A sexual dimorphism in the spatial vision of band-winged grasshoppers

Alex B. Duncan[1], Brae A. Salazar[1], Sara R. Garcia[2], and Nicholas C. Brandley[2,1,*]

1: Colorado College, Department of Organismal Biology & Ecology, 14 E, W Cache La Poudre
St., Colorado Springs CO 80903, USA

2: College of Wooster, Department of Biology, 1189 Beall Ave, Wooster OH 44691, USA

*: corresponding author: nbrandley@wooster.edu

Abstract

Visual acuity (VA) --- a measurement of the fineness or coarseness of vision ---

correlates with the size of an animal, with larger species often possessing sharper vision.

However, it is unknown whether the same relationship between visual acuity and size holds

within a species when individuals differ consistently and substantially in size, such as through a

sexual size dimorphism. Here we examine the visual acuity of three species of sexually

dimorphic band-winged grasshoppers, in which females are the larger sex (*Arphia*

pseudonietana, *Dissosteira carolina*, and *Spharagemon equale;* total n = 98). Using a radius of

curvature estimation method, we find that females have ~21% finer vision in the most acute

region and axis of the eye than do males. Further explorations of the eyes of the species

showing the greatest size dimorphism (*D. carolina*) suggest that this VA dimorphism is driven by

females having larger eyes with more ommatidia. In contrast to many flying insects where males

have finer vision to acquire mates, our study is one of the first to demonstrate a female-biased

sexual dimorphism in acuity. Given the number of species in which females are larger than

males, our results suggest that differences in VA between the sexes may be more common than

currently appreciated.

1. INTRODUCTION

Animal behavior is driven by what information they can perceive, which itself is limited by their sensory systems (Partan and Marler, 2002; Uexküll, 2013). Therefore, studying how an animal responds to stimuli requires understanding what information it can perceive (Caves et al., 2019; Jordan and Ryan, 2015). Notably, animals differ not just in the presence or absence of senses, but in the fineness with which they can parse information within a sensory modality. For example visual acuity (VA) --- defined as the ability to perceive static spatial detail and used as a measurement of the fineness or coarseness of vision --- varies by up to four orders of magnitude between species (Caves et al., 2018; Land and Nilsson, 2012), making it a promising parameter for further exploration of within sense variation.

Between species, VA changes with both the ecology and morphology of a given species. Ecologically, VA is finer in predatory animals (Veilleux and Kirk, 2014) and those in high-photon environments (Caves et al., 2017; Veilleux and Kirk, 2014). Morphologically, larger eyes (and through correlations between eye and body size, larger bodies) are typically associated with finer vision regardless of taxonomic level, with studies supporting this at the kingdom (Caves et al., 2018; Land and Nilsson, 2012), superclass/class (Caves et al., 2017; Kiltie, 2000; Veilleux and Kirk, 2014), order (Rutowski et al., 2009), and family (Jander and Jander, 2002) levels.

Despite numerous examples of VA varying with size between species, surprisingly fewer studies have examined whether the same principles hold within a species (although see Corral-López et al., 2017; Spaethe and Chittka, 2003). VA --- like other aspects of vision --- can vary within a species in other contexts. For example, numerous within-species VA studies have focused on mate pursuit in flying insects. In these cases, males have finer vision in specialized regions designed to spot potential mates (Kirschfeld and Wenk, 1976; Land and Eckert, 1985), which is accompanied by neurological and behavioral specializations. However, we know little about within-species variation in VA outside of this context. Another way VA may vary within a species is through a sexual size dimorphism. Size dimorphisms may have large effects on

vision in small animals with compound type eyes (such as insects). Unlike camera eyes that

have a single larger lens, compound eyes have numerous smaller lenses for each of their

ommatidia. As a result, the small overall size of compound eyes and their numerous lenses may

leave them diffraction-limited (Barlow, 1952; Kirschfeld, 1976; Land, 1997). Thus increases in

eye size often improve optical performance (Jander and Jander, 2002; Rutowski et al., 2009).

Based on both 1) the interspecific correlation between size and VA and 2) the optical

limitations outlined above, one could predict that sexual size dimorphisms may be accompanied

by the larger sex having finer vision. However, this may not always be the case, as larger eyes

can optimize other visual parameters (such as sensitivity) instead of VA. Additionally, eye size

may not scale with body size within a species. An ideal animal group to examine whether size

dimorphisms may be accompanied by VA dimorphisms are the band-winged grasshoppers.

Band-winged grasshoppers (subfamily Oedipodinae) are a morphologically diverse subfamily of

~200 diurnal species known for their colorful hindwing patterns (Otte, 1970; Otte, 1984). Many

species of band-winged grasshoppers show a size dimorphism; however, little is known about

their vision. Here, we first examine VA in the most acute region of the eye across three species

of band-winged grasshopper--- *Arphia pseudonietana* (Thomas, 1870)*, Dissosteira carolina*

(Linnaeus*, 1758) and *Spharagemon equale* (Say, 1825) --- to determine if the larger females

have finer vision. In a second experiment, we then further examine eye and receptor scaling in

the species that showed the greatest size dimorphism (*D. carolina*) to better understand whole

eye changes between the sexes.

72   **2. MATERIALS & METHODS**

2.1 Experiment I: VA variation by biological sex across three species

2.1.1 Study Organisms

To examine how biological sex influences visual acuity in band-winged grasshoppers,

individuals of three species were collected during the summer and early fall (June-October) of

2016 and 2017 in Colorado Springs, CO. *Dissosteira carolina* (n = 16 males and 16 females)

were collected from an urban site near manicured grass, while *Arphia pseudonietana* (n = 18

males and 18 females), and *Spharagemon equale* (n = 15 males and 15 females) were

collected at a high-altitude grassland site. Grasshoppers were euthanized using ethyl acetate

and then stored at ~ -20°C prior to imaging. Total length (head to the tip of the forewings; *mm*),

weight (g), and biological sex were recorded for each individual.

2.1.2 Estimating Visual Acuity

We measured visual acuity (VA) of all three species via a modified radius of curvature

estimation (Bergman and Rutowski, 2016). Briefly, this method estimates VA by calculating how

many ommatidia view a given angle of visual space.

To calculate VA, eyes were imaged at 40X magnification using AmScopeX for Mac MU

(MW Series 05/26/2016; United Scope LLC; Irvine, CA, USA) under diffuse lighting conditions

(LED312DS; Fotodiox Inc. USA; Gurnee, IL, USA) with a M28Z zoom stereo binocular

microscope (Swift Optical Instruments Inc.; Carlsbad, CA, USA) and AmScope 14MP USB3.0

digital camera (United Scope LLC; Irvine, CA, USA). Because grasshopper eyes are not

spherical and therefore have different curvatures in each axis, we used a lateral image to

measure curvature ---and ultimately VA--- in the axis perpendicular to the horizon (y axis), a

dorsal view for the axis parallel to the horizon (x axis), and one anterior view of the flattest

region of the eye to measure facet diameter (Figure 1). For consistent positioning, physical

attributes were used for orientation in each image (lateral image = the inside eye edge, dorsal

image = the top of the eye, anterior image = the center of the eye).

Using the lateral (for y curvature) or dorsal views (for x curvature), the localized

curvature of the eye ($b\,/\,\alpha$) was calculated in two axes via ImageJ (v. 1.50i; Schneider et al.

2012). First, we drew two lines perpendicular to the surface of the eye (Figure 1a*;* Bergman and

Rutowski 2016). From these lines, *b* was calculated as the distance between the two points

created by the intersection of the perpendicular lines to the eye edge, while the angular distance

covered between these points (*α*) was calculated using the ImageJ angle function. These

measurements were then combined (*b* / *α)* to calculate the distance of the eye's surface

covered in a given angle (*µm* per °).

Facet diameter (*D*) was calculated from two rows of ten facets in the flattest region of the

eye (Figure 1B). Similar to studies in other Oedipodinae (Horridge, 1978), preliminary results

found that facet diameter was relatively constant across the majority of the eye surface of each

individual grasshopper.

Using the previous measurements, the inter-ommatidial angle (∆Φ) in each axis was

then calculated as:

113                    $\Delta\Phi = D / (b / \alpha)$ (1)

114   Where *D* = the facet diameter and *b / α* = eye surface length in a given angle.

115   Finally, visual acuity (VA, in degrees, smaller values indicate finer vision) in each axis

116   was calculated as two times the inter-ommatidial angle:

117                    $VA = 2 * \Delta\Phi$ (2)

2.1.3 Statistical Analysis

All data was analyzed in R version 3.4.4 (R CORE Team, 2018). To quantify the

magnitude of the sexual size dimorphism, we first used a two-way ANOVA examining how total

length varies with species and biological sex.

VA data was analyzed via two different approaches that produced similar results. First,

we analyzed how VA in each axis was predicted by a variety of factors using generalized linear

models (GLMs). We used the Akaike information criterion (AIC) to select the most parsimonious

model(s). We treated models within 2 AIC as equally parsimonious. Second, because our GLM

results suggested that biological sex and species were the main predictors of VA, we used two-

way ANOVAs to examine how biological sex and species predict VA and its morphological

determinants (eye curvature and facet size). Normality for all data was first checked using the

Shapiro-Wilk normality test. In two cases data was either natural log transformed (VA_x) or had

an outlier excluded (vertical eye curvature) to restore normality. Student's T-tests or Tukey

HSDs were used for post-hoc analysis.

Figure 1: An example of the radius of curvature estimation used to estimate visual acuity

in band-winged grasshoppers. A lateral view of the eye with the edge in focus. Lines tangent

to the curvature of the eye (red and blue) have been drawn for the flattest region of the eye and

are used to calculate the curvature of the eye in the relevant axis (see methods and Bergman

and Rutowski 2016 for more details). B) an anterior view of the flattest region of the eye used to

calculate average facet diameter. b / α = eye surface length (*b*) in a given angle (*α*), D = facet

diameter.

2.2 Experiment II: Eye size and facet count variation in Dissosteira carolina

2.2.1 Study Organisms

To further explore the relationship between body size, eye size, and VA, we examined eye and

ommatidia scaling within the species that showed the greatest body size dimorphism (_D._

carolina). Unfortunately logistical issues prevented us from returning to the same study

population, so 48 _D. carolina_ (n = 25 male and 23 female) were collected during the summer

(June-September) of 2018 from a suburban site near manicured grass in Wooster, OH. Directly

prior to imaging, grasshoppers were euthanized in a freezer for ~1 hour. Weight (g), body length

(head to tip of the thorax; _mm_), head size (_mm_), and biological sex were recorded for each

individual.

2.2.2 Eye size measurements in Dissosteira carolina

All individuals (25 males, 23 females) were photographed at a lower resolution (30x)

then the previous experiment to ensure that all eyes completely fit within the images. Eye

images were taken using an AmScope stereo trinocular microscope (United Scope LLC; Irvine,

CA, USA) paired with a MU 1403 digital camera (United Scope LLC; MU1403; Irvine, CA, USA).

Because band-winged grasshoppers have non-spherically shaped eyes, calibrated images of

each eye were taken from three different angles (anterior, dorsal, lateral) and used to calculate

eye size in all three axes (x, y, z) in ImageJ (v 1.8.0, Schneider et al. 2012). Note that this

methodology only estimates the exposed eye depth in the z axis.

2.2.3 Facet counts in Dissosteira carolina

To examine if sexes vary in number of receptors, we calculated the total number of facets per

eye via eye castings of a subsample of the Ohio _D. carolina_ population (e.g. Narendra et al.,

2013). Individuals representative of the various eye sizes within each biological sex were used

(n=3 of each sex). A single thin layer of #800 crystal clear nail polish (Sally Hansen Inc., New

York, NY, USA) was applied to the entirety of the left eye and dried for 30-50 minutes. When

dry, the eye castings were removed and cut horizontally and vertically into multiple sections with

a #11 surgical scalpel blade (Swann-morton LTD., Sheffield, UK). Each section was then

flattened between two microscope slides overnight before imaging at 30X magnification

(Supplemental Figure 1) using an AmScope stereo trinocular microscope (United Scope LLC;

Irvine, CA, USA) paired with a MU1403 digital camera (United Scope LLC; Irvine, CA, USA).

Each eye section was assigned a non-identifying name, and facets were then counted by

individuals blinded to original sex and grasshopper ID of the images. Sections originating from

the same eye were then summed to determine the total facet count for each eye.

2.2.4 Statistical Analysis

We examined how biological sex and eye axis vary via a two-way ANOVA in R (R CORE

Team, 2018). Student's T-tests or Tukey HSDs were used for post-hoc analysis.

181   **3. RESULTS**

3.1 Experiment I: VA variation by biological sex across three species

3.1.1 Dimorphism in body size

Biological sex, species, and their interaction all significantly predicted total length in

band-winged grasshoppers (Figure 2). Biological sexes significantly differed in total body length

($p < 0.001$, df=1, F=257.93, two-way ANOVA), with females being on average 6.2-7.0 mm longer

than their male counterparts ($p < 0.001$, TukeyHSD). Additionally, all three species significantly

differed from each other in total length ($p < 0.001$, df=2, F=482.81, two-way ANOVA; all

TukeyHSD $p < 0.001$). The interaction between biological sex and species on total body length

was also significant ($p < 0.001$, df=2, F=12.71), with *Dissosteira carolina* showing the largest

dimorphism.

Figure 2: Body length in three species of band-winged grasshoppers. All three species show a sexual size dimorphism, with biological females being longer than males (p<0.001, df=1, F=257.93, two-way ANOVA). Additionally, total length varies between species, with all three showing significant differences from each other (p<0.001 in all comparisons, TukeyHSD). There is also a significant interaction between biological sex and species on total body length (p<0.001, df=2, F=12.71). Sample sizes (from left to right) are 18, 18, 16, 16, 15, and 15 individuals respectively. Error bars indicate 95% CI, significance symbols are based on post-hoc analyses (Student's T-test for biological sex differences within a species, TukeyHSD for species differences).

3.1.2 Visual acuity perpendicular to the horizon (VA_y)

Visual acuity varied depending on the axis of view in all three species; amongst all

individuals measured (n=98) visual acuity perpendicular to the horizon (VA_y; average of 2.17°)

was twice as sharp as visual acuity parallel to the horizon (VA_x; average of 4.34°; Figure 3). As

such, results are presented separately for each axis of view.

The most parsimonious models of VA_y all included biological sex, species, and their

interactions as factors (Table 1). Culmulatively, models including both sex and species as

predictors accounted for 92% of relative model probability (w_i). Some equally parsimonious

models included size measurments as well, but these did not significanty improve the model

(Table 1). In the GLM including biological sex and species, males were estimated to have

~0.416 degrees coarser vision (Std error=0.066, t=6.275, p<0.0001, GLM).

Results from a two-way ANOVA mirrored those of the GLM (Figure 3A). VA_y was

dependent on both biological sex (p<0.001, df=1, F=41.04, two-way ANOVA) and species

(p<0.05, df=2, F=3.44, two-way ANOVA). Across all individuals measured, female VA_y (average

= 1.96°) was estimated to be 21% finer (p<0.001, 95% CI = 0.29°-0.55°, TukeyHSD) than male

VA_y (average = 2.38°; Figure 2A). Additionally, the large *D. carolina* had significantly finer VA_y

than *S. equale* (p<0.05, 95% CI= 0.002°-0.39°, TukeyHSD), however other species

comparisons did not vary significantly (all p<0.05, TukeyHSD, Figure 2A). The interaction

between sex and species on VA_y trended towards significance (p=0.055, df=2, F=2.994, two-

way ANOVA). Within any biological sex and species combination examined, size did not

correlate with VA_y (all r^2 < 0.2, all p>0.1; Supplemental Figure 2), suggesting that differences in

size may act between the sexes but not within.

3.1.3 Visual acuity parallel to the horizon (VA_x)

In contrast to VA_y, VA_x showed no sexual dimorphism nor species-specific differences.

VA_x was not normally distributed (p<0.01, W = 0.95755, Shapiro-Wilk normality test), and

therefore a natural log transformation was used to restore normality. Neither biological sex

(p=0.18, df=1, F=1.86, two-way ANOVA), species (p=0.07, df=2, F=2.73, two-way ANOVA), nor

their interactions (p=0.99, df=2, F=0.013, two-way ANOVA) were significant predictors of the

natural log of VA_x (Figure 3B).

Table 1. Summary of predictor combinations explaining visual acuity perpendicular to the horizon (VA_y)

Model	ΔAIC	l_i	w_i
VA_y ~Sex*Species + Length	**0**	**1.00**	**0.28**
VA_y ~Sex*Species	**0.1**	**0.95**	**0.27**
VA_y ~Sex*Species + Weight	**1.3**	**0.52**	**0.15**
VA_y ~Sex + Species + Length	2.2	0.33	0.09
VA_y ~Sex + Species	2.3	0.32	0.09
VA_y ~Sex + Species + Weight	3.7	0.16	0.04
VA_y ~Sex + Length	4.5	0.11	0.03
VA_y ~Sex + Weight	4.6	0.10	0.03
VA_y ~ Sex	5	0.08	0.02
VA_y ~ Weight	15.3	0.00	0.00
VA_y ~ Length	24.3	0.00	0.00
VA_y ~ Species	34.6	0.00	0.00

ΔAIC values were calculated relative to the best fit model. Models in bold are equally

parsimonious (within 2 ΔAIC). l_i = model's relative likelihood, w_i = model probability

Figure 3: Visual acuity in the two axes of vision in band-winged grasshoppers. Visual

acuity in band-winged grasshoppers is sexually dimorphic, but only in one axis. Units are

presented in degrees, so finer vision is indicated by smaller values. **A)** Biological females

typically have finer visual acuity than males in the axis perpendicular to the horizon ($p<0.001$,

df=1, F=41.04, two-way ANOVA). Additionally, species differ significantly from one another

($p<0.05$, df=2, F=3.44) with *D. carolina* having finer vision than *S. equal*e ($p<0.05$, TukeyHSD).

B) In contrast to A, visual acuity parallel to the horizon is both coarser and shows no significant

sexual dimorphism or species-specific effect (see text for details). **A-B)** Sample sizes (from left

to right) are 18, 18, 16, 16, 15, and 15 individuals respectively in each panel. Error bars indicate

95% CI, significance symbols are based on post-hoc analyses (Student'sT-test for biological sex

differences within a species, TukeyHSD for species differences).

3.1.4 Dimorphism in the Morphological Determinants of VA

At the morphological level, compound eyes can achieve finer VA if they have either 1) a

flatter curvature or 2) smaller facets. In female band-winged grasshoppers, changes in eye

curvature --- and not facet size --- led to finer VA_y (Figure 4). Vertical eye curvature was not

normally distributed (p<0.01, W=0.95, Shapiro-Wilk normality test) because of the presence of

one female *S. equale* outlier with a particularly flat eye (perpendicular eye curvature = 51

µm/degree). Removal of the outlier restored normality (p=0.18, W=0.98, Shapiro-Wilk normality

test). Even with this especially flat eye removed, females still have a vertical curvature that is

~18% flatter than their male counterparts (p<0.001, df=1, F=49.56, two-way ANOVA, Figure

4A). Additionally, there are species-specific differences in vertical eye curvature (p<0.001, df=2,

F=14.58, two-way ANOVA) with post-hoc testing revealing that the larger *D. carolina* has a

flatter vertical eye curvature than either of the two other species (p<0.001 in both cases,

TukeyHSD). The interactions between biological sex and species on vertical eye curvature were

trending towards significance (p=0.06, df=2, F=2.87), suggesting that the dimorphism in vertical

curvature may vary between species.

Similarly to VA_x and VA_y, differences in eye curvature between the sexes were less

pronounced in the horizontal axis than in the vertical axis (Figure 4B). Both biological sex

(p<0.01, df=1, F=8.65, two-way ANOVA) and species (p<0.01, df=2, F=4.85) significantly

affected horizontal eye curvature. However, female horizontal curvature was only ~6.5% flatter

than males. There was no significant effect of the interaction between biological sex and

species on horizontal eye curvature (p=0.78, df=2, F=0.252).

Females had slightly larger facet diameters than males (~0.9 µm larger or 2.8%, Figure

4C) across all three species measured (p<0.01, df=1, F=11.47, two-way ANOVA). Additionally,

all three species differed significantly in facet size (p<0.001, df=2, F=96.06, two-way ANOVA)

with the larger *D. carolina* having the largest facets (all p<0.05, Tukey HSD). There was no

significant effect of the interaction between biological sex and species on facet diameter

(p=0.52, df=2, F=0.651, two-way ANOVA). The combination of changes in receptor spacing and

facet size led to an eye parameter (Figure 4D) that varied significantly between both sex

(p<0.001, df=1, F=29.15, two-way ANOVA) and species (p<0.001, df=2, F=2.92, two-way

ANOVA). The interactions between biological sex and species on the eye parameter were

trending towards significance (p=0.059, df=2, F=2.918).

Figure 4: Morphological determinants of visual acuity in band-winged grasshoppers. Eye curvature is the biggest morphological driver of the sexually dimorphic visual acuity in these three species of band-winged grasshoppers. **A)** Biological females had ~18% flatter vertical eye curvature in the region of the eye with the sharpest visual acuity (p<0.001, df=1, F=49.6, two-way ANOVA). **B)** Although significant, the differences in horizontal curvature were not as pronounced, as female curvature was only ~6.5% flatter than males (p<0.01, df=1, F=8.65, two-way ANOVA). **C)** Female facet diameters were slightly larger (~2.8%) than male facet diameters (p<0.01, df=1, F=1.52, two-way ANOVA). **D)** The observed eye parameter values are typical of diurnal insects and varied significantly between both sexes (p<0.001, df=1, F=29.15, two-way ANOVA) and species (p<0.001, df=2, F=2.92, two-way ANOVA). **A-D)** Sample sizes (from left to right) in each panel are 17, 18, 16, 16, 15, and 15 individuals respectively. Error bars indicate 95% CI, significance symbols are based on post-hoc analyses (Student'sT-test for biological sex differences, TukeyHSD for species differences).

3.2 Experiment II: Eye size and facet count variation in Dissosteira carolina

Further investigation of the VA_y dimorphism in the species that showed the greatest sexual size dimorphism (*D. carolina*) found that females typically have larger eyes than males, but not in every axis (Figure 5). Compared to males, females had significantly larger eye diameters in the horizontal (X; ~13% increase) and vertical (Y, ~23% increase) axes, but did not vary in exposed eye depth (Z; $p<0.001$, df=1, F=28.90, two-way ANOVA; sex*interaction $p<0.001$, df=2,F=7.98). Additionally, both male and female eyes varied significantly in diameter between the axes ($p<0.001$, df=2, F=240.73).

As expected based on differences of eye size and facet diameter, facet counts of a subset of representative individuals suggest that the female eyes have more facets than males (Table 2). Although there was variability in the number of facets seen in each sex, representative females (average = 5565 facets, n=3) had eyes with ~19% more facets than males (4679 facets, n=3).

306
Figure 5: Eye size in the band-winged grasshopper *Dissosteira carolina*. Eye size is both

asymmetrical and varies between the sexes. Females have larger eyes in the X and Y axis, but

not in exposed eye depth (Z; p<0.001, df=1, F=28.90, two-way ANOVA). Exposed eye depth (Z)

only measures exposed eye surface and may be an underestimate of total size. Sample size

are 23 females and 25 males. Error bars indicate 95% CI, significance symbols are based on

post-hoc analyses (Student's T-test for biological sex differences, TukeyHSD for species

differences).

314
315

Table 2: Whole eye facet counts in representative *Dissosteira carolina* individuals

eye size percentile	female	male
~25th	4901 facets	4571 facets
~50th	5514 facets	4453 facets
~75th	6280 facets	5013 facets

4. Discussion

Most band-winged grasshoppers have non-spherical eyes featuring an elongated vertical axis that gives them a jellybean-like shape. Similar to other invertebrates with non-spherical eyes (Bagheri et al., 2020; Kelber and Somanathan, 2019) this leads to better visual acuity in the axis of elongation: in band-winged grasshoppers visual acuity in the vertical axis (VA_y) is around twice as fine as that in the horizontal axis (VA_x; Figure 3). VA_y appears to be enhanced by a particularly flat vertical region near the center of the eye.

VA_y in this particularly flat region of the eye is sexually dimorphic, resulting in female grasshoppers --- the larger of the two sexes --- having finer vision (Figure 3a). Similar to other grasshopper species (Hochkirch and Gröning, 2008; Otte, 1984), female band-winged grasshoppers were substantially larger than males in this study (Figure 2). Similarly VA_y --- but not VA_x --- was sexually dimorphic, with females having ~21% finer vision than males in the sharpest region of the eye (Figure 3a-b). This VA_y dimorphism is similar in magnitude to the classic "love spot" seen in male flies (although without the accompanying sensitivity specializations, (Land and Eckert, 1985)) however it is in the opposite direction: females have finer vision rather than males. This suggests that much like we observe between species (Caves et al., 2017; Caves et al., 2018; Kiltie, 2000; Land and Nilsson, 2012; Veilleux and Kirk, 2014), size differences can lead to finer vision within a species. Notably, within either sex there was no further effect of size on VA_y (Supplemental Figure 2). Thus, in some cases the positive relationship between acuity and size could be constrained to individuals with consistently different developmental processes such as sex and/or morph differences.

The magnitude of the VA_y dimorphism varied by species as all of the top predictor models included a term for the interaction between sex and species (Table 1). This was driven by *Arphia pseudonietana* showing a much smaller dimorphism than either of the two other species (Figure 6) that when considered by itself was only trending towards significantly varying between the sexes (Figure 3A). Notably, *A. pseudonietana* was one of the species that showed

a smaller size dimorphism (Figure 2), which could lead to a less pronounced VA dimorphism.

However, *Spharagemon equale* has a similar size dimorphism but showed a substantial VA_y

dimorphism suggesting that selective pressures and/or developmental constraints on band-

winged vision could be acting differently across species.

Across all three species examined, the dimorphism was morphologically caused by a

flatter eye surface and not a reduction in facet diameter (Figure 4). Although both changes can

lead to finer acuity, decreasing facet diameter is not typically seen in insects because it may

also reduce overall sensitivity (Kirschfeld, 1976; Land, 1997). The acute zones of many insects

require both fine VA and heightened sensitivity (Warrant, 2016), and therefore often feature flat

regions with large facets. In band-winged grasshoppers, females either had similar or slightly

larger facets than males. However, their eye's flatter curvature led to a substantial VA

dimorphism. As a result, female band-winged grasshoppers have finer VA_y without sacrificing

sensitivity by decreasing facet diameter (Figure 4).

Further study in *D. carolina* suggests that the sexual size dimorphism corresponds with

both an eye size dimorphism (Figure 5) and a facet number dimorphism (Table 1). Altogether,

this suggests that female *D. carolina* improve VA_y by having larger eyes with more facets.

Surprisingly, we also found that females had a larger eye diameter in the horizontal axis, despite

showing no improvement in VA_x compared to males. This increase in X eye diameter suggests

that female *D. Carolina* may improve VA, sensitivity, and/or field of view in other regions of their

eye that were not measured within this study. Future studies should further explore the regional

variation within female and male eyes to elucidate how visual performance changes outside of

the central region of the eye.

It is unknown what --- if any --- selective advantage female band-winged grasshoppers

may gain from having larger eyes with finer vision. One possibility is that females could benefit

from finer vision by more accurately interpreting visual signals. Although mating systems and

behaviors are variable across the subfamily, many species use a variety of potential visual

signals including those involving either their colorful hindwings or leg movements (Kerr, 1974;

Otte, 1970; Otte, 1984; Willey and Willey, 1969). Females may therefore benefit from finer

vision by being better able to perceive and interpret these potential signals of species identity

and/or mate quality. Additionally, females could use their enhanced vision to initiate anti-

predator defenses at a greater distance. Band-winged grasshoppers have a suite of defensive

behaviors in response to approaching predators (Collier and Hodgson, 2017; Santer et al.,

2012). Initiating these defenses at greater distances could be especially beneficial to females

because they may take a longer time than males to reach sexual maturity even once in their

final adult instar (Pfadt, 1994). Alternatively, the VA dimorphism could be a byproduct of

increased body size and serve no beneficial function. Future studies utilizing the natural

variation in band-winged size dimorphism, mating systems, and development could elucidate

how these factors contribute to the visual dimorphism seen in this study.

Surprisingly, our results are one of the first studies to show females with finer vision than

males in species with the potential for high-resolution vision (Figure 6). In insects, females of the

tiny parasitic wasp *Trichogramma evanescens* have larger eyes with ~23% finer vision than

males. However, because of their small size, female VA (~20°) is much coarser than what was

measured in this study and likely limits their visually guided behaviors (Nilsson, 2009; Nilsson,

2013). In mammals, Seymoure and Juraska (1997) found that female rats behaviorally

outperform males for coarse stimuli but that this sex difference disappeared at finer stimuli. And

in fish, Corral-López et al. (2017) found that female artificially selected guppies have finer VA

than males. However, unlike what we report in this study, the difference disappears when

controlling for body size.

We believe that the documentation of females with finer vision than males is lacking—

not because it is a rare phenomenon—but rather little research has so far been conducted on

the topic. Aside from the well documented love-spot in species of flying insects, few visual

acuity studies have examined differences between the sexes and studies often do not report the

sexes of the individuals measured. Instead most studies have prioritized either an ecological

approach [sampling only a few individuals because they sample many species; *e.g.* (Collin and

Partridge, 1996; Lisney et al., 2012; Pettigrew et al., 1988)] or a retinal topography approach

[sampling only a few individuals because of the extensive work it takes in each individual; *e.g.*

(Coimbra et al., 2013; Landgren et al., 2014) making extensive comparisons between the sexes

not possible. More recent studies in insects have found variation in eye-scaling within insect

species but have so far focused on the morphs of eusocial insects (Perl and Niven, 2016a; Perl

and Niven, 2016b; Taylor et al., 2019). Because eye size is a major factor influencing visual

performance (Cronin et al., 2014; Kirschfeld, 1976; Land and Nilsson, 2012), body size

increases that lead to eye size increases could be utilized for VA improvements (Corral-López

et al., 2017). Based on the prevalence of sexual size dimorphisms throughout many animal

groups (Kuntner and Coddington, 2020; Lislevand et al., 2007; Parker, 1992; Teder, 2014), VA

differences between the sexes may be an under-documented --- yet not uncommon ---

phenomenon that warrants further consideration and exploration.

Figure 6: In insects with moderate spatial vision (VA < 10°), studies that report VA

separately for each sex have typically 1) not shown females with finer vision than males

and 2) been based on smaller sample sizes. Our study (colored circles) is one of the first to

suggest a female-biased VA dimorphism in insects with the potential for Type I spatial vision

(VA < 10°). Red = *Arphia pseudonietana,* Yellow = *Spharagemon equale,* gray = *Dissosteira*

carolina. Data and references can be found in Supplemental Table 1.

5. Acknowledgements

419   The authors thank Dr. Carrie Veilleux and Dr. Eleanor Caves for comments on an earlier draft of

420   this manuscript. The authors would also like to thank Cesar Nufio for initial help with species

421   identification. Eran Maina, Jack Redick, Jack Whalen, and Miura Wiley all provided help in the

422   field during experiment II.

Literature Cited:

Bagheri, Z. M., Jessop, A.-L., Kato, S., Partridge, J. C., Shaw, J., Ogawa, Y. and Hemmi, J. M. (2020). A new method for mapping spatial resolution in compound eyes suggests two visual streaks in fiddler crabs. *The Journal of Experimental Biology* **223**, jeb210195.

Barlow, H. B. (1952). The size of ommatidia in apposition eyes. *Journal of Experimental Biology* **29**, 667–674.

Bergman, M. and Rutowski, R. L. (2016). Eye morphology and visual acuity in the pipevine swallowtail (*Battus philenor*) studied with a new method of measuring interommatidial angles. *Biological Journal of the Linnean Society* **117**, 646–654.

Caves, E. M., Sutton, T. T. and Johnsen, S. (2017). Visual acuity in ray-finned fishes correlates with eye size and habitat. *J. Exp. Biol.* **220**, 1586.

Caves, E. M., Brandley, N. C. and Johnsen, S. (2018). Visual acuity and the evolution of signals. *Trends in Ecology & Evolution* **33**, 358–372.

Caves, E. M., Nowicki, S. and Johnsen, S. (2019). Von uexküll revisited: addressing human biases in the study of animal perception. *Integrative and Comparative Biology* **59**, 1451–1462.

Coimbra, J. P., Hart, N. S., Collin, S. P. and Manger, P. R. (2013). Scene from above: Retinal ganglion cell topography and spatial resolving power in the giraffe (*Giraffa camelopardalis*). *Journal of Comparative Neurology* **521**, 2042–2057.

Collier, A. and Hodgson, J. Y. S. (2017). A shift in escape strategy by grasshopper prey in response to repeated pursuit. *Southeastern Naturalist* **16**, 503–515.

Collin, S. P. and Partridge, J. C. (1996). Retinal specializations in the eyes of deep-sea teleosts. *Journal of Fish Biology* **49**, 157–174.

Corral-López, A., Garate-Olaizola, M., Buechel, S. D., Kolm, N. and Kotrschal, A. (2017). On the role of body size, brain size, and eye size in visual acuity. *Behavioral Ecology and Sociobiology* **71**, 179.

Cronin, T. W., Johnsen, S., Marshall, N. J. and Warrant, E. J. (2014). *Visual Ecology*. STU-Student edition. Princeton University Press.

Hochkirch, A. and Gröning, J. (2008). Sexual size dimorphism in Orthoptera (*sens. str* .) — a review. *Journal of Orthoptera Research* **17**, 189–196.

Horridge, G. A. (1978). The separation of visual axes in apposition compound eyes. *Philosophical Transactions of the Royal Society of London. B, Biological Sciences* **285**, 1–59.

Jander, U. and Jander, R. (2002). Allometry and resolution of bee eyes (Apoidea). *Arthropod Structure & Development* **30**, 179–193.

Jordan, L. A. and Ryan, M. J. (2015). The sensory ecology of adaptive landscapes. *Biol Lett*
11,.

Kelber, A. and Somanathan, H. (2019). Spatial vision and visually guided behavior in apidae.
Insects **10**, 418.

Kerr, G. E. (1974). Visual and acoustical communicative behaviour in *dissosteira carolina*
(orthoptera: acrididae). *The Canadian Entomologist* **106**, 263–272.

Kiltie, R. A. (2000). Scaling of visual acuity with body size in mammals and birds. *Functional*
Ecology **14**, 226–234.

Kirschfeld, K. (1976). The Resolution of Lens and Compound Eyes. In *Neural Principles in*
Vision (ed. Zettler, F.) and Weiler, R.), pp. 354–370. Berlin, Heidelberg: Springer.

Kirschfeld, K. and Wenk, P. (1976). The Dorsal Compound Eye of Simuliid Flies: *Zeitschrift für*
Naturforschung C **31**, 764–765.

Kuntner, M. and Coddington, J. A. (2020). Sexual Size Dimorphism: Evolution and Perils of
Extreme Phenotypes in Spiders. *Annual Review of Entomology* **65**, 57–80.

Land, M. F. (1997). Visual Acuity in Insects. *Annual Review of Entomology* **42**, 147–177.

Land, M. F. and Eckert, H. (1985). Maps of the acute zones of fly eyes. *J. Comp. Physiol.* **156**,
525–538.

Land, M. F. and Nilsson, D.-E. (2012). *Animal Eyes*. Oxford University Press.

Landgren, E., Fritsches, K., Brill, R. and Warrant, E. (2014). The visual ecology of a deep-
sea fish, the escolar *Lepidocybium flavobrunneum* (Smith, 1843) <sup/>. *Philosophical*
Transactions of the Royal Society B: Biological Sciences **369**, 20130039.

Lislevand, T., Figuerola, J. and Székely, T. (2007). Avian body sizes in relation to fecundity,
mating system, display behavior, and resource sharing: *ecological archives* e088-096.
Ecology **88**, 1605–1605.

Lisney, T. J., Iwaniuk, A. N., Kolominsky, J., Bandet, M. V., Corfield, J. R. and Wylie, D. R.
(2012). Interspecifc variation in eye shape and retinal topography in seven species of
galliform bird (Aves: Galliformes: Phasianidae). *Journal of Comparative Physiology A*
198, 717–731.

Narendra, A., Alkaladi, A., Raderschall, C. A., Robson, S. K. A. and Ribi, W. A. (2013).
Compound eye adaptations for diurnal and nocturnal lifestyle in the intertidal ant,
polyrhachis sokolova. *PLOS ONE* **8**, e76015.

Nilsson, D.-E. (2009). The evolution of eyes and visually guided behaviour. *Philosophical*
Transactions of the Royal Society B: Biological Sciences **364**, 2833–2847.

Nilsson, D.-E. (2013). Eye evolution and its functional basis. *Visual Neuroscience* **30**, 5–20.

Otte, D. (1970). A comparative study of communicative behavior in grasshoppers.
Miscellaneous Publications: Museum of Zoology, University of Michigan.

Otte, D. (1984). *The North American Grasshoppers, Volume 2, Acridiae, Oedipodinae*.
Cambridge (MA): Harvard University Press.

Parker, G. A. (1992). The evolution of sexual size dimorphism in fish*. *Journal of Fish Biology*
41, 1–20.

Partan, S. and Marler, P. (2002). The Umwelt and its relevance to animal communication:
Introduction to special issue. *Journal of Comparative Psychology* **116**, 116–119.

Perl, C. D. and Niven, J. E. (2016a). Differential scaling within an insect compound eye.
Biology Letters **12**, 20160042.

Perl, C. D. and Niven, J. E. (2016b). Colony-level differences in the scaling rules governing
wood ant compound eye structure. *Sci Rep* **6**, 1–8.

Pettigrew, J. D., Dreher, B., Hopkins, C. S., McCall, M. J. and Brown, M. (1988). Peak
density and distribution of ganglion cells in the retinae of microchiropteran bats:
implications for visual acuity. *Brain, Behavior and Evolution* **32**, 39–56.

Pfadt, R. E. (1994). *Field Guide to Common Western Grasshoppers*. Wyoming Agricultural
Experiment Station.

R CORE Team (2018). *R: A Language and Environment for Statistical Computing*. Vienna,
Austria: R Foundation for Statistical Computing.

Rutowski, R. L., Gislén, L. and Warrant, E. J. (2009). Visual acuity and sensitivity increase
allometrically with body size in butterflies. *Arthropod Structure & Development* **38**, 91–
100.

Santer, R. D., Rind, F. C. and Simmons, P. J. (2012). Predator versus prey: locust looming-
detector neuron and behavioural responses to stimuli representing attacking bird
predators. *PLoS ONE* **7**, e50146.

Seymoure, P. and Juraska, J. M. (1997). Vernier and grating acuity in adult hooded rats: The
influence of sex. *Behavioral Neuroscience* **111**, 792–800.

Spaethe, J. and Chittka, L. (2003). Interindividual variation of eye optics and single object
resolution in bumblebees. *Journal of Experimental Biology* **206**, 3447–3453.

Taylor, G. J., Tichit, P., Schmidt, M. D., Bodey, A. J., Rau, C. and Baird, E. (2019).
Bumblebee visual allometry results in locally improved resolution and globally improved
sensitivity. *eLife* **8**, e40613.

Teder, T. (2014). Sexual size dimorphism requires a corresponding sex difference in
development time: a meta-analysis in insects. *Functional Ecology* **28**, 479–486.

Uexküll, J. von (2013). *A Foray into the Worlds of Animals and Humans: with A Theory of*
Meaning. U of Minnesota Press.

Veilleux, C. C. and Kirk, E. C. (2014). Visual acuity in mammals: effects of eye size and
ecology. *Brain, Behavior and Evolution* **83**, 43–53.

Warrant, E. J. (2016). Sensory matched filters. *Current Biology* **26**, R976–R980.

Willey, R. B. and Willey, R. L. (1969). Visual and acoustical social displays by the grasshopper
arphia conspersa (orthoptera: acrididae). *Psyche: A Journal of Entomology* **76**, 280–305.