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RESVERATROL AND PTEROSTILBENE POTENTLY INHIBIT SARS-COV-2 INFECTION IN VITRO

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Abstract

The current COVID-19 pandemic is caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and has an enormous impact on human health and economy[1]. In search for therapeutic options, researchers have proposed resveratrol, a food supplement with known antiviral, anti-inflammatory and anti-oxidant properties as an advantageous antiviral therapy for SARS-CoV-2 infection[2–4]. Here, we provide evidence that both resveratrol and its metabolically more stable structural analog, pterostilbene, exhibits potent antiviral properties against SARS-CoV-2 *in vitro*. Resveratrol and pterostilbene showed antiviral activity in African green monkey kidney cells and in human primary bronchial epithelial cells cultured in an air-liquid interface system. Mechanistic analyses demonstrated that both compounds actively interfere with the post-entry steps of virus replication cycle and their antiviral activity is long-lasting. Collectively, our data indicate that resveratrol and pterostilbene are promising antiviral compounds to treat SARS-CoV-2 infection and advocate evaluation of these compounds in clinical trials

Introduction

Since its emergence in December 2019, the novel severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has infected over 23 million people worldwide[1]. Infection with SARS-CoV-2 leads to a wide range of manifestations ranging from an asymptomatic infection to a self-limiting mild disease to a potentially fatal disease. It is estimated that approximately 15% of the infected individuals develop severe pneumonia and about 5% develop an acute respiratory distress syndrome (ARDS), septic shock and/or multiple organ failure. Severe disease is a consequence of lung inflammation and damage caused by direct viral infection of the lung and/or by the immune response triggered to control virus dissemination[5–7]. Within less than a year, over 850,000 individuals have succumbed to SARS-CoV-2 infection worldwide[1]. To prevent and treat SARS-CoV-2 infection, vaccines and antiviral drugs are urgently needed.

The natural compounds resveratrol (trans-3,5,4'-trihydroxystilbene) and the structurally related pterostilbene (trans-3,5-dimethyoxy-4'-hydroxystilbene) share multiple bioactivities which are potentially beneficial for human health[8]. In recent years, resveratrol was described to exhibit antiviral activity towards a large number of viruses including human immunodeficiency virus[9], influenza virus[10], respiratory syncytium virus[11], as well as, Middle East respiratory syndrome coronavirus (MERS-CoV)[12]. In most cases, resveratrol was found to directly interfere with viral replication[13]. Next to the direct effect on virus replication, resveratrol was also reported to exhibit anti-inflammatory and anti-oxidant properties thereby having the potential to mitigate virus-induced disease pathogenesis[13,14]. Based on these findings, we and others have postulated that resveratrol and its structural analogs might be an advantageous treatment option for SARS-CoV-2 infected individuals[2–4].

In this study, we examined the efficacy of resveratrol and pterostilbene on suppression of viral replication in several *in vitro* models of SARS-CoV-2 infection. Specifically, we used African green monkey kidney cells (Vero E6), human lung epithelial cells (Calu-3), and human primary bronchial epithelial cells differentiated on air-liquid interface (ALI) cultures. To delineate the mode-of-action, we assessed the effect of the compounds when added prior to, during, and after virus cell entry had been established. The antiviral effect was also evaluated over time to assess the long-lasting effect of the compounds.

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Results

Resveratrol and pterostilbene inhibit SARS-CoV-2 infection in Vero E6 cells

Prior to assessing antiviral activity of resveratrol and pterostilbene, we determined the cellular cytotoxicity of the compounds in Vero E6 cells. We observed a dose-dependent cytotoxic effect of both resveratrol and pterostilbene (Extended data Fig. 1a,b). Limited cytotoxicity was observed up to a concentration of 200 µM resveratrol and 100 µM pterostilbene. At these conditions, no cytotoxicity was observed for the solvent control EtOH. However, the cells treated with compound appeared to be stressed based on morphology and therefore we decided to use 150 µM resveratrol and 60 µM pterostilbene as the highest concentration in follow-up experiments. Next, we investigated the antiviral effect of resveratrol and pterostilbene during infection with SARS-CoV-2 (isolate NL/2020). Vero E6 cells were inoculated with SARS-CoV-2 (MOI 1) in the presence of increasing concentrations of resveratrol, pterostilbene or an equivalent volume of EtOH corresponding to the highest concentration of the compound. Resveratrol and pterostilbene showed a dose-dependent antiviral effect on SARS-CoV-2 infection in Vero E6 cells (Fig. 1a,b). No effect on virus progeny production was observed for the EtOH solvent control. Subsequent non-linear regression analyses revealed that virus particle production is reduced by 50% (EC50) at a concentration of 66 µM resveratrol and 19 µM pterostilbene (Fig. 1c). Furthermore, 90% reduction (EC90) of virus progeny is observed at a concentration of 119 µM resveratrol and 47 µM pterostilbene (Fig. 1c). Thus, pterostilbene has a more efficacious antiviral effect at lower concentrations in comparison to resveratrol.

Next, we investigated how long resveratrol and pterostilbene maintain their antiviral activity in cell culture. We infected Vero E6 cells with SARS-CoV-2 at MOI 0.01 in the presence of the compound and harvested the supernatant at 16, 24, 40, 60 hours post-inoculation (hpi). For the non-treated (NT) and solvent control samples, no differences were observed in virus growth and virus particle production plateaued at 40 hpi at which time point all cells were dead (Fig. 1d,e). In the presence of resveratrol as well as pterostilbene a strong antiviral effect (~2 log reduction which corresponds to 99% reduction in virus production) was observed at 16 and 24 hpi. Significant antiviral activity was observed up to 40 hpi. As this time-point corresponds to roughly 5 rounds of replication[15], the results highlight the long-lasting antiviral effect of both compounds.

Resveratrol and pterostilbene directly interfere with SARS-CoV-2 replication.

To test whether the antiviral capacity of the compounds rely on lysis, inactivation or neutralization of the virion itself, we next performed a virucidal assay[16]. Briefly, 2.5×10^5 PFUs of SARS-CoV-2 was incubated with 150 µM resveratrol, 60 µM pterostilbene or equivalent volumes of EtOH for 2 h and subjected to plaque assay. No differences in viral titers were observed relative to the non-treated (NT) or EtOH control (Fig. 2a). This indicates that the compounds do not exhibit virucidal activity at these conditions, but rather interfere with viral replication in Vero E6 cells. To examine this further, we performed a time-of-drug-addition experiment. In this experiment, resveratrol or pterostilbene was added either prior, during or post SARS-CoV-2 inoculation and progeny virus particle production was evaluated at 8hpi (Fig. 2b). Comparable results were obtained for both compounds (Fig. 2c,d). No effect was observed when the compounds were solely present prior to infection. A mild, yet non-significant, reduction in virus particle production was observed when the compounds were present during virus inoculation. Importantly, a significant reduction in virus progeny production was observed when the compounds were added after removal of the virus inoculum (Fig. 2c,d). No significant effect was noted when resveratrol or pterostilbene were added at 4 or 6 hpi (Extended data Fig. 2a and 2b). Collectively, these results indicate that the compounds directly interfere with the viral infectious cycle at a stage after virus entry but prior to virus assembly and release.

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Resveratrol and pterostilbene do not exhibit significant anti-SARS-CoV-2 activity in Calu-3 cells.

We next sought to verify the findings in the human lung epithelial cell model, Calu-3 cells[17]. Resveratrol was more cytotoxic in Calu-3 cells when compared to Vero E6, since the highest non-toxic concentration was 50 µM (Extended data Fig. 3a). For pterostilbene, the highest non-toxic concentration was set at 60 µM, similar to Vero E6 cells (Extended data Fig. 3b). At these concentrations, no significant reduction in virus particle production was observed with both compounds in Calu-3 cells (Fig. 3a,b) although a negative trend in virus particle production was observed with pterostilbene. The inability of both compounds to induce antiviral effects in Calu-3 as observed in Vero E6, underline the inherent differences between these two cell line models.

Resveratrol and pterostilbene significantly inhibit SARS-CoV-2 infection in primary human bronchial epithelial cells cultured under ALI conditions.

Given the known variability of data obtained in SARS-CoV-2 cell line models[17], we decided to verify the antiviral activity of resveratrol and pterostilbene in a primary human bronchial epithelium cell (PBEC) model[18,19]. PBECs obtained from healthy individuals, were cultured on ALI conditions to induce differentiation into ciliated and secretory epithelial cells[20] (Fig. 4a). In this model, no cytotoxicity was found at a concentration of 150 µM resveratrol and 60 µM pterostilbene as determined by live/death staining using flow cytometry and by an LDH assay (Extended data Fig. 4a,b). Accordingly, fully differentiated PBECs were inoculated with SARS-CoV-2 (MOI 5) in the presence of resveratrol and pterostilbene, and progeny virus particle production was evaluated at 12, 24, 48 hpi (Fig. 4a). At 12 hpi, the viral titers were very low and in presence of the compounds often fell below the threshold of detection (Fig. 4b,c). Importantly, at 24 and 48 hpi, a significant antiviral effect was observed for both resveratrol and pterostilbene (Fig. 4b,c). At 48 hpi, resveratrol significantly reduced the virus titer with 2.1 Log (corresponding to 99.3% reduction) when compared to the EtOH control in PBECs. In presence of pterostilbene, the virus titer was reduced with 1.2 Log (corresponding to 87,5% reduction) when compared to the EtOH control in PBEC at 48 hpi. Collectively, our data demonstrate that resveratrol and pterostilbene exhibit potent antiviral activity towards SARS-CoV-2 in differentiated human primary bronchial epithelial cells.

Based on our data and studies reporting the pharmacological properties of resveratrol and pterostilbene, we conclude that both drugs have the potential to exhibit antiviral efficacy in the course of COVID-19. The concern of the relatively low metabolic stability and limited bioavailability of resveratrol following oral administration in humans[8,13] could be circumvented by use of other administration modes and routes such as the use of a resveratrol aerosolized suspension spray, resveratrol co-spray dried microparticles or nanosponges[21–23]. Alternatively, pterostilbene, the pharmacologically superior compound to resveratrol, is an attractive candidate worth further testing in clinical trials[8].

Finally, since both drugs are commercially available and could therefore be used as a self-medicative prophylactic, a word of caution must be considered. Our data represent promising laboratory findings in cells, and therefore do not indicate that these drugs will be of benefit to treat COVID-19 in patients. Randomized double-blind controlled clinical trials must first swiftly be conducted to prove whether or not these drugs are indeed advantageous for COVID-19 treatment.

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Author contributions

B.M.E., N.D.K, E.B, M.C.N, P.V., J.M., I.A.R.Z. and J.M.S designed the experiments. B.M.E., N.D.K, E.B, B.H.T., D.P.I.P., H.H.E., L.A., D.G., executed the experiments. M.B. selected and recruited donors, sampled donors, and analyzed donor data. B.M.E, I.A.R.Z and J.M.S wrote the manuscript. All authors edited the manuscript.

Material and methods

Compounds

100% trans-resveratrol was obtained from Bulkpowders and dissolved in absolute ethanol (EtOH) obtaining a stock solution of 100 mM which was used in the experiments. Pterostilbene was (Sigma Aldrich) dissolved in absolute EtOH obtaining a stock solution of 10 mM which was used in the experiments. The EtOH concentration was below 0.6 % in all infection experimental conditions.

Cell culture and differentiation

The African green monkey Vero E6 cell line (ATCC CRL-1586) was maintained in Dulbecco's minimal essential medium (DMEM) (Gibco), high glucose supplemented with 10% fetal bovine serum (FBS) (Life Science Production), penicillin (100 U/mL), and streptomycin (100 U/mL) (Gibco). The human lung epithelial cell line Calu-3 (ATCC HTB-55) was maintained in DMEM F-12 (Lonza, Switzerland) supplemented with 10% FBS, 1% Glutamax (Thermofisher), 1% non-essential amino acid (Thermofisher), penicillin (100 U/mL), and streptomycin (100 U/mL) (Gibco). All cells were mycoplasma negative and maintained at 37°C under 5% CO_2. Primary bronchial epithelial cells were cultured from bronchial bushing obtained by fibreoptic bronchoscopy performed using a standardized protocol during conscious sedation [24,25]. The medical ethics committee of the University Medical Center Groningen approved the study, and all subjects gave their written informed consent. The donors were 2 male and 2 female non-smoking healthy control volunteers (less than 2.5 packyears) with no history of respiratory disease aged 49-62. PBECs were cultured and fully differentiated under ALI conditions in transwell inserts, as previously described[26].

SARS-CoV-2 production and characterization

The SARS-CoV-2 strain NL/2020 was obtained from European Virus Archive global (EVAg -010V-03903). The original stock was passaged twice in Vero E6 cells to obtain a working stock. Infectious virus titers were determined by plaque assay on Vero E6 cells and defined as the number of plaque forming units (PFU) per mL. Briefly, Vero E6 cells were seeded at a density of 1.3×10^5 cells/well in a 12-well plate format. At 24 hr post-seeding, cells were infected with 10-fold serial dilutions of the sample in duplo. At 2 hpi, wells were overlaid with 1% seaplaque agarose (Lonza) prepared in 2x MEM. Plaques were counted at 44 hpi. One plaque in the lowest dilution corresponds to 150 PFUs/ml and was set as the detection limit of the assay.

Cytotoxicity Assays

MTS

In the context of Vero E6 and CaLu-3 cells, cells were exposed to increasing concentrations of resveratrol and pterostilbene for 8 hr. Subsequently, cellular cytotoxicity was evaluated using the CellTiter 96® AQueous One Solution Cell Proliferation Assay kit using manufacturer's instructions from Promega (Madison, WI, USA). Briefly, cells were seeded in a 96-well plate at a density of $1x10^4$. At 24 hr post-seeding, Vero E6 cells were treated with increasing concentrations of resveratrol and pterostilbene ranging from 2 to 250 µM or the

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equivalent volumes of EtOH for 8 hr at 37 °C. Calu-3 cells were treated with 50 to 150 µM resveratrol, 40 or 60 µM pterostilbene or the equivalent volumes of EtOH for 8 hr at 37 °C. At 8 hr post-treatment, 20 µl of MTS/PMS solution was added per well and incubated for 2 hr at 37 °C. Subsequently, 10% SDS was added to each well (2% end concentration) to stop the reaction and the absorbance was measured at 490 nm with a microplate reader. Values are displayed as percentage compared to normalized non-treated control. All individual experiments were performed in triplicate.

Live death staining flow cytometry

PBECs were exposed to 150 µM resveratrol and 60 µM pterostilbene at the basolateral side for 48 hr at 37 °C, and subsequently harvested and stained with fixable viability dye eFluor 780 for 20 min at 4 °C. After staining cells were washed in FACS buffer (PBS 2% FBS), centrifuged and subsequently fixed with 4% PFA for 10 min at 4 °C. After fixation, cells were washed, centrifuged and resuspended in FACS buffer. Cells were analyzed for viability with the LSR-2 flow cytometer (BD Bioscience). Data was analyzed using Kaluza software (Beckman Coulter).

LDH

PBECs were exposed to 150 µM resveratrol and 60 µM pterostilbene at the basolateral side for 48 hr at 37 °C. after incubation, apical sides of the inserts were incubated with medium for 30 min at 37 °C. The apical wash was collected and centrifuged 2000 x g at 4 °C to clear from cell debris. The commercially available kit (ThermoFisher, CyQUANT™ LDH Cytotoxicity Assay Kit) was used according to manufacturer protocol. The absorbance was measures ad 490 and 680 nm using a microplate reader. 680 nm absorbance OD values (background) were subtracted from 490 nm OD values. Cytotoxicity was calculated using the following formula:

$$\%\ cytotoxicity = \frac{(OD\ Compound\ treated\ LDH\ activity - OD\ spontaneous\ LDH\ activity)}{(OD\ maximum\ LDH\ activity - OD\ spontaneous\ activity)}$$

Antiviral assay in Vero E6 and Calu-3 cells

Vero E6 cells were seeded at a density of 1.3 x 10^5 in 12-well plates and Calu-3 cells were seeded at a density of 2x10^5 in 24-well plates. Cells were infected with SARS-CoV-2 at a multiplicity of infection (MOI) 1 and treated with increasing concentrations of resveratrol and pterostilbene or the equivalent volumes of EtOH corresponding to the highest concentration of compound for 2 hr at 37 °C. Infection was done in 250 µl DMEM (2% FBS) medium. After infection, virus inoculum was removed, cells were washed twice with plain DMEM media, and fresh DMEM 10% FBS containing the compound or the equivalent volumes of EtOH was added after which incubation was continued. Cell supernatant was collected at 8 hours post inoculation (hp)i, centrifuged to clarify from cell debris and the viral titer was determined using plaque assay. For the durability assay, Vero E6 cells were infected with SARS-CoV-2 at MOI 0.01 and treated with 150 µM resveratrol or 60 µM pterostilbene or the equivalent volume of EtOH as indicated above. Supernatants were collected at 16, 24, 40 and 60 hpi and analyzed as above.

Antiviral assay in primary bronchial epithelial ALI culture.

After 3 weeks culture under ALI conditions, cells were washed once with plain ALI medium and inoculated with SARS-CoV-2 at MOI 5 at the apical side. At the time of infection, 75 or 150 µM Resveratrol, 60 µM Pterostilbene or the equivalent volumes of EtOH corresponding with the highest concentration of compound were added at the basolateral side of the insert. At 2 hpi, cells were washed twice with ALI medium at the apical side and incubation was continued on air at 37 °C. Thirty min prior to harvesting (12, 24 and 48 hpi), OptiMEM (Gibco) was added to the apical side of the ALI cultures. At the time of harvest, the apical supernatant and 150 µl of basolateral medium was harvested. At the basolateral side, after each harvest new ALI culture media containing the compound or EtOH was added. The viral titer in the apical supernatant was determined using plaque assay.

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Time-of-drug-addition assay

For the time-of-drug-addition experiments, the cells were treated with 150 µM resveratrol or 60 µM pterostilbene at pre, during, or post-inoculation conditions (Fig. 2B depicts experimental set-up). For pre-treatment, cells were incubated with the compounds or the equivalent volume of EtOH for 2 h. At the time of infection, cells were washed three times before the addition of the virus inoculum. For the during condition, the compounds or the equivalent volumes EtOH were added together with the virus inoculum and was present for 2 h. At 2hpi, cells were washed three times with plain DMEM, fresh DMEM 10% FBS was added, and incubation was continued. For the post-infection conditions, the compounds or the equivalent volumes EtOH were added to the cell culture medium after removal of the virus inoculum. All supernatants were collected 8 hpi, centrifuged to clarify from cell debris and subjected to plaque assay to determine the viral titer.

Virucidal assay

$2.5x10^5$ PFUs of SARS-CoV-2 were incubated in 300 µl DMEM 2 % FBS in the presence or absence of 150µM Resveratrol, 60 µM Pterostilbene or the equivalent volume of EtOH for 2h at 37 °C. Subsequently, viral titer was determined by plaque assay.

Statistical analysis

All data is represented as mean ± SEM. The concentration that reduced virus particle production by 50 and 90% is referred to as EC50 and EC90, respectively. Dose-response curves were fitted by non-linear regression analysis employing a sigmoidal model. All data was analyzed in GraphPad Prism 8 software (La Jolla, CA, USA). Non-paired two-tailed Student T test was used to evaluate statistical differences and a p value ≤ 0.05 was considered significant with $*p \leq 0.05$, $**p \leq 0.01$ and $***p \leq 0.001$ and NS as non-significant.

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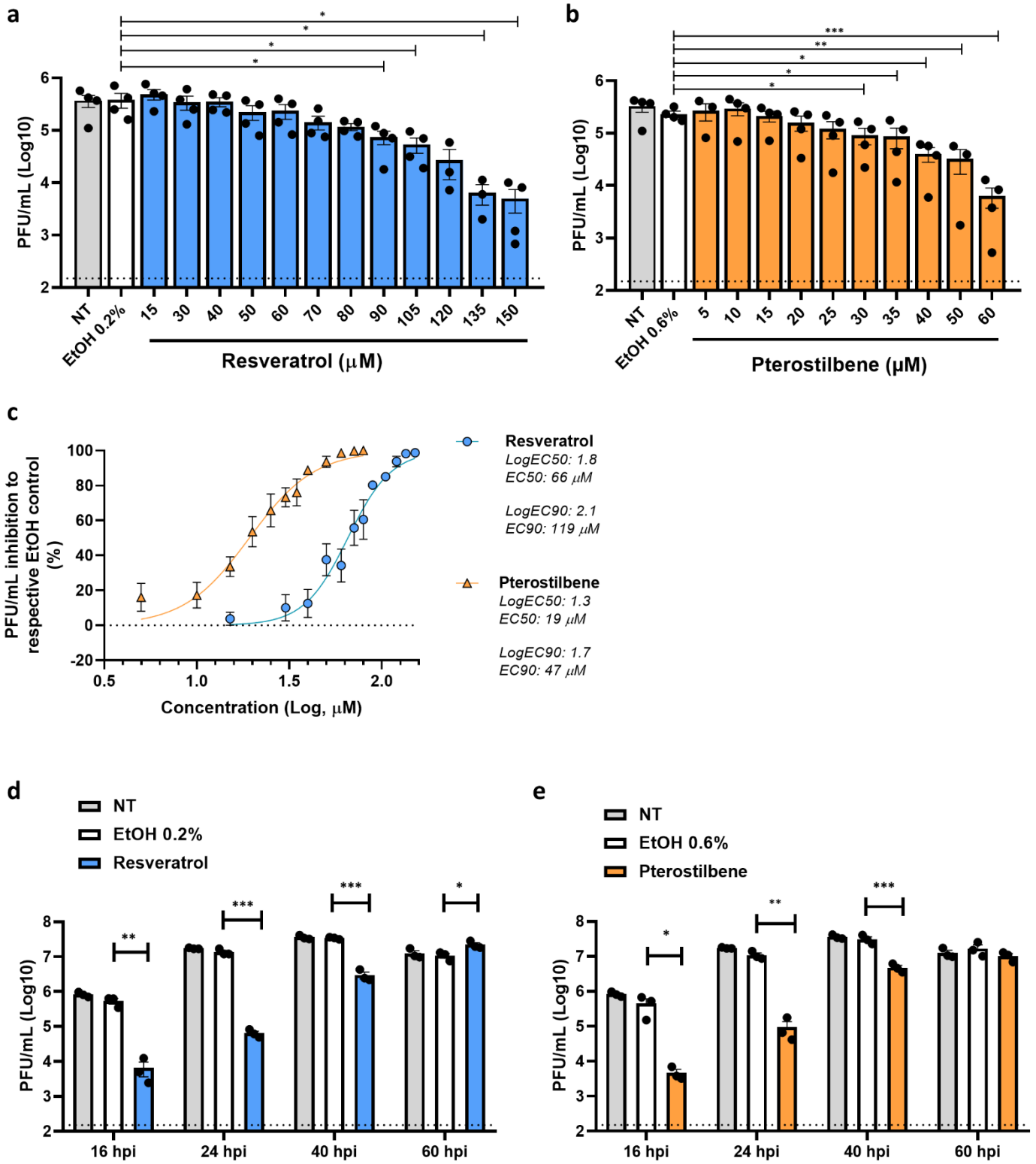

Figure 1. Antiviral effect of resveratrol and pterostilbene towards SARS-CoV-2 in Vero E6 cells. Production of infectious virus by Vero E6 cells inoculated with SARS-CoV-2 at MOI 1 in the absence (NT denotes for non-treated) or presence of increasing concentrations of **(a)** resveratrol, **(b)** pterostilbene or the EtOH solvent control. **(c)** The EC50 and EC90 values determined by non-linear regression analysis. **(d,e)** Durability of the antiviral effect of **(d)** resveratrol and **(e)** pterostilbene at 16, 24, 40 and 60 hours post-inoculation (hpi). Dotted line indicates the threshold of detection. Data are represented as mean ± SEM of at least three independent experiments. Each symbol represents data from a single independent experiment. Student T test was used to evaluate statistical differences and a p value ≤ 0.05 was considered significant with $*p \leq 0.05$, $**p \leq 0.01$ and $***p \leq 0.001$. In the absence of '*' the data is non-significant.

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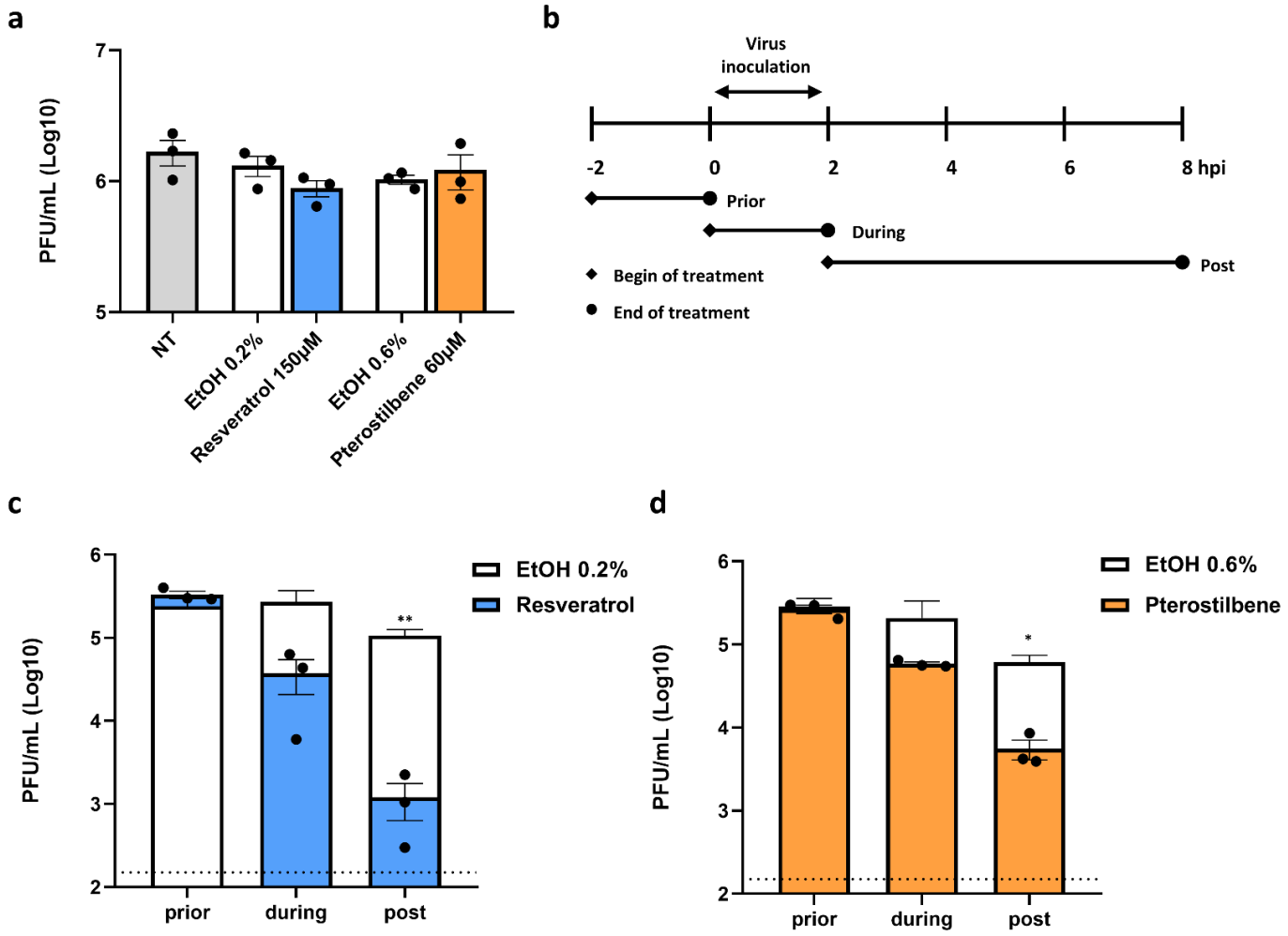

Figure 2. Resveratrol and pterostilbene interfere with SARS-CoV-2 infection when added post virus inoculation conditions. (a) Virucidal effect of resveratrol and pterostilbene on SARS-CoV-2. **(b)** Schematic representation of the experimental design. Vero E6 cells were inoculated with SARS-CoV-2 at MOI of 1 and treated with **(c)** 150 µM resveratrol, **(d)** 60 µM pterostilbene or **(c,d)** equivalent volumes of EtOH. Virus production was determined at 8 hpi via plaque assay. Data is represented as mean ± SEM from three independent experiments. Dotted line indicates the threshold of detection. Student T test was used to evaluate statistical differences and a p value ≤ 0.05 was considered significant with $*p \leq 0.05$, $**p \leq 0.01$ and $***p \leq 0.001$. In the absence of '*' the data is non-significant.

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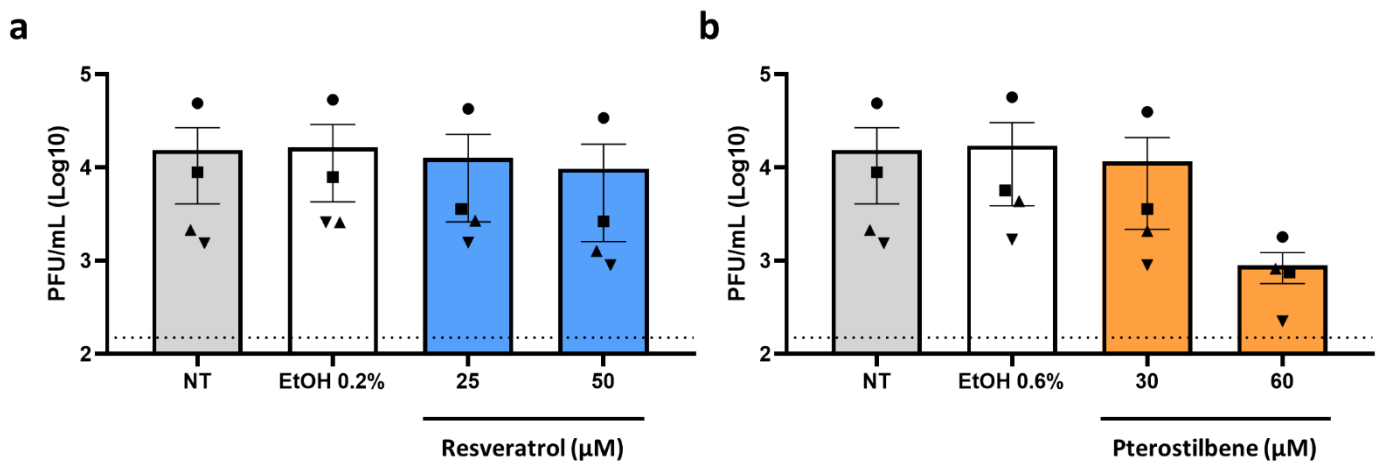

Figure 3. Resveratrol and pterostilbene do not significantly inhibit SARS-CoV-2 infection in Calu-3 cells. Calu-3 cells were inoculated with SARS-CoV-2 and treated with **(a)** 25 or 50 µM resveratrol, **(b)** 30 or 60 µM pterostilbene **(a,b)** or the EtOH solvent control. Progeny virus production was determined at 8 hpi by plaque assay. Data is represented as mean ± SEM from three independent experiments. Dotted line indicates the threshold of detection. Student T test was used to evaluate statistical differences and a p value ≤ 0.05 was considered significant with $*p \leq 0.05$, $**p \leq 0.01$ and $***p \leq 0.001$. In the absence of '*' the data is non-significant.

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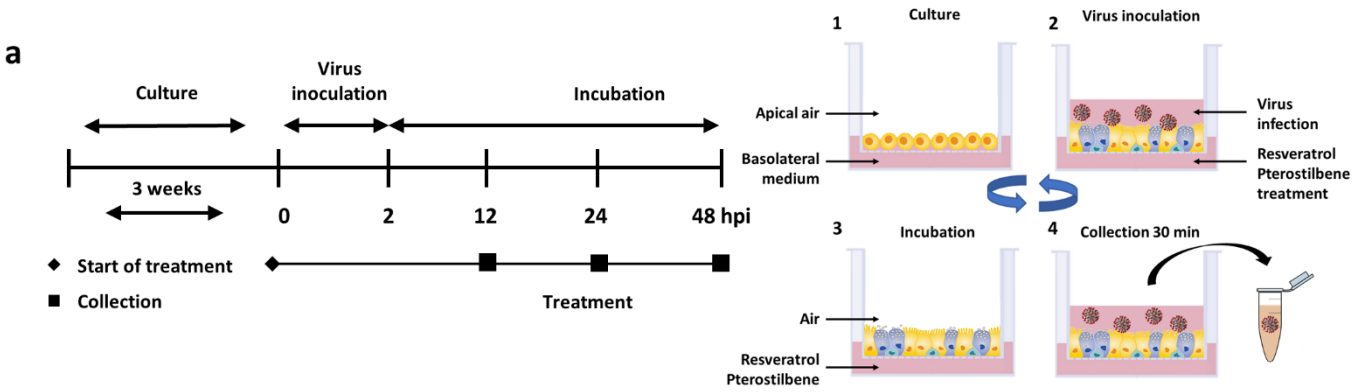

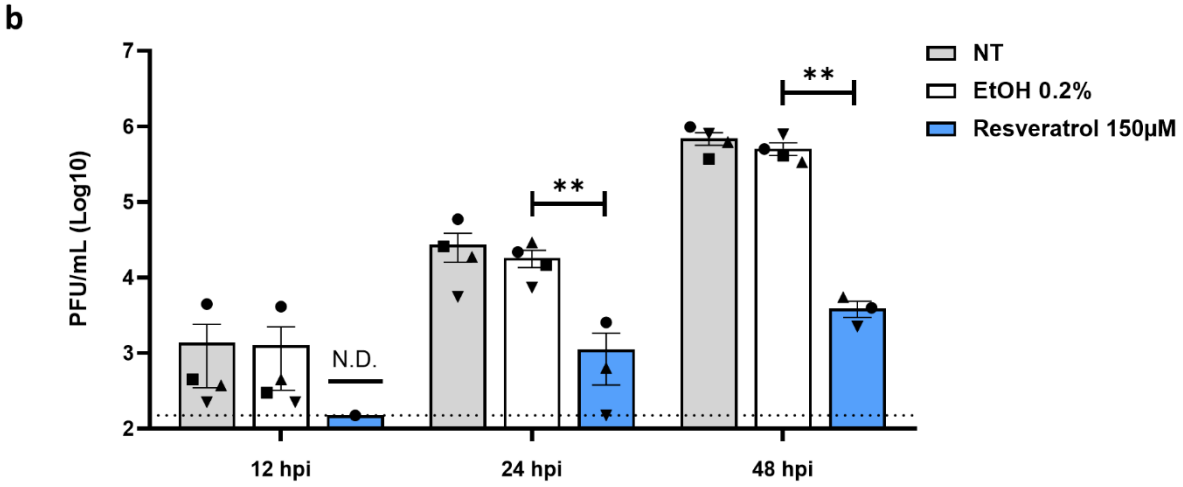

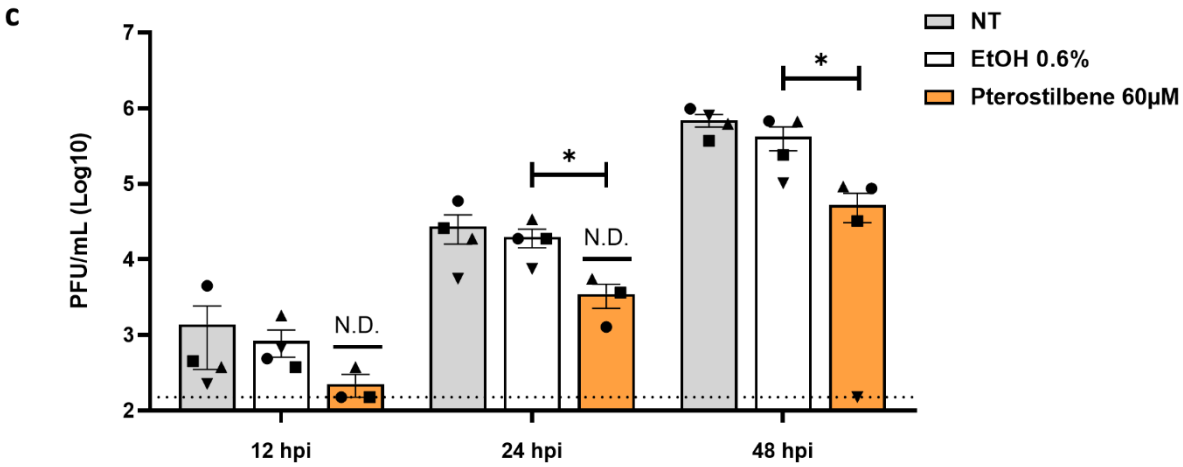

Fig 4. Resveratrol and pterostilbene inhibit SARS-CoV-2 infection in primary human bronchial epithelium cells. (a) Schematic representation of the experimental design. **(1)** Primary human bronchial epithelium cells (PBECs) were cultured on permeable inserts under Air Liquid Interface (ALI) conditions for 21 days. **(2)** Cells were inoculated with SARS-CoV-2 MOI of 5 at the apical side. **(3)** At 2 hpi, the virus inoculum was removed and cells were exposed to air until virus collection. **(4)** Medium was added to the apical side and collected after 30 min incubation. After supernatant collection, PBECs were exposed to air again **(3)** until the next collection time point. Step **4** and **3** were repeated until the end of the experiment. **(b,c)** Progeny virus production in PBECs in presence of **(b)** resveratrol or **(c)** pterostilbene. Data is represented as mean ± SEM from three or four different donors. Dotted line indicates the threshold of detection. Student T test was used to evaluate statistical differences and a p value ≤ 0.05 was considered significant with *$p \leq 0.05$, **$p \leq 0.01$ and ***$p \leq 0.001$. In the absence of '*' the data is non-significant.

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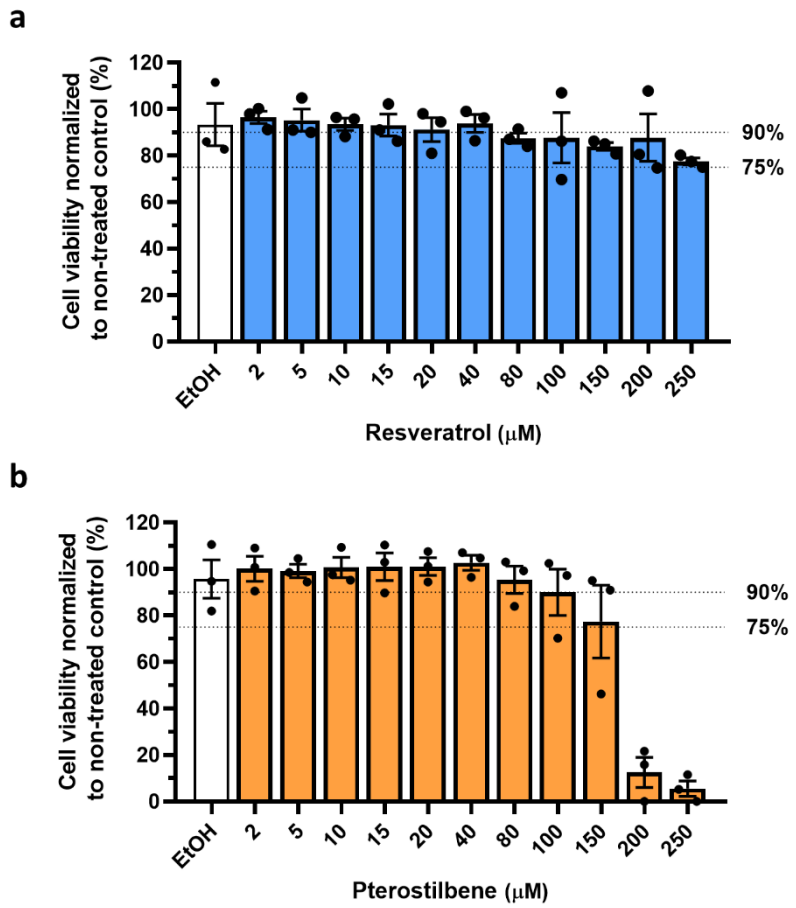

Extended data Fig. 1. Cellular cytotoxicity of resveratrol and pterostilbene in Vero E6 cells. Cell viability of Vero E6 cells incubated with increasing concentrations of **(a)** resveratrol, **(b)** pterostilbene or equivalent volumes of EtOH respective to the highest concentration of compound **(a,b)**. Cell viability was assessed using an MTS assay kit. Cell viability is expressed as percentage compared to the non-treated (NT) control. Data are represented as mean ± SEM from three independent experiments.

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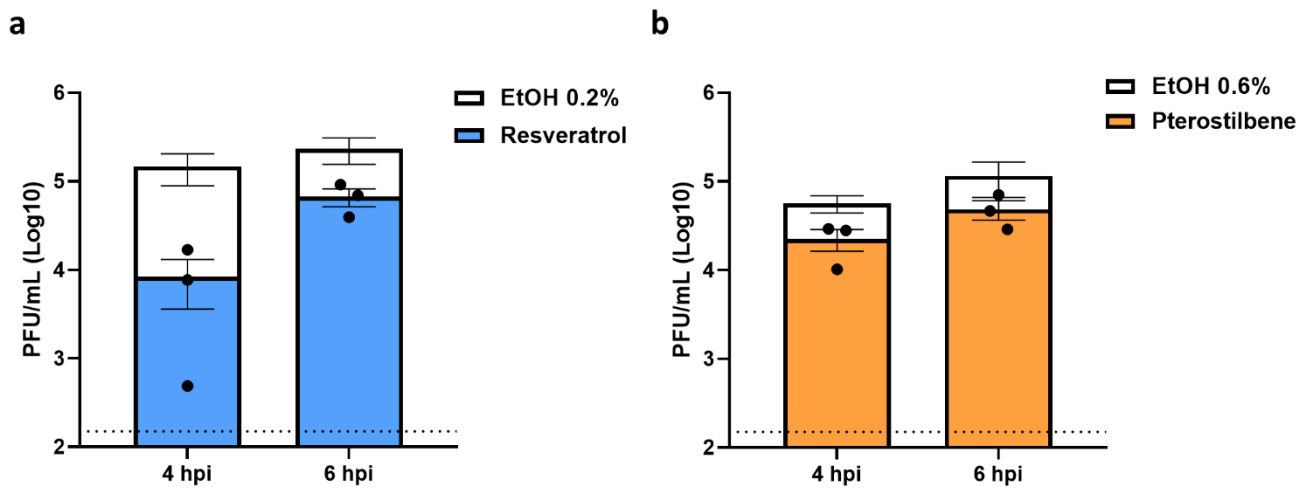

Extended data Fig. 2. Effect of resveratrol and pterostilbene when added late in the replication cycle of SARS-CoV-2. Vero E6 cells were inoculated with SARS-CoV-2 at MOI of 1 and treated with **(a)** 150 µM resveratrol, **(b)** 60 µM pterostilbene or **(a,b)** EtOH solvent control at 4 or 6 hpi. Virus production was determined at 8 hpi via plaque assay. Data is represented as mean ± SEM from three independent experiments. Dotted line indicates the threshold of detection. Student T test was used to evaluate statistical differences and a p value ≤ 0.05 was considered significant with *$p \leq 0.05$, **$p \leq 0.01$ and ***$p \leq 0.001$. In the absence of '*' the data is non-significant.

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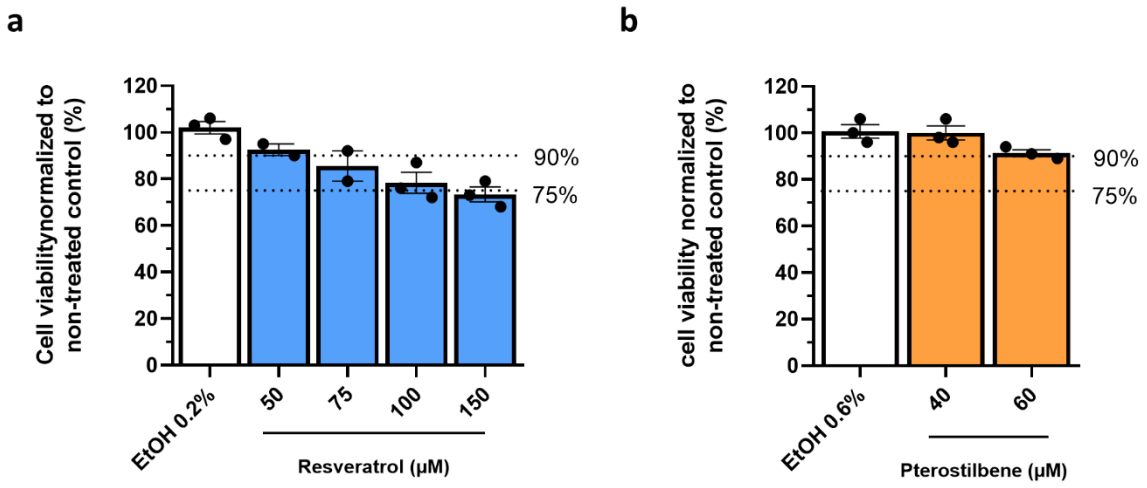

Extended data Fig. 3. Cellular cytotoxicity of resveratrol and pterostilbene in Calu-3 cells. Calu-3 cells were incubated with increasing concentrations **(a)** resveratrol, **(b)** pterostilbene **(a,b)** or equivalent volumes of EtOH corresponding to the highest used concentration, respectively. Cell viability was assessed using an MTS assay. Cell viability is expressed as percentage compared to non-treated control. Data is represented as mean ± SEM from two or three independent experiments each performed in duplicates.

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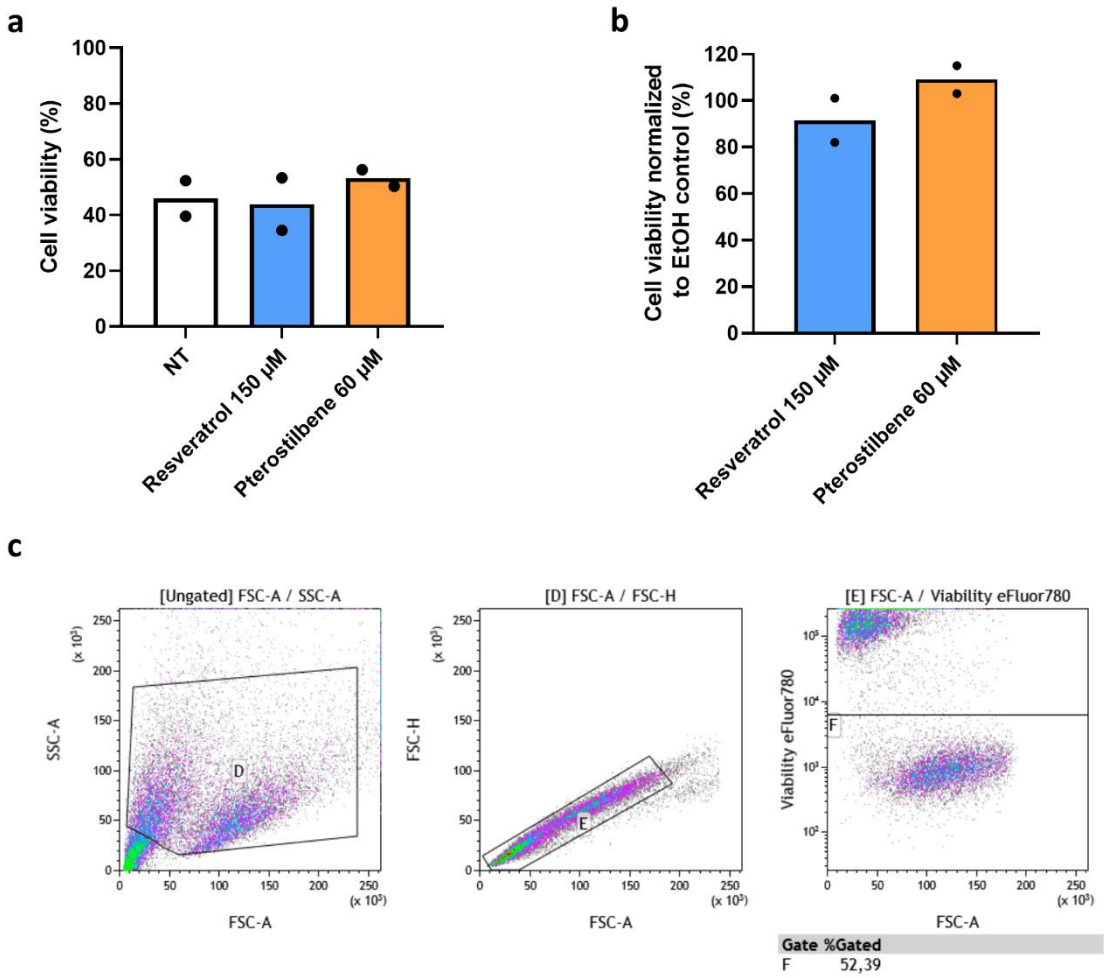

Extended data Fig. 4. Cellular cytotoxicity of resveratrol and pterostilbene in PBEC. PBECs were incubated with 150 µM resveratrol and 60 µM pterostilbene or NT at basolateral side for 8 hr. Cell viability was assessed by flow cytometry using **(a)** life/death staining and by **(b)** LDH assay. **(c)** Gating strategy flow cytometry. Data represents mean from two independent donors.