

1 Short title for running head: DeLong-Duhon and Bagley: Species delimitation in *Stereum*

2 **Phylogeny, morphology, and ecology resurrect previously synonymized species of North**  
3 **American *Stereum***

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10 **ABSTRACT**

11 *Stereum* is a globally widespread genus of basidiomycete fungi with conspicuous shelf-like  
12 fruiting bodies. Several species have been extensively studied as pathogens and for medically  
13 relevant secondary metabolites, but prior to this publication the molecular phylogeny of  
14 *Stereum* had not been analyzed, despite pervasive morphological cryptic speciation in the genus. Here,  
15 we provide a preliminary investigation into species boundaries among some North American  
16 *Stereum*. The nominal species *Stereum ostrea* has been referenced in field guides, textbooks,  
17 and scientific papers as a common fungus with a wide geographic range and even wider  
18 morphological variability. We use nuc rDNA ITS1-5.8S-ITS2 (ITS barcode) sequence data of  
19 specimens from midwestern and eastern North America, alongside morphological and  
20 ecological characters, to show that *Stereum ostrea* is a complex of at least three  
21 reproductively isolated species. Preliminary morphological analyses show that these three  
22 species correspond to three historical taxa that were previously synonymized with or  
23 confused for *S. ostrea*: *Stereum fasciatum*, *Stereum lobatum*, and *Stereum subtomentosum*.  
24 *Stereum hirsutum* ITS sequences taken from GenBank suggest that other *Stereum* species

25 may also be species complexes. Future work should apply a multilocus approach and global  
26 sampling strategy to better resolve the taxonomy and evolutionary history of this important  
27 fungal genus.

28 **KEY WORDS:** Stereaceae; ITS; systematics; taxonomy; biodiversity

## 29 INTRODUCTION

30 The most recent estimate of fungal diversity predicts that there are 2.2 to 3.8 million species  
31 of fungi, but only ~120 000 have been formally described, and fewer than 23 000 have ITS  
32 sequences available on NCBI GenBank (Hawksworth and Lücking 2017; Vu et al. 2014).  
33 New species of fungi are often discovered as cryptic species lumped under single, well-  
34 established names, such as with *Amanita muscaria* and *Cantharellus cibarius* (Geml et al.  
35 2006; Buyck and Hofstetter 2011). While the members of some species complexes may  
36 initially be difficult to separate morphologically, distinguishing features can become clear  
37 after phylogenetic analysis and trait-mapping, as was shown with *Fomes fomentarius* and its  
38 neglected sister species *F. inzengae* (Peintner et al. 2019). Critically, differences among  
39 cryptic or apparently cryptic species can be economically relevant, such as with the fungal  
40 plant pathogen *Magnaporthe grisea* and the morphologically indistinguishable *M. oryzae*,  
41 which infect crabgrass and rice, respectively (Couch and Kohn, 2002). Fungi are not only  
42 incredibly speciose, but are also morphologically and ecologically diverse, thus presenting  
43 fantastic systems to explore how divergent selection along a variety of ecological axes might  
44 influence the origin of new biodiversity. However, in the absence of robust taxonomic  
45 resources, it is difficult to even begin studying which ecological factors might drive  
46 divergence in fungi.

47 One understudied fungal group that may be a good candidate for future  
48 ecological speciation research is *Stereum*, a diverse genus of shelf-like wood-decay

49 fungi common in wooded biomes throughout the world. Though *Stereum* has been the focus  
50 of extensive bioprospecting (Doljak et al. 2006; Hybelhauerová et al. 2008; Tian et al. 2020),  
51 and some economically important species are relatively well-researched (Stenlid and  
52 Vasiliauskas 1998; Čermák et al. 2004), the below-genus level taxonomy of *Stereum* has not  
53 been subjected to phylogenetic analysis. This disconnect between the slow progress toward  
54 molecular taxonomy in *Stereum* and interest in the potentially useful properties of *Stereum*  
55 species can be problematic, especially where species descriptions include uninformative or  
56 deceptive morphological characters that may lead to misidentification of study specimens.

57 *Stereum ostrea*, like many other *Stereum* treated carelessly by systematists, is an  
58 exemplar of a species with a fraught taxonomic history (Lloyd 1913). While the name *S.*  
59 *ostrea* has been applied to collections around the world, it is unlikely that these varied  
60 collections are from a single phylogenetic species given pre-Anthropocene obstacles to  
61 dispersal. First used to describe a collection from the island of Java in Indonesia, the name *S.*  
62 *ostrea* has been consistently applied to specimens from North America since a publication by  
63 Lentz (1955) placed it in synonymy with two other putative species, *S. fasciatum* and *S.*  
64 *lobatum*, with which Welden (1971) and Chamuris (1985, 1988) concurred. Even before this,  
65 *S. ostrea* was considered either a synonym of *S. fasciatum* by some authors (Burt 1920; Pilát  
66 1930; Banerjee 1935; Hendrickx 1948) or of *S. lobatum* by other authors (Masse 1890;  
67 Cooke 1892; Hohnel and Litschauer 1907; Reinking 1920; Boedijn 1940). Critically,  
68 Demoulin (1985) argued in favor of *S. fasciatum* and *S. lobatum* as distinct species, outlining  
69 their differences, but the literature remained divided (Eicker and Louw 1998). Currently, *S.*  
70 *fasciatum* is considered a synonym of *S. ostrea* (according to Index Fungorum in October  
71 2020, <http://indexfungorum.org/>), and despite being an accepted name *S. lobatum* is rarely  
72 used. Another similar species, *S. subtomentosum*, has both been confused with *S. fasciatum*

73 and proposed to be in a complex with *S. hirsutum* (Pouzar 1964; Welden 1971;  
74 Chamuris 1988; Ginns and Lefebvre 1993).

75 We hypothesize that *S. ostrea* in North America is not a single species but a  
76 complex of several species. To test this, and to begin developing a more integrated  
77 approach to *Stereum* taxonomy, we collected over 50 *Stereum* specimens from eastern  
78 North America and used nuc rDNA ITS1-5.8S-ITS2 (ITS barcode) sequence data to  
79 create a phylogenetic tree. Because sequence information from a single gene is not  
80 always enough to reliably delineate species, we coupled our phylogenetic approach  
81 with morphological and ecological data. Following our hypothesis that *S. ostrea* is a  
82 species complex, we predicted that differences in ITS sequences would correlate with  
83 differences in morphology and ecology (e.g., substrate), and as such would reveal  
84 evidence of more than one reproductively isolated *Stereum* species.

## 85 **MATERIALS AND METHODS**

86 ***Collection and Identification.*** – We collected *Stereum* basidiocarps from locations in  
87 midwestern and eastern North America, with the most intensive sampling in Iowa and some  
88 supplemental collections made in Alabama and Florida (SUPPLEMENTARY TABLE 1).  
89 For identification we used the dichotomous key and morphological descriptions from  
90 Chamuris (1988), the most comprehensive and recent publication on *Stereum*. Based on  
91 morphology, we identified our collections as *S. ostrea*, *S. hirsutum*, *S. complicatum*, *S.*  
92 *gausapatum*, *S. sanguinolentum*, and *S. striatum*. We used iNaturalist to record photographs,  
93 dates of collection, and approximate GPS coordinates of collection location. We also  
94 recorded whether the substrate from which samples were collected was hardwood or conifer,  
95 and if the hymenium changed color when bruised. We air dried all collections and preserved

96 samples in polyethylene plastic bags. In total, we collected and analyzed 49 samples, 36 of  
97 which are included in this paper (SUPPLEMENTARY TABLE 1).

98 All *Stereum* species in this study are found primarily on angiospermous wood,  
99 except for *S. sanguinolentum* which is primarily found on conifers. *Stereum* are occasionally  
100 reported on rare hosts and have been sequenced from atypical host species (Ginns and  
101 Lefebvre 1993; Jusino 2014). SD50 *S. complicatum* was found growing on a dead branch of  
102 loblolly pine still attached to the tree.

103 **DNA Extraction, PCR and sequencing.** – We used a CTAB and liquid nitrogen method  
104 adapted from Chen et al. (2010) to extract DNA from 3 x 3 mm pieces of basidiocarp from  
105 each collection, taking care to exclude as much dirt and debris as possible. We used DNA  
106 diluted 1:20 with molecular grade water for PCR amplification using ITS1-F  
107 (CTTGGTCATTTAGAGGAAGTAA) and ITS4 (TCCTCCGCTTATTGATATGC) primers  
108 (Gardes and Bruns 1993; White et al. 1990), with the following thermocycler program: 3 min  
109 @ 94 C, 20-30 cycles (30 s @ 94 C, 30 s @ 55 C, 30 s @ 72 C), 2 min @ 72 C. We cleaned  
110 PCR products with Exo-SAP following manufacturer protocols. We sequenced PCR products  
111 in forward and reverse directions on an Applied Biosystems ABI 3730 DNA Analyzer  
112 (Thermo Fisher Scientific, Massachusetts) housed in the Roy J. Carver Center for Genomics  
113 in the University of Iowa Biology Department.

114 **Phylogenetic Trees.** – We used Geneious 8.1.7 (<http://www.geneious.com/>) for alignment of  
115 forward and reverse sequences for each collection, which we then manually checked and  
116 trimmed. We chose *Xylobolus subpileatus*, another member of the Stereaceae, as an  
117 outgroup. This specimen was collected in Florida by S.G.D. and sequenced by the Smith Lab  
118 at the University of Florida. We also obtained North American and Eurasian *S. hirsutum*  
119 sequences from BOLD and GenBank. We used MAFFT (Katoh et al. 2002) via the CIPRES  
120 server (Miller et al. 2010) to align chosen sequences, then generated a maximum likelihood

121 tree using RAxML (Stamatakis 2014) via CIPRES. For computation of the Bayesian tree, we  
122 used MrBayes 3.2.7 (Ronquist et al. 2012) with a GTR+I+ $\Gamma$  substitution model for 200 000  
123 generations, with sampling every 100 generations.

## 124 **RESULTS**

125 Maximum likelihood (FIG. 1) and Bayesian trees (SUPPLEMENTARY FIG. 1) were largely  
126 in agreement, except for the placement of the single *Stereum striatum* sequence. Both trees  
127 show *Stereum* sorting out into several clades. Specimens initially identified as *S. ostrea* were  
128 monophyletic but split into three well-differentiated clades (FIG. 1). Sequences within the  
129 same clade differed from one another by 1–3%, while sequences among the three clades  
130 differed from one another by 7–14%.

131         The “*S. ostrea*” specimens in the first clade (FIG. 1) all share features of the  
132 previously synonymized species *S. fasciatum*, as described by many authors including  
133 Burt (1920) and Demoulin (1985). These specimens feature a cap clothed in coarse  
134 hair that is resistant to wearing off in bands, and these hairs gather in individual  
135 clumps that are best observed with a hand lens or dissecting microscope (FIG. 2A–B).  
136 If bruised (when basidiocarps are fresh) or wetted (when dried) the hymenium  
137 (undersurface) does not stain color (FIG. 2A).

138         Similarly, the “*S. ostrea*” specimens in the second clade (FIG. 1) showed  
139 features consistent with the previously synonymized *S. lobatum* (Burt 1920;  
140 Demoulin 1985). The cap is clothed in matted, felted hairs that quickly begin wearing  
141 off, from the edge of the cap inwards, in concentric bands exposing the chestnut-  
142 brown context beneath (FIG. 2C–D). When bruised or wetted the hymenium stains a  
143 bright yellow color (FIG. 2B).

144 “*S. ostrea*” specimens in the third clade shared some features with both *S. fasciatum*  
145 and *S. lobatum*, however the basidiocarps were usually thicker, more irregular, and a richer  
146 brown color both cap and hymenium. The cap is covered clumping hairs that are typically  
147 longer and woolier than specimens of the first clade (*S. fasciatum*), and wear off in bands  
148 more readily but not to the extent of specimens in the second clade (*S. lobatum*) (FIG. 2E–F).  
149 When bruised or wetted the hymenium stains a bright yellow color like *S. lobatum* (FIG. 2B).  
150 Overall, the morphology agrees with descriptions of *S. subtomentosum* (Pouzar 1964;  
151 Chamuris 1988).

152 Specimens identified before sequencing as belonging to other species of *Stereum*  
153 formed clades with ITS sequences differing within clades by 15%. These included *S.*  
154 *complicatum*, *S. gausapatum*, and *S. sanguinolentum*. The single sequence of *S. striatum* was  
155 placed differently in the two trees but differed from all other sequences by more than 7%. *S.*  
156 *hirsutum* sequences acquired from GenBank and BOLD databases differed from one another  
157 by 1–6%, but only as much as 4% within the two geographically isolated (North America vs.  
158 Eurasia) clades.

## 159 **DISCUSSION**

160 Together, our results support the hypothesis that the North American fungi that have been  
161 lumped under the name “*S. ostrea*” are actually at least three distinct species, reproductively  
162 isolated and differing in morphology. The inference of reproductive isolation is based on the  
163 observation that while all three species can be found growing together on the same substrate,  
164 there is strong concordance between ITS sequence identity and morphology, such that they  
165 appear not to be hybridizing despite often living in sympatry. Demoulin (1985) argued that *S.*  
166 *fasciatum* and *S. lobatum* were distinct species based on differing morphology and lack of  
167 fusion while growing on the same substrate, and our results support this claim with new  
168 genetic evidence.



169           As a consequence of these new data, the *Stereum* in the three “*S. ostrea*”  
170 clades should be recognized as distinct species, and those in synonymy with *S. ostrea*  
171 should revert to their original names. Our FIGS. 2 and 3 along with images for each  
172 specimen on iNaturalist (SUPPLEMENTARY TABLE 1) are provided to assist future  
173 naturalists and researchers in distinguishing among these taxa. Most importantly,  
174 these three species in midwestern and eastern North America can be differentiated by  
175 1) texture of cap hairs, 2) presence and extent of banding, and 3) color staining of the  
176 hymenium when bruised or wetted (FIGS. 2 and 3).

177           A more general conclusion of this research is that an integrative approach to  
178 *Stereum* taxonomy that includes DNA sequence data will be helpful in resolving  
179 species boundaries. In many cases, original descriptions are only a few sentences long  
180 or overly vague. Further, *Stereum* taxonomy has a long history of synonymization  
181 efforts such that descriptions of *Stereum* species from different authors are often  
182 conflicting. Historically, *Stereum* has been used as a genus name to describe many  
183 corticioid fungi, many of which have been transferred to other genera both in the  
184 Stereaceae and to other genetically distant families (Larsson and Larsson 2003).  
185 Many *Stereum* are accepted as synonyms of better-known *Stereum* species, but if our  
186 study is of any indication, some of those names may also need to be resurrected.

187           Our work here demonstrates a need to explore the rest of the *Stereum* genus,  
188 so that we may better understand their ecological roles, how they might be useful to  
189 us, and how they evolved. Molecular phylogenetic approaches will be necessary as a  
190 supplemental tool to delineate *Stereum* species, as morphology alone has proved  
191 inadequate. By using a multilocus approach and worldwide sampling strategy, we can  
192 achieve greater resolution within this genus and other members of the Stereaceae.



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199 encouragement and donation of valuable literature on *Stereum*.

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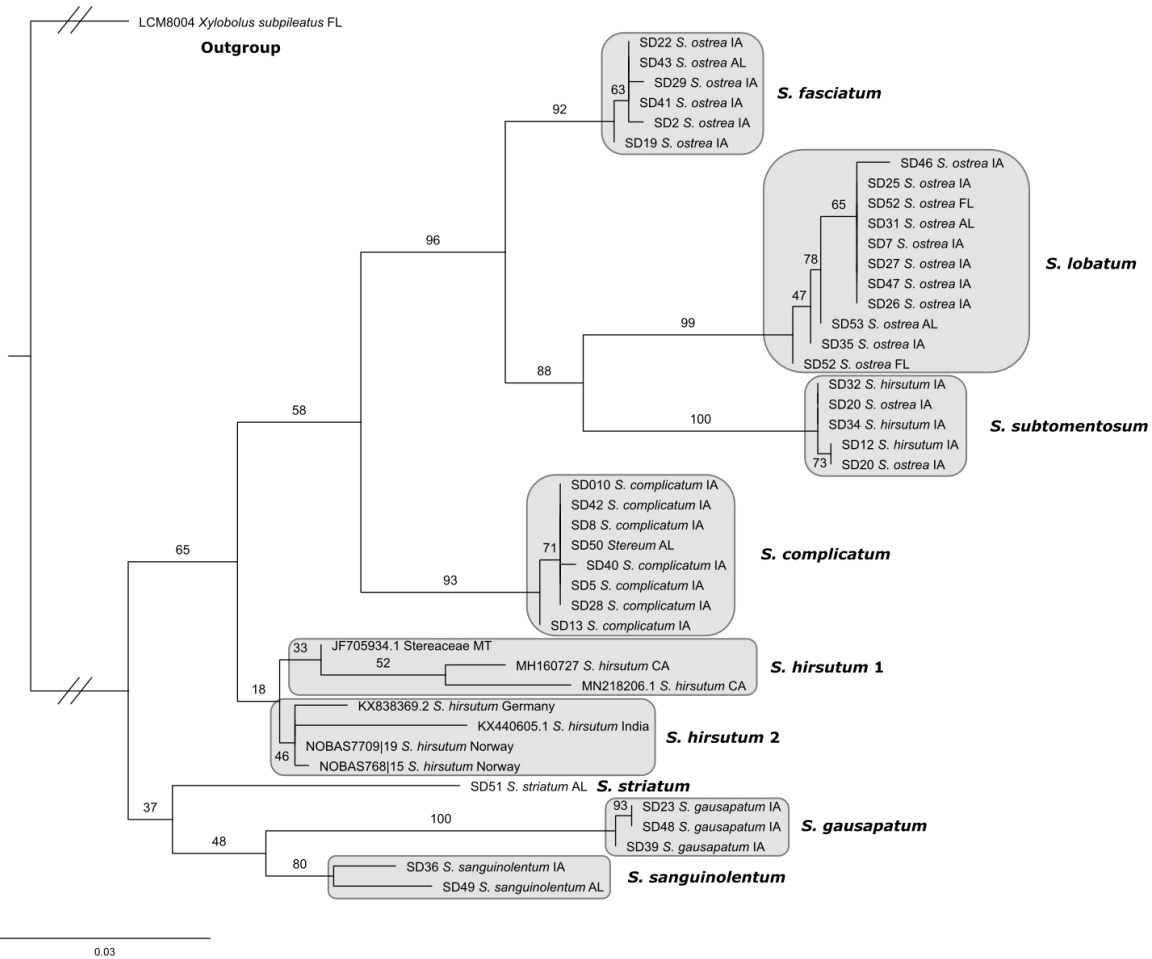
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## 296 LEGENDS

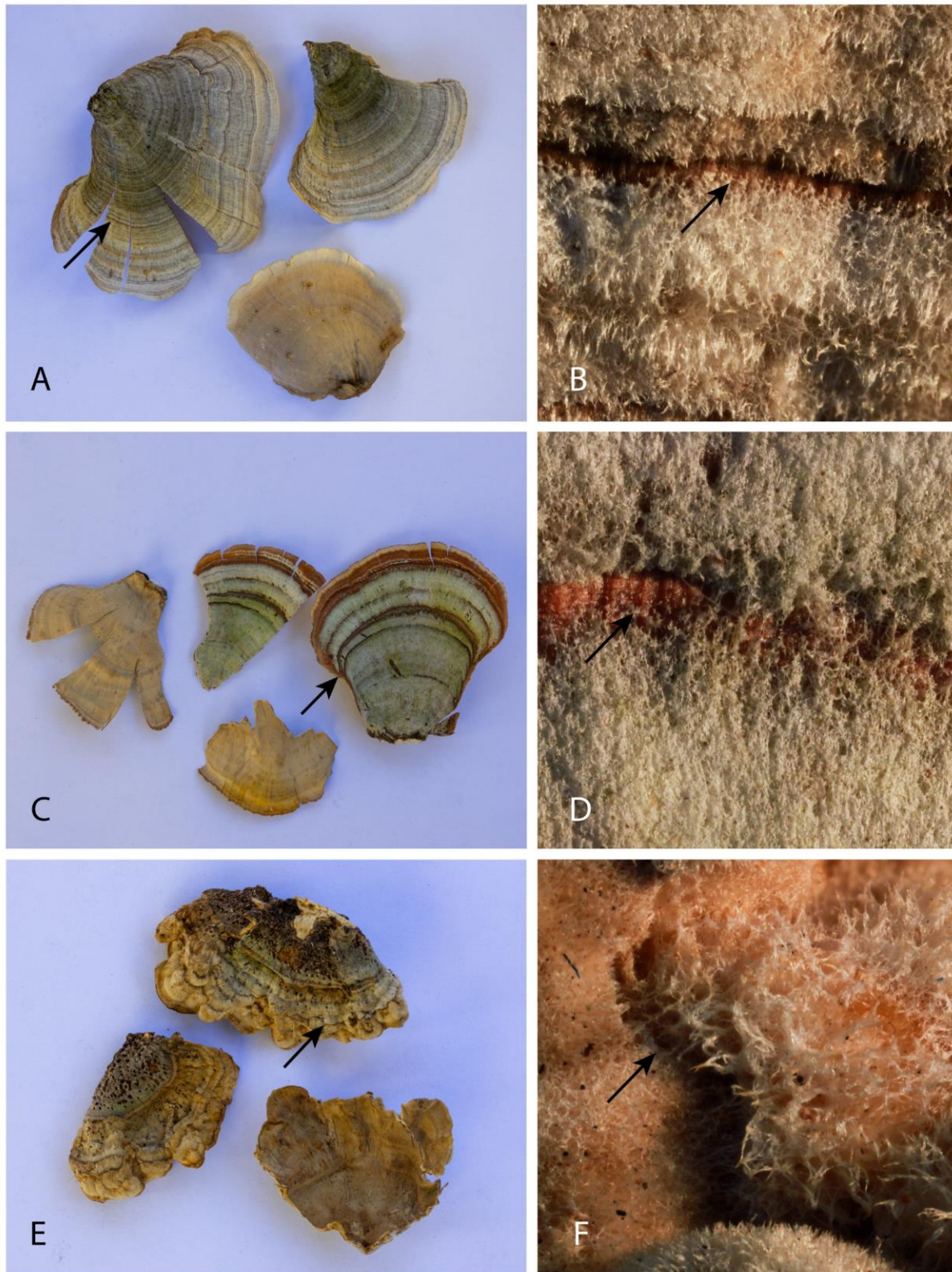
297 **Figure 1.** Maximum likelihood phylogeny generated from ITS sequence data. Bootstrap  
298 values are above branches. Scale bar represents the number of nucleotide changes per site.  
299 The first three clades show strong support, suggesting *S. ostrea* in midwestern and eastern  
300 North America consists of three distinct species, which we identify as *S. fasciatum*, *S.*  
301 *lobatum*, and *S. subtomentosum*.



302

303 **Figure 2.** Morphological comparison of a-b. SD01 *Stereum fasciatum*, c-d. SD03 *S. lobatum*,  
 304 e-f. SD12 *S. subtomentosum*. Arrows point to areas where hairs have worn away in  
 305 concentric bands, exposing the chestnut-brown context; a. few very thin bands, c. wide  
 306 bands, starting from cap edge, and e. irregular, uneven banding. The second column shows  
 307 differences between species in cap hair texture; b. coarse and clumped in tufts, d. short,  
 308 matted, and felted, and f. long, wooly and tufted.





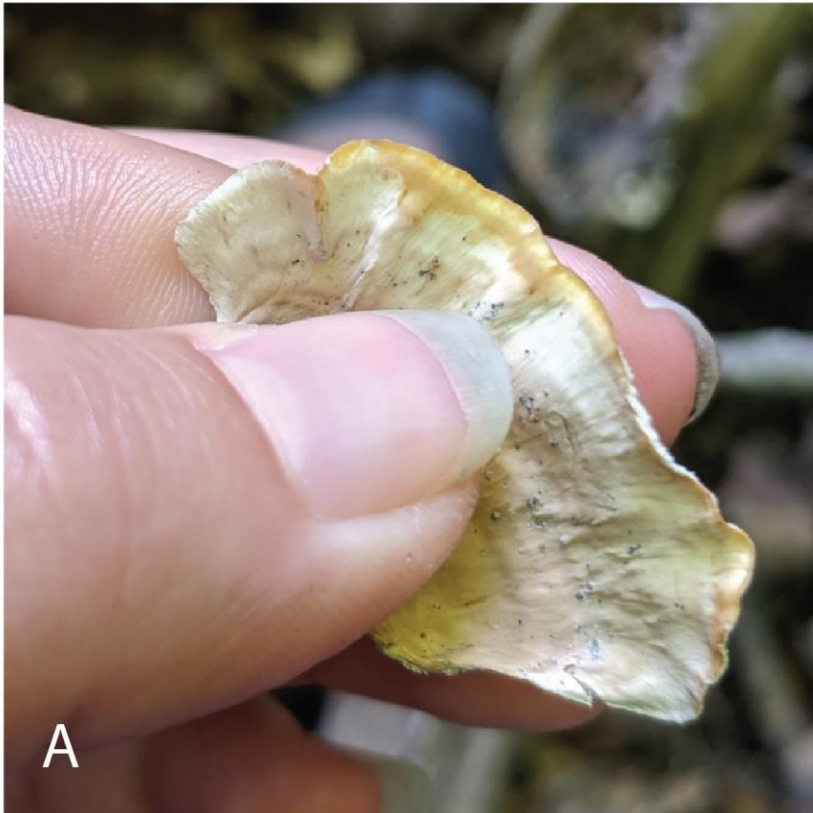
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310 **Figure 3.** Comparison of color staining between a. *S. fasciatum* and b. *S. lobatum*, which

311 show slight darkening, and bright yellow staining, respectively. *S. submentosum* (not



312 shown) exhibits the same bright yellow staining as *S. lobatum*. Note that both specimens  
313 were dry, and re-wetted to investigate color staining.



314