Evaluation of stability and inactivation methods of SARS-CoV-2 in context of laboratory settings

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Summary:

The novel coronavirus SARS-CoV-2 is the causative agent of the acute respiratory disease COVID-19, which has become a global concern due to its rapid spread. Laboratory work with SARS-CoV-2 in a laboratory setting was rated to biosafety level 3 (BSL-3) biocontainment level. However, certain research applications in particular in molecular biology require incomplete denaturation of the proteins, which might cause safety issues handling contaminated samples. In particular, it is critical to provide proof of inactivation before samples can be removed from the BSL-3.

In this study, we evaluated common lysis buffers that are used in molecular biological laboratories for their ability to inactivate SARS-CoV-2. We have found that guanidine thiocyanate, SDS, and Triton-X containing lysis buffers were effective in inactivation of SARS-CoV-2, however, the M-PER lysis buffer containing a proprietary detergent failed to inactivate SARS-CoV-2.

Furthermore, we compared chemical and non-chemical inactivation methods including ethanol, acetone-methanol mixture, PFA, UV-C light, and heat inactivation. In addition, the stability of the virus in cell culture media at 4°C and on surfaces used in laboratory environment was analyzed.

In conclusion, careful evaluation of the used inactivation methods are required and additional inactivation steps are necessary before removal of lysed viral samples from BSL-3.

Keywords:

SARS-CoV-2, COVID-19, bio safety, corona virus, stability, inactivation

Declaration of interest:

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1. Introduction:

The coronavirus disease 2019 (COVID-19) is caused by an infection with the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). The origin of SARS-CoV-2 outbreak was initially described in Wuhan, China (Zhu et al., 2020) and rapidly established a worldwide pandemic. There are globally 14,348,858 cases 603,691 deaths in total while 229,780 new cases and 5,111 deaths are reported within 24 h (WHO, 2020).

Numerous experimental studies are currently being carried out, which require appropriate inactivation methods to restrict laboratory born spread among scientists and lab personnel.

SARS-CoV-2 is a spherical beta coronavirus with a size of 120 nm in diameter, which has a lipid envelope. Persistence and inactivation of coronaviruses including the highly pathogenic SARS-CoV and MERS-CoV, which emerged in the last decades, was evaluated in several publications (reviewed in (Kampf et al., 2020a; Kampf et al., 2020b)). Thus, a whole series of chemical and physical inactivation methods such as UV radiation, heat inactivation and detergents are effective inactivation methods against SARS-CoV-2 (Kratzel et al., 2020b). However since human coronaviruses were shown to remain infectious on inanimate surfaces at room temperature for up to 9 days (reviewed in (Kampf et al., 2020a; Kampf et al., 2020b)), contamination of frequently touched surfaces in laboratory settings are therefore a potential source of viral transmission.

In addition, it might be assumed SARS-CoV-2 can remain infectious for much longer period of time when stored in liquid milieu at appropriate temperatures. This is the case in laboratory environments in which the condition of the sample has to be partially preserved for the subsequent molecular biological assays. Quantitative reverse transcriptase polymerase chain reaction (RT-qPCR) is the major detection method for SARS-CoV-2, in which guanidine thiocyanate containing chaotropic buffers are efficient in disrupting viral structures. However, there are applications in particular in molecular biology that require incomplete denaturation of the proteins (e.g. luciferase measurements).

Thus, in this study we evaluated common lysis buffers that are used in molecular biological laboratories for their ability to inactivate SARS-CoV-2. Furthermore, we investigated the stability of SARS-CoV-2 under laboratory conditions such as on different surfaces, temperatures, physical (UV) and chemical noxes.

2. Materials and methods:

4.1 Cell culture and virus propagation

Caco2 and Vero cells were cultured in Minimum Essential Medium (MEM) supplemented with 10% fetal calf serum (FCS), 100 IU/ml of penicillin and 100 g/ml of streptomycin. Viral titers of SARS-CoV isolate FFM-1 (Drosten et al., 2003) and SARS-CoV-2 isolate (Frankfurt 1, FFM1) (Hoehl et al., 2020; Toptan et al., 2020) were determined by $TCID_{50}$. All infectious work was performed under biosafety level 3 (BSL-3) conditions in a BSL-3 facility according to the Committee on Biological Agents (ABAS) and Central Committee for Biological Safety (ZKBS).

2.2 Determination of SARS-CoV-2 infectivity by $TCID_{50}$

Infectious titer of SARS-CoV-2 supernatant were determined by an end-point limiting dilution assay. Briefly, Caco-2 were seeded onto a 96-well plate and infected with untreated or treated SARS-CoV-2 containing supernatant in an initial 1:10 dilution (quadruplicates). Titration was performed on 96-well plate with ten serial 1:10 dilutions. Viral titer was determined 48h post infection as 50% tissue culture infectious dose ($TCID_{50}$) as described by Spearman and Kaerber (Kärber, 1931). To analyze inhibition capacity of different inactivation methods viral titers were normalized to the untreated control and reduction factor was calculated as described elsewhere (Rabenau et al., 2005b). All infection experiments were performed with initial viral titer of 1x 10^6 $TCID_{50}$/mL if not indicated differently.

2.3 Chemical Inactivation of cell culture supernatants

To test different chemical compound to inactivate SARS-CoV-2, virus containing supernatant was mixed 1:1 or 1:10 with test compound in lysis buffer or fixation solution assay, respectively. Mixture was incubated for 10 minutes at room temperature before further processed. Testing different lysis buffers, compound-virus mixture was further diluted 1:1000 to avoid cytotoxic effect before added to the seeded Caco-2 cells. Viral load and inactivation capacity were determined by RT-qPCR and CV staining, respectively. Analyzing different fixation solutions, compound-virus mixture was further diluted 1:100 before added to the seeded Vero cells. Viral titers and inactivation capacity were determined by RT-qPCR and compared to viral control.

2.4 Physical Inactivation of cell culture supernatants

Analyzing physical parameters to inactivate SARS-CoV-2, virus containing supernatant was treated either at different temperatures or with UV-C light or was applied to different surfaces and stored for a distinct period of time. For surface inactivation SARS-CoV-2 were applied to cell culture dish (polystyrole; plastic), used smartphone display and used protection film and dried at room temperature. Dried virus spots were incubated for 6 h or 5 days at room temperature before wiped off with a PBS soaked swab (2 cm^2) and eluted in culture medium. Viral titer was determined by $TCID_{50}$. Testing different temperatures for inactivation of SARS-CoV-2 virus containing supernatant was incubated on a thermo shaker for defined time at 56°C, 60°C or 90°C. Determination of viral load was performed by infecting Vero cells and measuring gene copies/reaction by RT-qPCR. Finally, SARS-CoV-2, SARS-CoV- and Influenza A containing supernatant was applied to cell culture dish and dried. Dried virus spots were treated with UV light of different light sources (UVA Cube and UV-C LED, Hönle AG, Germany; Holton LaminAir, Heto-Holten, Denmark) for a defined time before eluted in culture medium. Viral load and inactivation capacity were determined by $TCID_{50}$ and CV staining, respectively.

2.5 Rotitest Vital viability assay

To analyze the viability of cells treated different compounds to inactivate SARS-CoV-2, the intracellular dehydrogenase activity was tracked using the Rotitest Vital Kit (Carl Roth GmbH + Co. KG, Karlsruhe, Germany). Briefly, cells were seeded in 96-well plates and incubated with 1/1000 dilution of inactivating compound in culture medium in triplicates per sample. After 48 h of incubation at 37°C and 5% CO_2, Rotitest Vital solution was added and analyzed using a multimode reader at 450 nm.

2.6 RT-qPCR analysis and detection of SARS-CoV-2 genomic RNA

For detection of intracellular and extracellular SARS-CoV-2 genomic RNA Primer M-475-F (5'-TGTGACATCAAGGACCTGCC-3') and M-574-R (5'-CTGAGTCACCTGCTACACGC-3') were used together with the Fam-BHQ1 dual-labeled probe M-507-P (5'-TGTTGCTACATCACGAACGC-3') to detect SARS-CoV-2 specific RNAs as described previously (Toptan et al., 2020). For normalization to intracellular mRNA level human GAPDH was measured using primer GAPDH-fwd (5'-TGCACCACCAACTGCTTA) and GAPDH-rev (5'-GGATGCAGGGATGATGTTC-3').

2.7 Determination of SARS-CoV-2 inactivation by Crystal Violet Staining

Inactivation of SARS-CoV-2 by chemical and physical methods was determined by Crystal Violet Staining (CV). Therefore, Caco-2 cells were seeded on a 96-well plate infected with untreated or treated SARS-CoV-2 supernatant and cultured for 48 h at 37°C, 5% CO_2. Cells were fixed after incubation with 3% paraformaldehyde (PFA) for 20 minutes at room temperature before stained with 0.1% Crystal Violet for another 20 minutes. Staining solution was removed and plates were dried overnight. Crystal violet was dissolved with 100% ice-cold methanol and analyzed using a multimode reader at 560 nm. To determine inactivation capacity samples were normalized to untreated control.

2.8 Statistical analysis

If not indicated differently all infection experiments were repeated in three independent experiments. Data analysis was performed in Microsoft Excel and GraphPad Prism 6 (GraphPad Software, USA). Statistical significance compared to untreated control was determined using unpaired student's t-test. Asterisks indicated p-values as * ($p < 0.05$), ** ($p \leq 0.01$) and *** ($p \leq 0.005$).

3. Results:

3.1 SARS-CoV-2 is stable for several weeks

To investigate how long SARS-CoV-2 viruses are stable under laboratory storage conditions, we thawed different stock aliquots of a SARS-CoV-2 strain FFM1 (Hoehl et al., 2020; Toptan et al., 2020) (which were cryo-conserved at -80°C) at weekly intervals and stored them at 4°C in cell culture medium for up to 23 weeks. The long-stored virus stocks were then examined quantitatively for their infectivity using $TCID_{50}$ according to Spearmen and Kaerber. Remarkably, we found that even after 23 weeks under these storage conditions, there was no significant decrease in viral infectivity (***Figure 1a***).

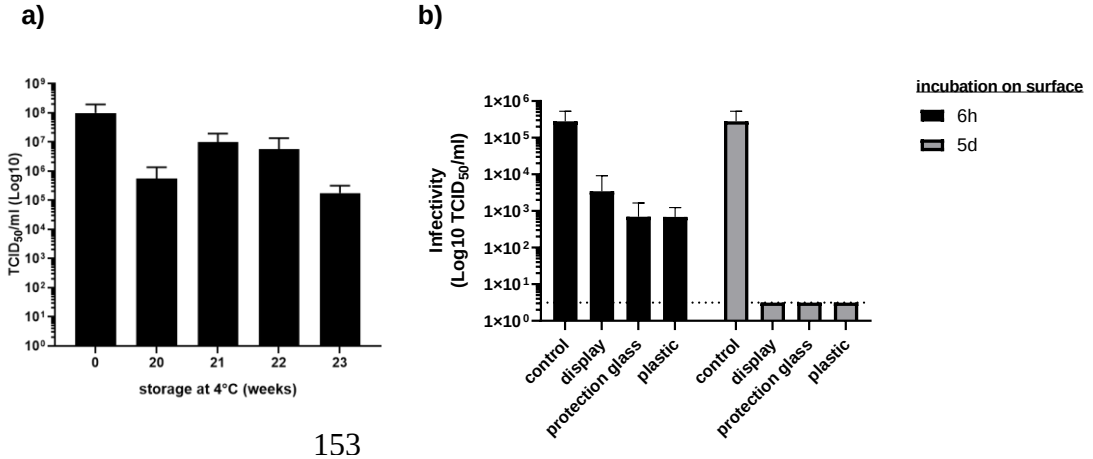

Figure 1 – Stability of SARS-CoV-2 under different storage conditions. a) SARS-CoV-2 virus stocks were stored in cell culture medium at 4°C for the indicated period of time. b) Stability of SARS-CoV-2 on surfaces (with and without protection glass).$TCID_{50}$ using CaCo2-cells was performed to determine viral infectivity.

Since coronaviruses were shown to remain infectious on commonly used touch surface materials (Warnes, Little, and Keevil, 2015), we next examined how long SARS-CoV-2 might be stable on standard laboratory surfaces. For this purpose, we coated plastic as well as touchscreen displays of mobile phones with and without protective glass with SARS-CoV-2 (2.8×10^5 TCID50/mL). After a short and long incubation time at room temperature corresponding to one working day (6 h) and one working week (5 days), the surfaces (2 cm^2) were wiped off with a PBS soaked swab and the viruses were eluted in culture medium and tested quantitatively for infectivity using $TCID_{50}$. After 6 hours we saw a decrease in $TCID_{50}$ of 1.91 log on the display and of approx. 2.6 log for displays with a protective glass and for plastic surfaces (***Figure 1b, and Table 1***). After 5 days we were unable to recover infectious virus particles on any of the surfaces indicating a minimal reduction factor of 4.95 log.

These data indicate that it is particularly important to inactivate contaminated material and instruments before continue working under less stringent safety requirements.

Table 1 – Stability of SARS-CoV-2 on surfaces in laboratory environment. Common laboratory surfaces plastic ware and touchscreen displays with and without protection glass were used. Detection limit was 3.2×10^0 $TCID_{50}$/mL. 4 log_{10} reduction was considered as antiviral activity.

Incubation time	Surface	Virus load ($TCID_{50}$/mL)	Minimal reduction factor (log10)
Viral Control		2.8×10^5	0.00
6 hours	display	3.4×10^4	1.91
	protection glass	7.0×10^2	2.60
	plastic	6.8×10^2	2.61
5 days	display	3.2×10^0	4.95
	protection glass	3.2×10^0	4.95
	plastic	3.2×10^0	4.95

3.2 Effectiveness of lysis buffers at inactivating SARS-CoV-2

Molecular biological detection methods require specific lysis methods to ensure subsequent partly enzymatic reactions. Buffers containing guanidine thiocyanate have proven useful for the isolation of viral nucleic acids and PCR-based analysis. We have therefore tested a set of 6 common lysis buffers for nucleic acid extraction whereof three are also used in routine diagnostic. Buffers AL, ATL and AVL (Qiagen, Hilden, Germany) and the cobas omni buffer (Roche, Germany) are commonly used to isolate nucleic acids from cell culture supernatants and patient material such as serum, blood, plasma, but also throat swab samples. The buffer RLT (Qiagen), however, is mainly used for research purpose for the isolation of total cellular RNA from infected cells. SARS-CoV-2 containing cell culture supernatants were mixed 1:1 with lysis buffers and diluted after 10 minutes incubation time. To evaluate the remaining infectivity viral outgrowth were performed in Caco2 cells and after 48 h cell culture supernatants were harvested for crystal violet staining and RT-qPCR analysis. To control that the readout was not affected by cell toxicity issues, in parallel we performed a cell viability assay (***Figure 2a***). In order to monitor virus production, we carried out an RT-qPCR targeting the SARS-CoV-2 M-gene as described previously (Toptan et al., 2020). As shown in ***Figure 2*** all tested buffers containing guanidine isothiocyanate including the cobas omni buffer (Roche) were able to completely inactivate samples containing SARS-CoV-2 as determined by crystal violet staining and intracellular SARS-CoV-2 RNA RT-qPCR (***Table 2***).

a) **b)**

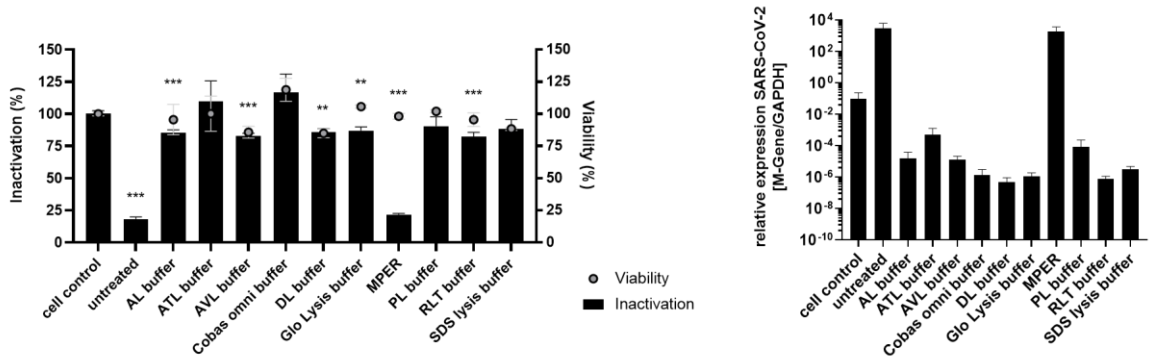

Figure 2 – Inactivation of SARS-CoV-2 by lysis buffers commonly used in molecular biology laboratories. a) For the Rotitest Vital viability assay. Cells were incubated with a mixture of cell culture medium and the depicted lysis buffer in the given concentration used for virus inactivation (1/1000). Cell viability was determined by commercial RotiTest Vital Assay measuring OD_{450}. For the untreated control no viability assay was performed to avoid handling with infectious samples outside the safety cabinet. Crystal violet staining. Samples were diluted 1/1000 to avoid cell toxicity. b) RT-qPCR analysis of RNA obtained from SARS-CoV-2 infected cells. Relative expression of SARS-CoV-2 RNA targeting M-Gene. Values were normalized to GAPDH expression. Error bars indicate SD from the mean of representative experiment performed in triplicates.

The IGEPAL CA-630 (MP Biomedicals Germany GmbH, Eschwege, Germany) containing DL buffer was developed in order to avoid expensive and time-consuming RNA extraction procedure while concomitantly inactivating infectious samples prior to PCR analysis. DL puffer was found to efficiently inactivate SARS-CoV-2 (***Table 2***; **Supplementary Figure 1**). Next, we also tested lysis buffers that are commonly used to analyze protein samples and used an SDS-containing buffer for Western blot

analysis, and Triton-X containing buffers, which are commonly used for luciferase-based assay. The latter have the critical property of maintaining protein function in order to allow enzymatic reactions with light emission. The buffers used therefore have a membrane-permeabilizing, but incompletely denaturing effect on cells. For SDS-containing lysis buffer and the commercially available Glo-Lysis buffer (Promega, Walldorf, Germany) as well as the PL buffer (lysis juice; p.j.k., Kleinblittersdorf, Germany) we were able to demonstrate complete inactivation (***Table 2***, **Supplementary Figure 1**). Importantly, Mammalian Protein Extraction Reagent (M-PER) (ThermoFisher) was not able to inactivate SARS-CoV-2 (**Figure 2**, ***Table 2***). As determined using crystal violet staining, only 22% of the cells were inactivated, which was comparable to the untreated control (18%). This data was confirmed by RT-qPCR since the relative SARS-CoV-2 M gene expression for M-PER treated samples (1.79×10^3) was as high as the untreated control (2.89×10^3).

These findings highlight the fact that careful evaluation of the used inactivation methods are required and that additional inactivation steps might be necessary to prepare the samples for safe processing.

Table 2 Inactivation of SARS-CoV-2 by common lysis buffers*.* Percent inactivation relative to OD_{570} in Crystal Violet staining, normalized to cell control.

Lysis buffer	Inactivation (%) (crystal violet staining)	Relative expression M-Gene (RT-qPCR)
cell control	n. d.	9.59×10^{-2}
untreated	18	2.89×10^{3}
AL buffer	86	1.54×10^{-5}
ATL buffer	109	4.97×10^{-4}
AVL buffer	83	1.22×10^{-5}
Cobas omni buffer	116	1.29×10^{-6}
DL buffer	86	4.83×10^{-7}
Glo Lysis buffer	87	1.13×10^{-6}
MPER	22	1.79×10^{3}
PL buffer	91	8.37×10^{-5}
RLT buffer	82	7.81×10^{-7}
SDS lysis buffer	88	2.99×10^{-6}

3.3 Physical Inactivation effectively eliminates infectious SARS-CoV-2

Since not all lysis buffers are able to completely inactivate SARS-CoV-2, we have tested further physical and chemical inactivation methods. First, we evaluated the effects of heat inactivation on SARS-CoV-2 infectivity and heated cell culture supernatants at different temperatures with different incubation times. Subsequently, the supernatants were used to infect Caco2 cells and after 48 h cell culture supernatants were harvested and subjected to RT-qPCR to evaluate viral outgrowth (***Table 3***). We found that heat inactivation using 56°C for 5 min was not effective (0.15 log), while increasing the incubation time to

30 min was a highly effective method of inactivation (4.35 log). Already 5 minutes at 60°C drastically reduced the infectivity of the virus suspension by 4.35 log. Short incubation time of 1 min at 90°C also achieved a very effective 4.28 log reduction in SARS-CoV-2 viral load.

Table 3 Heat inactivation, log reduction. Virus load was determined via RT-qPCR targeting SARS-CoV-2 M gene with a detection limit of 2x 10^3 copies/reaction. 4 $\log_{10}$ reduction was considered as antiviral activity.

Temperature (°C)	Incubation time (min)	Virus load (copies/reaction)	Minimal reduction factor (log10)
Viral Control		6.15×10^7	0.00
56	5	4.31×10^7	0.15
	30	2.74×10^3	4.35
60	5	3.48×10^3	4.25
	30	6.23×10^3	3.99
90	1	3.22×10^3	4.28
	5	1.19×10^4	3.71

Next, we evaluated chemical disinfectant and fixation solutions and treated virus supernatants in cell culture media in 1:10 dilution to the testing compound for 10 minutes at room temperature. acetone/methanol (40:60), ethanol (70%), and paraformaldehyde (PFA, 3%) were found suitable to completely inactivate the virus (**Table 4**). However, ethanol/PBS (50%, 1:1) did not inactivate SARS-CoV-2, which was in line with previously published data (Kratzel et al., 2020b).

Table 4 **Fixation of SARS-CoV-2**. Virus load was determined via RT-qPCR targeting SARS-CoV-2 M gene with a detection limit of 2x 10^3 copies/reaction. 4 $\log_{10}$ reduction was considered as antiviral activity.

Fixation solution	Virus load (copies/reaction)	Minimal reduction factor (Log10)
Viral Control	4.29×10^7	0.00
Acetone/Methanol (40:60)	1.77×10^3	4.38
70% Ethanol	4.12×10^3	4.02
50% Ethanol/PBS (1:1)	4.95×10^7	0.00
3% Paraformaldehyde	6.62×10^3	3.81

Finally, we investigated the influence of UV light on the stability of SARS-CoV-2. For comparison, we included two different strains of SARS-CoV (from 2003) and also H1N1 Influenza A virus (IAV) as a representative of a seasonally recurring respiratory pathogen. To mimic daily laboratory decontamination routines, we exposed SARS-CoV-2 contaminated samples in a biological safety

cabinet to UV light and irradiated for 15- 60 min. We found that 15 min were already highly efficient and 30 min as well as 60 min completely inactivated SARS-CoV-2 infectivity. In addition, we used two different UV light sources with a discharge lamp (low pressure radiation chamber) or a highly potent LED UV light to inactivate contaminated surfaces. We compared different exposure times ranging from 2 – 210 seconds for UV-C discharge lamp and 0.5 -3.5 seconds for UV-C LED with fixed distances of 20 to 40 mm to the light source, respectively. Using UV light from the discharge lamp, 2 seconds exposure time resulted in a minimal reduction factor of 3.5 log while after 4 seconds no infectious SARS-CoV-2 was detected in the viral outgrowth assay (***Figure 3***, **Table 5**). One second UV-C LED was sufficient to reduce viral infectivity by 3.9 log. 3.5 seconds completely reduced viral titers >4 log.

Table 5 - Inactivation of SARS-CoV-2 by UV-irradiation. Quantification of SARS-CoV-2 using $TCID_{50}$ method. After exposure to UV-C light for the indicated exposure time SARS-CoV-2 was inoculated on Caco2 cells. Detection limit: 3.2×10^0 $TCID_{50}$/mL. 4 log_{10} reduction was considered as antiviral activity.

Light source	Exposure time (s)	Virus load ($TCID_{50}$/mL)	Minimal reduction factor (Log10)
UV-C discharge lamp	Viral control	3.2×10^4	-
	2	1.0×10^1	3.5
	4	3.2×10^0	4.0
	15	3.2×10^0	4.0
	35	3.2×10^0	4.0
	210	3.2×10^0	4.0
UV-C LED	Viral control	3.2×10^4	-
	0.5 sec	1×10^1	1.7
	1.0 sec	3.2×10^0	3.9
	3.5 sec	3.2×10^0	4.0

These data demonstrate that UV-light exposure is a suitable method for complete inactivation of SARS-CoV-2 contaminated material. In particular, irradiation with UV-C LED light might also represent a fast and reliable inactivation method for contaminated material and surfaces.

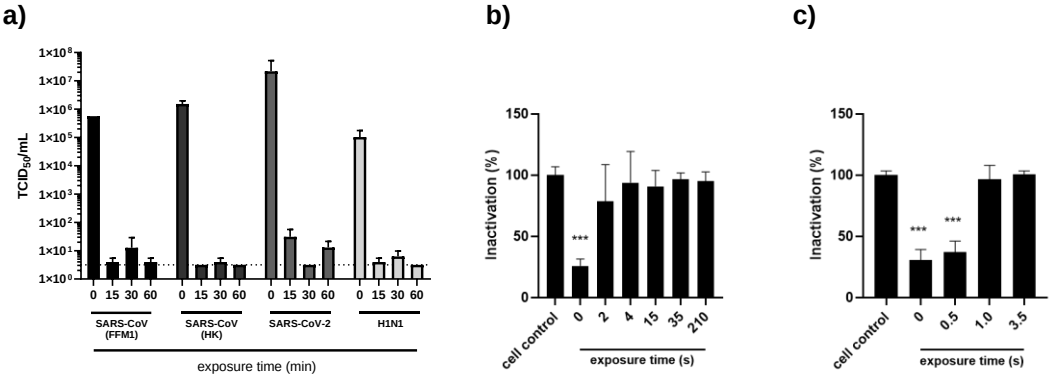

Figure 3 – Inactivation of SARS-CoV-2 using UV light. **a)** Quantification of SARS-CoV-2 using $TCID_{50}$. After exposure to UV light in the safety cabinet for the indicated exposure time SARS-CoV-2 was used to infect CaCo2 cells. SARS-CoV-strains FFM1 (Drosten et al., 2003) and Hongkong (HK), SARS-CoV-2 strain FFM1 (Hoehl et al., 2020; Toptan et al., 2020), H1N1 (PR8). **b) and c)** Cristal violet staining of CaCo2 cells infected with UV-irradiated SARS-CoV-2. UV-C discharge lamp and UV-C LED were used as light source for the indicated exposure time. s, seconds. Error bars indicate SD from the mean of three independent experiments.

4. Discussion:

Enveloped viruses are less resistant to chemical and physical noxes and other environmental influences compared to non-enveloped viruses (Geller, Varbanov, and Duval, 2012; Kramer, Schwebke, and Kampf, 2006). Depending on air temperature and relative humidity (Casanova et al., 2010) coronaviruses are stable for up to 9 days on inanimate surfaces like metal, glass or plastic (Kampf et al., 2020b; van Doremalen, Bushmaker, and Munster, 2013).

In this study mid-term and long term storages revealed a high stability of the virus during liquid storage over several weeks/months. We have found that even after months of storage under laboratory conditions at 4°C, the virus has hardly lost any of its infectivity. This suggests that laboratories must be absolutely certain that their inactivation methods used for subsequent research purposes will also provide sufficient inactivation. SARS-CoV-2 can be efficiently inactivated by surface disinfection procedures (Kampf et al., 2020b), however, several research methods in molecular biology require functional proteins for enzymatically catalyzed reactions or for the maintenance of secondary and tertiary structures. In this manuscript, we tested several lysis buffers and other chemical and physical inactivation methods that would allow downstream analysis of SARS-CoV-2 infection outside of the BSL3 laboratory. Among several lysis buffers we found the proprietary detergent M-PER lysis buffer (Thermo Fisher) to fail in the ability to inactivate SARS-CoV-2.

In agreement with previously studies we found that most inactivation methods were effective. In particular, our results were in line with previous studies that showed high temperature (Kratzel et al., 2020a), alcohols (Kratzel et al., 2020b), UV irradiation (Duan et al., 2003), and lysis by detergents are effective in inactivate corona virus (Rabenau et al., 2005a; Rabenau et al., 2005b). In line with previously published data on SARS-CoV (Rabenau et al., 2005a), fixation with PBS/ethanol (EtOH 50%; 1:1) over 5 min was not able to eliminate infectivity.

Of note, inactivation efficiency largely depends on the initial virus load and the reaction volume. Especially in the case of heat inactivation, the heating time of the respective medium must be considered. In addition, the higher the virus titer, the longer the inactivation time is necessary for

complete inactivation. UV-irradiation of infected cell culture samples was effective but varied between methods. The usual radiation takes place within the safety cabinets after work. Here, 15 minutes were already highly effective to eliminate infectious virus. In most laboratories, an exposure time of 30 minutes is suggested after infectious work. Thus, after correct use, inactivation of SARS-CoV-2 can be assumed, however, the physical lifespan of the light source must not be exceeded in order to generate sufficient emission. For fast inactivation, we evaluated two different UV-C light sources and found that UV-C discharge lamp with a radiation duration of 4 seconds was sufficient to inactivate SARS-CoV-2. The use of the high-potency UV-C LED light already results after 1 seconds. Although, all tested UV light source were UV-C rays the distance between light source and sample plays a critical role to define irradiation time. Furthermore, it has to take in account that lifespan of conventional UV lamps used in safety cabinets ranges between 6000 to 8000 hours. Therefore, intensity and effectiveness of the UV light is decreasing over time and when reaching maximum of expected useful life. Irradiation with UV-C LEDs thus represents a highly effective inactivation method for contaminated surfaces.

In comparison to the Triton-X-containing lysis buffers used for enzyme based assays (e.g. luciferase based measurements), we found that the commercially available M-PER buffer (Thermo Scientific) was not sufficient to inactivate SARS-CoV-2. M-PER is a proprietary detergent in 25 mM bicine buffer (pH 7.6) used for enzyme based reporter protein assays and immunoassays as well as protein purification. While bicine is an organic compound used as a buffering agent the exact composition of M-PER is not publicly known. However, according to the manufacturer, M-PER reagent has been tested on different cell lines showing complete lysis of adherent cells. Even if the manufacturers describe an efficient and fast cell lysis, the virus was able to withstand the mild detergent in our experiments.

In conclusion, inactivation methods including alcohols (except PBS/EtOH 1:1), acetone-methanol mixture, PFA, UV-light and heat inactivation efficiently inactivated SARS-CoV-2. Most but not all research laboratory used lysis buffers were suitable to inactivate SARS-CoV-2. However, since M-PER was not able to inactivate SARS-CoV-2, research buffers, especially if they are non-denaturating, must be carefully evaluated to protect laboratory personnel and must be substituted to maintain laboratory safety.

5. Acknowledgements

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6. References:

Casanova, L.M., Jeon, S., Rutala, W.A., Weber, D.J. and Sobsey, M.D., 2010. Effects of air temperature and relative humidity on coronavirus survival on surfaces. Applied and environmental microbiology 76, 2712-7.

Drosten, C., Gunther, S., Preiser, W., van der Werf, S., Brodt, H.R., Becker, S., Rabenau, H., Panning, M., Kolesnikova, L., Fouchier, R.A., Berger, A., Burguiere, A.M., Cinatl, J.,

Eickmann, M., Escriou, N., Grywna, K., Kramme, S., Manuguerra, J.C., Muller, S., Rickerts, V., Sturmer, M., Vieth, S., Klenk, H.D., Osterhaus, A.D., Schmitz, H. and Doerr, H.W., 2003. Identification of a novel coronavirus in patients with severe acute respiratory syndrome. The New England journal of medicine 348, 1967-76.

Duan, S.M., Zhao, X.S., Wen, R.F., Huang, J.J., Pi, G.H., Zhang, S.X., Han, J., Bi, S.L., Ruan, L., Dong, X.P. and Team, S.R., 2003. Stability of SARS coronavirus in human specimens and environment and its sensitivity to heating and UV irradiation. Biomedical and environmental sciences : BES 16, 246-55.

Geller, C., Varbanov, M. and Duval, R.E., 2012. Human coronaviruses: insights into environmental resistance and its influence on the development of new antiseptic strategies. Viruses 4, 3044-68.

Hoehl, S., Rabenau, H., Berger, A., Kortenbusch, M., Cinatl, J., Bojkova, D., Behrens, P., Boddinghaus, B., Gotsch, U., Naujoks, F., Neumann, P., Schork, J., Tiarks-Jungk, P., Walczok, A., Eickmann, M., Vehreschild, M., Kann, G., Wolf, T., Gottschalk, R. and Ciesek, S., 2020. Evidence of SARS-CoV-2 Infection in Returning Travelers from Wuhan, China. The New England journal of medicine.

Kampf, G., Todt, D., Pfaender, S. and Steinmann, E., 2020a. Corrigendum to "Persistence of coronaviruses on inanimate surfaces and their inactivation with biocidal agents" [J Hosp Infect 104 (2020) 246-251]. The Journal of hospital infection.

Kampf, G., Todt, D., Pfaender, S. and Steinmann, E., 2020b. Persistence of coronaviruses on inanimate surfaces and their inactivation with biocidal agents. The Journal of hospital infection 104, 246-251.

Kärber, G., 1931. Beitrag zur kollektiven Behandlung pharmakologischer Reihenversuche. Naunyn-Schmiedebergs Archiv für experimentelle pathologie und pharmakologie 162, 480-483.

Kramer, A., Schwebke, I. and Kampf, G., 2006. How long do nosocomial pathogens persist on inanimate surfaces? A systematic review. BMC infectious diseases 6, 130.

Kratzel, A., Steiner, S., Todt, D., V'Kovski, P., Brueggemann, Y., Steinmann, J., Steinmann, E., Thiel, V. and Pfaender, S., 2020a. Temperature-dependent surface stability of SARS-CoV-2. The Journal of infection.

Kratzel, A., Todt, D., V'Kovski, P., Steiner, S., Gultom, M., Thao, T.T.N., Ebert, N., Holwerda, M., Steinmann, J., Niemeyer, D., Dijkman, R., Kampf, G., Drosten, C., Steinmann, E., Thiel, V. and Pfaender, S., 2020b. Inactivation of Severe Acute Respiratory Syndrome Coronavirus 2 by WHO-Recommended Hand Rub Formulations and Alcohols. Emerging infectious diseases 26, 1592-1595.

Rabenau, H.F., Cinatl, J., Morgenstern, B., Bauer, G., Preiser, W. and Doerr, H.W., 2005a. Stability and inactivation of SARS coronavirus. Medical microbiology and immunology 194, 1-6.

Rabenau, H.F., Kampf, G., Cinatl, J. and Doerr, H.W., 2005b. Efficacy of various disinfectants against SARS coronavirus. The Journal of hospital infection 61, 107-11.

Toptan, T., Hoehl, S., Westhaus, S., Bojkova, D., Berger, A., Rotter, B., Hoffmeier, K., Cinatl, J., Jr., Ciesek, S. and Widera, M., 2020. Optimized qRT-PCR Approach for the Detection of Intra- and Extra-Cellular SARS-CoV-2 RNAs. International journal of molecular sciences 21.

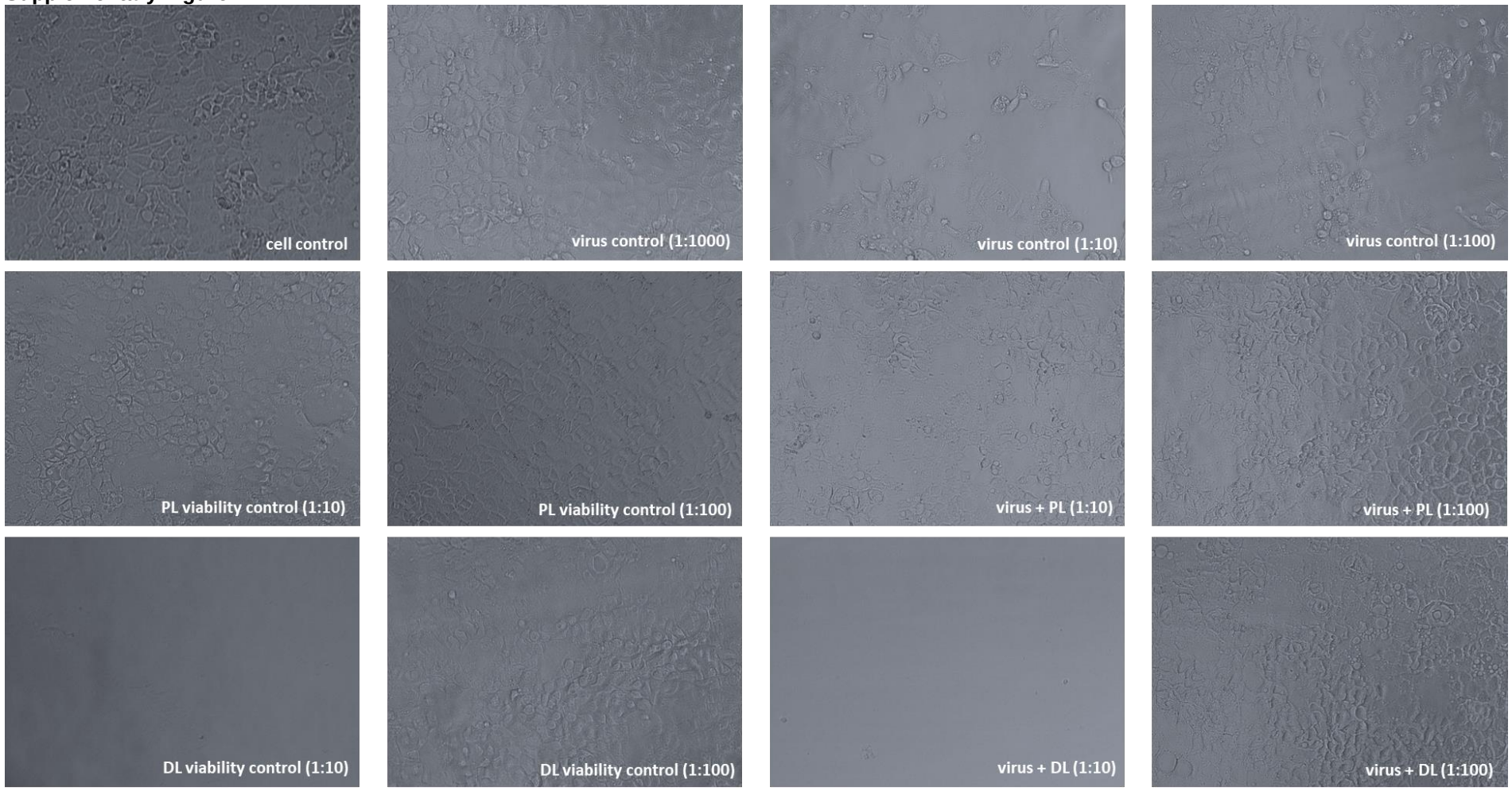
van Doremalen, N., Bushmaker, T. and Munster, V.J., 2013. Stability of Middle East respiratory syndrome coronavirus (MERS-CoV) under different environmental conditions. Eurosurveillance 18, 20590.

Warnes, S.L., Little, Z.R. and Keevil, C.W., 2015. Human Coronavirus 229E Remains Infectious on Common Touch Surface Materials. mBio 6, e01697-15.

WHO. 2020. WHO SARS-CoV-2 situation report 182, Report on coronavirus disease 2019 (COVID-19), WHO.

Zhu, N., Zhang, D., Wang, W., Li, X., Yang, B., Song, J., Zhao, X., Huang, B., Shi, W., Lu, R., Niu, P., Zhan, F., Ma, X., Wang, D., Xu, W., Wu, G., Gao, G.F., Tan, W., China Novel Coronavirus, I. and Research, T., 2020. A Novel Coronavirus from Patients with Pneumonia in China, 2019. The New England journal of medicine 382, 727-733.

Supplementary Figure 1:



Supplementary Figure 1 - SARS-CoV-2 inactivation with Triton-containing lysis buffer (PL) and IGEPAL-630 containing buffer (DL). Crystal violet staining of CaCo2 cells infected with SARS-CoV-2 previously inactivated with the indicated buffer. Virus and lysis buffer (1:1) were incubated for 15 min at ambient temperature. Samples were diluted as indicated in brackets to avoid cell toxicity