Activation by Cleavage of the Epithelial Na^+ Channel α and γ Subunits Independently Coevolved with the Vertebrate Terrestrial Migration

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Abstract:
Vertebrates evolved mechanisms for sodium conservation and gas exchange in conjunction with migration from aquatic to terrestrial habitats. Epithelial Na^+ channel (ENaC) function is critical to systems responsible for extracellular fluid homeostasis and gas exchange. ENaC is activated by cleavage at multiple specific extracellular polybasic sites, releasing inhibitory tracts from the channel's α and γ subunits. We found that proximal and distal polybasic tracts in ENaC subunits coevolved, consistent with the dual cleavage requirement for activation observed in mammals. Polybasic tract pairs evolved with the terrestrial migration and the appearance of lungs, coincident with the ENaC activator aldosterone, and appeared independently in the α and γ subunits. In summary, sites within ENaC for protease activation developed in vertebrates when renal Na^+ conservation and alveolar gas exchange was required for terrestrial survival.

The migration of vertebrates to a terrestrial habitat began ~380 Mya and was driven by competitive pressure and the relative availability of resources [1]. Survival in a terrestrial environment required several adaptations, including those for respiration, extracellular fluid volume balance and osmoregulation. Aldosterone signaling has a significant role in extracellular fluid homeostasis for tetrapods and depends on three proteins that appeared during the evolution of vertebrates: aldosterone synthase, the mineralocorticoid receptor (MR), and 11β-HSD2 [2]. One of the major endpoints of aldosterone signaling through MR is increased activity of the epithelial Na^+ channel (ENaC) in the kidney, which results in enhanced Na^+ and fluid retention. Having originally evolved in a marine environment, these mechanisms to achieve electrolyte homeostasis may have required adaptive modifications as the ancestors of today's terrestrial vertebrates moved to a relatively dry environment.

ENaCs are heterotrimers comprising α, β, and γ subunits that each have intracellular amino and carboxyl termini, two transmembrane domains, and a large extracellular region (Figs. 1A, B). ENaC subunits belong to the ENaC/Degenerin family of proteins that form trimeric cation-selective channels with similar extracellular folds [3-5]. Aldosterone regulation of ENaC activity and abundance depends on essential sequences in both the extracellular and intracellular domains. These include polybasic tracts in extracellular GRIP domains unique to ENaC subunits; however, the polybasic tracts of ENaC are not present in all vertebrate lineages (Fig. 1B) [4, 6-8]. Aldosterone promotes double cleavage of the α and γ subunits at polybasic tracts, which liberates embedded inhibitory tracts and converts dormant channels to constitutively open ones [9-12], thereby leading to increased fluid retention. ENaC subunits also feature PY motifs on their intracellular C-termini that are important for protein degradation (Fig. 1B) [13, 14]. These motifs recruit WW-domain containing Nedd4-2, whose binding results in the ubiquitylation of ENaC subunit N-termini and enhanced endocytosis and degradation [15, 16]. Nedd4-2 dependent reduction of ENaC abundance is inhibited by aldosterone [16]. These complementary regulatory mechanisms help aldosterone minimize Na^+ wasting and fluid loss during volume contraction.

Here we investigated the evolution of ENaC regulatory mechanisms to determine which features coevolved with the marine-terrestrial transition. We consistently found both activating cleavage sites in the ENaC α and γ subunits of terrestrial vertebrates, while they appeared only sporadically in fishes. Phylogenetic analysis and likelihood ratio tests showed a coevolutionary dependence of the polybasic tracts with each other. They also showed a coevolutionary dependence of tandem polybasic tracts with terrestrial status and with lungs. Analysis of ancestral reconstructions strongly suggest that the polybasic tracts appeared independently in the α and γ subunits. Similar analyses of the PY motif showed no coevolutionary pattern and that the PY motif first arose in an ancient ancestral ENaC subunit. Our data suggest that changes associated with adaptation to terrestrial life provided selective pressure for the development of ENaC activation by cleavage.

Results

ENaC subunit sequence conservation and evolution

The subphylum Vertebrata encompasses the vast majority of species in the phylum Chordata. We identified ENaC subunit sequences in each of the classes of Vertebrata. Notably, although we found ENaC subunits in Sarcopterygii (lobe-finned fishes) and the ray-finned Polypteriformes (ropefish), we did not find ENaC subunits in any other ray-finned fishes. Instead, BLAST searches using human ENaC subunits identified ENaC-related acid-sensing ion channels (ASICs) and uncharacterized ENaC-like proteins. This suggests ray-finned fishes lost all ENaC subunits after the early divergence of the Polypteriformes order and before the divergence of Chondrostei (sturgeons) and Neopterygii (gars and teleosts). Similarly, we only found ENaC-related proteins in non-vertebrate chordates (e.g. tunicates or cephalochordates). To examine sequence conservation within ENaC sequences, we assembled ENaC subunit sequences found in each of the classes, and top teleost and non-vertebrate chordate hits identified in a BLAST search using human ENaC subunits (Supplementary Table 1). Sequence conservation was highest in the transmembrane helices (TM1 and TM2) and defined extracellular secondary structures in the ENaC and ASIC1 structures (Fig. 1C and Supplementary Fig. 1) [3, 4]. Sequence conservation was lowest in the intracellular N- and C-termini, extracellular loops, and the GRIP domain unique to ENaC subunits. Sequences associated with key functions demonstrated varied

conservation. For example, G/S-X-S in TM2 contributing to ion selectivity [17] is well-conserved within a conserved region (Fig. 1C). The PY motif in the intracellular C-terminus is also well conserved, but in a poorly conserved region. In contrast, the region in the GRIP domain containing the embedded inhibitory tracts and cleavage sites is poorly conserved.

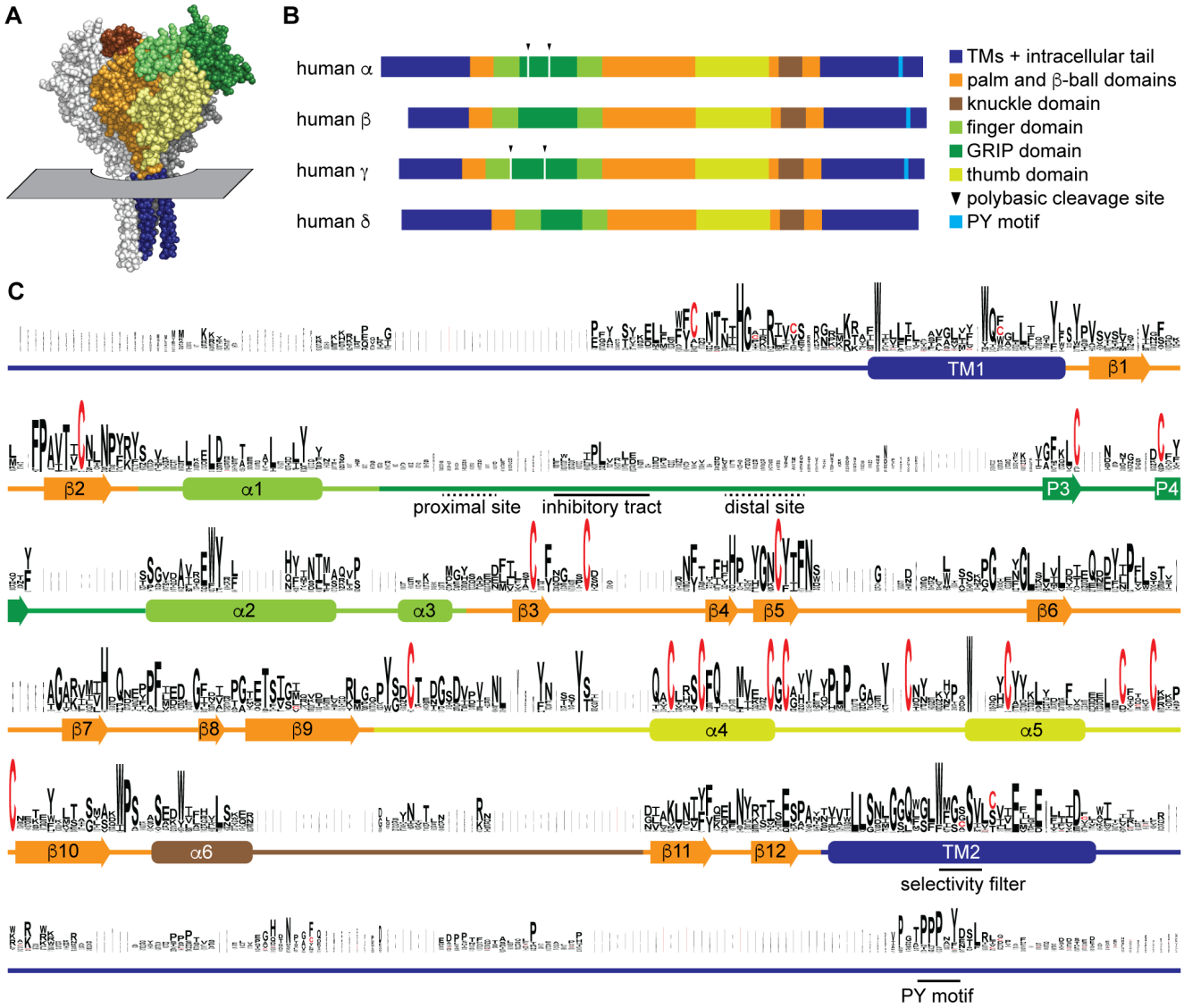

Figure 1. Sequence conservation in ENaC subunits. A, Space filling model of ENaC (pdb code: 6BQN) with plane indicating position of outer membrane border. The α and γ subunits are white and grey, respectively. The β subunit is colored by domain as indicated in panel B. Intracellular structures are absent in this structural model. B, Linear model of human ENaC subunits showing domain organization and highlighting position of polybasic cleavage sites and PY motifs. C, Sequences were aligned using MUSCLE [18]. Residue symbol sizes are proportional to frequency at a given position. Key features in the sequence are indicated, as are the approximate position of α helices (rounded rectangles) and β-strands (arrows). Colors correspond to protein domains, as indicated in panel B. The GRIP domain is unique to ENaC subunits in the ENaC/Deg family. The P1 and P2 β-strands in the GRIP domain are not indicated, but are likely near the inhibitory tract and distal site, respectively, if present.

To examine the evolution of ENaC subunits, we assessed the phylogenetic relationship of our 53 assembled sequences with maximum-likelihood methods (Fig. 2). In the resulting tree, we observed 5 well-supported clades: 1 each for the ENaC α, β, γ, and δ subunits, and one for the non-ENaC proteins. Each ENaC subunit clade largely recapitulated the expected phylogenetic relationships, with jawless fishes (lampreys) rooted closest to the base of the clade, followed by cartilaginous fishes (elephant shark), lobe-finned fishes (coelacanth and lungfishes), reptiles and birds, and mammals. Our evolutionary model suggests three gene duplication events for ENaC subunits, as previously reported [19]. The first duplication (node 1) gave ancestral β/γ subunits (node 2) and α/δ subunits (node 3). The second

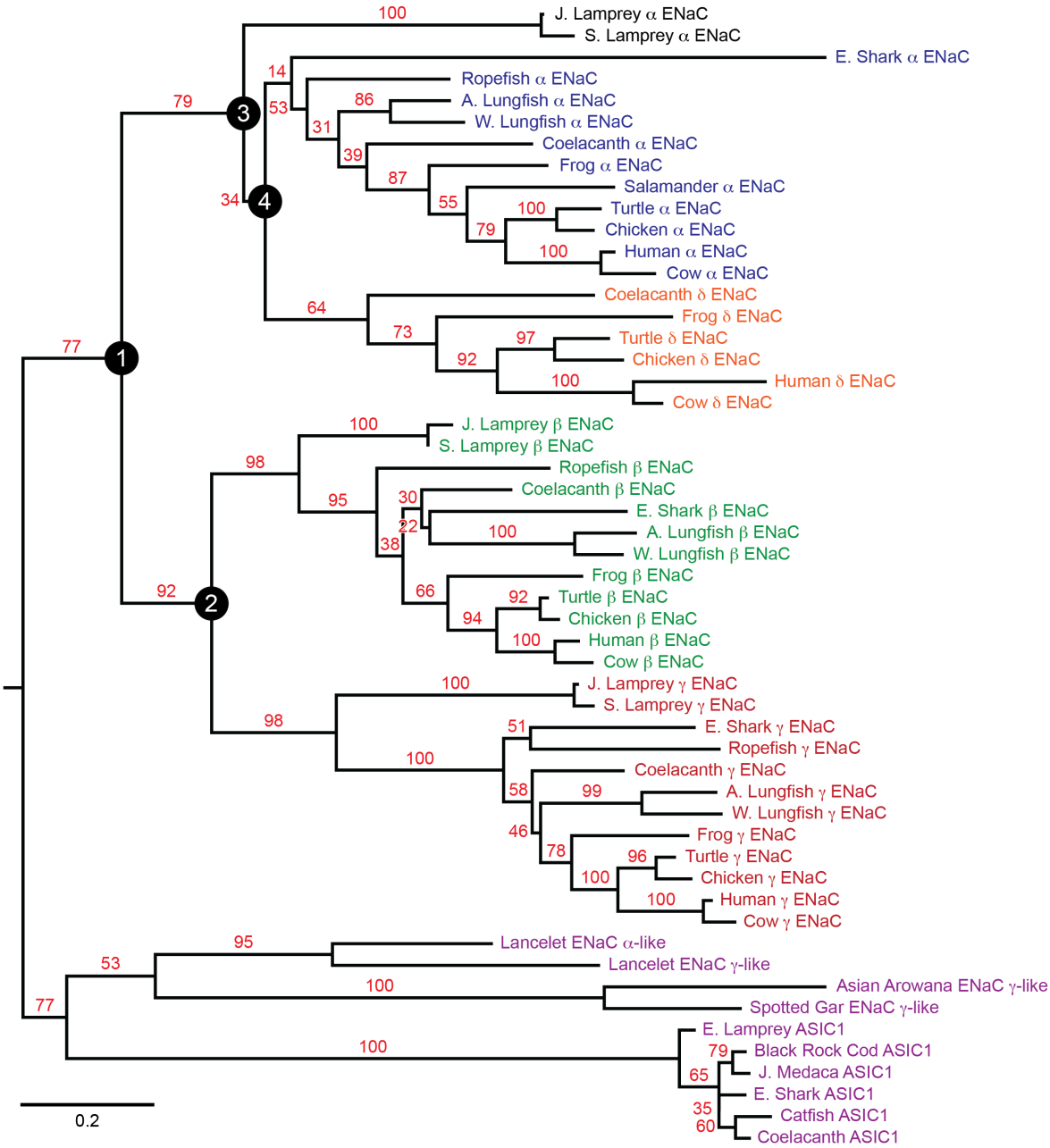

Figure 2. Phylogenetic tree of ENaC subunits. Maximum-likelihood tree calculated from ENaC subunit sequences of marine species and select terrestrial vertebrates, and ENaC-related proteins. Branch support bootstrap values are shown. Key ancestral nodes are indicated by circled numbers. A. Lungfish = Australian Lungfish, E. Shark = Elephant Shark, E. Lamprey = European River Lamprey, J. Lamprey = Japanese Lamprey, J. Medaca = Japanese Medaca, S. Lamprey = Sea Lamprey, W. Lungfish = West African Lungfish.

duplication (node 2) gave ancestral β and γ subunits. Both of these duplication events occurred in ancestors of jawless fishes at least 550 Mya. The third duplication (node 4) led to the divergence of α and δ subunits and occurred in an ancestor of jawed fishes.

Polybasic tracts in the ENaC GRIP domains varied over time

We then examined the sequence conservation of the polybasic tracts in the ENaC subunit GRIP domains required for ENaC activation by cleavage. In mammals, the GRIP domains of the α and γ subunits are each subject to double cleavage, leading to the release of embedded inhibitory tracts and channel activation. The β and δ subunits are not similarly processed. The proprotein convertase furin cleaves both the proximal and distal sites of the α subunit, and the proximal site of the γ subunit [6]. Cleavage distal to the γ subunit inhibitory tract can be catalyzed by several proteases at the cell surface, including prostasin at a polybasic tract [20, 21]. We inspected our multiple sequence alignment for polybasic tracts that aligned closely with the tracts in mammalian α and γ subunits (Supplementary Fig. 1). We found polybasic tracts aligning with the proximal and distal sites of the human α and γ subunits for all tetrapod α and γ subunits (Table 1). We also found both sites present in the γ subunit from *Erpetoichthys calabaricus* (Ropefish) and *Neoceratodus forsteri* (Australian lungfish), but not in *Protopterus annectens* (West African lungfish). In coelacanth, we identified single polybasic tracts in the α, β, and γ subunits. The elephant shark's γ subunit also had a single distal polybasic tract. The human δ subunit sequence also exhibits a polybasic tract in this region, but experimental evidence shows that it is not cleaved [22].

			α		δ		β		γ	
Animal	Terr.	Lungs	site 1	site 2	site 1	site 2	site 1	site 2	site 1	site 2
S. Lamprey			—	—	Ø	Ø	—	—	—	—
J. Lamprey			—	—	Ø	Ø	—	—	—	—
E. Shark			(···)	(···)	Ø	Ø	—	—	—	RQHR
Ropefish		X	—	—	Ø	Ø	—	—	RKRR	NRKR
Coelacanth			RSNR	—	—	—	KRER	—	—	VKQR
W. Lungfish		X	—	—	Ø	Ø	—	—	—	—
A. Lungfish		X	—	—	Ø	Ø	—	—	RKLR	RQYR
Frog	X	X	RVKR	RVSR	—	—	—	—	RSKR	KRTR
Salamander	X	X	RERR	RVRR	Ø	Ø	Ø	Ø	Ø	Ø
Turtle	X	X	RSPR	RHKR	—	—	—	—	KVRR	NKRK
Chicken	X	X	RTSR	RQKR	—	—	—	—	KVRR	RKRK
Cow	X	X	RSRR	RGVR	—	*RLQR*[1]	—	—	RKRR	RKRK
Human	X	X	RSRR	RRAR	—	*RLQR*[1]	—	—	RKRR	RKRK

Table 1. Polybasic sequences aligning with human ENaC subunit proximal (site 1) and distal (site 2) cleavage sites. Terr., terrestrial. Dashes indicate absence of polybasic tract. [1]The ENaC δ subunit in mammals has a polybasic sequence in the aligned region, but is not cleaved in human channels [22]. Ø, no sequence was available. (···), GRIP domain sequence was missing in the available sequence. Species abbreviations are as in Fig. 2.

GRIP domain cleavage sites coevolved with each other

Activation by cleavage requires two cleavage events within a single subunit and the release of embedded inhibitory tracts [7, 23]. The data in Table 1 suggest that sites 1 and 2 coevolved with each other. To test this idea, we calculated the probability that the sites coevolved with each other using nested likelihood models based on our model of ENaC evolution in BayesTraits (see Supplementary Table 2) [24, 25]. Each sequence was assigned the traits of the presence or absence of "site 1" and "site 2", as indicated in Table 1. All models considered the evolutionary gains and losses of these traits over the full phylogenetic tree capturing the 4 ENaC subunits (Fig. 2).

To test for dependence between the two sites, we contrasted two nested models, an independent model and a dependent model. The independent model contained two parameters: an appearance rate

for both sites and a loss rate for both sites. The loss or gain of a site did not depend on the status of the other site, and therefore served as the null hypothesis. The dependent model added two free parameters by having separate gain and loss rates depending on the status of the other site. A likelihood ratio test between nested models showed a strong preference for the dependent model (p=0.01). Fits to the dependent model suggest little selection pressure in the change from 0 sites to 1 site, with loss rates 11-fold higher than gain rates. In contrast, in the change from 1 site to 2 sites, gain rates were 7-fold higher than loss rates. This is congruent with the requirement for two cleavage sites within a single subunit's GRIP domain for channel activation, and supports the notion that selection pressure derives from the functional consequence of double cleavage [7, 23].

GRIP domain double cleavage coevolved with the terrestrial migration and with lungs

The data in Table 1 also suggest that the terrestrial migration or the development of lungs may have provided the selection pressure for channel activation by cleavage. ENaC function is also important in aldosterone-insensitive tissues, including the airway, where control of airway surface liquids is essential for lung function. We first calculated the likelihood of site 1, site 2, or both sites coevolving with the terrestrial transition using nested likelihood models. Each sequence was assigned the traits of the presence or absence of "site 1", "site 2", or "tandem sites", and "terrestrial or marine" states, as indicated in Table 1. The independent model contained three parameters: a site appearance and loss rate, and a rate for transition from marine to terrestrial status (see Supplementary Table 2). The independent model does not allow a relationship between rates of site gain or loss with respect to marine or terrestrial state, and therefore served as the null hypothesis. The dependent model added two free parameters by having separate site gain and loss rates in the marine and terrestrial states. A likelihood ratio test between nested models showed a strong preference for the dependent model for site 1 (p=0.0053) and for tandem sites (p=0.0098), but not for site 2 (p=0.26). Fits to the dependent model suggest that in the marine state, site appearance and disappearance was dynamic, with a disappearance rate being 5-fold higher than the appearance rate. However, once in the terrestrial state, both rates dropped to 0, supporting the notion of selection pressure favoring the presence of cleavage sites in the terrestrial state. We then calculated the likelihood of site 1, site 2, or both sites coevolving with the development of lungs using analogous procedures. A likelihood ratio test between nested models showed a preference for the dependent model for site 1 (p=0.031) and for tandem sites (p=0.018), but not for site 2 (p=0.94).

ENaC expression in fishes with lungs and amphibians

Lungs coevolved with the terrestrial migration of vertebrates (p=0.0004 in our dataset), and may have driven the coevolution of GRIP domain polybasic tracts with lungs that we observed. However, ENaC subunit transcripts were not detected in lung tissues from either of the lungfish species examined [26, 27]. To further investigate the role of ENaC in fishes with lungs, we examined the tissue distribution of ENaC transcripts in ropefish, which evolved lungs independently from tetrapods [28]. We observed clear bands for the transcripts of all three ENaC subunits in the gills and kidneys of the ropefish (Fig. 3A), which are important organs for ion homeostasis. In the lung, we observed a clear band for the β subunit, a faint band for the α subunit, and a barely perceptible band for the γ subunit. In the other tissues examined, we observed broad expression of β subunit transcripts, faint bands for the α subunit in the liver and intestine, and no other bands for the γ subunit. Taken together with the lungfish data [26, 27], these data suggest that transcripts for ENaC subunits with GRIP domain polybasic tracts (Table 1) are readily detected at important sites of ion exchange, but are difficult to detect in lungs.

We also investigated the tissue distribution of ENaC subunit transcripts from the African clawed frog, in which both the α and γ subunits have GRIP domain cleavage sites [29]. Similar to mammals, we detected bands for α, β, and γ subunit transcripts in both kidneys and lungs (Fig. 3B). In contrast to humans where bands for the δ subunit were relatively faint for both kidneys and lungs [30], we observed a strong band for the δ subunit transcript in the kidney, and no band in the lung. The expression pattern we observed is consistent with a previous investigation of α and δ subunit expression in *Xenopus laevis* tissues [29], and suggest that cleavage regulates ENaC function in both the lungs and kidneys of frogs, similar to mammals.

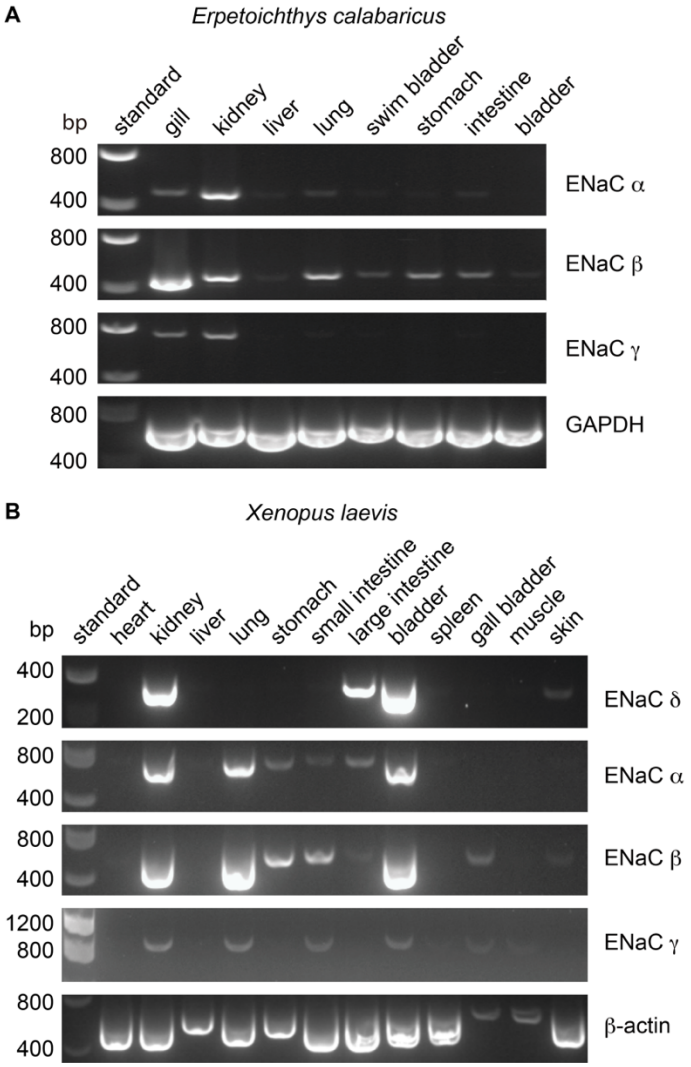

Figure 3. Tissue distribution of *Erpetoichthys calabaricus* (ropefish) and *Xenopus laevis* ENaC subunit transcripts by RT-PCR. cDNA libraries were generated from tissue homogenates. PCR reactions were performed using primers indicated in Supplementary Table 3.

α and γ subunit GRIP domain cleavage sites evolved independently

The absence of cleavage sites in species rooted closest to the early gene duplication events suggests that each cleavage site did not result from a common ancestor, but rather appeared independently in the α and γ subunits. To test this idea, we performed ancestral reconstruction of nodes 1-4 in our phylogenetic tree (Fig. 2) using BayesTraits and MCMC methods. In the unconstrained evolutionary model, each of these nodes were free to adopt any status for the site of interest. In the divergent evolutionary model, nodes 1-4 were constrained to contain the site of interest. In the convergent evolutionary model, node 1 was constrained to exclude the site of interest. Models were compared by converting marginal likelihoods resulting from MCMC runs of each model to log Bayes Factors, where values of >2, 5-10, and >10 support positive, strong, and very strong preferences for the better fitting model, respectively. Within the framework of sites 1 and 2 coevolving with each other, comparison of the unconstrained evolutionary model with the convergent evolutionary model resulted in log Bayes Factors of 0.38 for site 1 and 0.34 for site 2, supporting no preference between models. Comparison of the divergent evolutionary model to the convergent model for each of the sites resulted in log Bayes Factors of 6.5 in both cases, supporting a strong preference for the convergent evolutionary model for each of the cleavage sites. This suggests that, despite analogous molecular mechanisms in the α and γ subunits,

the appearance of cleavage sites in the α and γ subunits were independent events. Neither the proximal nor the distal cleavage site arose from a common pre-α/γ ENaC subunit.

ENaC PY motifs evolved through divergent evolution

Our data suggest that cleavage of the α and γ subunits evolved contemporaneously with aldosterone synthase. Indeed, aldosterone enhances activating cleavage of ENaC subunits [11, 12]. Due to the apparent connection to aldosterone, we examined the evolution of the PY motifs in the C-termini of ENaC subunits, which are also targets of aldosterone-dependent regulation. The PY motifs facilitate enhanced channel turnover via Nedd4-2, which can be inhibited by aldosterone and other hormones through kinase signaling [31]. Mutation or deletion of the PY motifs increases ENaC function, leading to Liddle syndrome which is characterized by hypertension and hypokalemia. We found PY motifs in all α, β, and γ subunits where sequences of the C-termini were available (Table 2, Supplemental Fig. 1), suggesting a divergent evolutionary model that did not depend on terrestrial status. Interestingly, δ subunits apparently lost their PY motifs after their divergence from the α subunit in the coelacanth ancestor. We tested whether appearance of the PY motif depended on the terrestrial migration or the development of lungs using nested likelihood models, as above. A likelihood ratio test between the nested models showed no preference for the dependent model for either terrestrial status (p=0.54) or lungs (p=0.28), suggesting their evolution was affected by neither. To test whether the PY motifs evolved from a common ancestor, we tested models of PY motif evolution using ancestral reconstruction at nodes 1-4 in Fig. 3 and MCMC methods. The unrestricted model suggested probabilities of a PY motif present at 98.1%, 99.3%, 91.5%, and 75.9% for nodes 1, 2, 3, and 4, respectively. We then tested a series of restricted models to test competing evolutionary models. In the divergent evolutionary model, nodes 1-4 were restricted to contain PY motifs. Comparison of the unrestricted and divergent evolutionary models resulted in a log Bayes Factor of 0.19, suggesting these models were nearly equivalent. In convergent evolutionary models, one of the four early nodes was restricted to not contain a PY motif, resulting in 4 models. Comparison of the divergent model to each of the convergent models resulted in log Bayes Factors ranging from 4.7 to 8.9, suggesting a strong preference for the divergent model over each of the non-divergent models. These data suggest that the PY motif in each subunit, in contrast to the polybasic tracts, arose from a common progenitor.

Animal	Terrestrial	Lungs	α	δ	β	γ
S. Lamprey			PPPSF	Ø	PPPHY	PPPQY
J. Lamprey			PPDY	Ø	PPPHY	PPPQY
E. Shark			(…)	Ø	PPPRY	PPPNY
Ropefish		X	PPPAY	Ø	PPPHY	PPPNY
Coelacanth			PPAY	(…)	PPPNY	PPPTY
W. Lungfish		X	PPPAY	Ø	PPPHY	PPPQY
A. Lungfish		X	PPPAY	Ø	PPPKY	PPPQY
Frog	X	X	PPPAY	—	PPPNY	PPPKY
Salamander	X	X	PPPAY	Ø	Ø	Ø
Turtle	X	X	LPSY	—	PPPNY	PPPNY
Chicken	X	X	LPSY	—	PPPNY	PPPNY
Cow	X	X	PPPAY	—	PPPNY	PPPRY
Human	X	X	PPPAY	—	PPPNY	PPPKY

Table 2. PY motifs (L/P-P-X-Y) in the C-terminal tails of ENaC subunits from various species. Ø, no sequence was available. (…), sequence for the C-terminal region was missing in the available sequence. Species abbreviations are as in Fig. 2.

Discussion

Our results suggest that the two ENaC GRIP domain cleavage sites in each of the α and γ subunits co-evolved with each other, but appeared independently in each subunit. Our results also suggest that double cleavage coevolved with both the terrestrial migration of vertebrates and the development of lungs (Fig. 4). Co-evolution of site 1 with site 2 makes sense given that functional consequences of cleavage in mammalian ENaCs only result from double cleavage of a single subunit followed by liberation of the intervening inhibitory tract. We found that the polybasic tracts appeared sporadically in the GRIP domain of ENaC subunits from marine species, but appeared and remained consistently in terrestrial vertebrates. Terrestrial life required adaptive changes to overcome the stresses of electrolyte regulation and gas exchange. Co-evolution of the terrestrial migration with tandem GRIP polybasic tracts, and consequently higher ENaC activity, suggests that selection pressure could have arisen from either stressor.

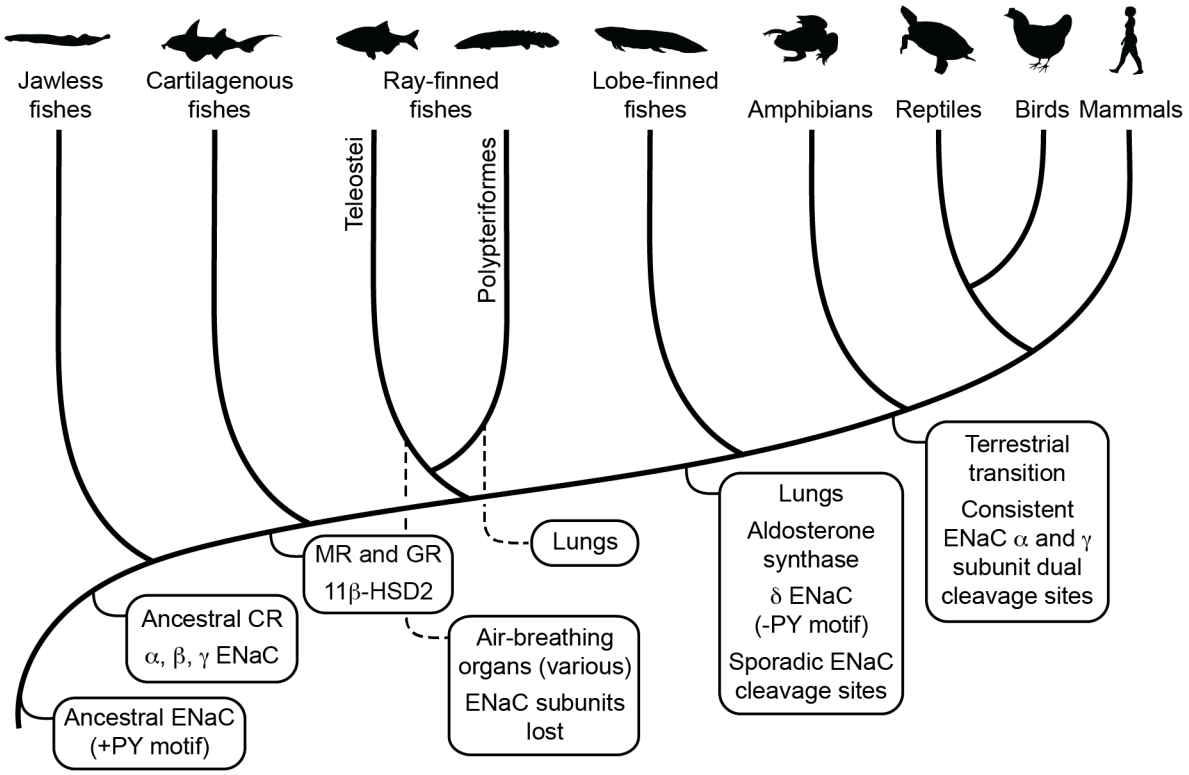

Figure 4. Schematic view of evolution of aldosterone signaling, air-breathing organs, ENaC, and ENaC regulatory motifs. ENaC subunits were not found in non-vertebrate chordates or in teleosts. The ancestral ENaC subunit likely had a PY motif and was a substrate for Nedd4-2-dependent regulation. ENaC α, β and γ subunits appeared before the emergence of jawless fishes, whereas ENaC δ subunits first appeared in an ancestor of the lobe-finned coelacanth. Proteins required for aldosterone signaling (MR, 11β-HSD2, and aldosterone synthase) evolved before the emergence of tetrapods. Lungs appeared in lobe-finned fishes on the lineage to tetrapods. Air-breathing organs (e.g. respiratory gas bladders and labyrinth organs) evolved independently in ray-finned fishes, including morphologically distinct lungs in Polypteriformes. Individual GRIP domain ENaC cleavage sites first appeared sporadically in marine species. Dual cleavage sites appeared consistently in the ENaC α and γ subunits in terrestrial vertebrates. CR, corticoid receptor; MR, mineralocorticoid receptor; GR, glucocorticoid receptor. Animal silhouettes courtesy of PhyloPic (http://www.phylopic.org).

Air-breathing organs evolved independently several times as an adaptation to chronic or periodic environmental hypoxia [28]. Lungs appeared in lobe-finned fishes after their divergence from other bony fishes, and developmental and morphological evidence support homology between the lungs of lungfishes and tetrapods. Extant deep-water coelacanths possess vestigial lungs that correspond to the likely functional lungs of the Cretaceous shallow-water coelacanth *Axelrodichthys* [32, 33]. Air-breathing organs evolved several times in ray-finned fishes, giving *Polypteridae* (ropefish) morphologically distinct lungs, and other ray-finned fishes air-breathing capabilities through modified gas bladders and labyrinth organs (Fig. 4). Notably, we identified ENaC subunits in ropefish and not in any other ray-finned fish,

suggesting ENaC subunits were lost soon after the divergence of Polypteriformes from ray-finned fishes in the Devonian period ~360 Mya [34]. Their loss reflects a lack of selective pressure to maintain the ENaC subunits, potentially because ray-finned fishes have an alternative method to maintain Na^+ and fluid homeostasis. Airway surface liquids are critical to the function of mammalian lungs and are regulated by ENaC and other ion channels [35]. ENaC knockout mice die of asphyxiation shortly after birth due to an inability to clear fluid from the lung [36], and ENaC polymorphisms have been associated with lung dysfunction [37, 38]. ENaC regulation by cleavage is relevant in primary airway epithelial cell cultures [39, 40], where ENaC is regulated by glucocorticoids rather than by aldosterone [41]. However, transcripts of ENaC subunits with GRIP domain polybasic tracts were not detected in the lungs of lungfishes [26, 27] and were faint in the lungs of the ropefish (Fig. 3). As lungs coevolved with the terrestrial migration of vertebrates, concurrent stresses may have provided selection pressure for ENaC activation by cleavage.

Emersion from an aquatic environment poses a risk of desiccation and a challenge to terrestrial life [42]. Aldosterone signaling through MR regulates electrolyte balance and total body volume in tetrapods. Key molecules required for aldosterone signaling appeared at various points during the evolution of vertebrates (Fig. 4) [2]. Interestingly, aldosterone synthase is absent in cartilaginous and ray-finned fishes, but appears in lobe-finned fishes on the lineage to terrestrial vertebrates [43]. The appearance of aldosterone synthase coincided with the appearance of the ENaC δ subunit, and preceded the consistent appearance of tandem polybasic tracts in the ENaC α and γ subunits. In ray-finned fishes, osmoregulation and euryhaline adaptation primarily occur through glucocorticoid receptor signaling [44]. Teleost MR signaling is implicated in central nervous system functions that include sodium appetite, but plays a secondary role in ion transporter regulation in osmoregulatory organs, in contrast to MR signaling in terrestrial vertebrates. Notably, aldosterone promotes cleavage of the ENaC α and γ subunits in mammals through MR signaling [11, 12]. Analysis of transcripts from ropefish and lobe-finned lungfishes show expression of ENaC α, β, and γ subunit transcripts in the gills and kidneys (Fig. 3), which are important sites of ion regulation in fishes [26, 27]. Furthermore, ENaC α subunit mRNA levels were positively correlated with plasma aldosterone in the West African lungfish *Protopterus annectins*, suggesting an important role for aldosterone and ENaC in electrolyte homeostasis in lobe-finned fishes [26].

The evolution of ENaC GRIP domain cleavage sites contrasts with the divergent evolution of PY motifs in the C-termini, which appear in all extant α, β, and γ subunits, but was lost in δ subunits that first appeared in a coelacanth ancestor. The presence of both ENaC subunit PY motifs and Nedd4-like proteins in ancestors of jawless fishes [45] suggests that aldosterone signaling co-opted Nedd4-2 dependent regulation. Differences in physiologic roles may underlie loss of the PY motif in ENaC δ subunits. ENaC δ subunit tissue distribution aligns poorly with aldosterone sensitivity, and is different than for the other ENaC subunits [46]. Despite the lack of PY motifs, δ subunits can be ubiquitylated by Nedd4-2, presumably through binding β or γ subunit PY motifs [47]. This is possible where subunit expression overlaps (e.g. lung and esophagus), but there are neuronal and reproductive tissues where only the δ subunit has been detected.

In summary, we found that two forms of ENaC regulation modulated by aldosterone signaling emerged through distinct evolutionary patterns. Channel activation by double cleavage of the α and γ subunits coevolved with the terrestrial migration of vertebrates, closely paralleling that of aldosterone synthase and likely as a mechanism to enhance Na^+ conservation and fluid retention. Regulation of cell-surface stability through PY-motif interactions with Nedd4-2 was likely present in Cambrian period ancestral ENaC subunits and preceded the development of aldosterone signaling.

Materials and Methods

Sequence retrieval, alignment, phylogenetic tree calculation

Human ENaC subunit protein sequences were used as query sequences in BLAST searches of databases (see Supplementary Table 1). Organism restrictions and organism specific databases were used to identify proteins in each of the major vertebrate classes. Alignment using MUSCLE 3.8.31 [48],

curation using Gblocks 0.91b [49], and phylogenetic tree calculation using PhyML [50] were performed using Phylogeny.fr [51]. The final tree was calculated using the *smaller final blocks* option in Gblocks and the *bootstrapping procedure* option (100 bootstraps) in PhyML. Residue frequency in the aligned sequences were visualized using WebLogo [52]. Trees were visualized using FigTree (http://tree.bio.ed.ac.uk/software/figtree/).

Motif analysis

Furin cleaves human α ENaC after RSRR[178] and RRAR[204], and human γ ENaC at RKRR[138] [6]. Furin requires Arg at P1 and has a preferred P4-P3-P2-P1↓ = R-X-R/K-R↓ substrate sequence, although deviations at P2 and P4 have been observed, e.g. Ala-203 at P2 in the ENaC α subunit [53, 54]. Prostasin cleaves the human ENaC γ subunit after RKRK[181] [7]. Prostasin cleavage requires Arg or Lys at P1, and prefers basic residues at P2, P3, and P4 [55]. We inspected the region in the alignment that contains the human ENaC cleavage sites (Supplementary Fig. 1) for ideal furin sites or alternatively, for polybasic tracts ending in Arg that aligned within 5 residues of either of the α subunit furin sites for α and δ subunits, the γ subunit furin site for β and γ subunits, or for any of the furin sites for ASIC subunits. We also inspected the alignment for polybasic tracts ending in Arg or Lys that aligned within 5 residues of the human γ subunit prostasin site for β and γ subunits. For the PY motif (L/P-P-X-Y) required for Nedd4-2 dependent regulation [56], we inspected the C-terminal region of the alignment. All sequences found aligned with human ENaC subunit PY motifs in the C-termini.

Testing phylogenetic models of site gain and loss

BayesTraits V3 [24, 25] was used to compare evolutionary models of two binary traits using maximum likelihood methods, as described above. Traits were assigned as indicated in Tables 1 and 2: absent, present, or ambiguous (in cases where the relevant region was missing). ASICs in our dataset were all assigned "marine", and only coelacanth ASIC2 was assigned as having a polybasic tract at site 2 (Supplementary Fig. 1). BayesTraits run parameters are provided in Supplementary Table 2. Nested models were compared using a likelihood ratio test, with degrees of freedom equal to the difference in the number of parameters for each model. The likelihood ratio statistic is calculated as: likelihood ratio = 2×{log-likelihood(dependent model) – log-likelihood(independent model)}, and was converted to a p-value using the *chisq.dist.rt* function of Excel. BayesTraits was also used to determine whether traits likely appeared independently (convergent model), or derived from a common ancestor (divergent model). Program parameters are provided in Supplementary Table 2. Values of a given trait at all key ancestral nodes were determined while trait values at specific nodes were fixed to reflect each of the hypothetical evolutionary models. Maximum likelihood runs of each model were used to determine likely average values for model parameters. Uniform priors were selected for parameters with large expected values, and exponentially distributed priors with a mean of 0.001 were selected for parameters with small expected values. MCMC runs for each model were performed for the default number of iterations (1,010,000), and generated model parameters with means similar to those from maximum likelihood runs, consistent with convergence. The stepping stone sampler (100 stones with 10,000 iterations) was used to calculate log marginal likelihood values from MCMC runs. Log marginal likelihood values were converted to log Bayes Factors {log Bayes Factors = 2×(log marginal likelihood model 1 – log marginal likelihood model 2)} for model comparisons. Values of log Bayes Factors were used as evidence for model preference, where <2 was weak evidence, >2 was positive evidence, 5-10 was strong evidence, and >10 was very strong evidence.

Tissue distribution of ENaC subunit transcripts

All animals were handled according to approved institutional animal care and use committee (IACUC) protocols of the University of Pittsburgh. Total RNA was extracted from various tissues isolated from *Erpetoichthys calabaricus and Xenopus laevis* using TRIzol Reagent (Invitrogen) and denatured by heating at 70 ºC for 10 min. Using 2 µg of RNA isolated from each tissue, cDNA libraries were synthesized using RevertAid First Strand cDNA Synthesis Kit (ThermoFisher). Specific primers for the frog ENaC β and γ subunits, each of the ropefish ENaC subunits, and glyceraldehyde-3-phosphate dehydrogenase

(GAPDH) as an internal standard (Supplementary Table 3) were designed using the NCBI Primer-Blast tool (https://www.ncbi.nlm.nih.gov/tools/primer-blast/). Primers for frog α and δ ENaC subunits and β-actin were previously described [29]. All primers were custom synthesized (Integrated DNA Technologies). PCR was run for 30 cycles using GoTaq® G2 Green master mix (Promega) and specific primers, with the annealing temperature set at 60 ºC. PCR products were visualized by agarose gel electrophoresis.

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Competing Interests

The authors declare no conflicts of interest.

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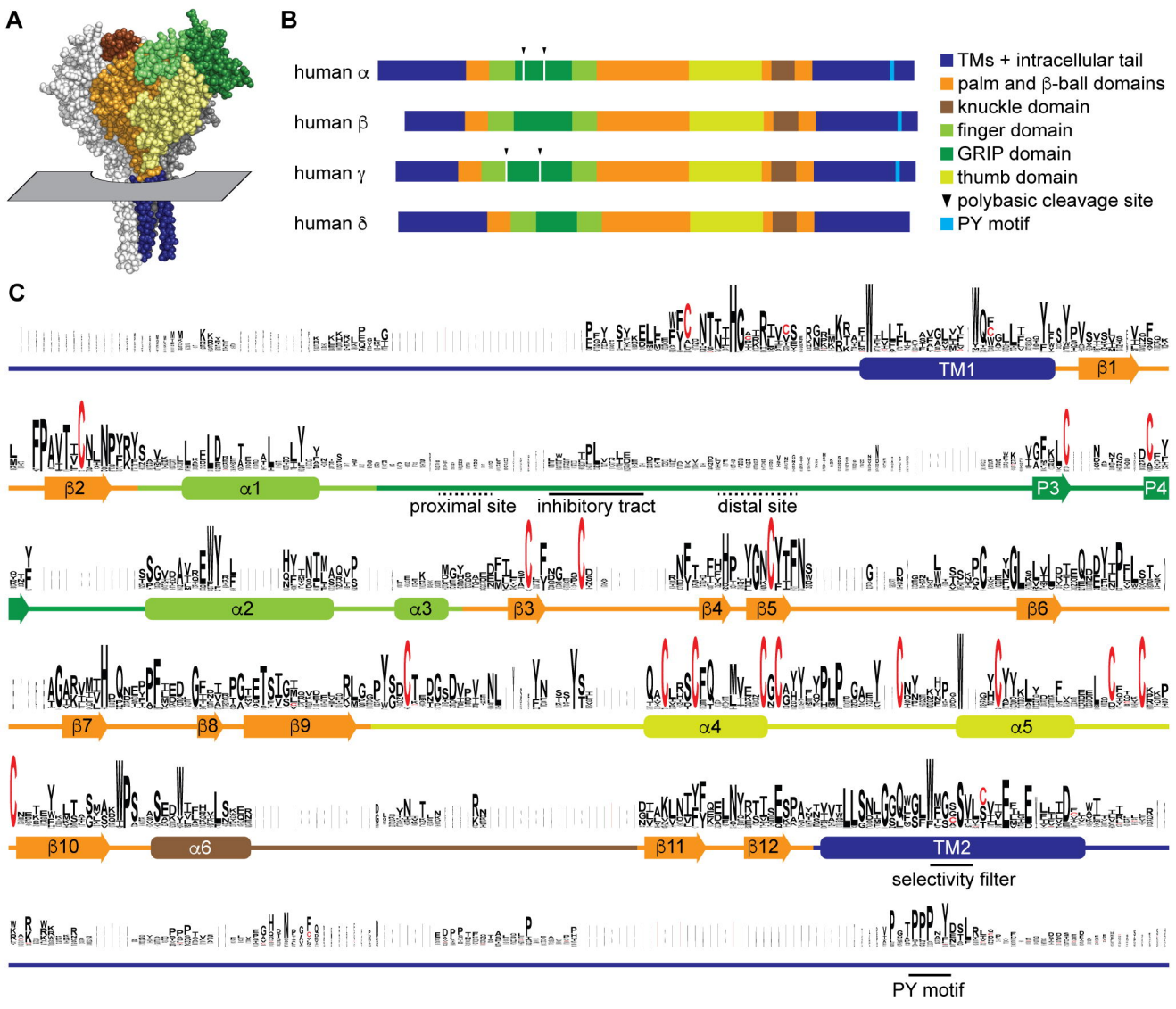
A
B
human α
human β
human γ
human δ
TMs + intracellular tail
palm and β-ball domains
knuckle domain
finger domain
GRIP domain
thumb domain
polybasic cleavage site
PY motif
C
TM1
β1
β2
α1
P3
P4
proximal site
inhibitory tract
distal site
α2
α3
β3
β4
β5
β6
β7
β8
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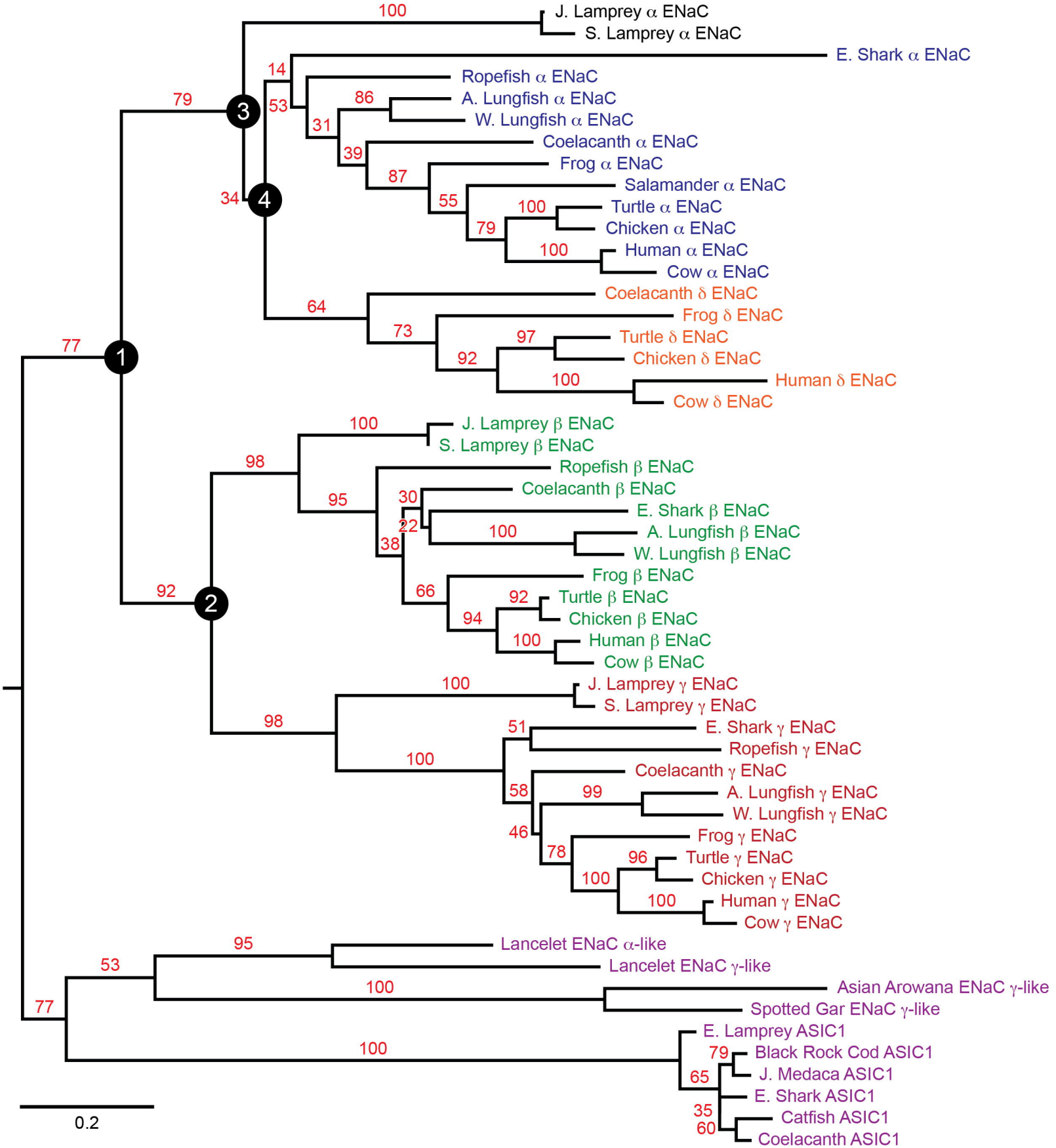
α6
β11
β12
TM2
selectivity filter
PY motif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S. Lamprey α ENaC
E. Shark α ENaC
Ropefish α ENaC
A. Lungfish α ENaC
W. Lungfish α ENaC
Coelacanth α ENaC
Frog α ENaC
Salamander α ENaC
Turtle α ENaC
Chicken α ENaC
Human α ENaC
Cow α ENaC
Coelacanth δ ENaC
Frog δ ENaC
Turtle δ ENaC
Chicken δ ENaC
Human δ ENaC
Cow δ ENaC
J. Lamprey β ENaC
S. Lamprey β ENaC
Ropefish β ENaC
Coelacanth β ENaC
E. Shark β ENaC
A. Lungfish β ENaC
W. Lungfish β ENaC
Frog β ENaC
Turtle β ENaC
Chicken β ENaC
Human β ENaC
Cow β ENaC
J. Lamprey γ ENaC
S. Lamprey γ ENaC
E. Shark γ ENaC
Ropefish γ ENaC
Coelacanth γ ENaC
A. Lungfish γ ENaC
W. Lungfish γ ENaC
Frog γ ENaC
Turtle γ ENaC
Chicken γ ENaC
Human γ ENaC
Cow γ ENaC
Lancelet ENaC α-like
Lancelet ENaC γ-like
Asian Arowana ENaC γ-like
Spotted Gar ENaC γ-like
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Black Rock Cod ASIC1
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Catfish ASIC1
Coelacanth ASIC1
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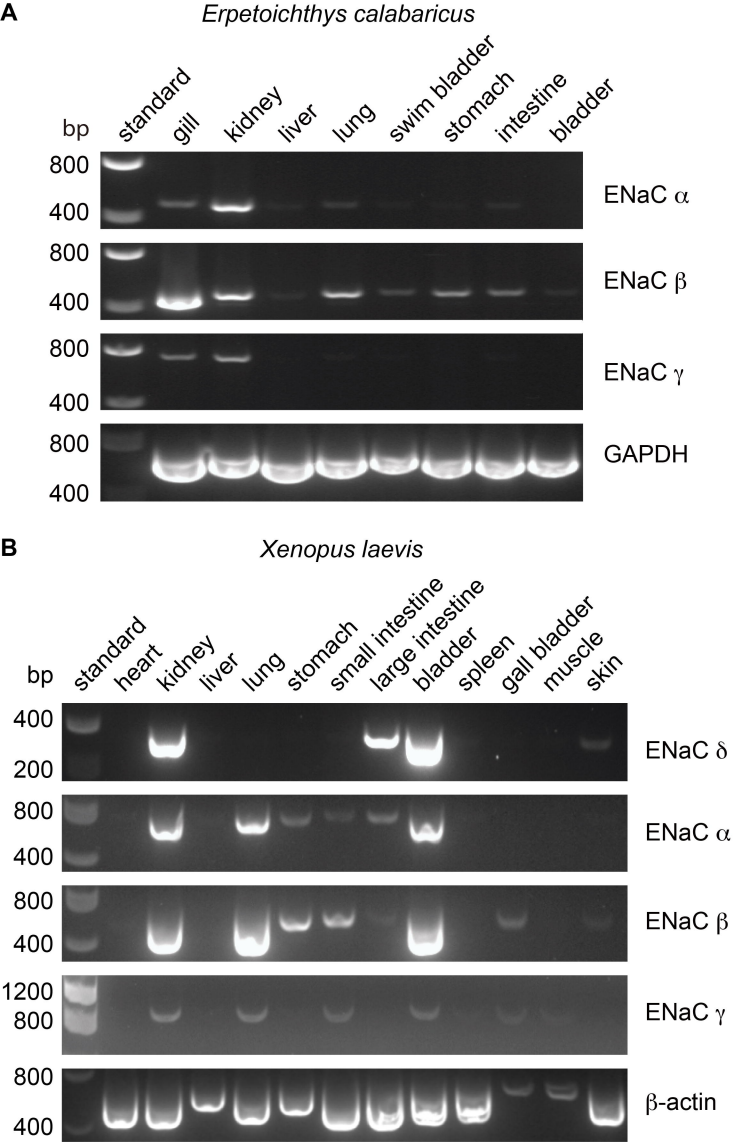
A
Erpetoichthys calabaricus
bp
standard
gill
kidney
liver
lung
swim bladder
stomach
intestine
bladder
800
400
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800
400
ENaC β
800
400
ENaC γ
800
400
GAPDH
B
Xenopus laevis
bp
standard
heart
kidney
liver
lung
stomach
small intestine
large intestine
bladder
spleen
gall bladder
muscle
skin
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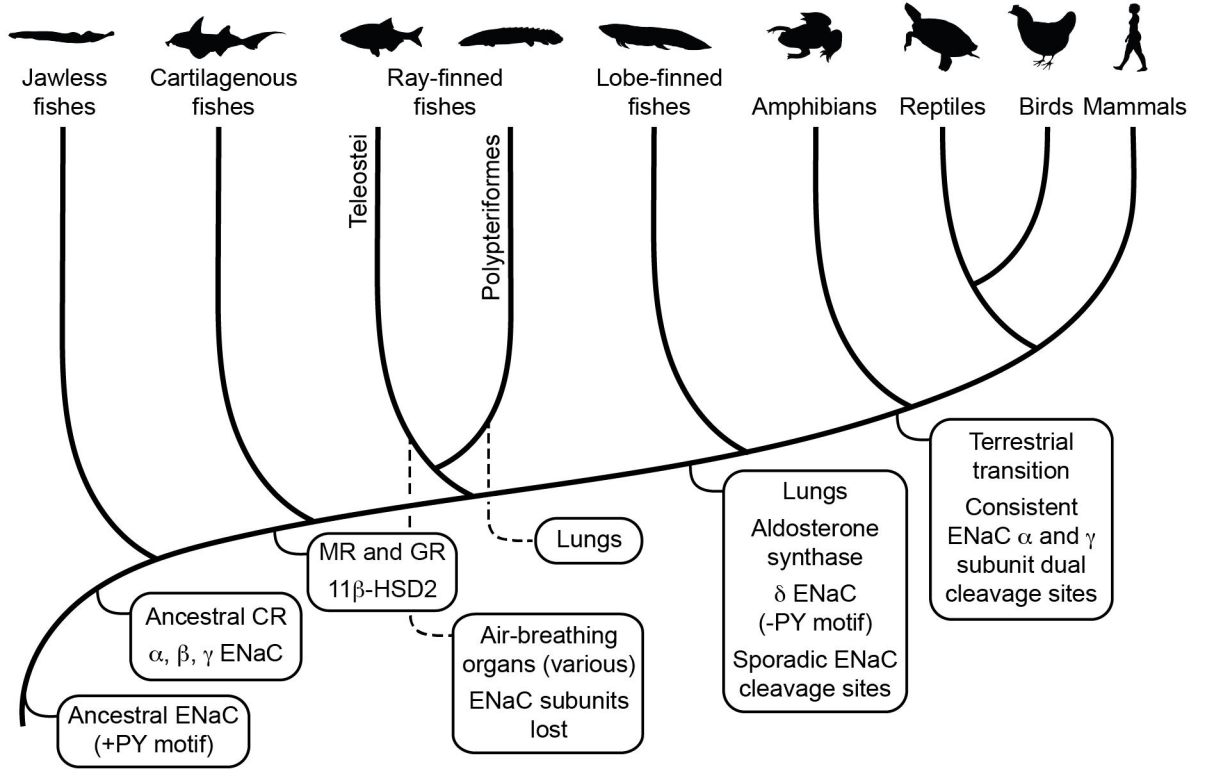
Jawless fishes
Cartilagenous fishes
Ray-finned fishes
Lobe-finned fishes
Amphibians
Reptiles
Birds
Mammals
Teleostei
Polypteriformes
Terrestrial transition
Consistent ENaC α and γ subunit dual cleavage sites
Lungs
Aldosterone synthase
δ ENaC (-PY motif)
Sporadic ENaC cleavage sites
Lungs
MR and GR
11β-HSD2
Air-breathing organs (various)
ENaC subunits lost
Ancestral CR
α, β, γ ENaC
Ancestral ENaC (+PY motif)