

1 **Deploying a novel tuberculosis molecular bacterial load assay to assess the elimination rate of**
2 ***Mycobacterium tuberculosis* in patients with multidrug-resistant tuberculosis in Tanzania**

3 Peter M. Mbebele^{1,2*}, Emmanuel A. Mpolya², Elingarami Sauli², Bariki Mtafya³, Nyanda E. Ntinginya³,
4 Kennedy K. Addo⁴, Katharina Kreppel², Sayoki Mfinanga⁵, Patrick P.J. Phillips⁶, Stephen H. Gillespie⁷,
5 Scott K. Heysell⁸, Wilber Sabiiti⁷ and Stellah G. Mpagama^{1,2}

6 **Affiliations**

- 7 1. Kibong'oto Infectious Diseases Hospital (KIDH), Siha, Kilimanjaro, Tanzania
- 8 2. Department of Global Health and Biomedical Sciences, School of Life Sciences and
9 Bioengineering, Nelson Mandela African Institution of Science and Technology (NM-
10 AIST), Arusha, Tanzania
- 11 3. National Institute for Medical Research, Mbeya Medical Research Centre, Tanzania
- 12 4. Department of Bacteriology, Noguchi Memorial Institute for Medical Research, University
13 of Ghana, Accra, Ghana
- 14 5. National Institute for Medical Research, Muhimbili Centre, Dar Es Salaam, Tanzania
- 15 6. UCSF Center for Tuberculosis, University of San Francisco, San Francisco, California, USA
- 16 7. School of Medicine, University of St Andrews, Scotland, UK.
- 17 8. Division of Infectious Diseases and International Health, University of Virginia,
18 Charlottesville, Virginia, USA

19 ***Corresponding author,**

20 Dr. Peter Mbebele

21 Kibong'oto Infectious Diseases Hospital (KIDH),

22 P.O BOX 12, Siha, Kilimanjaro, Tanzania

23 Email: mbebelepeter@yahoo.com

24 **Running title:** Monitoring MDR-TB treatment response by TB-MBLA

25 **Abstract**

26 **Background:** Rifampin or multidrug-resistant-tuberculosis (RR/MDR-TB) treatment has transitioned
27 to injectable-free regimens. We tested whether *M. tuberculosis* (*Mtb*) elimination rates measured by
28 molecular bacterial load assay (TB-MBLA) in sputa correlate with composition of the RR/MDR-TB
29 antibiotic regimen.

30 **Methods:** Serial sputa were collected from patients with RR/MDR- and drug-sensitive TB at day 0, 3,
31 7, 14, and then monthly for 4 months of anti-TB treatment. TB-MBLA was used to quantify viable *Mtb*
32 16S rRNA in sputum for estimation of colony-forming-unit per mL (eCFU/mL). *Mtb* elimination rates
33 were compared among regimens using nonlinear-mixed-effects modeling of repeated measures.

34 **Results:** Among 37 patients with a total of 296 serial sputa; 7 patients received
35 rifampin/isoniazid/pyrazinamide/ethambutol (RHZE), 8 an all-oral bedaquiline-based regimen, 9 an
36 injectable and bedaquiline-containing regimen, and 13 an injectable-containing but bedaquiline-free
37 regimen. The overall mean daily *Mtb* elimination was -0.24 [95% Confidence-Interval (CI); -0.39 to -
38 0.08] log₁₀ eCFU/mL, and it varied with treatment-regimen ($p < 0.001$). Compared to the adjusted *Mtb*
39 elimination of -0.17 (95% CI; -0.23 to -0.12) for the injectable-containing but bedaquiline-free reference
40 regimen, the elimination rates were -0.62 (95% CI; -1.05 to -0.20) log₁₀ eCFU/mL for the injectable and
41 bedaquiline-containing regimen ($p = 0.019$), -0.35 (95% CI; -0.65 to -0.13) log₁₀ eCFU/mL for the all-
42 oral bedaquiline-based regimen ($p = 0.054$), and -0.29 (95% CI; -0.78 to +0.22) log₁₀ eCFU/mL for
43 RHZE ($p = 0.332$)

44 **Conclusion:** TB-MBLA distinguished *Mtb* elimination rates in sputa from patients receiving different
45 treatment regimens, suggesting a reliable monitoring tool for RR/MDR-TB, that does not require
46 mycobacterial culture.

47 **Introduction**

48 Measurement of pulmonary tuberculosis (PTB) treatment response in endemic settings largely depends
49 on sputum smear microscopy ^[1]. While the sputum smear microscopy detection threshold is at least 10³
50 *Mycobacterium tuberculosis* (*Mtb*) in colony-forming-units in 1 mL (CFU/mL) per sputum sample,
51 many patients with PTB such as those with human immunodeficiency virus and the acquired
52 immunodeficiency syndrome (HIV/AIDS) present with paucibacillary disease and may be unable to
53 produce a good quality sputa for detection of acid-fast-bacilli (AFB) ^[2,3]. Besides, sputum smear
54 microscopy cannot differentiate drug susceptibility, thus it is not applicable for rifampicin and or
55 multidrug resistant (RR/MDR)-TB diagnosis or treatment monitoring. Furthermore, microscopy cannot
56 distinguish viable from non-viable *Mtb* which requires prolonged incubation in solid or liquid media ^[3].
57 Patients with RR/MDR-TB are typically monitored for cultured growth in Lowenstein-Jensen (LJ) solid
58 medium or the Mycobacterium Growth Indicator Tube liquid culture system. Culture is sensitive with
59 a detection limit of 10 – 100 CFU/mL of sputum, yet it is also prone to contamination and can take up
60 to 8 weeks to determine a definitive positive or negative result, thereby limiting the ability to take
61 appropriate and timely clinical action ^[4].

62 The novel TB molecular bacterial load assay (TB-MBLA) was developed by Gillespie et al and used
63 for monitoring clearance of *Mtb* from sputa, as a marker for TB treatment response ^[5]. TB-MBLA is a
64 real-time polymerase chain reaction (RT-qPCR) assay which detects and quantifies elimination of 16S
65 rRNA from both viable replicating and dormant *Mtb* in patient's sputa during treatment ^[6]. Previously,
66 TB-MBLA was assessed by the Pan-African Consortium for Evaluation of Anti-TB Antibiotics
67 (PanACEA) group in patients treated for drug-sensitive (DS)-TB, and demonstrated considerable
68 potential to replace both smear microscopy and culture for monitoring TB treatment response ^[6-8]. TB-
69 MBLA was found to be consistently read as positive for samples with as low as 10 CFU/mL of *M.*
70 *tuberculosis* and the cycle threshold for this read-out has been optimized at a value of 30 ^[6].

71 Recently, TB-endemic countries, including Tanzania, have adopted new and repurposed TB medicines,
72 such as bedaquiline, delamanid and linezolid, and constructed regimens with limited microbiological

73 evidence of effectiveness in patients with RR/MDR-TB. Hence, we deployed TB-MBLA to describe
74 elimination of *Mtb* in patients receiving RR/MDR-TB and DS-TB treatment. We tested the hypothesis
75 that *Mtb* elimination rates from the sputa, as measured by TB-MBLA, not only correlated with time-to-
76 culture conversion but were dependent upon the composition of the RR/MDR-TB antibiotic regimen.

77 **Materials and Methods**

78 **Patients, ethics and design**

79 From August 2018 to December 2019, longitudinal cohort study was conducted among patients with
80 RR/MDR- and DS-TB confirmed using Xpert® MTB /Rif^[9]. The study was approved by the National
81 Institute for Medical Research (NIMR) in Tanzania (NIMR/HQ/R.8a/Vol. IX/2662). Permission to
82 conduct the study was granted by authorities of the Kibong'oto Infectious Diseases Hospital (KIDH).
83 Inclusion criteria were patients aged at least 18 years who consented to provide quality early-morning
84 sputum and clinical information. Critically ill patients, pregnant women and those who interrupted
85 treatment were excluded. Each patient was followed for 16 weeks during which they provided sputum
86 for testing at day 0 (baseline), 3, 7, 14, 28, 56, 84 and 112 of treatment. The treatment regimens included
87 standard RHZE (rifampicin, isoniazid, pyrazinamide, ethambutol) for DS-TB; an all-oral bedaquiline
88 based regimen (bedaquiline, linezolid, levofloxacin, pyrazinamide and ethionamide), an injectable and
89 bedaquiline containing regimen (kanamycin, bedaquiline, levofloxacin, pyrazinamide and
90 ethionamide), and injectable-containing but bedaquiline free regimen (kanamycin, levofloxacin,
91 pyrazinamide, ethionamide and cycloserine) containing regimens for RR/MDR-TB.

92 **Study Setting**

93 Patients were recruited at KIDH, national centre of excellence for clinical management of drug resistant
94 (DR)-TB located in the Siha district of Kilimanjaro region in Tanzania^[9]. TB-MBLA testing was
95 performed at the National Institute for Medical Research, Mbeya Medical Research Centre branch,
96 given that laboratory's prior experience with the assay.

97 **Sample size determination**

98 The numbers of patients required to determine differences in bactericidal activity over time in 4
99 treatment regimens were calculated as previously reported by Guo et al ^[10]. We assumed a Spearman
100 correlation of 0.51, and a baseline *Mtb* burden of 5.5 log₁₀ eCFU/mL, as well as daily *Mtb* decline and
101 decay rate of 0.42 and 0.05 log₁₀ eCFU/mL respectively ^[6,8]. Hence, at least 9 patients were needed per
102 regimen to reach a power of 90% with a two-sided type I error of 5%. Considering a RR/MDR-TB
103 treatment success of 56% globally and 75% in Tanzania ^[11], at least 20% of patients were likely to lost
104 be to follow up and hence a minimum of 45 patients were desirable to be sampled.

105 **TB-MBLA and Culture**

106 *M. tuberculosis* quantification by TB-MBLA was performed as described by Gillespie et al ^[11]. In
107 summary, 1mL of homogenized sputum was treated using guanidine thiocyanate (GTC), and was frozen
108 at -80°C to preserve the *M. tuberculosis* RNA. Total *M. tuberculosis* RNA was extracted using the
109 RNA pro (FastRNA Pro BlueKit MP Biomedical) according to manufacturer's instructions. The extract
110 was treated with DNase I enzyme (TURBO DNA-Free Kit Ambion) to remove DNA. The *M.*
111 *tuberculosis* 16S rRNA was quantified by reverse transcriptase quantitative PCR (RT-qPCR) and the
112 cycle-threshold CT translated to bacterial load (estimated CFU per mL (eCFU/mL) using a standard
113 curve on a Rotor gene Q 5plex platform (Qiagen). The cut-off for TB-MBLA positivity is a 30 CT value
114 that corresponds to 1.0 log₁₀ eCFU/mL, beyond which the test was considered negative ^[8,11]. *Mtb* culture
115 was performed on LJ slants from the remaining sputum collected at baseline, 14 days then monthly for
116 4 months per previous instructions ^[13].

117 **Statistical analysis**

118 Data were recorded in a clinical case report form (CRF), entered and cleaned before statistical analysis.
119 Patients who completed 8 treatment visits and had positive pre-treatment TB-MBLA results were
120 analysed and visualised in R, version 4.0.2 (<http://www.R-project.org>). Continuous variables such as
121 age, body-mass-index (BMI) in Kg/m² and time to TB-MBLA negativity were described as median
122 with their 25th and 75th interquartile range (IQR), and were compared using a Kruskal–Wallis test.

123 Accordingly, proportions for HIV, gender, cavitory-disease and previous TB treatment were compared
124 across different regimens using Chi-Square or Fischer's exact test. The rate of *Mtb* elimination
125 (\log_{10} eCFU/mL) was fitted on quadratic polynomial nonlinear-mixed-effects (NLME) for repeated
126 measures as previous ^[14], using Baseline bacterial load, cavity, HIV, silicosis and gender as fixed
127 effects. Individual patients were accounted for random effect. A model was reliably selected if had low
128 Akaike-information-criterion but high intraclass-correlation-coefficient (Table 2). Effect size in mean
129 *Mtb* load between two treatment regimens at month 4 were compared using one-way analysis-of-
130 variance (ANOVA) and Tukey's test for repeated measures ^[15]. The median time to TB-MBLA and
131 culture conversion to negative was estimated using the Kaplan-Meier method, and was compared across
132 different regimens using a log-rank test ^[16]. Cox Proportional-Hazards regression models were used to
133 estimate the hazard ratios (HR) for *Mtb* elimination, and was adjusted for the effects of HIV, baseline
134 bacillary load, cavitory disease, silicosis, gender, prior history of treatment for drug sensitive TB and
135 clearance rate. The mean *Mtb* load at baseline was the cut-off that beyond 4.0 \log_{10} eCFU/mL was
136 considered as high bacterial load. Mean clearance was considered as high if it was above the overall
137 mean clearance rate and low if it was below. Similarly, the overall mean rate of *Mtb* clearance per day
138 was used as the cut-off for low and high rate of clearance. A p value < 0.05 was considered significance.
139 A 95% confidence interval (CI) of the mean clearance rate and HR was included.

140 **Results**

141 **Population**

142 Of 59 patients enrolled, 37 patients with a total of 296 serial sputa were analysed. Reasons for exclusion
143 and patient's distribution are outlined in Figure 1. In total, 30 (81%) and 7 (19%) of 37 patients analysed
144 had RR/MDR-TB and DS-TB respectively. Clinical and demographics are presented in Table 1.
145 Twenty-seven (73%) out of 37 patients were male. Their median (IQR) age was 37 (32 – 49) years.
146 Patients who received standard RHZE treatment were younger than those who received RR/MDR-TB
147 treatment regimens (p = 0.038). Also, 11 (30%) patients were living with HIV infections with a CD4 T

148 cell count of 208 (95% CI; 144 – 272) cells/ μ L. More patients with HIV received an all-oral than
149 injectable-based treatment regimen ($p = 0.001$).

150 **Bactericidal activity over time**

151 The *Mtb* load measured by TB-MBLA and culture in Figure 2 decreased significantly over time ($R = -$
152 0.77 , $p < 0.001$). The mean *Mtb* load in \log_{10} eCFU/mL (95% CI) was reduced from 5.19 (4.40 – 5.78)
153 at baseline to 3.10 (2.70 – 3.50) at day 14, then to 2.52 (2.13 – 2.90) at day 28, 1.88 (1.53 -2.22) at day
154 56 and <1.36 (1.03 – 1.70) at day 84 through 112 of treatment. The overall mean daily *Mtb* elimination
155 was -0.24 (95% CI; -0.39 to -0.08) \log_{10} eCFU/mL, and it varied with treatment-regimen (Table 3, $p <$
156 0.001). An injectable and bedaquiline containing regimen had the highest mean *Mtb* elimination rate
157 followed by an all-oral bedaquiline based regimen compared to injectable-containing but bedaquiline
158 free reference regimen (Table 3, $p = 0.019$). Kanamycin containing regimens in Figure 3 had rapid
159 bactericidal activity at day 14, but was not translated into long term bactericidal effect ($p < 0.001$). An
160 all-oral bedaquiline-based regimen had a sharp decline after day 28.

161 **Median time to *M. tuberculosis* elimination**

162 There was strong positive correlation in time-to sputum conversion between TB-MBLA and culture [r
163 $= 0.46$ (95% CI; $0.36 - 0.55$), $p < 0.001$]. The overall median time to sputum TB-MBLA conversion to
164 negative was 56 (IQR; 28-84) days. The median time to TB-MBLA conversion to negative were 28, 42
165 and 84 days among patients on injectable and bedaquiline, an all-oral bedaquiline-based regimen, and
166 injectable-containing but bedaquiline free regimens respectively. Percentage of patients who converted
167 to sputum negative by TB-MBA and culture are shown in Figure 4. Approximately, 24% (9/37) of
168 patients had negative TB-MBLA at day 14 compared to 51% (19/37) culture negative ($p = 0.019$), which
169 was respectively increased to 43% (16/37) and 65% (24/37) at day 28 of treatment ($p = 0.002$). At day
170 56, 68% (25/37) had sputum converted to negative by TB-MBLA compared to 89% (33/37) by culture
171 ($p = 0.897$). Despite that all patients on standard RHZE converted to negative at day 90 of treatment, 4
172 patients with RR/MDR-TB did not convert to negative. Three out of these 4 patients were on injectable-
173 containing but bedaquiline-free, and remained positive by TB-MBLA at day 112

174 **Hazard ratio (HR) of *M. tuberculosis* elimination**

175 The overall mean Mtb load \log_{10} eCFU/mL at baseline was 5.19 (95% CI; 4.40 – 5.78), and was similar
176 in all patients treated with any of the 4 regimens (Table 3, $p = 0.453$). The mean Mtb load (\log_{10}
177 eCFU/mL) among female was 5.6 (95% CI; 5.0 – 6.2) \log_{10} eCFU/mL compared to 4.7 (95% CI; 4.3 –
178 5.2) \log_{10} eCFU/mL among male ($p = 0.017$) patients. Patients with chest cavity had mean Mtb load of
179 5.26 (95% CI; 4.45 – 5.87) compared to 4.40 (95% CI; 3.91 – 4.75) \log_{10} eCFU/mL in those without
180 cavity ($p = 0.080$). Adjusting for bacterial load, initial elimination rate, silicosis, chest cavity, HIV and
181 gender, the hazard-ratios for *Mtb* elimination were 12.37 (95% CI, 2.87 – 53.30; $p = 0.001$) and 14.31
182 (95% CI, 3.49 – 58.65; $p < 0.001$) for patients who received an all-oral bedaquiline and injectable and
183 bedaquiline-containing regimens respectively (Table 4). Bacterial load at baseline strongly correlated
184 positively with median time to sputum conversion to negative by both TB-MBLA and culture [$r = 0.48$
185 (95%CI; 0.18 – 0.69), $p = 0.003$]. High Mtb load and TB/silicosis were independently predictor of slow
186 Mtb elimination compared to low Mtb load and TB without silicosis (Table 4, $p \leq 0.033$)

187 **Discussion**

188 This study shows for the first time to our knowledge that TB-MBLA is promising for monitoring
189 treatment response among patients treated with DS- and -RR/MDR-TB regimens, as well as those with
190 concomitant TB/silicosis. As measured by TB-MBLA, *M. tuberculosis* decreased significantly over
191 time on treatment, and this kinetic correlated with what was observed using LJ culture medium. For
192 decades, culture has been used as a routine microbiological tool for monitoring drug-resistant TB
193 treatment response ^[17,18], but in many TB endemic settings, culture is unavailable or limited to
194 specialized centres. Importantly, culture results can take up to 8 weeks from the time of sputum
195 collection, which when making treatment decisions based on a result from a two-months old specimen,
196 is akin to driving a car while only looking in the rear-view mirror. Given the continued decentralization
197 of RR/MDR-TB services, monitoring treatment response in laboratories capable of performing qPCR,
198 such as with Xpert MTB/RIF, will allow laboratory assays to impact treatment decisions closer to the

199 point-of-care. Therefore this study in RR/MDR-TB compliments the growing evidence base for the
200 application of TB-MBLA in routine clinical management [6,8,19].

201 Interestingly, our findings suggest that bactericidal activity at day 14 may not be a suitable predictor of
202 the long-term efficacy of a regimen, particularly when that regimen is bedaquiline containing. In this
203 cohort at day 14, more than 75% of people had a positive TB-MBLA and more than half had a positive
204 culture result. Whereas between 14-56 days we observed substantial *M. tuberculosis* elimination in
205 those treated with a bedaquiline containing regimens, suggesting that evaluation of bactericidal activity
206 be performed later, such as at day 56, for modern RR/MDR-TB regimens. These findings may contradict
207 those from a phase 2b trial where the bactericidal activity of a bedaquiline containing regimen as was
208 measured by culture media at day 56 proved an unreliable indicator of a regimen's ability to predict
209 long term treatment outcomes or shorten treatment duration, and rather raise the question of whether
210 TB-MBLA may in fact be a superior predictor to culture.^[20]

211 Another important finding from this study of TB-MBLA is that *M. tuberculosis* elimination kinetics
212 were regimen-dependent. Overall, more rapid elimination occurred during the first 28 days for all
213 regimens, yet that earlier rapid elimination was more prominent at day 14 for patients who received
214 kanamycin regardless of receipt of bedaquiline, followed by those who received an all-oral bedaquiline
215 containing regimen, which did not achieve these rates of elimination until 1 month or more of treatment.
216 This observation concurs with previous reports that the bactericidal activity of bedaquiline in MDR-TB
217 is delayed at the beginning, but accelerates later in therapy [21]. Despite the superior activity of
218 kanamycin containing regimens at day 14, this more rapid early elimination of *M. tuberculosis* was not
219 sustained as a long term-bactericidal effect, such that 3 patients on injectable containing but bedaquiline
220 free regimen remained positive after 4 months of treatment. These findings as measured by TB-MBLA
221 fit with the pharmacodynamical understanding that kanamycin and other aminoglycoside/polypeptides
222 if active against mycobacteria, primarily exert their effect against those extracellular organisms that are
223 rapidly dividing and may be more abundant early in the treatment course [22,23].

224 The shorter overall time to sputum conversion to negative, as measured by TB-MBLA and conventional
225 culture, for all patients who received bedaquiline regardless of kanamycin further supports arguments
226 that bedaquiline should be a cornerstone of regimens designed to shorten MDR-TB treatment duration
227 [24]. The conventional injectable-containing but bedaquiline free regimen has been in practice for
228 decades, even though more than 40% of patients treated with this regimen had unfavourable outcomes
229 in TB endemic settings [11]. Aminoglycosides such as kanamycin is no longer part of the current MDR-
230 TB treatment regimens not because of its lack of bactericidal activity, as our data would suggest the
231 contrary in the early treatment period, but rather because of the significant toxicity and patient
232 intolerances that led to treatment interruption [25,26]. While we do not advocate this approach, from
233 microbiological perspective alone, as demonstrated in this study and others such as Mpagama *et al.* [27],
234 kanamycin could be included for first month only for instance and then dropped before toxicities
235 accumulate. In a more patient-centered approach however, our findings demonstrate how potentially
236 important it will be to find tolerable substitutes for kanamycin that can match the early bactericidal
237 effect.

238 The main strengths in this study is that we have utilized TB-MBLA to model elimination rates among
239 patients with RR/MDR-TB and those with TB/silicosis. We have shown that patients with TB/silicosis
240 had slower *M. tuberculosis* elimination rates by TB-MBLA compared to those with TB and without
241 silicosis. This slow rate of elimination could partially be attributed to the underlying pulmonary
242 pathophysiology which can include progressive massive fibrosis [28,29], and anatomically, a blunted local
243 host immune response to *M. tuberculosis* infection [28]. We observed a similarly slower rate of *M.*
244 *tuberculosis* elimination among patients with RR/MDR-TB who had high initial bacterial load, which
245 supplements previous studies of TB-MBLA kinetics from patients with drug sensitive TB [6,8,19].
246 Limitations of the study include the endpoints, which were limited to 4 months such that predicting
247 long-term treatment success was beyond the scope of this study. Nevertheless, modelling *M.*
248 *tuberculosis* elimination for 4 months as we accomplished here has been used as marker for treatment
249 failure and relapse in several observational studies [18,30], and exceeds the duration of monitoring used

250 in other trials of R/MDR-TB regimens that have employed conventional culture based techniques. [20]
251 Additionally, this study had no control over the treatment regimens prescribed. However, given the
252 feasibility of TB-MBLA and the comparability of this study's findings to those prior with TB-MBLA
253 in drug-susceptible TB [8] , we plan to apply TB-MBLA systematically within an ongoing operational
254 research protocol for injectable-free RR/MDR-TB treatment in Tanzania, that employs standardized
255 regimens over varying treatment durations. Lastly, the number of patients per treatment regimen were
256 small such that findings should be cautiously interpreted with inference to other populations with
257 RR/MDR-TB. However, considering the low MDR-TB burden in countries like Tanzania as well as the
258 repeated measurements per patient, findings in this study are critical to inform how TB-MBLA may be
259 applied as a culture-independent method for RR/MDR-TB care locally.

260 In conclusion, patients who received bedaquiline-containing regimens exhibited higher *M. tuberculosis*
261 elimination-rates and had shorter time-to sputum TB-MBLA and culture conversion to negative. While
262 both kanamycin containing regimens had superior bactericidal activity during two weeks of RR/MDR-
263 TB treatment, the addition of bedaquiline allowed for improved elimination after 1 month of therapy.
264 Together, these findings provide insight into formulating optimal all-oral bedaquiline containing
265 regimens with the best potential to shorten MDR-TB treatment duration [20,26,31]. Given the ease of use
266 of TB-MBLA and the fact that it does not require laboratory procedures associated with culture or the
267 prolonged time to receive a culture-based result, we envision that TB-MBLA can be used to make
268 regimen adjustments, and enhance infection control practices for patients with RR/MDR-TB and health
269 workers in hospital and community settings

270 **Acknowledgements**

271 This study received financial support from the EDCTP2 programme supported by the European Union
272 project (grant number: TMA2016SF-1463-REMODELTZ) and DELTAS Africa Initiative (Afrique
273 One-ASPIRE /DEL-15-008). The Afrique One-ASPIRE is funded by a consortium of donors including
274 the African Academy of Sciences, Alliance for Accelerating Excellence in Science in Africa, the New
275 Partnership for Africa's Development Planning and Coordinating Agency, the Wellcome Trust

276 (107753/A/15/Z), and the UK Government. All funding bodies have had no role in the
277 conceptualization, methodology, data interpretation and writing of manuscript.

278 Furthermore, authors acknowledge Ms Batuli Mono, Taji Mnzava, Joseph Kachala and Dr Bibie Said
279 of KIDH for their assistant with recruitment and data collection from study participants. We also thank
280 Mr. Elisha S. Juma and Ms Sarapia P. Malya of KIDH, and Emmanuel Sichone and Joseph John of
281 NIMR Mbeya for assisting with laboratory work. In addition, we also acknowledge the KIDH
282 administration for granting permission to conduct this study.

283 **Transparency declarations.**

284 All authors have no conflict of interest to declare. PMM, EAM, ES, WS and SGM conceived the study,
285 designed the work and interpreted clinical and TB-MBLA results. PMM and BM acquired data. PMM,
286 KK, EAM, PPJP, WS, and SGM analyzed the data. PMM drafted the manuscript and responded to all
287 co-authors' inputs. SHG, NEN, and SKH reviewed the manuscript. All authors wrote, approved and
288 agreed to be accountable for all scientific aspects in the final version of this manuscript.

289 **References**

- 290 1. Mitnick CD, White RA, Lu C, Rodriguez CA, Bayona J, Becerra MC, et al. Multidrug-resistant
291 tuberculosis treatment failure detection depends on monitoring interval and microbiological
292 method. *Eur Respir J* 2016;48(4):1160–70. <http://dx.doi.org/10.1183/13993003.00462-2016>
- 293 2. Park JH, Choe J, Bae M, Choi S, Jung KH, Kim MJ, et al. Clinical characteristics and radiologic
294 features of immunocompromised patients with pauci-bacillary pulmonary tuberculosis receiving
295 delayed diagnosis and treatment. *Open Forum Infect Dis* 2019;6(2):1–9.
- 296 3. Prasanta Kumar Das, Somtirtha B. Ganguly BMS. Sputum Smear Microscopy in Tuberculosis:
297 It Is Still Relevant in the Era of Molecular Diagnosis When Seen from the Public Health
298 Perspective. *Biomed Biotechnol Res J* 2019;3:77–9.
- 299 4. van Zyl-Smit RN, Binder A, Meldau R, Mishra H, Semple PL, Theron G, et al. Comparison of
300 quantitative techniques including Xpert MTB/RIF to evaluate mycobacterial burden. *PLoS One*

- 301 2011;6(12):e28815.
- 302 5. World Health Organization. Global Tuberculosis Report 2017. 20 Avenue Appia, 1211 Geneva
303 27, Switzerland: World Health Organization; 2017.
- 304 6. Honeyborne I, McHugh TD, Phillips PPJ, Bannoo S, Bateson A, Carroll N, et al. Molecular
305 bacterial load assay, a culture-free biomarker for rapid and accurate quantification of sputum
306 Mycobacterium tuberculosis bacillary load during treatment. *J Clin Microbiol*
307 2011;49(11):3905–11.
- 308 7. Mtafya B, Sabiiti W, Sabi I, John J, Sichone E, Ntinginya NE, et al. Molecular bacterial load
309 assay concurs with culture on NaOH-induced loss of mycobacterium tuberculosis viability. *J Clin*
310 *Microbiol* 2019;57(7):1–9.
- 311 8. Sabiiti W, Azam K, Farmer ECW, Kuchaka D, Mtafya B, Bowness R, et al. Tuberculosis
312 bacillary load, an early marker of disease severity: The utility of tuberculosis Molecular Bacterial
313 Load Assay. *Thorax* 2020;0:1–3.
- 314 9. Mbelele PM, Aboud S, Mpagama SG, Matee MI. Improved performance of Xpert MTB/RIF
315 assay on sputum sediment samples obtained from presumptive pulmonary tuberculosis cases at
316 Kibong’oto infectious diseases hospital in Tanzania. *BMC Infect Dis* 2017;17:1–7.
- 317 10. Guo Y, Logan HL, Glueck DH, Muller KE. Selecting a sample size for studies with repeated
318 measures. *BMC Med Res Methodol* 2013;13:100. doi:10.1186/1471-2288-13-100
- 319 11. World Health Organization. Global tuberculosis report 2019. Geneva: World Health
320 Organization. <http://repositorio.unan.edu.ni/2986/1/5624.pdf>
- 321 12. Gillespie H Stephen SW and OK. Mybacterial Load Assay. *Methods Mol Biol*
322 2017;1616(3):155–70. <https://www.springer.com/gp/book/9781493970353>
- 323 13. Tripathi K, Tripathi PC, Nema S, Shrivastava AK, Dwiwedi K. Modified Petroff ’ s Method : an
324 Excellent Simplified Decontamination Technique in Comparison with Petroff ’ s Method. *Int J*
325 *Recent Trends Sci Technol* 2014;10(3):461–4.

- 326 14. Rustomjee R, Lienhardt C, Kanyok T, Davies GR, Levin J, Mthiyane T, et al. A phase II study
327 of the sterilising activities of ofloxacin, gatifloxacin and moxifloxacin in pulmonary tuberculosis.
328 *Int J Tuberc Lung Dis* 2008;12(2):128–38.
- 329 15. Hazra A, Gogtay N. Biostatistics series module 3: Comparing groups: Numerical variables.
330 *Indian J Dermatol* 2016;61(3):251–60.
- 331 16. Gillespie SH, Crook AM, McHugh TD, Mendel CM, Meredith SK, Murray SR, et al. Four-month
332 Moxifloxacin-based regimens for drug-sensitive tuberculosis. *N Engl J Med* 2014;371(17):1577–
333 87.
- 334 17. Rockwood N, Bruyn E, Morris T, Wilkinson RJ, Kingdom U, Infectious C, et al. Assessment of
335 treatment response in tuberculosis. *Expert Rev Respir Med* 2017;10:643–54.
- 336 18. Goletti D, Lindestam Arlehamn CS, Scriba TJ, Anthony R, Cirillo DM, Alonzi T, et al. Can we
337 predict tuberculosis cure? What tools are available? *Eur Respir J* 2018;52(5).
338 <http://dx.doi.org/10.1183/13993003.01089-2018>
- 339 19. Honeyborne I, Mtafya B, Phillips PPJ, Hoelscher M, Ntinginya EN, Kohlenberg A, et al. The
340 molecular bacterial load assay replaces solid culture for measuring early bactericidal response to
341 antituberculosis treatment. *J Clin Microbiol* 2014;52:3064–7.
- 342 20. Tweed CD, Dawson R, Burger DA, Conradie A, Crook AM, Mendel CM, et al. Bedaquiline,
343 moxifloxacin, pretomanid, and pyrazinamide during the first 8 weeks of treatment of patients
344 with drug-susceptible or drug-resistant pulmonary tuberculosis: a multicentre, open-label,
345 partially randomised, phase 2b trial. *Lancet Respir Med* 2019;7(12):1048–58.
346 [http://dx.doi.org/10.1016/S2213-2600\(19\)30366-2](http://dx.doi.org/10.1016/S2213-2600(19)30366-2)
- 347 21. Nguyen TVA, Cao TBT, Akkerman OW, Tiberi S, Vu DH, Alffenaar JWC. Bedaquiline as part
348 of combination therapy in adults with pulmonary multi-drug resistant tuberculosis. *Expert Rev*
349 *Clin Pharmacol* 2016;9(8):1025–37.
- 350 22. Krause KM, Serio AW, Kane TR, Connolly LE. Aminoglycosides: An overview. *Cold Spring*

- 351 Harb Perspect Med 2016;6(6):1–18.
- 352 23. Motta I, Calcagno A, Bonora S. Pharmacokinetics and pharmacogenetics of anti-tubercular
353 drugs: a tool for treatment optimization? *Expert Opin Drug Metab Toxicol* 2018;14(1):59–82.
354 <https://doi.org/10.1080/17425255.2018.1416093>
- 355 24. Doan TN, Cao P, Emeto TI, McCaw JM, McBryde ES. Predicting the outcomes of new short-
356 course regimens for multidrug-resistant tuberculosis using intrahost and pharmacokinetic-
357 pharmacodynamic modeling. *Antimicrob Agents Chemother* 2018;62(12):1–11.
- 358 25. World Health Organization. Consolidated guidelines on drug-resistant tuberculosis treatment.
359 Geneva, Switzerland: World Health Organization; 2019.
- 360 26. World Health Organization. Rapid Communication : Key changes to treatment of multidrug- and
361 rifampicin-resistant tuberculosis. Geneva: 2018.
- 362 27. Mpagama SG, Ndusilo N, Stroup S, Kumburu H, Peloquin CA, Gratz J, et al. Plasma Drug
363 Activity in Patients on Treatment for Multidrug-. *Antimicrob Agents Chemother*
364 2014;58(2):782–8.
- 365 28. Konečný P, Ehrlich R, Gulumian M, Jacobs M. Immunity to the dual threat of silica exposure
366 and mycobacterium tuberculosis. *Front Immunol* 2019;9:3069.
- 367 29. Skowroński M, Halicka A, Barinow-Wojewódzki A. Pulmonary tuberculosis in a male with
368 silicosis. *Adv Respir Med* 2018;86(3):121–5.
- 369 30. Ahmad N, Ahuja SD, Akkerman OW, Alffenaar JWC, Anderson LF, Baghaei P, et al. Treatment
370 correlates of successful outcomes in pulmonary multidrug-resistant tuberculosis: an individual
371 patient data meta-analysis. *Lancet* 2018;392(10150):821–34.
- 372 31. Silva DR, Mello FC de Q, Migliori GB. Shortened tuberculosis treatment regimens: what is new?
373 *J Bras Pneumol* 2020;46(2):e20200009.
- 374

375 **Table 1. Socio-demographic and clinical characteristics of patients per treatment regimen.**

Variable	All	RHZE (n = 7)	Injectable ± BDQ (n = 21)	All-oral BDQ (n = 9)	p-value
Median age (IQR)	37 (32 – 49)	30 (29 – 33)	42 (34-54)	36 (33- 44)	0.038
Male (%)	27 (73)	4 (57)	18 (86)	5 (56)	0.125
Chest cavity, n (%)	29 (78)	7 (100)	14 (67)	8 (89)	0.163
Probable TB, n (%)	34 (92)	7 (100)	18 (86)	9 (100)	0.568
HIV positive, n (%)	11 (20)	0 (0)	3 (14)	8 (89)	0.001
TB/Silicosis, n (%)	7 (19)	1 (14)	4 (19)	2 (22)	0.731
Malnourished, n (%)	22 (59)	4 (57)	11 (52)	7 (78)	0.432
Retreatment, n (%)	23 (62)	5 (71)	14 (67)	4 (44)	0.528
Median BMI (IQR)	18 (15 – 19)	17 (15 – 20)	18 (16 – 20)	17 (15 – 18)	0.301
Median days spent before care (IQR)	84 (60 – 196)	85 (68 – 93)	84 (56 – 196)	88 (68 – 365)	0.778

BDQ, bedaquiline; BMI, body-mass-index; injectable± BDQ, kanamycin with or without BDQ and IQR, interquartile range.

376

377 **Table 2. Fitting and selection of a reliable polynomial nonlinear mixed effects model for repeated measures**

Polynomial models (degree)	Intercepts (log₁₀ eCFU/mL)	Intraclass correlation coefficient (ICC)	Standard deviation (SD)	Akaike information criterion (AIC)	Likelihood ratio test	p value
Non-poly (model 1)	3.00	0.54	0.81	722.89	1 vs. 2	< 0.001
Squared (model 2)	2.99	0.63	0.67	634.63	2 vs.3	< 0.001
Cubic (model 3)	3.00	0.65	0.63	611.59	3 vs. 4	< 0.001
Quadratic (model 4)	3.20	0.67	0.61	592.7	4 vs. 5	0.020
Pentadric (model 5)	2.89	0.68	0.60	588.58		

Model 4 had the lowest AIC and within variability (SD) but high ICC values, the key selection criteria for a reliable model, and hence it was used to model *M. tuberculosis* elimination rates

378

379 **Table 3 Mean daily *M. tuberculosis* elimination rates (log₁₀ eCFU/mL) and corresponding burden at day 0 (baseline) and 112 of treatment**

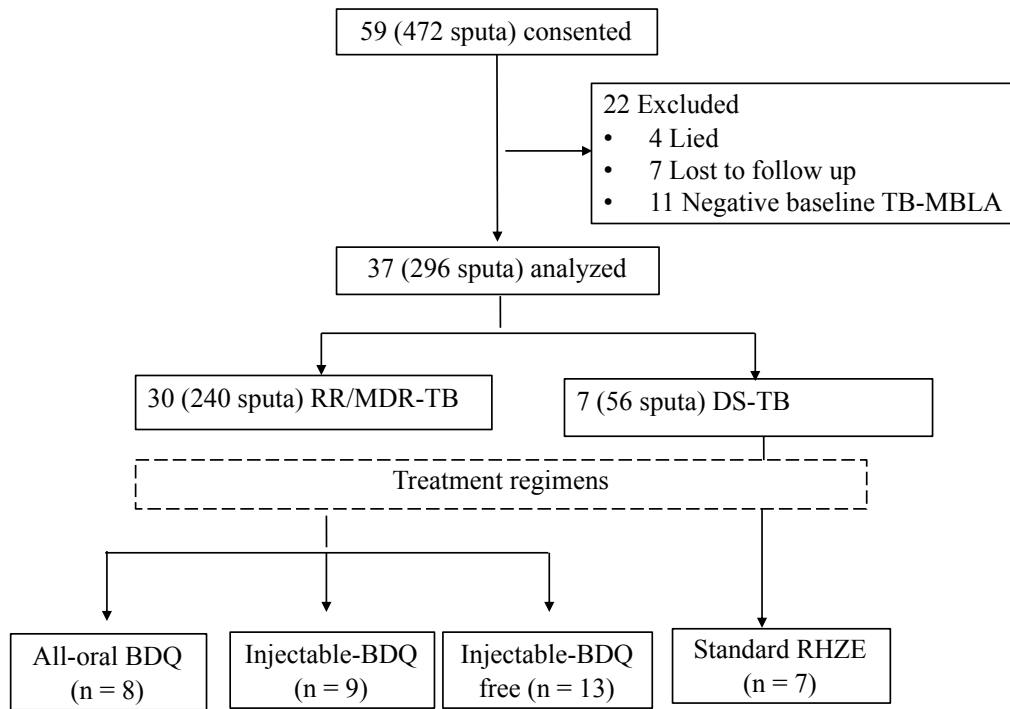
Treatment regimens	Mean <i>M. tuberculosis</i> elimination rates				Mean (95% CI) <i>M. tuberculosis</i> load	
	Unadjusted model for covariates		Adjusted model for covariates		Day 0 (baseline) †	Day 112 *
	Rates (95% CI)	p-value	Rates (95% CI)	p-value		
1. Reference (injectable-BDQ free)	-0.18 (-0.27 to -0.08)		-0.17 (-0.23 to -0.12)		4.73 (4.13 – 5.32)	2.77 (2.51- 3.04)
2. Injectable-bedaquiline	-0.48 (-1.25 to +0.28)	0.239	-0.62 (-1.05 to -0.20)	0.019	4.63 (3.95 – 5.47)	2.08 (1.81 - 2.36)
3. All-oral bedaquiline	-0.26 (-0.48 to +1.00)	0.507	-0.35 (-0.65 to -0.13)	0.054	5.36 (4.65 – 6.08)	2.47 (2.20 - 2.74)
4. Standard RHZE	-0.23 (-0.57 to +1.02)	0.593	-0.29 (-0.78 to +0.22)	0.332	5.17 (4.36 – 5.99)	2.51 (2.18 - 2.85)

†Baseline mean *M. tuberculosis* load in all regimens were comparable (ANOVA, p = 0.453). An asterisk (*) denotes p -values for mean difference in *M. tuberculosis* load for regimen pairwise comparison at day 112: regimen 1 & 2, p < 0.001; regimen 2 & 3, p = 0.031; regimen 1 & 3, p = 0.077; and regimen 2 & 4, p = 0.040. Reference regimen was the injectable-bedaquiline (BDQ) free regimen composed of kanamycin (KAN), levofloxacin (LFX), pyrazinamide (PZA), ethionamide (ETH) and Cycloserine (CS); Injectable-bedaquiline regimen was comprised of KAN, BDQ, LFX, PZA and ETH; All-oral bedaquiline regimen contained BDQ, LFX, linezolid (LZD), PZA and ETH; and the RHZE for rifampicin, isoniazid, PZA and ethambutol (E) Covariates adjusted included baseline bacterial load, cavity, gender, HIV and silicosis, *M. tuberculosis* elimination rates varied among regimens

381 **Table 4. Hazard ratio (HR) of *M. tuberculosis* (*Mtb*) elimination in Cox Proportion-Hazard model**

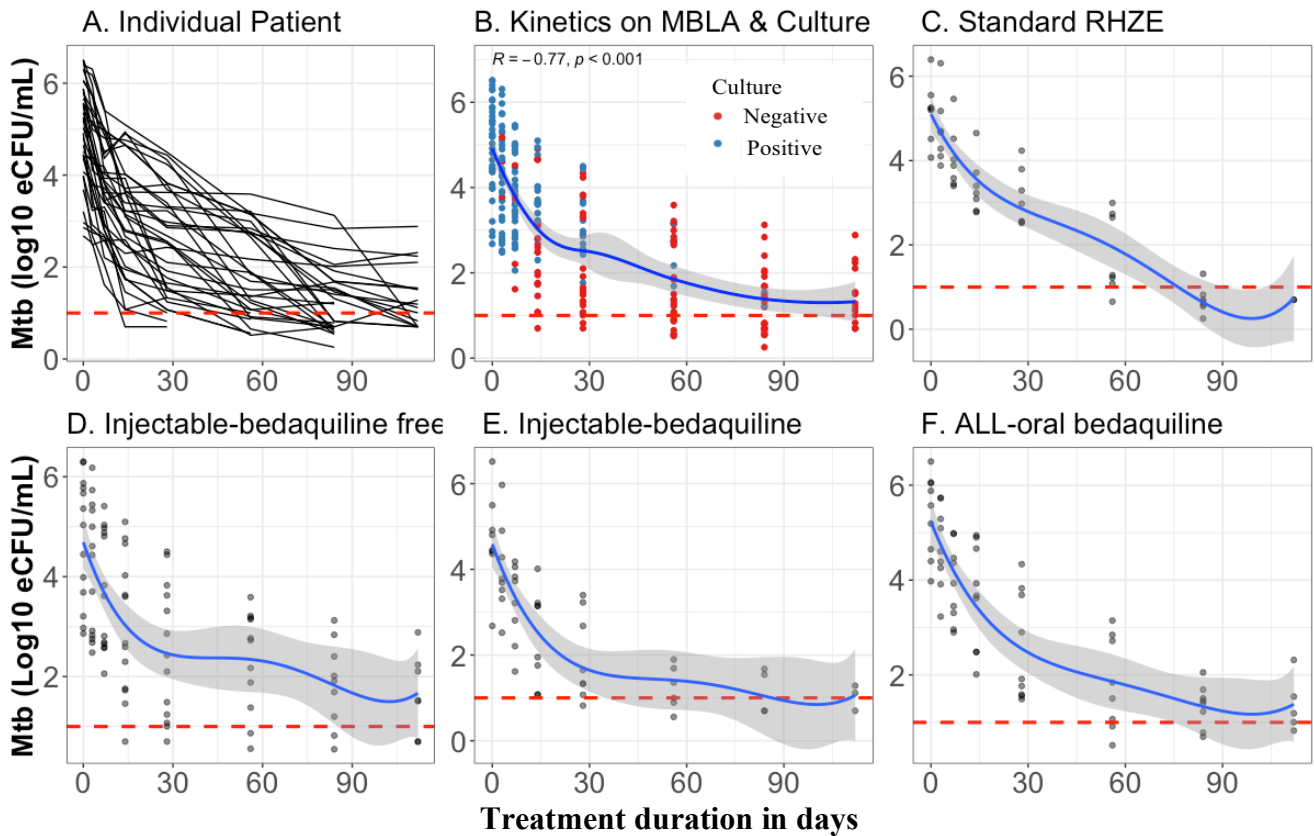
Predictor Variable	Unadjusted model		Adjusted model	
	HR (95% CI)	p-value	HR (95% CI)	p-value
Male gender	0.86 (0.40 – 1.85)	0.705	2.44 (0.82 – 7.24)	0.109
TB/Silicosis	0.20 (0.10- 0.88)	0.028	0.12 (0.03 – 0.49)	0.003
TB/HIV	2.26 (1.07 -4.77)	0.033	0.88 (0.31 – 2.50)	0.813
Cavitary disease	0.38 (0.17 - 0.86)	0.021	0.85 (0.17 – 2.70)	0.790
Positive chest x-ray	0.57 (0.17 – 1.88)	0.354	0.23 (0.03 – 1.62)	0.790
High <i>Mtb</i> load	0.72 (0.54 -0.97)	0.033	0.26 (0.13 – 0.54)	< 0.001
Retreatment	1.02 (0.51 - 2.05)	0.958	0.59 (0.24 – 1.44)	0.248
All-oral bedaquiline	1.58 (0.61 - 4.04)	0.344	12.37 (2.87 – 53.30)	0.001
Injectable- bedaquiline	4.63 (1.64 – 13.09)	0.004	14.31 (3.49 – 58.65)	< 0.001
Standard RHZE	1.43 (0.53 – 3.89)	0.482	3.25 (0.90 – 11.73)	0.072
High initial <i>Mtb</i> elimination rate	5.96 (2.03 – 17.48)	0.009	4.81 (1.39 – 16.65)	0.013

All-oral bedaquiline regimen was comprised of Bedaquiline (BDQ), levofloxacin (LFX), linezolid (LZD), pyrazinamide (PZA) and ethionamide (ETH). Injectable-bedaquiline is a modified regimen comprised of kanamycin (KAN), BDQ, LFX, PZA and ETH. Standard RHZE included rifampicin (H), isoniazid (H), PZA and ethambutol (E).



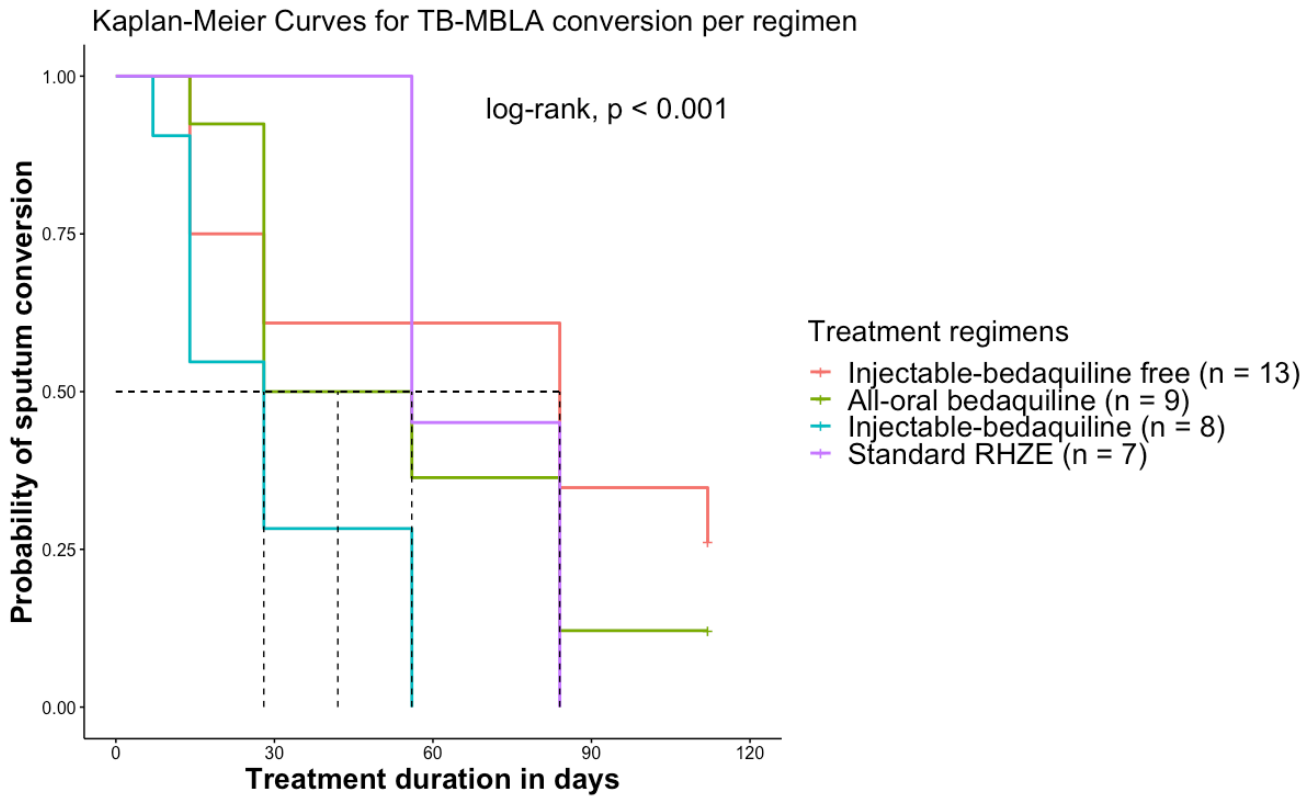
383

384 **Figure 1. Recruitment and patient's distributions in different treatment regimens.** DS-TB, drug
 385 sensitive TB; RR/MDR-TB, rifampicin or multidrug resistance, TB-MBLA; tuberculosis molecular
 386 bacterial load assay, Standard RHZE comprised of rifampicin, isoniazid, PZA & ethambutol).
 387 Injectable-BDQ free regimen was comprised of kanamycin (KAN), levofloxacin (LFX), pyrazinamide
 388 (PZA), ethionamide (ETH) and Cycloserine (CS). Injectable-BDQ regimen was comprised of KAN,
 389 Bedaquiline (BDQ), LFX, PZA and ETH; and All-oral BDQ regimen contained BDQ, LFX, linezolid
 390 (LZD), PZA and ETH.



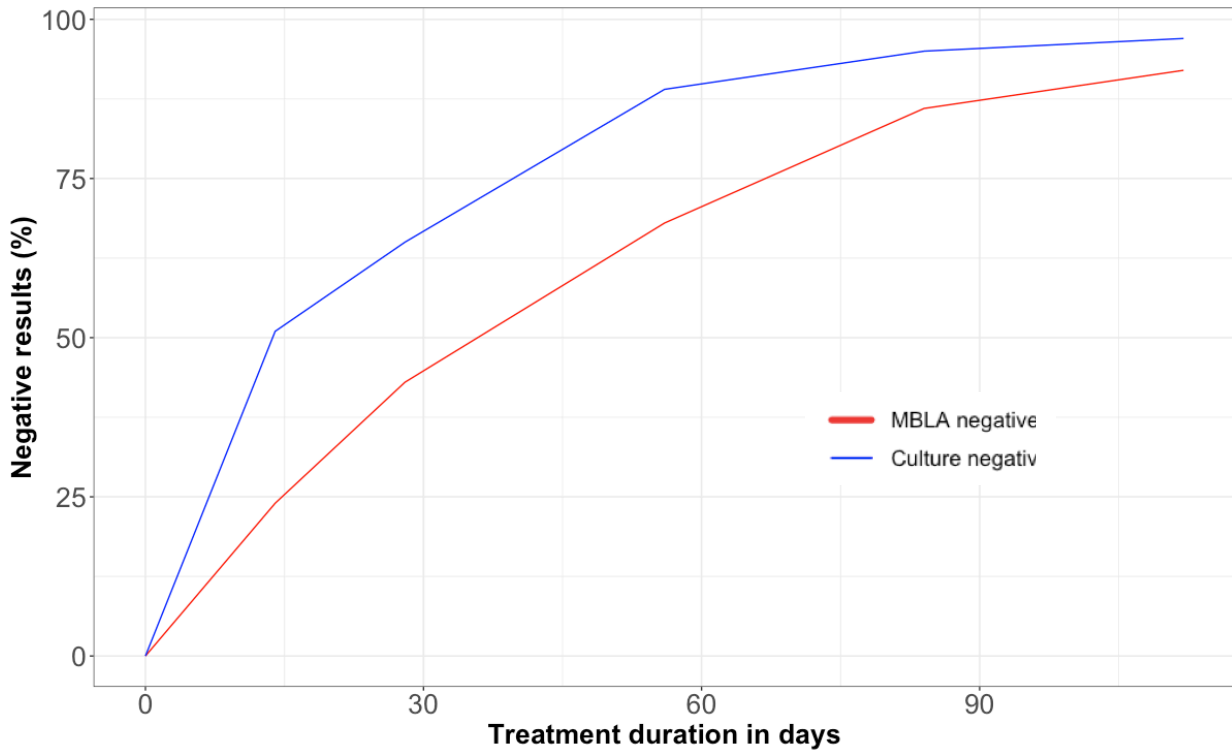
391

392 **Figure 2. *M. tuberculosis* elimination during treatment.** The plots A-F show *M. tuberculosis* (*Mtb*)
 393 kinetics between patients (A) as measured by TB-MBLA and culture (B) among patients treated with
 394 standard RHZE (C), injectable bedaquiline free regimen (D) containing kanamycin (KAN),
 395 levofloxacin (LFX), pyrazinamide (PZA), ethionamide (ETH) and Cycloserine (CS); Injectable-
 396 bedaquiline regimen (E) was comprised of KAN, Bedaquiline (BDQ), LFX, PZA and ETH; and an all-
 397 oral bedaquiline regimen (F) containing BDQ, LFX, linezolid (LZD), PZA and ET



398

399 **Figure 3. *M. tuberculosis* elimination per treatment regimen over time.** Bedaquiline containing
 400 regimens had short median time to TB-MBLA conversion to negative compared to injectable-containing
 401 but bedaquiline free regimen containing kanamycin (KAN), levofloxacin (LFX), pyrazinamide (PZA),
 402 ethionamide (ETH) and Cycloserine (CS). Injectable-bedaquiline was comprised of KAN, bedaquiline
 403 (BDQ), LFX, PZA and ETH; an all-oral bedaquiline regimen was composed of BDQ, LFX, linezolid
 404 (LZD), PZA and ETH, and Standard RHZE composed of rifampicin, isoniazid, PZA and ethambutol.



405

406 **Figure 4. Percentage of patients who converted to negative by TB-MBLA and culture over time**
 407 **(N = 37).** Changes in presentation of patients with negative results by tuberculosis molecular bacterial
 408 load assay (TB-MBLA, red line) and Lowenstein-Jensen culture medium (blue line). In the first 60
 409 days, high proportion of patients had negative culture results compared to TB-MBLA.