Ca^{2+} and Membrane Potential Transitions During Action Potentials are Self-Similar to Each Other and to Variability of AP Firing Intervals Across the Broad Physiologic Range of AP Intervals During Autonomic Receptor Stimulation

Dongmei Yang[1], Christopher H. Morrell[1,2], Alexey E. Lyashkov[1], Syevda Tagirova[1], Ihor Zahanich[1], Yael Yaniv[3], Tatiana M. Vinogradova[1], Bruce D. Ziman[1], Victor A. Maltsev[1], Edward G. Lakatta[1*]

[1] Laboratory of Cardiovascular Science, National Institute on Aging, National Institutes of Health, Baltimore, USA.

[2] Mathematics and Statistics Department, Loyola University Maryland, Baltimore, USA.

[3] Biomedical Engineering Faculty, Technion-IIT, Haifa, Israel.

Short title: pacemaker Ca^{2+} and Vm self-similarity

[*] **Correspondence:**

Edward G. Lakatta, M.D.
Laboratory of Cardiovascular Science, NIA/NIH
Biomedical Research Center,
251 Bayview Blvd., Baltimore, MD 21224, USA.
Tel.: 410-558-8202; Fax: 410-558-8150
E-mail : lakattae@grc.nia.nih.gov

Key words:

single sinoatrial nodal pacemaker cells; Local diastolic Ca^{2+} releases; diastolic depolarization; autonomic receptor stimulation; self-similarity of Ca^{2+} and membrane potential during action potentials; action potential; firing interval variability.

Abstract

Ca^{2+} and V_m transitions occurring throughout AP cycles in sinoatrial nodal (SAN) cells are **cues** that: (1) **not only regulate** activation states of molecules operating within criticality (Ca^{2+} domain) and limit-cycle (V_m domain) mechanisms of a coupled-clock system that underlies SAN cell automaticity; (2) but are also **regulated by** the activation states of the clock molecules they regulate. In other terms, these cues are **both** causes and effects of clock molecular activation (recursion). Recently, we demonstrated that Ca^{2+} and V_m transitions during AP cycles in single SAN cells isolated from mice, guinea pigs, rabbits and humans are self-similar (obey a power law) and are also self-similar to trans-species AP firing intervals of these cells in *vitro*, to heart rate in *vivo*, and to body mass.

Neurotransmitter stimulation of β adrenergic receptor or cholinergic receptor initiated signaling in SAN cells modulates their AP firing rate and rhythm by impacting on the degree to which SAN clocks couple to each other, creating the broad physiologic range of SAN cell mean AP firing intervals and firing interval variabilities. Here we show that Ca^{2+} and V_m domain kinetic transitions (time to AP ignition in diastole and 90% AP recovery) occurring within given AP, the mean AP firing intervals, and AP firing interval variabilities within time-series of APs in 230 individual SAN cells are self-similar (obey power laws). In other terms, these long-range correlations inform on self-similar distributions of order among SAN cells across the entire broad physiologic range of SAN AP firing intervals, regardless of whether autonomic receptors of these cells are stimulated or not, and regardless of the type (adrenergic or cholinergic) of autonomic receptor stimulation. These long-range correlations among distributions of Ca^{2+} and V_m kinetic functions that regulate SAN cell clock coupling during each AP cycle in different **individual**, isolated SAN cells not in contact with each other. Our numerical model simulations further extended our perspectives to the molecular scale and demonstrated that many ion currents also behave self-similar across autonomic states.

Thus, to ensure rapid flexibility of AP firing rates in response to different types and degrees of autonomic input, nature "did not reinvent molecular wheels within the coupled-clock system of pacemaker cells", but differentially engaged or scaled the kinetics of gears that regulate the rate and rhythm at which the "wheels spin" in a given autonomic input context.

Introduction

The heart is a central player within a hierarchical system of clocks operating within the autonomic neuro-visceral axis that creates and synchronizes rhythmic functions ranging from msec to days and beyond (Lakatta, 2021; Shivkumar et al., 2016). The heart's beating rate and rhythm are regulated by autonomic input to sinoatrial nodal (SAN) pacemaker cells that modulates functions within a coupled-clock system intrinsic to SAN cells (Lakatta, Maltsev, & Vinogradova, 2010).

What is the coupled-clock system within pacemaker cells and how do clocks couple to each other?

The SAN cell coupled-clock system is comprised of a calcium "clock", the sarcoplasmic reticulum (SR), that continuously oscillates Ca^{2+} via a *criticality* mechanism (Nivala, Ko, Nivala, Weiss, & Qu, 2012) and phase-like transitions (A.V. Maltsev et al., 2011); the Ca^{2+} clock is continuously but variably coupled to a "membrane clock", an ensemble of surface membrane ion channels that generates current oscillations via a *limit- cycle* mechanism (Weiss & Qu, 2020). The criticality mechanisms, in turn, are governed by power law and self-similarity across wide scales (Bak, 1999). The "biochemical engine" of the coupled-clock system is a constitutively active, Ca^{2+} calmodulin-dependent adenylyl cyclase (AC) that generates cyclic AMP, leading to modulation of cAMP-gated ion channels, EPAC signaling, and PKA and CAMKII-dependent kinase activities, mechanisms that regulate intracellular Ca^{2+} levels, Ca^{2+} dynamics and membrane potential within SAN cells (Lakatta, Maltsev, Bogdanov, Stern, & Vinogradova, 2003; Lakatta et al., 2010; Lakatta et al., 2006; Lakatta, Vinogradova, & Maltsev, 2008; V. A. Maltsev & Lakatta, 2008; Yaniv et al., 2015). Variable rates and rhythms at which SAN cells fire APs are controlled by the kinetics of sub-cellular and cell-wide transitions in [Ca^{2+}] gradients and the membrane potential (V_m), and the extent to which V_m and Ca^{2+} become coupled during AP cycles in any given epoch. For more details, see Supplementary Discussion.

The well-known variability of AP firing intervals of isolated SAN cells in *vitro*, of SAN tissue *ex vivo* or of heartbeat intervals in *vivo* (Monfredi et al., 2014; Yaniv, Ahmet, et al., 2014) indicates that coupled-clock system Ca^{2+} and V_m functions during AP cycles never achieve a true steady state from one AP to the next.

These time- dependent Ca^{2+} and V_m domain transitions during APs are **cues,** that **not only regulate** activation states of clock molecules, but are also **regulated by** the activation status of the very molecules they regulate. In other terms, changes in these cues cause changes in clock molecule activation that feedback to change the characteristics of activation cues. This recursive dynamic imparts robustness to SAN cell automaticity (Lyashkov, Behar, Lakatta, Yaniv, & Maltsev, 2018; V. A. Maltsev & Lakatta, 2009). The variability in the degree to which Ca^{2+} and membrane clock molecules couple to each other throughout AP cycles is due to transitions (changes) that occur in Ca^{2+} and V_m domain cues throughout AP cycles (Monfredi et al., 2013; Yaniv, Lyashkov, et al., 2014).

Spontaneous transitions in sub-cellular Ca^{2+} and V_m domains that emerge during the spontaneous diastolic depolarization (DD) phase of an AP cycle have been conceptualized as the AP "ignition phase" (Lyashkov et al., 2018). The ignition process in the Ca^{2+} domain is linked to the emergence of local spontaneous, diastolic oscillatory RyR activation, that generates local Ca^{2+} releases (LCRs) that self-organize to form Ca^{2+} wavelets that propagate locally (Bogdanov, Vinogradova, & Lakatta, 2001; Vinogradova et al., 2004). Ca^{2+}-dependent activation of the surface membrane electrogenic Na^+/Ca^{2+} exchanger generates inward current that accelerates diastolic V_m depolarization and couples the clocks. The time at which the rate of this feed-forward crosstalk acutely accelerates to 0.15V/s, marks the onset of the coupled-clock ignition process (Lyashkov et al., 2018).

Following ignition onset, the extent to which the Ca^{2+} and V_m clock become coupled continues to increase throughout the diastolic period as LCRs and Ca^{2+} wavelets emerge at remote areas across the cell and continue to self-organize in time throughout the cellular space, creating an explosive ensemble Ca^{2+} signal that progressively depolarizes the cell membrane, i.e., clock-coupling progressively increases. This Ca^{2+}-induced change in V_m increase in clock-coupling cues the activation of low-voltage activated Ca^{2+} channels ($Ca_v1.3$ and $Ca_v3.1$), resulting in Ca^{2+} influx

that contributes to the further organization of the ensemble LCR Ca^{2+} signal via feed-forward electrochemical (Ca^{2+}-V_m-Ca^{2+}) signaling, when the diastolic V_m enters a range that cues the activation of L type Ca^{2+} channels ($Ca_v1.2$). The ignition phase of the coupled-Ca^{2+} and V_m domain sub-cellular kinetic transitions culminates in the generation of cell-wide events; a marked transition in the rate of V_m depolarization, due to the activation of $Ca_v1.2$, results in the rapid AP upstroke and Ca^{2+} influx, which, via Ca^{2+}-induced Ca^{2+} release from the SR via RyRs, generates an AP-induced cytosolic Ca^{2+} transient. In other terms, spontaneous, cell-wide Ca^{2+} signals and APs in SAN cells emerge from spatio-temporal self-organization of spontaneous sub-cellular Ca^{2+} oscillations (the *criticality* mechanism) (Nivala et al., 2012). Serca 2a pumping Ca^{2+} into SR and K^+ channel repolarization of V_m return the Ca^{2+} and V_m domain cues toward their diastolic levels at which LCRs again begin to emerge, creating the ignition phase of the next AP cycle.

Self-organized criticality

Spatiotemporal self-organization across geometric scales (sub-cellular to cell-wide) is a manifestation of criticality that has been observed in excitable cells throughout nature (Stožer et al., 2019) including cultured astrocytes (Jung, Cornell-Bell, Madden, & Moss, 1998), immature oocytes (Lopez, Piegari, Sigaut, & Dawson, 2012) and mouse cardiac ventricular myocytes (Nivala et al., 2012) . Self-similar, scale-free distributions of parameters across wide scales that obey power law behavior (are ln-ln linear) are an indication of their self-ordered criticality (Bak, 1999).

It has recently been discovered that coupling of sub-cellular Ca^{2+} signals (cues) generated by the Ca^{2+} clock within isolated SAN cells to the cell surface membrane proteins during APs to elicit a change in V_m manifests long range power law correlations (are self-similar) across species (S. T. Sirenko et al., 2021). Specifically, Ca^{2+} and V_m domain kinetic transitions (cues) during AP cycles in single SAN cells isolated from mice, guinea pigs, rabbits and humans are self-similar to each other during APS and self-similar to trans-species AP firing intervals of these cells in *vitro*, to heart rate in *vivo*, and to body mass (S. T. Sirenko et al., 2021).

Neurotransmitter stimulation of βAR or CR-initiated signaling modulates SAN cells' AP firing rate and rhythm by impacting on coupled-clock protein functions, modulating the degree to which criticality (Ca^{2+} domain) and limit-cycle (V_m domain) mechanisms couple to each other during AP cycles (Lakatta et al., 2010; V. A. Maltsev & Lakatta, 2009). AP firing intervals in rabbit SAN cells during autonomic stimulation vary over a four-fold range, from about 200 msec during βAR stimulation (βARs) up to about 800 msec during CR stimulation (CRs) (Lyashkov et al., 2009; Vinogradova, Bogdanov, & Lakatta, 2002).

We hypothesized that transitions in V_m and Ca^{2+} domain cues during the diastolic AP ignition (Lyashkov et al., 2018) and recovery phases (Figure 1) of APs are: (1) not only self-similar to each other in cells without autonomic receptor stimulation (control cells), but are self-similar to V_m and Ca^{2+} cues in **other** cells during CRs and during βARs; and (2) that Ca^{2+} and V_m cues during APs are self-similar to AP firing interval variabilities (and therefore self-similar to mean AP firing intervals) regardless of the presence or absence or type of autonomic receptor stimulation. In other terms, we hypothesized that Ca^{2+} and V_m domain clock-coupling cues occurring during **all** APs are self-similar to each other i.e., manifest long-range correlations **in all** isolated SAN cells within populations of cells that differ with respect to autonomic input and, that these Ca^{2+} and V_m cues during APs are also self-similar to the rate and rhythm of AP firing across the **entire range** of AP firing intervals created by these cues in **all** isolated SAN cells.

To test these hypotheses: we studied a large population (n=230) of single rabbit SAN cells to which we applied: CRs (carbachol, CCh), to one subset of cells; βARs (isoproterenol, ISO) to another subset; and no autonomic receptor stimulation to a third subset of cells. This created three populations of SAN cells having APFIs distributed across the entire physiologic range. We measured intracellular Ca^{2+} or membrane potential in these cells to: (1) characterize the times to ignition onset, and times to 90% recovery of V_m and Ca^{2+} parameters during APs in AP time-series; and (2), to determine the correlations of these V_m and Ca^{2+} kinetic parameters to each other during APs, to AP firing interval variability (and therefore to mean AP firing intervals). Thus, the data set to be analyzed consisted of 12 different kinetic parameters in each cell population (control, CCh and ISO); 6 parameter means, 3 each in the Ca^{2+} and V_m domains; and 6 parameter variabilities (SDs) around the means. To determine the degree of self-similarity among V_m and Ca^{2+} domain parameters, we constructed density distribution plots and applied correlation, power law, and principal component (PC) analyses to Ca^{2+} and V_m domain data sets separately, and to the combined Ca^{2+} and V_m data sets. We further extended our perspectives from cell population and single cell levels downwards to the molecular scale by performing numerical modeling simulation and analyzing variabilities of ion currents and Ca^{2+} with respect to APFI to determine whether these ion currents and Ca^{2+} also obeyed a power law across autonomic states.

Materials and Methods

The study was performed in accordance with the Guide for the Care and Use of Laboratory Animals published by the National Institutes of Health (NIH Publication number. 85-23, revised 1996). The experimental protocols have been approved by the Animal Care and Use Committee of the National Institutes of Health (protocol #034LCS2016). Materials and methods briefly presented here are detailed in Supplementary Material.

Single, isolated rabbit SAN cells isolation.

Single, spindle-shaped, spontaneously beating SAN cells were isolated from the hearts of New Zealand rabbits (Charles River Laboratories, Wilmington, MA) as described previously (Vinogradova TM, 2000).

Spontaneous APs recordings.

Time series of spontaneous APs were recorded in subsets of freshly isolated SAN cells using the perforated patch-clamp technique with Axopatch 200B patch-clamp amplifier (Axon Instruments) (Bogdanov et al., 2001) at 34±0.5ºC. AP parameters (Figure 1) were measured via a customized program (Lyashkov et al., 2018) were APFI, APD_{90}, and the time to ignition onset (TTIO) measured by the time at which diastolic membrane potential dV/dt accelerates to 0.15 V/sec (Figure 1) which reflects the onset of the ignition phase of the AP cycle (Lyashkov et al., 2018).

Ca^{2+} measurements.

In another subset of SAN cells, AP-induced global Ca^{2+} transients and spontaneous LCRs (Figure 1) were measured at 34±0.5ºC with a confocal microscope (Zeiss LSM510, Germany) in the line-scan mode (Vinogradova et al., 2004; D. Yang, Lyashkov, Li, Ziman, & Lakatta, 2012). The interval between the peaks of two adjacent AP-induced Ca^{2+} transients (Figure 1) is defined as Ca^{2+} transient (CaT) firing interval (CaTFI), which is highly correlated with the APFI, as demonstrated by simultaneous recordings of V_m and Ca^{2+} in a separate subset of cells (Figure 1).

The LCR period is defined as the time from the peak of the prior AP-induced Ca^{2+} transient to an LCR peak in diastole (Figure 1); the time to 90% decay of the CaT was defined as CaT_{90}.

Numerical modeling.

We performed numerical simulations using a modified Maltsev-Lakatta model that features the coupled-clock mechanism (V. A. Maltsev & Lakatta, 2009). The computer code for the original model is freely available and can be downloaded and run in CellML format (http://models.cellml.org/workspace/maltsev_2009) using the Cellular Open Resource software developed by Alan Garny at Oxford University in the UK (Garny, Noble, Hunter, & Kohl, 2009) (for recent development of this software see http://www.opencor.ws/). The original model could not be directly used for APFI variability simulations because it is a system of first-order differential equations that is deterministic and showing no APFI variability in limit cycle oscillatory regime of AP steady firing. Thus, we modified the model to generate variability of AP waveforms by supplementing total membrane current (I_{tot}) with an additional randomly fluctuating current around its zero-mean value, known as perturbation current or Iper (as previously implemented by Henggui Zhang (Monfredi et al., 2014)). Furthermore, we also performed an additional set of simulations with I_{per} added to Ca^{2+} release flux to mimic the effect of stochastic LCRs (Bogdanov et al., 2006; Monfredi et al., 2013). Using the resultant stochastic dynamical system of SAN cell, we simulated fluctuating APFI, ion currents, and Ca^{2+} dynamics for three conditions: (i) basal AP firing, (ii) during βARs with ISO (100 nM), and (iii) during CRs with CCh (100 nM). The effects of autonomic modulation (conditions ii and iii) were modelled as previously described (V. A. Maltsev & Lakatta, 2010), except modulation of I_{CaL} current by CCh that was modelled as described by Zaza et al. (Zaza, Robinson, & Difrancesco, 1996). All model equations and parameters are provided in Supplementary Material.

Experimental design and statistics.

Supplementary Figure S1 illustrates schematic of the experimental design to assess long-range correlations of V_m and Ca^{2+} parameters during APs and APFI intervals in cells within and among populations of cells that differed with respect to autonomic input. V_m and Ca^{2+} parameter intervals (msec) are presented as Mean ± SD. AP firing interval variability within a time series is taken as standard deviation (SD) about the mean, or as the coefficient of variation (CV, the ratio of SD to the mean).

Analyses of V_m and Ca^{2+} parameter interval distributions measured in AP time-series in **different** cells determined the association among pairs of variables using Spearman's correlations for average data, and Pearson's correlation for both average and individual data (Howell, 2002); In cells in which Ca^{2+} was measured, the mean interval between AP- induced CaTs was usually longer than that the mean APFI in cells in which APs were recorded (due to slight buffering effects of the fluorescent Ca^{2+} probe). To allow all the variables to be combined into a single analysis, we matched on the APFI variable Z-scores in control, ISO, or CCh populations by 'matchit' function in R (Ho, Imai, King, & Stuart, 2011).

Density estimates of **within cell** standard deviations and means of parameters of cells within each autonomic state population, are presented as nonparametric kernel estimates of probability density functions, scaled so that the total area under each curve is unity (Silverman, 1986).

In order to determine whether distributions of parameter means and SDs are self-similar, i.e., obeyed a power law suggesting fractal-like behavior, we constructed ln-ln plots of distributions of the AP and Ca^{2+} function means and SDs measured across the broad range of apparent steady states in the absence of, or the presence of βARs or CRs. (Kucera, Heuschkel, Renaud, & Rohr, 2000; Yaniv, Lyashkov, & Lakatta, 2013).

The relationships among all distributions of all parameters (means and SDs), were also assessed in principal component analyses (Johnson & Wichern, 2008).

In a few cells in which experimentally measured parameter interval distributions measured in the V_m and Ca^{2+} domains in the same cell and for numerical simulation of ion currents and Ca^{2+} prior to and during autonomic receptor stimulation Poincaré indices were employed to define long-range correlations among variables. A Poincaré Plot graphs a parameter (*n*), in an AP time series on the *x*-axis versus the same parameter of the succeeding AP (n+1) on the *y*-axis, i.e. one takes a sequence of parameters and plots each one against the following parameter (Huikuri et al., 2000).

When statistical inference was performed, a p- value<0.05 was considered statistically significant.

Results

Assessment of self-similarity of Ca^{2+} and V_m kinetic interval parameters to each other during APs and to AP firing interval variability

Figure 1 A illustrates a time-series of APs in a SAN cell during which Ca^{2+} and V_m were simultaneously measured in the absence of autonomic receptor stimulation. Kinetic transitions in Ca^{2+} and V_m parameters as illustrated in Figure 1B, were assessed during each AP. Figure 1C shows that V_m and Ca^{2+} parameters during AP time series are self-similar to each other and are also self-similar to the AP firing intervals (r=0.844 in this cell).

Figure 2 illustrates the time series of AP intervals in Figure 1A plotted as phase-plane diagrams in which V_m is depicted as a function of Ca^{2+} throughout each cycle. The times at which various channels are activated throughout the V_m/Ca loop are indicated. The times of ignition and 90% recovery are also indicated. The point resolution is 3.072ms, and the point spread indicates the rates at which the electrochemical signal changes during each cycle. The dashed line in the figure marks the border between the disordered and ordered molecular activation. The arrows indicate the direction of the electrochemical signal emergence.

A Poincaré plot (scatter graph) constructed from consecutive data points in a time-series (Figure 3A) is a convenient tool that provides information on correlations (self-similarity) of data across the time-series. The X axis defines the parameter (n) occurrence in msec, and the Y axis defines the parameter occurrence at (n+1). The Poincaré plot in Figure 3A depicts the data of the time-series of the cell in Figure 1. Note that although the means vary over a three-fold range, all six parameter means (the Ca^{2+} and V_m interval parameters measured during APs and AP firing intervals) in the absence of autonomic receptor stimulation are described by a line of identity, indicating their self-similarity across the AP time series. Quantitative analysis of short and long term variability in a given time series of observations entails fitting an ellipse to each cloud of data points within the Poincaré Plot (Figure 3B): The length of a line describing the slope of the long axis of each ellipse is referred to as SD2 of the data points (c.f. Figure inset); the length of the line describing the slope of the short axis, which is perpendicular in direction to the long axis line, is referred to as SD1. Note in Figure 3C that the SD1 is self-similar to SD2 across the four-fold range

of Ca^{2+} and V_m parameters. The center point of each ellipse i.e., the intersection of SD1 and SD2, is the **average** interval between events (AP intervals or other parameters measured in the time series) within the time series. SD1/SD2 (Figure inset) informs on non-linear trends (unequal lengths of SD1 and SD2) across intervals within each ellipse.

Figure 4 illustrates combined Poincaré plots of TTIO, APD_{90}, during APs and AP firing intervals in time-series of APs of two representative cells: one cell in control and during CRs by CCh; and the other cell in control and during βARs stimulation by ISO.

Although the range of absolute values of kinetic interval parameters of cells depicted in the Poincaré Plot in Figure 4B vary by 20-fold, all points (n=5673) are self-similar, i.e. are fit by a single line (r=0.992) with a slope of unity, passing nearly through the origin.

Supplementary Table S1 lists the SD1s, SD2s, SD1/SD2, the means of the TTIO and APD_{90} intervals and AP firing intervals depicted in Figure 4B. Note also that the SD1s, SD2s, and SD1/SD2 of TTIO, APD_{90} and AP firing intervals progressively increase from ISO to control, and **markedly** increase from control to CCh, creating degrees of non-linearity across the combined control, ISO and CCh states, which is also reflected in the mean AP firing intervals across the three states (Supplementary Table S1). The V_m transitions during APs across different autonomic states are self-similar to APFIs across these states (Figure 5A). Figure 5B shows the self-similarity of parameter means of SD1s of V_m parameters to their SD2s across autonomic states, indicating self-similarity of short term (e.g., beat to beat) and long term (e.g., rhythm across more than 2 beats) variabilities across autonomic states within the time series. Figure 5C shows the self-similarity of all parameter means depicted in Figures 4, 5A and 5B to their SDs across autonomic states.

The data in Figures 4 and 5 demonstrate that over the entire range of physiologic AP firing intervals from 192.7ms in ISO to 305.5 ms in control, and to 910.3 ms in CCh, variabilities of TTIO and APD_{90} measured <u>during</u> APs are self-similar to each other and are also self-similar to the variability of AP firing intervals within the time-series, and therefore self-similar to the mean AP firing interval of the time-series.

Self-similarity among V_m variables in the cells in Figure 4 across autonomic states in control and during ISO and CCh in Figures 2 and 3A can also easily be ascertained from the shapes of their population density distributions (Figure 6). Note that in the cell super-fused with CCh the distributions of TTIO, APD_{90} and AP firing intervals (Panels A-C) are broader than in control or during ISO. Note also that the distributions of kinetic transitions during APs and APFI become more synchronized from CCh to ISO (Panels A-C). In other terms, the degree to which V_m and Ca^{2+} parameters are synchronized during APs increases from CCh to control to ISO, similar to the AP firing variabilities and mean AP firing intervals.

Self-Similarity of Ca^{2+} and V_m parameter <u>means</u> to each other during APs and to APFI variabilities and mean AP firing intervals across autonomic states in <u>different</u> cells.

We next determined whether the self-similarity (long-range correlations) of Ca^{2+} to V_m parameters measured within the same cells as depicted in Figures 1-6 extends to populations of **different** cells within and among different autonomic states. To accomplish this, we applied CRs (carbachol, CCh), to one subset of cells; βAR stimulation (isoproterenol, ISO) to another subset; and no autonomic receptor stimulation to a third subset of cells. This created populations of SAN cells having APFIs distributed across the entire physiologic range.

Table 1 lists descriptive statistics (means and SDs) of the variabilities and means of Ca^{2+} and V_m kinetic parameter intervals measured in **different** cells within and among populations of cells that **differ** with respect to autonomic input. The **mean SD** of V_m and Ca^{2+} domain parameters listed in Table 1A gives the average time-series variability of each parameter among cells within **each** of the three cell populations (control, ISO, or CCh). The standard deviation of the SDs (**SDSD**) in Table 1A tells us how variable the SDs of each parameter are among cells within **each** cell population. The **means of each parameter** measured within a time series (Table 1B) tells us the **average level** of the parameter among cells within each of the three populations; and the **SD of the means** tells us the **variability of the mean** parameter levels among cells within each cell population.

Figure 7 A illustrates the distributions of **SDs** of parameters within Ca^{2+} and V_m domains **during** APs and, of AP firing intervals in cells listed in Table 1A of each of the three populations of cells that differed in autonomic state: control cells (n=78); cells during super-fusion with ISO (n=27); and in cells super-fused with CCh (n=10). The self-similarity (long-range correlations) of the mean SDs of Ca^{2+} to V_m parameter transitions during ignition and recovery phases of APs across the wide range of AP firing intervals induced by the type of autonomic receptor stimulation, or lack thereof, is evident in the self-similarity of their mean **SD** distribution **shapes** (Figure 7A**)**.

The distribution of the **means** listed in Table 1B are illustrated in Figure 7B. Note that the **shapes** of the distributions are self-similar to each other across the three different autonomic states. Note also that the shapes of the distribution of the **means** of a given parameter in Figure 7B are similar to the distribution of that parameter's SDs in Figure 7A (because the interval distribution means stem from the distributions of their SDs). In other terms, the variability in the times at which parameters occur within a time series (their parameter SDs) determines what the mean interval of events in the time series will be.

Also note in Figure 7, that compared to cells not super-fused with an autonomic receptor agonist (control cells) and those super-fused with ISO, the shapes of the distributions of cells super-fused with CCh are broad, indicating marked variability among CCh cells within the parameter distributions of both interval means and interval SDs.

Figure 8A illustrates the self-similarity of V_m parameter means and SDs to Ca^{2+} means and SDs across autonomic states. Heat maps of the long-range correlations among V_m and Ca^{2+} parameter means are shown in Supplementary Figure S3. The long- range correlations (self-similarity) between the means of the means and means of SDs given in table 1 is shown in Figure 8B.

Self-similarity of Ca^{2+} and V_m parameters among a<u>ll individual cells</u> within and among the three autonomic states

Two by two correlations of V_m and Ca^{2+} parameter **means** and **SDs** of all (230) cells that comprised the three different cell populations in Figure 7 and Table 1 are highly significant (Table 2).

Figure 9 shows ln-ln plots of the distributions of means and SDs of Ca^{2+} and V_m parameters of all cells across the three autonomic states. The piecewise linear fit of the data in each panel is largely driven by the data from cells super-fused with CCh that manifested broad interval distributions and high mean APFIs in Figures 4 to 7. Figure 8C shows that the means and SDs of

V_m parameters measured during APs in individual cells (n=115) are self-similar to Ca^{2+} means and SDs measured during AP in other individual cells (n=115) across the three autonomic states.

Although, as noted above, the V_m and Ca^{2+} parameters within and among cells during CCh super-fusion were more broadly distributed than those during control or during ISO, the correlations between V_m and Ca^{2+} parameters among all cells **within** the CCh super-fused population of cells were extremely strong for most parameters (Supplementary Figure S3). Weaker but still significant correlations between times to 90% recovery and other variables are observed in CCh super-fused cells (likely because times to 90% are the most difficult parameters in the data set to measure accurately).

Figure 8D shows that V_m and Ca^{2+} parameters measured during APs are self-similar to AP firing interval means and SDs across the three autonomic states.

Correlation of Ca^{2+} and V_m domain parameter means in individual cells to their SDs within and across autonomic states

The relationship between mean AP firing rate and its SD is known to be non-linear (Monfredi et al., 2014). Although the relationships of **all** Ca^{2+} and V_m parameter **means** relative to their corresponding **SDs** measured in the combined set of data derived from different populations of cells in control or during ISO or CCh super-fusion are non-linear (Figure 10, A-C), the ln-ln plots of this combined data (Figure 10, Panels D-F), however, are linear, indicating their self-similarity across **all** 230 cells that differed by autonomic state.

Principal component analyses

Next, we employed principal component analyses to determine whether the self-similarity of parametric measures within the entire data set of variables could be summarized by a smaller set of principal components that contain most of the information in all the variables. PCs are linear combinations of the original variables, and each PC is statistically independent of the others: the first PC explains as much of the total variability in the data as possible, the second PC as much of the remaining variability, and so on. Highly self-similar parameters within the complete data set are explained by the sum of the first few PCs.

In a PC analysis of SDs of the six variables (3 in the V_m domain and 3 in the Ca^{2+} domain) the first 3 PCs explained 91.4% of the total variation within the entire SD data set (Supplementary Table S2, Figure 11A). Similarly, in a PC analysis of the means of all six measured variables means (3 in the V_m domain and 2 in the Ca^{2+} domain) in a PC analysis, the first 2 PCs explained 92.8% of the variability in the data set of all 6 means (Supplementary Table S2, Figure 11B). Finally, in a PC analysis of all 12 variables (6 means and 6 SDs), the first 3 PCs explained 88% of the variability within the total (means plus SDs) data set (Supplementary Table S2, Figure 11C).

Because a smaller set of PCs can explain a substantial proportion of the total variability in each set of V_m **or** Ca^{2+} domain means, SDs, and means and SDs, means that these distributions of V_m and Ca^{2+} parameters measured during an AP, and AP firing intervals in control cells and different cells super-fused with ISO or CCh are each self-similar to each other. In other terms, Ca^{2+} and V_m domain functions operative within the SAN cells coupled-clock system manifest self-similar-scale free characteristics i.e., are kinetic fractals of each other, across the entire physiologic range of AP firing intervals.

Numerical model simulations of APFIV, major ion currents, and Ca^{2+}.

Variability of V_m and Ca^{2+} parameters measured experimentally in cells within and across autonomic states is linked to the respective variabilities of clock molecular availability to respond to V_m and Ca^{2+} cues (Figure 2) that cannot be directly measured experimentally during AP firing. To gain further insight into the variability of these biophysical mechanisms, we performed numerical modeling simulations. The APFI variability was generated by SAN cell model described as a stochastic dynamical system, i.e., a dynamical system (deterministic Maltsev-Lakatta model (V. A. Maltsev & Lakatta, 2009)) subjected to the effects of noise current, I_{per} (see Methods and Supplementary Material for details). I_{per} amplitude was tuned for the model APFIV to match that measured experimentally under respective experimental conditions. We investigated two scenarios of noise generation: when I_{per} was added to I_{tot} or when I_{per} was added to Ca^{2+} release flux current in each of the three autonomic states: (i) basal AP firing, (ii) ISO 100 nM, and (iii) CCh (100 nM). Variability of 6 major currents was simulated and analyzed: I_f, I_{NCX}, I_{Kr}, I_{CaL}, I_{CaT} and I_{KACh}. Variability of [Ca] under cell membrane was also simulated during the three autonomic states. For all items we measured variability of their peak amplitudes and amplitudes at -40 mV during DD.

Model simulation results are presented in Figure 12, with their numerical values given in supplementary Tables S6 and S7. Regardless of the type of noise generation (via Ca^{2+} or I_{tot}), it affected the variability of ion currents and Ca^{2+} the same way and the predicted variabilities for many parameters differed substantially from that of APFI:

1) I_f variability was substantial: in the basal state and in ISO I_f variability was similar to or larger than APFI variability; The variability of I_f decreased in CCh.

(2) I_{NCX} variability was also substantial: at -40 mV it was substantially larger than that of APFI variability (except in CCh when I_{per} was added to I_{tot}); variability of I_{NCX} peak amplitude (negative, during AP upstroke) was similar to that of APFI in the basal state and ISO, but became reduced in CCh.

(3) I_{Kr} variability was substantially less than APFI variability under all conditions.

(4) I_{CaT} variability was the largest among ion currents, being similar to that of $I_{NCX,}$ in CCh when I_{per} was added to Ca^{2+} release.

5) Peak I_{CaL} variability was always less than that of APFI variability; at -40 mV it was greater in ISO than in basal state and CCh.

(6) Variability of Ca^{2+} release flux in the basal state and ISO at -40 mV was greater than or similar to APFI variability; in CCh variability of Ca^{2+} release flux was less than that of APFI.

Some components exhibited power law behavior over a wide range of APFI over all conditions tested (Figure 13).

We next determined whether self-similarity across autonomic states observed for experimental data during the ignition phase is also applied to simulated ion currents or Ca^{2+} data during this time of the cycle, i.e. at -40 mV. To this end, we applied the statistical tests utilized for experimental data to simulated data (for the scenario when I_{per} was added to I_{tot}).

Simulated ion currents and Ca^{2+} amplitudes during AP ignition (-40 mV) across the three autonomic states are self-similar to each other, strongly correlated to each other, as were experimentally measured parameters (Table 2). These two-by-two correlations of all the simulated components are listed in Supplementary Table S3. Selected examples of these correlations are shown in Figure 14. As I_{NCX} is fully determined by V_m and Ca^{2+}, at a fixed voltage (-40 mV) it is

fully determined by only Ca^{2+}. That is why we have 100% correlation of I_{NCX} and Ca^{2+}. I_{CaT} strongly correlated with Ca^{2+} variations, because the stronger Ca^{2+} signal is linked to the higher DD rate and hence stronger (time-dependent) activation of I_{CaT}. Surprisingly I_{KACh} amplitude was also highly correlated with Ca variations.

Variations in I_f and I_{Kr} at -40 mV did not depend on variations of Ca^{2+}, but their mean values strongly depended on Ca^{2+} across the autonomic states. I_f activation and I_{Kr} deactivation are early DD mechanisms and do not seem to interplay with Ca^{2+} at the ignition onset at -40 mV in a given cycle.

Figure 15A illustrates the Poincare plots of many simulated parameters (APFI and TTIO; I_{NCX}, I_{CaT}, I_{Kr}, [Ca], I_{CaL}, I_{KACh}, and I_f, all at -40 mV). Although the range of absolute values of these simulated parameters substantially vary, all simulated parameters are self-similar, i.e. are fit by a single line (r=0.998) with a slope of unity, resembling Poincare relationship of experimentally measured AP parameters (Figures 1 and 4).

Figure 15B illustrates ln-ln plots of the relationships of the means of simulated components to their SDs. Note that this relationship follows power law behavior just as did the relationship of experimentally measured means for AP parameters vs their SDs (Figure 10).

Finally, in PC analyses of the simulated parameters in Figure 15B the first 2 PCs accounted for 94% of the variation in the eight variables (Supplementary Figure S4).

Discussion

We measured membrane potential and Ca^{2+} times to onsets of AP ignition during diastole, and times to 90% recovery during APs, and AP firing intervals in 3 populations of single, isolated rabbit SAN cells that differed with respect to autonomic input: those in which CRs were stimulated by CCh; those in which βARs were stimulated with ISO; and in untreated (control cells). **Absolute values** of times to AP ignition onsets and to 90% recovery intervals in Ca^{2+} and V_m domains during APs and AP firing intervals differed within and among individual cells in cell populations and different autonomic states, and differed **markedly** among cells of the three populations of cells with differential autonomic receptor stimulation. Our novel finding is that although differing markedly in absolute values, Ca^{2+} and V_m parameters were self-similar to each other during APs and self-similar to AP firing intervals, not only within and among different cells **within** each of the three populations of cells studied, but, remarkably, **among all** cells, regardless of the autonomic receptor stimulation profile. Thus, Ca^{2+} and V_m domain kinetic transitions (intervals) during APs, individual AP firing interval and mean AP firing intervals within AP time series manifest long-range correlations (self-similar scale-free correlations, i.e., obey power law) across the entire broad range of AP firing intervals, regardless of whether autonomic receptors of these cells are stimulated or not, and regardless of the type of autonomic receptor stimulation.

The degree to which molecular activation states within each clock and between clocks are synchronized during APs determines when the next AP will occur, i.e. the AP firing interval variability and mean AP firing intervals within a cell and across the entire population of single SAN cells studied: the higher the degree of order (self-organized activation of clock molecules) the more ordered and less variable the aggregate of kinetic functions, the least variability of AP firing intervals and the shorter the **mean** AP firing interval; vice-versa, the lower the degree of order among clock molecular activation states the lower the aggregate synchronization among

clock molecular functions, the greater the variability of AP firing intervals, the longer the **mean** AP cycle interval.

Self-similar or fractal-like beating rate variability among cardiac cells in culture has been previously identified in a number of studies **but only when cells were confluent**, or electrically connected to each other. This behavior has been attributed to influences of tonic or phasic resetting of membrane potential, or to mechanical factors via cell-to-cell connections (Clay & DeHaan, 1979; Jongsma, Tsjernina, & de Bruijne, 1983; Kucera et al., 2000). Our novel observation is self-similarity of V_m and Ca^{2+} domain intervals during APs and AP firing intervals across diverse populations of single SAN ells that were not physically connected to each other.

Thus, self-similar distributions of order that have been demonstrated to occur in other instances throughout nature (Bak, 1999), also exist within SAN cell coupled-clock system functions. We interpret this power law behavior of SAN cell functions to result from concordant gradations of self-organized order (synchronization) of clock molecular activation across the entire physiologic range of AP firing intervals.

Clock molecular activation cues

Voltage, time, Ca^{2+}, cAMP signaling, and PKA and CaMKII-dependent clock protein phosphorylation, are the cues regulate the activation kinetics of molecules that control pacemaker functions in single SAN cells (Supplementary Figure S2) (Lakatta et al., 2003; Lakatta et al., 2010; Lakatta et al., 2006; Lakatta et al., 2008; V. A. Maltsev & Lakatta, 2008; Yaniv et al., 2015). Some coupled-clock system proteins are activated by $Ca^{2+,}$ e.g. SERCA 2; others by V_m and cAMP binding, e.g. HCN channels (DiFrancesco & Tortora, 1991) and other cyclic nucleotide-regulated channels; or by Ca^{2+} **and** V_m, e.g. NCX, or by phosphorylation **and** Ca^{2+} e.g. phospholamban and Ryanodine Receptors and AC type 8; while the activation states of still other coupled-clock system proteins are modulated by V_m, Ca^{2+} **and** phosphorylation, e.g. L type and some K^+ channels.

Both voltage and Ca^{2+} activation cues oscillate in amplitude throughout each AP cycle and command rapid responses from clock molecules. The degree to which activation status of molecules of a given species is synchronized at any given time following the prior AP determines the ensemble response of that molecular species to its activation cues. It is well documented that following a synchronizing event, e.g. the occurrence of an AP, activation states of molecules underlie AP cycle transition through variably inactivated states, altering the availability to respond to a subsequent activation cue. Our new concept of synchronization of functional cues is based on the idea that the coupled-clock system inheres (inevitably) some degree of disorder that stems from its key constituent proteins operating (stochastically switching) intrinsically within their conformational flexibilities and heterogeneity. The balance of order/disorder is linked to molecule interactions (i.e. effectiveness of their respective cues) that allows them to operate cooperatively as an ensemble or system with various degree of synchronization (i.e. order) that is reflected in respective variability of the output function of the system, i.e. APFI variability in our case.

Thus, we interpret the experimentally measured concordant behavior of surface membrane and Ca^{2+} regulatory functions during AP cycles across the entire physiologic range of AP cycles to reflect a concordance in the degrees of activation of molecules that drive these regulatory functions. Importantly Ca^{2+} and V_m cues **not only** regulate the synchronization clock of molecular activation states but are also **regulated by** the degree of synchronized activation of molecules determined by these cues (recursion). Because membrane and Ca^{2+} clocks become coupled in the context of the electrochemical signal that waxes and wines to cause the AP cycle, the extent of

self-organized molecular activation **within** each clock indirectly affects self-organization of molecular activation of the **other** clock operating within the coupled-clock system. And because scaling of **mean** APFIs among all cells is self-similar to APFI **variability** among cells, AP firing variability and mean APFI are determinants of the Ca^{2+} and V_m cues that determine kinetic intervals during an APs: a recursive, feed-forward process.

Mean APFI and APFIV are not only regulated by, but also regulate the degree to which clock molecular functions are synchronized

Changes in Ca^{2+} and V_m cues during an AP, not only determine the characteristics of that AP, but also determine when the next AP will occur, and the mean AP firing interval within an AP time-series.

A prolongation of the mean APFIs, itself, contributes to the concurrent increase in the AP firing interval variability at long mean APFI: because an increase in mean APFI reduces net Ca^{2+} influx, and indirectly reduces Ca^{2+}/cAMKII-AC-dependent phosphorylation of Ca^{2+} cycling proteins, reducing the SR Ca^{2+} cycling kinetics and increasing the variability of LCR periods.

Characteristics of the AP that are determined by availability of M clock molecules to respond to a change in membrane potential, both directly and indirectly entrain the Ca^{2+} and M clock activation: As the mean AP interval shortens, less time elapses between APs, and therefore at shorter intervals less time is required than at longer intervals for molecules to retain (remember) the synchronizing influences imparted by the preceding AP. This causes the relationship of mean APFI to APFIV of isolated SAN cells to be non-linear (Figure 5 and Supplementary Figure S2), as originally demonstrated by ZaZa (Zaza & Lombardi, 2001) and later by Monfredi et al (Monfredi et al., 2014). Conversely, as time following a prior AP increases, the effectiveness of the Ca^{2+} activation cue, itself, wanes, because the cell Ca^{2+} level and SR Ca^{2+} load become reduced, due to time-dependent Ca^{2+} efflux from the cell. We may speculate, therefore, that during long AP cycles, fewer molecules of some molecular species are available to respond to Ca^{2+} activation cues.

Gradations of self-organized molecular activation within and between clocks, regulate the APFI rhythm i.e., the APFI variability. In other terms, the average APFI, kinetics of the AP, AP-triggered Ca^{2+}-transient, LCR periods and diastolic depolarization kinetics, and beat-to-beat variability of these parameters measured in the present study are readouts of the relative extents to which of clock molecules become activated and the degree to which the clocks are coupled. When the degree to which Ca^{2+} and M clocks kinetics are coupled or synchronized is low, the AP firing rate is slow, and AP firing interval variability is high, e.g., during CRs. Conversely when the degree of coupling or synchronization of the Ca^{2+} and V_m kinetics of the two clocks is high e.g., during βAR stimulation, AP firing is rapid and AP firing interval variability is low.

So, what factors affect the degree of synchronization of clock molecules?

Concordant degrees of self-similar synchronization of M and Ca^{2+} clock kinetic functions reflect concordant gradations of activation states of specific molecules that govern these functions and how these cues change throughout an AP cycle.

AP firing rate and rhythm synchronization of clock molecules

The AP that emerges from the diastolic ignition events is, itself, the most potent integrator or synchronizer, not only of surface membrane electrogenic molecules, but also of Ca^{2+}-clock functions: a synchronized global cytosolic Ca^{2+} transient that ensues following synchronous

activation of voltage-dependent L-type Ca^{2+} channels is created by synchronized Ca^{2+}-induced, Ca^{2+} release from SR via ryanodine receptor activation. (Lakatta, 2004; Song, Sham, Stern, Lakatta, & Cheng, 1998; Wang, Song, Lakatta, & Cheng, 2001; Zhou et al., 2009).

The efficacy of V_m and Ca^{2+} activation cues that oscillate as electrochemical signal that underlies the V_m change during AP cycle varies with the AP cycle interval or period: shorter periods (i.e., faster AP firing rates or shorter APFIs) are more effective than longer periods (i.e., slower AP firing rates or longer APFIs), because during very long AP cycles, Ca^{2+} activation states of some molecules become more unsynchronized. At very short times following a large voltage oscillation (i.e., an AP), many molecules of a given molecular species in relatively inactivated state may not optimally respond to activation cues (e.g., impaired excitability/non-excitability). As the time following a prior activation increases, although a sub-population of molecules of given species may regain full ability to respond to activation cues, substantial variability in the activation status of other molecules of that species still may exist, limiting the number of molecules that can respond to (be recruited by) an activation cue. Our results provide novel clues to the cellular basis for the observation that an AP occurrence, itself, influences the range of APFIs that immediately follow it (Nolasco & Dahlen, 1968). The AP, itself, indirectly affects all Ca^{2+}-clock functions because it regulates net cell Ca^{2+} balance. Functions of M-clock molecules that underlie the generation of an AP *indirectly* regulate the availability for SR Ca^{2+} cycling by modulation of the level of cell Ca^{2+}, the SR "oscillatory substrate". Thus, M clock functions also *indirectly* regulate LCR periods and sizes via their impact on the "steady state" intracellular Ca^{2+} level. When the average interval between APs becomes prolonged, a reduction net Ca^{2+} influx into: efflux from the cell (Lakatta, 2004) reduces the cytosolic [Ca^{2+}], the rate of Ca^{2+} pumping into SR, and the SR Ca^{2+} load. These reductions, in turn, prolong the average time from the prior AP occurrence for spontaneous local diastolic ryanodine receptor activation to occur within SAN cell local micro-domains; the randomness of spontaneous local diastolic ryanodine receptor activation occurring within these micro-domains also increases, broadening the distribution of LCR periods and shifting these to longer times at long AP cycle (Figure 4).

Thus, the degree of variability in activation states of M and Ca^{2+} clock molecules that emerges over time following their synchronization by the prior AP is implicated in the cycle length dependence of variability of Ca^{2+} and M clock functions measured here (Table 1, Figures 3-5). Heartbeat variability in *vivo*, and AP firing interval variability (APFIV) of isolated SAN cells in *vitro* indicate that, neither autonomic input to SAN cells, nor functions intrinsic to the SAN cell coupled-clock system, respectively, achieve a steady-state from one beat to the next.

Ca^{2+}-dependent synchronization of clock molecules

The local [Ca^{2+}], itself, also serves as a powerful synchronizer of clock molecular function: ordered/disordered Ca^{2+} regulation has been recently reported for ryanodine receptor -mediated Ca^{2+} releases (A. V. Maltsev, Stern, & Maltsev, 2019).

Studies in **permeabilized** SAN cells, in which Ca^{2+}-clock function is preserved, but M clock function is abolished, and therefore AP's cannot occur and do not influence LCR periodicity, clearly demonstrate that: in a fixed, physiologic, free [Ca^{2+}], LCR occurrences are random when the free [Ca^{2+}] is low; and that LCR periodicity emerges as the free [Ca^{2+}] in the system is increased, due in part, to an increase in the Ca^{2+} charge of the SR capacitor (S. Sirenko et al., 2013). The intracellular concentration of the oscillatory substrate, Ca^{2+}, itself is regulated, in part, by the SAN

cell transmembrane Na^+ gradient and membrane potential (S. Sirenko et al., 2016; S. Sirenko et al., 2013).

cAMP activation or phosphorylation of clock proteins modulate the synchronization of and response to activation cues

Autonomic receptor stimulation modulates both the activation cues and responses of clock molecules to these cues. The impact of autonomic receptor signaling on the effectiveness of clock-coupling occurs over several AP cycles, and is reflected in time-dependent transitions in the AP firing rate and rhythm. The kinetics and stoichiometry of increases in PKA activity in response to gradations in βAR stimulation predict the kinetics and stoichiometry of concurrent time-dependent increases in AP firing rate (Yaniv, Lyashkov, et al., 2014). Prior studies (Lyashkov et al., 2009; D. Yang et al., 2012) have demonstrated that gradations in the phosphorylation status of phospholamban at Ser^{16} across the 3 autonomic state mean APFI's of cell populations in the present study (Supplementary Figure S3) strikingly resemble gradations of the means of APFIs and APFIVs observed across these autonomic states measured in the present study (Table 1).

The extent to which clock molecules respond to Ca^{2+} or V_m activation cues during an AP is **modulated** by βARs or CRs dependent-phosphorylation of many of the **same** proteins that drive SAN cell automaticity in the absence of autonomic receptor activation. This βARs or CRs impacts on the memory of the extent to which clock molecules activation had been synchronized during the prior AP. βARs or CRs modulation has two facets: (1) a direct effect, due to cAMP or phosphorylation-dependent activation of clock proteins and (2); an indirect effect by altering the APFI, which alters the cell Ca^{2+} activation cues that are directly modulated by autonomic receptor stimulation. Specifically, βARs and CRs not only, respectively, reduce or increase mean APFI, but also, respectively, shift variability within distributions of Ca^{2+} and V_m functions in the same direction (S. Sirenko et al., 2016) as the shift in mean APFI.

Numerical modeling

Because we experimentally measured characteristics of APs in populations of single cells that differed by autonomic input status, we were able to glean insights not only into APFI variability in an "average" cell, but also into populations of cells isolated from SAN tissue. Embracing SAN function at the cell population level resonates with recent studies of SAN function at the tissue level (Bychkov et al., 2020) (Fenske et al., 2020) that have revealed a novel understanding of the SAN impulse as an emergent property created by a collective of variable interactions among heterogeneous cell populations within the SAN tissue. On the other end, APFI variability per se, also emerges at smaller, subcellular scales, due to variability in the functions of individual molecules, such as ion channels, transporters, and pumps, individually and in complex cooperation with each other. These molecular functions cannot be measured directly in our single cell experiments. Thus, we employed numerical modeling to extend our perspectives from cell populations and single cell levels downwards to the molecular scale. Such broader consideration of variability makes sense when we approach pacemaker function as a multiscale phenomenon (Qu, Garfinkel, Weiss, & Nivala, 2011) featuring free scale and fractal-like characteristics (Weiss & Qu, 2020). Considering the SAN cell as a stochastic dynamical system, we examined variability of major ion currents and submembrane $[Ca^{2+}]$ during different autonomic states that created a broad range of AP firing intervals, that were measured experimentally.

Our simulations indicate that the APFI variability of some ion currents and sub-membrane Ca^{2+} can be close to that of the APFI itself, but also can be substantially lower or higher than the

APFI variability, depending on the presence or absence of autonomic receptor stimulation, and the time during the AP cycle (Figure 12): components such as I_f, I_{NCX}, I_{CaT} and Ca^{2+} exhibit substantial cycle to cycle variability, whereas I_{CaL}, I_{KACh}, and I_{Kr} show less, or moderate variability. This behavior reflects complex non-linear recursive interactions of V_m and Ca^{2+} that couple the clocks that drive the system (Lyashkov et al., 2018) and, as such, cannot be directly and definitively interpreted in cause-effect terms. Nevertheless, our simulations confirmed ion channel behavior that could have been envisioned. For example, independent of the nature of the noise added (to I_{tot} or to Ca^{2+} release flux): components contributing to DD (I_f, I_{NCX}, and I_{CaT}) exhibit larger cycle to cycle variability, whereas components contributing to generation of all-or-none AP characteristics exhibit less variability (peak I_{CaL} and I_{Kr}). This result is in accord with the idea of order/disorder transitions during AP cycle (Figure 2, order/disorder dash line), i.e., the DD manifests disorder and transition to order, and hence, a larger variability; then following the AP upstroke, the AP itself manifests order and hence, less variability.

But our simulations also provided some unexpected, interesting results: I_{NCX} and sub-membrane Ca^{2+} amplitudes during DD followed a power law relationship over a wide range of APFI under all conditions tested, indicating that self-similar scale-free or fractal-like behavior is operative within the coupled-clock mechanism via I_{NCX} (Figure 13). There is also likely to be a secondary effect of I_{NCX} amplitude itself on DD acceleration in a recursive fashion. Indeed, an increase in I_{NCX} is expected to accelerate DD, but at the same time, is further accelerated by the very same acceleration it imparts to the ignition mechanism (i.e. diastolic I_{CaL} and Ca^{2+}-induced Ca^{2+} release). This self-acceleration ignition mechanism results in the power law behavior predicted by the model.

Peak I_f also followed a power function over a broad range of APFIs, albeit in a noisy manner manifesting some extremely long APFIs in CCh**.** The current that fluctuated most with respect to variabilities in APFI turned out to be I_{CaT}, the peak amplitude of which also reflects DD dynamics. When DD is rapidly accelerating (as it does at shorter cycles) I_{CaT} peak quicky activates and achieves a higher peak amplitude; and vice-versa, when DD is slower at longer APFIs I_{CaT} becomes inactivated over time without achieving a peak. In log-log plots this forms a straight-line for almost the entire range of APFIs (power law behavior), except extremely long APFIs when variations in (already) slow DD dynamics have almost no effect on I_{CaT} peak. Thus, simulations of biophysical components within the pacemaker cell system exhibited a power law behavior over a wide range of APFIs that encompasses the broad range of APFIs measured experimentally.

Observation of model simulations through analytic lenses applied to experimental data indicated that (as was the case for experiment data) the simulated variables are self-similar to each other across broad range of APFI within the three autonomic states (Figure 15, Supplementary Figure S4, Supplementary Table S3).

Model limitations:

Our I_{CaL} model was adopted from the Kurata model (Kurata, Hisatome, Imanishi, & Shibamoto, 2002), and while it does include Ca^{2+}-dependent inactivation mechanism, it lacks I_{CaL} facilitation described in 2000 by Mangoni et al.(Mangoni et al., 2000). Also, our numerical model of I_f lacks dynamic regulation by cAMP (DiFrancesco, 1999), Ca^{2+} activated K^+ channels and store-operated channels that may potentially contribute to APFIV.

Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Author Contributions

EGL and VAM designed the project; DY, AEL, IZ, YY, TMV and BDZ performed experiments and analyzed the results; DY, CHM and ST did the statistical analyses; DY and VAM performed the numerical simulation; DY and EGL wrote the manuscript with the support from YY and VAM. All authors contributed to the article and approved the submitted version.

Funding

This work was supported partially by the Intramural Research Program of the NIH, National Institute on Aging, and by the ISF [No. **330/19**].

Abbreviations:

AP: action potential

APD_{90}: action potential duration from AP overshoot to 90% repolarization

APFI: AP firing interval

APFIV: variability of AP firing intervals

TTIO: time from the previous AP overshoot to the ignition onset when dV/dt=0.15 (V/S)

CaTFI: firing internal of AP-induced Ca^{2+} transient

CaT_{90}: 90% decay time of AP-induced Ca^{2+} transient

LCR: local Ca^{2+} releases

SR: sarcoplasmic reticulum

SD: standard deviation

CV: coefficient of variation

PC: principal component

I_{CaL}: high voltage-activated, L-type Ca^{2+} current

I_f: hyperpolarization-activated funny current

I_{KACh}: acetylcholine-activated K^+ current

I_{Kr}: K^+ current exhibiting strong inward rectification

I_{NCX}: Na^+-Ca^{2+} exchanger (NCX) current

βARs: β-adrenergic receptor stimulation

CRs: Cholinergic receptor stimulation

CCh: carbachol

ISO: isoproterenol

HR: heart rate

SAN: sinoatrial node

Acknowledgements

The authors wish to acknowledge the assistance of Jia-Hua Qu, MD, PhD for assistance with the PC analysis illustrations, and sincerely appreciate Loretta Lakatta, R.N., B.S.N., Robert Monticone, B.S., Tracy Oppel, and Ruth Sadler, B.A. for their editorial assistance. This manuscript has been released as a pre-print at bioRxiv (https://www.biorxiv.org/content/10.1101/2020.09.01.277756v1) (D. Yang et al., 2020).

References

Bak, P. (1999). *How Nature Works: the science of self-organized criticality.* : New York: Springer-Verlag.

Bogdanov, K. Y., Maltsev, V. A., Vinogradova, T. M., Lyashkov, A. E., Spurgeon, H. A., Stern, M. D., & Lakatta, E. G. (2006). Membrane potential fluctuations resulting from submembrane Ca^{2+} releases in rabbit sinoatrial nodal cells impart an exponential phase to the late diastolic depolarization that controls their chronotropic state. *Circ Res, 99*(9), 979-987. doi:10.1161/01.RES.0000247933.66532.0b

Bogdanov, K. Y., Vinogradova, T. M., & Lakatta, E. G. (2001). Sinoatrial nodal cell ryanodine receptor and Na^{+}-Ca^{2+} exchanger: molecular partners in pacemaker regulation. *Circ Res, 88*(12), 1254-1258. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/11420301

Bychkov, R., Juhaszova, M., Tsutsui, K., Coletta, C., Stern, M. D., Maltsev, V. A., & Lakatta, E. G. (2020). Synchronized cardiac impulses emerge from multi-scale, heterogeneous local calcium signals within and among cells of heart pacemaker tissue. *JACC: Clinical Electrophysiology, 6*(8), 907-931.

Clay, J. R., & DeHaan, R. L. (1979). Fluctuations in interbeat interval in rhythmic heart-cell clusters. Role of membrane voltage noise. *Biophys J, 28*(3), 377-389. doi:10.1016/S0006-3495(79)85187-5

DiFrancesco, D. (1999). Dual allosteric modulation of pacemaker (f) channels by cAMP and voltage in rabbit SA node. *J Physiol Pharmacol, 515*, 367-376.

DiFrancesco, D., & Tortora, P. (1991). Direct activation of cardiac pacemaker channels by intracellular cyclic AMP. *Nature, 351*, 145-147.

Fenske, S., Hennis, K., Rotzer, R., Brox, V. F., Becirovic, E., Scharr, A., . . . Wahl-Schott, C. (2020). cAMP-dependent regulation of HCN4 controls the tonic entrainment process in sinoatrial node pacemaker cells. *Nat Commun., 11*(1), 5555. doi:10.1038/s41467-020-19304-9

Garny, A., Noble, D., Hunter, P. J., & Kohl, P. (2009). CELLULAR OPEN RESOURCE (COR): current status and future directions. . *Philos Trans A Math Phys Eng Sci, 367*, 1885-1905. doi:doi: 10.1098/rsta.2008.0289

Ho, D., Imai, K., King, G., & Stuart, E. (2011). MatchIt: Nonparametric Preprocessing for Parametric Causal Inference. *Journal of Statistical Software, 42*(8).

Howell, D. C. (2002). *Statistical Methods for Psychology* (5th ed.). Duxbury, Thomson Learning.

Huikuri, H. V., Makikallio, T. H., Peng, C. K., Goldberger, A. L., Hintze, U., & Moller, M. (2000). Fractal correlation properties of R-R interval dynamics and mortality in patients with

depressed left ventricular function after an acute myocardial infarction. *Circulation, 101*(1), 47-53. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/10618303

Johnson, R. A., & Wichern, D. W. (2008). *Applied Multivariate Statistical Analysis* (6th Edition ed.): Pearson.

Jongsma, H. J., Tsjernina, L., & de Bruijne, J. (1983). The establishment of regular beating in populations of pacemaker heart cells. A study with tissue-cultured rat heart cells. *J Mol Cell Cardiol, 15*(2), 123-133. Retrieved from https://www.ncbi.nlm.nih.gov/pubmed/6854658

Jung, P., Cornell-Bell, A., Madden, K. S., & Moss, F. (1998). Noise-induced spiral waves in astrocyte syncytia show evidence of self-organized criticality. *J. Neurophysiol., 79*, 1098-1102. doi:10.1152/jn.1998.79.2.1098

Kucera, J. P., Heuschkel, M. O., Renaud, P., & Rohr, S. (2000). Power-law behavior of beat-rate variability in monolayer cultures of neonatal rat ventricular myocytes. *Circ Res, 86*(11), 1140-1145. Retrieved from https://www.ncbi.nlm.nih.gov/pubmed/10850965

Kurata, Y., Hisatome, I., Imanishi, S., & Shibamoto, T. (2002). Dynamical description of sinoatrial node pacemaking: improved mathematical model for primary pacemaker cell. *The American journal of physiology, 283*, H2074-2101.

Lakatta, E. G. (2004). Beyond Bowditch: the convergence of cardiac chronotropy and inotropy. *Cell Calcium, 35*(6), 629-642. doi:10.1016/j.ceca.2004.01.017

Lakatta, E. G. (2021). Heartbeat music. *Heart Rhythm, 18*(5), 811-812. doi:10.1016/j.hrthm.2021.01.011

Lakatta, E. G., Maltsev, V. A., Bogdanov, K. Y., Stern, M. D., & Vinogradova, T. M. (2003). Cyclic variation of intracellular calcium: a critical factor for cardiac pacemaker cell dominance. *Circ Res, 92*(3), e45-50. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/12595348

Lakatta, E. G., Maltsev, V. A., & Vinogradova, T. M. (2010). A coupled SYSTEM of intracellular Ca^{2+} clocks and surface membrane voltage clocks controls the timekeeping mechanism of the heart's pacemaker. *Circ Res, 106*(4), 659-673. doi:10.1161/CIRCRESAHA.109.206078

Lakatta, E. G., Vinogradova, T., Lyashkov, A., Sirenko, S., Zhu, W., Ruknudin, A., & Maltsev, V. A. (2006). The integration of spontaneous intracellular Ca2+ cycling and surface membrane ion channel activation entrains normal automaticity in cells of the heart's pacemaker. *Ann N Y Acad Sci, 1080*, 178-206. doi:10.1196/annals.1380.016

Lakatta, E. G., Vinogradova, T. M., & Maltsev, V. A. (2008). The missing link in the mystery of normal automaticity of cardiac pacemaker cells. *Ann N Y Acad Sci, 1123*, 41-57. doi:10.1196/annals.1420.006

Lopez, L., Piegari, E., Sigaut, L., & Dawson, S. P. (2012). Intracellular calcium signals display an avalanche-like behavior over multiple lengthscales. *Front. Physiol., 3*, 350. doi:10.3389/fphys.2012.00350

Lyashkov, A. E., Behar, J., Lakatta, E. G., Yaniv, Y., & Maltsev, V. A. (2018). Positive Feedback Mechanisms among Local Ca Releases, NCX, and ICaL Ignite Pacemaker Action Potentials. *Biophys J, 114*(5), 1176-1189. doi:10.1016/j.bpj.2017.12.043

Lyashkov, A. E., Vinogradova, T. M., Zahanich, I., Li, Y., Younes, A., Nuss, H. B., . . . Lakatta, E. G. (2009). Cholinergic receptor signaling modulates spontaneous firing of sinoatrial nodal cells via integrated effects on PKA-dependent Ca^{2+} cycling and IK_{ACh}. *Am J Physiol Heart Circ Physiol, 297*(3), H949-959. doi:10.1152/ajpheart.01340.2008

Maltsev, A. V., Maltsev, V. A., Mikheev, M., Maltseva, L. A., Sirenko, S. G., Lakatta, E. G., & Stern, M. D. (2011). Synchronization of stochastic Ca2+ release units creates a rhythmic Ca2+ clock in cardiac pacemaker cells. *Biophys J, 100*, 271-283.

Maltsev, A. V., Stern, M. D., & Maltsev, V. A. (2019). Mechanisms of Calcium Leak from Cardiac Sarcoplasmic Reticulum Revealed by Statistical Mechanics. *Biophys J, 116*(11), 2212-2223. doi:10.1016/j.bpj.2019.04.031

Maltsev, V. A., & Lakatta, E. G. (2008). Dynamic interactions of an intracellular Ca2+ clock and membrane ion channel clock underlie robust initiation and regulation of cardiac pacemaker function. *Cardiovasc Res, 77*(2), 274-284. doi:10.1093/cvr/cvm058

Maltsev, V. A., & Lakatta, E. G. (2009). Synergism of coupled subsarcolemmal Ca^{2+} clocks and sarcolemmal voltage clocks confers robust and flexible pacemaker function in a novel pacemaker cell model. *Am J Physiol Heart Circ Physiol, 296*(3), H594-615. doi:10.1152/ajpheart.01118.2008

Maltsev, V. A., & Lakatta, E. G. (2010). A novel quantitative explanation for the autonomic modulation of cardiac pacemaker cell automaticity via a dynamic system of sarcolemmal and intracellular proteins. *Am J Physiol Heart Circ Physiol, 298*(6), H2010-2023. doi:10.1152/ajpheart.00783.2009

Mangoni, M. E., Fontanaud, P., Noble, P. J., Noble, D., Benkemoun, H., Nargeot, J., & Richard, S. (2000). Facilitation of the L-type calcium current in rabbit sino-atrial cells: effect on cardiac automaticity. *Cardiovasc Res, 48*, 375-376.

Monfredi, O., Lyashkov, A. E., Johnsen, A. B., Inada, S., Schneider, H., Wang, R., . . . Boyett, M. R. (2014). Biophysical characterization of the underappreciated and important relationship between heart rate variability and heart rate. *Hypertension, 64*(6), 1334-1343. doi:10.1161/HYPERTENSIONAHA.114.03782

Monfredi, O., Maltseva, L. A., Spurgeon, H. A., Boyett, M. R., Lakatta, E. G., & Maltsev, V. A. (2013). Beat-to-Beat Variation in Periodicity of Local Calcium Releases Contributes to Intrinsic Variations of Spontaneous Cycle Length in Isolated Single Sinoatrial Node Cells. *PLoS One, 8*(6), e67247. doi:10.1371/journal.pone.0067247

Nivala, M., Ko, C. Y., Nivala, M., Weiss, J. N., & Qu, Z. (2012). Criticality in intracellular calcium signaling in cardiac myocytes. *Biophys J, 102*(11), 2433-2442. doi:10.1016/j.bpj.2012.05.001

Nolasco, J. B., & Dahlen, R. W. (1968). A graphic method for the study of alternation in cardiac action potentials. *J Appl Physiol, 25*(2), 191-196. doi:10.1152/jappl.1968.25.2.191

Qu, Z., Garfinkel, A., Weiss, J. N., & Nivala, M. (2011). Multi-scale modeling in biology: how to bridge the gaps between scales? *Progress in biophysics and molecular biology, 107*, 21-31.

Shivkumar, K., Ajijola, O. A., Anand, I., Armour, J. A., Chen, P. S., Esler, M., . . . Zipes, D. P. (2016). Clinical neurocardiology defining the value of neuroscience-based cardiovascular therapeutics. *J. Physiol., 594*(14), 3911-3954. doi:doi: 10.1113/JP271870

Silverman, B. W. (1986). *Density Estimation for Statistics and Data Analysis*: London: Chapman & Hall/CRC.

Sirenko, S., Maltsev, V. A., Yaniv, Y., Bychkov, R., Yaeger, D., Vinogradova, T. M., . . . Lakatta, E. G. (2016). Electrochemical Na+ and Ca2+ Gradients Drive Coupled-Clock Regulation of Automaticity of Isolated Rabbit Sinoatrial Nodal Pacemaker Cells. *Am J Physiol Heart Circ Physiol*, ajpheart 00667 02015. doi:10.1152/ajpheart.00667.2015

Sirenko, S., Yang, D., Li, Y., Lyashkov, A. E., Lukyanenko, Y. O., Lakatta, E. G., & Vinogradova, T. M. (2013). Ca^{2+}-dependent phosphorylation of Ca^{2+} cycling proteins generates robust rhythmic local Ca^{2+} releases in cardiac pacemaker cells. *Sci Signal, 6*(260), ra6. doi:10.1126/scisignal.2003391

Sirenko, S. T., Tsutsui, T., Tarasov, K. V., Yang, D., Wirth, A. N., Maltsev, V. A., . . . Lakatta., E. G. (2021). Self-Similar Synchronization of Calcium and Membrane Potential Transitions During Action Potential Cycles Predict Heart Rate Across Species. *JACC Clin Electrophysiol, S2405-500X(21)00200-0*. doi:10.1016/j.jacep.2021.02.016

Song, L. S., Sham, J. S., Stern, M. D., Lakatta, E. G., & Cheng, H. (1998). Direct measurement of SR release flux by tracking 'Ca^{2+} spikes' in rat cardiac myocytes. *J Physiol, 512(Pt 3)*, 677-691. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/9769413

Stožer, A., Markovic, R., Dolensek, J., Perc, M., Marhl, M., Rupnik, M. S., & Gosak, M. (2019). Heterogeneity and Delayed Activation as Hallmarks of Self-Organization and Criticality in Excitable Tissue. *Front Physiol, 10*, 869.

Vinogradova, T. M., Bogdanov, K. Y., & Lakatta, E. G. (2002). beta-Adrenergic stimulation modulates ryanodine receptor Ca^{2+} release during diastolic depolarization to accelerate pacemaker activity in rabbit sinoatrial nodal cells. *Circ Res, 90*(1), 73-79. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/11786521

Vinogradova, T. M., Zhou, Y. Y., Maltsev, V., Lyashkov, A., Stern, M., & Lakatta, E. G. (2004). Rhythmic ryanodine receptor Ca^{2+} releases during diastolic depolarization of sinoatrial pacemaker cells do not require membrane depolarization. *Circ Res, 94*(6), 802-809. doi:10.1161/01.RES.0000122045.55331.0F

Vinogradova TM, Z. Y., Bogdanov KY, Yang D, Kuschel M, Cheng H and Xiao RP. (2000). Sinoatrial Node Pacemaker Activity Requires Ca2+ / Calmodulin-Dependent Protein Kinase II Activation. *Circ Res, 87*, 760-767.

Wang, S. Q., Song, L. S., Lakatta, E. G., & Cheng, H. (2001). Ca^{2+} signalling between single L-type Ca^{2+} channels and ryanodine receptors in heart cells. *Nature, 410*(6828), 592-596. doi:10.1038/35069083

Weiss, J. N., & Qu, Z. (2020). The Sinus Node: Still Mysterious After All These Years. *JACC Clin Electrophysiol, 6*(14), 1841-1843.

Yang, D., Lyashkov, A. E., Li, Y., Ziman, B. D., & Lakatta, E. G. (2012). RGS2 overexpression or G_i inhibition rescues the impaired PKA signaling and slow AP firing of cultured adult rabbit pacemaker cells. *J Mol Cell Cardiol, 53*(5), 687-694. doi:10.1016/j.yjmcc.2012.08.007

Yang, D., Lyashkov, A. E., Morrell, C. H., Zahanich, I., Yaniv, Y., Vinogradova, T. M., . . . Lakatta, E., G. (2020). Self-similar action potential cycle-to-cycle variability of Ca2+ and current oscillators in cardiac pacemaker cells. *bioRxiv*. doi:https://doi.org/10.1101/2020.09.01.277756

Yaniv, Y., Ahmet, I., Liu, J., Lyashkov, A. E., Guiriba, T. R., Okamoto, Y., . . . Lakatta, E. G. (2014). Synchronization of sinoatrial node pacemaker cell clocks and its autonomic modulation impart complexity to heart beating intervals. *Heart Rhythm, 11*(7), 1210-1219. doi:10.1016/j.hrthm.2014.03.049

Yaniv, Y., Ganesan, A., Yang, D., Ziman, B. D., Lyashkov, A. E., Levchenko, A., . . . Lakatta, E. G. (2015). Real-time relationship between PKA biochemical signal network dynamics and increased action potential firing rate in heart pacemaker cells: Kinetics of PKA activation

in heart pacemaker cells. *J Mol Cell Cardiol, 86*, 168-178. doi:10.1016/j.yjmcc.2015.07.024

Yaniv, Y., Lyashkov, A. E., & Lakatta, E. G. (2013). The fractal-like complexity of heart rate variability beyond neurotransmitters and autonomic receptors: signaling intrinsic to sinoatrial node pacemaker cells. *Cardiovasc Pharm Open Access, 2*. Retrieved from https://www.ncbi.nlm.nih.gov/pubmed/26709383

Yaniv, Y., Lyashkov, A. E., Sirenko, S., Okamoto, Y., Guiriba, T. R., Ziman, B. D., . . . Lakatta, E. G. (2014). Stochasticity intrinsic to coupled-clock mechanisms underlies beat-to-beat variability of spontaneous action potential firing in sinoatrial node pacemaker cells. *J Mol Cell Cardiol, 77*, 1-10. doi:10.1016/j.yjmcc.2014.09.008

Zaza, A., & Lombardi, F. (2001). Autonomic indexes based on the analysis of heart rate variability: a view from the sinus node. *Cardiovasc Res, 50*(3), 434-442. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/11376619

Zaza, A., Robinson, R. B., & Difrancesco, D. (1996). Basal responses of the L-type Ca2+ and hyperpolarization-activated currents to autonomic agonists in the rabbit sino-atrial node. . *J Physiol Pharmacol, 491*, 347-355.

Zhou, P., Zhao, Y. T., Guo, Y. B., Xu, S. M., Bai, S. H., Lakatta, E. G., . . . Wang, S. Q. (2009). Beta-adrenergic signaling accelerates and synchronizes cardiac ryanodine receptor response to a single L-type Ca^{2+} channel. *Proc Natl Acad Sci U S A, 106*(42), 18028-18033. doi:10.1073/pnas.0906560106

Figure legends

Figure 1. Simultaneous recording of AP and Ca^{2+} in a 10 beat time series of APs (A) and the measured APFI and CaTFI (B); the definition of parameters measured in Vm and Ca^{2+} domains (C), and the self-similarity of the Vm and Ca^{2+} parameters during APs to each other and to the APFI (D).

Figure 2A illustrates the time series of AP intervals in Figure 1A plotted as phase-plane diagrams in which Vm is depicted as a function of Ca^{2+} throughout each cycle. The times at which various channels are activated throughout the Vm/Ca phase-plane loop are indicated. The times of ignition and 90% recovery are also indicated (B). The point resolution is 3.072 ms, and the spread indicates the rates at which the electrochemical signal changes during each cycle. The dashed line in the figure marks the border between the disordered and ordered molecular activation. The arrows indicate the direction of the electrochemical signal emergence.

Figure 3. Poincare plots (A) and fitting of the Poincare plot Ellipse clouds (B), the relation of sd2s to sd1s (C) of the 6 parameters from simultaneously recorded during the 10 beats time series from Figure 1.

Figure 4. Self-control AP recordings in 2 cells during CCh or ISO (A), Poincare plots of the 3 measured Vm domain parameters in 3 autonomic states: CCh, control (2 cells) and ISO (B, number of beats in each time series was 197, 397, 397 and 900, respectively, in carbochol, carbochol control, isoproterenol control and isoproterenol). The inset shows an example of Ellipse fitting.

Figure 5. Correlation between Mean and SD of the 3 Vm parameters (A), correlation of Poincare Ellipse fitting sd1 and sd2 (B), and the self-similarity of APD90 and TTIO to APFI (C) across the 3 autonomic states, the same as in figure 4.

Figure 6. Density distributions of the self-control Vm parameters measured in the same cell as in figure 4 and 5 (2 control cells are combined). The density distributions are presented as nonparametric kernel estimates of probability density functions (Silverman, 1986), scaled so that the total area within each curve is unity.

Figure 7. Density distributions of selected parameter means (B) and SDs (A) of M and Ca^{2+} clock functions measured in different cells prior to and during autonomic receptor stimulation across the 3 groups of mean APFI steady states in Table 1. The density distributions are presented as nonparametric kernel estimates of probability density functions (Silverman, 1986), scaled so that the total area within each curve is unity. Both the mean and variability about the mean are concordant with each other across the 3 experimental groups in control and shift concordantly in response to autonomic receptor stimulation in all cells measured.

Figure 8. A. Plot of mean of Means and SDs of Vm vs. Ca^{2+} from Table 1; B. Mean of means in Table 1 vs. mean of the SDs in Table 1; C. Means and SDs of V_m parameters vs. Means and SDs of corresponding Ca^{2+} parameters for all 230 cells; D. Means and SDs of Ignition and Recovery vs. Means and SDs of Cycle Length for all 230 cells (Note: In Panel A, M.M = mean of means; M.SD = mean of SDs. CL= cycle length, Ign = Ignition, and Rec = Recovery).

Figure 9. Distributions of SDs and Means over all groups illustrating self-similarity (Fractal-like behavior). The regression lines are fit as a single piecewise linear model with a join point at the center of the interval with the highest frequency.

Figure 10: Means vs SDs for all cells on a linear scale for A. V_m, B. Ca^{2+}, C. V_m and Ca^{2+}; and using a logarithmic scale for D. V_m, E. Ca, F. V_m and Ca^{2+}.

Figure 11: Plots of Principal Components: A) SDs only; B) Means only; C) Means and SDs.

Figure 12. Results of analysis of coefficient of variation (CV) of major ion currents I_f, I_{Kr}, I_{CaL}, I_{CaT}, I_{KACh}, and Ca^{2+} simulated by Maltsev-Lakatta coupled-clock model with noise current (I_{per}) added to either total current I_{tot} (panels A~ C) or Ca release flux (panels D~F).

Figure 13. I_{NCX}(A), I_{CaT}(B), I_f,C), and $[Ca]_{sub}$ (D) exhibited power law behavior (a linear dependence in the ln-ln plot here) over a wide range of APFI over all conditions tested. Noise (I_{per}) was added to Itot in Maltsev-Lakatta coupled-clock model. Similar dependencies were found when I_{per} was added to Ca release flux (not shown).

Figure 14. The correlation between I_{NCX}(A), I_{CaT}(B), I_f, (C) I_{Kr}(D) or I_{KACh}(E) vs. $[Ca]_{Sub}$ at -40mV. Note that I_{KACh} only presents during CCh.

Figure 15. Poincaré plots of the simulated I_{NCX}, I_{CaT}, I_{CaL}, I_{Kr}, I_{KACh}, I_f, $[Ca]_{Sub}$ at -40mV, APFI and TTIO from 1 model cell during ISO, CCh or at basal condition in ln-ln plot (A) and the correlation between the means and SDs of these parameters (B). Note that I_{KACh} only presents during Carbochol.

Tables

Table 1. A. Mean of SDs and SD of SDs of AP and Ca^{2+} domain intervals among individual cells in each of the 3 steady state populations that differ with respect to autonomic receptor stimulation; **B.** Mean ± SD of means of AP and Ca^{2+} domain interval in each cell population (A).

A. Mean SD ± SD of SDs (ms, individual cells within each population)			
AP recordings	**ISO (n=27)**	**Control (n=78)**	**CCh (n=10)**
APFI, SD	8.43 ± 1.90	13.30 ± 5.18	99.08 ± 57.28
TTIO, SD	13.14 ± 4.54	23.72 ± 13.33	132.78 ± 88.24
APD_{90}, SD	4.52 ± 1.91	9.23 ± 4.56	78.40 ± 58.07
Ca^{2+} recordings	**ISO (n=27)**	**control (n=78)**	**CCh (n=10)**
CaTFI, SD	17.52 ± 14.30	16.23 ± 11.24	86.08 ± 54.28
CaT_{90}, SD	17.31 ± 12.34	13.17 ± 7.81	30.79 ± 13.35
LCR period, SD	38.11 ± 24.77	40.34 ± 23.90	153.31 ± 93.59
B. Mean ± SD of Means (ms)			
AP recordings	**ISO (n=27)**	**Control (n=78)**	**CCh (n=10)**
APFI, Mean	244.87 ± 22.13	321.57 ± 63.08	786.45 ± 374.35
TTIO, Mean	188.01 ± 18.69	239.45 ± 60.45	690.44 ± 334.27
APD_{90}, Mean	123.55 ± 14.06	169.06 ± 35.37	249.72 ± 81.38
Ca^{2+} recordings	**ISO (n=27)**	**control (n=78)**	**CCh (n=10)**
CaTFI, Mean	357.80 ± 64.94	390.64 ± 83.37	872.84 ± 284.33
CaT_{90}, Mean	160.76 ± 43.49	174.73 ± 42.73	289.94 ± 70.24
LCR period, Mean	302.92 ± 55.65	342.68 ± 72.87	734.92 ± 191.01

Table 2. Correlation matrices of Vm or Ca^{2+} parameters measured in different cells within 3 different autonomic states (summary data listed in Table 1).

			Action Potential (V_m) (n = 115)						Ca^{2+} (n = 115)					
			Means			SDs			Means			SDs		
			APFI	TTIO	APD90	APFI	TTIO	APD90	CaTFI	CaT90	LCR Period	CaTFI	CaT90	LCR Period
Action Potential (V_m)	Means	APFI: r	1											
		APFI: p												
		TTIO: r	0.9817	1										
		TTIO: p	2.7E-83											
		APD90: r	0.8550	0.7879	1									
		APD90: p	5.3E-34	1.5E-25										
	SDs	APFI: r	0.8947	0.8954	0.6540	1								
		APFI: p	2.3E-41	1.6E-41	2.2E-15									
		TTIO: r	0.9139	0.8777	0.7132	0.9284	1							
		TTIO: p	4.7E-46	6.8E-38	3.8E-19	2.2E-50								
		APD90: r	0.8164	0.7868	0.6705	0.7742	0.8414	1						
		APD90: p	1.8E-28	3.2E-25	4.4E-16	5.5E-24	1.0E-31							
Ca^{2+}	Means	CaTFI: r	0.9573	0.9373	0.8307	0.8408	0.8702	0.8375	1					
		CaTFI: p	9.9E-63	1.4E-53	1.6E-30	6.8E-32	1.6E-36	3.6E-31						
		CaT90: r	0.6574	0.6797	0.5982	0.5532	0.4897	0.5245	0.7017	1				
		CaT90: p	1.3E-15	6.7E-17	1.7E-12	1.4E-10	2.8E-08	2.1E-09	2.4E-18					
		LCR Period: r	0.9380	0.9162	0.8342	0.8257	0.8442	0.8137	0.9792	0.6962	1			
		LCR Period: p	7.7E-54	1.1E-46	5.5E-31	7.2E-30	2.2E-32	3.7E-28	3.7E-80	5.8E-18				
	SDs	CaTFI: r	0.6349	0.6170	0.5236	0.7077	0.6678	0.6416	0.7064	0.4362	0.7200	1		
		CaTFI: p	4.3E-14	3.4E-13	2.7E-09	1.9E-18	6.5E-16	2.5E-14	2.3E-18	1.4E-06	2.6E-19			
		CaT90: r	0.3194	0.3367	0.2951	0.3146	0.2598	0.3078	0.4448	0.5409	0.4531	0.6040	1	
		CaT90: p	0.0006	0.0003	0.0015	0.0007	0.0055	0.0010	8.0E-07	6.2E-10	4.7E-07	2.3E-12		
		LCR Period: r	0.7174	0.7321	0.6283	0.6234	0.5874	0.6696	0.7665	0.6262	0.7282	0.6704	0.4619	1
		LCR Period: p	1.9E-19	1.5E-20	5.6E-14	1.0E-13	5.2E-12	4.4E-16	1.8E-23	7.2E-14	2.9E-20	4.6E-16	2.6E-07	

Figure 1.

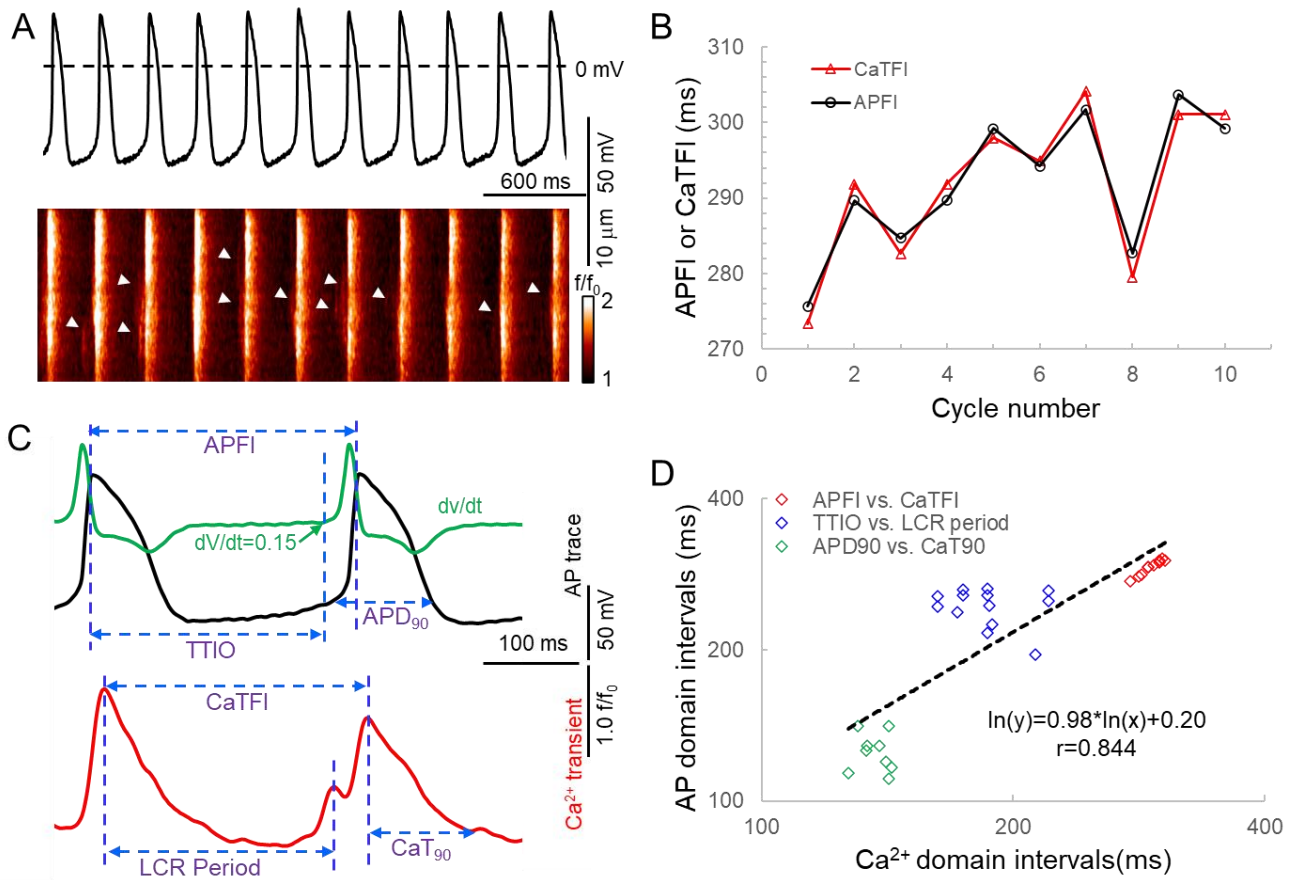

A
0 mV
600 ms
50 mV
10 µm
f/f0
2
1
B
CaTFI
APFI
APFI or CaTFI (ms)
Cycle number
C
APFI
dv/dt
dV/dt=0.15
AP trace
APD90
TTIO
100 ms
50 mV
CaTFI
Ca2+ transient
1.0 f/f0
LCR Period
CaT90
D
APFI vs. CaTFI
TTIO vs. LCR period
APD90 vs. CaT90
ln(y)=0.98*ln(x)+0.20
r=0.844
AP domain intervals (ms)
Ca2+ domain intervals(ms)

Figure 2.

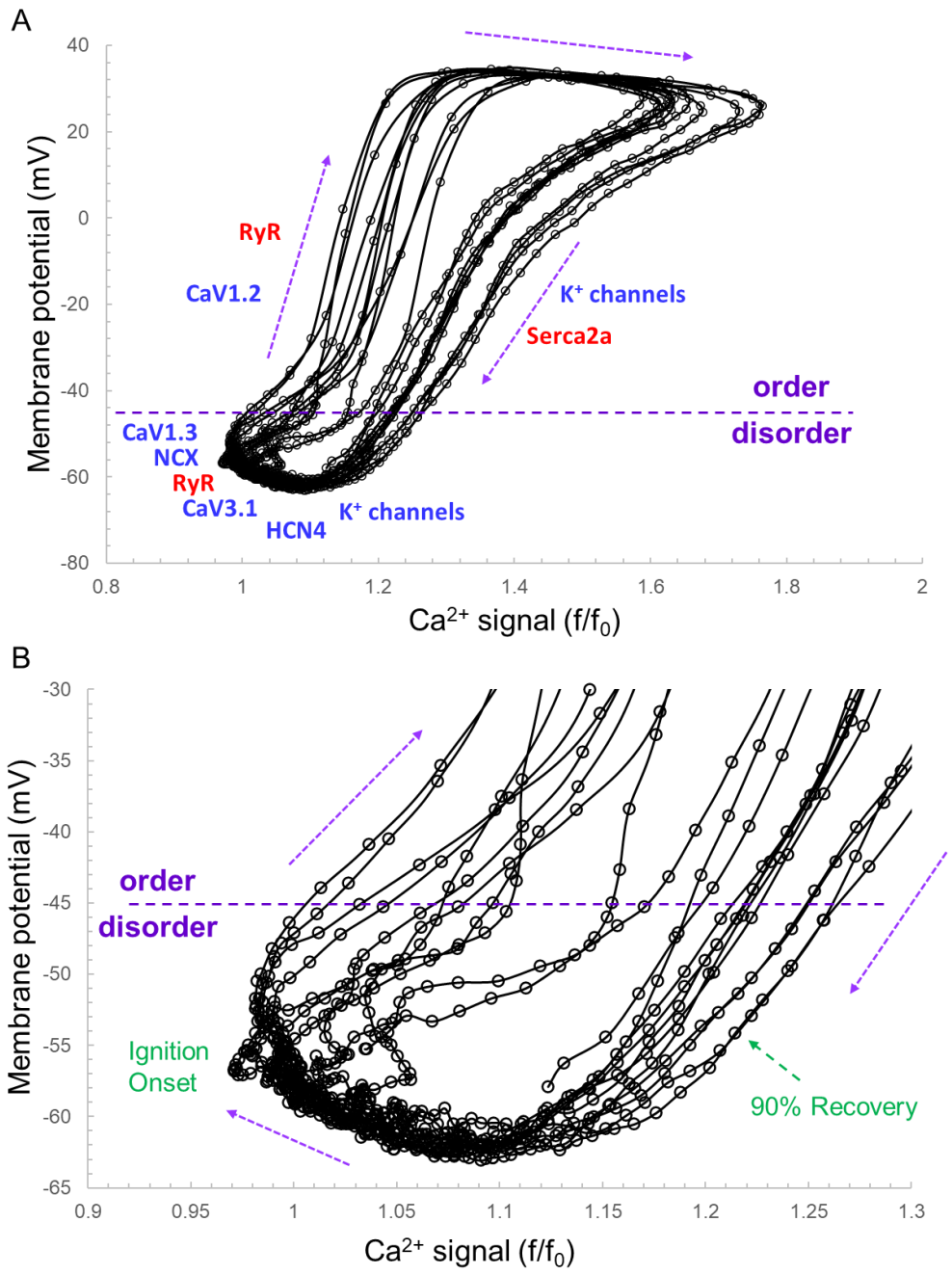

A
Membrane potential (mV)
Ca2+ signal (f/f0)
RyR
CaV1.2
K+ channels
Serca2a
order
disorder
CaV1.3
NCX
RyR
CaV3.1
HCN4
K+ channels
B
Membrane potential (mV)
Ca2+ signal (f/f0)
order
disorder
Ignition
Onset
90% Recovery

Figure 3.

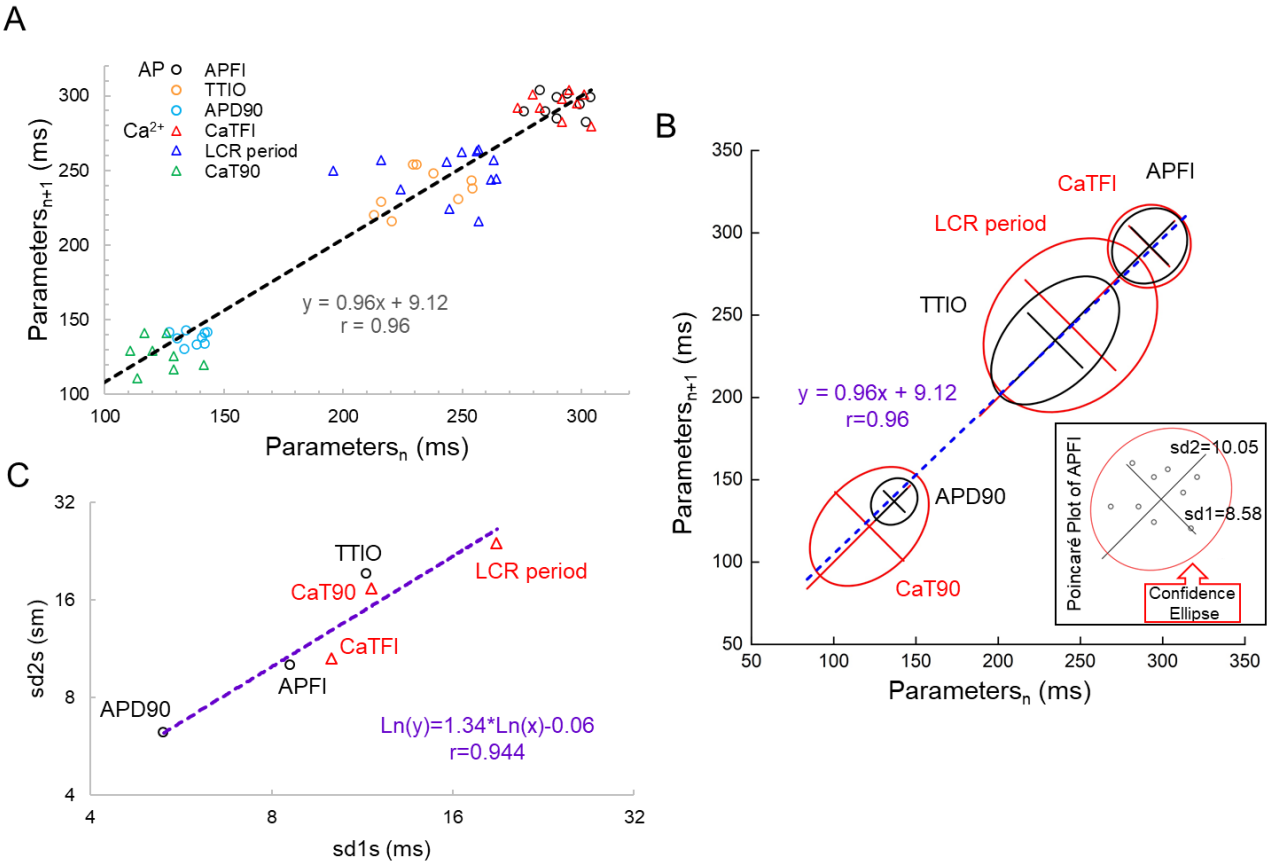

A
AP
APFI
TTIO
APD90
Ca2+
CaTFI
LCR period
CaT90
y = 0.96x + 9.12
r = 0.96
Parameters_n+1 (ms)
Parameters_n (ms)
B
CaTFI
APFI
LCR period
TTIO
y = 0.96x + 9.12
r=0.96
APD90
CaT90
Poincaré Plot of APFI
sd2=10.05
sd1=8.58
Confidence Ellipse
Parameters_n+1 (ms)
Parameters_n (ms)
C
TTIO
CaT90
LCR period
CaTFI
APFI
APD90
Ln(y)=1.34*Ln(x)-0.06
r=0.944
sd2s (sm)
sd1s (ms)

Figure 4.

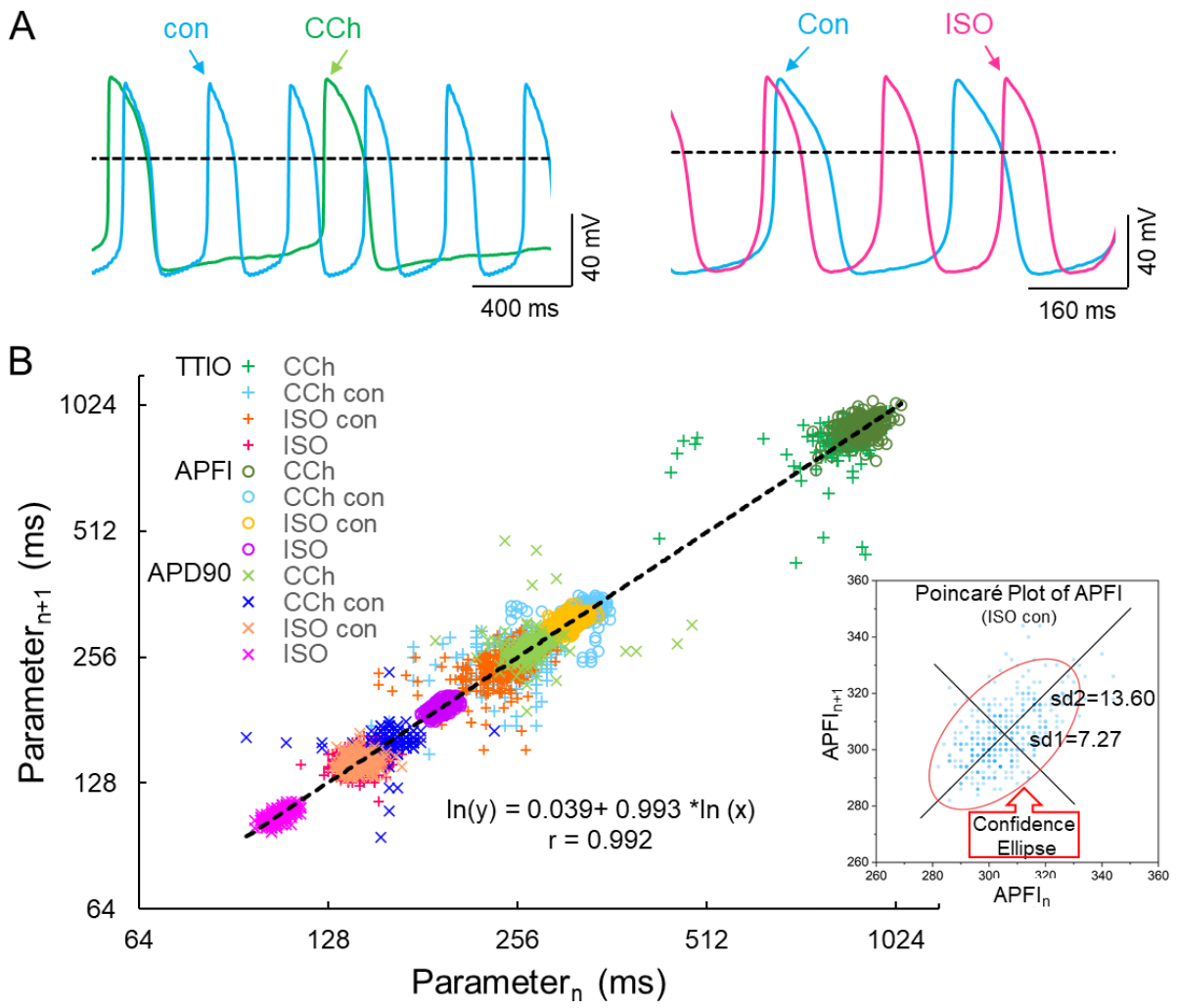

A
con
CCh
40 mV
400 ms
Con
ISO
40 mV
160 ms
B
TTIO
CCh
CCh con
ISO con
ISO
APFI
CCh
CCh con
ISO con
ISO
APD90
CCh
CCh con
ISO con
ISO
ln(y) = 0.039+ 0.993 *ln (x)
r = 0.992
Parameter$_{n+1}$ (ms)
Parameter$_n$ (ms)
64
128
256
512
1024
Poincaré Plot of APFI
(ISO con)
sd2=13.60
sd1=7.27
Confidence Ellipse
APFI$_{n+1}$
APFI$_n$
260
280
300
320
340
360

Figure 5.

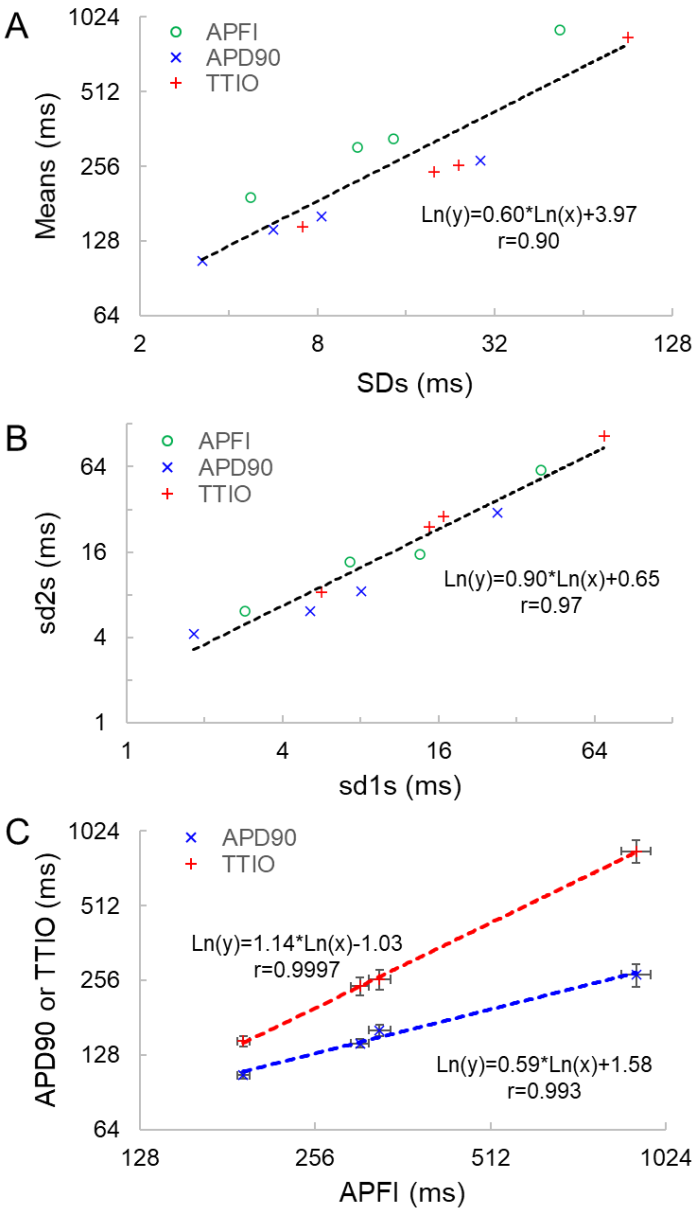

A
1024
512
256
128
64
Means (ms)
APFI
APD90
TTIO
Ln(y)=0.60*Ln(x)+3.97
r=0.90
2
8
32
128
SDs (ms)
B
64
16
4
1
sd2s (ms)
APFI
APD90
TTIO
Ln(y)=0.90*Ln(x)+0.65
r=0.97
1
4
16
64
sd1s (ms)
C
1024
512
256
128
64
APD90 or TTIO (ms)
APD90
TTIO
Ln(y)=1.14*Ln(x)-1.03
r=0.9997
Ln(y)=0.59*Ln(x)+1.58
r=0.993
128
256
512
1024
APFI (ms)

Figure 6.

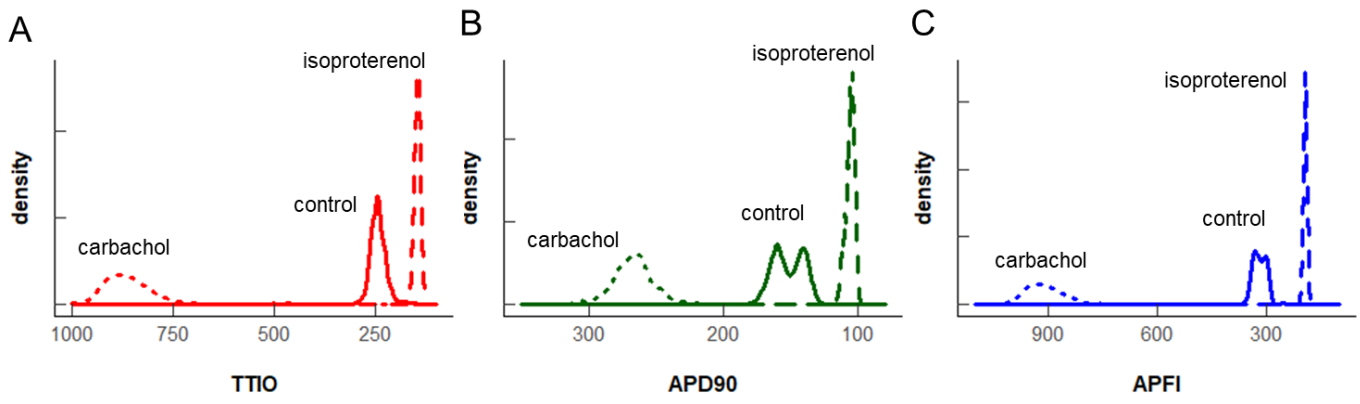

A
isoproterenol
control
carbachol
density
1000
750
500
250
TTIO
B
isoproterenol
control
carbachol
density
300
200
100
APD90
C
isoproterenol
control
carbachol
density
900
600
300
APFI

Figure 7.

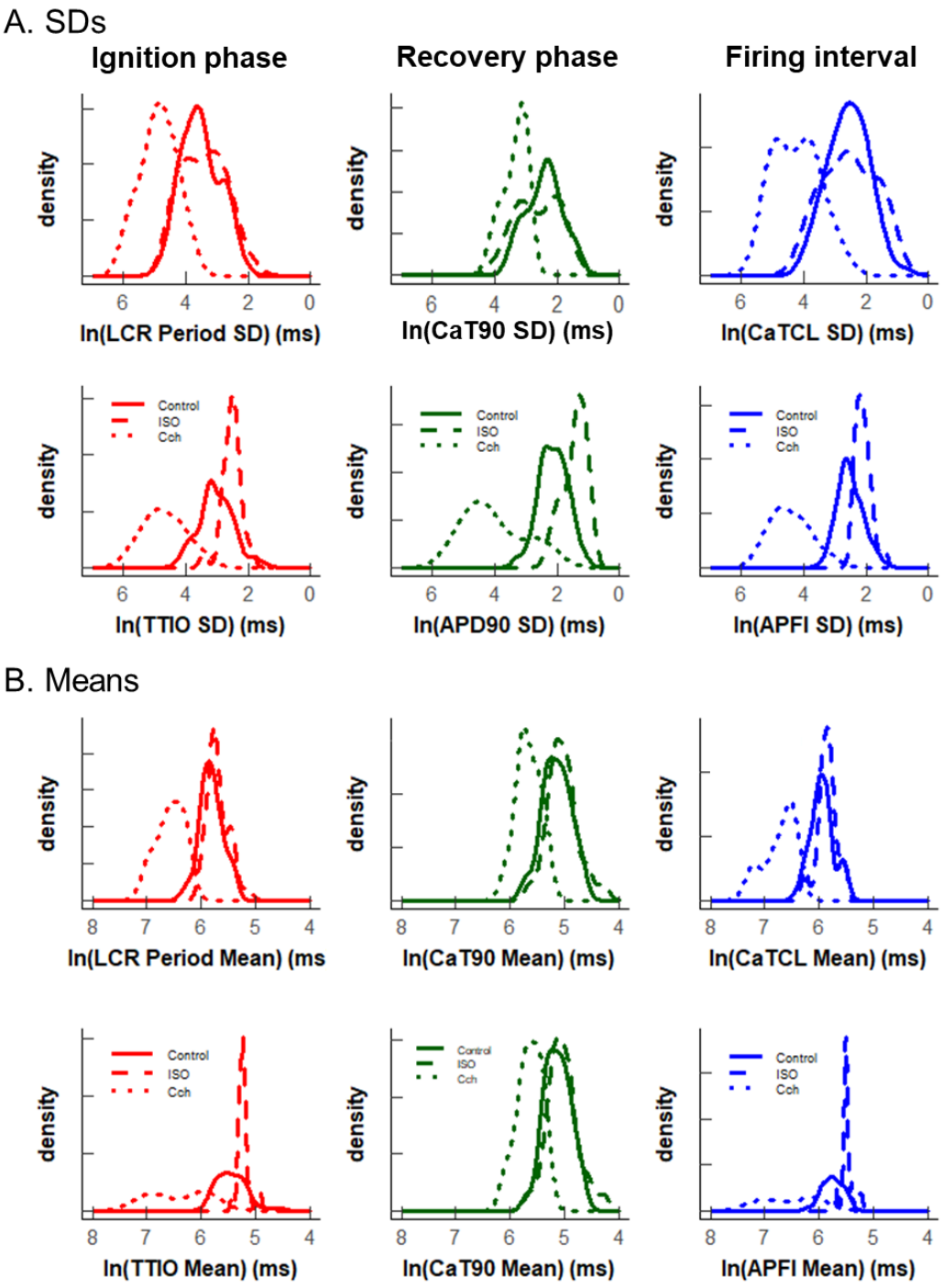

A. SDs
Ignition phase
Recovery phase
Firing interval
density
ln(LCR Period SD) (ms)
ln(CaT90 SD) (ms)
ln(CaTCL SD) (ms)
Control
ISO
Cch
ln(TTIO SD) (ms)
ln(APD90 SD) (ms)
ln(APFI SD) (ms)
B. Means
ln(LCR Period Mean) (ms
ln(CaT90 Mean) (ms)
ln(CaTCL Mean) (ms)
ln(TTIO Mean) (ms)
ln(CaT90 Mean) (ms)
ln(APFI Mean) (ms)

Figure 8.

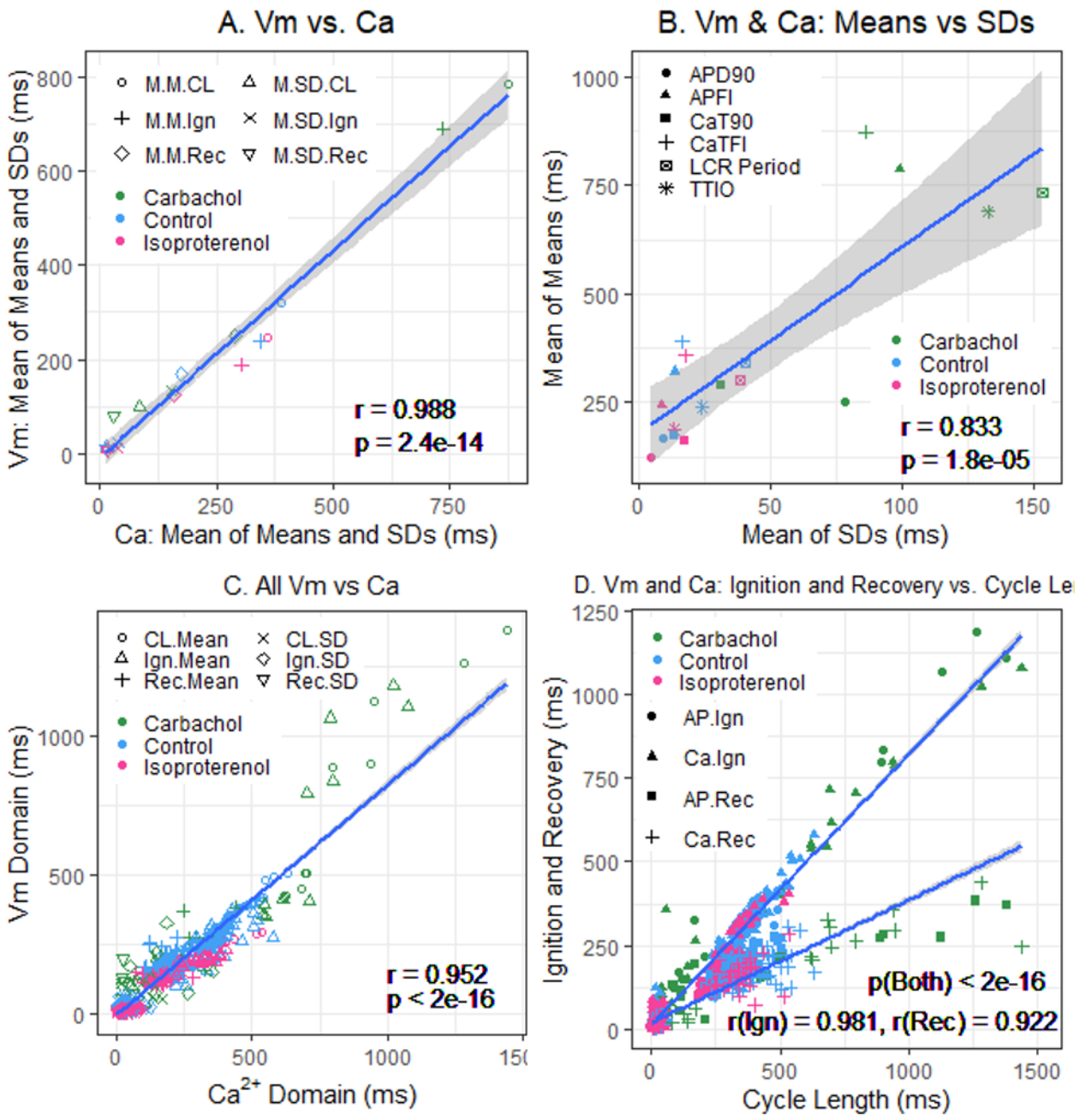

A. Vm vs. Ca
M.M.CL
M.SD.CL
M.M.Ign
M.SD.Ign
M.M.Rec
M.SD.Rec
Carbachol
Control
Isoproterenol
r = 0.988
p = 2.4e-14
Vm: Mean of Means and SDs (ms)
Ca: Mean of Means and SDs (ms)
B. Vm & Ca: Means vs SDs
APD90
APFI
CaT90
CaTFI
LCR Period
TTIO
Carbachol
Control
Isoproterenol
r = 0.833
p = 1.8e-05
Mean of Means (ms)
Mean of SDs (ms)
C. All Vm vs Ca
CL.Mean
CL.SD
Ign.Mean
Ign.SD
Rec.Mean
Rec.SD
Carbachol
Control
Isoproterenol
r = 0.952
p < 2e-16
Vm Domain (ms)
Ca2+ Domain (ms)
D. Vm and Ca: Ignition and Recovery vs. Cycle Le
Carbachol
Control
Isoproterenol
AP.Ign
Ca.Ign
AP.Rec
Ca.Rec
p(Both) < 2e-16
r(Ign) = 0.981, r(Rec) = 0.922
Ignition and Recovery (ms)
Cycle Length (ms)

Figure 9.

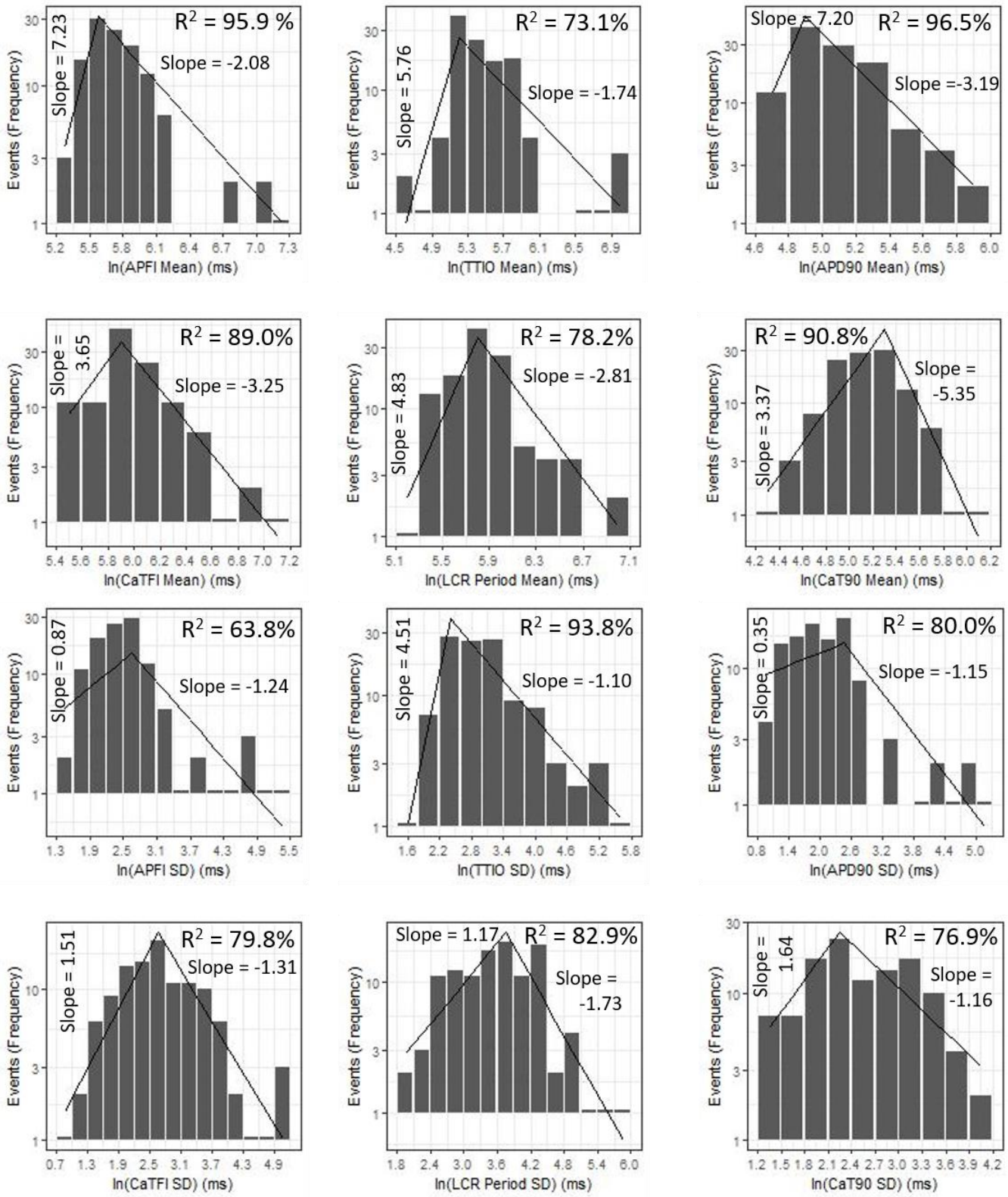

R2 = 95.9 %
Slope = 7.23
Slope = -2.08
Events (Frequency)
ln(APFI Mean) (ms)
R2 = 73.1%
Slope = 5.76
Slope = -1.74
ln(TTIO Mean) (ms)
Slope = 7.20
R2 = 96.5%
Slope =-3.19
ln(APD90 Mean) (ms)
R2 = 89.0%
Slope = 3.65
Slope = -3.25
ln(CaTFI Mean) (ms)
R2 = 78.2%
Slope = 4.83
Slope = -2.81
ln(LCR Period Mean) (ms)
R2 = 90.8%
Slope = 3.37
Slope = -5.35
ln(CaT90 Mean) (ms)
R2 = 63.8%
Slope = 0.87
Slope = -1.24
ln(APFI SD) (ms)
R2 = 93.8%
Slope = 4.51
Slope = -1.10
ln(TTIO SD) (ms)
R2 = 80.0%
Slope = 0.35
Slope = -1.15
ln(APD90 SD) (ms)
R2 = 79.8%
Slope = 1.51
Slope = -1.31
ln(CaTFI SD) (ms)
Slope = 1.17
R2 = 82.9%
Slope = -1.73
ln(LCR Period SD) (ms)
R2 = 76.9%
Slope = 1.64
Slope = -1.16
ln(CaT90 SD) (ms)

Figure 10:

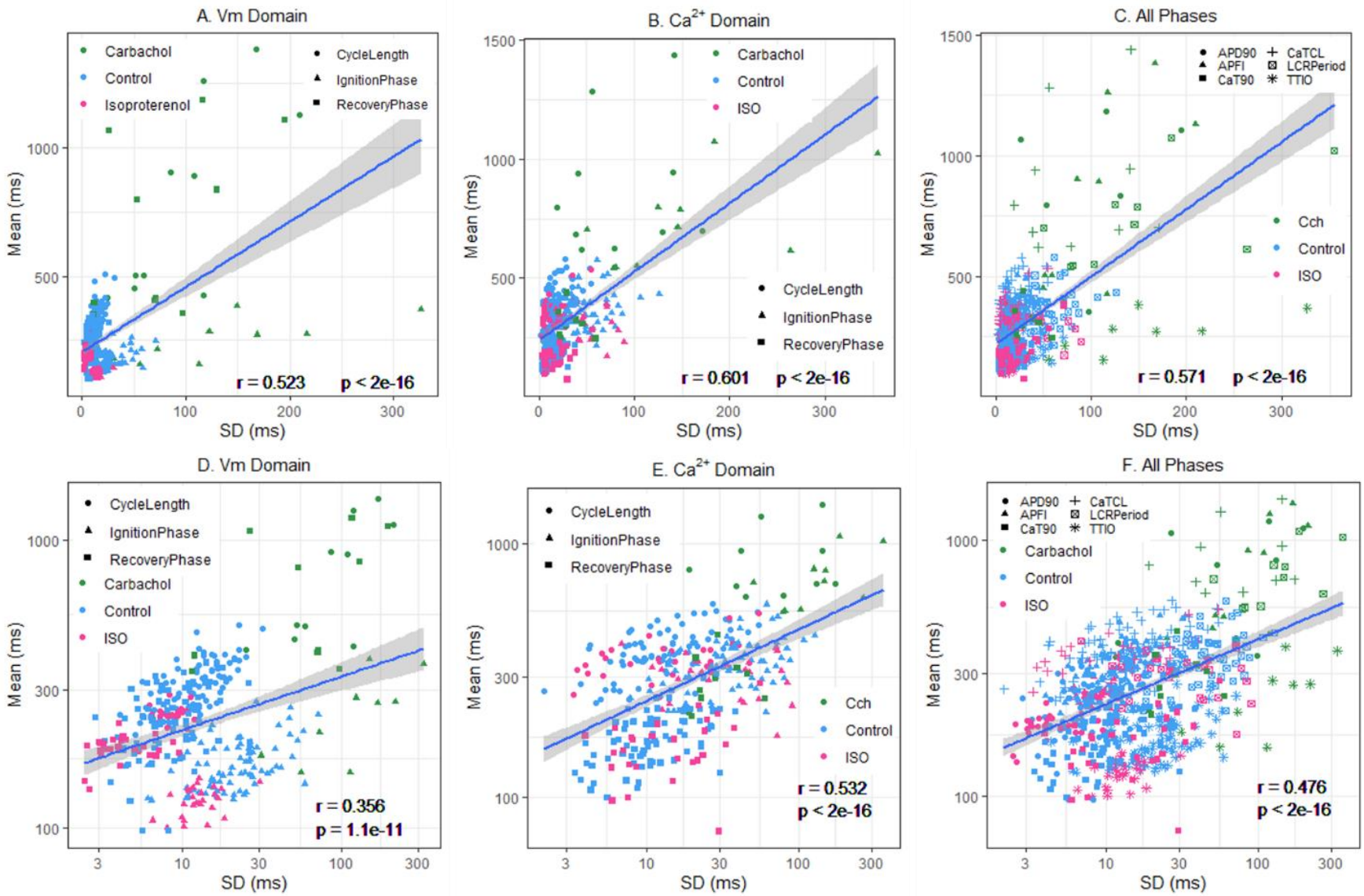

A. Vm Domain
Carbachol
Control
Isoproterenol
CycleLength
IgnitionPhase
RecoveryPhase
r = 0.523
p < 2e-16
B. Ca2+ Domain
Carbachol
Control
ISO
CycleLength
IgnitionPhase
RecoveryPhase
r = 0.601
p < 2e-16
C. All Phases
APD90
APFI
CaT90
CaTCL
LCRPeriod
TTIO
Cch
Control
ISO
r = 0.571
p < 2e-16
D. Vm Domain
CycleLength
IgnitionPhase
RecoveryPhase
Carbachol
Control
ISO
r = 0.356
p = 1.1e-11
E. Ca2+ Domain
CycleLength
IgnitionPhase
RecoveryPhase
Cch
Control
ISO
r = 0.532
p < 2e-16
F. All Phases
APD90
APFI
CaT90
CaTCL
LCRPeriod
TTIO
Carbachol
Control
ISO
r = 0.476
p < 2e-16
Mean (ms)
SD (ms)

Figure 11:

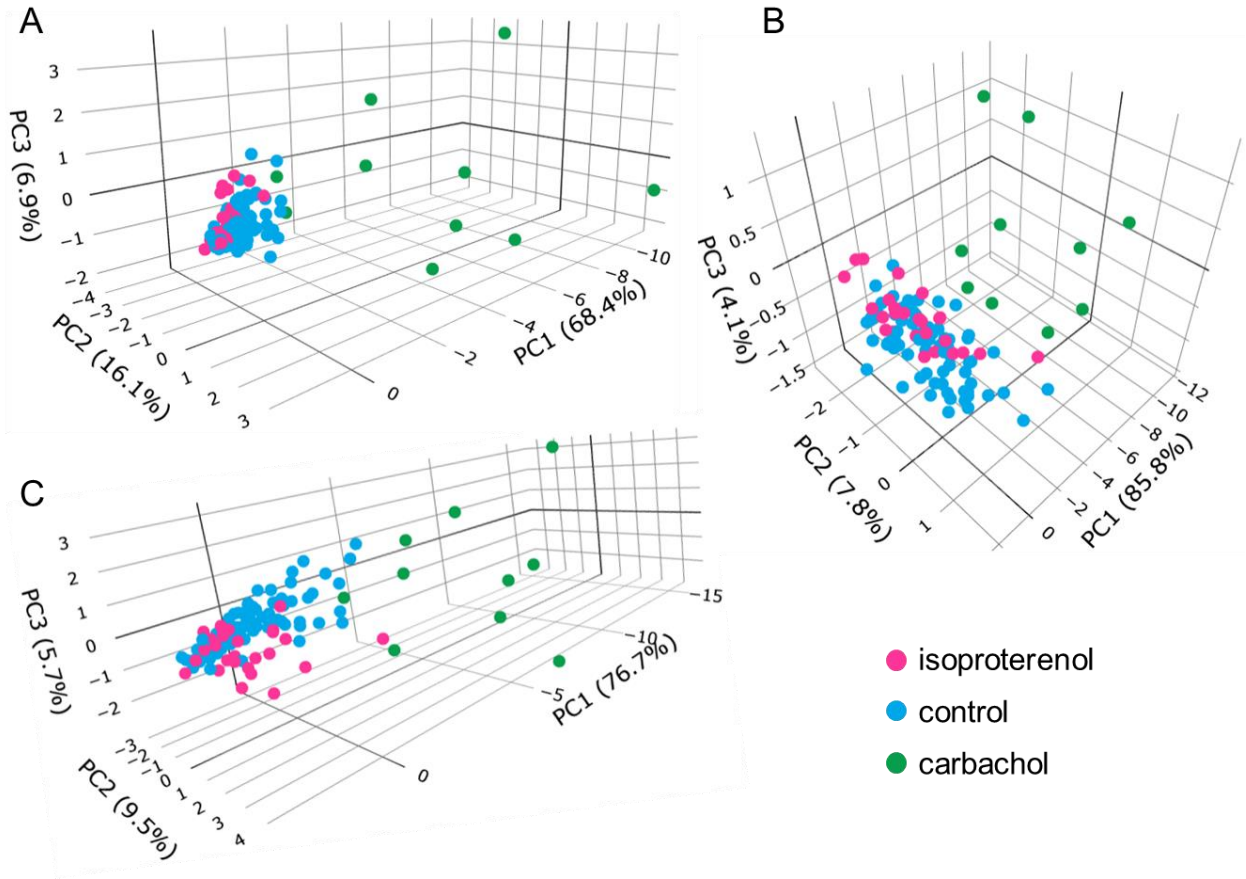

A
PC3 (6.9%)
PC2 (16.1%)
PC1 (68.4%)
B
PC3 (4.1%)
PC2 (7.8%)
PC1 (85.8%)
C
PC3 (5.7%)
PC2 (9.5%)
PC1 (76.7%)
isoproterenol
control
carbachol

Figure 12.

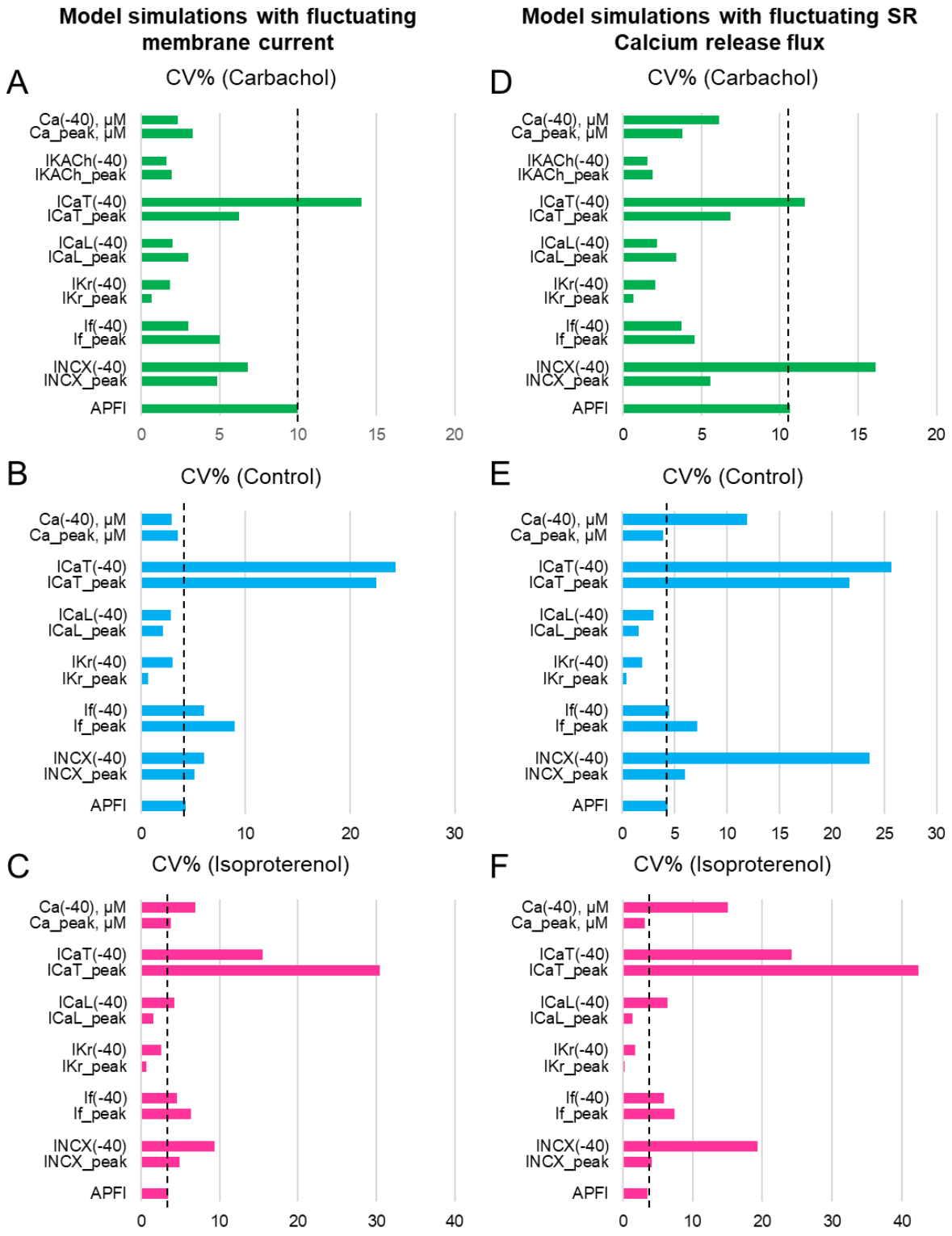

Model simulations with fluctuating membrane current
Model simulations with fluctuating SR Calcium release flux
A
CV% (Carbachol)
B
CV% (Control)
C
CV% (Isoproterenol)
D
CV% (Carbachol)
E
CV% (Control)
F
CV% (Isoproterenol)
Ca(-40), μM
Ca_peak, μM
IKACh(-40)
IKACh_peak
ICaT(-40)
ICaT_peak
ICaL(-40)
ICaL_peak
IKr(-40)
IKr_peak
If(-40)
If_peak
INCX(-40)
INCX_peak
APFI

Figure 13.

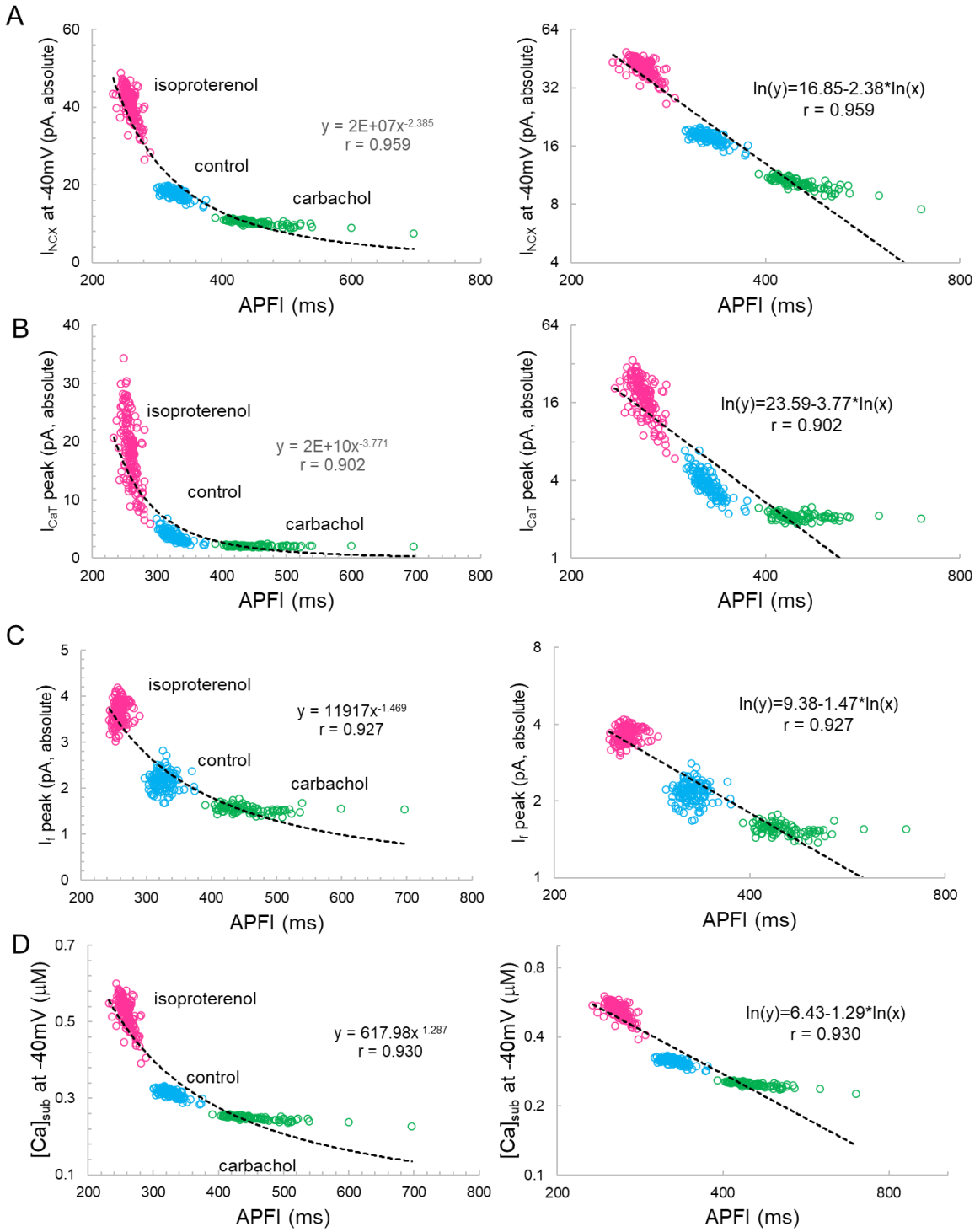

A
I_{NCX} at -40mV (pA, absolute)
isoproterenol
control
carbachol
y = 2E+07x^-2.385
r = 0.959
APFI (ms)
ln(y)=16.85-2.38*ln(x)
r = 0.959
B
I_{CaT} peak (pA, absolute)
y = 2E+10x^-3.771
r = 0.902
ln(y)=23.59-3.77*ln(x)
r = 0.902
C
I_f peak (pA, absolute)
y = 11917x^-1.469
r = 0.927
ln(y)=9.38-1.47*ln(x)
r = 0.927
D
$[Ca]_{sub}$ at -40mV (μM)
y = 617.98x^-1.287
r = 0.930
ln(y)=6.43-1.29*ln(x)
r = 0.930

Figure 14.

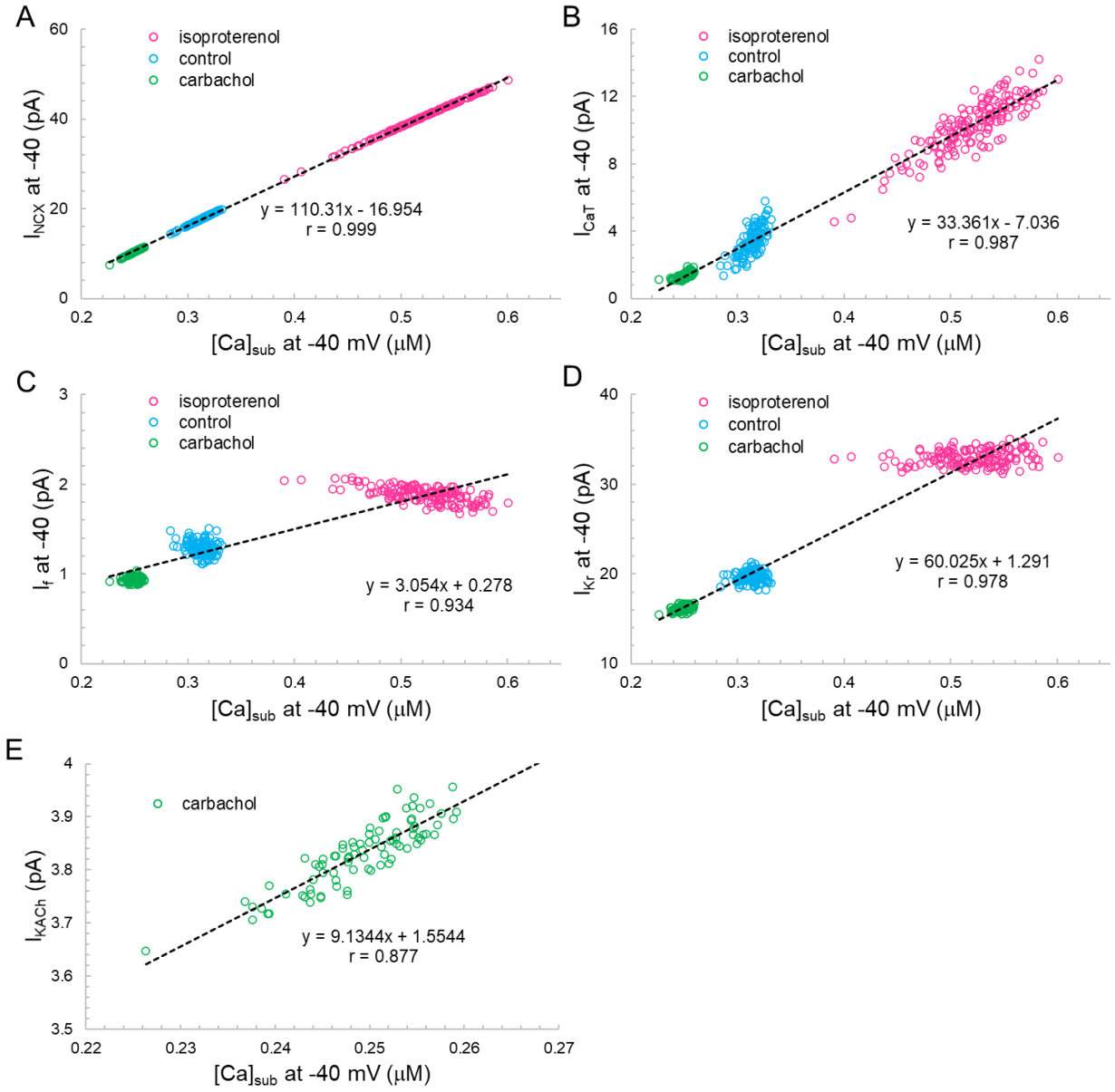
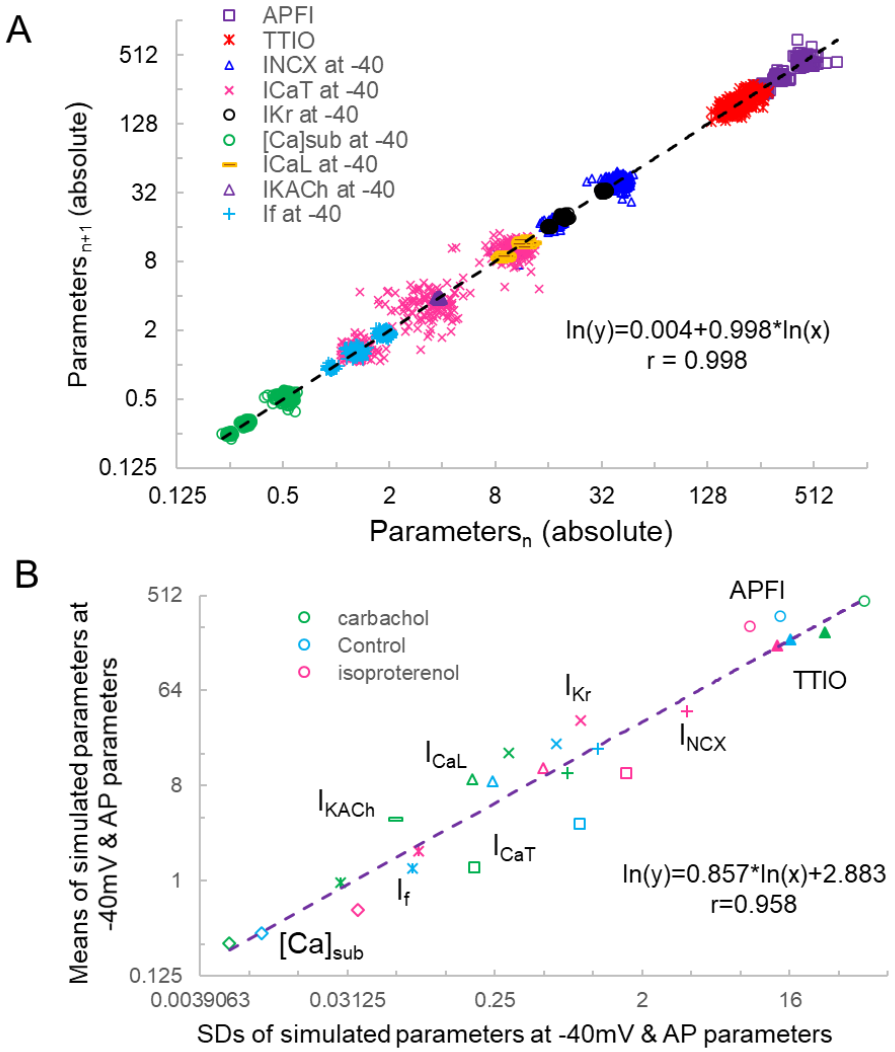

A
isoproterenol
control
carbachol
y = 110.31x - 16.954
r = 0.999
I_{NCX} at -40 (pA)
$[Ca]_{sub}$ at -40 mV (μM)
B
isoproterenol
control
carbachol
y = 33.361x - 7.036
r = 0.987
I_{CaT} at -40 (pA)
$[Ca]_{sub}$ at -40 mV (μM)
C
isoproterenol
control
carbachol
y = 3.054x + 0.278
r = 0.934
I_f at -40 (pA)
$[Ca]_{sub}$ at -40 mV (μM)
D
isoproterenol
control
carbachol
y = 60.025x + 1.291
r = 0.978
I_{Kr} at -40 (pA)
$[Ca]_{sub}$ at -40 mV (μM)
E
carbachol
y = 9.1344x + 1.5544
r = 0.877
I_{KACh} (pA)
$[Ca]_{sub}$ at -40 mV (μM)

Figure 15.


A
APFI
TTIO
INCX at -40
ICaT at -40
IKr at -40
[Ca]sub at -40
ICaL at -40
IKACh at -40
If at -40
512
128
32
8
2
0.5
0.125
Parameters$_{n+1}$ (absolute)
ln(y)=0.004+0.998*ln(x)
r = 0.998
0.125
0.5
2
8
32
128
512
Parameters$_n$ (absolute)
B
carbachol
Control
isoproterenol
APFI
TTIO
I_{Kr}
I_{NCX}
I_{CaL}
I_{KACh}
I_{CaT}
I_f
$[Ca]_{sub}$
512
64
8
1
0.125
Means of simulated parameters at -40mV & AP parameters
ln(y)=0.857*ln(x)+2.883
r=0.958
0.0039063
0.03125
0.25
2
16
SDs of simulated parameters at -40mV & AP parameters