Effectively control of viral disease outbreak in shrimp culture system by selective predation

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Abstract:

Due to the limited understanding of the characteristics of predator-pathogen-prey interactions, few attempts to use selective predation for controlling diseases in prey populations have been successful. The global pandemic of white spot syndrome (WSS), caused by white spot syndrome virus (WSSV), causes devastating economic losses in farmed shrimp production. Currently, there is no effective control for WSS. Here, we determined the transmission dynamics of WSSV and the feeding ability and selectivity of fish on healthy, infected and dead shrimp by experiments and mathematical modeling. Accordingly, we developed a novel and convenient shrimp cultural ecosystem, which that effectively prevented WSS outbreaks, by introducing aquaculture fish species. This provides a scheme for developing control strategies for viral diseases with high transmission rate.

Predators prefer to eat infected preys in nature, and it is predicted that selective

predation on infected individuals can reduce the prevalence of diseases in prey

population (*1, 2*). The effectiveness of selective predation on disease prevention is

determined by the interplay of several factors (*3, 4*). Without a comprehensive

understanding of characters of predator, pathogen and prey, there are few successful

attempts of using selective predation to control disease in prey population, especially

diseases with high transmission rate (*5-7*).

Aquaculture is the third largest source of animal protein that accounts for 17% protein

consumed by the global population, and shrimps are the global aquaculture species (*8*).

However, white spot syndrome (WSS), caused by white spot syndrome virus (WSSV),

led to huge economic losses to the global shrimp aquaculture industry (*9*). Currently,

there is no effective prevention and control strategies for WSS, though several methods

have been tested in farmed shrimp system (*10-13*). WSS pandemic mainly occurs

depending on the sequential transmission of WSSV from healthy shrimps of consuming

WSSV-infected shrimps to other healthy shrimps (*14, 15*), lacking a predator to remove

the infected individuals from the high density single-species culture system (**fig. S1**).

The prevalence of diseases in prey population is positively correlated with disease

transmission rate, while negatively correlated with predation pressure and predator

selectivity (*16, 17*). Thus, to develop a shrimp culture system which controls WSSV

outbreak by selective predation, we studied WSSV transmission dynamic in the Pacific

white shrimp (*Litopenaeus vanmamei*) population and elucidated the feeding ability

and selectivity of diverse species of aquaculture fishes on shrimps.

To unravel the transmission dynamic of WSS, we carried out the studies on the

relationship among the body weight of one initial WSSV-infected shrimp, number of

deaths, and death time distribution. One piece of WSSV-infected dead shrimp infected

several healthy shrimps with the same bodyweight via ingestion. The infected number

of shrimps in groups of 1.98g, 6.13g, and 7.95g of average body weight were 57.3, 64.7

and 71.3, respectively. (**fig S2 and table S1**). This suggests the basic reproduction

number (R_0) of WSSV increases with the body weight of WSSV-infected dead shrimps.

Time to death was consistent across the three groups of body weight for WSSV-infected

shrimps, with the majority on the third to sixth day and the peak number of deaths on

the fourth and fifth days (**Figs. 1A and S3, and tables S2 to S4**). A mathematical model

(Model 1) was developed to describe the transmission dynamic of WSS (**Figs. 1A and**

1B, **supplementary text**). In addition, the changes of live and dead shrimp numbers

during WSSV transmission were determined by artificial infection experiments. The

number of live shrimps began to decrease after 2 days of WSSV infection, and

drastically decreased after 4 days of WSSV infection (**Fig. 1C**). The dynamic changes

of healthy, infected, and dead shrimps could be expressed by a mathematical model

(Model 2) (**figs. S4 and S5**, **supplementary text**). Model 2 predicts that it is possible

to cut off the transmission route of WSSV by removing infected and dead shrimps, but

the time window of prevention is about 2 days. Thus, the fish used as predator has to

ingest infected and dead shrimps promptly and continuously.

To identify the fish species for controlling WSSV transmission, we examined the

feeding ability and selectivity of diverse aquaculture fish species that ingest Pacific

white shrimps, including grass carp (*Ctenopharyngodon idella*), African sharptooth

catfish (*Clarias gariepinus*) (hereafter catfish), red drum (*Sciaenops ocellatus*), and

brown-marbled grouper (*Epinephelus fuscoguttatus*). The brown-marbled grouper did

not ingest dead shrimp continuously, and its daily dead shrimp ingestion rate is low

(**table S5**). The daily dead shrimp ingestion rates of grass carp, catfish and red drum is

8.26%, 4.99%, and 11.63% respectively (**tables S6 to S8**). The daily healthy shrimp

ingestion rate of grass carp, catfish and red drum is 2.09%, 1.01% and 6.04%

respectively, indicating the relatively high healthy shrimp ingestion rate of red drum

(**tables S9 to S11**). Grass carp and catfish have high feeding selectivity of dead shrimps

over infected and healthy shrimps (**Figs. 2A and S6, and tables S12 and S13**).

Additionally, grass carp and catfish move quickly and swallow infected and dead

shrimps completely. This suggests grass carp and catfish have high feeding selectivity

and ability, which can cut off the WSSV transmission route in which the healthy

shrimps ingest infected dead shrimps.

The capacity and specification of fish were determined for effective control of WSS.
Firstly, we identified the minimum body weight of co-cultured grass carp and catfish.
Four experimental ponds were set up, in which 600 healthy and 3 WSSV-infected
shrimps with body weight of 5 g were cultured with one grass carp of different body
weights. After 13 days in culture, the ponds co-cultured with one grass carps weighted
0.3 kg, 0.5 kg, 1 kg, 1.5 kg showed 0, 0, 82.4%, 79.4% of shrimp survival rate,
respectively (**Fig. 2B and table S14**). This indicates that the minimum body weight of
grass carp for effective control of WSS is 1 kg. Furthermore, experiments showed co-
culturing of catfish of body weight greater than 0.5 kg could effectively control the
WSS transmission in the shrimp population. (**fig. S7 and table S15**). Secondly, we
determined the capacity of grass carp for WSS prevention in shrimp populations of
different shrimp body weights based on experimental results. In a 10 m^2 pond with 750
shrimps, one 1 kg grass carp could control 70 pieces of 2.5 g WSSV-infected shrimps,
50 pieces of 5.0 g WSSV-infected shrimps, or 30 pieces of 7.8 g WSSV-infected
shrimps, which is consistent with the result of mathematical simulation (Model 3) (**Fig.
2C and table S16, supplementary text**). This suggests the capacity of grass carp for
control of WSS is negatively correlated with the body weight of shrimps. Thus,
releasing fishes in the early stages of shrimp production should improve their power for
controlling WSS. Lastly, the minimum stocking quantity of grass carp was determined
for effectively controlling WSS. We released 45-, 150-, 225-, 300-, 450-, 600- and 750-
grass-carps/hm^2, of bodyweight of 1 kg~1.25 kg in the 40 ponds (13.57 hm^2) at a
demonstration farm (**Figs. 2D and S8, and table S17**). When grass carp stocking

quantities were greater than or equal to 300-grass-carp/hm^2, the success rate of WSS

control was 100%.

We next translated the knowledge gleaned from experiments to an applied technology

scale. Firstly, the effectiveness of controlling WSSV transmission by co-culturing

shrimps with grass carp was tested in two experimental zones at a farm in Maoming,

Guangdong province, China (Farm 1) in 2011 (**Fig. 3A**). In the 18 ponds (6.03hm^2) of

zone A, we cultured 9x10^5/hm^2 of shrimps for 20 days, and then introduced 317–

450/hm^2 of grass carps with average body weight of 1 kg. We did not observe the WSS

outbreak in 17 ponds and harvested with 7,332 ± 2,059 kg/hm^2 in 110 days of culture

(**Fig. 3B and table S18**). One pond was unsuccessful due to pathogenic bacterium

(vibrio) infection. Shrimps were cultured without grass carps in 28 ponds (11.30hm^2)

of zone B. A total of 20 ponds in zone B had WSS outbreak, resulting in an average

yield of 1,844 ± 2,304 kg/hm^2. In 2012, we switched zones A and B, cultured shrimps

with grass carps in zone B but without fishes in zone A (**Fig. 3B and table S18**). In all

ponds of zone B, we did not observe the WSS outbreak and harvested with 8,587 ±

1,655 kg/hm^2 in 110 days of culture. Whereas, the average yield of zone A is 1,953 ±

2,188 kg/hm^2 due to the WSS outbreak occurred in 12 of 18 ponds. Secondly, to

evaluate the effectiveness of controlling WSSV transmission by co-culturing shrimps

with catfish, two experimental zones were designed at a farm in Qinzhou, Guangxi

province, China (Farm 2) in 2011 (**Fig. 3C**). In zone A, we cultured 7.5x10^5/hm^2 of

shrimps in 38 ponds (21.20hm^2) for 10 days, and then introduced 525–750/hm^2 of

catfish with average body weight of 0.5kg. Shrimps were cultured in 57 ponds (67.00hm^2) without fish in zone B. In zone A, we did not observe the WSS outbreak in all the ponds and harvested with 8730 ± 1187 kg/hm^2 in 110 days of culture (**Fig. 3E and table S19**). However, the WSS outbreak occurred in 53 ponds of zone B, resulting in an average yield of 450 ± 1420 kg/hm^2. In 2012, we split zone B into zone B1 and B2. Shrimps were cultured with catfish in 38 ponds of zone A and 25 ponds (27.00 hm^2) of zone B1, while shrimps were cultured without fish in 32 ponds (40.00 hm^2) of zone B2 (**Fig. 3D**). We did not see a WSS outbreak and harvested with 9628 ± 1471 kg/hm^2 in zone A and 6375 ± 1000 kg/hm^2 in zone B1. Nevertheless, the WSS outbreak occurred in 29 of 32 ponds of zone B2, resulting in the average yield of 500 ± 1900 kg/hm^2 (**Fig. 3E and table S19**). Moreover, releasing grass carp and/or catfish effectively controlled WSS outbreak and substantially increased the shrimp production at Farm 1 from 2013 to 2019 (**Fig. 3F and data S1**).

Theory predicted that selective predation on infected individuals can prevent outbreaks of diseases in prey population (*1, 18*). By elucidating the transmission dynamics of WSSV, we developed a cultural system to control disease outbreak in shrimp population by restoring the interaction of predator and prey. In addition, the species, body weight, and density of co-cultured fishes were determined by resolving the feeding ability and selectivity of fishes on shrimps through experiments. This, to our knowledge, is the first artificial system that successfully controls viral disease with high transmission rate by selective predation. Furthermore, the use of aquaculture fish as

predator results in production of shrimps and fishes simultaneously in the system. It highlights the significance and provides a scheme of unraveling the characters of predator, pathogen, and prey in developing disease control strategies by selective predation.

References and Notes:

1. C. Packer, R. D. Holt, P. J. Hudson, K. D. Lafferty, A. P. Dobson, Keeping the herds healthy and alert: implications of predator control for infectious disease. *Ecol Lett* **6**, 797-802 (2003).
2. P. T. J. Johnson, D. E. Stanton, E. R. Preu, K. J. Forshay, S. R. Carpenter, Dining on disease: How interactions between infection and environment affect predation risk. *Ecology* **87**, 1973-1980 (2006).
3. S. R. Hall, M. A. Duffy, C. E. Caceres, Selective predation and productivity jointly drive complex behavior in host-parasite systems. *Am Nat* **165**, 70-81 (2005).
4. R. D. Holt, M. Roy, Predation can increase the prevalence of infectious disease. *Am Nat* **169**, 690-699 (2007).
5. M. A. Duffy, Selective predation, parasitism, and trophic cascades in a bluegill-Daphnia-parasite system. *Oecologia* **153**, 453-460 (2007).
6. F. D. Uehlinger, A. C. Johnston, T. K. Bollinger, C. L. Waldner, Systematic review of management strategies to control chronic wasting disease in wild deer populations in North America. *BMC Vet Res* **12**, 173 (2016).
7. S. R. Hall, A. J. Tessier, M. A. Duffy, M. Huebner, C. E. Caceres, Warmer does not have to mean sicker: temperature and predators can jointly drive timing of epidemics. *Ecology* **87**, 1684-1695 (2006).
8. FAO, "The state of world fisheries and aquaculture 2020: Sustainability in action," (Rome, 2020).
9. K. F. Clark, "Nimaviruses of Crustaceans" in *Aquaculture Virology,* F. S. B. Kibenge, M. G. Godoy, Eds. (Academic Press, Amsterdam, 2016), chap. 26, pp. 397-413.
10. J. G. He *et al.*, Theory and strategies for controlling white spot syndrome (WSS) of cultured *Penaeus monodon* in South China. *Acta Scientiarum Naturalium Universitatis Sunyatseni* **39**, 147-153 (2000).
11. J. Witteveldt, C. C. Cifuentes, J. M. Vlak, M. C. van Hulten, Protection of *Penaeus monodon* against white spot syndrome virus by oral vaccination. *J Virol* **78**, 2057-2061 (2004).
12. Y. Wu *et al.*, Inhibition of white spot syndrome virus in *Litopenaeus vannamei* shrimp by sequence-specific siRNA. *Aquaculture* **271**, 21-30 (2007).

13.   V. Alday-Sanz, Specific Pathogen Free (SPF), Specific Pathogen Resistant
(SPR) and Specific Pathogen Tolerant (SPT) as Part of the Biosecurity Strategy
for Whiteleg Shrimp (*Penaeus vannamei* Boone 1931). *Asian Fisheries Science*
31S, 112–120 (2018).
14.   C. F. Lo *et al.*, White spot syndrome baculovirus (WSBV) detected in cultured
and captured shrimp, crabs and other arthropods. *Dis. Aquat. Org.* **27**, 215-225
(1996).
15.   J. G. He, H. M. Zhou, B. Yao, X. M. Yang, M. Deng, White spot syndrome
baculovirus (WSBV) host range and transmission route *Acta Scientiarum*
Naturalium Universitatis Sunyatseni **38**, 65-69 (1999).
16.   M. A. Duffy, S. R. Hall, A. J. Tessier, Selective predators and their parasitized
prey: Are epidemics in zooplankton under top-down control? *Limnol. Oceanogr.*
50, 412-420 (2005).
17.   M. Roy, R. D. Holt, Effects of predation on host-pathogen dynamics in SIR
models. *Theor Popul Biol* **73**, 319-331 (2008).
18.   R. M. Anderson, R. M. May, The population dynamics of microparasites and
their invertebrate hosts. *Phil. Trans. R. Soc. Lond. B* **291**, 451–524 (1981).

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Fig. 1. Transmission dynamics of WSSV. (A) Daily death percentage of shrimp populations with initial WSSV-infected shrimps of different body weights. The red points with error bars are the results from experiments, and the solid blue lines are the results of model 1. **(B)** The illustration shows the death percentages of infected shrimps with different body weight each day. We estimate the parameters in the mode 1 and draw the above 3D surface to show the relationship between death percentage concerning shrimp weights and time. For all weights of shrimps, the death percentage rises at the beginning and then drops. The peak time for death is around the fourth day. **(C)** The relationship between the number of survived shrimps and days after healthy shrimps were co-cultured with WSSV-infected shrimps. Means and standard errors are shown.

Fig. 2. Specification of grass carp for control of WSSV transmission. (A) Feeding
selectivity of grass carp on dead, infected (endopod and exopod removed), and healthy
shrimps. The diseased shrimps which infected WSSV would have reduction of activity
and die within one day. The activity of shrimps reduced after the endopod and exopod
removed. Thus, the endopod and exopod removed shrimps were used to resemble
WSSV-infected shrimps. Grass carp ingested significantly more dead shrimps than
infected (endopod and exopod removed) and healthy shrimps. **(B)** The effect of the
different body weights of grass carps on the control of WSS outbreak. Means and
standard errors are shown. **(C)** Capacity of 1kg grass carp for controlling WSS. Blue
asterisks represent the number of infected shrimps successfully controlled by one 1-kg
grass carp, while clear circles represent the number of infected shrimps that failed to be
controlled by one 1-kg grass carp. The red line is simulated highest value of one 1-kg
fish that can control the number of infected shrimps with different body weights of
infected shrimps based on the model 3. **(D)** The relationship of the number of co-
culturing grass carp and the occurrence rate of WSS. More than 300 grass carps of about
1.0kg per hectare can completely control the outbreaks of WSS, but less than 225 grass
carp cannot fully control outbreaks of WSS.

Fig. 3. Control of WSS in shrimp production by fish. (A) The design of field study for control of WSS using grass carp. The satellite map of the farm at Maoming, Guangdong Province, China (Farm 1) is shown. The 46 experimental ponds were divided into zone A (red) and zone B (blue). In 2011, shrimps were cultured with grass carps in the ponds in area A, while shrimps were cultured without fishes in the ponds in area B. In 2012, shrimps were co-cultured with grass carp in area B but without fishes in area A. **(B)** Total yield of shrimp production in ponds with (red) or without grass carp (blue) at Farm 1. Means and standard errors are shown. **(C)** The design of field study for control of WSS using catfish in 2011. The satellite map of the farm in Qinzhou, Guangxi province, China (Farm 2) is shown. The 95 experimental ponds were divided into zone A (red) and zone B (blue). Shrimps were cultured with catfishes in the ponds in area A, while shrimps were cultured without fishes in the ponds in area B. **(D)** The design of field study for control of WSS using catfish in 2012. Shrimps were continued to be cultured with catfishes in the ponds in area A, while area B was divided into two: shrimps were cultured with catfishes in the ponds in area B1, and shrimps were cultured without fishes in the ponds in area B2. **(E)** Total yield of shrimp production in ponds with (red) or without catfish (blue) at Farm 2. Means and standard errors are shown. **(F)** Total yield of shrimp production in ponds with (red) or without fishes (blue) at Farm 1 from 2013 to 2019. Means and standard errors are shown.