SUBFUNCTIONALISATION OF PARALOGOUS GENES AND EVOLUTION OF DIFFERENTIAL CODON USAGE PREFERENCES: THE SHOWCASE OF POLYPYRIMIDINE TRACT BINDING PROTEINS

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ABSTRACT

Gene paralogs are copies of an ancestral gene that appear after gene or full genome duplication. When the two sister gene copies are maintained in the genome, redundancy may release certain evolutionary pressures, allowing one of them to access novel gene functions. Here we focused on the evolutionary history of the three polypyrimidine tract binding protein (*PTBP*) paralogs and their concurrent evolution of differential codon usage preferences in vertebrate species.

PTBP1-3 show high identity at the amino acid level (up to 80%), but display strongly different nucleotide composition, divergent CUPrefs and distinct tissue-specific expression levels. Phylogenetic inference suggests that the duplication events leading to the three extant *PTBP1-3* lineages predate the basal diversification within vertebrates. We identify a distinct substitution pattern towards GC3-enriching mutations in *PTBP1*, concurrent with a trend for the use of common codons and for a tissue-wide expression. Genomic context analysis shows that GC3-rich nucleotide composition for *PTBP1*s is driven by local mutational processes. In contrast, *PTBP2*s are enriched in AT-ending, rare codons, and display tissue-restricted expression. Nucleotide composition and CUPrefs of *PTBP2* are only partly driven by local mutational forces, and could have been shaped by selective forces. Interestingly, trends for use of UUG-Leu codon match those of AT-ending codons.

Our interpretation is that a combination of directional mutation–selection has differentially shaped CUPrefs of *PTBPs* in Vertebrates: GC-enrichment of *PTBP1* is linked to the strong and broad tissue-expression, while AT-enrichment of *PTBP2* and *PTBP3* are linked to rare CUPrefs and specialized spatio-temporal expression. This scenario is compatible with a gene subfunctionalisation process by differential expression regulation associated to the evolution of specific CUPrefs.

Keywords Codon usage bias, codon usage preferences, gene duplication, paralog, ortholog, evolution, mutation-selection, nucleotide composition, tissue-specific expression

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1 Significance Statement

In vertebrates, PTBP paralogs display strong differences in gene composition, gene expression regulation, and their expression in cell culture depends on their codon usage preferences. We show that placental mammals PTBP1 have become GC-rich because of local mutational pressures, resulting in an enrichment of frequently used codons and in a strong, tissue-wide expression. On the contrary, PTBP2 in vertebrates are AT-rich, with a lower contribution of local mutational processes to their specific nucleotide composition, show high frequency of rare codons and in placental mammals display a restricted expression pattern contrasting to that of PTBP1. The systematic study of composition and expression patterns of gene paralogs can help understand the complex mutation-selection interplay that shape codon usage bias in multicellular organisms.

2 Introduction

During gene translation, ribosomes assemble proteins by specific amino acid linear polymerisation guided by the successive reading of mRNA nucleotide triplets, called codons. Each time a codon is read, it is chemically compared to the set of available tRNAs' anticodons. Upon codon-anticodon match, the ribosome loads the tRNA and adds the associated amino acid to the nascent protein. The main 20 amino acids are decoded by 61 codon-anticodon combinations, so that multiple codons are associated with the same amino acid. These are named synonymous codons (Nirenberg and Matthaei, 1961; Khorana et al., 1966). Codon Usage Preferences (CUPrefs) refer to the differential usage of synonymous codons, between species, or between genes and genomic regions in the same genome (Grantham et al., 1980; Carbone et al., 2003). Mutation and selection are the two main forces shaping CUPrefs (Duret, 2002; Chamary et al., 2006; Plotkin and Kudla, 2011). Mutational biases relate to directional mechanistic biases during genome replication (Reijns et al., 2015; Apostolou-Karampelis et al., 2016), during genome repair (Lujan et al., 2012), or during recombination (Pouyet et al., 2017), preferentially introducing one nucleotide over others or inducing recombination and maintaining genomic regions depending on their composition. Mutational biases are well known in prokaryotes and eukaryotes, ranging from simple molecular preferences towards 3'A-ending in the *Taq* polymerase (Clark, 1988) to the complex GC-biased gene conversion in vertebrates (Pouyet et al., 2017). Selective forces shaping CUPrefs are often described as translational selection. This notion refers to the ensemble of mechanistic steps and interactions during translation that are affected by the particular CUPrefs of the mRNA, so that the choice of certain codons at certain positions may actually enhance the translation process and can be subject to selection (Bulmer, 1991). Translational selection covers thus codon-mediated effects acting on mRNA maturation, secondary structure and overall stability (Presnyak et al., 2015; Novoa and Ribas de Pouplana, 2012), subcellular localisation, programmed frameshifts, translation speed and accuracy, or protein folding (Caliskan et al., 2015; Mordstein et al., 2020; Spencer and Barral, 2012).

Translational selection has been demonstrated in prokaryotes and some eukaryotes (Satapathy et al., 2016; Percudani et al., 1997; Duret and Mouchiroud, 1999; Whittle and Extavour, 2016), often in the context of tRNA availability (Ikemura, 1981). However, its very existence in Vertebrates remains highly debated (Pouyet et al., 2017; Galtier et al., 2018).

Homologous genes share a common origin either by speciation (orthology) or by duplication events (paralogy) (Sonnhammer and Koonin, 2002). Upon gene (or full genome) duplication, the new genome will contain two copies of the original gene, referred to as in-paralogs. After speciation, each daughter cell will inherit one couple of paralogs, *i.e.* one copy of each ortholog (Koonin, 2005). The emergence of paralogs upon duplication releases the evolutionary constraints on the individual genes. Evolution can thus potentially lead to function specialisation, such as evolving a particular substrate preferences, or engaging each paralog on specific enzyme activity preferences in the case of promiscuous enzymes (Copley, 2020). Gene duplication can also allow one paralog to explore broader sequence space and to evolve radically novel functions, while the remaining counterpart can assure the original function.

The starting point for our research are the experimental observations by Robinson and coworkers reporting differential expression of the polypyrimidine tract binding protein (*PTBP*) human paralogs as a function of their nucleotide com-

position (Robinson et al., 2008). Vertebrates genomes encode for three in-paralogous versions of the *PTBP* genes, all of them fulfilling similar functions in the cell: they form a class of hnRNP RNA-Binding Proteins that are involved in the modulation of mRNAs alternative splicing (Pina et al., 2018). Within the same genome the three paralogs display high amino-acid sequence similarity, around 70% in humans and with similar overall values in vertebrates (Pina et al., 2018).

Despite the high resemblance at the protein level, the three *PTBP* paralogs sharply differ in nucleotide composition, CUPrefs and tissue expression pattern. In humans, *PTBP1* is enriched in GC3-rich synonymous codons and is widely expressed in all tissues, while *PTBP2* and *PTBP3* are AT3-rich and display an enhanced expression in the brain and in hematopoietic cells respectively (Supplementary Material S1). Robinson and coworkers studied the expression in human cells in culture of all three human *PTBP* paralogous genes placed under the control of the same promoter. They showed that the GC-rich paralog *PTBP1* was more highly expressed than the AT-rich ones, and that the expression of the AT-rich paralog *PTBP2* could be enhanced by synonymous codons recoding towards the use of GC-rich codons (Robinson et al., 2008). Here we have built on the evolutionary foundations of this observation and extended the analyses of CUPrefs to *PTBP* paralogs in vertebrate genomes. Our results suggest that paralog-specific directional changes in CUPrefs in mammalian *PTBP* concurred with a process of subfunctionalisation by differential tissue pattern expression of the three paralogous genes.

3 Material and Methods

Sequence retrieval

We assembled a dataset of DNA sequences from 47 mammalians and 27 non-mammalians Vertebrates and 3 protostomes using the BLAST function on the nucleotide database of NCBI (NCBI Resource Coordinators, 2018) taking the human *PTBP* paralogs as references (see supplementary Material S2 for accession numbers). We could identify the corresponding three orthologs in all Vertebrate species screened, except for the European rabbit *Oryctolagus cuniculus*, lacking *PTBP1* and from the rifleman bird *Acanthisitta chloris*, lacking *PTBP3* (Supplementary Material S2). The final vertebrate dataset contained 75 *PTBP1*, 76 *PTBP2* and 75 *PTBP3* sequences. As outgroups for the analysis, we retrieved the orthologous genes from three protostome genomes, which contained a single *PTBP* homolog per genome (Supplementary Material S2). From the original dataset, we identified a subset of nine mammalian and six non-mammalian vertebrates species with a good annotation of the *PTBP* chromosome context, and we retrieved synteny and composition information on the flanking regions and introns (Supplementary Material S3). Because of annotation hazards, intronic and flanking regions information were missing for some *PTBP*s in the African elephant *Loxodonta africana*, Schlegel's Japanese Gecko *Gekko japonicus* and the whale shark *Rhincodon typus* assemblies. For the selected 15 species the values for codon adaptation index (CAI) (Sharp and Li, 1987) and codon usage similarity index (COUSIN) (Bourret et al., 2019) were calculated using the COUSIN server (available at https://cousin.ird.fr).

Clustering PTBPs by their CUPrefs

For each *PTBP* paralog we calculated codon composition and CUPrefs analyses via the COUSIN tool (Bourret et al., 2019). For each *PTBP* gene we constructed a vector of 59 positions with the relative frequencies of all synonymous codons. To reduce information dimension for the analysis of CUPrefs, we applied on the 229 59-dimension vectors: i) a k-means clustering; ii) a hierarchical clustering; and iii) a principal component analysis (PCA).

Alignment and phylogenetic analyses

To generate robust alignments without introducing artefacts due to large evolutionary distances between in-paralogs we proceeded stepwise, as follows: i) we aligned separately at the amino acid level each set of *PTBP* paralog sequences of mammals and non-mammalians Vertebrates; ii) for each *PTBP* paralog we merged the alignments for mammals and for non mammals, obtaining the three *PTBP1*, *PTBP2* and *PTBP3* alignments for all Vertebrates; iii) we combined the three alignments for each paralog into a single one; iv) we aligned the outgroup sequences to the global Vertebrate *PTBP*s alignment. All alignments steps were performed using MAFFT (Katoh et al., 2002). The final amino acid alignment was back-translated to obtain the codon-based nucleotide alignment. The codon-based alignment was trimmed using Gblocks (Castresana, 2000) (Supplementary Material)

Phylogenetic inference was performed at the amino acid and at the nucleotide level using RAxML v8.2.9 and bootstrapping over 1000 cycles (Stamatakis, 2014). For nucleotides we used codon-based partitions and applied the GTR+G4 model while for amino acids we applied the LG+G4 model. For the 79 species used in the analyses we retrieved a species-tree from the TimeTree tool (Kumar et al., 2017). Distances between phylogenetic trees were computed using the Robinson-Foulds index, which accounts for differences in topology (Robinson and Foulds, 1981), and the K-tree score, which accounts for differences in topology and in branch length (Soria-Carrasco et al., 2007). After phylogenetic inference we computed marginal ancestral states for the respectively most recent common ancestors at the nucleotide level of each paralog using RAxML. Using these ancestral sequences we estimated the number of synonymous and non-synonymous mutations of each extant sequence to the corresponding most recent common ancestor.

Statistical analyses

Correlation between matrices was assessed via the Mantel test. Non-parametric comparisons were performed using the Wilcoxon-Mann-Whitney test for population medians and the Wilcoxon signed rank test for paired comparisons. Statistical analyses were performed using the *ape* and *ade4* R packages and JMP v14.3.0.

4 Results

Vertebrate PTBP paralogs differ in nucleotide composition

In order to understand the evolutionary history of *PTBP* genes we performed first a nucleotide composition and CUPrefs analysis on the three paralogs in 79 species. Overall, *PTBP1* are GC-richer than *PTBP2* and *PTBP3* (respective mean percentages 55.9, 42.3 and 44.9 for GC content and 69.5, 33.4 and 38.3 for GC3 content; Figure 1, Supplementary Material S2). In addition, *PTBP1*s show a difference in GC3 between mammalian and non-mammalian gene (respectively 79.8 against 59.9 mean percentages). A linear regression model followed by a Tukey's honest significant differences analysis for GC3 using as explanatory levels paralog (*i.e. PTBP1-3*), taxonomy (*i.e.* mammalian or non-mammalian) and their interaction identifies three mains groups of *PTBP*s (Table 1): a first one corresponding to mammalian *PTBP1*, a second one grouping non-mammalian *PTBP1* and a third one spanning all *PTBP2* and *PTBP3*. The largest explanatory factor for GC3 was the paralog *PTBP1-3*, accounting alone for 65% of the variance, while the interaction between the levels taxonomy and paralog captured around 15% of the remaining variance (Table 1). These trends are confirmed when performing paired comparisons between paralogs present in the same mammalian genome, with significant differences in GC3 content in the following order: *PTBP1* > *PTBP3* > *PTBP2* (Wilcoxon

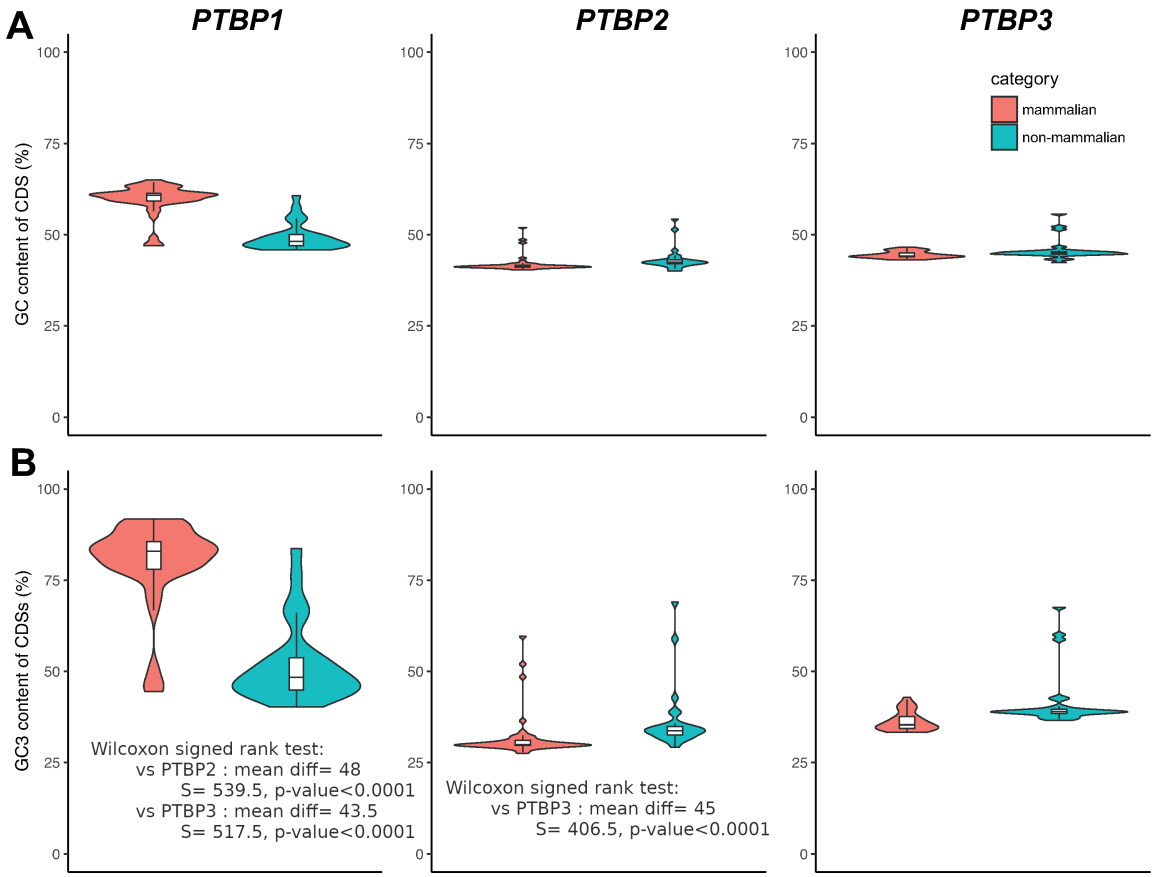

Figure 1: **GC content (A) and GC3 content (B) of Vertebrates *PTBP*s.** Violin plots display the overall distribution while box and whiskers display median, quartiles and 95% of the corresponding values for mammalian (red) and non-mammalian (blue) individual genomes. Results of Wilcoxon signed rank test between overall GC3 content of paralogs is indicated

signed rank test: *PTBP1* vs *PTBP2*, mean diff=48.0, S=539.50, p-value <0.0001; *PTBP1* vs *PTBP3*, mean diff=43.5, S=517.50, p-value <0.0001; *PTBP3* vs *PTBP2*, mean diff=4.5, S=406.50, p-value <0.0001). Note that even if all of them significantly different, the mean paired differences in GC3 between *PTBP1* and *PTBP2-3* are ten times larger than the corresponding mean paired differences between *PTBP2* and *PTBP3*.

The distribution of the residuals between observed and expected values after our model fit to the data allows to identify a number of outliers species with interesting taxonomical patterns in compositional deviation (Table 2). For non mammals, the three *PTBP* paralogs in the rainbow trout *Oncorhynchus mykiss* genome display high GC3 content (between 67% and 76%), all of them significantly higher than model-predicted values (expected values between 36% and 51%). A similar case occurs for the zebrafish *Danio rerio* genome: the three paralogs display GC3 values around 58%, which for *PTBP2* and *PTBP3* paralogs are significantly higher than predicted by the model (expected values around 38%). Very interestingly, for the monotrema platypus *Ornithorhynchus anatinus* as well as for the three marsupials in the dataset the Tasmanian devil *Sarcophilus harrisii*, the koala *Phascolarctos cinereus* and the grey

short-tailed opossum *Monodelphis domestica* their *PTBP1* genes present similar GC3 content around 47%, which is significantly lower than predicted by the model (expected values around 79%).

In many vertebrate species, strong compositional heterogeneities are observed along chromosomes and are often referred to as "isochores". To explore the influence of the genomic environment on the nucleotide composition of *PTBP*s, for 15 species with well-annotated genomes we analyzed the correlation of paralog GC3 with two local compositional variables of the corresponding gene (GC content of intronic and flanking regions) and with three global compositional variables for the corresponding genomes (global GC3 in the complete genomic ORFome, global GC content in all introns, and global GC content in all flanking regions) (Table 3 and Figure 2). First, for *D. rerio* the GC3 composition of *PTBP2* and *PTBP3* is clearly different from the rest, in line with the outlier results presented in Table 2. We have thus excluded the zebra fish values and performed an individual as well as a stepwise linear fit to explain the variance in GC3 composition by the variance in the local and global compositional variables mentioned above (Table 3). For all three *PTBP*s the local GC content explains best the corresponding GC3 content, but with strong differences between paralogs: while variation in the local composition captures almost perfectly variation in the GC3 content of *PTBP1* (R^2=0.97) and relatively well in the case of *PTBP2* (R^2=0.46), the fraction of variance explained by the local composition significantly drops for *PTBP3* (R^2=0.15).

Vertebrate *PTBP* paralogs differ in CUPrefs

Table 1: Global linear regression model and post-hoc Tukey's honest significant differences (HSD) test for GC3 composition as explained variable and the explanatory levels paralog (*PTBP1-3*), taxonomy (*i.e.* mammalian or non-mammalian) and their interactions. Overall goodness of the fit: Adj Rsquare=0.83; F ratio=205.7; Prob > F: <0.0001.Individual effects for the levels: i) paralog: F ratio=274.3; Prob > F: <0.0001; ii) taxonomy: F ratio=27.2; Prob > F: <0.0001; iii) interaction paralog*taxonomy: F ratio=87.9; Prob > F: <0.0001.

Level	**Least Sq. Mean (GC3%)**	**Standard error**	**Tukey's HSD group**
	Paralog		
PTBP1	65.87	1.00	A
PTBP3	39.00	1.01	B
PTBP2	34.03	1.00	C
	Taxonomy		
mammalian	49.32	0.70	A
non-mammalian	43.28	0.92	B
	Paralog*Taxonomy		
PTBP1, mammalian	79.81	1.22	A
PTBP1, non-mammalian	51.93	1.59	B
PTBP3, non-mammalian	41.64	1.62	C
PTBP3, mammalian	36.36	1.22	C, D
PTBP2, non-mammalian	36.27	1.59	C, D
PTBP2, mammalian	31.79	1.20	D

For each *PTBP* coding sequence we extracted the relative frequencies of synonymous codons and performed different approaches to reduce information dimension and visualise CUPrefs trends. The results of a principal component analysis (PCA) are shown in Figure 3. The first PCA axis captured 68.9% of the variance, far before the second and the third axes (respectively 6.7% and 3.2%). Codons segregate in the first axis by their GC3 composition, the only exception being the UUG-Leu codon, which grouped together with AT-ending codons. This first axis differentiates mammalian *PTBP1*s on the one hand and *PTBP2*s and *PTBP3*s on the other hand. Non-mammalian *PTBP1*s scatter between mammalian *PTBP1*s and *PTBP3*s, along with the protostomates *PTBP*s. In the second PCA axis the only obvious (but nevertheless cryptic) codon-structure trends are: i) the split between C-ending and G-ending codons, but not between A-ending and U-ending codons; and ii) the large contribution in opposite directions to this second axis of the AGA and AGG-Arginine codons. This second PCA axis differentiates *PTBP2*s from *PTBP3*s paralogs, consistent with these composition trends. A paired-comparison confirms that *PTBP3*s are richer in C-ending codons than *PTBP2*s, respectively 21.7% against 15.4% (Wilcoxon signed rank test: mean diff=6.2, S=1184.0, p-value <0.0001).

As an additional way to identify groups of genes with similar CUPrefs we applied a hierarchical clustering and a k-means clustering. Both analyses mainly aggregate *PTBP* genes by their GC3 richness. The *PTBP* dendrogram resulting of the hierarchical clustering (rows in clustering in Figure 3; Kappa-Fleiss consistency score = 0.76) shows five main clades that cluster the paralogs with a good match to the following groups: mammalian *PTBP1*s, non-mammalian *PTBP1*s, *PTBP2*s, *PTBP3*s and a fifth group containing the protostomata *PTBP*s and a few individuals of all three paralogs. Regarding codon clustering, the hierarchical stratification sharply splits GC-ending codons from AT-ending codons, with the only exception again of the UUG-Leu codon, which consistently groups within the AT-

Table 2: Individual genes with outlier values with respect to the linear regression expected values for the levels paralog (*PTBP1-3*), taxonomy (mammalian or non-mammalian) and their interactions.

Species	**paralog**	**observed GC3 (%)**	**expected GC3 (%)**	**deviation GC3 (%)**
		mammalian		
Desmodus rotundus	*PTBP2*	59.60	31.79	27.81
Miniopterus natalensis	*PTBP2*	48.52	31.79	16.72
Monodelphis domestica	*PTBP1*	44.49	79.81	-35.32
Ornithorhynchus anatinus	*PTBP1*	51.14	79.81	-28.67
Ornithorhynchus anatinus	*PTBP2*	52.00	31.79	20.21
Phascolarctos cinereus	*PTBP1*	47.53	79.81	-32.28
Sarcophilus harrisii	*PTBP1*	45.44	79.81	-34.37
		non-mammalian		
Danio rerio	*PTBP2*	58.89	36.27	22.62
Danio rerio	*PTBP3*	60.08	41.64	18.44
Lepisosteus oculatus	*PTBP3*	58.73	41.64	17.10
Oncorhynchus mykiss	*PTBP1*	76.27	51.93	24.34
Oncorhynchus mykiss	*PTBP2*	69.03	36.27	32.76
Oncorhynchus mykiss	*PTBP3*	67.58	41.64	25.95
Pogona vitticeps	*PTBP1*	83.68	51.93	31.75

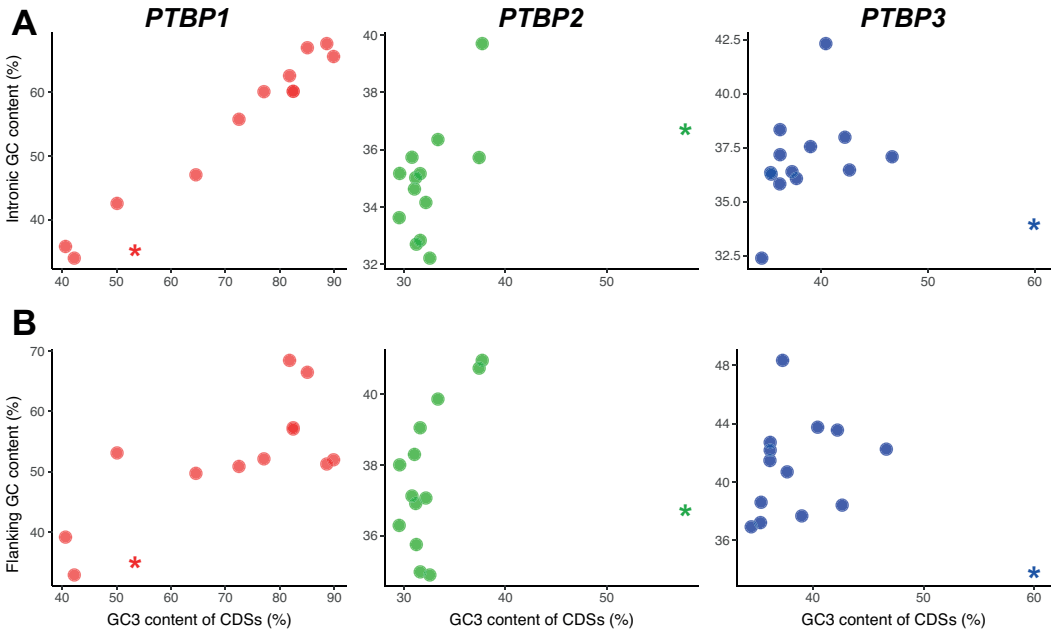

Figure 2: **Variation in GC3 content of *PTBP*s (x-axis) and in the GC content of the corresponding introns (A, y axis) or flanking regions (B, y axis)**. Each dot represents one of the 15 individual used for the genomic context analysis. The asterisk indicates the values for the species *Danio rerio*, which shows peculiar results for *PTBP2* and *PTBP3*, consistent with its outlier behaviour in the global model.

ending codons. The elbow approach of k-means clustering identifies an optimal number of four clusters and separates the paralog genes with a good match as following: *PTBP1*, *PTBP2*, *PTBP3* and a group containing the protostoma and individuals from all paralogs (Kappa-Fleiss consistency score = 0.75).

Overall, k-means clustering and hierarchical clustering, both based on the 59-dimensions vectors of the CUPrefs, are congruent with one another (Kappa-Fleiss consistency score = 0.83), and largely concordant with the PCA results. CUPrefs define thus groups of *PTBP* genes consistent with their orthology and taxonomy. It is interesting to note that for some species the *PTBP* paralogs display unique distributions of CUPrefs, such as an overall similar CUPrefs in the three *PTBP* genes of the whale shark *Rhincodon typus*, or again some shifts in nucleotide composition between paralogs in the Natal long-fingered bat *Miniopterus natalensis*.

In order to characterise the directional CUPrefs bias of the different paralogs, we have analysed for the 15 species with well-annotated genomes described above, the match between each individual *PTBP* and the average CUPrefs of the corresponding genome (Table 4). Our results highlight strong differences for mammalian paralogs: *PTBP1*s display COUSIN values above 1 while *PTBP2*s display COUSIN values below zero. Given the interpretation of COUSIN values (Bourret et al., 2019) these results mean that in mammals *PTBP1*s are enriched in commonly used codons in a higher proportion than the average in the genome, while *PTBP2*s are enriched in rare codons to the extent that their CUPrefs go in the opposite direction to the average in the genome. As for *PTBP3*, in mammals we observe COUSIN values below 0 in most cases or very close to 0 in the case of the horse *Equus caballus* and house mouse *Mus*

musculus, implying a tendency towards rare codons. In non-mammals however, PTBPs show an overall similarity to their respective reference CUPrefs.

Phylogenetic reconstruction of *PTBP*s

We explored the evolutionary relationships between *PTBP*s by phylogenetic inference at the amino acid and at the nucleotide level (Figure 4, Supplementary Material Figure S10). Our final dataset contained 74 *PTBP* sequences from mammals (47 species within 39 families) and non mammal vertebrates (27 species within 24 families). We used the

Table 3: Results for an individual (left) or for a sequential (right) least squares regression for explaining variation in GC3 composition of *PTBP*s genes, by variation of different local (introns or flanking regions of the corresponding gene) or of global (all coding CDS, all introns and all flanking regions in the corresponding genome) compositional variables in 14 well-annotated vertebrate genomes. For the sequential fit, variables are ordered according to their contribution to the sequentially better model, and the order may thus differ between paralogs. Variables labelled with "n.s." (not significant) do not contribute with significant additional explanatory power when added to the sequential model. BIC, Bayesian information content.

PTBP1					
Individual contributions			**Sequential contribution**		
Parameter	**R^2**	**P value F test**	**Parameter**	**R^2**	**BIC**
Local_GC_intron	0.9726	<0.001	Local_GC_intron	0.9726	66.4765
Local_GC_flanking	0.5345	0.0069	Local_GC_flanking	0.974 (n.s.)	68.3142
Global_GC3_exome	0.7279	0.0004	Global_GC3_exome	0.9749 (n.s.)	70.3842
Global_GC_introns	0.116	0.2786	Global_GC_flanking	0.9803(n.s.)	69.9886
Global_GC_flanking	0.1041	0.3065	Global_GC_introns	0.9806(n.s.)	72.2531
PTBP2					
Individual contributions			**Sequential contribution**		
Parameter	**R^2**	**P value F test**	**Parameter**	**R^2**	**BIC**
Local_GC_intron	0.3738	0.0264	Local_GC_intron	0.4558	60.1257
Local_GC_flanking	0.4558	0.0113	Global_GC_introns	0.4895(n.s.)	61.8583
Global_GC3_exome	0.0943	0.3075	Global_GC3_exome	0.4914(n.s.)	64.3761
Global_GC_introns	0.0488	0.4684	Global_GC_flanking	0.4934(n.s.)	66.8894
Global_GC_flanking	0.0287	0.5801	Local_GC_flanking	0.4974(n.s.)	69.35
PTBP3					
Individual contributions			**Sequential contribution**		
Parameter	**R^2**	**P value F test**	**Parameter**	**R^2**	**BIC**
Local_GC_intron	0.1554	0.1825	Local_GC_intron	0.1554	74.7338
Local_GC_flanking	0.0522	0.4528	Local_GC_flanking	0.2095(n.s.)	76.4388
Global_GC3_exome	0.0504	0.461	Global_GC_introns	0.2718(n.s.)	77.9368
Global_GC_introns	0.0002	0.9661	Global_GC3_exome	0.2938(n.s.)	80.1032
Global_GC_flanking	0.0024	0.8744	Global_GC_flanking	0.2938(n.s.)	82.667

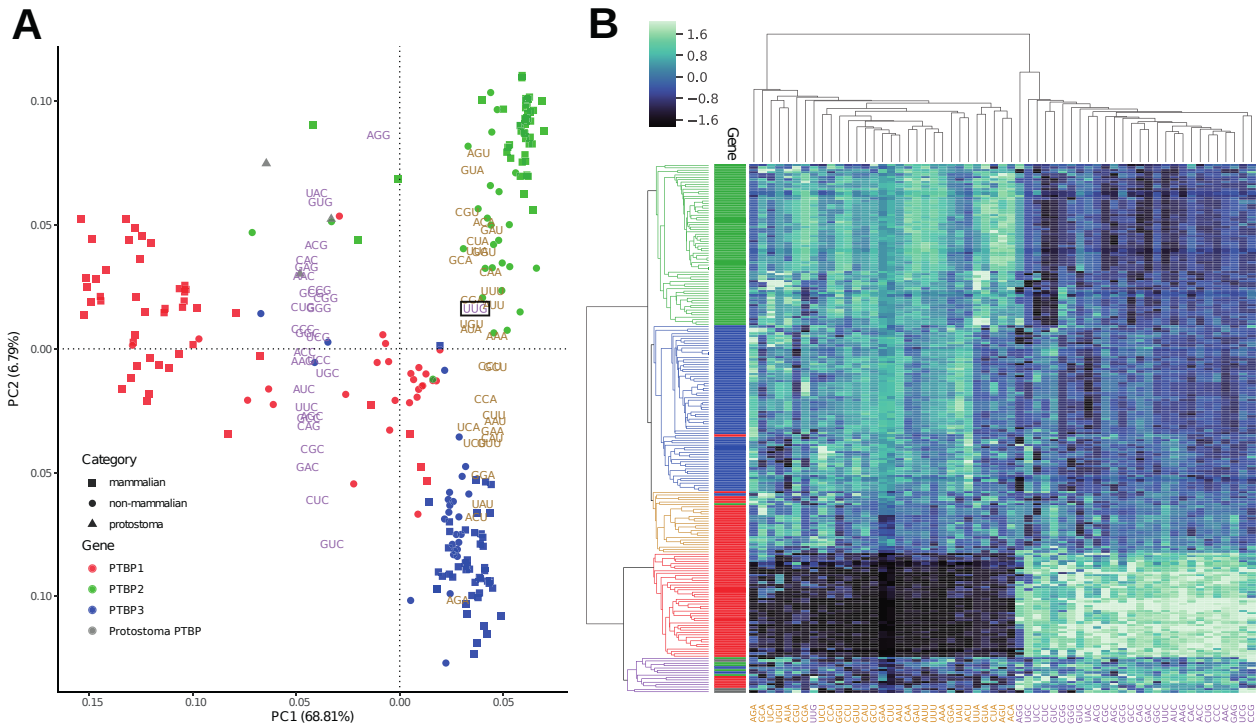

Figure 3: **CUPrefs analysis of *PTBP*s**. A) Plot of the two first dimensions of a PCA analysis based on the codon usage preferences of *PTBP1*s (red), *PTBP2*s (green), *PTBP3*s (blue) and protostoma (grey) individuals. Taxonomic information is included as mammals (squares), non-mammals (circles) and protostomes (triangles). The PCA was created using as variables the vectors of 59 positions (representing the relative frequencies of the 59 synonymous codons) for each individual gene. The eigenvalues of the individual codon variables are given by their position on the graph. Each codon variable is identified by its name and by a colour code, purple for GC-ending codons and orange for AT-ending codons. The percentage of the total variance explained by each axis is shown in parenthesis. B) Heatmap of *PTBP*s individuals (rows) and synonymous codons (columns). Left dendrogram represents the hierarchical clustering of *PTBP*s based on their CUPrefs with colour codes that stand for the clusters created from this analysis. Side bars give information on heatmap individuals regarding i) their origin : *PTBP1* (red), *PTBP2* (green), *PTBP3* (blue) or protostoma (grey). Note the position of the UUG-Leu codon, in both the PCA and the codon dendrogram, as the sole GC-ending codon clustering with all other AT-ending codons)

PTBP genes from three protostome species as outgroups. Both amino acid and nucleotide phylogenies rendered three main clades grouping the *PTBP*s by orthology. In both topologies, *PTBP1* and *PTBP3* orthologs cluster together, although the protostome outgroups are linked to the tree by a very long branch, hampering the proper identification of the Vertebrate *PTBP* tree root. Amino acid and nucleotide subtrees were largely congruent (see topology and branch length comparisons in Table5). The apparently large nodal and split distance values between nucleotide and amino acid *PTBP2* trees stem from disagreements in very short branches, as evidenced by the lowest K-tree score for this ortholog (as a reminder, the Robinson-Foulds index exclusively regards topology while the K-tree score combines topological and branch-length dependent distance between trees, see Material and Methods). In all three cases, internal structure of the ortholog trees essentially recapitulates species taxonomy at the higher levels (Table5). Some of the species

identified by the mathematical model as displaying a largely divergent nucleotide composition present accordingly long branches in the phylogenetic reconstruction, such as *PTBP3* for *O. mykiss*.

Table 4: Global linear regression model and post-hoc Tukey's honest significant differences (HSD) test, the explained variable being the COUSIN value of the each *PTBP* gene against the average of the corresponding genome and the explanatory levels paralog (*PTBP1-3*), taxonomy (*i.e.* mammalian or non-mammalian) and their interactions. Overall goodness of the fit: Adj Rsquare=0.82; F ratio=36.84; Prob > F: <0.0001.Individual effects for the levels: i) paralog: F ratio=40.72; Prob > F: <0.0001; ii) taxonomy: F ratio=10.87; Prob > F: =0.0021; iii) interaction paralog*taxonomy: F ratio=28.11; Prob > F: <0.0001.

Level	**Least Sq. Mean (COUSIN)**	**Standard error**	**Tukey's HSD group**
	Paralog		
PTBP1	1.45	0.11	A
PTBP3	0.29	0.11	B
PTBP2	0.19	0.11	B
	Taxonomy		
mammalian	0.44	0.080	A
non-mammalian	0.85	0.098	B
	Paralog*Taxonomy		
PTBP1, mammalian	1.90	0.14	A
PTBP1, non-mammalian	0.99	0.17	B
PTBP2, non-mammalian	0.81	0.17	B
PTBP3, non-mammalian	0.75	0.17	B
PTBP3, mammalian	-0.16	0.14	C
PTBP2, mammalian	-0.43	0.14	C

Table 5: Comparison between species tree and subtrees of the nucleotide based maximum likelihood tree. Each subtree corresponds to a paralog. The K-tree score compares topological and pairwise distances between trees after re-scaling overall tree length, with higher values corresponding to more divergent trees. The Robinson-Foulds score compares only topological distances between trees, the values shown correspond to the number of partitions that are not shared between two trees.

Reference tree	**Comparison tree**	**K-tree score**	**Robinson-Foulds score**
	Nucleotide tree VS species tree		
PTBP1	Species tree	0.759	42
PTBP2	Species tree	0.762	24
PTBP3	Species tree	1.700	28
	Nucleotide tree VS Amino acid tree		
PTBP1-AA	*PTBP1*-NT	0.149	78
PTBP2-AA	*PTBP2*-NT	0.129	110
PTBP3-AA	*PTBP3*-NT	0.380	40

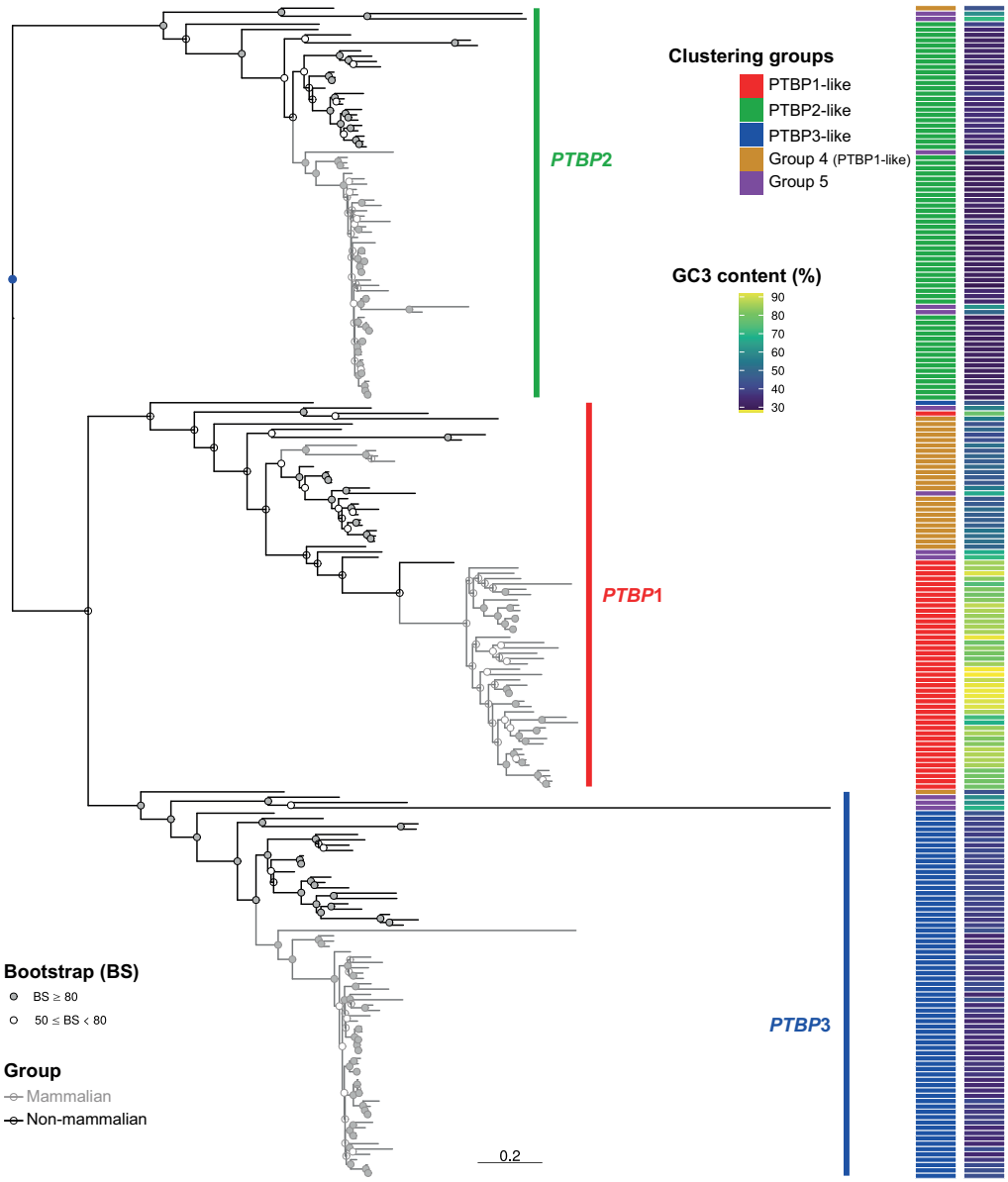

Figure 4: **Maximum-likelihood nucleic acid phylogeny of *PTBP*s genes.** The phylogram depicts *PTBP2*s (green side bar), *PTBP1*s (red side bar) and *PTBP3*s (blue side bar) clades. The outgroup genes from protostomata are not shown to focus on the scale for vertebrate *PTBP*s, but their placement on the tree and the polarity they provide for vertebrate *PTBP*s is given by the blue dot. Gray branches indicate mammalian *PTBP*s, while black branches indicate non-mammalian species. Note the lack of monophyly for mammals for *PTBP1*s. Filled dots on nodes indicate bootstrap values above 80, and empty dots indicate lower support values. Side bar on the left identifies the classification of each gene into the five groups identified by the hierarchical clusters, with the colour code in the inset. Side bar on the right displays GC3 content of the corresponding genes, with the gradient for the colour code ranging from 0 (blue) to 100% (yellow).

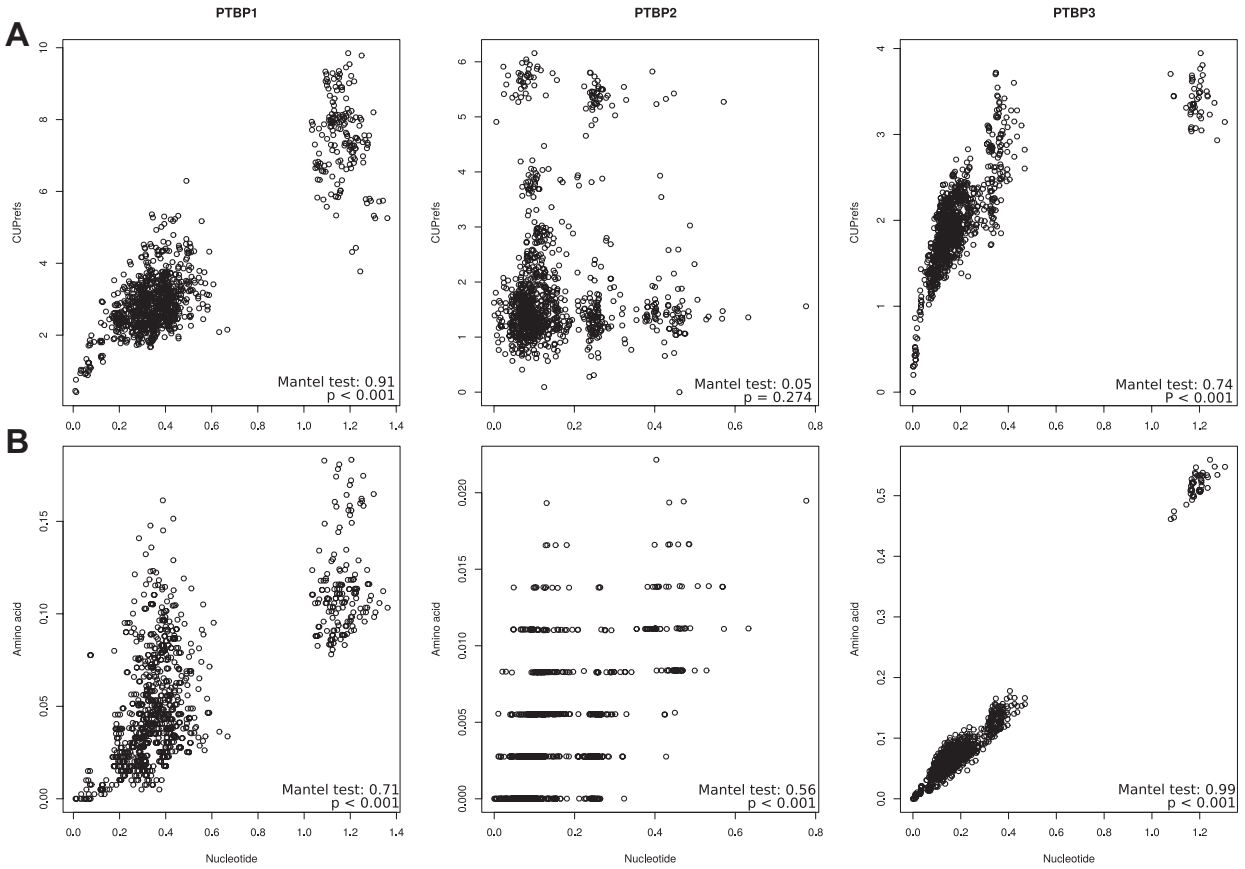

Figure 5: **Nucleotide-based pairwise distances against A) CUPrefs-based and B) amino acid-based pairwise distances for the different mammalian *PTBP* orthologs.** The results for a Mantel test assessing the correlation between the corresponding matrices are shown in each inset.

We have then analysed the correspondence between nucleotide-based and amino acid-based pairwise distances to measure the extent of codon usage bias impact on the obtained phylogeny. We observe a good correlation between both reconstructions for all paralogs, except for mammalian *PTBP2*s, which display extremely low divergence at the amino acid level (Figure 5 B, Supplementary Material S8 B). For mammalian *PTBP1*s, the plot allows to clearly differentiate a cloud with the values corresponding to the monotremes+marsupial mammals, split apart from placental mammals in terms of both amino acid and nucleotide distances. This distribution matches well the fact that monotremes+marsupials cluster separately from placental mammals in *PTBP1* phylogeny (see grey branches being paraphyletic for *PTBP1* in Figure 4). The same holds true for the platypus *PTBP3*, extremely divergent from the rest of the mammalian orthologs. For mammalian paralogs, the plots allow to see the increased number of overall mutations in general and of non-synonymous mutations in particular in *PTBP3*s compared with *PTBP1*. The precise mutational patterns are analysed in detail below. The histograms describing the accumulation of synonymous and non-synonymous mutations confirm that mammalian *PTBP1*s have selectively accumulated the largest number of synonymous mutations compared to non-mammalian *PTBP1*s and to other orthologs.

We have finally analysed the connection between nucleotide-based evolutionary distances within *PTBP* paralogs and CUPrefs-based distances (Figure 5A, Supplementary Material S8 A). A trend showing increased differences in CUPrefs as evolutionary distances increase is evident only for *PTBP1*s and *PTBP3*s in mammals. For mammalian *PTBP1*s the plot clearly differentiates a cloud with the values corresponding to the monotremes+marsupials splitting apart from placental mammals in terms of both evolutionary distance and CUPrefs. For mammalian *PTBP2*s the plot captures the divergent CUPrefs of the platypus and of the bats *M. natalensis* and *Desmodus rotundus*, while for non-mammalian *PTBP2*s the divergent CUPrefs of the rainbow trout are obvious. Finally, for mammalian *PTBP3*s the large nucleotide divergence of the platypus paralog is evident. Importantly, all these instances of divergent behaviour (except for the platypus *PTBP3*) are consistent with the deviations described above from the expected composition by the mathematical modelling of the ortholog nucleotide composition.

Mammalian PTBP1s accumulate GC-enriching synonymous substitutions

We have shown that *PTBP1* genes are GC-richer and specifically GC3-richer than the *PTBP2* and *PTBP3* paralogs in the same genome, and that this enrichment is of a larger magnitude in placental *PTBP1*s. We have thus assessed whether a directional mutational pattern underlies this enrichment, especially regarding synonymous mutations. For this we have inferred the ancestral sequences of the respective most recent common ancestors of each *PTBP* paralog, recapitulated synonymous and non-synonymous mutations between extant sequences and these ancestors, and constructed the corresponding mutation matrices (table S11). The two first axes of a principal component analysis using these mutational matrices capture, with a similar share, 66.95% of the variance between individuals (Figure 6). The first axis of the PCA separates synonymous from non-synonymous substitutions. Intriguingly though, while T<->C transitions are associated to synonymous mutations, as expected, G<->A transitions are associated to non-synonymous mutations. The second axis separates substitutions by their effect on nucleotide composition: GC-stabilizing/enriching on one direction, AT-stabilizing/enriching on the other one. Strikingly, the mutational spectrum of mammalian *PTBP1*s sharply differs from the rest of the paralogs. Substitutions in mammalian *PTBP1* towards GC-enriching changes, in both synonymous and non-synonymous compartments, are the main drivers of the second PCA axis. In contrast, synonymous mutations in *PTBP3* as well as all mutations in *PTBP2* tend to be AT-enriching. Finally, the mutational trends for *PTBP1* in mammals are radically different from those in non-mammals, while for *PTBP2* and *PTBP3*s the substitution patterns are similar in mammals and non-mammals for each of the compartments synonymous and non-synonymous.

5 Discussion

The non equal use of synonymous codons has puzzled biologists since first described. It gave rise to fruitful (and unfruitful) controversies between defenders of *all-is-neutralism* and defenders of *all-is-selectionism*, and launched further the quest for additional molecular signaling beyond codons themselves (Callens et al., 2021). The main questions around CUPrefs are twofold. On the one hand, their origin: to what extent they are the result of fine interplay between mutation and selection processes. On the other hand, their functional implications: whether and how particular CUPrefs can be linked to specific gene expression regulation processes, by modifying the kinetics and dynamics of DNA transcription, mRNA maturation and stability, mRNA translation, and/or protein folding and stability. In the present work we have built on the experimental results of Robinson and coworkers, which display the differential ex-

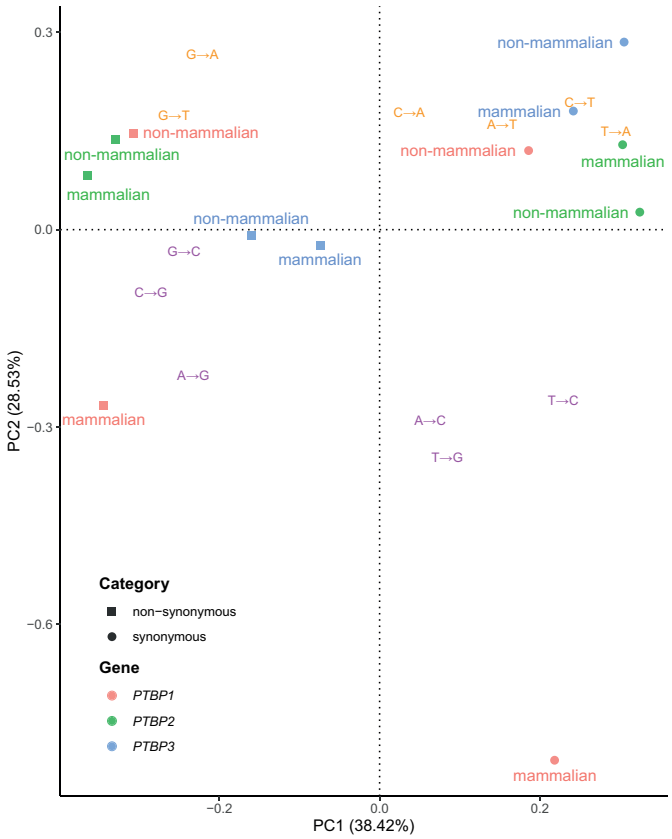

Figure 6: **Mutational spectra of synonymous and non-synonymous substitutions for *PTBP*s.** This principal component analysis (PCA) has been built using the observed nucleotide synonymous and non-synonymous substitution matrices for each *PTBP* paralog, inferred after phylogenetic inference and comparison of extant and ancestral sequences. The variables in this PCA are the types of substitution (*e.g.* A->G), identified by a colour code as GC-enriching / stabilizing substitutions (purple) or AT-enriching / stabilizing substitutions (orange). Variables are plotted according to their eigenvalues. Individuals in this PCA are the mutation categories in *PTBP* genes, stratified by their nature (synonymous or non-synonymous), by orthology (colour code for the different *PTBP*s is given in the inset) and by their taxonomy (mammals, or non-mammals).

pression of the *PTBP* human gene paralogs as a function of their CUPrefs (Robinson et al., 2008). From this particular example, we have aimed at exploring the nature of the connection between paralogous gene evolution and CUPrefs. Our results show that the three *PTBP* paralogous genes of Vertebrates, with divergent expression patterns, also have divergent nucleotide composition and CUPrefs. We carry on Robinson and coworkers suppositions and propose here that this evolutionary pattern could be compatible with a phenomenon of phenotypic evolution by sub-functionalisation (in this case specialisation in tissue-specific expression levels), linked to genotypic evolution by association to specific CUPrefs patterns. Such conclusions invite to pursue Robinson and coworkers efforts by comparing *PTBP*s CUPrefs-modulated expression among numerous Vertebrates cell lines, especially between mammalians and non-mammalians ones.

We have reconstructed the phylogenetic relationships and analysed the evolution and diversity of CUPrefs among *PTBP* paralogs within 74 vertebrate species. The phylogenetic reconstruction shows that the genome of ancestral vertebrates already contained the three extant *PTBP* paralogs. This is consistent with the ortholog and paralog identification in the databases ENSEMBL and ORTHOMAM (Yates et al., 2020; Scornavacca et al., 2019; Pina et al., 2018). Although our results suggest that *PTBP1* and *PTBP3* are sister lineages, the distant relationship of the vertebrate genes with the protostome outgroup precludes the inference of a clear polarity between vertebrate *PTBP*s. We identify no instance of replacement between paralogs, and the evolutionary histories of the different *PTBP*s comply well with those of the corresponding species. The most blatant mismatch between gene and species trees is the polyphyly of mammalian *PTBP1*. In fact, monotremes and marsupials constitute a monophyletic clade, without placental mammals and not basal to them. Multiple findings in our results show sharp, contrasting patterns between *PTBP1* and the *PTBP2-3* paralogs: i) the excess of accumulation of synonymous mutations in mammalian *PTBP1*s for a similar total number of mutations (Figure 5 B); ii) the larger differences in CUPrefs between genes with a similar total number of nucleotide changes in the case of *PTBP1*s in mammals (Figure 5 A); iii) the explicitly different mutational spectrum of synonymous mutations in *PTBP1*s, enriched in A->C, T->G and T->C substitutions (Figure 6); iv) the sharp difference of CUPrefs between *PTBP1*s, and *PTBP2-3*s; and v) the clustering of *PTBP1* genes in monotremes and marsupials together with *PTBP1* genes in non-mammals according to their CUPrefs (Figure3 A). Overall, the particular nucleotide composition and the associated CUPrefs in mammalian *PTBP1* genes are most likely associated to specific mutational biases as shown by the strong correlation between coding and non-coding GC content in *PTBP1* orthologs (Figure 2; Table 3).

While GC3-rich nucleotide composition and CUPrefs of mammalian *PTBP1*s are dominated by local mutational biases, this is not the case for mammalian PTBP2, overall AT3-richer without any clear correlation between coding and non-coding GC content among studied species (Figure 2; 3). In vertebrates, nucleotide composition varies strongly along chromosomes, so that long stretches, historically named "isochores", appear enriched in GC or in AT nucleotides and present particular physico-chemical profiles (Caspersson et al., 1968). Local mutational biases and GC bias gene conversion mechanism, underlying such heterogeneity, predominantly shape local nucleotide composition in numerous Vertebrates genomes, so that the physical location of a gene along the chromosome largely explains its CUPrefs (Holmquist, 1989). In agreement with this mutational bias hypothesis, variation in GC3 composition of *PTBP1*s is almost totally (R2=0.97) explained by the variation in local GC composition (Figure 2; Table 3), suggesting that a same mutational bias has shaped the GC-rich composition of the flanking, intronic and coding regions of *PTBP1*s. The same trend, but to a lesser degree holds also true for PTBP2s (R2=0.45). GC-biased gene conversion is often invoked as a powerful mechanism underlying such local GC-enrichment processes, leading to the systematic replacement of the alleles with the lowest GC composition by a GC richer homolog (Marais, 2003). It has been proposed that gene expression during meiosis prevents GC-biased gene conversion during meiotic recombination (Pouyet et al., 2017). Expression of *PTBP1* in human cells is documented during meiosis in the ovocite germinal line and expression of the AT-rich *PTBP2* has been observed during spermatogenic meiosis (Zagore et al., 2015; Hannigan et al., 2017). With the assumption that *PTBP*s patterns of expression are shared between mammalian species, the GC-richness of *PTBP1* is not due to any GC-biased conversion and the low GC content of *PTBP2* can be explained by an accumulation of GC->AT and AT->AT mutations. The low GC-content observed in *PTBP3*, coupled with a lack of correlation with

neither coding nor non-coding GC-content could indicate that other mechanisms may shape the observed CUPrefs for this paralog.

In mammals, global GC-enriching genomic biases strongly impact CUPrefs, so that the most used codons in average tend to be GC-richer (Hershberg and Petrov, 2009). For this reason, mammalian GC3-rich *PTBP1*s match better the average genomic CUPrefs than AT3-richer *PTBP2* and *PTBP3*, which display CUPrefs in the opposite direction to the average of the genome. In the case of humans, *PTBP1* presents a COUSIN value of 1.747, consistent with an enrichment in preferentially-used codons, while on the contrary, the COUSIN value of -0.477 for *PTBP2* and of -0.235 for *PTBP3* points towards a strong enrichment in rare codons (Supplementary Material S4). The poor match between human *PTBP2* CUPrefs and the human average CUPrefs could result in low expression of these genes in different human and murine cell lines, otherwise capable of expressing *PTBP1* at high levels and *PTBP3* at a lesser degree (Robinson et al., 2008). The barrier to *PTBP2* expression seems to be the translation process, as *PTBP2* codon-recoding towards GC3-richer codons results in strong protein production in the same cellular context, without significant changes in the corresponding mRNA levels (Robinson et al., 2008). Such codon recoding strategy towards preferred codons has become a standard practice for gene expression engineering, despite our lack of understanding the whole impact of local and global gene composition, nucleotide CUPrefs or mRNA structure on gene expression (Brule and Grayhack, 2017).

The poor expressibility of *PTBP2* in human cells, the increase in protein production by the introduction of common codons, along with mutational biases failing to explain entirely *PTBP2* nucleotide composition and CUPrefs, raise the question of the adaptive value of poor CUPrefs in this paralog. Specific tissue-dependent or cell-cycle dependent gene expression regulation patterns have been invoked to explain the codon usage-limited gene expression for certain human genes, such as *TLR7* or *KRAS* (Newman et al., 2016; Lampson et al., 2013; Fu et al., 2018). In humans, the expression levels of the three *PTBP* paralogs are tissue-dependent (Supplementary Material S1), and these differences are conserved through mammals (Keppetipola et al., 2012). In the case of the duplicated genes, subfunctionalisation through specialisation in spatio-temporal gene expression has often been proposed as the main evolutionary force driving conservation of paralogous genes (Ferris and Whitt, 1979). Such differential gene expression regulation in paralogs has actually been documented for a number of genes at very different taxonomic levels (Donizetti et al., 2009; Guschanski et al., 2017; Freilich et al., 2006). Specialised expression patterns in time and space can result in antagonistic presence/absence of the paralogous proteins (Adams et al., 2003). This is precisely the case of *PTBP1* and *PTBP2* during central nervous system development: in non-neuronal cells, *PTBP1* represses *PTBP2* expression by the skip of the exon 10 during *PTBP2* mRNA maturation, while during neuronal development, the micro RNA miR124 down-regulates *PTBP1* expression, which in turn leads to up-regulation of *PTBP2* (Keppetipola et al., 2012; Makeyev et al., 2007). Further, despite the high level of amino acid similarity between both proteins, *PTBP1* and *PTBP2* seem to perform complementary activities in the cell and to display different substrate specificity, so that they are not directly inter-exchangeable by exogenous manipulation of gene expression patterns (Vuong et al., 2016).

In addition to local genomic context analyses, we explored *PTBP* chromosomal location and local synteny (Supplementary Material S13). The results show that, while it is clear that the position of human *PTBP1* is telomeric, and thus in one of the GC-richer region of human chromosome 9, most *PTBP*s are randomly positioned among species. Thus, while the specific location of human *PTBP1* may have influenced its CUPrefs, it is unclear whether the chro-

mosomic location of *PTBPs* have an impact on observed nucleotide composition. Synteny of *PTBP*s genes seems to be conserved, with some exceptions: most mammalian *PTBP1*s have a conserved synteny block that differs from non-mammalian species ; with the exception of *D. rerio*. *PTBP2* and *PTBP3* synteny seems conserved between mammalian and non-mammalian species with only *D. rerio* lackig the *SUSD1* gene between *PTBP3* and *UGCG*. Such results could indicate that vertebrate radiation has been followed up by a changing of *PTBP1* genomic context, with a swapping in flanking genes in mammalian branches. Such results could be related to the observed GC-content drift in *PTBP1* between mammalian and non-mammalian species.

In a different subject, we want to drive the attention of the reader towards the puzzling trend of the UUG-Leu codon in our CUPrefs analyses. This UUG codon is the only GC-ending codon systematically clustering with AT-ending codons in all our analyses, and does not show the expected symmetrical behaviour with respect to UUA (see Figure 3). Such behaviour for UUG has been depicted, but not discussed, in other analyses of CUPrefs in mammalian genes (see figure 7 in Laurin-Lemay et al. (2018)), in coronavirus genomes (Daron and Bravo, 2021), as well as for AGG-Arg and GGG-Gly in a global study of codon usages across the tree of life (see figure 1 in (Novoa et al., 2019)). The reasons underlying the clustering of UUG with AT-ending codons are unclear. A first line of thought could be functional: the UUG-Leu codon is particular because it can serve as alternative starting point for translation (Peabody, 1989). However, other codons such as ACG or GUG act more efficiently than UUG as translation initiation, and do not display any noticeable deviation (Ivanov et al., 2011). A second line of thought could be related to the tRNA repertoire, but both UUG and UUA are decoded by similar numbers of dedicated tRNAs in the vast majority of genomes (*e.g.* respectively six and seven tRNA genes in humans (Palidwor et al., 2010)). Finally, another line of thought suggests that UUG and AGG could be disfavoured if mutational pressure towards GC is very high, despite being GC-ending codons (Palidwor et al., 2010). Indeed, the series of synonymous transitions UUA->UUG->CUG for Leucine and the substitution chain AGA->AGG->CGG for Arginine are expected to lead to a depletion of UUG and of AGG codons when increasing GC content. Both UUG and ACG codons would this way display a non-linear, non-monotonic response to GC-mutational biases (Palidwor et al., 2010). In our data-set, however, AGG maps with the rest of GC-ending codons, symmetrically opposed to AGA as expected, and strongly contributing to the second PCA axis. Thus, only UUG presents frequency use patterns similar to those of AT-ending codons. We humbly admit that we do not find a satisfactory explanation for this behaviour and invite researchers in the field to generate alternative explanatory hypotheses.

We have presented here an evolutionary analysis of the *PTBP* paralogs family, as a showcase of evolution upon gene duplication. Our results show that CUPrefs in *PTBP*ss have evolved in parallel with specific gene expression regulation patterns. In the case of *PTBP1*, the most tissue-wise expressed of the paralogs, we have potentially identified compositional, mutational biases as the driving force leading to strong enrichment in GC-ending codons. In contrast, for *PTBP2* the enrichment in AT-ending codons is rather compatible with selective forces related to specific spatio-temporal gene expression pattern, antagonistic to those of *PTBP1*. Our results suggest that the systematic study of composition, genomic location and expression patterns of paralogous genes can contribute to understanding the complex mutation-selection interplay shaping CUPrefs in multicellular organisms.

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7 Data Availability Statement

All data required to reproduce our findings is provided in the tables in the main text or in the Supplementary Material section.

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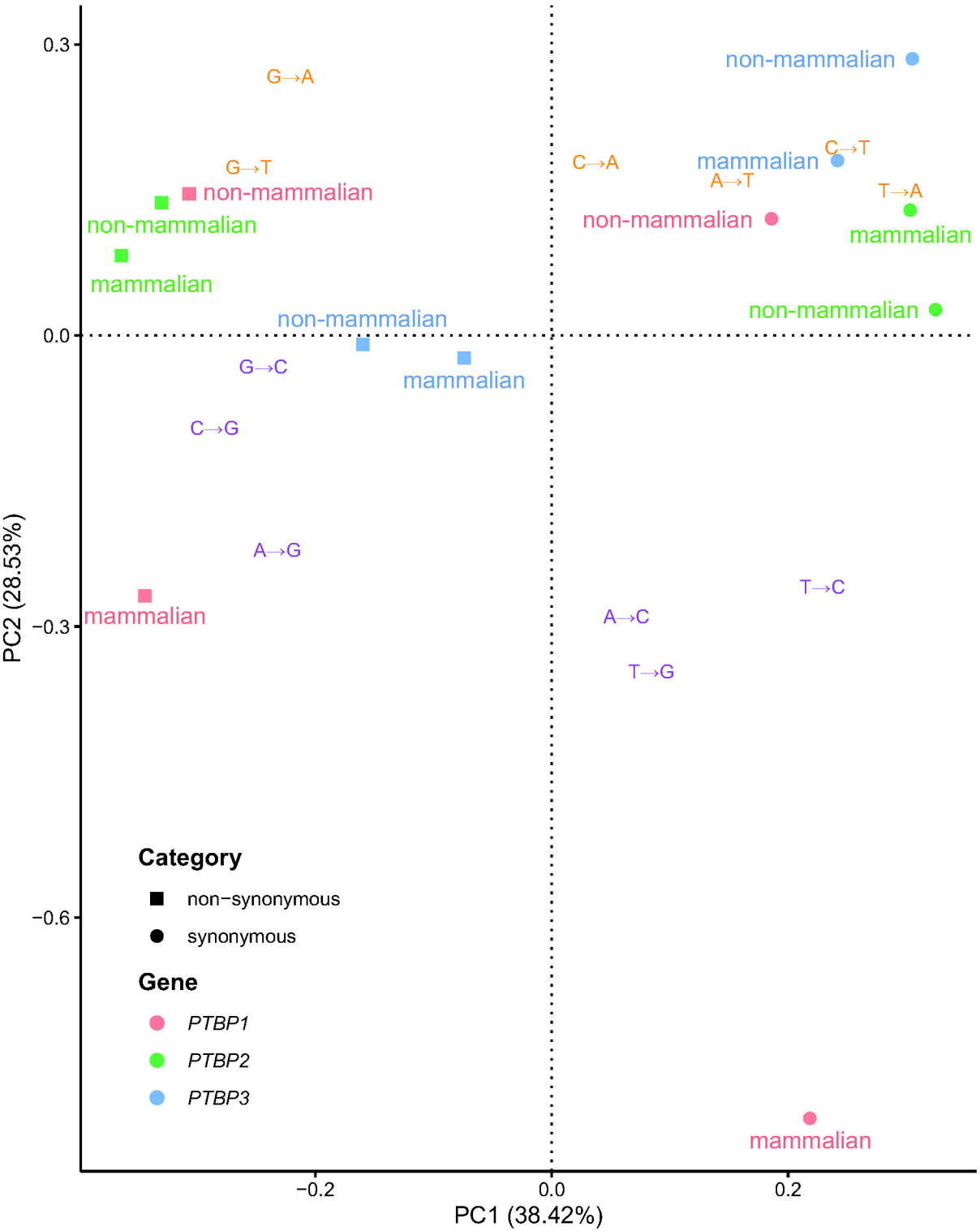
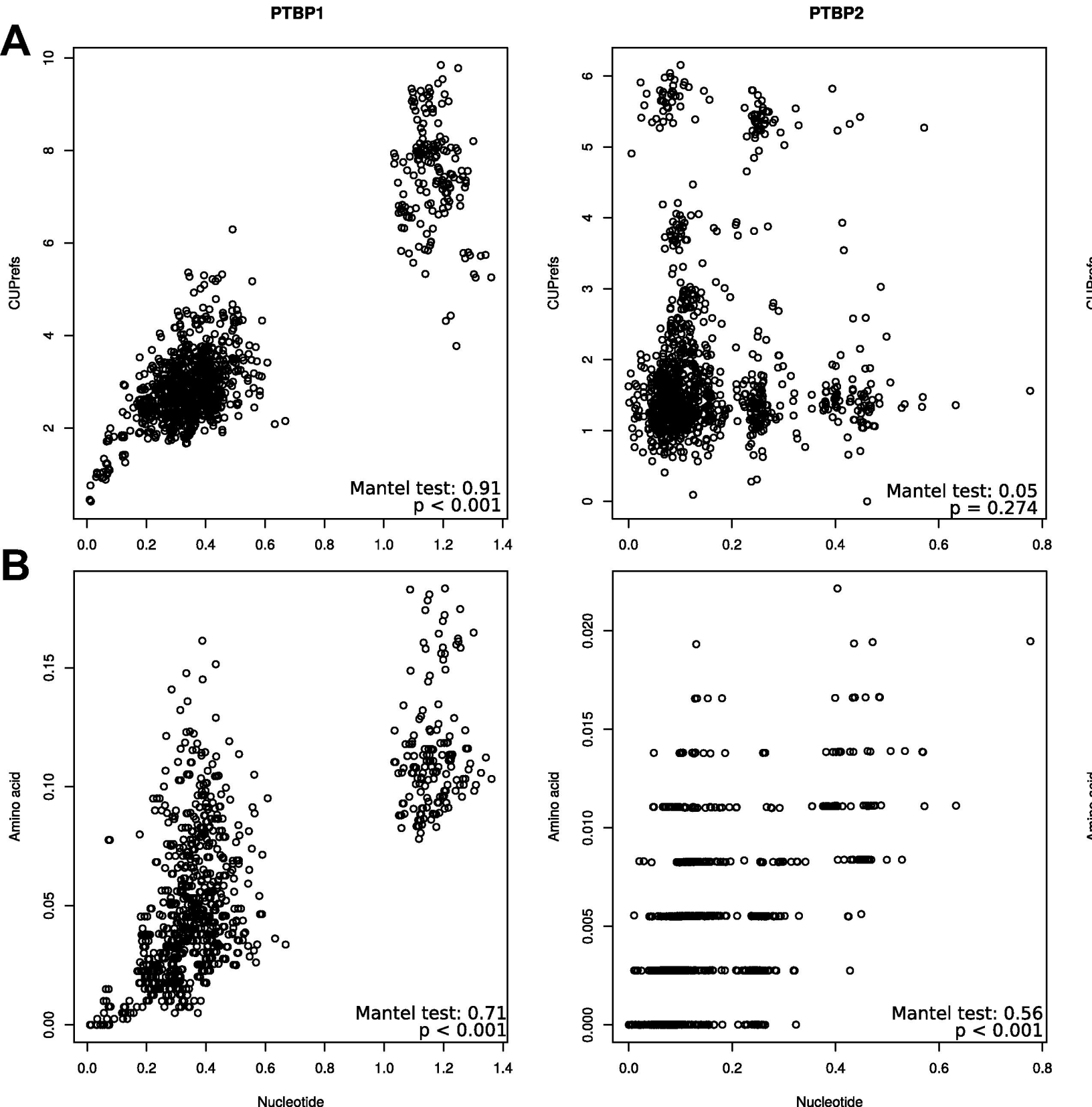
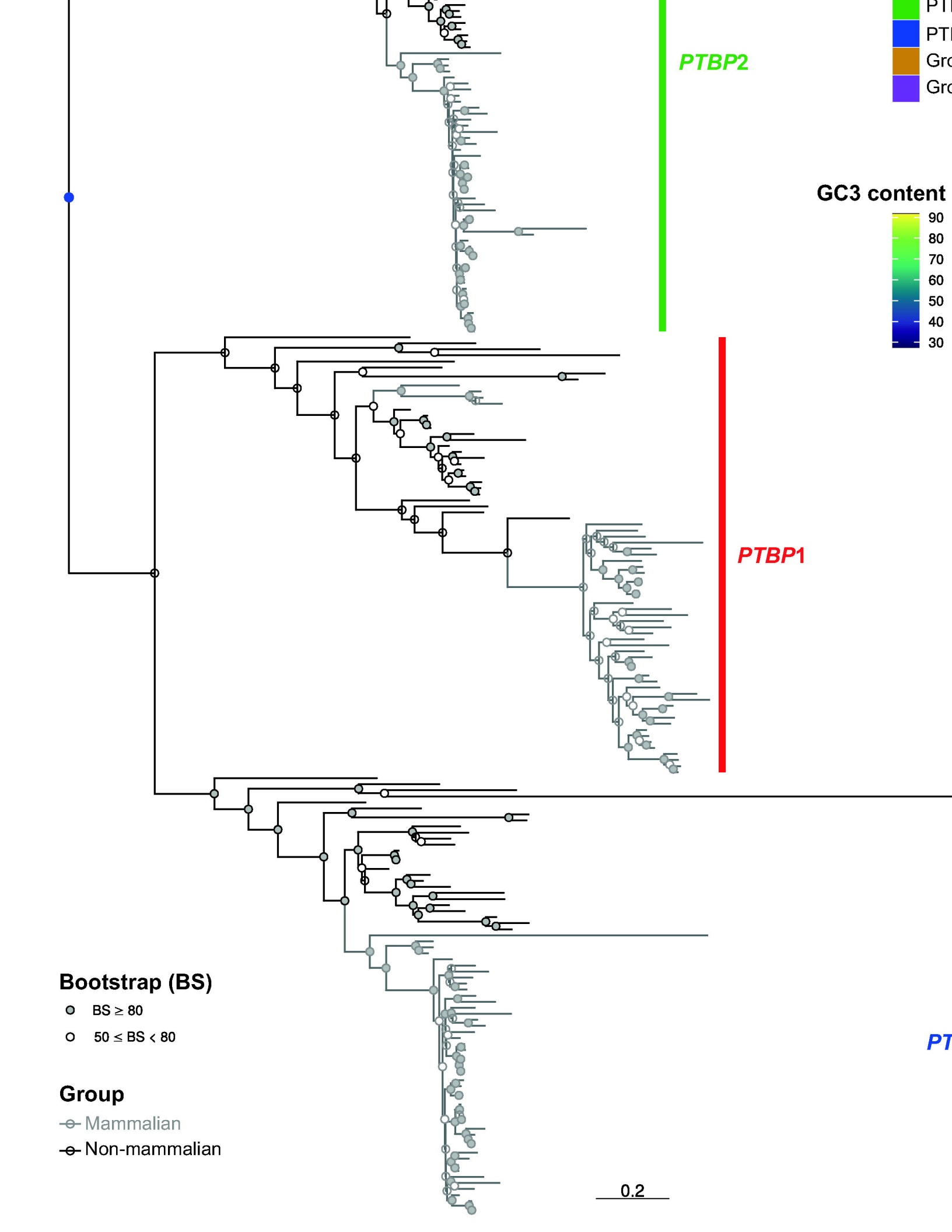
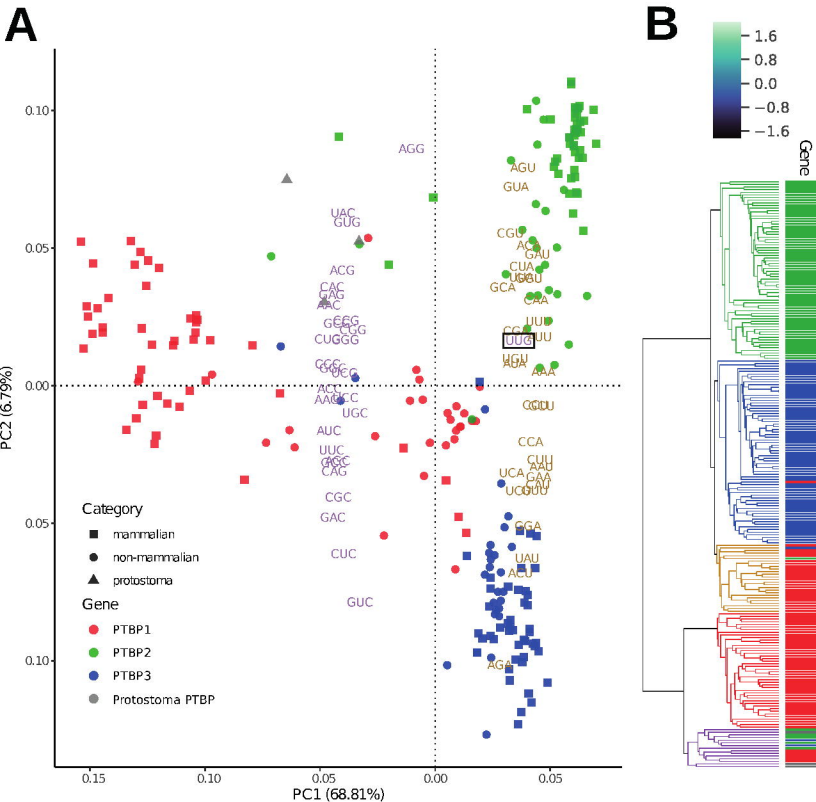
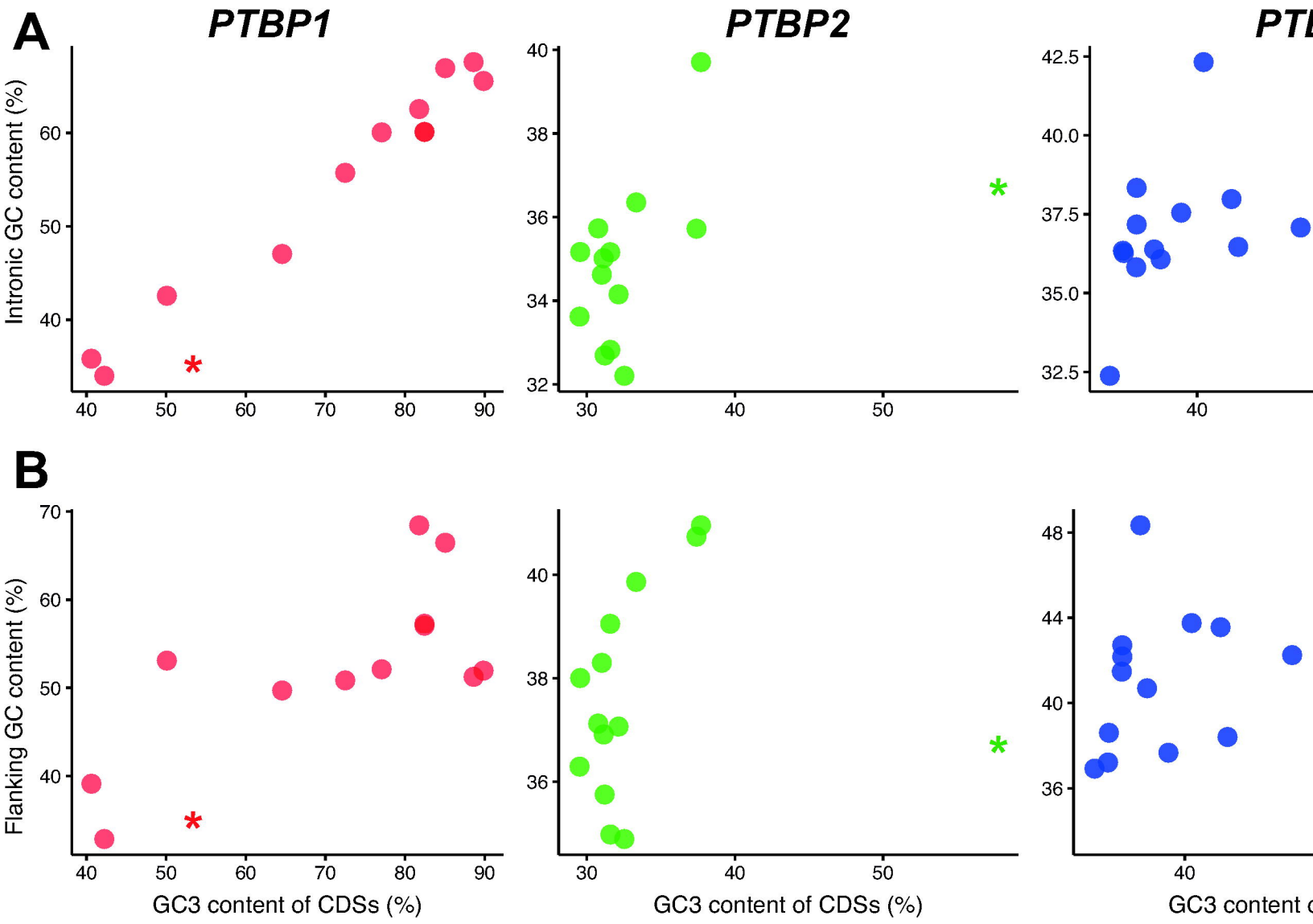
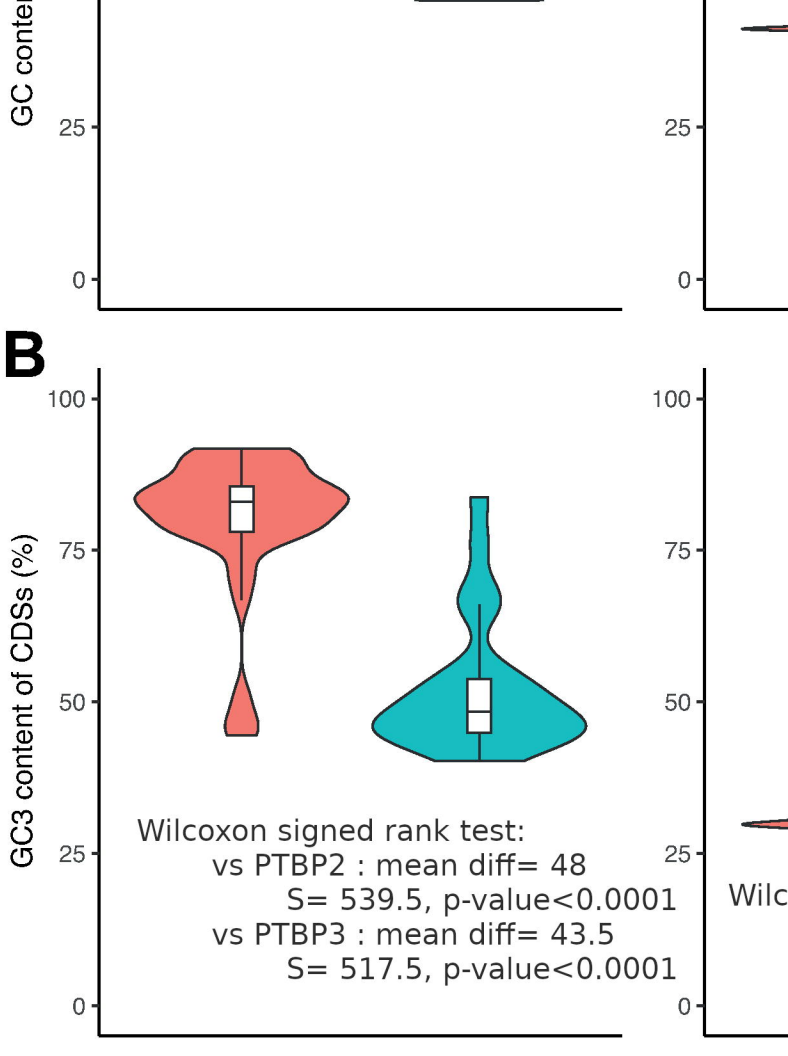

0.3
0.0
−0.3
−0.6
−0.2
0.0
0.2
PC1 (38.42%)
PC2 (28.53%)
G→A
G→T
non-mammalian
non-mammalian
mammalian
non-mammalian
mammalian
G→C
C→G
A→G
mammalian
non-mammalian
C→A
mammalian
C→T
A→T
T→A
non-mammalian
mammalian
non-mammalian
T→C
A→C
T→G
mammalian
Category
non−synonymous
synonymous
Gene
PTBP1
PTBP2
PTBP3


A
B
PTBP1
PTBP2
CUPrefs
Amino acid
Nucleotide
Mantel test: 0.91
p < 0.001
Mantel test: 0.05
p = 0.274
Mantel test: 0.71
p < 0.001
Mantel test: 0.56
p < 0.001


PTBP2
PTBP1
PT
Gro
Gro
GC3 content
90
80
70
60
50
40
30
Bootstrap (BS)
BS ≥ 80
50 ≤ BS < 80
Group
Mammalian
Non-mammalian
0.2


A
B
PC1 (68.81%)
PC2 (6.79%)
Category
mammalian
non-mammalian
protostoma
Gene
PTBP1
PTBP2
PTBP3
Protostoma PTBP
Gene
1.6
0.8
0.0
−0.8
−1.6


A
PTBP1
PTBP2
PTB
Intronic GC content (%)
B
Flanking GC content (%)
GC3 content of CDSs (%)
GC3 content of CDSs (%)
GC3 content o


GC conte
25
0
25
0
B
100
75
50
25
0
GC3 content of CDSs (%)
Wilcoxon signed rank test:
vs PTBP2 : mean diff= 48
S= 539.5, p-value<0.0001
vs PTBP3 : mean diff= 43.5
S= 517.5, p-value<0.0001
100
75
50
25
0
Wilc