κ B, following either LPS- or cytokine-induced inflammation (Figure 2C).

3.3 Exosomes secreted by OECs contain CryAB, which moderates intercellular immune response

Since it has been shown that CryAB secretion can occur via exosomes (Sreekumar *et al.*, 2010; Kore *et al.*, 2014; Guo *et al.*, 2019), exosomes were isolated from OECs to determine whether they were positive for CryAB and whether the CryAB secreted via OEC-exosomes had the ability to attenuate astrocyte reactivity. For these experiments, exosome fractions were isolated from culture supernatants of OECs generated from both *CryAB*^{-/-} mice and WT (*CryAB*^{+/+}) controls. To ensure the quality of fractions used, exosomes and whole cell lysates (CL) from WT OECs were analyzed by immunoblotting for the following proteins: the structural protein, β -actin; a mitochondrial protein, Tomm20; a nuclear protein, histone H3; and the extravesicular protein Flotilin-1. The exosome fraction was devoid of β -actin, Tomm20 and histone H3, but was positive for Flotilin-1 (Figure 3A), consistent with published information for exosome fractions (Jeppesen *et al.*, 2019). Competitive ELISA against CryAB confirmed the presence of CryAB in OEC exosomes. Exosomes derived from 1×10^6 OECs contained 9.22 ± 0.2 ng CryAB, whereas CryAB protein was undetectable in CM from which exosomes were depleted (Supplemental Figure 1A). Next, exosomes from both genotypes were immunoblotted for CryAB and the endocytosis protein, Alix, which is concentrated in exosomes (Figure 3B; Jeppesen *et al.*, 2019). CryAB was present in WT OECexo fractions but was absent in *CryAB*^{-/-}OECexo fractions; while the exosome marker Alix was present in exosome fractions from OECs of both genotypes. Finally, to assay whether CryAB present in OEC exosomes could suppress astrocyte reactivity, the exosomes were added to immortalized C8D30 astrocytes for 24 hrs, and treated with LPS for

the last 2 hours of this incubation. As shown in Figure 3C, quantitative immunoblotting demonstrated that: a) exosomes from WT OECs were able to suppress astrocyte reactivity, as measured by reduced nuclear translocation of NF κ B; b) astrocytes treated with *CryAB*^{-/-}OECexo remained reactive; and c) the reactivity of astrocytes treated with *CryAB*^{-/-}OECexo was reduced by the presence of recombinant CryAB protein. Immunostaining OEC-astrocyte co-cultures for NF κ B further showed strong nuclear NF κ B immunostaining exhibited by astrocytes co-cultured with *CryAB*^{-/-}OECs (Figure 3D, lower right), whereas astrocytes co-cultured with WT OECs showed little if any nuclear NF κ B immunostaining (Figure 3D, lower left). Together, these results are consistent with CryAB, secreted by OECs in exosomes being an important protein for OEC-astrocyte crosstalk, and functioning as an anti-inflammatory molecule for astrocytes.

3.4 Astrocytes internalize CryAB-containing OEC exosomes

To determine whether astrocytes take up CryAB-containing exosomes secreted by OECs, *CryAB*^{-/-} astrocytes were cultured with exosome fractions from OEC cultures generated from WT mice. Uptake was visualized by immunostaining of GFAP-positive astrocytes (Figure 4, magenta); colabeled with antibodies to endosome/exosome marker CD63 (red) and CryAB (green). CryAB and CD63 colocalized in *CryAB*^{-/-} astrocytes treated with exosomes for 4 hrs (Figure 4, insets). Neither untreated *CryAB*^{-/-} astrocytes (Figure 4B) nor WT astrocytes (Figure 4C) showed such specific colocalization, consistent with the uptake of CryAB-containing OEC exosomes by astrocytes.

3.5 OEC secreted factors, including CryAB, reduce astrocytes' expression of genes associated with neurotoxic reactivity

To evaluate the effects of OEC-secreted CryAB on expression of “neurotoxic” genes, astrocytes were exposed to LPS alone; or WT OEC-CM, *CryAB*^{-/-}OEC-CM, or CryAB immunoprecipitated from isolated OEC-exosome fractions (IP-CryAB) together with LPS. mRNA from treated astrocytes was then analyzed for 12 transcripts known to be associated with neurotoxic astrocyte reactivity (Liddelow *et al.*, 2017). Q-RT-PCR analysis (Figure 5) showed that all tested transcripts were reduced in expression in the presence of WT OEC-CM, and this effect was significant for 9 of the 12 (Figure 5, second row, white arrows, $p \leq 0.05$ A vs B). In contrast, 4 of the transcripts showed increased expression when treated with *CryAB*^{-/-}OEC-CM (Figure 5C, black arrows). The analysis also suggests that suppression of expression of *Ggta1*, *Serp11*, *ligp1*, *Gbp2* and *Amigo2* was CryAB-dependent, for the following reasons: a) suppression of expression failed to occur with *CryAB*^{-/-}OEC-CM treatment, while still taking place with IP-CryAB treatment (Figure 5, C vs D); or b) expression was upregulated in the *CryAB*^{-/-}OEC-CM group (Figure 5, C vs A). Suppression of expression of 4 genes (*H2-T23*, *Srgn*, *H2D1* and *C3*) appeared to be independent of CryAB, since it still occurred in

astrocytes treated with *CryAB*^{-/-}-OEC-CM (Figure 5, C vs A). In contrast to either OEC-CM treatments (*CryAB*^{-/-} or WT), a significant increase in *Fbln5* was detected in astrocytes treated with IP-CryAB (Figure 5, D vs A). These results are consistent with the finding that CryAB, secreted by OECs, functions as an anti-inflammatory agent for astrocytes. In addition, comparison of OEC-CM treatment to IP-CryAB for transcripts *Ugt1a1*, *C3* and *Fbln5* suggest that there are factors in OEC-CM, in addition to CryAB, that suppress neurotoxic astrocyte reactivity.

4. Discussion

4.1 OEC-secreted factors that moderate astrocyte reactivity

Astrocyte reactivity is a pathological response that occurs in a wide range of CNS injuries, inflammation and diseases. *In vivo* studies show that some reactive astrocytes induced by ischemia can promote neural recovery and repair (reviewed in Rossi *et al.*, 2007); in contrast, reactive astrocytes induced by bacterial endotoxins such as LPS are neurotoxic (Zamanian *et al.*, 2012). This harmful, neurotoxic astrocyte reactivity appears to be driven by pro-inflammatory cytokines secreted by activated microglia (Liddel *et al.*, 2017). However, anti-inflammatory factors that suppress neurotoxic astrocyte reactivity are largely unknown. Olfactory system is one of the few niches in the mammalian CNS that supports neuronal regeneration (Forni *et al.*, 2013). Olfactory sensory neurons are vulnerable to damage due to their exposed location in the nasal cavity and have a remarkable capacity for regeneration (Calof *et al.*, 1996, Forni *et al.*, 2013), suggesting the presence of robust anti-inflammatory factors in the olfactory system. OECs wrap and guide axons of the olfactory sensory neurons en route to the OB, where they establish new connections. Moreover, OECs directly interact with astrocytes at the entry point into the CNS, enabling regenerating olfactory axons to make new connections (Williams *et al.*, 2004; Li *et al.*, 2005; Raisman & Li, 2007). Notably, both transplanted OECs (Lakatos *et al.*, 2003; reviewed in Roet & Verhaagen, 2014) and co-cultured OECs (Hale *et al.*, 2011) have been shown to intermingle with astrocytes and to moderate astrocyte activation.

The studies in this report investigate anti-inflammatory factors secreted by OECs participating in OEC-astrocyte crosstalk. Mass spectrometry was used to analyze proteins in CM from primary OECs and compared to the CM of immortalized OEC lines with different anti-inflammatory capacities. Two proteins were identified as potential factors that could suppress neurotoxic astrocyte reactivity: MCAM and CryAB. MCAM (also called CD146 or MUC18) is a signaling receptor that can be cleaved from the cell membrane, generating a soluble form that is associated with increased cell migration and invasion (Seftalioglu & Karakoc, 2000), and primarily has been studied in endothelial cell angiogenesis and cancer metastasis (reviewed in Dye *et al.*, 2013). Studies on Multiple Sclerosis (MS) patients showed no association between MCAM expression and disease activity (Petersen *et al.*, 2019). In contrast, CryAB treatment of MS patients was associated with a therapeutic outcome, downregulation of T cell proliferation and pro-inflammatory cytokine production (Quach *et al.*, 2013; van Noort *et al.*, 2015). In addition, CryAB has been shown to have neuroprotective and regenerative effects in neuroinflammatory animal model systems (Arac *et al.*, 2011; Ousman *et al.*, 2007; van Noort *et al.*, 2015). Moreover, there is a correlation between glial activation and increased CryAB levels in Alexander, Alzheimer's and Parkinson's diseases, as well as traumatic brain injury and stroke (reviewed in Dulle & Fort, 2016). Thus, we investigated the role of OEC-secreted CryAB in OEC-astrocyte crosstalk.

4.2 Exosomal release of CryAB

Our results show that OECs secrete CryAB via exosomes, and that these exosomes produce an intercellular anti-inflammatory effect following their uptake by astrocytes. Exosomes are small vesicles packaged inside multivesicular endosomes (MVE) and released into the extracellular matrix when MVE fuse with the plasma membrane (Jeppesen *et al.*, 2019). Subsequent uptake by a neighboring cell initiates intercellular communication. Notably, released exosomes are functional components of the extracellular matrix that can be induced by stress signals but are associated with cell-cell communication rather than apoptosis (Jeppesen *et al.*, 2019; Gupta *et al.*, 2014). Consistent with our findings, recent studies suggest that CryAB can be secreted by glia in an autocrine manner (Kore *et al.*, 2014; Guo *et al.*, 2019) and play a protective role (Ousman *et al.*, 2007). Although the downstream effects of CryAB in astrocytes that we demonstrate have yet to be fully explored, one important role of CryAB may be its interaction with transcription factors, including NF κ B, to suppress inflammation by inhibition of their nuclear translocation (Shao *et al.*, 2013; Zhang *et al.*, 2015; Qiu *et al.*, 2016). However, exosome-mediated regulation of the astrocytic immune response by OECs may also be a unique interaction, as membrane composition and protein content of exosomes is cell type (Kalra *et al.*, 2012; Keerthikumar *et al.*, 2015; Kim *et al.*, 2015) and context specific (György *et al.*, 2011; Müller *et al.*, 2012). The robust anti-inflammatory response induced by OEC-secreted CryAB, shown in the present report, may also be a function of concentration, as our results indicate the concentration of CryAB in OEC exosomes to be higher than that found in astrocyte exosomes. We found that CryAB in OEC secreted exosomes was ~21% higher than that of astrocyte exosomes (Supplemental Figure 1A) and exposure to LPS increased OEC secreted CryAB concentration by ~25% (Supplemental Figure 1B), indicating OECs actively respond to stress by either secreting more exosomes or increasing CryAB concentration in exosomes, or both. In addition, exposure to Anosmin1 increased CryAB concentration by ~77% (Supplemental Figure 1C), compared to control protein recombinant myoglobin, suggesting OECs can actively respond to extracellular astrocyte signals by increasing CryAB concentrations.

4.3 CryAB, as well as other factors secreted by OECs, can suppress neurotoxic astrocyte reactivity

Stress causes denaturation of correctly folded proteins that can result in their aggregation and binding to CryAB (Muranova *et al.*, 2018) and subsequent changes in gene expression (Singh *et al.*, 2019). Our results show that OEC-secreted CryAB suppressed expression of a number of genes associated with neurotoxic astrocyte reactivity and suggest there are additional factor(s) in OEC-CM that further suppress this harmful reactivity. In fact, complement component-3 (C3) expression in IP-CryAB +LPS-treated astrocytes, although not significantly different than that observed in astrocytes treated with LPS alone, was significantly greater

than expression in astrocytes treated with either OEC-CM groups (Figure 5). C3 is an important marker for neurotoxic astrocytes, evident by knockout mice showing reduced activity in microglia and astrocytes, as well as neuron loss (Shi *et al.*, 2017). Moreover, C3 is found colocalized with astrocyte markers in regions of neurodegeneration in human post-mortem tissue (Liddelow *et al.*, 2017). Therefore, more experiments are required to identify OEC-secreted factor(s) that can suppress C3. In this regard, other proteins identified in our mass spectrometric screen, showing smaller changes, may be worthwhile to evaluate. Certainly, crosstalk mechanisms between OECs and surrounding niche cells, including astrocytes, are highly complicated (Chuah *et al.*, 2011). In addition to unidentified anti-inflammatory factors, absence or suppression of pro-inflammatory factors might also play a role in OEC-CM's effect on astrocyte reactivity. For example, our mass spectrometry screening showed higher concentration of S100A4 and S100A6 proteins in C4-CM compared to pOEC-CM or C7-CM (Figure 2, Supplemental Data-1 & 2). S100 proteins are known to modulate neuroinflammation (Donato, 2001), and as such are also candidate molecules that may contribute to the observed difference in the inflammatory reactivity of astrocytes treated with C4-CM versus pOEC-CM or C7-CM.

Recent studies have transformed our perception of astrocytes from a passive structural support network for neurons, to active effectors in the regulation of synaptic transmission, neural excitability, plasticity and recovery. In this paper, we identify OEC secreted CryAB as an anti-inflammatory factor that can moderate astrocyte reactivity, suppressing both transcription of neurotoxic classified genes and nuclear translocation of pro-inflammatory factor NF κ B. Improving our understanding of the crosstalk between astrocytes and OECs may inform strategies to identify other endogenous repair mechanisms that facilitate CNS repair, and consequently impact function, in the injured nervous system.

Figure Legends

Figure 1: OEC-CM alone is sufficient to suppress LPS-induced astrocyte reactivity, an effect mimicked by a subset of immortalized OEC lines.

(A) Quantitative immunoblotting for NF κ B was performed on the nuclear fraction of C8D30 astrocyte lysates (inset). All groups were compared using a one-way ANOVA (N=3). Treatment with LPS for 2 hrs significantly increased NF κ B activity (* indicates $p \leq 0.05$, gray bars). The presence of OECs blocked the effect of LPS (purple bars). OEC conditioned medium (OEC-CM) alone also blocked the increase in nuclear NF κ B (red bars). (B and C) CM from six immortalized OEC lines were investigated for their ability to block the effect of LPS on nuclear NF κ B translocation in C8D30 astrocytes, as measured by the ratio of fluorescence intensity of nuclear to cytoplasmic NF κ B. (B) Photomicrograph of images of C8D30 astrocytes cocultured with CM of immortalized cell lines. (C) Fluorescent NF κ B nuclear and cytoplasmic intensities were measured and ratios plotted. Pink dashed line depicts value of astrocytes treated with primary OEC-CM +LPS and red dashed line depicts value of astrocytes +LPS. A median value was calculated from ~ 100 cells per field, and then a mean/group was calculated. C7-CM and D6-CM decreased NF κ B nuclear translocation (N = 3; * $p \leq 0.05$; one-way ANOVA), while C4-CM treated groups was not significantly different than astrocytes +LPS alone (N = 3; * $p \leq 0.05$; one-way ANOVA). (D) Photomicrograph from line C7. Multiple morphologies were found in all the OEC cell lines: (a) Schwann Cell-like, (b) astrocyte-like type1, and (c) astrocyte-like type2. Schwann cell-like spindle cells (a) predominated in C7. (E) C7 and C4 olfactory cell lines share multiple markers with OECs including BLBP (magenta), Sox10 (green) and P75 (red). Scale bars represent 25 and 20 μ m.

Figure 2: OEC-secreted anti-inflammatory protein CryAB and recombinant CryAB is sufficient to suppress astrocyte reactivity measured by NF κ B.

(A, B) Comparison of factors secreted from C7 line, C4 line and primary OECs (pOECs), analyzed by mass spectrometry. Proteins detected in CM following LPS treatment (A) or Anosmin1 treatment (B) were ranked by their relative abundance indicated by color code (heat map, inset). Relative abundance of detected proteins in C7-CM compared to pOEC-CM is graphed on the Y axes, and relative absence of the same proteins from C4-CM ($1 - P(\text{C4-CM})$ = probability of not being found in C4-CM) is graphed on the X axes. Proteins of similar abundance in CMs from C7 cells and pOECs (horizontal red-dashed lines), and not likely to be present in C4-CM (Y axes) were identified. Alpha crystallin B chain (CryAB) had the highest C7/C4 expression ratio and was equally abundant in CM from C7 cells and pOECs. (C) Recombinant CryAB alone suppressed the inflammatory response, quantified as the ratio of nuclear to cytoplasmic NF κ B (Y axis) in astrocytes exposed to either LPS or a cocktail of the cytokines Il-1 α , TNF α and C1q for 2 hrs. Group values were obtained from

triplicate wells in which a median value was calculated from ~50 cells per field. N = 3; (*p ≤ 0.05; **p ≤ 0.01; ***p ≤ 0.0001); two-way ANOVA.

Figure 3: CryAB, secreted by primary OECs into exosomes, suppresses inflammatory response in an astrocyte cell line.

(A) Immunoblot of exosome (Exo, left) and whole cell lysate (CL, right) fractions from WT primary OECs were screened for β -actin, Tomm20, histone H3, and Flotilin-1. The exosome fraction from WT exosomes was devoid of cellular β -actin, Tomm20 and histone H3, but contained the extravesicular protein Flotilin-1. (B) Immunoblots for CryAB and the exosome marker Alix were performed on exosome fractions made from *CryAB*^{-/-} and WT (*CryAB*^{+/+}) OEC cultures. CryAB was absent in exosome fractions from *CryAB*^{-/-} OEC culture medium, whereas the exosome marker Alix was present. (C) C8D30 astrocytes were treated for 24 hours with exosomes isolated from WT or *CryAB*^{-/-} OECs. Astrocytes were exposed to 1 μ g/ml LPS for the last 2 hrs of exosome treatment. Nuclear fractions of astrocytes were analyzed via quantitative immunoblotting for NF κ B and Histone H3. Inset shows a representative immunoblot and graph shows mean \pm SD of NF κ B/histone ratio. All conditions were compared to astrocyte alone group (Control, N = 3; p ≤ 0.05; one-way ANOVA). Treatment of astrocytes with WT OEC-exosomes (exo) + LPS, blocked nuclear NF κ B translocation. In contrast, *CryAB*^{-/-}OEC-exosomes failed to suppress nuclear NF κ B translocation. Recombinant CryAB (50ng/ml) added to *CryAB*^{-/-}OEC-exosomes was sufficient to attenuate NF κ B translocation induced by LPS, with levels comparable to WT OEC-exo +LPS. (D) C8D30 astrocytes treated with LPS for 2 hrs (top right: "+ LPS") showed stronger immunostaining for NF κ B in the nucleus (magenta) compared to untreated controls (top left). Astrocytes co-cultured with *CryAB*^{-/-} OECs (Sox 10-positive cells with blue nuclei) had increased levels of NF κ B immunostaining in the nucleus (bottom right: "+*CryAB*^{-/-}OECs +LPS") compared to astrocytes co-cultured with WT OECs (bottom left: "+OECs +LPS"). Scale bar represents 40 μ m.

Figure 4: CryAB in OEC exosomes is internalized by astrocytes.

(A) OEC exosomes from WT mice were co-cultured with primary astrocytes from CryAB KO mice. (B) Untreated astrocytes from CryAB KO. (C) Untreated astrocytes from WT mice. All groups were stained for CD63 (endosomes; red), CryAB (green), GFAP (magenta) and Dapi (blue). Uptake of OEC secreted CryAB (green) is detected in *CryAB*^{-/-} astrocytes and is often associated with endosomes (red) (A, arrows, top arrow area shown in inset, arrowhead points to CryAB positive endosome). No CryAB staining (green) is detected in untreated astrocytes from CryAB KO (B, inset). CryAB (green) is present in untreated astrocytes from WT

mice but rarely associated with endosomes (C, inset, arrowhead). Scale bar represents 5 μ m in low mag and 1 μ m in insets.

Figure 5: OEC-CM suppresses multiple transcripts associated with neurotoxic astrocyte reactivity in LPS-treated astrocytes.

Heat map illustrating results of Q-RT-PCR (Supplemental Data 3) to detect neurotoxic astrocyte transcripts in primary astrocytes treated with LPS (A); LPS plus WT OEC-CM (B); LPS plus *CryAB*^{-/-}OEC-CM (C); or IP-CryAB for 24 hrs (D). Compared to LPS, OEC-CM (B) significantly suppressed 9 transcripts, whereas *CryAB*^{-/-}OEC-CM (C) significantly suppressed 4. (D) IP-CryAB suppressed 6 of the 9 transcripts suppressed by OEC-CM. In addition, *CryAB*^{-/-}OEC-CM+LPS caused a significant increase in 4 transcripts, while IP-CryAB caused a significant increase in one. Finally, one transcript, C3, was significantly suppressed by both OEC-CM treatments (*CryAB*^{-/-} or WT) but was elevated by IP-CryAB treatment. Q-RT-PCR experiments were analyzed by one-way ANOVA followed by Dunnett's multiple post hoc test (n= 3; arrows p \leq 0.05).

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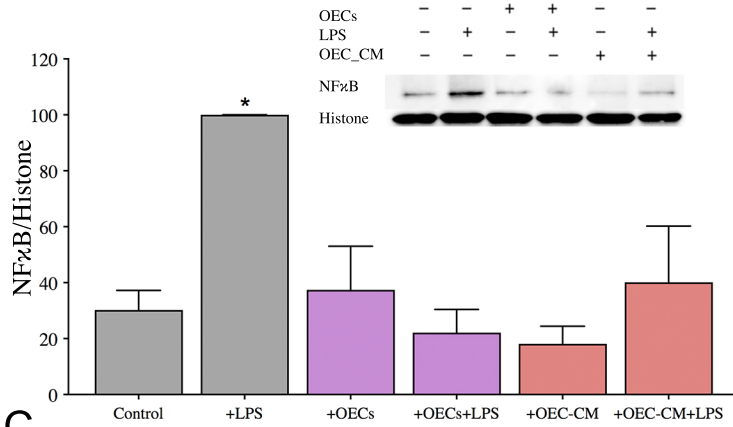
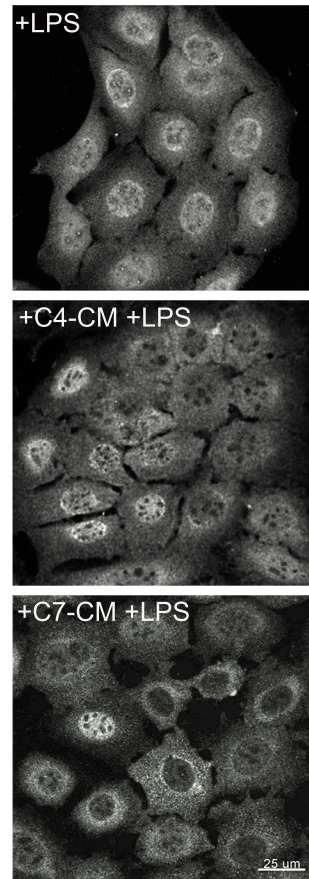
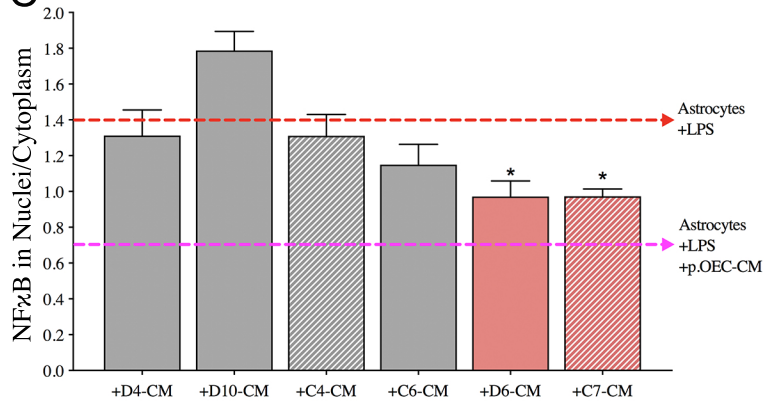
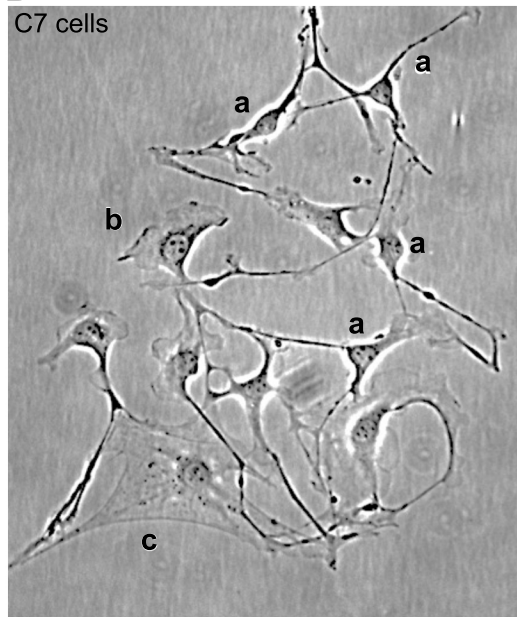
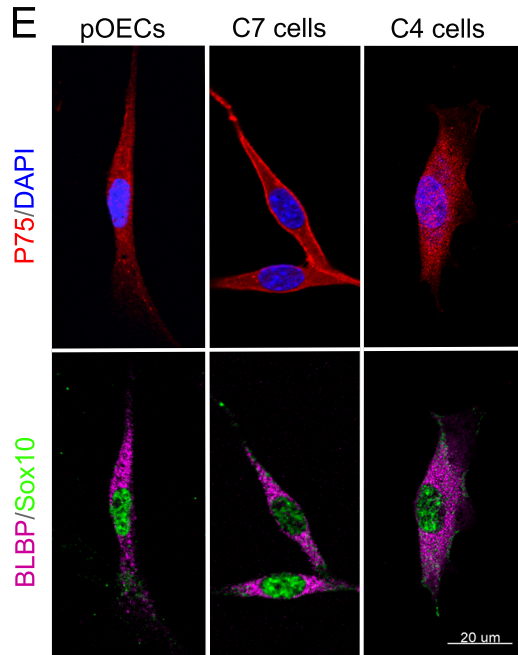
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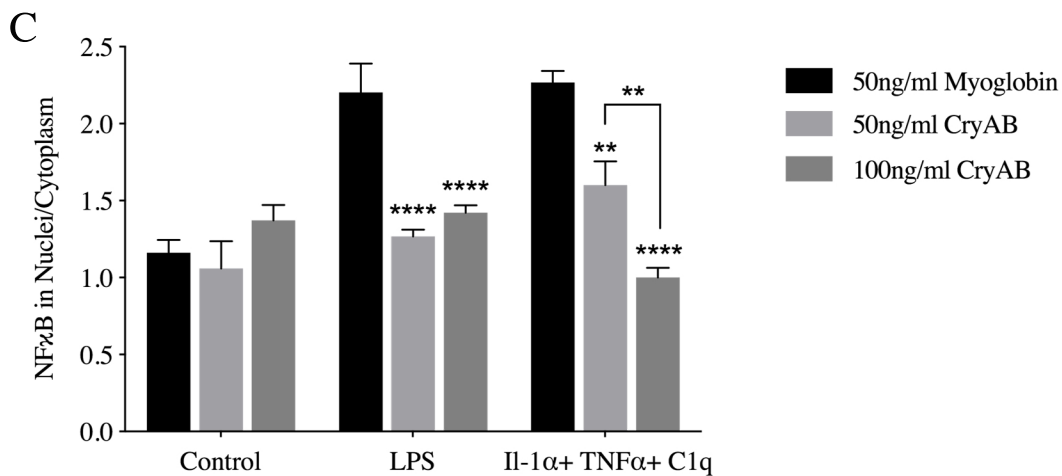
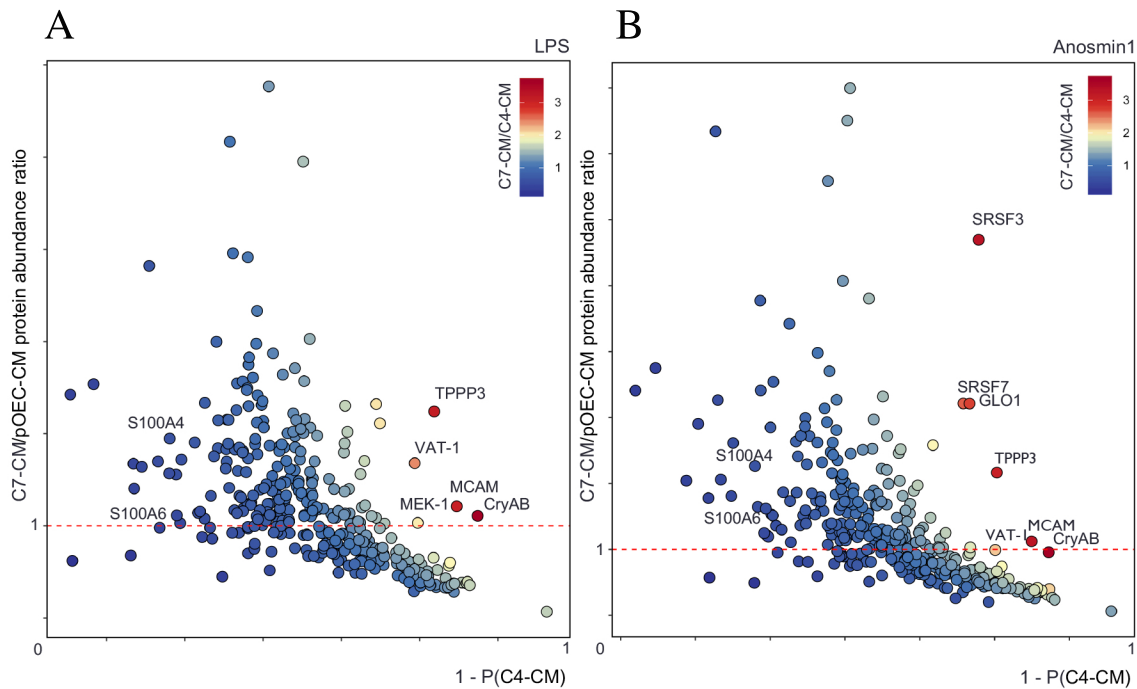
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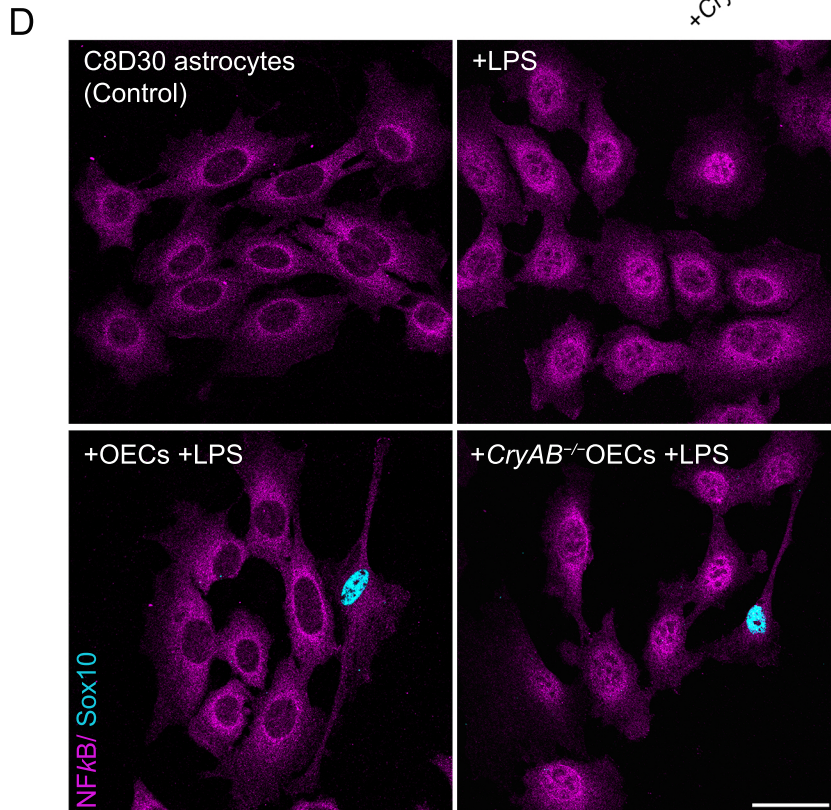
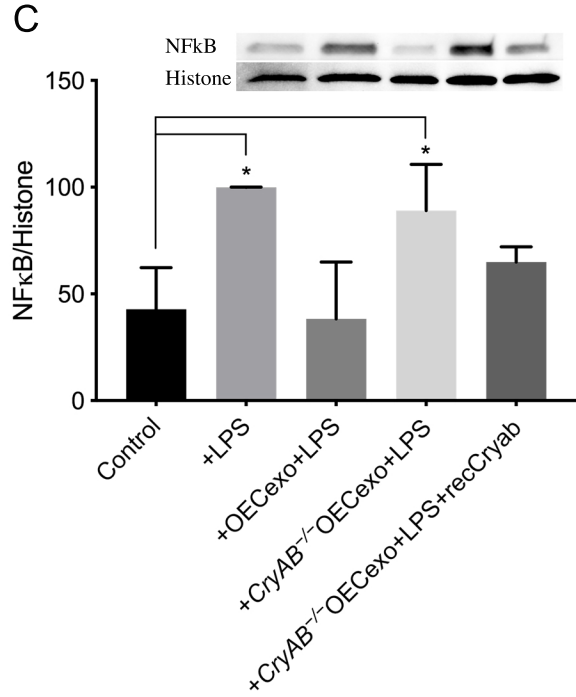
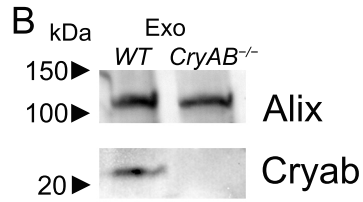
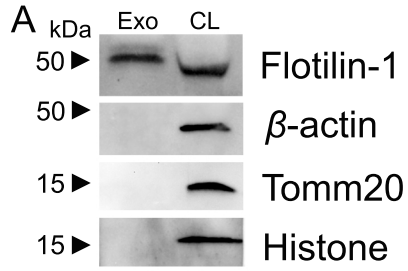
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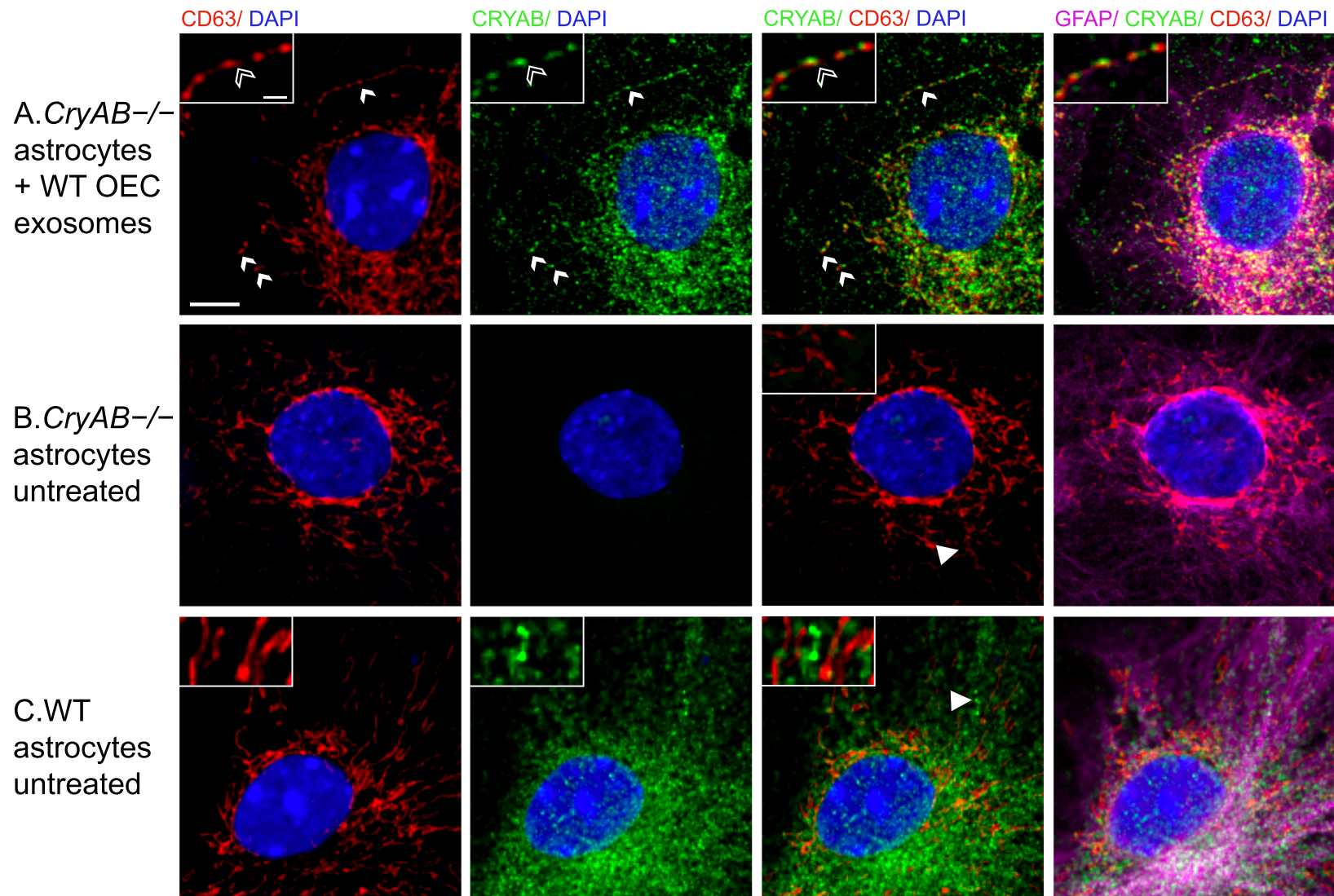
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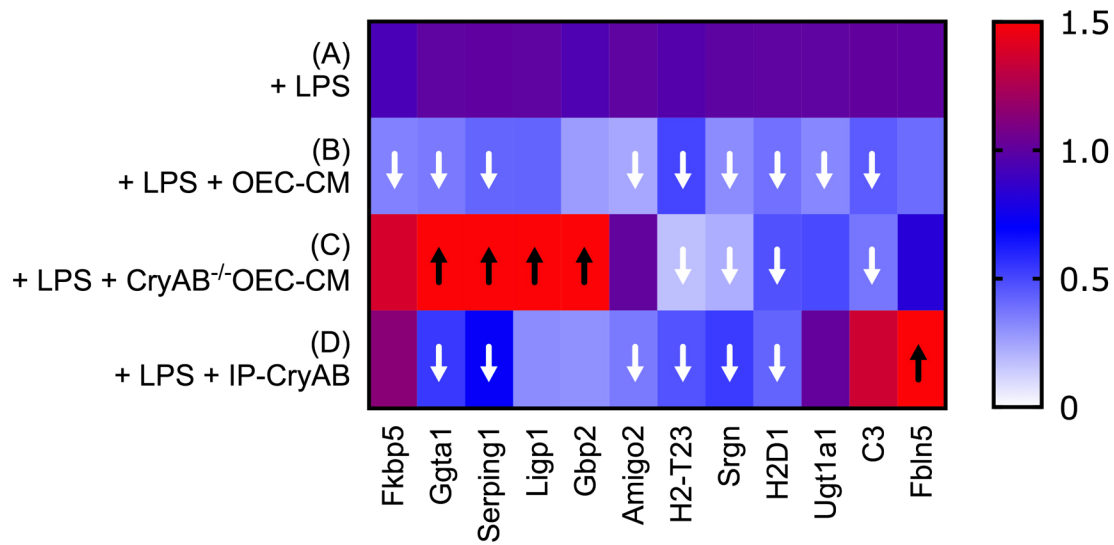
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A**B****C****D****E**

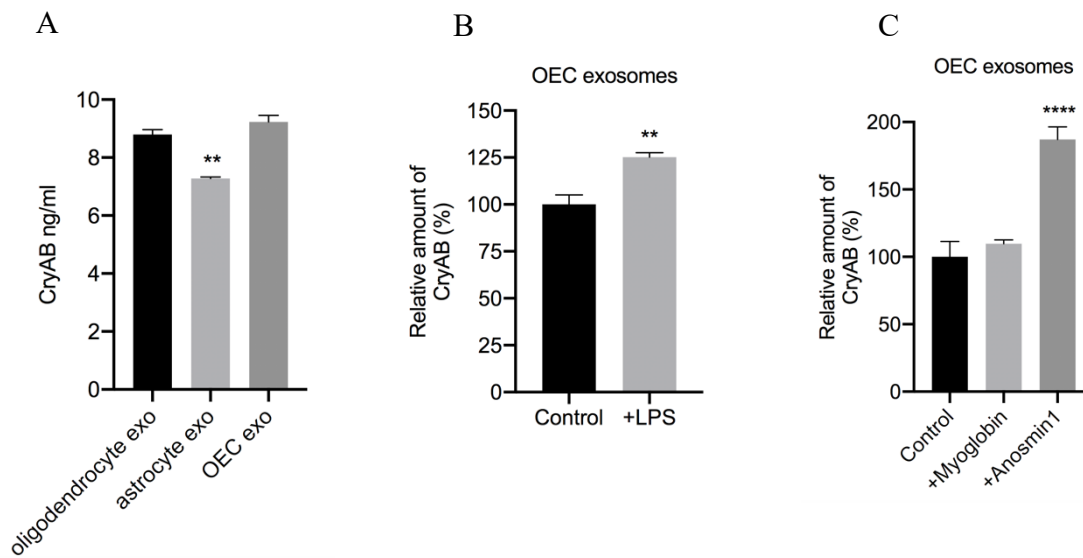








Supplementary Figure 1: Competitive ELISA against CryAB was used to examine exosomes. (A) Exosomes (exo) derived from 1×10^6 OECs were determined to have 9.22 ± 0.2 ng CryAB. CryAB in OEC secreted exosomes was not significantly different than the concentration in oligodendrocyte exosomes but 21.1% higher than that of astrocyte exosomes. (B) Exposure to LPS increased OEC secreted CryAB concentration by 25.17% (C). Although OECs' exposure to LPS or astrocytic signals were not necessary for the suppressive effect of OECs on NF κ B translocation in C8D30 astrocytes, whether the CryAB secretion would be facilitated by external signals was investigated by stimulating with LPS or Anosmin1. Exposure to Anosmin-1 increased CryAB concentration by 77.17%, compared to control protein recombinant myoglobin.



SUPPLEMENTARY ONLINE MATERIAL

SUPPLEMENTARY TABLE 1

| +D4-CM | +D10-CM | +C4-CM | +C6-CM | +D6-CM | +C7-CM | +pOECs | LPS control |
|--------|---------|--------|--------|--------|--------|--------|-------------|
| 102 | 74 | 81 | 83 | 113 | 127 | 79 | 123 |
| 129 | 78 | 67 | 64 | 126 | 77 | 84 | 100 |
| 92 | 107 | 74 | 69 | 131 | 80 | 95 | 88 |

SUPPLEMENTARY TABLE 2

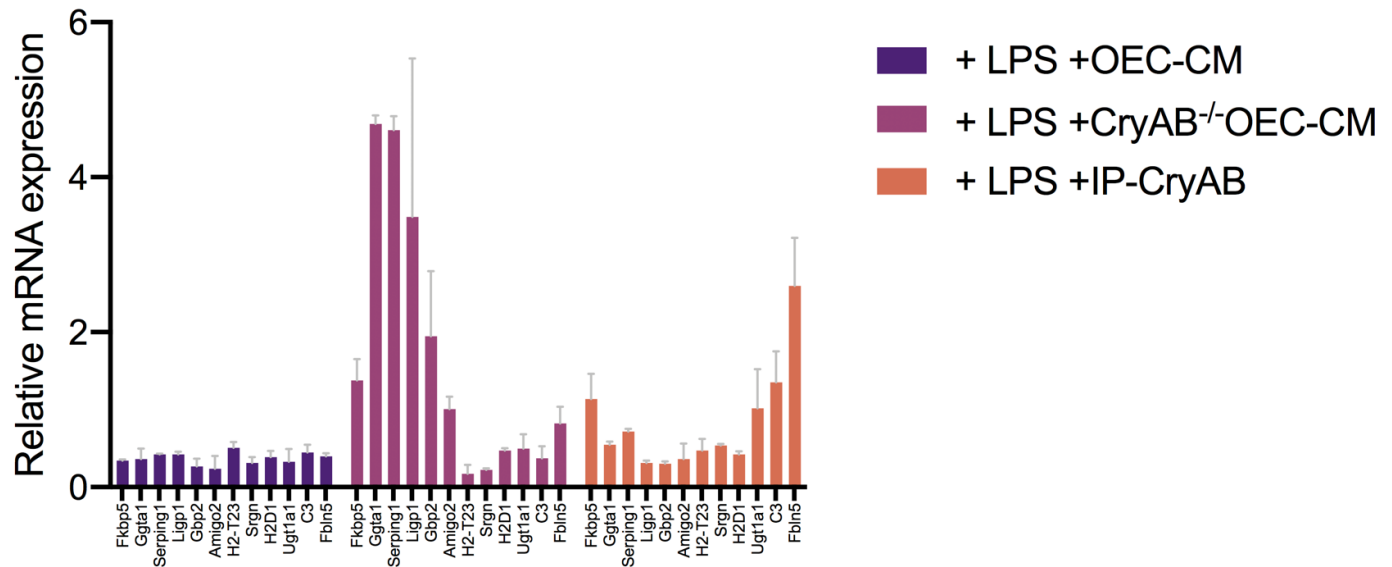
| | Medium | 50ng/ml Cryab | 100ng/ml Cryab |
|------------------------------------|--------|---------------|----------------|
| Medium | 50 | 52 | 43 |
| | 55 | 28 | 36 |
| | 44 | 43 | 51 |
| LPS | 53 | 50 | 47 |
| | 52 | 25 | 53 |
| | 56 | 40 | 54 |
| Il-1 α + TNF α + C1q | 52 | 46 | 59 |
| | 41 | 29 | 65 |
| | 49 | 59 | 61 |

Median values were calculated for each biological replicate obtained from multiple images containing around 100 cells for Figure 1C and 50 cells for Figure 2B.

SUPPLEMENTARY ONLINE MATERIAL

SUPPLEMENTARY TABLE 3

| Sample | dCt | dCt | dCt | mean | SD | SEM | ddCt | 2 ^Δ -(ddCt) | 2 ^Δ -(ddCt-SEM) | 2 ^Δ -(ddCt+SEM) |
|----------------------------|--------|--------|--------|--------|-------|-------|--------|------------------------|----------------------------|----------------------------|
| LPS | | | | | | | | | | |
| Fkbp5 | 7.831 | 7.461 | 8.015 | 7.769 | 0.282 | 0.163 | 0.000 | 1.000 | 1.120 | 0.893 |
| Ggta1 | 6.781 | 6.949 | 7.044 | 6.925 | 0.134 | 0.077 | 0.000 | 1.000 | 1.055 | 0.948 |
| Serping1 | 4.895 | 4.539 | 4.655 | 4.697 | 0.181 | 0.105 | 0.000 | 1.000 | 1.075 | 0.930 |
| Ligp1 | 7.240 | 7.012 | 7.063 | 7.105 | 0.120 | 0.069 | 0.000 | 1.000 | 1.049 | 0.953 |
| Gbp2 | 1.580 | 1.415 | 1.755 | 1.583 | 0.170 | 0.098 | 0.000 | 1.000 | 1.070 | 0.934 |
| Amigo2 | 9.793 | 9.748 | 9.844 | 9.795 | 0.048 | 0.028 | 0.000 | 1.000 | 1.019 | 0.981 |
| H2-T23 | 3.418 | 3.566 | 3.830 | 3.605 | 0.209 | 0.120 | 0.000 | 1.000 | 1.087 | 0.920 |
| Srgn | 5.651 | 5.635 | 5.730 | 5.672 | 0.051 | 0.030 | 0.000 | 1.000 | 1.021 | 0.980 |
| H2D1 | -4.439 | -4.486 | -4.382 | -4.436 | 0.052 | 0.030 | 0.000 | 1.000 | 1.021 | 0.979 |
| Ugt1a1 | 7.087 | 7.195 | 7.317 | 7.200 | 0.115 | 0.066 | 0.000 | 1.000 | 1.047 | 0.955 |
| C3 | 6.603 | 6.681 | 7.080 | 6.788 | 0.256 | 0.148 | 0.000 | 1.000 | 1.108 | 0.903 |
| Fbln5 | 11.742 | 11.483 | 11.806 | 11.677 | 0.171 | 0.099 | 0.000 | 1.000 | 1.071 | 0.934 |
| LPS +OEC-CM | | | | | | | | | | |
| Fkbp5 | 9.217 | 9.269 | 9.114 | 9.200 | 0.079 | 0.046 | 1.431 | 0.371 | 0.383 | 0.359 |
| Ggta1 | 8.041 | 8.182 | 9.227 | 8.484 | 0.648 | 0.374 | 1.559 | 0.339 | 0.440 | 0.262 |
| Serping1 | 5.899 | 5.969 | 5.964 | 5.944 | 0.039 | 0.022 | 1.248 | 0.421 | 0.428 | 0.415 |
| Ligp1 | 8.242 | 8.474 | 8.401 | 8.372 | 0.119 | 0.069 | 1.267 | 0.416 | 0.436 | 0.396 |
| Gbp2 | 3.030 | 3.230 | 4.131 | 3.463 | 0.587 | 0.339 | 1.880 | 0.272 | 0.344 | 0.215 |
| Amigo2 | 11.367 | 11.378 | 14.133 | 12.293 | 1.594 | 0.920 | 2.498 | 0.177 | 0.335 | 0.094 |
| H2-T23 | 4.164 | 4.506 | 4.531 | 4.400 | 0.205 | 0.119 | 0.796 | 0.576 | 0.625 | 0.531 |
| Srgn | 7.426 | 7.711 | 7.011 | 7.383 | 0.352 | 0.203 | 1.711 | 0.306 | 0.352 | 0.265 |
| H2D1 | -3.297 | -3.152 | -2.696 | -3.048 | 0.314 | 0.181 | 1.387 | 0.382 | 0.433 | 0.337 |
| Ugt1a1 | 8.151 | 9.198 | 9.449 | 8.932 | 0.688 | 0.397 | 1.733 | 0.301 | 0.396 | 0.228 |
| C3 | 8.288 | 7.970 | 7.645 | 7.968 | 0.322 | 0.186 | 1.179 | 0.442 | 0.502 | 0.388 |
| Fbln5 | 13.173 | 12.940 | 12.920 | 13.011 | 0.141 | 0.081 | 1.334 | 0.397 | 0.420 | 0.375 |
| LPS +CryABKO OEC-CM | | | | | | | | | | |
| Fkbp5 | 7.034 | 7.021 | 7.562 | 7.205 | 0.309 | 0.178 | -0.563 | 1.478 | 1.672 | 1.306 |
| Ggta1 | 4.680 | 4.735 | 4.673 | 4.696 | 0.034 | 0.020 | -2.229 | 4.687 | 4.752 | 4.623 |
| Serping1 | 2.442 | 2.555 | 2.483 | 2.493 | 0.057 | 0.033 | -2.203 | 4.605 | 4.711 | 4.501 |
| Ligp1 | 5.831 | 6.017 | 5.579 | 5.809 | 0.220 | 0.127 | -1.296 | 2.455 | 2.681 | 2.248 |
| Gbp2 | 0.146 | 1.509 | 0.293 | 0.650 | 0.748 | 0.432 | -0.933 | 1.910 | 2.576 | 1.416 |
| Amigo2 | 9.983 | 9.845 | 9.550 | 9.793 | 0.221 | 0.128 | -0.002 | 1.002 | 1.094 | 0.917 |
| H2-T23 | 7.413 | 5.966 | 5.216 | 6.198 | 1.117 | 0.645 | 2.594 | 0.166 | 0.259 | 0.106 |
| Srgn | 6.305 | 7.722 | 8.020 | 7.349 | 0.916 | 0.529 | 1.677 | 0.313 | 0.451 | 0.217 |
| H2D1 | -3.402 | -3.416 | -3.233 | -3.351 | 0.102 | 0.059 | 1.085 | 0.471 | 0.491 | 0.452 |
| Ugt1a1 | 7.791 | 8.941 | 8.113 | 8.282 | 0.593 | 0.342 | 1.082 | 0.472 | 0.599 | 0.373 |
| C3 | 8.070 | 7.776 | 9.057 | 8.301 | 0.671 | 0.387 | 1.513 | 0.350 | 0.458 | 0.268 |
| Fbln5 | 11.586 | 12.106 | 12.276 | 11.989 | 0.359 | 0.207 | 0.313 | 0.805 | 0.930 | 0.697 |
| LPS +IP-CryAB | | | | | | | | | | |
| Fkbp5 | 7.057 | 7.839 | 7.602 | 7.499 | 0.401 | 0.232 | -0.269 | 1.205 | 1.415 | 1.026 |
| Ggta1 | 7.713 | 7.937 | 7.758 | 7.803 | 0.119 | 0.069 | 0.878 | 0.544 | 0.571 | 0.519 |
| Serping1 | 5.114 | 5.172 | 5.249 | 5.178 | 0.068 | 0.039 | 0.482 | 0.716 | 0.736 | 0.697 |
| Ligp1 | 8.687 | 8.955 | 8.787 | 8.810 | 0.135 | 0.078 | 1.704 | 0.307 | 0.324 | 0.291 |
| Gbp2 | 3.079 | 3.347 | 3.246 | 3.224 | 0.135 | 0.078 | 1.641 | 0.321 | 0.338 | 0.304 |
| Amigo2 | 10.597 | 11.388 | 12.252 | 11.412 | 0.828 | 0.478 | 1.617 | 0.326 | 0.454 | 0.234 |
| H2-T23 | 4.391 | 5.143 | 4.150 | 4.561 | 0.518 | 0.299 | 0.957 | 0.515 | 0.634 | 0.419 |
| Srgn | 6.580 | 6.512 | 6.626 | 6.573 | 0.057 | 0.033 | 0.901 | 0.536 | 0.548 | 0.524 |
| H2D1 | -3.032 | -3.279 | -3.256 | -3.189 | 0.136 | 0.079 | 1.247 | 0.421 | 0.445 | 0.399 |
| Ugt1a1 | 8.277 | 6.647 | 7.045 | 7.323 | 0.850 | 0.490 | 0.123 | 0.918 | 1.290 | 0.654 |
| C3 | 6.097 | 6.949 | 6.158 | 6.401 | 0.476 | 0.275 | -0.387 | 1.308 | 1.582 | 1.081 |
| Fbln5 | 10.750 | 10.186 | 10.053 | 10.329 | 0.370 | 0.214 | -1.347 | 2.544 | 2.950 | 2.194 |



| Checked | Protein FDI | Master | Accession | Description | Exp. q-val | Sum PEP | Sc | Coverage [%] | # Peptides | # PSMs | # Unique P. | # Protein G | # AAs | MW [kDa] | calc. pI | Score Sequ | # Peptides | Abundance Ratio: (F1, 130) / (F1, 129) | Abundance Ratio: (F1, 131) / (F1, 129) |
|---------|-------------|------------|-----------|--|------------|---------|----|--------------|------------|--------|-------------|-------------|-------|----------|----------|------------|------------|--|--|
| FALSE | High | Master Pro | Q90X51-1 | plectin [OS=Mus musculus] | 0 | 168.56 | 11 | 45 | 87 | 45 | 1 | 4691 | 533.9 | 5.96 | 306.49 | 45 | 1.227 | 1.431 | |
| FALSE | High | Master Pro | P58252 | Elongation factor 2 [OS=Mus musculus] | 0 | 99.744 | 21 | 16 | 66 | 16 | 1 | 858 | 95.3 | 6.83 | 247.29 | 16 | 2.718 | 3.012 | |
| FALSE | High | Master Pro | P11276 | fibronectin [OS=Mus musculus] | 0 | 97.128 | 11 | 20 | 50 | 20 | 1 | 2477 | 272.4 | 5.59 | 188.54 | 20 | 0.747 | 1.133 | |
| FALSE | High | Master Pro | P52480 | Pyruvate kinase PKM [OS=Mus musculus] | 0 | 86.418 | 35 | 14 | 45 | 2 | 1 | 531 | 57.8 | 7.47 | 180.9 | 14 | 1.745 | 1.431 | |
| FALSE | High | Master Pro | P63017 | Heat shock cognate 71 kDa protein [OS=Mus musculus] | 0 | 75.674 | 21 | 13 | 55 | 10 | 1 | 646 | 70.8 | 5.52 | 210.48 | 13 | 2.528 | 2.014 | |
| FALSE | High | Master Pro | P09411 | phosphoglycerate kinase 1 [OS=Mus musculus] | 0 | 68.779 | 34 | 13 | 46 | 13 | 1 | 417 | 44.5 | 7.9 | 157.04 | 13 | 4.423 | 4.896 | |
| FALSE | High | Master Pro | P60710 | Actin, cytoplasmic 1 [OS=Mus musculus] | 0 | 68.401 | 39 | 12 | 43 | 5 | 1 | 375 | 41.7 | 5.48 | 164.18 | 12 | 1.544 | 1.161 | |
| FALSE | High | Master Pro | Q88TM8 | Filamin-A [OS=Mus musculus] | 0 | 68.209 | 8 | 19 | 44 | 15 | 1 | 2647 | 281 | 6.04 | 160.39 | 19 | 1.186 | 1.359 | |
| FALSE | High | Master Pro | P17182 | alpha-enolase [OS=Mus musculus] | 0 | 63.905 | 33 | 11 | 43 | 11 | 1 | 434 | 47.1 | 6.8 | 172.55 | 11 | 2.32 | 2.511 | |
| FALSE | High | Master Pro | P20152 | Vimentin [OS=Mus musculus] | 0 | 58.757 | 28 | 13 | 49 | 13 | 1 | 466 | 53.7 | 5.12 | 176.17 | 13 | 2.621 | 2.81 | |
| FALSE | High | Master Pro | P52480-2 | Isoform M1 of Pyruvate kinase PKM [OS=Mus musculus] | 0 | 58.552 | 29 | 13 | 38 | 1 | 1 | 531 | 57.9 | 7.14 | 136.08 | 13 | 1.092 | 0.948 | |
| FALSE | High | Master Pro | P10107 | annexin A1 [OS=Mus musculus] | 0 | 57.805 | 29 | 10 | 28 | 10 | 1 | 346 | 38.7 | 7.37 | 107.18 | 10 | 2.352 | 2.329 | |
| FALSE | High | Master Pro | P26041 | Moesin [OS=Mus musculus] | 0 | 56.362 | 28 | 15 | 40 | 10 | 1 | 577 | 67.7 | 6.6 | 131.47 | 15 | 1.893 | 1.637 | |
| FALSE | High | Master Pro | P57780 | Alpha-actinin-4 [OS=Mus musculus] | 0 | 48.178 | 14 | 12 | 35 | 3 | 1 | 912 | 104.9 | 5.41 | 113.6 | 12 | 0.834 | 0.863 | |
| FALSE | High | Master Pro | P05064 | fructose-bisphosphate aldolase A [OS=Mus musculus] | 0 | 48.053 | 29 | 10 | 30 | 9 | 1 | 364 | 39.3 | 8.09 | 104.94 | 10 | 1.843 | 1.574 | |
| FALSE | High | Master Pro | P11087-1 | Collagen alpha-1(I) chain [OS=Mus musculus] | 0 | 46.912 | 11 | 14 | 28 | 13 | 1 | 1453 | 137.9 | 5.85 | 97.19 | 14 | 0.696 | 0.492 | |
| FALSE | High | Master Pro | P10126 | Elongation factor 1-alpha 1 [OS=Mus musculus] | 0 | 45.662 | 21 | 9 | 28 | 9 | 1 | 462 | 50.1 | 9.01 | 109.97 | 9 | 1.6 | 1.98 | |
| FALSE | High | Master Pro | Q64727 | Vinculin [OS=Mus musculus] | 0 | 45.505 | 11 | 10 | 15 | 10 | 1 | 1066 | 116.6 | 6 | 54.03 | 10 | 0.493 | 0.603 | |
| FALSE | High | Master Pro | P01027-1 | Complement C3 [OS=Mus musculus] | 0 | 45.355 | 5 | 8 | 28 | 8 | 1 | 1663 | 186.4 | 6.73 | 112.15 | 8 | 0.328 | 0.232 | |
| FALSE | High | Master Pro | Q07235 | Glia-derived nexin [OS=Mus musculus] | 0 | 44.736 | 24 | 9 | 21 | 9 | 1 | 397 | 44.2 | 9.85 | 77.39 | 9 | 1.072 | 1.139 | |
| FALSE | High | Master Pro | P26039 | Talin-1 [OS=Mus musculus] | 0 | 43.551 | 6 | 12 | 23 | 12 | 1 | 2541 | 269.7 | 6.18 | 67.37 | 12 | 0.649 | 0.731 | |
| FALSE | High | Master Pro | Q77PR4 | Alpha-actinin-1 [OS=Mus musculus] | 0 | 43.441 | 12 | 10 | 29 | 1 | 1 | 892 | 103 | 5.38 | 94.42 | 10 | 0.752 | 0.757 | |
| FALSE | High | Master Pro | Q8VDD5 | Myosin-9 [OS=Mus musculus] | 0 | 41.956 | 6 | 11 | 22 | 9 | 1 | 1960 | 226.2 | 5.66 | 73.14 | 11 | 0.944 | 1.328 | |
| FALSE | High | Master Pro | Q61171 | Peroxioredoxin-2 [OS=Mus musculus] | 0 | 41.525 | 32 | 6 | 21 | 6 | 1 | 198 | 21.8 | 5.41 | 86.85 | 6 | 2.01 | 2.002 | |
| FALSE | High | Master Pro | P18760 | Cofilin-1 [OS=Mus musculus] | 0 | 41.122 | 54 | 7 | 24 | 7 | 1 | 166 | 18.5 | 8.09 | 106.74 | 7 | 3.541 | 4.467 | |
| FALSE | High | Master Pro | Q80X90 | Filamin-B [OS=Mus musculus] | 0 | 39.005 | 4 | 9 | 21 | 5 | 1 | 2602 | 277.7 | 5.71 | 75.34 | 9 | 0.691 | 0.51 | |
| FALSE | High | Master Pro | Q61598 | Rab GDP dissociation inhibitor beta [OS=Mus musculus] | 0 | 38.652 | 25 | 8 | 18 | 6 | 1 | 445 | 50.5 | 6.25 | 68.74 | 8 | 2.459 | 1.689 | |
| FALSE | High | Master Pro | Q61703 | Inter-alpha-trypsin inhibitor heavy chain H2 [OS=Mus musculus] | 0 | 38.479 | 8 | 9 | 22 | 9 | 1 | 946 | 105.9 | 7.27 | 78.85 | 9 | 0.364 | 0.232 | |
| FALSE | High | Master Pro | P05213 | Tubulin alpha-1B chain [OS=Mus musculus] | 0 | 36.125 | 19 | 7 | 15 | 7 | 1 | 451 | 50.1 | 5.06 | 70.82 | 7 | 0.896 | 0.664 | |
| FALSE | High | Master Pro | P63101 | 14-3-3 protein zeta/delta [OS=Mus musculus] | 0 | 35.393 | 38 | 8 | 23 | 5 | 1 | 245 | 27.8 | 4.79 | 85.23 | 8 | 2.05 | 2.084 | |
| FALSE | High | Master Pro | P62962 | profilin-1 [OS=Mus musculus] | 0 | 34.186 | 45 | 5 | 28 | 5 | 1 | 140 | 14.9 | 8.28 | 114.32 | 5 | 1.548 | 1.489 | |
| FALSE | High | Master Pro | Q88569 | heterogeneous nuclear ribonucleoproteins A2/B1 [OS=Mus musculus] | 0 | 33.745 | 16 | 7 | 18 | 5 | 1 | 353 | 37.4 | 8.95 | 65.38 | 7 | 2.474 | 2.507 | |
| FALSE | High | Master Pro | Q60847 | Collagen alpha-1(XII) chain [OS=Mus musculus] | 0 | 33.473 | 3 | 10 | 14 | 10 | 1 | 3120 | 340 | 5.64 | 46.12 | 10 | 0.626 | 0.698 | |
| FALSE | High | Master Pro | P09405 | Nucleolin [OS=Mus musculus] | 0 | 32.325 | 14 | 11 | 28 | 11 | 1 | 707 | 76.7 | 4.75 | 88.5 | 11 | 1.778 | 3.878 | |
| FALSE | High | Master Pro | P15532 | Nucleoside diphosphate kinase A [OS=Mus musculus] | 0 | 32.292 | 53 | 6 | 26 | 3 | 1 | 152 | 17.2 | 7.37 | 90.28 | 6 | 3.703 | 3.418 | |
| FALSE | High | Master Pro | P62737 | Actin, aortic smooth muscle [OS=Mus musculus] | 0 | 31.81 | 24 | 8 | 25 | 1 | 1 | 377 | 42 | 5.39 | 77.32 | 8 | 0.91 | 0.587 | |
| FALSE | High | Master Pro | Q9JMH6-1 | Thioredoxin reductase 1, cytoplasmic [OS=Mus musculus] | 0 | 31.631 | 11 | 5 | 11 | 5 | 1 | 613 | 67 | 7.44 | 52.36 | 5 | 1.334 | 0.981 | |
| FALSE | High | Master Pro | Q9WV44 | Transgelin-2 [OS=Mus musculus] | 0 | 31.513 | 42 | 8 | 21 | 8 | 1 | 199 | 22.4 | 8.24 | 69.09 | 8 | 1.223 | 1.594 | |
| FALSE | High | Master Pro | Q64433 | 10 kDa heat shock protein, mitochondrial [OS=Mus musculus] | 0 | 31.398 | 68 | 8 | 18 | 8 | 1 | 102 | 11 | 8.35 | 67.39 | 8 | 3.75 | 8.955 | |
| FALSE | High | Master Pro | P27773 | Protein disulfide-isomerase A3 [OS=Mus musculus] | 0 | 30.8 | 19 | 10 | 19 | 10 | 1 | 505 | 56.6 | 6.21 | 62.51 | 10 | 1.89 | 1.286 | |
| FALSE | High | Master Pro | P06745 | glucose-6-phosphate isomerase [OS=Mus musculus] | 0 | 30.442 | 16 | 7 | 14 | 7 | 1 | 558 | 62.7 | 8.13 | 44.29 | 7 | 1.155 | 0.81 | |
| FALSE | High | Master Pro | Q01768 | nucleoside diphosphate kinase b [OS=Mus musculus] | 0 | 29.932 | 53 | 6 | 19 | 3 | 1 | 152 | 17.4 | 7.5 | 75.76 | 6 | 2.156 | 2.261 | |
| FALSE | High | Master Pro | P35441 | thrombospondin-1 [OS=Mus musculus] | 0 | 29.845 | 8 | 9 | 17 | 9 | 1 | 1170 | 129.6 | 4.96 | 60.03 | 9 | 0.436 | 0.391 | |
| FALSE | High | Master Pro | P25444 | 40S ribosomal protein S2 [OS=Mus musculus] | 0 | 28.66 | 18 | 5 | 22 | 5 | 1 | 293 | 31.2 | 10.24 | 79.24 | 5 | 1.557 | 1.683 | |
| FALSE | High | Master Pro | Q05793 | Basement membrane-specific heparan sulfate proteoglycan core protein [OS=Mus musculus] | 0 | 28.565 | 3 | 9 | 15 | 9 | 1 | 3707 | 398 | 6.32 | 38.48 | 9 | 0.48 | 0.564 | |
| FALSE | High | Master Pro | Q62009 | Periostin [OS=Mus musculus] | 0 | 27.848 | 8 | 6 | 16 | 6 | 1 | 838 | 93.1 | 7.53 | 59.25 | 6 | 0.417 | 0.515 | |
| FALSE | High | Master Pro | Q9JKF1 | Ras GTPase-activating-like protein IQGAP1 [OS=Mus musculus] | 0 | 27.598 | 5 | 6 | 7 | 6 | 1 | 1657 | 188.6 | 6.48 | 29.32 | 6 | 0.977 | 0.892 | |
| FALSE | High | Master Pro | P07356 | Annexin A2 [OS=Mus musculus] | 0 | 27.585 | 28 | 9 | 16 | 9 | 1 | 339 | 38.7 | 7.69 | 50.66 | 9 | 1.691 | 1.226 | |
| FALSE | High | Master Pro | P06151 | L-lactate dehydrogenase A chain [OS=Mus musculus] | 0 | 27.541 | 16 | 5 | 16 | 5 | 1 | 332 | 36.5 | 7.74 | 60.26 | 5 | 1.529 | 2.075 | |
| FALSE | High | Master Pro | P48678-1 | Prelamin-A/C [OS=Mus musculus] | 0 | 27.457 | 11 | 8 | 22 | 8 | 1 | 665 | 74.2 | 6.98 | 77.17 | 8 | 1.68 | 1.5 | |
| FALSE | High | Master Pro | P61979-2 | Isoform 2 of Heterogeneous nuclear ribonucleoprotein K [OS=Mus musculus] | 0 | 27.04 | 16 | 5 | 13 | 5 | 1 | 464 | 51 | 5.33 | 53.94 | 5 | 1.741 | 2.143 | |
| FALSE | High | Master Pro | P17225 | Polypyrimidine tract-binding protein 1 [OS=Mus musculus] | 0 | 26.803 | 10 | 4 | 11 | 4 | 1 | 527 | 56.4 | 8.34 | 52.51 | 4 | 1.514 | 2.107 | |
| FALSE | High | Master Pro | Q61704 | Inter-alpha-trypsin inhibitor heavy chain H3 [OS=Mus musculus] | 0 | 26.569 | 4 | 4 | 14 | 4 | 1 | 889 | 99.3 | 6.05 | 60.83 | 4 | 0.351 | 0.216 | |
| FALSE | High | Master Pro | P99029 | Peroxioredoxin-5, mitochondrial [OS=Mus musculus] | 0 | 26.55 | 24 | 4 | 17 | 4 | 1 | 210 | 21.9 | 8.85 | 61.02 | 4 | 2.042 | 4.8 | |
| FALSE | High | Master Pro | Q01853 | Transitional endoplasmic reticulum ATPase [OS=Mus musculus] | 0 | 26.494 | 9 | 7 | 12 | 7 | 1 | 806 | 89.3 | 5.26 | 42.11 | 7 | 0.89 | 1.269 | |
| FALSE | High | Master Pro | Q99020 | Heterogeneous nuclear ribonucleoprotein A/B [OS=Mus musculus] | 0 | 26.267 | 25 | 6 | 15 | 5 | 1 | 285 | 30.8 | 7.91 | 50.92 | 6 | 2.721 | 1.949 | |
| FALSE | High | Master Pro | Q88844 | Isoctrate dehydrogenase [NADP] cytoplasmic [OS=Mus musculus] | 0 | 26.07 | 21 | 8 | 14 | 7 | 1 | 414 | 46.6 | 7.17 | 50.2 | 8 | 0.819 | 1.562 | |
| FALSE | High | Master Pro | Q08709 | Peroxioredoxin-6 [OS=Mus musculus] | 0 | 25.626 | 28 | 5 | 24 | 5 | 1 | 224 | 24.9 | 6.01 | 86.32 | 5 | 1.215 | 2.257 | |
| FALSE | High | Master Pro | P35700 | peroxiredoxin-1 [OS=Mus musculus] | 0 | 25.532 | 34 | 7 | 21 | 5 | 1 | 199 | 22.2 | 8.12 | 77.24 | 7 | 1.626 | 1.807 | |
| FALSE | High | Master Pro | P63038-1 | 60 kDa heat shock protein, mitochondrial [OS=Mus musculus] | 0 | 25.225 | 14 | 6 | 10 | 6 | 1 | 573 | 60.9 | 6.18 | 42.24 | 6 | 1.533 | 2.52 | |
| FALSE | High | Master Pro | P97298 | Pigment epithelium-derived factor [OS=Mus musculus] | 0 | 25.151 | 15 | 5 | 10 | 5 | 1 | 417 | 46.2 | 6.98 | 44.23 | 5 | 0.254 | 0.215 | |
| FALSE | High | Master Pro | P14152 | Malate dehydrogenase, cytoplasmic [OS=Mus musculus] | 0 | 24.795 | 19 | 7 | 16 | 7 | 1 | 334 | 36.5 | 6.58 | 56.85 | 7 | 3.319 | 2.319 | |
| FALSE | High | Master Pro | Q9D8N0 | elongation factor 1-gamma [OS=Mus musculus] | 0 | 24.487 | 15 | 6 | 11 | 6 | 1 | 437 | 50 | 6.74 | 36.57 | 6 | 0.898 | 1.293 | |
| FALSE | High | Master Pro | Q61207 | Prosaposin [OS=Mus musculus] | 0 | 24.46 | 11 | 6 | 19 | 6 | 1 | 557 | 61.4 | 5.19 | 61.01 | 6 | 1.645 | 2.828 | |
| FALSE | High | Master Pro | Q9DBJ1 | Phosphoglycerate mutase 1 [OS=Mus musculus] | 0 | 24.139 | 24 | 4 | 19 | 4 | 1 | 254 | 28.8 | 7.18 | 72.08 | 4 | 2.29 | 2.702 | |
| FALSE | High | Master Pro | P16546 | Spectrin alpha chain, non-erythrocytic 1 [OS=Mus musculus] | 0 | 24.188 | 3 | 5 | 6 | 5 | 1 | 2472 | 284.4 | 5.33 | 24.36 | 5 | 0.872 | 0.635 | |
| FALSE | High | Master Pro | P49312-1 | Heterogeneous nuclear ribonucleoprotein A1 [OS=Mus musculus] | 0 | 24.171 | 13 | 5 | 15 | 3 | 1 | 320 | 34.2 | 9.23 | 53.75 | 5 | 1.545 | 1.597 | |
| FALSE | High | Master Pro | P62259 | 14-3-3 protein epsilon [OS=Mus musculus] | 0 | 23.994 | 26 | 6 | 15 | 3 | 1 | 255 | 29.2 | 4.74 | 51.55 | 6 | 1.208 | 1.05 | |
| FALSE | High | Master Pro | P32261 | Antithrombin-III [OS=Mus musculus] | 0 | 23.178 | 16 | 9 | 23 | 9 | 1 | 465 | 52 | 6.46 | 68.54 | 9 | 0.315 | 0.196 | |
| FALSE | High | Master Pro | Q9CQV8-1 | 14-3-3 protein beta/alpha [OS=Mus musculus] | 0 | 22.817 | 26 | 6 | 17 | 2 | 1 | 246 | 28.1 | 4.83 | 61.24 | 6 | 1.392 | 1.627 | |
| FALSE | High | Master Pro | P16627 | Heat shock 70 kDa protein 1-like [OS=Mus musculus] | 0 | 22.668 | 11 | 4 | 13 | 1 | 1 | 641 | 70.6 | 6.24 | 53.95 | 4 | 0.398 | 0.203 | |
| FALSE | High | Master Pro | Q60864 | stress-induced-phosphoprotein 1 [OS=Mus musculus] | 0 | 21.795 | 8 | 6 | 11 | 6 | 1 | 543 | 62.5 | 6.8 | 43.05 | 6 | 1.134 | 1.46 | |
| FALSE | High | Master Pro | P17751 | Triosephosphate isomerase [OS=Mus musculus] | 0 | 21.404 | 17 | 5 | 19 | 5 | 1 | 299 | 32.2 | 5.74 | 69.32 | 5 | 2.904 | 5.766 | |
| FALSE | High | Master Pro | Q93092 | Transaldolase [OS=Mus musculus] | 0 | 21.211 | 19 | 7 | 17 | 7 | 1 | 337 | 37.4 | | | | | | |

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|-------|------|---------------------|---|---|--------|----|---|-----|---|---|------|-------|-------|--------|---|-------|-------|
| FALSE | High | Master Pro P20029 | 78 kDa glucose-regulated protein [OS=Mus musculus] | 0 | 19.584 | 5 | 3 | 10 | 1 | 1 | 655 | 72.4 | 5.16 | 44.78 | 3 | 0.376 | 0.265 |
| FALSE | High | Master Pro P50396 | Rab GDP dissociation inhibitor alpha [OS=Mus musculus] | 0 | 19.466 | 13 | 5 | 8 | 3 | 1 | 447 | 50.5 | 5.08 | 27.29 | 5 | 2.205 | 1.682 |
| FALSE | High | Master Pro Q92204 | Heterogeneous nuclear ribonucleoproteins C1/C2 [OS=Mus musculus] | 0 | 19.302 | 15 | 4 | 8 | 4 | 1 | 313 | 34.4 | 5.05 | 35.37 | 4 | 1.513 | 2.478 |
| FALSE | High | Master Pro P13020-1 | Gelsolin [OS=Mus musculus] | 0 | 18.991 | 13 | 9 | 23 | 9 | 1 | 780 | 85.9 | 6.18 | 65.04 | 9 | 0.51 | 0.41 |
| FALSE | High | Master Pro Q60668-1 | heterogeneous nuclear ribonucleoprotein D0 [OS=Mus musculus] | 0 | 18.914 | 12 | 4 | 7 | 3 | 1 | 355 | 38.3 | 7.81 | 23.83 | 4 | 5.072 | 4.098 |
| FALSE | High | Master Pro Q61553 | Fascin [OS=Mus musculus] | 0 | 18.859 | 10 | 4 | 14 | 4 | 1 | 493 | 54.5 | 6.89 | 56.49 | 4 | 2.059 | 3.991 |
| FALSE | High | Master Pro P17710-1 | Hexokinase-1 [OS=Mus musculus] | 0 | 18.788 | 7 | 7 | 10 | 7 | 1 | 974 | 108.2 | 6.8 | 31.7 | 7 | 0.846 | 1.107 |
| FALSE | High | Master Pro Q6GQT1 | Alpha-2-macroglobulin-P [OS=Mus musculus] | 0 | 18.785 | 2 | 3 | 22 | 2 | 1 | 1474 | 164.2 | 6.61 | 73.42 | 3 | 0.362 | 0.22 |
| FALSE | High | Master Pro P62908 | 40S ribosomal protein S3 [OS=Mus musculus] | 0 | 18.757 | 23 | 6 | 14 | 6 | 1 | 243 | 26.7 | 9.66 | 45.78 | 6 | 1.831 | 1.904 |
| FALSE | High | Master Pro P10649 | Glutathione S-transferase Mu 1 [OS=Mus musculus] | 0 | 18.605 | 23 | 5 | 16 | 2 | 1 | 218 | 26 | 7.94 | 57.61 | 5 | 2.614 | 4.41 |
| FALSE | High | Master Pro P99024 | tubulin beta-5 chain [OS=Mus musculus] | 0 | 18.548 | 20 | 7 | 12 | 7 | 1 | 444 | 49.6 | 4.89 | 38.22 | 7 | 0.678 | 0.471 |
| FALSE | High | Master Pro P24527 | leukotriene A-4 hydrolase [OS=Mus musculus] | 0 | 18.241 | 6 | 4 | 10 | 4 | 1 | 611 | 69 | 6.42 | 36.68 | 4 | 1.681 | 1.407 |
| FALSE | High | Master Pro Q04857 | Collagen alpha-1(VI) chain [OS=Mus musculus] | 0 | 17.902 | 5 | 4 | 11 | 4 | 1 | 1025 | 108.4 | 5.36 | 37.8 | 4 | 0.628 | 0.729 |
| FALSE | High | Master Pro P18242 | Cathepsin D [OS=Mus musculus] | 0 | 17.858 | 17 | 5 | 9 | 5 | 1 | 410 | 44.9 | 7.15 | 33.76 | 5 | 1.313 | 1.306 |
| FALSE | High | Master Pro Q99K51 | Plastin-3 [OS=Mus musculus] | 0 | 17.561 | 9 | 5 | 10 | 3 | 1 | 630 | 70.7 | 5.62 | 36.31 | 5 | 0.762 | 0.819 |
| FALSE | High | Master Pro Q9CZD3 | Glycine-tRNA ligase [OS=Mus musculus] | 0 | 17.534 | 8 | 5 | 12 | 5 | 1 | 729 | 81.8 | 6.65 | 33.51 | 5 | 1.972 | 1.291 |
| FALSE | High | Master Pro P68254-1 | 14-3-3 protein theta [OS=Mus musculus] | 0 | 17.51 | 21 | 5 | 15 | 1 | 1 | 245 | 27.8 | 4.78 | 53.21 | 5 | 1.731 | 1.757 |
| FALSE | High | Master Pro Q00493 | Carboxypeptidase E [OS=Mus musculus] | 0 | 17.46 | 9 | 3 | 9 | 3 | 1 | 476 | 53.2 | 5.19 | 37.97 | 3 | 0.971 | 0.777 |
| FALSE | High | Master Pro P15116 | Cadherin-2 [OS=Mus musculus] | 0 | 17.422 | 6 | 4 | 7 | 4 | 1 | 906 | 99.7 | 4.78 | 26.23 | 4 | 0.626 | 0.516 |
| FALSE | High | Master Pro P43277 | Histone H1.3 [OS=Mus musculus] | 0 | 17.402 | 14 | 4 | 8 | 3 | 1 | 221 | 22.1 | 11.03 | 29.78 | 4 | 1.792 | 2.131 |
| FALSE | High | Master Pro Q91ZK7 | prolow-density lipoprotein receptor-related protein 1 [OS=Mus musculus] | 0 | 16.975 | 1 | 5 | 8 | 5 | 1 | 4545 | 504.4 | 5.36 | 19.79 | 5 | 0.475 | 0.413 |
| FALSE | High | Master Pro Q6P1B1 | xxa-Pro aminopeptidase 1 [OS=Mus musculus] | 0 | 16.921 | 8 | 3 | 6 | 3 | 1 | 623 | 69.5 | 5.54 | 23.46 | 3 | 1.101 | 1.158 |
| FALSE | High | Master Pro P17742 | peptidyl-prolyl cis-trans isomerase A [OS=Mus musculus] | 0 | 16.884 | 28 | 5 | 34 | 5 | 1 | 164 | 18 | 7.9 | 110.96 | 5 | 3.269 | 2.981 |
| FALSE | High | Master Pro Q9CPU0 | lactoylglutathione lyase [OS=Mus musculus] | 0 | 16.871 | 23 | 6 | 18 | 6 | 1 | 184 | 20.8 | 5.47 | 59.73 | 6 | 3.214 | 1.344 |
| FALSE | High | Master Pro P62827 | GTP-binding nuclear protein RAN [OS=Mus musculus] | 0 | 16.869 | 20 | 3 | 5 | 3 | 1 | 216 | 24.4 | 7.49 | 19.06 | 3 | 1.666 | 1.481 |
| FALSE | High | Master Pro P19324 | Serpin H1 [OS=Mus musculus] | 0 | 16.823 | 12 | 5 | 9 | 5 | 1 | 417 | 46.5 | 8.82 | 31.93 | 5 | 1.006 | 0.805 |
| FALSE | High | Master Pro Q88G05 | Heterogeneous nuclear ribonucleoprotein A3 [OS=Mus musculus] | 0 | 16.653 | 6 | 3 | 9 | 1 | 1 | 379 | 39.6 | 9.01 | 33.45 | 3 | 1.038 | 0.988 |
| FALSE | High | Master Pro P68510 | 14-3-3 protein eta [OS=Mus musculus] | 0 | 16.587 | 17 | 5 | 17 | 1 | 1 | 246 | 28.2 | 4.89 | 59.37 | 5 | 2.04 | 1.476 |
| FALSE | High | Master Pro P26043 | radixin [OS=Mus musculus] | 0 | 16.521 | 11 | 7 | 13 | 2 | 1 | 583 | 68.5 | 6.2 | 43.24 | 7 | 1.292 | 1.235 |
| FALSE | High | Master Pro P51885 | Lumican [OS=Mus musculus] | 0 | 16.356 | 17 | 5 | 13 | 5 | 1 | 338 | 38.2 | 6.43 | 47.21 | 5 | 0.364 | 0.23 |
| FALSE | High | Master Pro P57776-3 | Isoform 3 of Elongation factor 1-delta [OS=Mus musculus] | 0 | 16.32 | 4 | 2 | 7 | 2 | 1 | 660 | 72.9 | 6.43 | 29.37 | 2 | 0.782 | 1.261 |
| FALSE | High | Master Pro P39061-3 | Collagen alpha-1(XVIII) chain [OS=Mus musculus] | 0 | 16.292 | 3 | 6 | 15 | 6 | 1 | 1774 | 182.1 | 5.62 | 46.76 | 6 | 0.935 | 1.182 |
| FALSE | High | Master Pro P30412 | peptidyl-prolyl cis-trans isomerase C [OS=Mus musculus] | 0 | 16.284 | 17 | 3 | 7 | 3 | 1 | 212 | 22.8 | 7.5 | 21.67 | 3 | 0.776 | 0.776 |
| FALSE | High | Master Pro P15626 | Glutathione S-transferase Mu 2 [OS=Mus musculus] | 0 | 16.26 | 23 | 4 | 9 | 1 | 1 | 218 | 25.7 | 7.39 | 33.94 | 4 | 0.349 | 0.432 |
| FALSE | High | Master Pro P80315 | T-complex protein 1 subunit delta [OS=Mus musculus] | 0 | 16.246 | 10 | 3 | 4 | 3 | 1 | 539 | 58 | 8.02 | 15.57 | 3 | 0.663 | 0.455 |
| FALSE | High | Master Pro P14206 | 40S ribosomal protein SA [OS=Mus musculus] | 0 | 16.091 | 17 | 5 | 6 | 5 | 1 | 295 | 32.8 | 4.87 | 22.24 | 5 | 1.252 | 1.061 |
| FALSE | High | Master Pro Q61316 | Heat shock 70 kDa protein 4 [OS=Mus musculus] | 0 | 16.089 | 4 | 3 | 8 | 3 | 1 | 841 | 94.1 | 5.24 | 31.23 | 4 | 1.071 | 1.165 |
| FALSE | High | Master Pro Q91VM5 | RNA binding motif protein, X-linked-like-1 [OS=Mus musculus] | 0 | 16.048 | 10 | 4 | 9 | 4 | 1 | 388 | 42.1 | 9.99 | 31.49 | 4 | 1.821 | 1.641 |
| FALSE | High | Master Pro Q922R8 | Protein disulfide-isomerase A6 [OS=Mus musculus] | 0 | 15.839 | 7 | 2 | 4 | 2 | 1 | 440 | 48.1 | 5.14 | 13.45 | 2 | 0.411 | 0.329 |
| FALSE | High | Master Pro Q60605 | Myosin light polypeptide 6 [OS=Mus musculus] | 0 | 15.68 | 25 | 3 | 8 | 3 | 1 | 151 | 16.9 | 4.65 | 26.86 | 3 | 1.128 | 1.089 |
| FALSE | High | Master Pro P10639 | thioredoxin [OS=Mus musculus] | 0 | 15.323 | 21 | 2 | 9 | 2 | 1 | 105 | 11.7 | 4.92 | 44.43 | 2 | 1.414 | 1.453 |
| FALSE | High | Master Pro P16045 | Galectin-1 [OS=Mus musculus] | 0 | 15.04 | 30 | 4 | 11 | 4 | 1 | 135 | 14.9 | 5.49 | 35.77 | 4 | 2.722 | 3.159 |
| FALSE | High | Master Pro Q9CR57 | 60S ribosomal protein L14 [OS=Mus musculus] | 0 | 14.814 | 12 | 2 | 5 | 2 | 1 | 217 | 23.5 | 11.02 | 23.51 | 2 | 0.498 | 0.48 |
| FALSE | High | Master Pro Q6ZWN5 | 40S ribosomal protein S9 [OS=Mus musculus] | 0 | 14.652 | 23 | 5 | 9 | 5 | 1 | 194 | 22.6 | 10.65 | 29.22 | 5 | 2.994 | 3.308 |
| FALSE | High | Master Pro P06801 | NADP-dependent malic enzyme [OS=Mus musculus] | 0 | 14.516 | 10 | 4 | 6 | 4 | 1 | 572 | 63.9 | 7.44 | 15.73 | 4 | 0.631 | 0.426 |
| FALSE | High | Master Pro P33434-1 | 72 kDa type IV collagenase [OS=Mus musculus] | 0 | 14.437 | 9 | 6 | 8 | 6 | 1 | 662 | 74.1 | 5.53 | 24.84 | 6 | 0.664 | 0.708 |
| FALSE | High | Master Pro Q9D0F9 | Phosphoglucomutase-1 [OS=Mus musculus] | 0 | 14.399 | 10 | 5 | 8 | 5 | 1 | 562 | 61.4 | 6.57 | 26.94 | 5 | 0.856 | 0.75 |
| FALSE | High | Master Pro P48036 | annexin A5 [OS=Mus musculus] | 0 | 14.367 | 14 | 4 | 9 | 4 | 1 | 319 | 35.7 | 4.96 | 29.55 | 4 | 2.519 | 1.699 |
| FALSE | High | Master Pro P68040 | Receptor of activated protein C kinase 1 [OS=Mus musculus] | 0 | 14.207 | 14 | 5 | 12 | 5 | 1 | 317 | 35.1 | 7.69 | 35.75 | 5 | 2.082 | 2.088 |
| FALSE | High | Master Pro Q70251 | Elongation factor 1-beta [OS=Mus musculus] | 0 | 13.931 | 17 | 4 | 13 | 4 | 1 | 225 | 24.7 | 4.69 | 39.29 | 4 | 1.733 | 2.458 |
| FALSE | High | Master Pro Q91I16 | alcohol dehydrogenase [NADP(+)] [OS=Mus musculus] | 0 | 13.906 | 14 | 4 | 7 | 4 | 1 | 325 | 36.6 | 7.39 | 24.78 | 4 | 1.093 | 0.97 |
| FALSE | High | Master Pro P26040 | Ezrin [OS=Mus musculus] | 0 | 13.824 | 10 | 6 | 12 | 1 | 1 | 586 | 69.4 | 6.1 | 38.8 | 6 | 0.56 | 0.714 |
| FALSE | High | Master Pro Q3TCN2 | Putative phospholipase B-like 2 [OS=Mus musculus] | 0 | 13.818 | 5 | 3 | 9 | 3 | 1 | 594 | 66.2 | 6.13 | 34.86 | 3 | 1.122 | 1.112 |
| FALSE | High | Master Pro P07724 | Serum albumin [OS=Mus musculus] | 0 | 13.806 | 4 | 2 | 118 | 2 | 1 | 608 | 68.6 | 6.07 | 412.31 | 2 | 0.268 | 0.178 |
| FALSE | High | Master Pro P62301 | 40S ribosomal protein S13 [OS=Mus musculus] | 0 | 13.77 | 29 | 4 | 8 | 4 | 1 | 151 | 17.2 | 10.54 | 27.15 | 4 | 1.175 | 1.44 |
| FALSE | High | Master Pro P21460 | Cystatin C [OS=Mus musculus] | 0 | 13.737 | 25 | 4 | 7 | 4 | 1 | 140 | 15.5 | 9 | 24.31 | 4 | 1.27 | 1.561 |
| FALSE | High | Master Pro P62855 | 40S ribosomal protein S26 [OS=Mus musculus] | 0 | 13.687 | 21 | 2 | 6 | 2 | 1 | 115 | 13 | 11 | 24.23 | 2 | 1.669 | 1.815 |
| FALSE | High | Master Pro P40142 | Transketolase [OS=Mus musculus] | 0 | 13.564 | 5 | 3 | 10 | 3 | 1 | 623 | 67.6 | 7.5 | 35.43 | 3 | 0.945 | 0.591 |
| FALSE | High | Master Pro P47754 | F-actin-capping protein subunit alpha-2 [OS=Mus musculus] | 0 | 13.559 | 15 | 3 | 6 | 2 | 1 | 286 | 32.9 | 5.85 | 20.59 | 3 | 1.24 | 1.366 |
| FALSE | High | Master Pro P70168 | Importin subunit beta-1 [OS=Mus musculus] | 0 | 13.449 | 3 | 2 | 7 | 2 | 1 | 876 | 97.1 | 4.78 | 30.55 | 2 | 0.823 | 0.916 |
| FALSE | High | Master Pro P14869 | 60S acidic ribosomal protein P0 [OS=Mus musculus] | 0 | 13.334 | 11 | 3 | 5 | 3 | 1 | 317 | 34.2 | 6.25 | 16.44 | 3 | 0.657 | 0.692 |
| FALSE | High | Master Pro Q9R0P5 | Destrin [OS=Mus musculus] | 0 | 13.148 | 25 | 3 | 9 | 3 | 1 | 165 | 18.5 | 7.97 | 26.96 | 3 | 0.646 | 0.54 |
| FALSE | High | Master Pro Q9Z1N5 | spliceosome RNA helicase DDX39B [OS=Mus musculus] | 0 | 13.105 | 9 | 4 | 8 | 4 | 1 | 428 | 49 | 5.67 | 23.6 | 4 | 0.958 | 0.726 |
| FALSE | High | Master Pro Q99P71 | rho GDP-dissociation inhibitor 1 [OS=Mus musculus] | 0 | 13.101 | 11 | 2 | 8 | 2 | 1 | 204 | 23.4 | 5.2 | 26.74 | 2 | 1.034 | 0.773 |
| FALSE | High | Master Pro P62204 | Calmodulin [OS=Mus musculus] | 0 | 13.02 | 27 | 4 | 7 | 4 | 1 | 149 | 16.8 | 4.22 | 24.8 | 4 | 2.809 | 2.663 |
| FALSE | High | Master Pro Q9CPV4-1 | Glyoxalase domain-containing protein 4 [OS=Mus musculus] | 0 | 12.993 | 12 | 3 | 4 | 3 | 1 | 298 | 33.3 | 5.47 | 16.06 | 3 | 0.702 | 0.789 |
| FALSE | High | Master Pro P09103 | Protein disulfide-isomerase [OS=Mus musculus] | 0 | 12.894 | 9 | 5 | 9 | 5 | 1 | 509 | 57 | 4.88 | 30.54 | 5 | 1.224 | 1.489 |
| FALSE | High | Master Pro Q91ZJ5-1 | UTP-glucose-1-phosphate uridylyltransferase [OS=Mus musculus] | 0 | 12.811 | 9 | 4 | 9 | 4 | 1 | 508 | 56.9 | 7.61 | 27.8 | 4 | 1.462 | 1.442 |
| FALSE | High | Master Pro Q8R2Y2-1 | Cell surface glycoprotein MUC18 [OS=Mus musculus] | 0 | 12.719 | 5 | 3 | 6 | 3 | 1 | 648 | 71.5 | 5.83 | 23.08 | 3 | 1.122 | 0.376 |
| FALSE | High | Master Pro Q8K423 | NAD(P)H-hydrate epimerase [OS=Mus musculus] | 0 | 12.695 | 13 | 3 | 8 | 3 | 1 | 282 | 31 | 7.69 | 24.84 | 3 | 0.99 | 1.033 |
| FALSE | High | Master Pro P63242 | Eukaryotic translation initiation factor 5A-1 [OS=Mus musculus] | 0 | 12.553 | 33 | 4 | 14 | 4 | 1 | 154 | 16.8 | 5.24 | 33.44 | 4 | 1.641 | 1.597 |
| FALSE | High | Master Pro Q99K10 | Aconitate hydratase, mitochondrial [OS=Mus musculus] | 0 | 12.34 | 5 | 4 | 8 | 4 | 1 | 780 | 85.4 | 7.93 | 28.5 | 4 | 1.867 | 2.674 |
| FALSE | High | Master Pro Q9Z2X1-1 | Heterogeneous nuclear ribonucleoprotein F [OS=Mus musculus] | 0 | 12.083 | 8 | 2 | 3 | 1 | 1 | 415 | 45.7 | 5.49 | 10.03 | 2 | 3.265 | 5.654 |
| FALSE | High | Master Pro Q920E5 | Farnesyl pyrophosphate synthase [OS=Mus musculus] | 0 | 12.059 | 8 | 2 | 4 | 2 | 1 | 353 | 40.6 | 5.66 | 14.02 | 2 | 0.888 | 1.297 |
| FALSE | High | Master Pro Q9CWX9 | bifunctional purine biosynthesis protein purH [OS=Mus musculus] | 0 | 12.046 | 3 | 3 | 5 | 3 | 1 | 592 | 64.2 | 6.76 | 18.24 | 3 | 0.556 | 0.627 |
| FALSE | High | Master Pro P50580 | proliferation-associated protein 2G4 [OS=Mus musculus] | 0 | 11.912 | 8 | 3 | 7 | 3 | 1 | 394 | 43.7 | 6.86 | 26.93 | 3 | 2.511 | 2.117 |
| FALSE | High | Master Pro Q64674 | spermidine synthase [OS=Mus musculus] | 0 | 11.838 | 10 | 3 | 7 | 3 | 1 | 302 | 34 | 5.5 | 23.99 | 3 | 0.887 | 0.925 |
| FALSE | High | Master Pro Q8CGC7 | Bifunctional glutamate/proline-tRNA ligase [OS=Mus musculus] | 0 | 11.544 | 4 | 5 | 6 | 5 | 1 | 1512 | 170 | 7.66 | 12.79 | 5 | 1.318 | 1.679 |

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|-------|------|---------------------|--|---|--------|----|---|----|---|---|------|-------|-------|-------|---|-------|-------|
| FALSE | High | Master Pro P22777 | Plasminogen activator inhibitor 1 [OS=Mus musculus] | 0 | 11.226 | 10 | 3 | 5 | 3 | 1 | 402 | 45.1 | 6.64 | 18 | 3 | 0.556 | 0.45 |
| FALSE | High | Master Pro Q8VEK3 | Heterogeneous nuclear ribonucleoprotein U [OS=Mus musculus] | 0 | 11.19 | 4 | 3 | 5 | 3 | 1 | 800 | 87.9 | 6.24 | 17 | 3 | 1.193 | 2.216 |
| FALSE | High | Master Pro Q9WUM4 | coronin-1C [OS=Mus musculus] | 0 | 11.189 | 7 | 3 | 5 | 3 | 1 | 474 | 53.1 | 7.08 | 17.93 | 3 | 1.182 | 1.022 |
| FALSE | High | Master Pro Q99K85 | phosphoserine aminotransferase [OS=Mus musculus] | 0 | 11.14 | 15 | 4 | 4 | 4 | 1 | 370 | 40.4 | 8.03 | 11.19 | 4 | 0.634 | 0.577 |
| FALSE | High | Master Pro Q9Z175 | Lysyl oxidase homolog 3 [OS=Mus musculus] | 0 | 10.979 | 6 | 4 | 5 | 4 | 1 | 754 | 83.7 | 7.01 | 19.05 | 4 | 0.511 | 0.879 |
| FALSE | High | Master Pro Q00612 | Glucose-6-phosphate 1-dehydrogenase X [OS=Mus musculus] | 0 | 10.86 | 5 | 2 | 4 | 2 | 1 | 515 | 59.2 | 6.49 | 14.96 | 2 | 1.481 | 0.992 |
| FALSE | High | Master Pro P62900 | 60S ribosomal protein L31 [OS=Mus musculus] | 0 | 10.816 | 18 | 2 | 7 | 2 | 1 | 125 | 14.5 | 10.54 | 25.37 | 2 | 1.528 | 1.986 |
| FALSE | High | Master Pro Q7TQ03 | Ubiquitin thioesterase otub1 [OS=Mus musculus] | 0 | 10.756 | 12 | 3 | 6 | 3 | 1 | 271 | 31.3 | 4.94 | 22.35 | 3 | 1.108 | 1.888 |
| FALSE | High | Master Pro Q6ZQ38 | cullin-associated nedd8-dissociated protein 1 [OS=Mus musculus] | 0 | 10.753 | 3 | 3 | 4 | 3 | 1 | 1230 | 136.2 | 5.78 | 12.17 | 3 | 0.856 | 0.948 |
| FALSE | High | Master Pro Q68FD5 | Clathrin heavy chain 1 [OS=Mus musculus] | 0 | 10.646 | 3 | 5 | 8 | 5 | 1 | 1675 | 191.4 | 5.69 | 21.14 | 5 | 0.535 | 0.613 |
| FALSE | High | Master Pro Q9D1A2 | cytosolic non-specific dipeptidase [OS=Mus musculus] | 0 | 10.569 | 13 | 4 | 7 | 4 | 1 | 475 | 52.7 | 5.66 | 26.48 | 4 | 0.569 | 0.429 |
| FALSE | High | Master Pro Q99KC8 | von Willebrand factor A domain-containing protein 5A [OS=Mus musculus] | 0 | 10.519 | 4 | 3 | 7 | 3 | 1 | 793 | 87.1 | 6.58 | 26.04 | 3 | 2.451 | 2.228 |
| FALSE | High | Master Pro P54728 | UV excision repair protein RAD23 homolog B [OS=Mus musculus] | 0 | 10.414 | 8 | 4 | 7 | 4 | 1 | 416 | 43.5 | 4.83 | 24.51 | 4 | 1.072 | 1.52 |
| FALSE | High | Master Pro Q5X1Y5 | Coatomer subunit delta [OS=Mus musculus] | 0 | 10.296 | 4 | 2 | 7 | 2 | 1 | 511 | 57.2 | 6.21 | 24.81 | 2 | 1.135 | 1.418 |
| FALSE | High | Master Pro P24369 | peptidyl-prolyl cis-trans isomerase B [OS=Mus musculus] | 0 | 10.239 | 16 | 4 | 9 | 4 | 1 | 216 | 23.7 | 9.55 | 31.27 | 4 | 2.971 | 2.249 |
| FALSE | High | Master Pro P60843 | Eukaryotic initiation factor 4A-II [OS=Mus musculus] | 0 | 10.122 | 7 | 4 | 5 | 1 | 1 | 406 | 46.1 | 5.48 | 15.41 | 4 | 0.736 | 0.59 |
| FALSE | High | Master Pro P99027 | 60S acidic ribosomal protein P2 [OS=Mus musculus] | 0 | 9.854 | 27 | 2 | 4 | 2 | 1 | 115 | 11.6 | 4.54 | 11.68 | 2 | 0.813 | 1.072 |
| FALSE | High | Master Pro Q61990 | Poly(rC)-binding protein 2 [OS=Mus musculus] | 0 | 9.798 | 6 | 2 | 6 | 1 | 1 | 362 | 38.2 | 6.79 | 18.49 | 2 | 1.36 | 2.288 |
| FALSE | High | Master Pro P61089 | ubiquitin-conjugating enzyme E2 N [OS=Mus musculus] | 0 | 9.783 | 16 | 2 | 10 | 2 | 1 | 152 | 17.1 | 6.57 | 35.43 | 2 | 1.364 | 2.013 |
| FALSE | High | Master Pro P19096 | Fatty acid synthase [OS=Mus musculus] | 0 | 9.593 | 1 | 4 | 6 | 4 | 1 | 2504 | 272.3 | 6.58 | 18.91 | 4 | 1.268 | 1.21 |
| FALSE | High | Master Pro Q9D8E6 | 60S ribosomal protein L4 [OS=Mus musculus] | 0 | 9.585 | 6 | 2 | 2 | 2 | 1 | 419 | 47.1 | 11 | 10.19 | 2 | 0.659 | 0.795 |
| FALSE | High | Master Pro Q61035 | Histidine-tRNA ligase, cytoplasmic [OS=Mus musculus] | 0 | 9.571 | 4 | 2 | 7 | 2 | 1 | 509 | 57.4 | 6 | 22.62 | 2 | 1.589 | 1.213 |
| FALSE | High | Master Pro Q99KK7 | dipeptidyl peptidase 3 [OS=Mus musculus] | 0 | 9.569 | 7 | 3 | 3 | 3 | 1 | 738 | 82.8 | 5.38 | 10.65 | 3 | 0.459 | 0.439 |
| FALSE | High | Master Pro Q61581 | Insulin-like growth factor-binding protein 7 [OS=Mus musculus] | 0 | 9.498 | 7 | 2 | 6 | 2 | 1 | 281 | 29 | 8.31 | 23.92 | 2 | 0.361 | 0.279 |
| FALSE | High | Master Pro Q35737 | Heterogeneous nuclear ribonucleoprotein H [OS=Mus musculus] | 0 | 9.332 | 8 | 2 | 7 | 2 | 1 | 449 | 49.2 | 6.3 | 7.46 | 2 | 0.43 | 0.445 |
| FALSE | High | Master Pro P05202 | Aspartate aminotransferase, mitochondrial [OS=Mus musculus] | 0 | 9.294 | 7 | 3 | 7 | 3 | 1 | 430 | 47.4 | 9 | 17.06 | 3 | 0.839 | 0.618 |
| FALSE | High | Master Pro P10630-2 | Isoform 2 of Eukaryotic initiation factor 4A-II [OS=Mus musculus] | 0 | 9.235 | 7 | 4 | 4 | 1 | 1 | 408 | 46.5 | 5.48 | 11.24 | 4 | 1.368 | 1.738 |
| FALSE | High | Master Pro P97855 | Ras GTPase-activating protein-binding protein 1 [OS=Mus musculus] | 0 | 9.162 | 6 | 2 | 5 | 2 | 1 | 465 | 51.8 | 5.59 | 17.32 | 2 | 0.617 | 0.675 |
| FALSE | High | Master Pro P62983 | Ubiquitin-40S ribosomal protein S27a [OS=Mus musculus] | 0 | 9.095 | 22 | 3 | 7 | 3 | 1 | 156 | 17.9 | 9.64 | 27.96 | 3 | 1.915 | 1.986 |
| FALSE | High | Master Pro Q9D819 | Inorganic pyrophosphatase [OS=Mus musculus] | 0 | 9.062 | 9 | 3 | 7 | 3 | 1 | 289 | 32.6 | 5.6 | 23.87 | 3 | 1.299 | 2.162 |
| FALSE | High | Master Pro Q11011 | puromycin-sensitive aminopeptidase [OS=Mus musculus] | 0 | 9.056 | 4 | 4 | 6 | 4 | 1 | 920 | 103.3 | 5.9 | 21.17 | 4 | 1.252 | 1.525 |
| FALSE | High | Master Pro Q60854 | serpin B6 [OS=Mus musculus] | 0 | 9.048 | 9 | 2 | 4 | 2 | 1 | 378 | 42.6 | 5.74 | 12.46 | 2 | 1.031 | 0.615 |
| FALSE | High | Master Pro P62821 | Ras-related protein Rab-1A [OS=Mus musculus] | 0 | 9.041 | 13 | 2 | 5 | 2 | 1 | 205 | 22.7 | 6.21 | 19.57 | 2 | 0.654 | 0.71 |
| FALSE | High | Master Pro P46638 | Ras-related protein Rab-11B [OS=Mus musculus] | 0 | 9.024 | 15 | 3 | 6 | 3 | 1 | 218 | 24.5 | 5.94 | 19.06 | 3 | 0.765 | 0.794 |
| FALSE | High | Master Pro P70296 | phosphatidylethanolamine-binding protein 1 [OS=Mus musculus] | 0 | 8.867 | 17 | 3 | 3 | 3 | 1 | 187 | 20.8 | 5.4 | 11.23 | 3 | 1.851 | 2.28 |
| FALSE | High | Master Pro Q9CR86 | Tubulin polymerization-promoting protein family member 3 [OS=Mus musculus] | 0 | 8.868 | 13 | 2 | 4 | 2 | 1 | 176 | 19 | 9.11 | 13.15 | 2 | 2.165 | 0.776 |
| FALSE | High | Master Pro P62852 | 40S ribosomal protein S25 [OS=Mus musculus] | 0 | 8.532 | 22 | 3 | 5 | 3 | 1 | 125 | 13.7 | 10.11 | 18.92 | 3 | 1.682 | 1.709 |
| FALSE | High | Master Pro Q3UGR5-1 | haloacid dehalogenase-like hydrolase domain-containing protein 2 [OS=Mus musculus] | 0 | 8.527 | 8 | 2 | 4 | 2 | 1 | 259 | 28.7 | 6.05 | 16.04 | 2 | 1.252 | 0.814 |
| FALSE | High | Master Pro P14131 | 40S ribosomal protein S16 [OS=Mus musculus] | 0 | 8.493 | 12 | 2 | 8 | 2 | 1 | 146 | 16.4 | 10.21 | 30.49 | 2 | 1.73 | 2.081 |
| FALSE | High | Master Pro Q3U114 | DNA damage-binding protein 1 [OS=Mus musculus] | 0 | 8.469 | 3 | 4 | 8 | 4 | 1 | 1140 | 126.8 | 5.26 | 24.14 | 4 | 1.21 | 1.078 |
| FALSE | High | Master Pro Q9CQ60 | 6-phosphogluconolactonase [OS=Mus musculus] | 0 | 8.288 | 9 | 2 | 5 | 2 | 1 | 257 | 27.2 | 5.85 | 18.63 | 2 | 3.98 | 3.859 |
| FALSE | High | Master Pro Q91W90 | Thioredoxin domain-containing protein 5 [OS=Mus musculus] | 0 | 8.009 | 8 | 3 | 4 | 3 | 1 | 417 | 46.4 | 5.78 | 13.01 | 3 | 1.421 | 1.123 |
| FALSE | High | Master Pro Q06138 | Calcium-binding protein 39 [OS=Mus musculus] | 0 | 7.962 | 10 | 4 | 6 | 4 | 1 | 341 | 39.8 | 6.89 | 16.14 | 4 | 1.21 | 1.235 |
| FALSE | High | Master Pro P17918 | proliferating cell nuclear antigen [OS=Mus musculus] | 0 | 7.956 | 11 | 2 | 5 | 2 | 1 | 261 | 28.8 | 4.77 | 15.79 | 2 | 1.994 | 1.555 |
| FALSE | High | Master Pro Q9Z247 | Peptidyl-prolyl cis-trans isomerase FKBP9 [OS=Mus musculus] | 0 | 7.892 | 7 | 2 | 2 | 2 | 1 | 570 | 63 | 5.21 | 7.38 | 2 | 0.55 | 0.343 |
| FALSE | High | Master Pro P68037 | Ubiquitin-conjugating enzyme E2 L3 [OS=Mus musculus] | 0 | 7.8 | 16 | 2 | 4 | 2 | 1 | 154 | 17.9 | 8.51 | 8.33 | 2 | 0.871 | 0.781 |
| FALSE | High | Master Pro Q9Z1Q9 | Valine-tRNA ligase [OS=Mus musculus] | 0 | 7.799 | 2 | 3 | 6 | 3 | 1 | 1263 | 140.1 | 7.77 | 17.11 | 3 | 1.009 | 0.931 |
| FALSE | High | Master Pro Q69ZM7-1 | Myoferlin [OS=Mus musculus] | 0 | 7.695 | 1 | 2 | 2 | 2 | 1 | 2048 | 233.2 | 6.16 | 8.14 | 2 | 1.248 | 2.322 |
| FALSE | High | Master Pro Q08807 | Peroxioredoxin-4 [OS=Mus musculus] | 0 | 7.598 | 11 | 3 | 5 | 1 | 1 | 274 | 31 | 7.15 | 17.49 | 3 | 1.722 | 1.776 |
| FALSE | High | Master Pro Q61033-1 | Lamina-associated polypeptide 2, isoforms alpha/zeta [OS=Mus musculus] | 0 | 7.57 | 4 | 2 | 2 | 2 | 1 | 693 | 75.1 | 8.05 | 5.93 | 2 | 0.203 | 0.316 |
| FALSE | High | Master Pro P01029 | Complement C4-B [OS=Mus musculus] | 0 | 7.52 | 1 | 3 | 4 | 3 | 1 | 1738 | 192.8 | 7.53 | 13.95 | 3 | 0.379 | 0.219 |
| FALSE | High | Master Pro Q9R0Y5-2 | Isoform 2 of Adenylate kinase isoenzyme 1 [OS=Mus musculus] | 0 | 7.49 | 12 | 2 | 3 | 2 | 1 | 210 | 23.1 | 5.83 | 8.51 | 2 | 0.733 | 0.674 |
| FALSE | High | Master Pro Q61RU2 | Tropomyosin alpha-4 chain [OS=Mus musculus] | 0 | 7.454 | 10 | 3 | 6 | 3 | 1 | 248 | 28.5 | 4.68 | 18.25 | 3 | 0.679 | 0.699 |
| FALSE | High | Master Pro Q61233 | Plastin-2 [OS=Mus musculus] | 0 | 7.425 | 4 | 3 | 7 | 1 | 1 | 627 | 70.1 | 5.33 | 23.79 | 3 | 0.455 | 0.383 |
| FALSE | High | Master Pro Q9WU78-3 | Isoform 3 of Programmed cell death 6-interacting protein [OS=Mus musculus] | 0 | 7.314 | 2 | 2 | 3 | 2 | 1 | 874 | 96.7 | 6.52 | 9.98 | 2 | 0.606 | 0.867 |
| FALSE | High | Master Pro Q9D154 | Leukocyte elastase inhibitor A [OS=Mus musculus] | 0 | 7.229 | 7 | 2 | 2 | 2 | 1 | 379 | 42.5 | 6.21 | 7.32 | 2 | 0.493 | 1.625 |
| FALSE | High | Master Pro P37804 | transgelin [OS=Mus musculus] | 0 | 7.226 | 15 | 3 | 3 | 3 | 1 | 201 | 22.6 | 8.81 | 7.7 | 3 | 0.063 | 0.049 |
| FALSE | High | Master Pro Q88FR4 | N-acetylglucosamine-6-sulfatase [OS=Mus musculus] | 0 | 7.22 | 6 | 3 | 4 | 3 | 1 | 544 | 61.1 | 8.24 | 8.77 | 3 | 0.81 | 1.042 |
| FALSE | High | Master Pro P07214 | Sparc [OS=Mus musculus] | 0 | 7.21 | 8 | 2 | 5 | 2 | 1 | 302 | 34.4 | 4.86 | 16.88 | 2 | 0.64 | 0.394 |
| FALSE | High | Master Pro P63280 | SUMO-conjugating enzyme ubc9 [OS=Mus musculus] | 0 | 7.13 | 14 | 2 | 3 | 2 | 1 | 158 | 18 | 8.66 | 12.26 | 2 | 7.995 | 5.828 |
| FALSE | High | Master Pro P63001 | Ras-related C3 botulinum toxin substrate 1 [OS=Mus musculus] | 0 | 7.119 | 12 | 3 | 3 | 3 | 1 | 192 | 21.4 | 8.5 | 11.91 | 3 | 0.819 | 0.772 |
| FALSE | High | Master Pro P23927 | Alpha-crystallin B chain [OS=Mus musculus] | 0 | 7.045 | 11 | 2 | 3 | 2 | 1 | 175 | 20.1 | 7.33 | 10.11 | 2 | 0.955 | 0.286 |
| FALSE | High | Master Pro Q9CQE8 | UPF0568 protein C14orf166 homolog [OS=Mus musculus] | 0 | 7.036 | 10 | 2 | 2 | 2 | 1 | 244 | 28.1 | 6.89 | 6.02 | 2 | 1.009 | 1.166 |
| FALSE | High | Master Pro A6X935-1 | inter alpha-trypsin inhibitor, heavy chain 4 [OS=Mus musculus] | 0 | 6.946 | 2 | 2 | 9 | 2 | 1 | 942 | 104.6 | 6.4 | 31.46 | 2 | 0.353 | 0.229 |
| FALSE | High | Master Pro Q8R081 | Heterogeneous nuclear ribonucleoprotein L [OS=Mus musculus] | 0 | 6.868 | 4 | 2 | 4 | 2 | 1 | 586 | 63.9 | 8.1 | 10.56 | 2 | 2.864 | 3.207 |
| FALSE | High | Master Pro Q61398 | Procollagen C-endopeptidase enhancer 1 [OS=Mus musculus] | 0 | 6.851 | 5 | 2 | 4 | 2 | 1 | 468 | 50.1 | 8.41 | 11.68 | 2 | 0.771 | 0.811 |
| FALSE | High | Master Pro P60335 | Poly(RC)-binding protein 1 [OS=Mus musculus] | 0 | 6.781 | 6 | 2 | 6 | 1 | 1 | 356 | 37.5 | 7.09 | 17.36 | 2 | 3.412 | 4.643 |
| FALSE | High | Master Pro P61961 | Ubiquitin-fold modifier 1 [OS=Mus musculus] | 0 | 6.766 | 51 | 2 | 3 | 2 | 1 | 85 | 9.1 | 9.31 | 12.5 | 2 | 0.593 | 0.499 |
| FALSE | High | Master Pro Q08997 | Copper transport protein ATOX1 [OS=Mus musculus] | 0 | 6.727 | 32 | 2 | 6 | 2 | 1 | 68 | 7.3 | 6.51 | 20.4 | 2 | 1.727 | 1.823 |
| FALSE | High | Master Pro P62897 | Cytochrome c, somatic [OS=Mus musculus] | 0 | 6.677 | 18 | 2 | 7 | 2 | 1 | 105 | 11.6 | 9.58 | 25.33 | 2 | 6.587 | 5.559 |
| FALSE | High | Master Pro P63323 | 40S ribosomal protein S12 [OS=Mus musculus] | 0 | 6.624 | 20 | 3 | 6 | 3 | 1 | 132 | 14.5 | 7.24 | 17.84 | 3 | 1.049 | 1.122 |
| FALSE | High | Master Pro P62849-1 | 40S ribosomal protein S24 [OS=Mus musculus] | 0 | 6.611 | 26 | 3 | 3 | 3 | 1 | 133 | 15.4 | 10.78 | 8.44 | 3 | 0.807 | 0.795 |
| FALSE | High | Master Pro Q9D019 | arginine-tRNA ligase, cytoplasmic [OS=Mus musculus] | 0 | 6.535 | 5 | 2 | 2 | 2 | 1 | 660 | 75.6 | 7.55 | 8.25 | 2 | 1.263 | 1.262 |
| FALSE | High | Master Pro Q08553 | Dihydropyrimidinase-related protein 2 [OS=Mus musculus] | 0 | 6.508 | 3 | 2 | 7 | 2 | 1 | 572 | 62.2 | 6.38 | 22.03 | 2 | 0.68 | 0.694 |
| FALSE | High | Master Pro Q08638-1 | Myosin-11 [OS=Mus musculus] | 0 | 6.482 | 2 | 3 | 5 | 1 | 1 | 1972 | 226.9 | 5.45 | 12.47 | 3 | | |
| FALSE | High | Master Pro P16858 | glyceraldehyde-3-phosphate dehydrogenase [OS=Mus musculus] | 0 | 6.331 | 9 | 2 | 3 | 2 | 1 | 333 | 35.8 | 8.25 | 8.7 | 2 | 1.306 | 1.243 |
| FALSE | High | Master Pro Q9Z2T6 | Keratin, type II cuticular Hb5 [OS=Mus musculus] | 0 | 6.276 | 4 | 2 | 3 | 2 | 1 | 507 | 55.7 | 6.42 | 7.99 | 2 | 0.258 | 0.424 |
| FALSE | High | Master Pro Q8R3Q6 | coiled-coil domain-containing protein 58 [OS=Mus musculus] | 0 | 6.224 | 18 | 2 | 2 | 2 | 1 | 144 | 16.7 | 8.16 | 6.04 | 2 | 0.393 | 0.236 |
| FALSE | High | Master Pro P45 | | | | | | | | | | | | | | | |

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|-------|------|---------------------|---|---|-------|----|---|---|---|---|------|-------|-------|-------|---|-------|--------|
| FALSE | High | Master Pro P02088 | Hemoglobin subunit beta-1 [OS=Mus musculus] | 0 | 6.18 | 13 | 2 | 9 | 2 | 1 | 147 | 15.8 | 7.65 | 27.25 | 2 | 0.329 | 0.253 |
| FALSE | High | Master Pro Q9CPY7-1 | cytosol aminopeptidase [OS=Mus musculus] | 0 | 6.163 | 5 | 3 | 7 | 3 | 1 | 519 | 56.1 | 7.72 | 19.81 | 3 | 1.238 | 1.229 |
| FALSE | High | Master Pro Q921Q5 | chloride intracellular channel protein 1 [OS=Mus musculus] | 0 | 6.118 | 11 | 2 | 3 | 2 | 1 | 241 | 27 | 5.17 | 10.34 | 2 | 1.171 | 1.039 |
| FALSE | High | Master Pro P47199 | Quinone oxidoreductase [OS=Mus musculus] | 0 | 6.103 | 8 | 2 | 2 | 2 | 1 | 331 | 35.2 | 8.07 | 11.15 | 2 | 1.347 | 1.41 |
| FALSE | High | Master Pro Q02053 | Ubiquitin-like modifier-activating enzyme 1 [OS=Mus musculus] | 0 | 6.1 | 3 | 2 | 2 | 2 | 1 | 1058 | 117.7 | 5.66 | 7.77 | 2 | 0.43 | 0.85 |
| FALSE | High | Master Pro Q88207 | Collagen alpha-1(V) chain [OS=Mus musculus] | 0 | 6.038 | 2 | 2 | 5 | 2 | 1 | 1838 | 183.6 | 4.98 | 17.42 | 2 | 1.383 | 1.543 |
| FALSE | High | Master Pro Q88342 | WD repeat-containing protein 1 [OS=Mus musculus] | 0 | 6.036 | 4 | 3 | 7 | 3 | 1 | 606 | 66.4 | 6.6 | 22.73 | 3 | 1.927 | 1.858 |
| FALSE | High | Master Pro Q8R1F1 | Niban-like protein 1 [OS=Mus musculus] | 0 | 6.028 | 3 | 2 | 3 | 2 | 1 | 749 | 84.8 | 5.94 | 8.06 | 2 | 1.287 | 0.981 |
| FALSE | High | Master Pro P62264 | 40S ribosomal protein S14 [OS=Mus musculus] | 0 | 5.995 | 14 | 2 | 3 | 2 | 1 | 151 | 16.3 | 10.05 | 11.37 | 2 | 1.85 | 1.815 |
| FALSE | High | Master Pro Q9C230-1 | obg-like ATPase 1 [OS=Mus musculus] | 0 | 5.98 | 7 | 2 | 2 | 2 | 1 | 396 | 44.7 | 7.81 | 6.85 | 2 | 0.61 | 0.541 |
| FALSE | High | Master Pro Q03958 | Prefoldin subunit 6 [OS=Mus musculus] | 0 | 5.974 | 17 | 2 | 3 | 2 | 1 | 127 | 14.4 | 8.88 | 9.59 | 2 | 0.972 | 1.372 |
| FALSE | High | Master Pro P26369 | Splicing factor U2AF 65 kDa subunit [OS=Mus musculus] | 0 | 5.961 | 4 | 2 | 4 | 2 | 1 | 475 | 53.5 | 9.09 | 14.78 | 2 | 2.411 | 2.942 |
| FALSE | High | Master Pro P97371 | Proteasome activator complex subunit 1 [OS=Mus musculus] | 0 | 5.938 | 8 | 2 | 3 | 2 | 1 | 249 | 28.7 | 5.97 | 10.2 | 2 | 0.572 | 2.182 |
| FALSE | High | Master Pro Q64437 | Alcohol dehydrogenase class 4 mu/sigma chain [OS=Mus musculus] | 0 | 5.851 | 8 | 2 | 4 | 2 | 1 | 374 | 39.9 | 7.85 | 13 | 2 | 7.339 | 11.192 |
| FALSE | High | Master Pro Q9CWS0 | N(G),N(G)-dimethylarginine dimethylaminohydrolase 1 [OS=Mus musculus] | 0 | 5.848 | 6 | 2 | 3 | 2 | 1 | 285 | 31.4 | 5.97 | 11.03 | 2 | 0.431 | 0.328 |
| FALSE | High | Master Pro Q8CIE6 | coatamer subunit alpha [OS=Mus musculus] | 0 | 5.812 | 2 | 2 | 2 | 2 | 1 | 1224 | 138.3 | 7.65 | 12.04 | 2 | 1.176 | 1.458 |
| FALSE | High | Master Pro Q921M3-1 | Splicing factor 3B subunit 3 [OS=Mus musculus] | 0 | 5.773 | 2 | 2 | 2 | 2 | 1 | 1217 | 135.5 | 5.26 | 5.56 | 2 | 0.528 | 0.439 |
| FALSE | High | Master Pro P97351 | 40S ribosomal protein S3a [OS=Mus musculus] | 0 | 5.703 | 9 | 3 | 8 | 3 | 1 | 264 | 29.9 | 9.73 | 23.16 | 3 | 1.183 | 1.146 |
| FALSE | High | Master Pro P47738 | Aldehyde dehydrogenase, mitochondrial [OS=Mus musculus] | 0 | 5.643 | 4 | 2 | 3 | 2 | 1 | 519 | 56.5 | 7.62 | 9.63 | 2 | 0.951 | 1.875 |
| FALSE | High | Master Pro Q99K18 | dynactin subunit 2 [OS=Mus musculus] | 0 | 5.634 | 8 | 2 | 2 | 2 | 1 | 402 | 44.1 | 5.26 | 7.39 | 2 | 0.671 | 0.528 |
| FALSE | High | Master Pro Q8VIJ6 | splicing factor, proline- and glutamine-rich [OS=Mus musculus] | 0 | 5.552 | 4 | 3 | 3 | 3 | 1 | 699 | 75.4 | 9.44 | 9.35 | 3 | 1.291 | 1.095 |
| FALSE | High | Master Pro Q9JM76 | Actin-related protein 2/3 complex subunit 3 [OS=Mus musculus] | 0 | 5.524 | 13 | 2 | 4 | 2 | 1 | 178 | 20.5 | 8.59 | 9.7 | 2 | 0.806 | 0.716 |
| FALSE | High | Master Pro Q8BV14 | dihydropteridine reductase [OS=Mus musculus] | 0 | 5.51 | 7 | 2 | 3 | 2 | 1 | 241 | 25.6 | 7.81 | 8.85 | 2 | 1.392 | 1.974 |
| FALSE | High | Master Pro Q05186 | Reticulocalbin-1 [OS=Mus musculus] | 0 | 5.494 | 6 | 2 | 2 | 2 | 1 | 325 | 38.1 | 4.84 | 5.93 | 2 | 0.403 | 0.284 |
| FALSE | High | Master Pro P05201 | Aspartate aminotransferase, cytoplasmic [OS=Mus musculus] | 0 | 5.486 | 6 | 3 | 5 | 3 | 1 | 413 | 46.2 | 7.14 | 11.76 | 3 | 0.681 | 0.626 |
| FALSE | High | Master Pro P09671 | Superoxide dismutase [Mn], mitochondrial [OS=Mus musculus] | 0 | 5.448 | 10 | 2 | 3 | 2 | 1 | 222 | 24.6 | 8.62 | 9.29 | 2 | 0.923 | 0.842 |
| FALSE | High | Master Pro P42208 | septin-2 [OS=Mus musculus] | 0 | 5.404 | 6 | 2 | 2 | 2 | 1 | 361 | 41.5 | 6.55 | 5.51 | 2 | 0.609 | 0.585 |
| FALSE | High | Master Pro P31938 | Dual specificity mitogen-activated protein kinase kinase 1 [OS=Mus musculus] | 0 | 5.36 | 6 | 2 | 2 | 2 | 1 | 393 | 43.4 | 6.7 | 5.94 | 2 | 0.74 | 0.409 |
| FALSE | High | Master Pro Q35887 | Calumenin [OS=Mus musculus] | 0 | 5.351 | 8 | 2 | 3 | 2 | 1 | 315 | 37 | 4.67 | 8.15 | 2 | 1.251 | 0.97 |
| FALSE | High | Master Pro P42932 | T-complex protein 1 subunit theta [OS=Mus musculus] | 0 | 5.332 | 3 | 2 | 4 | 2 | 1 | 548 | 59.5 | 5.62 | 9.65 | 2 | 0.495 | 0.373 |
| FALSE | High | Master Pro P11499 | Heat shock protein HSP 90-beta [OS=Mus musculus] | 0 | 5.307 | 4 | 3 | 3 | 2 | 1 | 724 | 83.2 | 5.03 | 8.92 | 3 | 0.993 | 1.174 |
| FALSE | High | Master Pro Q91Y97 | fructose-bisphosphate aldolase B [OS=Mus musculus] | 0 | 5.282 | 5 | 2 | 5 | 1 | 1 | 364 | 39.5 | 8.27 | 14.43 | 2 | 0.236 | 0.168 |
| FALSE | High | Master Pro P47753 | F-actin-capping protein subunit alpha-1 [OS=Mus musculus] | 0 | 5.197 | 9 | 2 | 4 | 1 | 1 | 286 | 32.9 | 5.55 | 14.49 | 2 | 0.843 | 0.866 |
| FALSE | High | Master Pro P21107 | Tropomyosin alpha-3 chain [OS=Mus musculus] | 0 | 5.146 | 6 | 2 | 3 | 2 | 1 | 285 | 33 | 4.72 | 9.99 | 2 | 0.811 | 1.479 |
| FALSE | High | Master Pro P32067 | Lupus La protein homolog [OS=Mus musculus] | 0 | 4.95 | 5 | 2 | 2 | 2 | 1 | 415 | 47.7 | 9.77 | 7.66 | 2 | 2.433 | 2.444 |
| FALSE | High | Master Pro P07091 | Protein S100-A4 [OS=Mus musculus] | 0 | 4.943 | 16 | 2 | 7 | 2 | 1 | 101 | 11.7 | 5.31 | 22.41 | 2 | 2.264 | 3.54 |
| FALSE | High | Master Pro Q08879 | Fibulin-1 [OS=Mus musculus] | 0 | 4.941 | 3 | 2 | 3 | 2 | 1 | 705 | 78 | 5.16 | 11.06 | 2 | 0.277 | 0.233 |
| FALSE | High | Master Pro Q922W0 | Aspartyl aminopeptidase [OS=Mus musculus] | 0 | 4.937 | 4 | 2 | 3 | 2 | 1 | 473 | 52.2 | 7.25 | 10.29 | 2 | 0.8 | 1.151 |
| FALSE | High | Master Pro Q9DAR7 | M7GpppX diphosphatase [OS=Mus musculus] | 0 | 4.873 | 11 | 2 | 3 | 2 | 1 | 338 | 39 | 6.48 | 8 | 2 | 0.638 | 0.518 |
| FALSE | High | Master Pro P54822 | adenylosuccinate lyase [OS=Mus musculus] | 0 | 4.838 | 4 | 2 | 4 | 2 | 1 | 484 | 54.8 | 7.27 | 10.43 | 2 | 1.041 | 1.103 |
| FALSE | High | Master Pro Q11136 | xa-Pro dipeptidase [OS=Mus musculus] | 0 | 4.828 | 3 | 2 | 3 | 2 | 1 | 493 | 55 | 5.78 | 5.95 | 2 | 0.728 | 0.845 |
| FALSE | High | Master Pro Q05142 | 60S ribosomal protein L35a [OS=Mus musculus] | 0 | 4.822 | 21 | 3 | 5 | 3 | 1 | 110 | 12.5 | 10.89 | 12.11 | 3 | 1.146 | 1.041 |
| FALSE | High | Master Pro Q62465 | Synaptic vesicle membrane protein VAT-1 homolog [OS=Mus musculus] | 0 | 4.804 | 4 | 2 | 4 | 2 | 1 | 406 | 43.1 | 6.37 | 13.27 | 2 | 0.991 | 0.495 |
| FALSE | High | Master Pro P14069 | protein S100-A6 [OS=Mus musculus] | 0 | 4.78 | 37 | 2 | 2 | 2 | 1 | 89 | 10 | 5.48 | 6.5 | 2 | 1.619 | 2.681 |
| FALSE | High | Master Pro Q61838 | Pregnancy zone protein [OS=Mus musculus] | 0 | 4.778 | 1 | 2 | 5 | 1 | 1 | 1495 | 165.7 | 6.68 | 14.14 | 2 | 0.361 | 0.243 |
| FALSE | High | Master Pro P50247 | Adenosylhomocysteinase [OS=Mus musculus] | 0 | 4.702 | 4 | 2 | 2 | 2 | 1 | 432 | 47.7 | 6.54 | 5.72 | 2 | 1.134 | 1.064 |
| FALSE | High | Master Pro Q9CQI3 | glia maturation factor beta [OS=Mus musculus] | 0 | 4.696 | 11 | 2 | 4 | 2 | 1 | 142 | 16.7 | 5.16 | 13.19 | 2 | 0.781 | 0.688 |
| FALSE | High | Master Pro P56399 | Ubiquitin carboxyl-terminal hydrolase 5 [OS=Mus musculus] | 0 | 4.628 | 3 | 2 | 4 | 2 | 1 | 858 | 95.8 | 5.01 | 9.79 | 2 | 0.537 | 0.514 |
| FALSE | High | Master Pro Q9CQM5 | Thioredoxin domain-containing protein 17 [OS=Mus musculus] | 0 | 4.483 | 15 | 2 | 5 | 2 | 1 | 123 | 14 | 4.77 | 17.41 | 2 | 3.139 | 3.088 |
| FALSE | High | Master Pro Q91WQ3 | Tyrosine-tRNA ligase, cytoplasmic [OS=Mus musculus] | 0 | 4.448 | 3 | 2 | 2 | 2 | 1 | 528 | 59.1 | 7.01 | 4.88 | 2 | 0.789 | 0.67 |
| FALSE | High | Master Pro Q8BF56-1 | Serine/threonine-protein phosphatase CPPED1 [OS=Mus musculus] | 0 | 4.405 | 8 | 2 | 3 | 2 | 1 | 312 | 35.2 | 5.34 | 9.26 | 2 | 1.288 | 1.195 |
| FALSE | High | Master Pro Q9CXW3 | Calcyclin-binding protein [OS=Mus musculus] | 0 | 4.389 | 7 | 2 | 3 | 2 | 1 | 229 | 26.5 | 7.87 | 10.06 | 2 | 3.41 | 9.381 |
| FALSE | High | Master Pro P08071 | Lactotransferrin [OS=Mus musculus] | 0 | 4.378 | 3 | 2 | 3 | 1 | 1 | 707 | 77.8 | 8.53 | 9.98 | 2 | 0.409 | 0.306 |
| FALSE | High | Master Pro P07901 | Heat shock protein HSP 90-alpha [OS=Mus musculus] | 0 | 4.32 | 2 | 2 | 3 | 1 | 1 | 733 | 84.7 | 5.01 | 8.48 | 2 | 1.371 | 1.656 |
| FALSE | High | Master Pro Q6NZ80 | DnaI homolog subfamily C member 8 [OS=Mus musculus] | 0 | 4.275 | 7 | 2 | 2 | 2 | 1 | 253 | 29.8 | 9.06 | 3 | 2 | 0.668 | 0.409 |
| FALSE | High | Master Pro P01942 | Hemoglobin subunit alpha [OS=Mus musculus] | 0 | 4.184 | 6 | 2 | 5 | 2 | 1 | 142 | 15.1 | 8.22 | 14.58 | 2 | 0.317 | 0.242 |
| FALSE | High | Master Pro P62245 | 40S ribosomal protein S15a [OS=Mus musculus] | 0 | 4.152 | 13 | 2 | 4 | 2 | 1 | 130 | 14.8 | 10.13 | 10.56 | 2 | 2.93 | 2.895 |
| FALSE | High | Master Pro P17426 | AP-2 complex subunit alpha-1 [OS=Mus musculus] | 0 | 4.13 | 3 | 2 | 3 | 2 | 1 | 977 | 107.6 | 7.03 | 8.39 | 2 | 0.794 | 0.708 |
| FALSE | High | Master Pro Q921X4-3 | Isoform 3 of interleukin enhancer-binding factor 3 [OS=Mus musculus] | 0 | 4.129 | 3 | 2 | 2 | 2 | 1 | 911 | 97.7 | 8.95 | 3.68 | 2 | 0.89 | 0.876 |
| FALSE | High | Master Pro Q76MZ3 | serine/threonine-protein phosphatase 2A 65 kDa regulatory subunit A alpha isoform [OS=Mus musculus] | 0 | 4.12 | 4 | 2 | 3 | 2 | 1 | 589 | 65.3 | 5.11 | 9.16 | 2 | 1.134 | 0.909 |
| FALSE | High | Master Pro P43276 | Histone H1.5 [OS=Mus musculus] | 0 | 4.098 | 9 | 2 | 5 | 1 | 1 | 223 | 22.6 | 10.92 | 15.33 | 2 | 1.362 | 1.441 |
| FALSE | High | Master Pro Q8BML9 | glutamine-tRNA ligase [OS=Mus musculus] | 0 | 4.034 | 3 | 2 | 4 | 2 | 1 | 775 | 87.6 | 7.31 | 10.28 | 2 | 0.818 | 0.856 |
| FALSE | High | Master Pro Q3UEB3-1 | poly(U)-binding-splicing factor PUF60 [OS=Mus musculus] | 0 | 4.011 | 3 | 2 | 3 | 2 | 1 | 564 | 60.2 | 5.29 | 8.93 | 2 | 0.728 | 0.841 |
| FALSE | High | Master Pro P50608 | Fibromodulin [OS=Mus musculus] | 0 | 4.001 | 6 | 2 | 4 | 2 | 1 | 376 | 43 | 6.04 | 10.3 | 2 | 0.413 | 0.322 |
| FALSE | High | Master Pro Q9CZ88 | 40S ribosomal protein S19 [OS=Mus musculus] | 0 | 3.987 | 13 | 2 | 4 | 2 | 1 | 145 | 16.1 | 10.4 | 10.04 | 2 | 4.773 | 6.091 |
| FALSE | High | Master Pro P47941 | Crk-like protein [OS=Mus musculus] | 0 | 3.928 | 7 | 2 | 2 | 2 | 1 | 303 | 33.8 | 6.74 | 5.61 | 2 | 0.681 | 0.608 |
| FALSE | High | Master Pro P59999 | Actin-related protein 2/3 complex subunit 4 [OS=Mus musculus] | 0 | 3.926 | 13 | 2 | 3 | 2 | 1 | 168 | 19.7 | 8.43 | 8.66 | 2 | 1.202 | 1.281 |
| FALSE | High | Master Pro P14824 | annexin A6 [OS=Mus musculus] | 0 | 3.83 | 3 | 2 | 2 | 2 | 1 | 673 | 75.8 | 5.5 | 5.81 | 2 | 0.487 | 0.369 |
| FALSE | High | Master Pro Q9WV32 | Actin-related protein 2/3 complex subunit 1B [OS=Mus musculus] | 0 | 3.611 | 5 | 2 | 2 | 2 | 1 | 372 | 41 | 8.35 | 6.22 | 2 | 1.268 | 1.265 |
| FALSE | High | Master Pro Q9JII5 | DA2-associated protein 1 [OS=Mus musculus] | 0 | 3.576 | 8 | 2 | 2 | 2 | 1 | 406 | 43.2 | 8.56 | 2.07 | 2 | 0.413 | 0.347 |
| FALSE | High | Master Pro P70349 | Histidine triad nucleotide-binding protein 1 [OS=Mus musculus] | 0 | 3.554 | 27 | 2 | 2 | 2 | 1 | 126 | 13.8 | 6.87 | 2.26 | 2 | 1.029 | 0.746 |
| FALSE | High | Master Pro Q8CIA5 | thimet oligopeptidase [OS=Mus musculus] | 0 | 3.359 | 2 | 2 | 4 | 2 | 1 | 687 | 78 | 6.06 | 11.25 | 2 | 1.394 | 1.72 |
| FALSE | High | Master Pro Q91WJ8-2 | Isoform 2 of Far upstream element-binding protein 1 [OS=Mus musculus] | 0 | 3.356 | 2 | 2 | 5 | 1 | 1 | 672 | 70.8 | 7.69 | 16.77 | 2 | 2.141 | 2.122 |
| FALSE | High | Master Pro P80318 | T-complex protein 1 subunit gamma [OS=Mus musculus] | 0 | 3.334 | 3 | 2 | 3 | 2 | 1 | 545 | 60.6 | 6.7 | 8.04 | 2 | 0.763 | 0.612 |
| FALSE | High | Master Pro Q9CQC6 | Basic leucine zipper and W2 domain-containing protein 1 [OS=Mus musculus] | 0 | 3.263 | 3 | 2 | 2 | 2 | 1 | 419 | 48 | 5.92 | 6.12 | 2 | 1.863 | 2.484 |
| FALSE | High | Master Pro P62751 | 60S ribosomal protein L23a [OS=Mus musculus] | 0 | 3.243 | 10 | 2 | 5 | 2 | 1 | 156 | 17.7 | 10.45 | 13.08 | 2 | 0.931 | 1.24 |
| FALSE | High | Master Pro P84104-1 | Serine/arginine-rich splicing factor 3 [OS=Mus musculus] | 0 | 3.225 | 14 | 2 | 3 | 1 | 1 | 164 | 19.3 | 11.65 | 5.11 | 2 | 5.696 | 1.903 |
| FALSE | High | Master Pro Q60972 | Histone-binding protein RBBP4 [OS=Mus musculus] | 0 | 3.076 | 4 | 2 | 3 | 2 | 1 | 425 | 47.6 | 4.89 | 8.79 | 2 | 1.727 | 1.477 |
| FALSE | High | Master Pro Q9EQH3 | Vacuolar protein sorting-associated protein 35 [OS=Mus musculus] | 0 | 3.073 | 4 | 2 | 3 | 2 | 1 | 796 | 91.7 | | | | | |

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|-------|------|---------------------|---|---|-------|----|---|---|---|---|------|-------|-------|-------|---|-------|-------|
| FALSE | High | Master Pro P63005-1 | Platelet-activating factor acetylhydrolase IB subunit alpha [OS=Mus musculus] | 0 | 2.924 | 3 | 2 | 4 | 2 | 1 | 410 | 46.6 | 7.37 | 11.2 | 2 | 1.811 | 1.34 |
| FALSE | High | Master Pro P35979 | 60S ribosomal protein L12 [OS=Mus musculus] | 0 | 2.886 | 10 | 2 | 5 | 2 | 1 | 165 | 17.8 | 9.42 | 13.54 | 2 | 1.233 | 1.113 |
| FALSE | High | Master Pro Q8VCT3 | aminopeptidase B [OS=Mus musculus] | 0 | 2.847 | 2 | 2 | 4 | 2 | 1 | 650 | 72.4 | 5.35 | 9.59 | 2 | 0.593 | 1.236 |
| FALSE | High | Master Pro Q8BL97-1 | serine/arginine-rich splicing factor 7 [OS=Mus musculus] | 0 | 2.767 | 9 | 2 | 3 | 1 | 1 | 267 | 30.8 | 11.9 | 4.16 | 2 | 3.212 | 1.283 |
| FALSE | High | Master Pro P63163 | small nuclear ribonucleoprotein-associated protein N [OS=Mus musculus] | 0 | 2.765 | 6 | 2 | 2 | 2 | 1 | 240 | 24.6 | 11.19 | 5.7 | 2 | 4.804 | 3.38 |
| FALSE | High | Master Pro Q91VW3 | SH3 domain-binding glutamic acid-rich-like protein 3 [OS=Mus musculus] | 0 | 2.717 | 18 | 2 | 4 | 2 | 1 | 93 | 10.5 | 5.14 | 9.44 | 2 | 1.593 | 2.146 |
| FALSE | High | Master Pro Q8CIG0 | Protein argonaute-2 [OS=Mus musculus] | 0 | 2.703 | 3 | 2 | 2 | 2 | 1 | 860 | 97.2 | 9.19 | 5.11 | 2 | 0.88 | 0.869 |
| FALSE | High | Master Pro Q3U962 | Collagen alpha-2(V) chain [OS=Mus musculus] | 0 | 2.679 | 1 | 2 | 3 | 1 | 1 | 1497 | 144.9 | 6.7 | 7.27 | 2 | 0.502 | 0.425 |
| FALSE | High | Master Pro Q92111 | Serotransferrin [OS=Mus musculus] | 0 | 2.599 | 3 | 2 | 9 | 1 | 1 | 697 | 76.7 | 7.18 | 23.82 | 2 | 0.417 | 0.236 |
| FALSE | High | Master Pro Q9JHH6 | Carboxypeptidase B2 [OS=Mus musculus] | 0 | 2.595 | 3 | 2 | 4 | 2 | 1 | 422 | 48.8 | 7.97 | 10.36 | 2 | 0.383 | 0.238 |
| FALSE | High | Master Pro P54071 | Isocitrate dehydrogenase [NADP], mitochondrial [OS=Mus musculus] | 0 | 2.581 | 4 | 2 | 3 | 1 | 1 | 452 | 50.9 | 8.69 | 7.79 | 2 | 1.222 | 1.084 |
| FALSE | High | Master Pro Q99LX0 | protein/nucleic acid deglycase DJ-1 [OS=Mus musculus] | 0 | 2.562 | 8 | 2 | 4 | 2 | 1 | 189 | 20 | 6.77 | 12.52 | 2 | 1.721 | 1.686 |
| FALSE | High | Master Pro Q91VH6 | Protein MEMO1 [OS=Mus musculus] | 0 | 2.418 | 10 | 2 | 3 | 2 | 1 | 297 | 33.7 | 7.14 | 5.8 | 2 | 2.85 | 3.844 |
| FALSE | High | Master Pro Q9DBP5 | UMP-CMP kinase [OS=Mus musculus] | 0 | 2.363 | 10 | 2 | 2 | 2 | 1 | 196 | 22.2 | 5.83 | 4.91 | 2 | 0.496 | 0.349 |
| FALSE | High | Master Pro P14211 | Calreticulin [OS=Mus musculus] | 0 | 2.342 | 3 | 2 | 4 | 2 | 1 | 416 | 48 | 4.49 | 10.01 | 2 | 1.018 | 0.762 |
| FALSE | High | Master Pro P24452 | Macrophage-capping protein [OS=Mus musculus] | 0 | 2.238 | 4 | 2 | 3 | 2 | 1 | 352 | 39.2 | 7.2 | 8.42 | 2 | 7.503 | 5.594 |
| FALSE | High | Master Pro P02468 | Laminin subunit gamma-1 [OS=Mus musculus] | 0 | 2.036 | 1 | 2 | 3 | 2 | 1 | 1607 | 177.2 | 5.21 | 7.54 | 2 | 0.414 | 0.505 |

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|------------------------|-------------|
| Sample | TMT reagent |
| Primary OEC+ 5nMA1 sup | 129 |
| OEmyc790_C7+ 5nMA1 sup | 130 |
| OEmyc790_C4+ 5nMA1 sup | 131 |