

Inferring the Total-Evidence Timescale of Marattialean Fern Evolution in the Face of Model Sensitivity

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Abstract.—Phylogenetic divergence-time estimation has been revolutionized by two recent developments: 1) total-evidence dating (or “tip-dating”) approaches that allow for the incorporation of fossils as tips in the analysis, with their phylogenetic and temporal relationships to the extant taxa inferred from the data, and 2) the fossilized birth-death (FBD) class of tree models that capture the processes that produce the tree (speciation, extinction, and fossilization), and thus provide a coherent and biologically interpretable tree prior. To explore the behaviour of these methods, we apply them to marattialean ferns, a group that was dominant in Carboniferous landscapes prior to declining to its modest extant diversity of slightly over 100 species. We show that tree models have a dramatic influence on estimates of both divergence times and topological relationships. This influence is driven by the strong, counter-intuitive informativeness of the uniform tree prior and the inherent nonidentifiability of divergence-time models. In contrast to the strong influence of the tree models, we find minor effects of differing the morphological transition model or the morphological clock model. We compare the performance of a large pool of candidate models using a combination of posterior-predictive simulation and Bayes factors. Notably, an FBD model with epoch-specific speciation and extinction rates was strongly favored by Bayes factors. Our best-fitting model infers stem and crown divergences for the Marattiales in the mid-Devonian and Late Cretaceous, respectively, with elevated speciation rates in the Mississippian and elevated extinction rates in the Cisuralian leading to a peak diversity of ~ 2800 species at the end of the Carboniferous, representing the heyday of the Psaroniaceae. This peak is followed by the rapid decline and ultimate extinction of the Psaroniaceae, with their descendants, the Marattiaceae, persisting at approximately stable levels of diversity until the present. This general diversification pattern appears to be insensitive to potential biases in the fossil record; despite the preponderance of available fossils being from Pennsylvanian coal balls, incorporating fossilization-rate variation does not improve model fit. In addition, by incorporating temporal data directly within the model and allowing for the inference of the phylogenetic position of the fossils, our study makes the surprising inference that the clade of extant Marattiales is relatively young, younger than any of the fossils historically thought to be congeneric with extant species. This result is a dramatic demonstration of the dangers of node-based approaches to divergence-time estimation, where the assignment of fossils to particular clades are made *a priori* (earlier node-based studies that constrained the minimum ages of extant genera based on these fossils resulted in much older age estimates than in our study) and of the utility of explicit models of morphological evolution and lineage diversification.

[Bayesian model comparison; Carboniferous; divergence-time estimation; fossil record; fossilized birth-death; lineage diversification; Marattiales; models of morphological evolution; *Psaronius*; RevBayes]

1 The ability to infer phylogenies with branch lengths
2 in units of time (“divergence-time estimation”) is an ex-
3 tremely powerful tool of evolutionary biology. Beyond
4 simply allowing for the inference of the timing of evo-
5 lutionary divergences, it enables studies of diversifica-
6 tion rates and rates of molecular evolution, permits test-
7 ing of the drivers of global patterns of biodiversity and

biogeography (*e.g.*, the roles of vicariance and disper- 8
sal), and allows us to examine the evolutionary impact 9
of major events in the Earth’s history. Extensive research 10
over the past two decades has dramatically improved our 11
ability to infer time-scaled phylogenies (see [Donoghue 12](#)
and [Yang 2016](#)), such that researchers today can choose 13
from a wide variety of models that relax the molecu- 14

lar clock (Sanderson 1997; Thorne et al. 1998; Magallón 2004; Drummond et al. 2006; Drummond and Suchard 2010; Lartillot et al. 2016) and from sophisticated methods for associating fossil data with nodes on a phylogeny (Marshall 2008; Ho and Phillips 2009; Heath 2012; Rothfels et al. 2015a). Molecular dating techniques, however, are fraught with controversy, and their application remains contentious (e.g., Graur and Martin 2004; Wheat and Wahlberg 2013; Wilf and Escapa 2014; Cracraft et al. 2015; Mitchell et al. 2015; Wang and Mao 2015). Much of this controversy is due to the difficulties inherent in accurately associating data from the fossil record with particular nodes in a phylogenetic tree, as is required by the dominant method of divergence-time estimation, the “node-dating” approach. In a node-dating analysis, the investigator associates fossils (or other sources of temporal information, such as island ages, or age estimates from prior studies) with particular nodes in a phylogenetic tree, and provides a calibration density for each node that reflects the investigator’s belief about the temporal relationship between the calibrated node and its constraining fossil (Ho and Phillips 2009).

In a conceptual departure from node-dating approaches, the increasingly popular “tip-dating” or “total-evidence dating” (TED) methods treat fossils as their own terminals in the phylogenetic analysis and jointly infer the placement of the fossils, the patterns of morphological evolution, and a time-calibrated phylogeny. These methods use a dataset containing molecular characters for extant taxa and morphological characters for both extant and fossil taxa (Lee et al. 2009; Pyron 2011; Ronquist et al. 2012; Sterli et al. 2013). TED approaches thus overcome the main weaknesses of node-based methods: the phylogenetic affinities of calibrating fossils are inferred from the morphological data rather than depending on researchers’ implicit assumptions or separate cladistic analyses; the temporal connection between a fossil and a portion of the tree of extant species is not determined in advance; and many more fossils can be used, including fragmentary ones and ones that are members of wholly extinct clades. More generally, these approaches allow for the incorporation of a much greater proportion of the fossil record and shift divergence-time estimation toward a less subjective treatment of fossil data.

For these reasons, TED methods offer great promise for more transparent inferences of divergence time. However, inferring the topological and temporal position of fossils in a Bayesian framework requires the investigator to specify a model of discrete morphological evolution and a tree model that includes non-contemporaneous tips. The morphological model most frequently used—the Mk model (Lewis 2001)—assumes that rates of change between character states are the same for all characters, and has been criticized as inadequate for mod-

eling morphological evolution (e.g., Sterli et al. 2013; Goloboff et al. 2019). Among the tree models, the fossilized birth-death models (Heath et al. 2014; Zhang et al. 2016; Gavryushkina et al. 2017) are an advance over the earlier uniform tree model (Ronquist et al. 2012) in the sense that they provide a coherent mechanistic description of diversification and preservation (Marshall 2019), but they are still limited in biologically important ways, and the relative impacts of these tree models on inference under TED have not been exhaustively explored.

In addition to the modeling concerns specific to total-evidence dating, TED methods share an unusual statistical pathology with other divergence-time estimation methods: nonidentifiability (dos Reis and Yang 2012; Zhu et al. 2015). Under the standard models of character evolution (continuous-time Markov chains), there is no information in molecular or morphological data about either rate or time individually; it is only their product (e.g., number of substitutions per site) that can be estimated. There are thus an infinite number of combinations of rate and time that have identical likelihoods for a given dataset, and the goal of divergence-time estimation—to isolate time from rate—depends on the priors on node ages and clock rates when performed in a Bayesian framework. This fundamental nonidentifiability of relaxed-clock models is apparent in node-dating analyses (Zhu et al. 2015), but is shared by TED analyses, too: the same data (character alignments) can be equivalently fit by very different combinations of model parameters, and those different model parameters could potentially lead to very different branch length inferences. There is thus a need for rigorous analyses of the behaviour of TED methods, for biologically meaningful priors, and for the development of general tools for evaluating model performance (Wilf and Escapa 2016).

Here we explore the sensitivity of TED analyses to these modeling choices, focusing on marattialean ferns (Marattiales: Polypodiopsida). These ferns have a deep fossil record that extends back more than 320 million years (see extensive review by Rothwell et al. 2018a); their relatively modest extant diversity (approximately 110 species; Murdock 2008a; Schuettpelz et al. 2016) belies their former dominance, especially during the Pennsylvanian when they were canopy dominants in both clastic and peat swamp communities (Clea 2015; DiMichele and Phillips 1996, 2002; Phillips et al. 1985). Apparent diversity then declined in the Triassic through Cretaceous, and no unequivocal records exist from the Cenozoic (Lundgren et al. 2019; Rothwell et al. 2018a). This temporal pattern—clusters of extinct and extant diversity separated by a depauperate intermediate sample—as well as extensive morphological homoplasy among the extant genera (Murdock 2008b; Lehtonen et al. 2020) not only make traditional node-dating approaches

123 effectively impossible, but may also challenge many of
124 the assumptions of total-evidence dating models. In this
125 study, we employ the statistical tools of sensitivity anal-
126 ysis, model adequacy, and model comparison to evalu-
127 ate the impact of modeling choices, to improve estimates
128 of marattialean phylogeny and divergence times, and to
129 learn about the processes driving marattialean evolution
130 and diversification.

131 MARATTIALES

132 Marattialean ferns constitute one of the eusporangiate
133 fern clades, and while their relationship to other ferns
134 has historically been unclear (e.g., Schuettpelz et al. 2006;
135 Qiu et al. 2007; Lehtonen 2011), an emerging consensus
136 places them as the sister group to the largest group of ex-
137 tant ferns, the leptosporangiates (Rai and Graham 2010;
138 Kuo et al. 2011; Grewe et al. 2013; Knie et al. 2015; Roth-
139 fels et al. 2015b; Kuo et al. 2018; Qi et al. 2018; Lehtonen
140 et al. 2020; but see Wickett et al. 2014; Shen et al. 2018;
141 One Thousand Plant Transcriptomes Initiative 2019). The
142 approximately 110 extant species of Marattiales are di-
143 vided among six genera (Murdock 2008a; Schuettpelz
144 et al. 2016). These ferns are homosporous, tropical in
145 distribution, and range in size from megaherbs (species
146 of “king fern” in the genus *Angiopteris* can have leaves
147 exceeding 3m in length) to the small dimorphic-leaved
148 species of *Danaea* (Fig. S1).

149 Fossil taxa of the Marattiales are generally attributed
150 to one of two families: the extinct Psaroniaceae and the
151 still living Marattiaceae (see Rothwell et al. 2018a). The
152 Marattiales fossil record includes spores, compressions,
153 and impressions, but they are best known from Carbonif-
154 erous taxa that were carbonate permineralized and de-
155 scribed in anatomical detail, including whole-plant re-
156 constructions (Fig S1E, M) The extensive record of the
157 Marattiales, in its richness, extensiveness, and temporal
158 extent (Millay 1997; Liu et al. 2000; Rothwell et al. 2018a),
159 makes this lineage an ideal test case for divergence-time
160 estimation.

161 In addition, the morphology of marattialeans is com-
162 paratively well-studied (e.g., Stidd 1974; Millay 1979;
163 Millay and Taylor 1984; Hill and Camus 1986; Millay
164 1997; Liu et al. 2000; Murdock 2008b; Cleal 2015; Rothwell
165 et al. 2018b), the phylogeny of the extant lineages is fairly
166 well understood (Li and Lu 2007; Murdock 2008b; Sen-
167 terre et al. 2014; Lehtonen et al. 2020), and there are pre-
168 existing matrices of molecular and morphological (in-
169 cluding strong fossil representation) characters available
170 for a broad taxon sample Hill and Camus 1986; Murdock
171 2008b; Rothwell et al. 2018b; see also Lehtonen et al. 2020,
172 which was published after our analyses were completed).

DATA

173
174 *Taxon samples.*—Our taxon sample includes fossil and ex-
175 tant members of the Marattiales and their sister clade, the
176 leptosporangiate ferns (Fig S1; summarized in Table S.2).
177 The ingroup sample comprises 45 fossil taxa representing
178 either the Psaroniaceae or Marattiaceae, along with 26
179 extant Marattiaceae; the outgroup samples include nine
180 extant species selected for phylogenetic breadth and six
181 well-understood fossil reconstructions spanning the di-
182 versity of leptosporangiate ferns. We chose from among
183 fossil taxa coded by Rothwell et al. (2018b), excluding
184 taxa with large amounts of missing data that were pri-
185 marily of interest for informing *Scolecoperis* classifica-
186 tion. Additionally, we added three Marattiales species
187 to expand our fossil age representation: *Floratheca apoka-*
188 *lyptica* (early Permian), *Rothwellopteris pectopteroides* (late
189 Permian), and an un-named species from the Lower Cre-
190 taceous assigned to the Marattiaceae by Vera and Césari
191 (2016). Our extant-taxon sample is also based on Roth-
192 well et al. (2018b) but altered to maximize the number
193 of taxa that had both morphological and DNA data by
194 adding five species of *Ptisana* and three species of *Danaea*.

195 In some of the analyses that follow, we used three ad-
196 ditional plant fossils: 1) *Psilophyton crenulatum* (early De-
197 vonian; Doran 1980); 2) *Pertica quadrifaria* (early Devo-
198 nian; Kasper and Andrews 1972), and; 3) *Rhacophyton cer-*
199 *atangium* (late Devonian; Cornet et al. 1976; Dittrich et al.
200 1983; see Supplemental Table S.2). We chose these taxa
201 because they are among the most complete fossil vascular
202 plants and are outside of the Marattiales + leptospor-
203 angiate clade.

204 *Morphological and molecular data.*—Our morphological
205 dataset was largely derived from Rothwell et al. (2018b),
206 which itself relied heavily on Hill and Camus (1986) and
207 Murdock (2008b), and amended as necessary. Our final
208 morphological matrix comprised 98 discrete charac-
209 ters describing anatomy and gross morphology; in total,
210 there were 79 binary characters, 10 three-state char-
211 acters, four four-state characters, three five-state charac-
212 ters, one six-state character, and one seven-state charac-
213 ter. We provide the details of how we assembled and
214 scored our morphological dataset in the Supplemental
215 Material (S§1.1).

216 We used available chloroplast DNA sequences from
217 Murdock (2008b) augmented with additional data for our
218 outgroup taxa. The final dataset comprised 33 species
219 with sequences from four chloroplast markers: *aptB*, *rbcL*,
220 *rps4 + rps4-trnS* spacer, *trnS-trnG* spacer + *trnG* intron
221 (Table S.1).

222 The complete morphological and molecular matrices
223 are available in the Data Dryad repository DOI:X and the
224 GitHub repository <https://github.com/mikeryanmay/>

225 [marattiales_supplemental/releases/tag/1.0](#).

226 METHODS

227 *Models*

228 The Bayesian total-evidence dating model consists of five
229 main components: 1) the molecular substitution model;
230 2) the molecular clock model; 3) the morphological tran-
231 sition model; 4) the morphological clock model, and;
232 5) the tree model. There is a long history of study-
233 ing the impact of substitution and molecular-clock mod-
234 els on phylogenetic inference (Huelsenbeck and Ran-
235 nala 2004; Schenk and Hufford 2010; dos Reis and Yang
236 2012; Zhu et al. 2015). By contrast, the influence of
237 the morphological-transition, morphological-clock, and
238 tree models have received less attention, and earlier
239 studies suggest that these model components can have
240 strong effects on divergence-time estimates (Rothfels and
241 Schuettpehl 2014; Condamine et al. 2015).

242 We performed analyses under a range of
243 morphological-transition, morphological clock, and
244 tree models to explore the relative impact of these three
245 model components on phylogenetic estimates of the
246 marattialean ferns. In total, these analyses comprised
247 72 model combinations: four morphological-transition
248 models \times two morphological-clock models \times nine
249 tree models, as described below (a graphical-model
250 schematic is shown in Fig. 1; graphical model repre-
251 sentations of specific model component are available in
252 Supplemental Material S82).

253 *Substitution model and molecular clock.*—In all analyses,
254 we partitioned the molecular dataset by locus, and then
255 by intronic and exonic regions, and for exonic regions,
256 among codon positions. We assigned an independent
257 GTR+I+ Γ substitution model with four rate categories to
258 each data subset. For the molecular clock, we assumed
259 that branch-specific rates of molecular evolution were
260 uncorrelated and drawn from a shared lognormal prior
261 distribution (the UCLN model; Drummond et al. 2006).

262 *Morphological transition models.*—As with substitution
263 models for molecular evolution (e.g., GTR+I+ Γ), the mor-
264 phological transition model is composed of a model that
265 describes how characters are partitioned, a model that
266 describes how rates vary among characters within a par-
267 tition (the among-character rate variation, or “ACRV”,
268 model), and a model that describes the relative rates of
269 change among character states (the morphological ma-
270 trix model) within a partition.

271 For the morphological partition scheme, we parti-
272 tioned characters based on the number of states—such
273 that there was one subset for binary characters, one for

three-state characters, etc.—for all analyses. Within each
subset, we assumed the characters evolved according to
one of the ACRV and morphological matrix models de-
scribed below; for a given combination of ACRV and ma-
trix model, we assumed each subset evolved under the
same type of model, but with parameter values that dif-
fered among subsets.

We used two different morphological matrix models:
an Mk model (Lewis 2001) that assumes that rates of
change—and therefore stationary frequencies—are equal
among states, and a model that allows the stationary fre-
quencies to vary among character states; the latter model
is equivalent to the F81 model commonly applied to
molecular data (Felsenstein 1981), and we therefore refer
to this model as an F81 model. The Mk model assumes
that relative rates of transition are the same among char-
acter states, such that the stationary frequency of each
state is the same among states *and* among characters; for
example, for binary characters, state “0” in one character
has the same frequency as state “0” in another charac-
ter. Because the relative rates are the same among char-
acters, and the overall rate of evolution is described by
the morphological clock model (described below), the
Mk model has no free parameters. For the F81 model,
we further relaxed the assumption that stationary fre-
quencies are shared across characters using a mixture
model (Wright et al. 2016); we refer to this as the F81
mixture model. In this model, state “0” in one charac-
ter is permitted to have a different stationary frequency
than state “0” in another character. This feature is partic-
ularly important in morphological data because, unlike
molecular data where, for example, a “T” indicates par-
ticular features regardless of which alignment site it oc-
curs in, the naming of morphological character states is
arbitrary, with no commonalities across characters. For
binary characters, we discretized a Beta distribution into
five mixture categories and defined an F81 transition ma-
trix using the value of each category as π_0 . We assumed
a symmetrical Beta distribution with shape parameter
 γ_m ; the symmetry of the distribution guarantees that the
likelihood does not depend on the labeling of the binary
states (i.e., which state is labeled “0” or “1”), and the five
mixture categories guarantees that the middle category
corresponds to a symmetric (Mk) model. For each mul-
tistate subset we drew five sets of stationary frequencies
from a symmetrical Dirichlet distribution with parameter
 γ_m , and also a vector of mixture weights, ω , which define
the prior probability that a character evolves according
to each of the stationary frequencies (Pagel and Meade
2004). We assumed the parameter γ_m was shared among
the Beta distribution for the binary characters and the
Dirichlet distributions for the multistate characters, and
estimated γ_m and ω (one per number of states greater
than two) from the data (Figure S6). For both models we

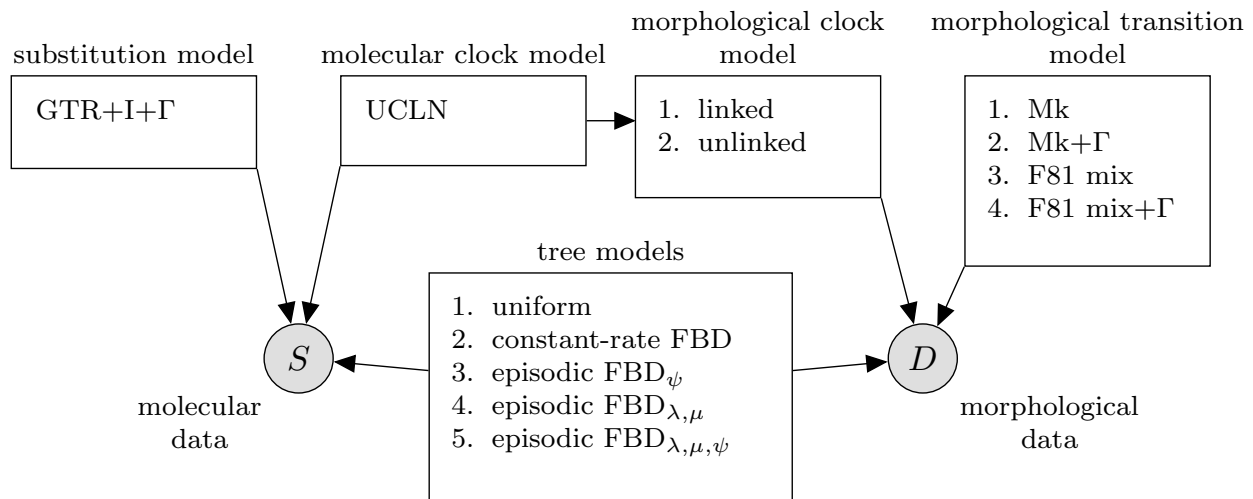


Figure 1: A graphical model representation of the total-evidence dating model. The total-evidence model includes five separate model components: 1) the substitution model; 2) the molecular-clock model; 3) the morphological-transition model; 4) the morphological-clock model, and; 5) the tree model. Each box shows the dependence between each module and the molecular (S) and morphological (D) datasets. GTR+I+ Γ : General time-reversible substitution model with a proportion of invariant sites and gamma-distributed among-site rate variation. UCLN: Uncorrelated lognormal relaxed molecular clock. Mk: Markov transition model with k states. F81 mix: Markov transition model with a mixture of unequal stationary frequencies. FBD: Fossilized birth-death model.

328 corrected for the fact that only variable characters were
329 included (Lewis 2001).

330 In addition to the two morphological transition mod-
331 els, we also modeled how rates of evolution varied
332 among characters within each subset. We used two alter-
333 native among-character rate-variation models: a shared-
334 rate model and a variable-rate model. The shared-rate
335 model assumed that the rate of evolution is the same
336 for all characters with the same number of states. For
337 the variable-rate model, we assumed character-specific
338 rates for characters with i states drawn from a discretized
339 Gamma distribution with four categories and parameter
340 α_i , which we also estimated from the data.

341 In total, we used four different morphological tran-
342 sition models: Mk, Mk+ Γ , F81 mixture, and F81
343 mixture+ Γ .

344 *Morphological clock models.*—The morphological clock
345 model describes how rates of evolution vary among
346 branches in the tree. We used two variants of this model:
347 a linked model where the rate of morphological evolu-
348 tion on a given branch is proportional to the rate of
349 molecular evolution on that branch, and an unlinked
350 model where the morphological and molecular rates are
351 independent. For the linked model, we included a single
352 free parameter, β_m , that defines the relative rate of
353 morphological to molecular evolution across all of the
354 branches (Figure S8). For the unlinked model, the free

parameter β_m describes the relative *mean* rate of mor-
355 phological to molecular evolution. We then drew each
356 branch-specific rate, $r_{m,i}$, independently from a lognor-
357 mal distribution with standard deviation σ_m , which we
358 also estimated from the data (Figure S9).
359

Tree models.—The tree model defines the probability of
360 the tree topology and node ages. We used five differ-
361 ent tree models: 1) a uniform model (Ronquist et al.
362 2012); 2) a constant-rate fossilized birth-death model
363 (CRFBD; Heath et al. 2014); 3) an episodic fossilized
364 birth-death model where fossilization rates were allowed
365 to vary over time (EFBD $_{\psi}$); 4) an episodic fossilized
366 birth-death model where diversification rates (speciation
367 and extinction rates, λ and μ) were allowed to vary
368 over time (EFBD $_{\lambda,\mu}$), and; 5) an episodic fossilized birth-
369 death model where all rates were allowed to vary over
370 time (EFBD $_{\lambda,\mu,\psi}$). For the variable-rate episodic models,
371 we divided time into 33 geological epochs (beginning
372 with the Terreneuvian and ending with the Holocene)
373 defined per the International Chronostratigraphic Chart
374 (updated from Cohen et al. 2013), and allowed one or
375 more parameters to vary among epochs according to a
376 mixture model. Specifically, we assumed that there were
377 three mixture categories, each with their own rate param-
378 eter and a mixture weight κ_i . The assignments of epochs
379 among mixture categories were treated as independent
380 random variables (with values 1 through 3) such that
381

each epoch was assigned to one of the three mixture categories with prior probability κ_i . For a given assignment, each epoch was associated with a specific rate parameter; we then estimated the rate and mixture weight for each category, and the assignment of each epoch among categories. By averaging over the assignment of epochs to each mixture category, this model also provides an estimate of the epoch-specific parameters. For models where multiple rates varied, we assumed that they varied independently (*i.e.*, that they were drawn from separate mixture models). This model differs from previous work where each time slice was allowed to have an independent rate parameter (rather than being drawn from a mixture model), but used a small number of time slices (see Zhang et al. 2016; Wright et al. 2020). Our approach balances model complexity (the number of rate parameters) and the temporal resolution over which rates vary (the number of time slices). For all models, we specified a uniform prior distribution between 410 and 550 Ma on the age of the origin of the tree. We conservatively based the maximum age on the age of the earliest plant fossil evidence (cryptospores from the Ordovician \sim 470 Ma Rubinstein et al. 2010), and the minimum age on the age of the oldest sample in our extended “ancient plants” dataset (see below). Additionally, we accommodated uncertainty in the ages of each fossil, which has been demonstrated to be important for accurate divergence-time estimates (Barido-Sottani et al. 2019).

The final component of the FBD tree models is the mechanism for accounting for incomplete sampling of extant taxa. Our taxon sample includes a relatively well-sampled extant ingroup (27 of 111 extant taxa), and a very sparsely sampled outgroup (nine of \sim 12000; Schuettelpelz et al. 2016). To accommodate this heterogeneous taxon sampling we therefore conducted each fossilized birth-death analysis twice, once with a sampling fraction, ρ , corresponding to the ingroup portion of the tree ($\rho = 27 \div 111$), and once corresponding to the full tree ($\rho = 36 \div 12000$).

In total, we used nine different tree models: the uniform tree model, plus four fossilized birth-death models \times two taxon-sampling fractions.

Analyses

MCMC analyses.—For each combination of the above model components, we estimated the posterior distribution using four replicate Metropolis-coupled MCMC with five coupled chains in RevBayes (Höhna et al. 2016). We performed all of our analyses on the University of California, Berkeley HPC cluster, savio (run times and additional computation details are described in Supplemental Material S§3.2). We assessed whether each chain failed to converge to, or sample adequately from, the joint posterior distribution using protocols described in

the Supplemental Material (S§3.1). All of the scripts used for analysis and post-processing are available in the Data Dryad repository DOI:X and the GitHub repository https://github.com/mikeryanmay/marattiales_supplemental/releases/tag/1.0.

Comparing estimates of topology and branch lengths.—We employed two techniques to summarize differences in phylogenetic estimates under the model combinations that we explored. First, we used multidimensional scaling (MDS) of tree-distance metrics (following Hillis et al. 2005) to compare the distributions of tree topologies and branch lengths inferred under these models. MDS projects the pairwise distance between each tree in a sample of trees into a lower dimensional—and therefore easier to visualize—representation of tree space. To facilitate automated MDS for a large number of comparisons, we implemented these MDS analyses using the R packages phangorn and smacof (de Leeuw and Mair 2009; Schliep et al. 2017; R Core Team 2019). We compared the distribution of phylogenies using MDS with three different metrics: 1) the Robinson-Foulds distance (RF, a measure of topological distance; Robinson and Foulds 1981); 2) the Kühner-Felsenstein distance (KF, a distance metric that incorporates both topology and branch lengths; Kühner and Felsenstein 1994) between time-scaled phylogenies (chronograms), and; 3) the KF distance between phylogenies with branch lengths proportional to the expected amount of morphological evolution (“morphograms”). Computing distances among phylogenies with sampled ancestors is difficult, since the number of tips and branches depends on the inferred number of sampled ancestors. We therefore resolved all sampled ancestors onto zero-length branches before computing distances among trees. We included 100 trees from the posterior distribution of each of the models in our MDS plots, and colored each point in the resulting tree space according to one of the TED model components to visually compare the relative impact of each model component on the posterior distribution of trees.

Second, we used lineage-through-time (LTT) curves to summarize divergence-time estimates under each model. An LTT curve displays the number of branches in the inferred tree at any given time, and therefore provides a less abstract summary of divergence-time estimates than MDS; however, in contrast to MDS plots, the LTT approach is unable to capture differences in topology. We compute the average LTT curve under a given model by calculating the average number of branches present at each time point for each tree in the posterior distribution; similarly, we compute the 95% credible interval (CI) of the number of branches at a given time. To compare the relative impact of the three model components, we compute the average LTT curve for each model, then com-

488 pute the average of these curves among all model combi-
489 nations that share the focal model component.

490 *Stochastic character mapping.*—We used stochastic charac-
491 ter mapping (Huelsenbeck et al. 2003) to visualize pat-
492 terns of morphological character evolution across the
493 phylogeny. In these analyses, we conditioned on the
494 maximum-clade-credibility (MCC) tree inferred for each
495 model, as well as posterior mean estimates of all relevant
496 model parameters, and simulated stochastic maps using
497 the R package *phytools* (Revell 2012).

498 *Implied total diversity over time.*—We simulated lineages
499 under each of the fossilized birth-death models, with-
500 out fossilization, to generate the implied total number of
501 lineages present at each point in time. For each model,
502 we sampled the origin time and diversification paramet-
503 ers from the corresponding posterior distribution, then
504 simulated lineages forward in time until the present to
505 generate the posterior-predictive distribution of the full
506 diversification process. We computed the median and
507 95% CI of the number of lineages at a large number of
508 evenly spaced time points to summarize the distribution
509 of the implied total diversity over time. These simula-
510 tions do not condition on the sampled tree, and therefore
511 should not be interpreted as the posterior distribution of
512 the number of missing taxa in our inferred tree. Rather,
513 these simulations represent the distribution of the num-
514 ber of lineages if we were to repeat the inferred lineage-
515 diversification process many times.

516 *Assessing absolute model adequacy.*—We assessed the ade-
517 quacy of each model using posterior-predictive simula-
518 tion (PPS; Bollback 2002; Höhna et al. 2018). The premise
519 of PPS is that, if a given model provides an adequate de-
520 scription of the true process that generated the observed
521 data, then datasets simulated by the model should re-
522 semble the observed data. The degree of resemblance
523 for a given simulated dataset is described by a summary
524 statistic that is designed to capture relevant aspects of the
525 data-generating process. If the distribution of this statis-
526 tic computed across simulated datasets (the posterior-
527 predictive distribution, PPD) contains the statistic for the
528 observed data with high probability, then the model is
529 deemed adequate, *i.e.*, the model provides a reasonable
530 description of the true data-generating process.

531 We assessed the adequacy of our models from the per-
532 spective of the morphological data. For a given model,
533 we drew random samples from the posterior distribution
534 of model parameters (including the phylogeny and pa-
535 rameters of morphological evolution). For each sample,
536 we simulated a morphological dataset, the same size as
537 the original and with the same patterns of missing data,

538 given the model parameters and computed two sum- 538
539 mary statistics: 1) S , the total parsimony score (number 539
540 of steps computed on the sampled tree) for the simulated 540
541 characters minus the total parsimony score for the ob- 541
542 served characters, and; 2) V , the variance in parsimony 542
543 scores among simulated characters minus the variance 543
544 in parsimony scores among observed characters. The 544
545 S statistic is intended to assess whether the model ade- 545
546 quately characterized the average rate of evolution, while 546
547 the V statistic is intended to assess if the model captures 547
548 how the rate and process of evolution vary among char- 548
549 acters. If the 95% interval of the posterior-predictive dis- 549
550 tribution of these statistics for a given model did not in- 550
551 clude 0, we deemed that model inadequate. We provide 551
552 further details of these simulations in the Supplemental 552
553 Material (S§4). 553

554 The structure of the TED model provides some expecta- 554
555 tions about the sensitivity of PPS to the different model 555
556 components we explored. Specifically, the PPD will be 556
557 sensitive to model components that have a strong im- 557
558 pact on the likelihood function, but insensitive to non- 558
559 identifiable model components. We therefore expect the 559
560 morphological-transition model to influence the PPD be- 560
561 cause this model component is identifiable. By contrast, 561
562 for a given morphological-clock model, the tree model 562
563 may have little influence on the PPD because rate and 563
564 time are nonidentifiable: tree models that prefer dif- 564
565 ferent node ages may nonetheless have similar likeli- 565
566 hoods because the clock model can compensate for dif- 566
567 ferent node ages. However, for a given tree model, the 567
568 two clock models we used can in principle influence the 568
569 PPD because the models with separate relaxed clocks can 569
570 achieve sets of branch rates that models with one relaxed 570
571 clock cannot (Zhu et al. 2015). For example, if the un- 571
572 linked model is true (*i.e.*, if rates of morphological and 572
573 molecular evolution are not proportional), then estimates 573
574 of branch-specific rates of morphological evolution un- 574
575 der the linked model will be influenced by the molecular 575
576 data and therefore be unable to attain values that fit the 576
577 morphological data. 577

578 *Assessing relative model fit.*—We compared the relative 578
579 fit of competing models using Bayes factors, which are 579
580 the ratio of the marginal likelihood of each model (Kass 580
581 and Raftery 1995). We estimated the marginal likelihood 581
582 for a given model using four replicate power-posterior 582
583 analyses in RevBayes, and computing the path-sampling 583
584 (Lartillot and Philippe 2006) and stepping-stone estima- 584
585 tors (Xie et al. 2011). Given the computational expense 585
586 of marginal-likelihood estimation, we only calculated 586
587 Bayes factors among the tree models (conditional on the 587
588 preferred morphological-transition and morphological- 588
589 clock models). 589

590 *Assessing the effect of the taxon sample and rooting.*—Our
591 fossil dataset is dominated by a cluster of late Car-
592 boniferous taxa whose relationships to each other and
593 to surviving lineages are apt to be highly uncertain,
594 which may limit our ability to infer ancient divergences.
595 Additionally, the inclusion of a sparsely sampled out-
596 group makes it difficult to specify an appropriate taxon-
597 sampling fraction for the fossilized birth-death models.
598 We performed additional analyses to understand the ro-
599 bustness of divergence-time estimates within the Marat-
600 tiales to different taxon samples and rooting strategies
601 (described in more detail in the Supplemental Material
602 S§5). In particular, in addition to our “standard” taxon set
603 (the ingroup Marattiales plus an outgroup of leptospor-
604 angiate ferns) we performed analyses with only ingroup
605 taxa, with the addition of the ancient land-plant fossils
606 described above, and by polarizing a subset of charac-
607 ters for which we are confident in their ancestral states *a*
608 *priori* (our “ingroup-only”, “ancient plants”, and “polar-
609 ized” analyses, respectively). In each of these analyses,
610 we assumed the best-performing morphological transi-
611 tion model, morphological clock model, and tree model,
612 as determined in the core analyses.

613 RESULTS

614 *Modeling results*

615 Here, for the fossilized birth-death models, we present
616 the results for the analyses that assume the ingroup sam-
617 pling fraction, for a total of 40 distinct model combina-
618 tions (four morphological transition models \times two mor-
619 phological clock models \times five tree models); results us-
620 ing the overall sampling fraction are qualitatively simi-
621 lar, and are presented in the Supplemental Material S§6.2.
622 LTT curves for each individual model combination are
623 available in the Supplemental Material. We also com-
624 pared the ages of individual clades for each pair of mod-
625 els (Figs. S20, S21, S22), and between ingroup and overall
626 sampling fractions (Fig. S59).

627 *Topological distance.*—The greatest differentiation in topo-
628 logical space is between models with and without the
629 uniform tree model (Fig. 2, top row); the individual fos-
630 silized birth-death tree models each have weak effects on
631 topology (Fig. 2, top right). The morphological transi-
632 tion model has a strong influence on topologies (Fig. 2,
633 top left), whereas the morphological clock models have a
634 mild effect (Fig. 2, top middle).

635 *Chronogram distance.*—Similar to the pattern observed in
636 topological distances, the uniform tree model has the
637 most striking impact on Kühner-Felsenstein distances
638 among chronograms (Fig. 2, middle row). However,

in contrast to topological distances, KF distances among
chronograms are differentiated by the FBD models, with
the EFBD $_{\lambda,\mu}$ and EFBD $_{\lambda,\mu,\psi}$ models sampling from a re-
gion of tree space (Fig. 2, middle right, central region)
that is rarely visited by the other models. The morpho-
logical transition and clock models appear to have a mild
effect on KF distances (Fig. 2, middle row, left and mid-
dle columns).

647 *Morphogram distance.*—In contrast to its weak effect on
648 topological and chronogram distances, the morphologi-
649 cal clock model has the strongest impact on KF distances
650 among morphograms (Fig. 2, bottom middle). Within the
651 clusters defined by the clock models, there is clear differ-
652 entiation among transition models, with the Mk+ Γ and
653 F81 mixture models being intermediate between the Mk
654 and F81 mixture+ Γ models (Fig. 2, bottom left). The tree
655 models have a mild effect on distances among chrono-
656 grams (Fig. 2, bottom right).

657 *Lineages through time.*—Consistent with our MDS plots,
658 the greatest differences among LTT curves are between
659 model combinations with and without the uniform tree
660 model; however, there are also consistent differences in
661 LTT curves among the fossilized birth-death models (Fig.
662 3, right). In particular, the EFBD $_{\lambda,\mu}$ and EFBD $_{\lambda,\mu,\psi}$ mod-
663 els indicate a later origin for the Marattiales and a more
664 rapid increase to peak diversity at the end of the Car-
665 boniferous, whereas the EFBD $_{\psi}$ model predicts a more
666 gradual accumulation of diversity. Overall, the EFBD $_{\lambda,\mu}$
667 and EFBD $_{\lambda,\mu,\psi}$ models estimate very similar LTT curves,
668 and the influence of the tree model on LTT curves de-
669 creases toward the present.

670 The morphological transition model has a mild but
671 consistent impact on lineage-accumulation leading up to
672 the Carboniferous (Fig. 3, left). Following the Carbonifer-
673 ous, the transition models generally result in very similar
674 LTT curves, with the exception of the Mk model, which
675 infers generally fewer lineages through the Cenozoic.

676 Whether rates of morphological evolution are linked or
677 unlinked to rates of molecular evolution has almost no
678 impact on LTT curves in the early history of the Marat-
679 tiales. However, LTT curves for the clock models begin
680 to diverge beginning about 175 Ma, after which the un-
681 linked model infers younger clade ages (Fig. 3, middle).

682 *Absolute model fit.*—The morphological transition model
683 had a strong impact on model adequacy (Fig. S23, col-
684 ored boxplots). In particular, transition models with-
685 out among-character rate variation did the worst ac-
686 cording to the parsimony variance statistic, *V*; addition-
687 ally, the F81 mixture+ Γ model did the best at describ-
688 ing the amount of evolution, according to the total par-
689 simony score statistic, *S*. We therefore identified the

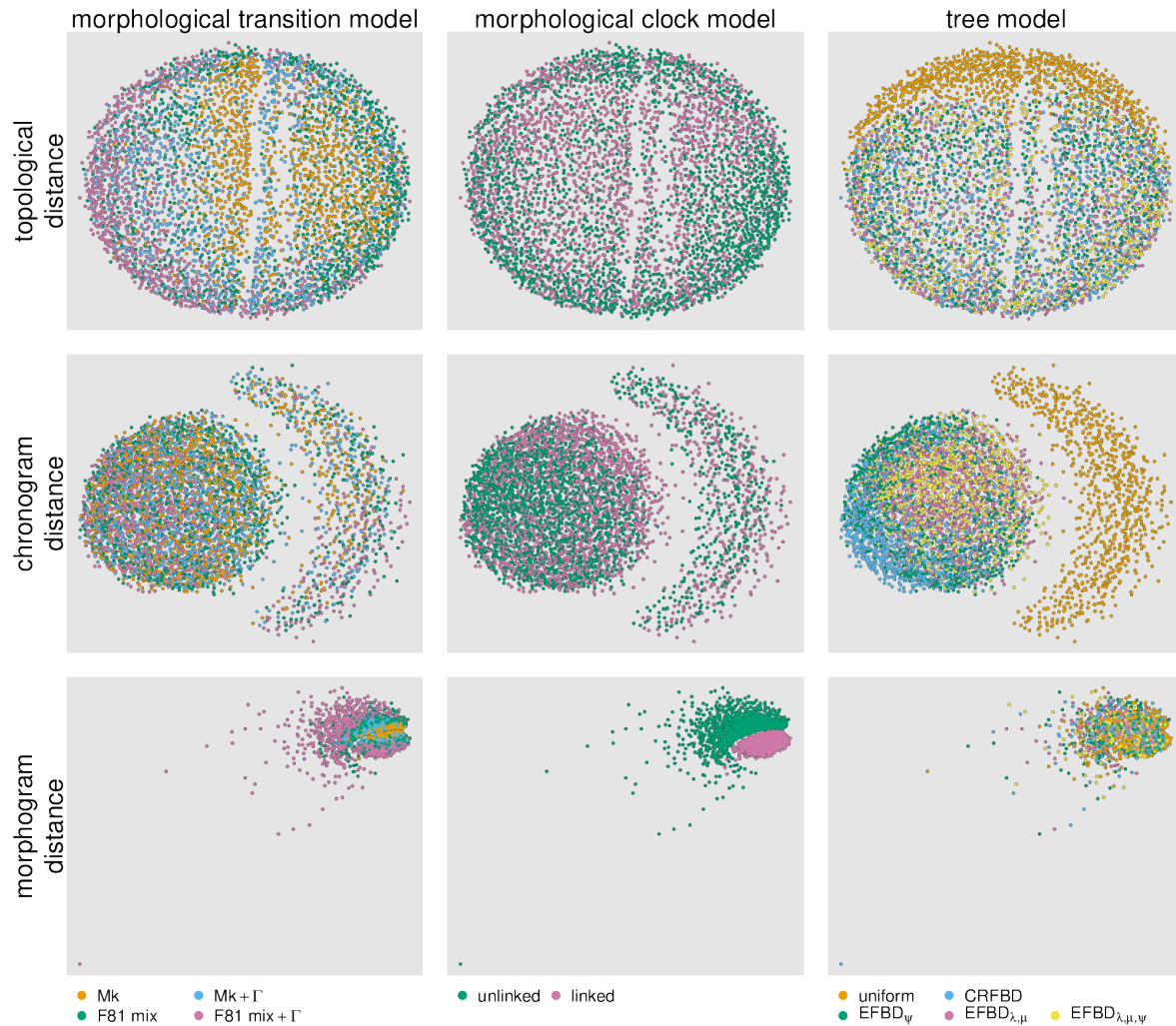


Figure 2: Comparing distributions of trees among model combinations. We compute the Robinson-Foulds distance (RF, a measure of topological distance, top row), the Kühner-Felsenstein distance (KF, a distance metric that incorporates both topology and branch lengths) between chronograms (middle row) and morphological phylograms (“morphograms”, bottom row). We then plot the (square-root transformed) distances in two-dimensional space using multi-dimensional scaling (MDS); each point represents the location of a given sampled tree in tree space according to the distance metric. We color points according to the morphological transition model (column 1), the morphological clock model (column 2), or the tree model (column 3). These results assume the ingroup sampling fraction for all of the fossilized birth-death models; see Figure S30 for the results with the overall sampling fraction.

690 F81 mixture+ Γ as the preferred morphological transition model.
691

692 Posterior-predictive distributions were largely insensi-
693 tive to the morphological clock model (Fig. S23, shaded
694 regions), a result that is in strong contrast to the influence
695 of the clock model on the posterior distributions of morpho-
696 grams (Fig. 2, bottom middle). The strong effect of
697 the clock models on the morphograms indicates that the
698 absence of a signal in the PPS is not related to nonidenti-
699 fiability. Because this model component had a modest ef-
700 fect on divergence-time estimates, we preferred the sim-
701 pler, linked morphological clock model for further anal-
702 yses.

703 As predicted based on nonidentifiability, the tree
704 model had essentially no impact on posterior-predictive
705 distributions (Fig. S23, columns), consistent with its
706 very limited impact on posterior distributions of morpho-
707 grams (Fig. 2, bottom right). We therefore compared
708 the relative fit of the tree models using Bayes factors.

709 We also assessed absolute model fit with either the ex-
710 tinct or extant taxa pruned from the tree and morpholog-
711 ical data before computing the summary statistics. When
712 extinct taxa were removed, the simulated datasets con-
713 sistentlly overpredicted the parsimony score (Fig. S24);
714 by contrast, when extant taxa were removed, simulated
715 datasets had lower parsimony scores than the observed

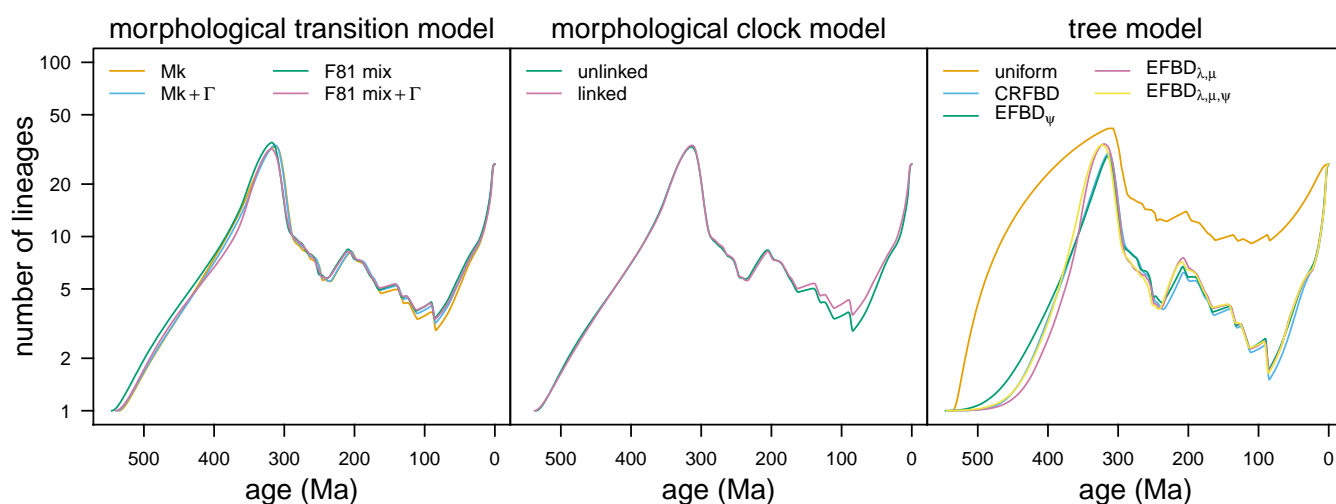


Figure 3: Comparing lineage-through-time curves among model combinations. For each focal model component (morphological transition model, and morphological clock model, and tree model, respectively), we compute the LTT curve averaged over the remaining model components, *i.e.*, the average number of branches in the phylogeny at a given time, averaged over all model combinations that share the focal model component. We removed outgroup taxa to emphasize the influence of model specification on age estimates within our ingroup. Left) We compute the average LTT for each of the 40 models (from 2000 sampled trees for each model), then compute the mean of the resulting average LTT among models with the same morphological-transition model. Middle) As in left, but we compute the mean of the average LTT among models with the same morphological clock model. Right) As in left, but we compute the mean of the average LTT among models with the same tree model.

716 morphological dataset (Fig. S25). The variance in the
 717 parsimony score across characters, V , was relatively ins-
 718 sensitive to the exclusion of either extant or extinct taxa.

719 *Relative model fit.*—Bayes factor comparison of the tree
 720 models were decisive (Table 1). The uniform tree model
 721 is by far the worst model: it is very strongly outper-
 722 formed by the second worst model, the constant-rate
 723 FBD model ($2 \ln \text{BF} > 40$). Interestingly, models that al-
 724 low fossilization rates to vary among epochs do not im-
 725 prove model fit: the EFBD_ψ is disfavored compared to
 726 the constant-rate FBD, and the $\text{EFBD}_{\lambda,\mu,\psi}$ model is dis-
 727 favored compared to the $\text{EFBD}_{\lambda,\mu}$ model. Overall, the
 728 $\text{EFBD}_{\lambda,\mu}$ model is the best-performing model, and is very
 729 strongly favored over the second-best model, $\text{EFBD}_{\lambda,\mu,\psi}$
 730 ($2 \ln \text{BF} = 14.25$; Table 1).

731 *Taxon sampling fractions.*—Results under fossilized birth-
 732 death models assuming the overall sampling fraction
 733 ($\rho = 36 \div 12000$; S§6.2) are qualitatively similar to those
 734 assuming the ingroup sampling fraction ($\rho = 27 \div 111$);
 735 in particular, clade-age estimates under the overall sam-
 736 pling fraction are modestly older than those under the
 737 ingroup sampling fraction (S§6.3).

738 *Taxon sample and rooting analyses.*—The broad topologi-
 739 cal patterns and timing of divergences are similar among
 740 these analyses (Figs. S57, S58, S59). Nevertheless, there
 741 are some notable trends in the effect of the different ap-
 742 proaches on inferred ages of specific clades, which we

discuss in the Supplemental Material (S§5). Notably, the
 similarity between ages inferred from our primary analy-
 ses with the ingroup sampling fraction and those inferred
 under the ingroup-only analysis suggests that age esti-
 mates based on the ingroup sampling fraction are reli-
 able. By contrast, those under the overall sampling frac-
 tion are probably overestimates (S§6.3). Because these
 analyses used different datasets, we can not correctly
 compare among them with posterior predictive simula-
 tion or Bayes factors. However, given that these analyses
 provided broadly consistent results, we ultimately base
 our discussion of marattialean phylogeny on the most in-
 clusive dataset (*i.e.*, the “ancient plants” analysis).

Phylogenetic results

Topological results.—Topologies varied among analyses,
 but all analyses except those under the uniform tree prior
 resolved a clade of extant Marattiaceae (exclusive of any
 fossil representatives) and a grade of fossil taxa diverging
 from the stem of this extant clade that includes a Psar-
 oniaceae clade comprising many Carboniferous to Trias-
 sic (– Cretaceous) taxa (Figs. 4, S26, S43, S52, S55). In
 the MCC tree for our focal phylogeny (from the “ancient
 plants” analysis), *Scolecoperis* species do not fall within
 a single clade (Fig. 4). Instead, most *Scolecoperis* repre-
 sentatives, along with a few other traditional psaronia-
 ceous fossil genera, fall in a “core” Psaroniaceae clade,
 while other *Scolecoperis* species along with *Grandeuryella*
renaulti and *Floratheca apokalyptica* form a small grade of
 lineages at the base of the remaining Marattiales with

Table 1: Comparing the fit of alternative tree models with Bayes factors. We report the average marginal likelihood \pm the standard deviation among four runs computed using the stepping-stone estimator, and the $2 \ln$ Bayes factor between each pair of models. Marginal likelihoods computed using the path-sampling estimator were essentially identical.

MODEL	MARGINAL LIKELIHOOD	2 LN BF AGAINST ALTERNATIVE				
		Uniform	CRFBD	EFBD $_{\psi}$	EFBD $_{\lambda,\mu}$	EFBD $_{\lambda,\mu,\psi}$
Uniform	-25941.74 ± 0.60	–				
CRFBD	-25920.73 ± 0.78	42.03	–			
EFBD $_{\psi}$	-25922.00 ± 0.59	39.49	–2.54	–		
EFBD $_{\lambda,\mu}$	-25908.05 ± 1.02	67.39	25.36	27.90	–	
EFBD $_{\lambda,\mu,\psi}$	-25915.17 ± 0.59	53.14	11.11	13.65	–14.25	–

772 which they share a reduced number of sporangia per
773 synangium (Fig. S70) and spore ornamentation charac-
774 teristics (Figs. S73, S74). The analyses with polarized-
775 character rooting and the ingroup-only dataset produced
776 similar results but with different arrangement of taxa in
777 the grade (Figs. S51, S55).

778 The “standard dataset” results weakly support an al-
779 ternative hypothesis, in which there is an expanded Psar-
780 oniaceae clade that consists of all *Scolecoperis* species
781 plus representatives of other genera, most of which
782 have traditionally been considered to belong to Psar-
783 oniaceae (e.g., *Araiangium pygmaeum*, *Acaulangium bul-*
784 *baceum*, *Grandeuryella renaulti*, *Convexocarpus distichus*,
785 *Floratheca apokalyptica*, *Buritranopteris costata*, and *Gemel-*
786 *litheca saudica*; Fig. S26). This topology does not appear
787 to be supported by obvious character-state transitions.
788 In each case, support for the topology is low, as it is in
789 all fossil-rich regions of the tree (Figs. 4, S26, S51, S55).
790 Support for the monophyly of the ingroup was greatest
791 for the ancient and polarized analyses (i.e., the ones with
792 additional information available to inform the root po-
793 sition; posterior probabilities 0.43 and 0.85, respectively,
794 compared to 0.33 with the standard dataset; Figs. 4, S51).
795 Conversely, the ingroup-only analysis was the most topo-
796 logically distinct in this fossil-rich region of the tree (Figs.
797 S57, S55), and in general has lower posterior support for
798 divergences along the backbone of the tree.

799 The “early diverging” stem lineages related to *Marat-*
800 *tropsis* and extant Marattiaceae consistently include *Rad-*
801 *stockia kidstonii*, *Qasimia shyfsmae*, and *Rothwellopteris*
802 *pecopteroides*, as well as a clade comprising *Millaya tu-*
803 *larosana*, *Eoangiopteris goodii*, and *Danaeites rigida* (Figs. 4,
804 S26, S27, S51, S52, S54, S55, S56). *Escapia christensenioides*
805 and *Danaeopsis fecunda* were highly unstable and inferred
806 in various positions among these stem lineages (Figs. 4,
807 S26, S55) or with *Scolecoperis* species (Fig. S51). Pruning
808 these two taxa did not have an appreciable effect on clade
809 support (not shown).

810 *Marattropsis* fossils, from the Triassic through Jurassic,
811 are morphologically very similar to extant marattialeans
812 and, with “Marattiaceae indet. Vera (2016)”, form a

813 grade of lineages most closely related to the extant taxa.
814 *Qasimia shyfsmae* and/or *Rothwellopteris pecopteroides* are
815 often inferred to be directly ancestral to the *Marattropsis*
816 + extant Marattiaceae clade. Notably, the phyloge-
817 netic separation of the extant Marattiaceae genera from
818 the morphologically nearly indistinguishable *Marattropsis*
819 species (see Fig. S1) and the superficially similar *Dane-*
820 *opsis fecunda* only occurs when temporal data are in-
821 corporated: in a separate analysis under a non-dated tree
822 model (i.e., without temporal data), the support for the
823 monophyly for the extant clade dropped dramatically (to
824 < 0.001 posterior probability, Fig. S76). Relationships
825 within crown Marattiaceae are consistent among analy-
826 ses (Figs. 4, S26, S43, S51, S55). *Danaea* is sister to the
827 remaining genera and *Marattia* is sister to a weakly sup-
828 ported *Christensenia* + *Angiopteris* clade; they are in turn
829 sister to *Eupodium* + *Ptisana*. Our leptosporangiate out-
830 group taxa are themselves also monophyletic, although
831 the position of fossils within the group varies among
832 analyses (Figs. 4, S26, S43, S51), with the Permian *Szea*
833 *sinensis* and in some cases the Triassic *Hopetedia praeter-*
834 *missa* inferred to be directly ancestral to extant taxa.

835 *Divergence-time estimates.*—The overall temporal pattern
836 we infer suggests that the Marattiales and leptosporan-
837 giate fern lineages diverged from each other in the Lower
838 or Middle Devonian (posterior mean 414.84 Ma, 95% CI
839 = [350.90 – 505.74]; Fig. S58, Table S.3).

840 Early-diverging stem lineages of Marattiales, mostly
841 involving taxa traditionally assigned to Psaroniaceae, di-
842 verged through the earliest Permian. Fossil taxa at-
843 tributed to *Marattropsis* begin to diverge in the Triassic
844 (Fig. 4, S51, S55), followed by a gap of ~ 115 million
845 years (Table S.5) before the inferred divergence of the
846 crown Marattiaceae in the Upper Cretaceous, although
847 there is considerable uncertainty around the age of this
848 crown node (posterior mean 85.50 Ma, 95% CI = [37.44 –
849 173.74], Table S.4). While all extant genera are inferred to
850 have diverged from each other by the Eocene, the extant
851 species diverged from each other beginning in the Neo-
852 gene (Figs. 4, S26, S51, S55).

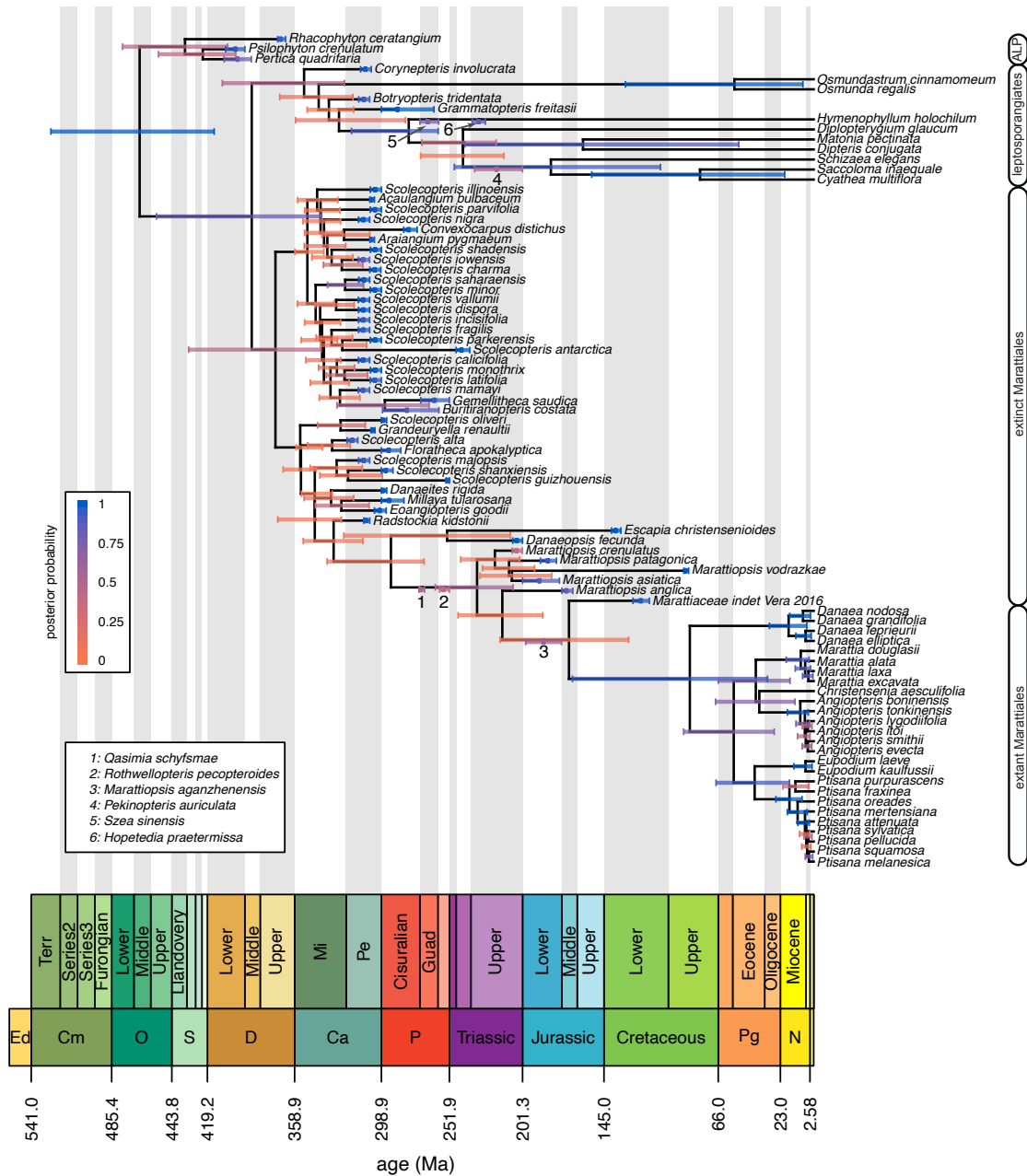


Figure 4: The maximum clade credibility tree under the preferred model with the “ancient plants” analysis. Bars correspond to the 95% credible interval of clade ages (for internal nodes) and tip ages (for fossils). Numbers and associated age bars along internal branches correspond to sampled ancestors (key bottom left). Bars are colored in proportion to the posterior probability of the clade for internal nodes, by the probability that the specimen is not a sampled ancestor for tips, and by the probability that the specimen is a sampled ancestor for sampled ancestors (legend, left). Major groups are labeled (right; ALP = ancient land plants). We plot time intervals according to the International Chronostratigraphic Chart (2020, updated from [Cohen et al. 2013](#)).

DISCUSSION

Our results bear both on broad methodological issues, and on details of marattialean phylogeny and biology. We begin by discussing the consequences of model specification on total-evidence dating, as well as prospects for future model and method development. Next, we dis-

cuss the implications of our results in the context of existing work on the phylogeny of Marattiales. Finally, we reconcile our methodological results with prior knowledge about the fossil record and marattialean paleoecology to draw a synthetic inference about the evolutionary history of the group, and to demonstrate the harmony between our inferences—particularly under the fossilized

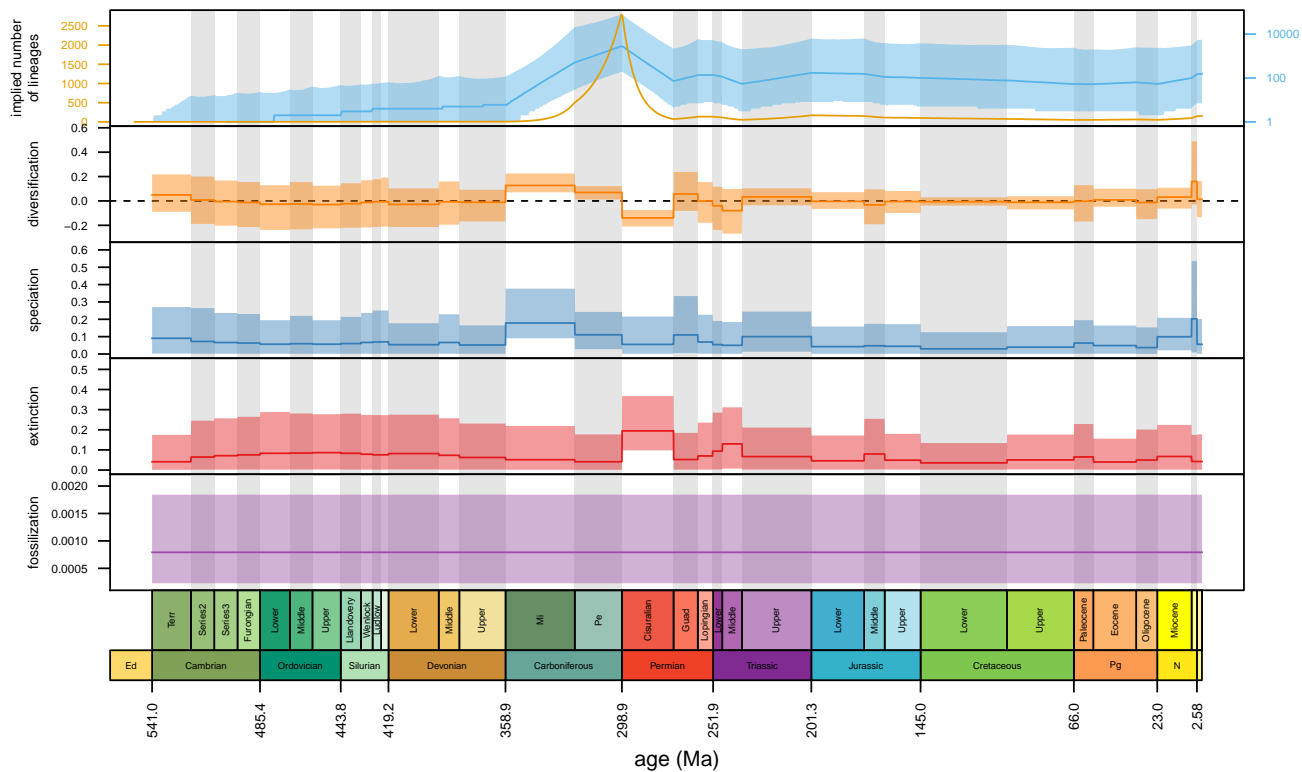


Figure 5: Diversification over time under the preferred model. The top panel shows the implied total number of lineages over time (before pruning unsampled lineages); the median value is shown on the linear scale in orange, and the median value and 95% credible interval are shown on a log scale, in blue. The next four panels correspond to the posterior distribution of epoch-specific net-diversification rates, speciation rates, extinction rates, and fossilization rates, respectively. In the top panel, the dark line represents the posterior median estimate, in the remaining panels it represents the posterior mean estimate. In all cases, and the shaded region corresponds to the 95% credible interval. Note that the preferred tree model, $\text{EFBD}_{\lambda,\mu}$, assumes that fossilization rates are constant (contrast against the model with fossilization-rate variation, Figure S28).

866 birth-death model—and the fossil record.

867 *Model specification and total-evidence dating*

868 The fundamental advance of total-evidence dating is that
 869 fossil specimens are included as extinct samples in the
 870 tree, and their topological and temporal relationships
 871 to extant lineages are inferred from the available data,
 872 rather than assumed *a priori* (or inferred in separate
 873 cladistic analyses) as in traditional node-based fossil
 874 calibration approaches. Estimating the phylogenetic posi-
 875 tion of fossils requires that we specify models that de-
 876 scribe how morphological characters evolve, how rates of
 877 morphological evolution vary among lineages, and how
 878 lineages are distributed in time (Warnock and Wright
 879 2020). As with any model-based method, the inferences
 880 we make will naturally be influenced by these modeling
 881 choices. However, this sensitivity is of special concern
 882 for divergence-time estimation analyses because some
 883 parameters in the model—namely, the node ages and
 884 clock rates—are nonidentifiable (Rannala 2002; Stadler
 885 and Yang 2013): the data cannot distinguish, for example,
 886 between a branch having a fast rate and a short duration,

887 or a slow rate and a long duration. This pathology makes
 888 it difficult to use standard maximum-likelihood methods
 889 to estimate divergence times because the ML divergence-
 890 time estimates will not be unique (though see Sanderson
 891 1997, 2002; Paradis 2013, for almost-Bayesian ML solu-
 892 tions to this problem). In a Bayesian framework the prob-
 893 lem is less acute, but at the cost of posterior estimates
 894 of time and rate that are very prior sensitive, even with
 895 large datasets. Specifically, the clock and tree models are
 896 the priors on time and rate in the Bayesian total-evidence
 897 dating model, and it is therefore critical to understand
 898 their role in total-evidence dating analyses.

899 We discuss the influence of each model component—
 900 and the prospects for elaborating upon these models—in
 901 the following sections.

902 *The relative impact of model components: Which models*
 903 *matter?*—Among the three model components, the tree
 904 model had the greatest influence on divergence-time esti-
 905 mates (Fig. 2, row 2, column 3), consistent with the
 906 expected prior sensitivity of this portion of the model.
 907 In particular, chronograms inferred under the uniform

908 model strongly differed from those under the other mod-
909 els, which we attribute, counter-intuitively, to the uni-
910 form prior being very informative (as we discuss in the
911 next section). However, distributions of chronograms
912 were markedly different even among the fossilized birth-
913 death models (Fig. S17). Lineage-through-time curves
914 indicate that the influence of these models was greatest
915 in (but not restricted to) the older, fossil-rich parts of the
916 tree (Fig. 3, right), presumably because there is less in-
917 formation available from the molecular data to constrain
918 older branch lengths, particularly for extinct clades.

919 Consistent with previous simulation results (Klopf-
920 stein et al. 2019), the model of morphological evolution
921 had a negligible impact on divergence-time estimates in
922 our study (Fig. 2, row 2). This result held after pruning
923 extant taxa (Fig. S16), suggesting that this phenomenon
924 is not the result of the information in the molecular
925 data overwhelming the information in the morphologi-
926 cal data. However, the morphological transition model
927 had an obvious impact on the posterior distribution of
928 tree topologies (Fig. 2, row 1 column 1), especially for
929 fossil taxa (Fig. S16). The discrepancy between the im-
930 pact of the transition model on chronograms (negligi-
931 ble) and topology (considerable) may be due to the fact
932 that our fossil dataset is dominated by a large number
933 of closely related and similarly-aged taxa: there may be
934 subtle (and weakly supported) differences about inferred
935 relationships among these taxa that ultimately have little
936 impact on clade ages. Similarly, the two morphological
937 clock models we compared (rates linked to the molecu-
938 lar clock, or not) had a profound influence on inferred
939 *morphograms* (Fig. 2, row 3 column 2) but only a minor
940 impact on chronograms (Fig. 2, row 2, column 2). As ex-
941 pected, the linked model had the largest consequences on
942 the inferred ages of young clades (Fig. 3, middle), where
943 the influence of the molecular data should be strongest.

944 The contrasting influence of the tree model and
945 morphological-clock model on old and young clades
946 makes sense in light of what we know about divergence-
947 time estimation under relaxed-clock models. Specifically,
948 divergence-time estimates are a compromise between the
949 plausibility of the node ages (from the perspective of
950 the tree model) and the plausibility of the branch rates
951 (from the perspective of the relaxed-clock model) that
952 are needed to explain the effective branch lengths (the
953 product of rate on time of each branch). Where there is a
954 lot of information about effective branch lengths (*e.g.*, in
955 clades with extant species and therefore abundant molec-
956 ular data), the tree model and clock model must come to
957 a compromise about how to explain the effective branch
958 lengths, and the constraints of the clock model may re-
959 duce differences among the tree models. Where there
960 is less information about effective branch lengths (*e.g.*,
961 in fossil clades), then the tree model will be less con-

strained by the clock model and will have a larger role
in determining the branch lengths; in the extreme case
when there are no character data, the tree model will
completely determine the length of branches subtend-
ing fossils (as in Heath et al. 2014). However, the rel-
ative impact of these models in our analyses could be
a consequence of the topological and temporal distribu-
tion of our fossil accessions, and should not be taken
as a general pattern. We emphasize that we did not
compare different (prior) forms of clock models, as the
only difference in the morphological clock models that
we examined was whether the morphological rates were
linked to the molecular ones or not. As molecular clock
models are known to have a strong potential influence
on divergence-time estimation—both from first princi-
ples (the nonidentifiability of rate and time) and from
empirical results (*e.g.*, Ronquist et al. 2012; Rothfels and
Schuettpeitz 2014; Crisp et al. 2014; Zhang et al. 2016)—
further studies are needed to examine the impact of these
models, and their interaction with the tree models.

The uniform tree model.—A noninformative prior is one
that maximizes the ability of the data to express its pref-
erence for different parameter values, a property often
attributed to “uniform” priors. However, the informa-
tiveness of a particular prior can depend on what as-
pect of the model is being conceived; for example, a uni-
form prior over tree topologies is not uniform over clades
(Pickett and Randle 2005). Technically, a noninforma-
tive prior expresses the same amount of prior evidence
regardless of how we parameterize the model (Jaynes
1968); this is rarely (if ever) the case with uniform priors
(Zwickl and Holder 2004, and references therein).

The uniform tree model (*sensu* Ronquist et al. 2012)
is likewise informative (*i.e.*, not noninformative) about
node ages (Warnock and Wright 2020); however, our re-
sults suggest that the strength of the uniform tree prior
on node-age estimates has been underappreciated. Con-
ceptually, this prior distribution on node ages is con-
structed as follows: for each tip, draw a uniform random
variable between the age of the tip and the origin time
(stem age) of the tree, order the resulting uniform ran-
dom variables, then use the smallest (*i.e.*, youngest) uni-
form random variable as the first node age (looking back-
ward in time), the second smallest as the second node
age, and so forth. Perhaps counter-intuitively, ordering
the uniform random variables results in a prior distribu-
tion on node ages that becomes increasingly informative
as the number of tips increases. To develop an intuition
for why this is the case, we can imagine the procedure for
a set of n contemporaneous (extant) tips with stem age
of 1 arbitrary time unit, so that each (unordered) node
age is a uniform random variable between 0 and 1. As
the number of random variables increases, the variance

1015 around any given (ordered) node age shrinks (Figs. S46,
1016 S47). Formally, the i^{th} node age is the i^{th} order statistic
1017 of a uniform distribution, which is a Beta random vari-
1018 able with $\alpha = i$ and $\beta = n + 1 - i$. The variance of this
1019 Beta random variable approaches zero as $n \rightarrow \infty$, indi-
1020 cating that the prior distribution on node ages becomes
1021 increasingly narrow as the number of tips increases (Figs.
1022 S46, S47). The exact distribution is somewhat different
1023 with extinct tips because the uniform random variables
1024 are not identically distributed, but the same general logic
1025 applies.

1026 The distribution of node ages in a birth-death tree (of
1027 extant taxa) can also be represented by order statistics
1028 (Yang and Rannala 1997) and one may worry that they
1029 could exhibit similar behavior. However, in the case of
1030 birth-death models, the distribution of order statistics de-
1031 pends on the hyperparameters of the model (speciation
1032 and extinction rates), which are typically (effectively al-
1033 ways) estimated from the data. The birth-death (and the
1034 fossilized birth-death) prior should therefore be able to
1035 adjust the distribution of order statistics to suit the data,
1036 especially if rates are allowed to vary over time. Indeed,
1037 the fossilized birth-death model with fixed hyperparam-
1038 eters also appears to be very informative, and can mimic
1039 the uniform tree model when rates are arbitrarily low
1040 (Fig. S48).

1041 Consistent with its high informativeness, the uniform
1042 tree model had a profound impact on divergence times
1043 in our analyses (Figs. 2 and 3), and resulted in system-
1044 atically increased estimates of clade ages (to the point of
1045 absurdity, such as Cambrian age for the Marattiales stem
1046 and Silurian for the crown group). On a finer scale, when
1047 compared to the fossilized birth-death models, the uni-
1048 form tree model tends to spread node ages out as evenly
1049 as possible, which resulted in older ages for large, closely
1050 related clades, such as *Scolecoperis* and the extant Marat-
1051 tiaceae. These results are consistent with previous work,
1052 which reported unexpectedly ancient divergences under
1053 the uniform tree prior (Slater 2013; Wood et al. 2013; Ar-
1054 cila et al. 2015). As a consequence of clades being very
1055 old, there was also an increased opportunity for fossils
1056 to be nested within clades that would otherwise be too
1057 young to contain them, resulting in large differences in
1058 the inferred topologies (Figs. 2 and S50).

1059 *Model evaluation: Which model is best?.*—There is strong
1060 evidence that rates of morphological evolution vary
1061 among characters: transition models that exclude
1062 gamma-distributed rates fail to generate patterns of vari-
1063 ation among characters that are similar to those in the
1064 observed morphological dataset (Fig. S23). There is also
1065 evidence that the process of evolution is not the same
1066 among characters and states, as the F81 mixture mod-
1067 els generally simulate more realistic datasets than the

Mk models. Additionally, there is some evidence that 1068
the process of morphological evolution may vary among 1069
branches of the tree: removing either extinct or ex- 1070
tant lineages from the simulated and observed morpho- 1071
logical datasets results in somewhat different patterns 1072
of model adequacy, particular among morphological- 1073
transition models (Figs. S24 and S25). 1074

1075 Posterior-predictive simulations were unable to detect 1076
differences among the morphological clock models (Fig. 1077
S23). This result is somewhat surprising because, in prin- 1078
ciple, different posterior distributions of morphograms 1079
(as observed under the morphological-clock models, Fig. 1080
2) could affect model adequacy. The apparent insen- 1081
sitivity to the morphological-clock model persists even 1082
after removing extinct lineages from the morphological 1083
datasets (Fig. S24), indicating that this is not simply a 1084
consequence of the topological and temporal distribution 1085
of fossils in our tree (*i.e.*, a large clade of ancient fossils 1086
with branch lengths that do not depend on any molecu- 1087
lar data).

1088 All of the tree models estimated very similar mor- 1089
phograms (Fig. 2), and consequently, the posterior- 1090
predictive distribution for these models were effectively 1091
the same. The failure of the PPS approach to differenti- 1092
ate among the tree models, despite the extreme effect of these 1093
models on the resulting inference (Figs. 2, 3) is a manifes- 1094
tation of the nonidentifiability of this model component: 1095
differences in time (node heights) can be compensated 1096
for by differences in rates, with no effect on the expected 1097
distribution of the data. However, Bayes factors (which 1098
are sensitive to differences in prior distributions) clearly 1099
differentiated among the tree models (Table 1). The low 1100
marginal likelihood under the uniform tree model pre- 1101
sumably reflects the fact that the clock rates necessary to 1102
achieve a reasonable fit to the data given the highly dis- 1103
torted node ages are implausible from the perspective of 1104
the relaxed-clock model.

1105 The other major, and surprising, result of our Bayes 1106
factor comparisons was the relatively poor performance 1107
of the FBD models that allow fossilization rates to vary. 1108
Considering that our fossils are overwhelmingly from 1109
the Pennsylvanian, and that this concentration appears 1110
to be due (at least in part) to the unique preservation 1111
potential of that epoch (via the formation of coal balls), 1112
we expected that fossilization-rate variation would be an 1113
important model component. However, the number of 1114
fossils in a given time interval is a function of both the 1115
fossilization rate and the number of lineages in that time 1116
interval. The EFBD $_{\psi}$ model struggles to produce enough 1117
lineages in the Pennsylvanian to generate a sufficient 1118
number of fossils, even with an elevated fossilization rate 1119
(Fig. S29); presumably, increasing the diversification rate 1120
would increase the number of lineages available to fos- 1121
silize, but would also predict many more extant taxa than

1122 we observe. By contrast, the EFBD $_{\lambda,\mu}$ model implies a
1123 spike of diversity in the Pennsylvanian (Fig. 5, top), and
1124 is therefore able to explain both the large number of fos-
1125 sils in that time period and the observed diversity at the
1126 present. Allowing fossilization rates to vary on top of
1127 diversification-rate variation does not appear to improve
1128 model fit, *i.e.*, the epoch-specific diversification-rate vari-
1129 ation appears to be sufficient to describe the observed
1130 patterns of fossil and extant diversity.

1131 *Modeling prospects.*—While divergence-time estimates ap-
1132 pear to be relatively insensitive to the morphological
1133 transition model—despite the fact that we investigated
1134 arguably the most biologically realistic morphological
1135 models applied in a TED analysis to date—these mod-
1136 els still provide the opportunity to learn about the pro-
1137 cesses that govern morphological evolution. It is no-
1138 table, for example, that the best overall morphological-
1139 transition model—the F81 mixture+ Γ model—is also the
1140 most parameter-rich, and that even this model struggles
1141 to adequately model variation in the process of morpho-
1142 logical evolution among branches of the tree (Figs. S24
1143 and S25). These results may indicate that our morpho-
1144 logical dataset could support even more complex mod-
1145 els, for example, ones that allow the process of evolution
1146 to vary among branches (Beaulieu et al. 2013; Goloboff
1147 et al. 2019), or that accommodate correlated evolution
1148 among characters (Pagel and Meade 2006; Meyer et al.
1149 2019) and other complex dependencies (Maddison 1993;
1150 Tarasov 2019). However, such models generally require
1151 increasing the state-space of the characters, which com-
1152 plicates the calculations used for correcting for variable-
1153 only characters, so more work is necessary before they
1154 should be used in total-evidence dating analyses. Our
1155 models also assume that morphological data subsets (de-
1156 fined by the number of states) evolve at different rela-
1157 tive rates, and that rate variation within data subsets fol-
1158 lows a gamma distribution. Models that accommodate
1159 variation in the rate and process of evolution using bio-
1160 logically meaningful data partitions (for example, parti-
1161 tioning between feeding and non-feeding characters, as
1162 in Wright et al. 2020, or between reproductive and veg-
1163 etative characters, etc.) provide another opportunity to
1164 improve model realism.

1165 Ultimately, our choice of candidate morphological
1166 clock models was guided by practical considerations,
1167 but, as with the morphological transition models, it is
1168 easy to imagine more biologically rich and meaning-
1169 ful models. Autocorrelated relaxed clocks, which as-
1170 sume that rates of evolution themselves evolve over the
1171 branches of the tree, are a biologically natural class of
1172 models with a long history in Bayesian divergence-time
1173 estimation (Heath and Moore 2014), and can accom-
1174 modate gradual patterns of rate change (Thorne et al.

1175 1998; Thorne and Kishino 2002), rare episodic changes
1176 (Huelsenbeck et al. 2000), clade-specific rates (Drum-
1177 mond and Suchard 2010), and complex mixtures of grad-
1178 ual and episodic change (Lartillot et al. 2016). We at-
1179 tempted to use autocorrelated Brownian motion and
1180 random-local-clock models (Thorne et al. 1998; Thorne
1181 and Kishino 2002; Drummond and Suchard 2010) in the
1182 early stages of our study, but were unable to make the
1183 MCMC analyses function adequately: as the number of
1184 branches in the tree grows, the dependency between the
1185 autocorrelated rates and the tree topology makes it in-
1186 creasingly difficult to efficiently sample over tree space.
1187 As total-evidence-dating approaches gain in popularity,
1188 it will be desirable to develop more efficient MCMC pro-
1189 cedures for sampling tree and branch rates under auto-
1190 correlated models.

1191 Consistent with previous studies (Zhang et al. 2016;
1192 Wright et al. 2020), our results suggest that the tree model
1193 can have a large—or, in the case of the uniform tree
1194 model, overwhelming—impact on total-evidence dating
1195 analyses. While the fossilized birth-death model im-
1196 proves our ability to jointly model extinct and extant di-
1197 versity, and to have a biologically meaningful tree model,
1198 it is nonetheless limited in some important ways. For
1199 example, our results assume that the sampling fraction
1200 applies uniformly to all extant taxa. While the qual-
1201 itative impact of each model component was consist-
1202 ent regardless of whether we assumed the total extant
1203 sampling fraction ($\rho = 36 \div 12000$) or the ingroup frac-
1204 tion ($\rho = 27 \div 111$; Supplemental Material S§6.2), exact
1205 age estimates are different between the two assumed
1206 sampling fractions, with the outgroup sampling fraction
1207 driving older age estimates (see Supplementary Material
1208 S§6.2). Unfortunately, neither of these sampling fractions
1209 is realistic, given that the sampling intensities for our in-
1210 group and outgroup are highly imbalanced.

1211 While a model of “diversified” taxon sampling exists
1212 for the fossilized birth-death process (Zhang et al. 2016),
1213 this model assumes that all unsampled extant lineages
1214 attach to the tree more recently than the youngest in-
1215 ternal node or fossil. This assumption is clearly inap-
1216 propriate for the Marattiales, where most extant species
1217 arose relatively recently (Fig. 4). Given that empirical
1218 datasets frequently exhibit imbalanced taxon sampling
1219 schemes, and some degree of “diversified” or “deep-
1220 node” sampling (Matschiner 2019), an important avenue
1221 of development for the fossilized birth-death process will
1222 be to derive more flexible models of incomplete taxon
1223 sampling. Beyond the details of accommodating incom-
1224 plete taxon sampling, the fossilized birth-death models
1225 we used in this study make many simplifying assump-
1226 tions about the diversification and sampling process. For
1227 example, they assume that diversification and fossiliza-
1228 tion rates are the same among lineages and within each

epoch, and do not depend on ecology, morphology, geography, etc. Tree models that allow for state-dependent and lineage-specific rates exist (in principle) for phylogenetic (epidemiological) models (Kühnert et al. 2016), but while the theoretical machinery underlying the phylogenetic and fossilized birth-death models is nearly identical, to our knowledge these models have not been adapted for the fossilized birth-death model. They therefore represent a major untapped resource for future development and application.

Finally, our use of posterior-predictive simulation to assess the adequacy of morphological models assumes that the statistics we have chosen—the total parsimony score, S , and the variance in parsimony scores, V —are sensitive to realistic violations of our models. More work is needed to determine the power and utility of these model evaluation tools to assess different components of the total-evidence model.

Marattiales phylogeny and divergence times

Extant relationships.—Topologically, our results for extant relationships are generally consistent with other molecular studies of Marattiales phylogeny, including in supporting *Danaea* as sister to the rest of the extant marattialeans (Murdock 2008b; Lehtonen et al. 2020), and supporting the monophyly of each genus (Murdock 2008b; Rothwell et al. 2018b; Lehtonen et al. 2020). Among the extant taxa, the main point of uncertainty remains the position of *Christensenia*. Whereas Murdock (2008b) found *Christensenia* sister to the remainder of the *Christensenia* + *Marattia* s.str. + *Angiopteris* clade, in our results *Marattia* is in that position. However, support is relatively weak for these relationships in our results as well as in Murdock (2008b). Lehtonen et al. (2020), while likewise having only weak support, resolve the third possible relationship: *Christensenia* sister to *Marattia*, and that clade sister to *Angiopteris*. This phylogenetic uncertainty is mirrored in previous morphological cladistic studies (see Hill and Camus 1986), as well as the sampling-focused analyses of Rothwell et al. (2018b), which have supported each of these positions for *Christensenia*, in addition to placing it sister to *Danaea* or to all other extant genera. Morphologically, our topological resolution of *Christensenia* sister to *Angiopteris* is supported by spherical spores (Fig. S64), synangia oval in longitudinal section (Fig. S67), and a raised stomatal complex (Fig. S75). Regardless of its precise position, *Christensenia* is clearly nested within extant Marattiaceae and the characters that it shares with Psaroniaceae species (most notably, its radially symmetrical synangia; Fig. S1K) are independently derived, a result that contrasts strongly with pre-phylogenetic and early morphological cladistic hypotheses (e.g., Campbell 1911; Hill and Camus 1986).

The extant Marattiales are deeply isolated from their

closest extant relatives, the leptosporangiate ferns, as has been repeatedly demonstrated (Pryer et al. 2001; Qiu et al. 2007; Murdock 2008b; Rai and Graham 2010; Lehtonen 2011; Rothfels et al. 2015b; Lehtonen et al. 2020). Given this situation—a very long stem branch connecting to a series of much shorter branches within the Marattiales crown group—one might expect uncertainty within the crown group relationships driven by uncertainty in the attachment position of the stem branch (Huelsenbeck et al. 2002; Rothfels et al. 2012), as in the maximum likelihood results of Murdock (2008b) and as has been seen in other similarly isolated groups (e.g., *Isoetes*, *Equisetum*, and *Cycas*; Des Marais et al. 2003; Schuettpelz and Hoot 2006; Nagalingum et al. 2011). With this concern in mind, we questioned whether *Danaea* was correctly inferred as sister to the remaining extant taxa. Our analyses allow for the long stem branch to be broken up by fossils, which may additionally provide important information about the polarity of morphological characters (i.e., which character states are ancestral for crown Marattiaceae), potentially allowing for more reliable inference of the “root” position for this crown clade (Doyle and Donoghue 1987; Gauthier et al. 1988; Donoghue et al. 1989; Huelsenbeck 1991; Smith 1998; Wills and Fortey 2000; Mongiardino Koch and Parry 2020). Nonetheless, our results further support the growing consensus (Pryer et al. 2001; Schuettpelz et al. 2006; Qiu et al. 2007; Murdock 2008b; Rai and Graham 2010; Lehtonen et al. 2020) that *Danaea* is sister to the remaining extant lineages; it shares synapomorphic states of the extant clade (e.g., the presence of stipules), and has apomorphic states (e.g., once-pinnate, dimorphic leaves), but few character states argue for it being nested among the other extant taxa; those that do appear to be homoplastic.

Overall relationships.—Our results depart in significant ways from classic interpretations of Marattiales evolution (see Rothwell et al. 2018a), the topologies inferred by previous total-evidence analyses (Rothwell and Good 2000; Lehtonen et al. 2020), and the morphological cladistic analysis by Liu et al. (2000). Namely, in our MCC trees Psaroniaceae appears paraphyletic, comprising a grade of early-diverging lineages in Marattiales, a component of which eventually gave rise to the Marattiaceae (Figs. 4, S26). To a lesser extent, Lehtonen et al. (2020) found a similar pattern, with a polytomy comprised of *Araiangium*, *Danaeites rigida* + *Millaya tularosana*, and *Sydneia* + *Radstockia* sister to the remaining Psaroniaceae and Marattiaceae clades, and Liu et al. (2000) also recovered a non-monophyletic Psaroniaceae, even though they used a much different fossil sample.

In contrast to Rothwell et al. (2018b), we find little evidence for the alternative hypothesis that Psaroniaceae and Marattiaceae are reciprocally monophyletic clades

1335 that diverged from a common ancestor and thereafter
1336 had separate evolutionary histories (Figs. S54, S27).
1337 Nonetheless, we do resolve a large Psaroniaceae clade
1338 that includes most (Fig. 4) or all (Fig. S26) of the
1339 *Scolecoperis* species along with, at least, *Araiangium pyg-*
1340 *maeum*, *Acaulangium bulbaceum*, *Convexocarpus distichus*,
1341 and *Buritranopteris costata* + *Gemellitheca saudica*. [Lehtonen et al. \(2020\)](#) and [Rothwell et al. \(2018b\)](#) found sev-
1342 eral other genera to be included in the large Psaroniaceae
1343 clade—such as *Zhutheca*, *Taiyuanitheca*, *Acitheca*, *Pecti-*
1344 *nanangium*, *Acrogoetheca*, *Symopteris*, and *Sydneia*—but we
1345 did not include these taxa in our analyses. Notably, *Rad-*
1346 *stockia* is inferred by both [Rothwell and Good \(2000\)](#) and
1347 [Lehtonen et al. \(2020\)](#) (the latter with *Sydneia*), to be sister
1348 to the rest of Marattiales (Psaroniaceae + Marattiaceae),
1349 whereas our analyses place *Radstockia* well within the
1350 Marattiales, consistent with [Hill and Camus \(1986\)](#).
1351

1352 Overall, the grade of lineages recovered in our MCC
1353 trees (which, other than *Scolecoperis* species, was largely
1354 consistent among our different empirical consideration
1355 analyses) captures the morphological transitions accom-
1356 panying the floristic turnover in the late Pennsylvanian
1357 and Permian that resulted in the modern Marattiaceae.
1358 These critical transitional forms, for example, *Eoangiopteris*
1359 *goodii*, *Millaya tularosana*, and *Radstockia kid-*
1360 *stonii*, could be assignable to a broad interpretation of
1361 Marattiaceae, owing to their morphological departure
1362 from most Psaroniaceae, and specifically in their ses-
1363 sile bilateral synangia that contain numerous sporangia
1364 borne on flattened rather than downturned pinnules, and
1365 by having spores with an ornamented exine (Fig. S71). In
1366 other respects, however, these plants retain traits charac-
1367 teristic of Psaroniaceae, such as highly dissected leaves
1368 (Fig. S63). The picture that emerges is of a remnant
1369 of the formerly diverse Psaroniaceae that, through a se-
1370 ries of morphological changes (some of which are cap-
1371 tured in the fossil record) evolves into the ecologically
1372 and morphologically distinct modern Marattiaceae ([Liu et al. 2000](#)).
1373

1374 The position of *Escapia*—another potentially “transi-
1375 tional” taxon—is highly uncertain in our analyses, and
1376 its alternative phylogenetic resolutions have notable im-
1377 plications for the interpretation of the temporal history
1378 of the Marattiales, particularly the persistence of Psar-
1379 oniaceae. *Escapia* is a fragmentary fossil ([Rothwell et al. 2018a](#)) that exhibits a unique suite of character states, including autapomorphies such as synangia served by transfusion tracheids (Fig. S68), and states that are otherwise found in the *Scolecoperis* clade, namely having both radial and bilateral synangia (Fig. S66), with ovate eusporangium cavities (Fig. S72), and extended sporangium tips (Fig. S69). In long-section, the synangia shape appear as two crescents (Fig. S67), a state that otherwise only occurs within our dataset in *Sco-*

1380 *copteris alta*. In consequence, in some of our analyses *Es-*
1381 *capia* is resolved sister to *Scolecoperis alta* (Fig. S51), or
1382 nested among *Scolecoperis* species (Fig. S43), support-
1383 ing the hypothesis that multiple lineages of Pennsylvanian
1384 Psaroniaceae persisted into the early Cretaceous and
1385 that psaroniaceae species may have extensively coexisted
1386 with members of the Marattiaceae ([Rothwell et al. 2018a,b](#)). However, in our favored model *Escapia* is resolved among stem groups more closely related to the extant clade (Fig. 4; see also Figs. S26, S55). In this case, *Escapia* would not be interpreted as the last vestige of the Psaroniaceae extending into the Cretaceous, but instead as another unusual transitional form, which converged upon synangial morphology similar to *Scolecoperis alta*.
1401

1402 The Permian *Qasimia schyfsmae* has been generally re-
1403 garded as the oldest representative of the Marattiaceae
1404 (e.g., [Hill et al. 1985](#); [Hill and Camus 1986](#); [Rothwell et al. 2018a](#)) based on synangial characters and foliage similarities with *Marattiopsis* species. Similar to other analyses ([Rothwell and Good 2000](#); [Lehtonen et al. 2020](#)), we found *Qasimia schyfsmae* to be most closely related to the clade of *Marattiopsis* + extant Marattiaceae (Fig. 4), and in many cases we reconstruct it as a direct ancestor of the extant species (as was suggested by [Hill and Camus 1986](#)).
1413

1414 Our other major phylogenetic result, and one of our
1415 most surprising inferences, is that the extant marattiaceans form a clade (with 0.96 posterior support) phylogenetically apart from any of the fossils (Fig. 4), despite the fact that a number of these fossils closely resemble extant taxa and have in the past even been considered congeneric with extant species (see discussions in [Bomfleur et al. 2013](#); [Escapa et al. 2015](#)). This result only holds when temporal data are incorporated; in the non-dated analyses (Fig. S77) some of these fossils are resolved among the extant species, as they were in, e.g., [Rothwell et al. \(2018b\)](#) and [Lehtonen et al. \(2020\)](#).
1425

1426 The position of these fossil taxa outside the crown
1427 group is not completely unexpected: the extant genera
1428 mostly lack unique apomorphies and are instead defined
1429 by suites of states (see [Murdock 2008a](#); [Escapa et al. 2015](#)).
1430 *Marattiopsis* species, in turn, are mostly based on frag-
1431 mentary fossils, and generally exhibit combinations of
1432 states that do not match any of the extant genera. There
1433 is, therefore, enough homoplasy that the fossils could be
1434 placed in a number of positions among extant taxa or on
1435 the stem, as exemplified by their alternative placements
1436 in other analyses ([Rothwell and Good 2000](#); [Lehtonen et al. 2020](#); [Liu et al. 2000](#)).
1437

1438 *Timescale of Marattiales evolution.*—Our mean Marattiales
1439 stem-age estimate (414 Ma, 95% CI = [351, 505]; Fig. 4,
1440 Table S.3) is older than, but broadly consistent with, ear-
1441 lier studies that have inferred ages for this node using

node-dating approaches (e.g., ~ 325 Ma (Rothfels et al. 2015b), ~ 366 Ma (Testo and Sundue 2016), and ~ 360 Ma (Qi et al. 2018); Shen et al. (2018) infer a much younger date, ~250 Ma, but they also infer a different topology). Similarly, our inferred stem age for the Marattiaceae is consistent with other analyses, both when defined as the clade including extant taxa + *Marattiopsis* spp. + *Qasimia*, or more narrowly as the extant taxa + *Marattiopsis* spp. (Lehtonen et al. 2020; Rothwell et al. 2018b). Previous node-based estimates of the crown group age, however, have varied tremendously. On the younger end, Qi et al. (2018) estimated an age of ~ 75 Ma, and Testo and Sundue (2016) place this node at ~ 160 Ma. Even the 160 Ma estimate, however, is dramatically younger than the 185 – 224 Ma or ~ 200 Ma inferences from Lehtonen et al. (2017) and Smith et al. (2010), respectively. The latter two inferences differed from the studies that inferred a younger crown age in assuming that a fossil marattialean was congeneric with an extant taxon, and thus they constrained the Marattiales crown node to be older than the fossil. Based on morphological similarities, this assumption is well-justified: parsimony analyses of morphology (e.g., Rothwell et al. 2018b; Lehtonen et al. 2020) consistently resolve fossils among the extant taxa, as do our non-clock trees (Fig. S77); the (effectively non-clock) parsimony-based dating analyses of Lehtonen et al. 2020 also infer a very old date, of ~ 220 Ma).

The influence of temporal data on estimates of topology and divergence times

Discrepancies between the clock and non-clock analyses (see Fig. S77), and between node-based and TED analyses, provide a strong example of the potential for temporal data to alter our inferences of phylogenetic relationships (Drummond et al. 2006; Gavryushkina et al. 2017; Lee and Yates 2018; Wright et al. 2020). TED analyses, by co-estimating the position of the fossils and the divergence times of the tree, allow for both the morphological characteristics and the temporal data associated with the fossils (their ages) to influence their position in the phylogeny. Effectively, if the full model—incorporating morphological and temporal information from all the samples, as well the influences of the tree and clock priors—prefers a clade age that is younger, such that it overwhelms the specific morphological data that might resolve the fossil inside the clade, the model can place the fossil elsewhere. This outcome might be more likely if there is considerable homoplasy in the morphological data (as there is in extant Marattiales, and in fossil *Marattiopsis*; see Escapa et al. 2015), resulting in a high inferred rate of morphological evolution and a reasonable chance of repeated evolution of particular character states. We infer strong support for a monophyletic crown group comprising all extant taxa to the exclusion of any

fossils (see Marattiales phylogeny and divergence times, above) and a relatively young age for the crown Marattiales (mean = 84 Ma, CI = [46, 169]; Fig. 4, Table S.4), in marked contrast to studies that assumed that fossils fell among the extant species. The Marattiales, then, provide a powerful illustration of dangers of the *a priori* assignment of fossils to nodes that is required for node-based divergence-time dating: phylogenetic positions of fossils that seem compelling based on morphology may not be supported in the context of the morphological and temporal data of the full sample (for a similar result, see Lee and Yates 2018).

Reconciling model inferences with marattialean biology

Inferred diversification dynamics.—While not the primary focus of our analyses, our application of the FBD model allowed us to infer the fine-scale (epoch-level) diversification dynamics of this clade, informed by the fossil record. Throughout its history, we estimate a diversification rate for the Marattiales near zero, with both speciation and extinction rates being low. There is some notable variation around this trend, however. Specifically, speciation rates increase in the Carboniferous (and particularly in the Mississippian), the middle Permian (Guadalupian), and to a lesser extent, the late Triassic, whereas extinction rates spike in the early Permian and lower and middle Triassic (the latter potentially reflecting the end-Permian mass extinction; Fig. 5). In combination, these rates result in a picture of Marattiales evolution where species-level diversity increases rapidly in the Pennsylvanian, culminating in a peak of ~ 2800 species (median estimate) before crashing abruptly in the early Permian (Fig. 5). However, allowing fossilization rates to vary reduces the inferred peak diversity: the EFB $D_{\lambda,\mu,\psi}$ model, while disfavored over our optimal tree model by Bayes factors [Table 1], infers a peak of “only” ~ 1200 species, which may be more biologically plausible. The other major pattern in our diversification inferences—the sudden spike in speciation rates in the Pliocene—coincides with the modern taxa and likely relates to a switch in species concepts rather than underlying biology.

Instead of reflecting the reality of the fossil record, the apparent lack of a signal of fossilization-rate variation may be due to a combination of confounded epoch-specific diversification and fossilization rates, taxonomic practice, our sampling regime, or simply a lack of power. Given that most preserved Pennsylvanian marattialeans were wetland species (DiMichele and Phillips 2002), the early Permian aridification of the tropics would have reduced both their fossilization potential and their diversity (Montañez et al. 2007); by modeling the latter, we may have been able to simultaneously account for the former. In addition, fossilization-rate variation is likely

1548 muted by the tendency for taxonomists to pay greater
1549 attention to taxa that are unexpected or otherwise inter-
1550 esting. In the case of the Marattiales, this bias results in
1551 small fragmentary Cretaceous fossils being described in
1552 great detail owing to their rarity (*i.e.*, *Escapia*; Rothwell
1553 et al. 2018a) and therefore included in our dataset, while
1554 the great bulk of the fossils—fine anatomical preserva-
1555 tions in Pennsylvanian coal balls—are relatively under-
1556 represented. Our sampling regime likely exacerbates this
1557 bias: of the many Carboniferous and Permian Marattiales
1558 fossils, we included relatively few (*e.g.*, only 12 of the
1559 over 30 described genera; Lundgren et al. 2019; Rothwell
1560 et al. 2018a). A final potential explanation is that we sim-
1561 ply do not have enough information to distinguish be-
1562 tween models with and without fossilization-rate varia-
1563 tion, and that with more fossil data we would have re-
1564 jected the constant-fossilization-rate model. Regardless,
1565 our primary results are nearly the same when we allow
1566 fossilization rates to vary (Figs. 2, 3).

1567 *A paleoecological perspective.*—While the Devonian origin
1568 time inferred here for the Marattiales significantly pre-
1569 dates any known fossils, we find that the major initial
1570 diversification of the Marattiales likely began in the Mis-
1571 sissippian (359 – 323 Ma). Although there is not a lot
1572 of unequivocal evidence of Mississippian marattialeans
1573 in the macrofossil record, such a pattern is expected for
1574 the initial diversification of a group; the existence of only
1575 a few lineages, likely with small geographical ranges,
1576 would lead to exceedingly low probability of fossil recov-
1577 ery, which is compounded by the difficulty in recognizing,
1578 or preserving, the defining characteristics of a group
1579 early in its evolution (Marshall 2019).

1580 The first potential macrofossil evidence of marat-
1581 tialeans, from the earliest stage of the Mississippian
1582 (Tournaisian, 359 – 347 Ma), is *Burnitheca pusilla*, an iso-
1583 lated permineralized synangium (Meyer-Berthaud and
1584 Galtier 1986). Compression fossils of trunks with dis-
1585 tichous or helically arranged leaf scars have also been
1586 described from Mississippian localities (Crookall 1959);
1587 these fossils have a growth habit similar to early Marat-
1588 tiales taxa, but, like *Burnitheca pusilla*, lack the details
1589 needed to confidently assign them to the order. Spec-
1590 imens of the genus *Megaphyton*, stem compression and
1591 impressions from latest Mississippian-age sediments, are
1592 generally regarded as the oldest evidence for Marattiales
1593 (Pfefferkorn et al. 1976).

1594 The relatively small size of the early specimens, and
1595 the absence of more delicate leaf fragments, indicate that
1596 they likely represent allochthonous plant material, mean-
1597 ing they were transported from their habitat before end-
1598 ing up in the depositional environment in which they
1599 were preserved (Greenwood 1991). This pattern im-
1600 plies that marattialeans initially grew in habitats with a

low preservation potential, which could reconcile their
scarcity in the fossil record with the high diversification
rates we infer during the Mississippian (Fig. 5). Dur-
ing the Mississippian-to-Pennsylvanian transition, the
climate in the Euramerican tropics during the glacial in-
tervals changed from seasonally dry to tropical everwet
(Calder and Gibling 1994); both the wetter climate and
the associated expansion of peat swamp habitats sub-
stantially increased chances of structural preservation of
the swamp vegetation (Cecil 2003; Gastaldo and Demko
2011). The earliest commonly accepted, unequivocal
marattialeans are stems known from these conditions of
high fossilization potential.

All finds suggest that the early Psaroniaceae were
trees of relatively small stature, with monocyclic steles,
distichous leaf arrangement, and a small root mantle
(DiMichele and Phillips 1977; Millay 1997). It was not
until the middle Pennsylvanian that Psaroniaceae species
became important understory elements and common
canopy trees (with thick root mantles and large decom-
pound leaves) in lowland clastic swamp communities
and more widely distributed in Euramerica (DiMichele
and Phillips 2002; Millay 1979, 1997). After an initial de-
cline during the late Middle Pennsylvanian lycopod col-
lapse, Psaroniaceae further increased in size and ecolog-
ical importance, became canopy dominants in the clas-
tic swamps, and firmly established themselves in the
drier part of the peat swamps (Cleal 2015; DiMichele
and Phillips 1996, 2002; Phillips et al. 1985). These, and
the earlier taxa, had highly dissected leaves and synan-
gia with small numbers of sporangia, traits that we re-
construct as persisting until the early Permian (Figs. S62,
S65). The well-known members of the Psaroniaceae, in-
cluding most in our analysis, are plants from these peat
swamps, depositional environments where chances of
preservation were high. Given that we have a good idea
how plant communities changed over time in the wet-
ter parts of the landscape, less information from the local
drier habitats, and very little information on the evolu-
tion of plants and their communities in the extrabasinal
environments (Looy et al. 2014), the drop in diversifica-
tion in the early Permian (Cisuralian; Fig. 5) likely repre-
sents the extinction of the swamp taxa, associated with
the aridification of the Euramerican tropics (Montañez
et al. 2007).

Marattialeans produce massive amounts of spores and
more than a dozen dispersed spore taxa have been found
in situ in marattialean sporangia (for an overview see
Balme 1995). The taxonomic resolution of these spores is
quite variable; some are known from multiple distantly
related taxa, while others have only been recorded from
members of the Psaroniaceae or Marattiaceae (see *e.g.*,
Millay and Taylor 1984; Lesnikowska 1989; Lesnikowska
and Willard 1997). The more taxonomically restricted

1655 taxa, including the genera *Fabasporites*, *Spinospores*, *Thy-*
1656 *mospora*, *Torisporea*, and smaller forms of *Cyclogranis-*
1657 *porites* and *Laevigatosporites*, can be used as evidence for
1658 Marattiales in the absence of distinctive larger fossils
1659 (Lesnikowska 1989; Looy and Hotton 2014). Combined
1660 with the initial rarity of the marattialeans in the fossil
1661 record, the spore record corroborates our inferred diver-
1662 sification patterns (Fig. 5). Small amounts of minute *Cy-*
1663 *clogranisporites* and *Punctatisporites* (Lesnikowska 1989)
1664 spores have been described from the Tournasian on, and
1665 are followed by a stepwise increase in species diversity
1666 and abundance when marattialeans become dominant el-
1667 ements of the swamp communities.

1668 CONCLUSION

1669 While our focus was on the Marattiales, our results per-
1670 tain to the general power and utility of the total-evidence
1671 dating framework, in conjunction with the fossilized
1672 birth-death class of tree models. The arguments in sup-
1673 port of TED approaches and FBD models are not trivial:
1674 as our results show, different modeling choices can cause
1675 dramatic differences in the resulting phylogenetic infer-
1676 ences, owing in large part to the nonidentifiability inher-
1677 ent to these models. Fortunately, statistical models are
1678 accompanied by a robust toolkit that allows us to assess
1679 the influence of model and prior specification on param-
1680 eter estimates, to compare the fit of competing models,
1681 to assess their ability to describe the true data-generating
1682 process, and to identify ways in which the models can
1683 be made more realistic. These features are most avail-
1684 able in biologically interpretable models, like the FBD
1685 model, because the parameter estimates can be compared
1686 against empirical expectations. With the support of this
1687 toolkit, these modeling choices transform from an ana-
1688 lytical nuisance into an opportunity to learn about the
1689 processes that produced our data, and subsequently to
1690 identify avenues for increasing the biological realism of
1691 our models.

1692 By applying this toolkit to the Marattiales, we were
1693 able to infer a nuanced picture of the evolution and di-
1694 versification of this clade over its ~ 400-million-year his-
1695 tory. This inference was possible despite the fact that
1696 much of the abundant Marattiales fossil record was left
1697 by lineages without extant descendants, the extant taxa
1698 are a young clade very distant from their closest liv-
1699 ing relatives, and the placement of fossils is compro-
1700 mised by their often fragmentary nature and morpholog-
1701 ical homoplasy among fossil and extant species. We infer
1702 that the Marattiales began to diversify in the Mississip-
1703 pian, prior to a well-established fossil record. Considera-
1704 tions of the ecology of potential early marattialeans sug-
1705 gest that this timescale may be reasonable: early marat-
1706 tialeans appear to be small, uncommon taxa, which oc-

curred in habitats with low preservational potential. We
also infer that the Marattiales experienced peak diver-
sity at the end of the Pennsylvanian, before a sharp de-
cline in the Permian to relatively stable levels of standing
diversity that persisted throughout the Mesozoic to the
present. Again, this pattern makes sense in light of the
ecology of marattialeans and our understanding of paleo-
climate: the relatively wet climate in Euramerica during
the Pennsylvanian would have been ideal for the prolif-
eration of wetland-adapted marattialeans, while subse-
quent aridification of that region during the early Per-
mian would have driven high rates of extinction. The
broad concordance of phylogenetic, ecological, and paleo-
climatological evidence demonstrates the potential for
total-evidence dating—particularly in conjunction with
the fossilized-birth-death model—not only to harmonize
“rocks and clocks”, but also to elucidate macroevolution-
ary processes.

SUPPLEMENTARY MATERIAL

Supplementary scripts and data can be found in the
Data Dryad repository DOI:10.5061/dryad.10.5061 and the GitHub repos-
itory [https://github.com/mikeryanmay/marattiales_](https://github.com/mikeryanmay/marattiales_supplemental/releases/tag/1.0)
[supplemental/releases/tag/1.0](https://github.com/mikeryanmay/marattiales_supplemental/releases/tag/1.0).

FUNDING

This research was supported by the National Science
Foundation (NSF) grant DEB-1754705 to CJR and CVL,
DEB-1754723 to NSN, and DEB-1754385 to MAS.

ACKNOWLEDGEMENTS

We thank Andy Murdock, members of the Rothfels
lab, Jack Tseng, Ixchel González Ramírez, Ziad Khouri,
Xavier Meyer, and Brian Moore for their thoughtful dis-
cussion, Bill DiMichele, Scott Elrick, and Ignacio Escapa
for images of fossil specimens, and associate editor Ryan
Folk and three anonymous reviewers for providing sug-
gestions that greatly improved the manuscript.

This research used the Savio computational cluster re-
source provided by the Berkeley Research Computing
program at the University of California, Berkeley (sup-
ported by the UC Berkeley Chancellor, Vice Chancellor
for Research, and Chief Information Officer).

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