An amplified derepression controller with multisite inhibition and positive feedback

Gorana Drobac⁽²⁾, Qaiser Waheed⁽²⁾, Behzad Heidari, Peter Ruoff^{*}

Department of Chemistry, Bioscience, and Environmental Engineering, University of Stavanger, Stavanger, Norway

These authors contributed equally to this work. *peter.ruoff@uis.no

Abstract

How organisms are able to maintain robust homeostasis has in recent years received increased attention by the use of combined control engineering and kinetic concepts, which led to the discovery of robust controller motifs. While these motifs employ kinetic conditions showing integral feedback and homeostasis for step-wise perturbations, the motifs' performance differ significantly when exposing them to time dependent perturbations. One type of controller motifs which are able to handle exponentially and even hyperbolically growing perturbations are based on derepression. In these controllers the compensatory reaction, which neutralizes the perturbation, is derepressed, i.e. its reaction rate is increased by the decrease of an inhibitor acting on the compensatory flux. While controllers in this category can deal well with different time-dependent perturbations they have the disadvantage that they break down once the concentration of the regulatory inhibitor becomes too low and the compensatory flux has gained its maximum value. We wondered whether it would be possible to bypass this restriction, while still keeping the advantages of derepression kinetics. In this paper we show how the inclusion of multisite inhibition and the presence of positive feedback loops lead to an amplified controller which is still based on derepression kinetics but without showing the breakdown due to low inhibitor concentrations. By searching for the amplified feedback motif in natural systems, we found it as a part of the plant circadian clock where it is highly interlocked with other feedback loops.

Introduction

The concept of homeostasis [1], defined by Cannon in 1929 [2], is fundamental to our understanding how organisms, including our body, work [3]. According to Cannon homeostasis refers to the automatic, self-regulating processes that keep steady states within certain, but narrow limits, despite internal or environmental perturbations [1–3]. Although (negative) feedback is recognized as a central part in homeostatic regulation [1,4,5], it is not the only dynamic component. The 'homeostatic response' may also include features such as anticipatory mechanisms [6], feedforward loops [7], or positive feedbacks [3,8,9].

In control engineering robust regulation of a variable A with set-point A_{set} can be achieved by so-called integral control [10], which is able to (precisely) correct for step-wise perturbations acting on a controlled variable A [10] (Fig 1).

10

11



Fig 1. Scheme of a negative feedback with integral control regulating variable A. The difference between the set-point A_{set} and the actual value of A (the error ϵ) is integrated over time and fed back into the process which generates A. This procedure ensures that A will precisely reach A_{set} for step-wise perturbations. Colors correspond to molecular reactions shown later.

While in engineering integral control began to be applied in the beginning of the twentieth century with the power steering of ships [11], its usage in physiology/biology first appeared once cybernetics [12–14] made the analogies between engineered and biological systems more explicit. Physiological models during this first era showed the dynamical processes, loops, integrated errors, etc. mostly in terms of flow diagrams, transfer functions, as engineered systems are described [15,16]. By the turn of the century, when the molecular biology behind physiological processes became better understood researchers began to describe the control process in terms of their reaction kinetics, such as in integral rein control [17] (focussing on that physiological controllers come in antagonistic pairs; see also the later analogous notion of inflow/outflow controllers [18]), in the integral feedback formulation of robust bacterial chemotaxis [19,20], or in the integral control approach of blood calcium homeostasis [21].

It became evident that certain kinetic conditions within a negative feedback loop, such as zero-order kinetics [18–20, 22–24], autocatalysis [25–27], or second-order (bimolecular/antithetic) kinetics [28, 29] can lead to robust adaptation [30, 31] by integral control where an intrinsic integration of the error between set-point and the actual value of the controlled variable is automatically performed. When these feedback motifs were investigated towards time-dependent perturbations, it turned out that controller performances can differ significantly, either due to the structure of the feedback loop or due to the kinetics of how the integral controller is implemented [32, 33].

Feedback structures which have been found to perform well when exposed to different time-dependent perturbations are based on derepression kinetics [32].



Fig 2. Inflow controller based on derepression kinetics (motif 2, Ref. [18]). Red color: reactions determining the set-point; orange: perturbation; blue: *A*-signaling; green: *E*-signaling.

Fig 2 shows one of the motifs (motif 2) with derepression kinetics acting as an inflow35controller [18]. Reactions are color-coded and relate to the scheme in Fig 1. This motif,36unlike those not using derepression, is able to adapt perfectly to exponentially37increasing perturbations. Motif 2 can even balance hyperbolically increasing38

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

disturbances with doubling times which decrease exponentially [32]. In Fig 2 k_1 represents a (time-dependent or constant) perturbation and k_2 is the maximum fully derepressed compensatory flux. E is subject to an enzymatic zero-order degradation described by the rate parameters k_3 (V_{max}) and K_M , while k_4 represents a zero-order synthesis rate with respect to E. K_I is an inhibition constant.

The rate equations of the motif 2 controller are:

$$\dot{A} = \frac{k_2}{1 + \frac{E}{K_L}} - k_1 \cdot A \tag{1}$$

39

41

42

43

44

45

46

47

48

49

50

51

52

53

54

57

58

59

$$\dot{E} = k_4 \cdot A - \frac{k_3 \cdot E}{K_M + E} \tag{2}$$

where $k_2/(1 + (E/K_I))$ describes the compensatory flux, which opposes the perturbing flux k_1A . For step-wise changes in k_1 and for low K_M values $(K_M \ll E), E/(K_M + E) \approx 1$ and the steady state value in A is described by the set-point (setting E=0 and solving for A_{ss})

$$A_{ss} = A_{set} = \frac{k_3}{k_4} \tag{3}$$

which will also be defended against time-dependent (increasing) k_1 values (see later).

Metaphorically speaking, the activation of the compensatory flux by derepression is somewhat like the static takeoff of an airliner, when the plane stands still at the beginning of the runway, but having engines in full thrust with the breaks on. When the brakes are released the plane starts to accelerate and rapidly reaches takeoff speed.

The controller in Fig 2 reaches its maximum compensatory flux when $E \leq K_I$; any further increase in the perturbation k_1 cannot be opposed and will lead to the controller's breakdown. This is illustrated in Fig 3 for different K_I values with k_1 increasing exponentially. The k_1 value at breakdown, k_1^{bd} , can be estimated by setting $E=K_I$ and solving for k_1 from Eq 1 with $\dot{A}=0$, i.e.,

$$k_1^{\rm bd} \approx \frac{k_2}{2A_{set}} \tag{4}$$



Fig 3. Behavior of the motif 2 zero-order controller (Fig 2) upon an exponential increase of k_1 and the influence of K_I . Left panel shows k_1 as a function of time. Phase 1: $k_1=k_{1,p1}=2.0$ and constant. The controller is at steady-state at its set-point $A_{set}=5.0$. Phase 2: k_1 starts to increase exponentially at time $t_{p1}=10.0$ according to the inset Equation. Right panel shows the controller's A and E values for different K_I 's, where A_1 is A when $K_I=10.0$, A_2 is A when $K_I=1.0$, A_3 is A when $K_I=0.1$, and A_4 is A when $K_I=1\times10^{-3}$. The other rate constants are: k_2 (max compensatory rate)= 1×10^5 , $k_3=5\times10^3$, $k_4=1\times10^3$, $K_M=1\times10^{-6}$. Initial concentrations: A_1 - A_4 =5.0, $E_1=1\times10^5$, $E_2=1\times10^4$, $E_3=1\times10^3$, and $E_4=10.0$. Controller starts to break down when k_1 is reaching 1×10^4 indicated by the red arrows in the left panel.

The curves A_i and E_i in the right panel show the different A and E values when K_I takes the values 10.0 (A_1) , 1.0 (A_2) , 0.1 (A_3) , and 1×10^{-3} (A_4) . While the controller's "lifetime" (the time span until the controller breaks down) is practically not affected by the different K_I values, the controller's "aggressiveness", i.e. its ability to rapidly respond to perturbations and to keep A at A_{set} , is improved with decreasing K_I values.

Goal of this work

While motif 2 and related controllers based on derepression can keep the controlled variable A at its set-point even for rapidly increasing time-dependent perturbations [32, 33] (Fig 3), they suffer from breakdown once the controller variable E becomes close to or lower than K_I . We wondered whether it would be possible to circumvent this restriction to a controller where the control species' concentration increases with increasing perturbation strength, while still keeping the controller properties based on derepression. Using the motif 2 controller as an example, we show that implementation of a positive feedback loop based on autocatalysis combined with multisite inhibition kinetics can avoid controller breakdown by low E values, but still shows the properties of a controller based on derepression.

Materials and methods

Computations were carried out by using the Fortran subroutine LSODE [34]. Plots were generated with gnuplot (www.gnuplot.info) and slightly edited with Adobe Illustrator (adobe.com). To make notations simpler, concentrations of compounds are denoted by compound names without square brackets. Time derivatives are generally indicated by the 'dot' notation. Concentrations and rate parameter values are given in arbitrary units (au). In the Supporting Information a set of MATLAB (mathworks.com) scripts are provided for illustration in comparison with corresponding Fortran calculations (S1 Matlab).

Results and Discussion

Effect of multisite inhibition on controller performance

In this section we investigate the effect of multisite inhibition. In mechanistic terms, we consider an enzyme or transporter, which is responsible for the compensatory flux [33]. In multisite inhibition E can bind to the enzyme or transporter at multiple sites with different binding constants K_I . To make things more straightforward, we assume that one, two, or four molecules of E can bind to the enzyme/transporter, but always with the same binding constant K_I . In this case, Eq 1 is replaced by (see for example Ref [35])

$$\dot{A} = \frac{k_2}{1 + (\frac{E}{K_I})^n} - k_1 \cdot A \tag{5}$$

where n is the number of inhibiting E molecules (n=1, 2, or 4). Taking Fig 3 with $K_I=0.1$ as a starting point, Fig 4 shows the results when n is changed from 1 to 2 and to 4.

65

66

67

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

94



Fig 4. Behavior of the motif 2 zero-order controller (Fig 2) upon an exponential increase of k_1 (left panel Fig 3 and the influence of multisite inhibition (Eq 5)). Phase 1: the controller is at its set-point/steady state with constant $k_1=2.0$. Phase 2: Response of the controller with the different n values upon the exponential increase of k_1 . Rate constants are: $K_I=0.1$, k_2 (max compensatory rate)=1×10⁵, $k_3=5\times10^3$, $k_4=1\times10^3$, $K_M=1\times10^{-6}$. Initial concentrations: A=5.0 (for all n), E (n=2) =9.9, E (n=4)=0.9.

From Fig 4 it is clearly seen that multisite inhibition improves the controller's performance, i.e., makes the controller more aggressive by showing a more rapid response and by keeping A closer to the controller's set-point. However, despite the better responsiveness of the controller when using multisite inhibition, the "lifetime" of the controller is not improved, i.e., the breakdown occurs at the same time/ k_1^{bd} value as in Fig 3.

Increasing controller lifetime by increasing the maximum compensatory flux

Eq 4 indicates that increasing the maximum compensatory flux k_2 will increase k_1^{bd} and thereby increase the lifetime of the controller (upon increasing values of k_1). To automate this we have added a new variable C, shown in Fig 5, which is activated by derepression from E, but with the effect to increase k_2 .



Fig 5. Increasing controller lifetime by an *E*-dependent increase of the maximum compensatory flux k_2 .

For the sake of simplicity the inhibition constant of the derepression of C by E is assumed to be the same (K_I) as for the *E*-induced derepression of the compensatory

97

98

99

100

101

102

103

104

105

106

107

flux. The rate equations for this controller are

5.2

$$\dot{A} = \frac{k_2 C}{1 + (\frac{E}{K_I})^n} - k_1 \cdot A \tag{6}$$

$$\dot{E} = k_4 \cdot A - \frac{k_3 \cdot E}{K_M + E} \tag{2}$$

$$\dot{C} = \frac{k_5}{1 + \frac{E}{K_I}} - k_6 \cdot C \tag{7}$$

Note that the rate equation for E remains unaltered. We also keep the multisite 112 inhibition of the compensatory flux by E, but consider only a single E-binding site for 113 the inhibition of the zero-order generation (k_5) of C. Fig 6 shows the lifetime of 114 controllers with n = 4 and with different k_5 and k_6 values when k_1 increases 115 exponentially as indicated in the left panel of Fig 3. As a reference, the blue solid and 116 dashed lines show respectively the A_3 and E_3 values from the results of Fig 4 with 117 n = 4, i.e., in the absence of C. For high k_5/k_6 ratios the lifetime of the controller is 118 clearly increased (see traces A_1, E_1 and A_2, E_2), while for a ratio of one ($k_5 = k_6 = 0.1$, 119 traces A_4, E_4) the lifetime of the controller is slightly reduced. 120



1000

Fig 6. Activation of the compensatory flux for reaction scheme of Fig 5 and corresponding Eqs 6, 2, and 7 with n=4. Left panel: Concentrations of A and E as a function of time with different k_5/k_6 rate constants. Blue curves A_3 and E_3 correspond to the calculations without C (Fig 4 with n=4). Right panel: Concentrations of C with different k_5/k_6 rate constants as a function of time. Phase 1: the controller is at steady state at its set-point $A_{set}=5.0$ at constant $k_1=2.0$. Phase 2: k_1 increases exponentially according to the inset in the left panel of Fig 3. k_5 and k_6 values and initial concentrations for the different A_i , E_i , C_i curves: i=1, $k_5=10.0$, $k_6=0.01$, $A_0=5.0$, $E_0=2.412$, $C_0=39.81$; i=2, $k_5=1.0$, $k_6=0.1$, $A_0=5.0$, $E_0=0.9$, $C_0=1.0$; i=3, no C (Fig 4 with n=4); i=4, $k_5=k_6=0.1$, $A_0=5.0$, $E_0=0.531$, $C_0=0.158$. Other rate constant values: $k_2=1\times10^5$, $k_3=5\times10^3$, $k_4=1\times10^3$, $K_M=1\times10^{-6}$, $K_I=0.1$.

Opposing *E* decrease by positive feedback

While the inclusion of C increases the compensatory flux $k_2C/(1+(E/K_I)^n)$ and leads to an improvement in the controller's lifetime, the controller still suffers from the general limitation that once E is driven down to values approaching K_I , i.e., the controller breaks down when (setting $E=K_I$)

$$k_1^{\rm bd} \approx \frac{k_2 C}{2A_{set}} \tag{8}$$



111

121

123

124

(Fig 6). We found, that this trend can be circumvented by including a positive feedback 126 in the generation of C without loosing the dynamic properties of the motif 2 controller. 127 The positive feedback in C can be generated by first-order or second-order autocatalysis. 128 Fig 7 shows the scheme for both first-order and second-order autocatalysis 129



Fig 7. Inclusion of autocatalysis in the generation of C.

In case of first-order autocatalysis Eq 7 is changed to

$$\dot{C} = \frac{k_5 C}{1 + \frac{E}{K_I}} - k_6 \cdot C \tag{9}$$

while for the second-order case both the synthesis and degradation terms are second-order with respect to C

$$\dot{C} = \frac{k_5 C^2}{1 + \frac{E}{K_I}} - k_6 \cdot C^2 \tag{10}$$

Note that for step-wise changes in $k_1 E$ becomes homeostatic controlled in addition ¹³³ to A, because $\dot{C}=0$ in Eqs 9 or 10 implies that ¹³⁴

$$E_{ss} = \left(\frac{k_5}{k_6} - 1\right) K_I \tag{11}$$

independent of the perturbation k_1 .

Comparing the influence of first-order and second-order autocatalysis in C on controller performance

We have tested the influence of the first-order and second-order autocatalytic terms on 138 the controller performance for n=4. Considered were step-wise changes in k_1 , together 139 with linear, exponential, and hyperbolic increases of k_1 . For all these perturbation types 140 the controllers with both first-order and second-order autocatalysis show robust 141 homeostasis and defend their set-points $A_{set} = k_3/k_4$ successfully. It is interesting to 142 note that E no longer decreases, but approaches a steady state during the 143 time-dependent increase of k_1 ! We show here the results for exponentially and 144 hyperbolically increasing k_1 values. The controller's behavior for step-wise and linear 145 changes are described in S1 Text. 146

130

131

132

135

136

First-order autocatalysis in C

The controller is described by Eqs 2, 6, and 9. The perturbation k_1 increases exponentially as described in the left panel of Fig 3.

Fig 8, left panel, shows that C follows the exponential increase in k_1 closely, while the right panel shows that E goes into a steady state with A kept at its set-point $A_{set}=k_3/k_4=5.0$. The steady state value in E can be calculated from Eq 9 by noting that this equation can be written as

$$\frac{1}{C} \cdot \frac{\mathrm{d}C}{\mathrm{d}t} = \frac{\mathrm{d}\ln C}{\mathrm{d}t} = \frac{k_5}{1 + \frac{E}{K_L}} - k_6 \tag{12}$$

Inserting into Eq 12 the value of $d \ln C/dt$ (which is equal to $k_1/k_1=0.2$ and using L'Hôpital's rule) together with the values of the other rate constants and solving for E, gives

$$E_{ss} = \left(\frac{k_5}{\frac{d\ln C}{dt} + k_6} - 1\right) K_I = 0.7333 \tag{13}$$

in agreement with the numerical calculation.

1



Fig 8. Controller performance with first-order autocatalysis in C (Eq. 9) and exponential increase of k_1 . Phase 1: the controller is at steady state at its set-point $A_{set}=5.0$ with constant $k_1=2.0$. Initial concentrations: $A_0=5.0$, $E_0=0.9$, $C_0=1.0$. Phase 2: k_1 increases exponentially (left panel of Fig 3), $k_2=1 \times 10^5$, $k_3=5 \times 10^2$, $k_4=1 \times 10^2$, $k_5=10.0$, and $k_6=1.0$. $K_M=1 \times 10^{-6}$, $K_I=0.1$, n=4 (Eq 6). Left panel: k_1 and C as a function of time; right panel: A and E as a function of time.

One of the characteristic properties of the motif 2 derepression controller is its capability to tackle rapidly increasing perturbations, like hyperbolic growth [32]. While exponential growth in k_1 has a constant doubling time the doubling time in hyperbolic growth decreases exponentially and k_1 will (formally) reach infinity at a certain time point. We wondered whether the controller based on rate equations 2, 6, and 9 would still show this property. 159

In Eq 14 k_1 increases hyperbolically according to

1

$$k_1 = \frac{40.5}{\frac{40.5}{k_{1,p_1}} - (t - t_{p_1})} \tag{14}$$

where $k_{1,p1}$ is the constant value of k_1 during phase 1 ($k_1=2.0$ as in the previous calculations), while t_{p1} is the duration of phase 1 (here 1 time unit). When time treaches 21.25 (the infinity limit) the value of k_1 goes formally to infinity. Fig 9 shows the behavior of the controller close to the infinity limit. In the calculation t and k_1 reach 21.249997 and 1.4×10^7 , respectively. During the last 0.25 time units k_1 increases

164

154

155

156

157

147

148

149

150

151

152

> by approximately 5-orders of magnitude. Despite this rapid increase in k_1 the controller is able to maintain homeostasis, but finally breaks down before the infinity limit is reached.

It is interesting to note that in case of hyperbolic growth the first-order autocatalytic growth in C (left panel, Fig 9) is not able to maintain a constant E, i.e., E decreases, but A is still kept at its set-point (right panel, Fig 9).



Fig 9. Controller performance with first-order autocatalysis in C (Eq. 9) and hyperbolic increase of k_1 . Phase 1 (not shown): the controller is at steady state at its set-point $A_{set}=5.0$ with constant $k_1=2.0$. Phase 1 lasts 1 time unit. Initial concentrations: $A_0=5.0$, $E_0=0.9$, $C_0=1.0$. Phase 2: k_1 increases in a hyperbolic fashion (Eq 14), $k_2=1 \times 10^5$, $k_3=5 \times 10^5$, $k_4=1 \times 10^5$, $k_5=10.0$, and $k_6=1.0$. $K_M=1 \times 10^{-6}$, $K_I=0.1$, n = 4 (Eq 6). Left panel: k_1 and C as a function of time just before k_1 reaches the infinity limit; right panel: corresponding A and E concentrations as a function of time.

Second-order autocatalysis in C

We asked the question, how would the controller respond when the autocatalytic 177 generation of C becomes second-order (Eq 10), with other words, when the generation of C is itself due to hyperbolic growth (S2 Text)? 178



Fig 10. Controller performance with second-order autocatalysis in C (Eq. 10) and exponential increase of k_1 . Phase 1: the controller is at steady state at its set-point $A_{set}=5.0$ with constant $k_1=2.0$. Initial concentrations: $A_0=5.0$, $E_0=0.9$, $C_0=1.0$. Phase 2: k_1 increases exponentially (left panel of Fig 3), $k_2=1 \times 10^5$, $k_3=5 \times 10^2$, $k_4=1 \times 10^2$, $k_5=10.0$, and $k_6=1.0$. $K_M=1 \times 10^{-6}$, $K_I=0.1$, n=4 (Eq 6). Left panel: k_1 and C as a function of time; right panel: corresponding A and E concentrations as a function of time.

173

174

175

> Fig 10 shows the results when C is subject to second-order autocatalysis and k_1 100 increases exponentially. Although there is no apparent change in A in comparison with Fig 8 now E itself is under homeostatic control, besides A. E's set-point can be calculated by setting Eq 10 to zero, which leads to 103

$$E_{set} = K_I \left(\frac{k_5}{k_6} - 1\right) \tag{15}$$

Inserting the rate constant values from Fig 10 into Eq 15 gives a value for E_{set} of 0.9 in agreement with the numerical values for E_{ss} .



Fig 11. Controller performance with second-order autocatalysis in C (Eq. 10) and hyperbolic increase of k_1 (Eq 14). Phase 1 (not shown): the controller is at steady state at its set-point A_{set} =5.0 with constant k_1 =2.0. Phase 1 lasts 1 time unit. Initial concentrations: A_0 =5.0, E_0 =0.9, C_0 =1.0. Phase 2: k_1 increases hyperbolically. Rate constant values: k_2 =1 × 10⁵, k_3 =5 × 10², k_4 =1 × 10², k_5 =10.0, and k_6 =1.0. K_M =1 × 10⁻⁶, K_I =0.1, n = 4 (Eq 6). Left panel: k_1 and C as a function of time just before k_1 reaches the infinity limit (blue dashed line). At time 21.249997 k_1 =1.4 × 10⁷, C=5.4 × 10⁶. Right panel: A and E concentrations as a function of time.

Fig 11 shows the controller's behavior when k_1 increases hyperbolically with the same second-order autocatalysis as in Fig 10. However, while the controller is still able to keep A at its theoretical set-point, the value of E_{ss} shows now an offset below E_{set} (Eq 15).

In the case when k_1 increases hyperbolically, we wondered how well motif 2 would perform without the help of C in comparison with controllers that have first- or second-order autocatalysis in C?

189 190 191



Fig 12. Behavior of motif 2 controller (Fig 2) in comparison with a controller including first-order and second-order autocatalysis in C (Fig 7). Rate constants for all three controllers: $k_2=1 \times 10^5$, $k_3=5 \times 10^5$, $k_4=1 \times 10^5$, $K_M=1 \times 10^{-6}$, $K_I=0.1$, n=4. Additional rate constants for the autocatalytic controllers: $k_5=10.0$, and $k_6=1.0$. Initial concentrations: $A_0=5.0$, $E_0=0.9$, $C_0=1.0$ (when autocatalysis in C is included).

An answer to this question is given in Fig 12, which shows the performance of a single motif 2 controller without C in comparison with controllers having first- and second-order autocatalysis in C. Since the motif 2 controller without C is subject to the limitation described by Eq 4, this controller breaks down earlier in comparison with those controllers having autocatalysis in C. The second-order autocatalytic controller performs best and keeps A longest at its set-point before breakdown occurs near the infinity limit.

Oscillatory homeostats

This section is inspired by the fact that in physiology many cellular compounds or tissues show oscillatory behaviors [36–40], but are also under a homeostatic regulation. For example, circadian oscillations regulate hormones, blood glucose, and adapt organisms to the light/dark and seasonal changes on earth [41–46]. Another interesting example is the homeostatic stabilization of complex neural oscillations [47].

The motif 2 controller (Fig 2) becomes oscillatory when degradations with respect to A and E become zero-order. The resulting oscillator can maintain robust homeostasis [48] of $\langle A \rangle$, where

$$\langle A \rangle = \frac{1}{\tau} \int_0^\tau A(t) \, \mathrm{d}t \to A_{set}$$
 (16)

and integration is taken over a certain time interval τ .

This oscillator is identical to Goodwin's 1963 oscillator [49–51], although Goodwin 210 was probably not aware of the oscillator's homeostatic property. The promotion of 211 oscillations by zero-order kinetics have also been recognized by Kurosawa and Iwasa [52] 212 in an alternative version of Goodwin's equations [53]. 213

In case of motif 2, the oscillatory reaction scheme is shown in Fig 13

209

214

193

194

195

196

197

198

199

200

201

202

203

204



Fig 13. Motif 2 becomes an oscillatory homeostat when $K_M \ll E$ (condition for integral control) and, in addition, $K_M \ll A$. The resulting oscillator is identical to Goodwin's equations/oscillator from 1963 [49].

with the altered rate equation in A:

$$\dot{A} = \frac{k_2}{1 + (\frac{E}{K_I})^n} - \frac{k_1 \cdot A}{K_M + A} \tag{17}$$

When $K_M \ll A$ and n=1, Eqs 2 and 17 can be combined into a quasi-harmonic form [48] ²¹⁶

$$\frac{\ddot{A}}{\frac{k_2k_4K_I}{(K_I+E)^2}} + A = A_{set} = \frac{k_3}{k_4}$$
(18)

When the values of k_1 and k_2 are close to each other, the resulting oscillations are practically harmonic (sinusoidal) and the period P of the oscillator can be estimated as 217

$$P_{harm} = \frac{2\pi}{\sqrt{\frac{k_2 k_4 K_I}{(K_I + \langle E \rangle)^2}}}$$
(19)

with

$$\langle E \rangle = \frac{1}{\tau} \int_0^\tau E(t) \,\mathrm{d}t \tag{20}$$

Table 1 shows three examples of numerically calculated periods P_{num} in comparison with the corresponding harmonic periods P_{harm} . It may be noted that when k_1 and k_2 become significantly different, the resulting oscillations become highly nonlinear and P_{harm} can only qualitatively indicate the period's dependence on rate constants and $\langle E \rangle$.

Table 1. Harmonic and numerical periods.

k_1	k_2	P_{harm}	P_{num}			
9×10^{-3}	1×10^{-2}	22.0	22.1			
9×10^{-2}	0.1	7.01	6.99			
0.9	1.0	2.21	2.22			
$K_I = 0.1, K_M = 1 \times 10^{-8}, k_2 = k_4 = 1.0,$						
$A_0 = 0.96, E_0 = 3.3 \times 10^{-3}.$						

Fig 14 shows, as a reference, the behavior of the oscillatory m2 controller (Fig 13) 225 when k_1 increases exponentially. The value of $\langle A \rangle$ is at the controller's set-point (5.0) 226 and kept there, until, as in the non-oscillatory case (Fig 3), the controller breaks down 227 when E values become too low. 228

219



Fig 14. Response of oscillatory motif 2 (Fig 13) when k_1 increases exponentially in phase 2 by the growth law described in the left panel of Fig 3. Phase 1: k_1 is at 2.0; phase 2: At time t=80 k_1 starts to increase exponentially. At the end of phase 2 k_1 is 1.8×10^6 . Left panel: values of A, $\langle A \rangle$, and E as a function of time. Right panel: Calculated period as a function of time. Rate constants: $k_2=1.0 \times 10^5$, $k_3=5.0$, $k_4=1.0$, $K_I=1.0$, $K_M=1.0 \times 10^{-6}$, n=4. Initial concentrations: $A_0=1.56$, $E_0=20.55$. The rate equations are given by Eqs 2 and 17.

In the case when C is included to improve controller performance (Fig 15), the resulting controller shows an increased lifetime. This is shown in Fig 16.



Fig 15. Inclusion of C in the oscillatory m2 controller.

The presence of C had no significant effect on the period (Fig 16, right panel) which decreased practically in the same manner as in Fig 14 when C is absent. Despite the controller's increased lifetime in the presence of C, the controller will also in this case, due to the decrease in E, eventually break down as in the nonoscillatory case (Fig 6). 232 233 234 235 235 236 236 237 237 238

229



Fig 16. Response of oscillatory motif 2 containing C (Fig 15) when k_1 increases exponentially in phase 2 by the growth law described in the left panel of Fig 3. Rate equations for A, E, and C are given by Eqs 17, 2, and 7, respectively. Phase 1: k_1 is kept constant at 2.0; phase 2: At time t=80 k_1 starts to increase exponentially. At the end of phase 2 k_1 is 1.8×10^6 . Controller breakdown occurs just after 160 time units (data not shown). Left panel: values of A, $\langle A \rangle$, and E as a function of time. Right panel: Calculated period as a function of time. Rate constants: $k_2=1.0 \times 10^5$, $k_3=5.0$, $k_4=1.0$, $k_5=50.0$, $k_6=1.0$, $K_I=1.0$, $K_M=1.0 \times 10^{-6}$, n=4. Initial concentrations: $A_0=1.56$, $E_0=20.55$, $C_0=1.0$.

When C is generated by first-order autocatalysis (described by Eq 9) the controller is able to defend A_{set} for an extended time period (Fig 17a-c) and keeps $\langle E \rangle$ constant (Fig 17d). After an induction period, the controller is able to follow the exponentially increasing k_1 by C (Fig 17e) and thereby becoming capable to defend A_{set} . Fig 17f shows the exponential decrease of the calculated period. Since in phase $2 \langle E \rangle$ is now kept at a constant level the controller will remain operative as long as C can be increased and the activation of A by C is maintained.

When C is generated by second-order autocatalysis (Eq 10) the resulting controller is, as for first-order autocatalysis, able to defend A_{set} . Fig 18 shows the case when k_1 increases exponentially. We found that an increase of k_2 by one order of magnitude during phase 2 was beneficial for the controller's homeostatic behavior. To avoid overcompensation, k_5 was decreased by one order of magnitude during phase 2. Fig 18a shows the time profiles of A and $\langle A \rangle$. Once the controller is able to follow the increasing k_1 values (indicated by the downward arrows in the different panels) $\langle A \rangle$ is at A_{set} , as indicated in panel a by the white outlined $\langle A \rangle$.

In Fig 18b E is shown as a function of time going into a steady state once control over the increasing k_1 values have been taken. The takeover of control is most clearly seen in Fig 18c when after the induction period C is able to follow the increasing k_1 . Fig 18d shows that the period is decreasing exponentially in line with the exponential increase of k_1 . Fig 18e shows the high frequency oscillations in A near the end of phase 2 having a period of approximately 10^{-4} time units. Calculating the A-average over the 10^{-3} time units shows that $\langle A \rangle$ is at $A_{set}=5.0$.

Finally we have tested a controller with second-order autocatalytic generation of C257 when k_1 increases hyperbolically according to Eq 14. We found that a reduction of the 258 unperturbed period in phase 1 to approximately 2 time units gave a good illustration of 259 an operational controller under these conditions. The reduced period was achieved by 260 increasing k_4 to 50.0. This led to a decrease in A_{set} ($=k_3/k_4$) to 0.1. Fig 19a shows a 261 semilogarithmic plot of A and $\langle A \rangle$ as a function of time. At time t=50.0 k_1 starts to 262 grow (Eq 14) with $k_{p,1}=2.0$. The infinity limit is reached at 70.25 time units. Also here 263 we observe an induction period in which the controller adapts to the increasing k_1 . At 264 about 60 time units the controller is able to oppose the increasing k_1 values. At the 265 same time $\langle E \rangle$ goes into a steady state (Fig 19b) and C is able to follow k_1 (Fig 19c). 266

235

236

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

The period decreases in a corresponding manner as k_1 increases (Fig 19d), and the controller is able to defend A_{set} . Fig 19e shows that $\langle A \rangle = 0.0997$ when $\langle A \rangle$ is determined between 60 time units and close to the infinity limit.



Fig 17. Response of oscillatory m2 controller with first-order autocatalytic generation of C and exponential increase of k_1 . Phase 1 (0-80 time units): $k_1 = 2.0$. Phase 2 (80-180 time units): k_1 increases exponentially as shown in the left panel of Fig 3. Rate equations are given by Eqs 17, 2, and 9. (a) A (in purple) and overall $\langle A \rangle$ (in green) as a function of time. The white outlined $\langle A \rangle$ is the $\langle A \rangle$ value calculated from 120 (white vertical line) to 180 time units showing that $\langle A \rangle = A_{set} = 5.0$. (b) A, $\langle A \rangle$, and E for the time interval 179.0-180.0. (c) A, $\langle A \rangle$, and E for the time interval 179.9-180.0. (d) E as a function of time. (e) k_1 and C as a function of time. (f) The period as a function of time. Rate constants: $k_2=1\times10^5$, $k_3=5.0$, $k_4=1.0$, $k_5=5.0$ (phase 1), $k_5=50.0$ (phase 2), $k_6=0.1$ (phase 1), $k_6=1.0$ (phase 2), $K_I=1.0$, $K_M=$ 1×10^{-6} . Initial concentrations: $A_0=2.684$, $E_0=61.55$, $C_0=86.21$, n=4.

267

268



Fig 18. Response of oscillatory m2 controller with second-order autocatalytic generation of C and exponential increase of k_1 . Phase 1 (0-40 time units): k_1 =2.0. Phase 2 (40-160 time units): k_1 increases exponentially as shown in the left panel of Fig 3. Rate equations are given by Eqs 17, 2, and 10. Downward arrows indicate the starting point when the controller is able to compensate for the increasing k_1 values. (a) A (in purple) and overall $\langle A \rangle$ (in green) as a function of time. The white outlined $\langle A \rangle$ is the $\langle A \rangle$ value calculated from 140 (white vertical line) to 160 time units showing that $\langle A \rangle = A_{set} = 5.0$. (b) E as a function of time. (c) k_1 and C as a function of time. (d) The period as a function of time. (e) High frequency oscillations near the end of phase 2. $\langle A \rangle$ is close to $A_{set} = 5.0$. Rate constants: $k_2 = 1 \times 10^5$ (phase 1), $k_2 = 1 \times 10^6$ (phase 2), $k_3 = 5.0$, $k_4 = 1.0$, $k_5 = 5 \times 10^{-2}$ (phase 1), $k_5 = 1 \times 10^{-3}$ (phase 2), $k_6 = 1 \times 10^{-3}$, $K_I = 1.0$, $K_M = 1 \times 10^{-6}$. Initial concentrations: $A_0 = 9.6$, $E_0 = 55.4$, $C_0 = 101.0$, n = 4.



Fig 19. Response of oscillatory m2 controller with second-order autocatalytic generation of C and hyperbolic increase of k_1 . Rate equations are given, as in Fig 18, by Eqs 17, 2, and 10. Phase 1 (0-50 time units): $k_1 = 2.0$. Phase 2 (50-70.24999992 time units) k_1 increases hyperbolically (Eq 14) from 2.0 to 5.8673×10^8 . $A_{set}=0.1$. (a) A and average $\langle A \rangle$ as a function of time. $A_{set}=0.1$. (b) Concentration of E as a function of time. (c) k_1 and C time profiles. (d) The period as a function of time. (e) Concentration of A and calculated $\langle A \rangle$ (=0.0997) during phase 2 (time interval 60.0-70.24999992) when oscillations are present. Rate constants: $k_2=1\times10^5$ (phase 1), $k_2=1\times10^6$ (phase 2), $k_3=5.0$, $k_4=50.0$, $k_5=5\times10^{-2}$ (phase 1), $k_5=1\times10^3$ (phase 2), $k_6=1\times10^{-3}$ (phase 1), $k_6=1.57\times10^2$ (phase 2), $K_I=1.0$, $K_M=1\times10^{-6}$. Initial concentrations: $A_0=2.736\times10^{-4}$, $E_0=4.793\times10^1$, $C_0=1.489\times10^2$, n=4.

Period homeostasis with respect to step-wise perturbations in k_1 270

Since in some oscillatory physiologies, like circadian rhythms, period homeostasis is observed with respect to certain step-wise environmental perturbations, for example in temperature or pH [54, 55], we wondered whether it would be possible to include an additional variable to one of the above oscillatory controllers which would give homeostasis not only in <A> but also in the oscillator's period. We have previously shown [48] how the basic oscillatory m2 motif (Fig 13) can show period homeostasis by the addition of controller variables that keep E and the chemical fluxes through E at a constant level. Here we show that we can do the same by taking, as an example, the controller described in Fig 15 (including autocatalytic formation of C). 278



Fig 20. Reaction scheme of oscillatory m2 controller with autocatalytic generation of C and period homeostasis with respect to step-wise perturbations in k_1 .

Fig 20 shows the controller's reaction scheme with the additional variable I_1 , which keeps $\langle C \rangle$ and thereby $\langle E \rangle$ under homeostatic control. The rate equations are: 280

$$\dot{A} = \frac{k_2 C}{1 + (\frac{E}{K_I})^n} - \frac{k_1 \cdot A}{K_M + A} + k_9 \cdot I_1$$
(21)

$$\dot{E} = k_4 \cdot A - \frac{k_3 \cdot E}{K_M + E} \tag{22}$$

$$\dot{C} = \frac{k_5 C}{1 + \frac{E}{K_I}} - k_6 \cdot C \tag{23}$$

$$\dot{I}_1 = k_7 \cdot C - \frac{k_8 \cdot I_1}{K_M + I_1} \tag{24}$$

 I_1 acts as an additional inflow controller with the property to keep $\langle C \rangle$ at a constant level. The two compensatory fluxes

$$j_2 = \frac{k_2 C}{1 + (\frac{E}{K_I})^n}$$
(25)

and

$$j_9 = k_9 \cdot I_1 \tag{26}$$

act together such that $\langle A \rangle$, $\langle E \rangle$, and $\langle C \rangle$ are under homeostatic control, which leads to regulated fluxes through A, E, and C such that the period of the oscillator becomes constant.

October 16, 2020

18/26

282

283

284

285

286

> Fig 21 illustrates the period homeostasis for step-wise changes of k_1 from 2.0 to 10.0. The period length during phase 1 (k_1 =2.0) is approximately 18 time units. Directly 289 after the step in k_1 the period drops to approximately 4 time units but then increases to 290 its original value due to the increase in I_1 (panels a, e, and f). The set-point of $\langle C \rangle$ 291 can be calculated from Eq 24 by setting $\langle I_1 \rangle = 0$, which leads to 292

$$\langle \dot{I}_1 \rangle = \langle k_7 \cdot C \rangle - \left\langle \frac{k_8 \cdot I_1}{K_M + I_1} \right\rangle = 0 \quad \Rightarrow \quad \langle C \rangle = \frac{k_8}{k_7} \tag{27}$$

by assuming that $\langle I_1/(K_M + I_1) \rangle = 1$, i.e., $K_M \ll I_1$.



Fig 21. Example of period homeostasis of the controller in Fig 20 when k_1 changes from 2.0 (phase 1) to 10 at time t=500. Panel a shows oscillatory A and the calculated average $\langle A \rangle$ as a function of time. (b) Concentration of E as a function of time (in blue) and $\langle E \rangle$ (orange lines). For phase $1 \langle E \rangle$ was calculated for the entire phase, i.e., for the time interval 0-500, while in phase $2 \langle E \rangle$ was calculated for the time interval 2000-2500. (c) Step-wise change of k_1 from 2 to 10 (left ordinate, outlined in black). Right ordinate: concentration of C as a function of time. Orange lines: calculated average $\langle C \rangle$ for the time intervals given in panel b. (d) Time plot (in purple) and average value (in green) of respectively $K_I/(K_I+E)$ and $\langle K_I/(K_I+E) \rangle$. The average values are calculated for the time intervals as stated for panel b. For visibility, ordinate values, which have maximum values of 1 are cut off at 0.02. (e) Calculated period length as a function of time showing that I_1 manages to keep period homeostatically controlled. (f) I_1 as a function of time. Rate constants: $k_2=1\times10^5$, $k_3=500, k_4=k_5=100, k_6=1.0, k_7=100, k_8=0.1, k_9=8\times10^{-2}, K_I=1.0, K_M=1\times10^{-6}$. Initial concentrations: $A_0=2.683, E_0=4.463\times10^3, C_0=4.108\times10^{-6}, I_{1,0}=2.705, n=1$.

Fig 21c shows that the calculated $\langle C \rangle$ value (=0.001) is in perfect agreement with Eq 27. Applying $\langle \dot{C} \rangle = 0$ in Eq 23 leads to the condition

$$\left\langle \frac{K_I}{K_I + E} \right\rangle = \frac{k_6}{k_5} = 0.01 \tag{28}$$

which is obeyed, as seen in Fig 21d. However, despite the fact that Eq 28 is fulfilled, we were not able to extract an analytical value for $\langle E \rangle$. To get $\langle E \rangle$ (Fig 21b) we used the numerical solution of the rate equations. 299

An interesting aspect is whether period homeostasis can also be obtained when the perturbation becomes time-dependent. We will deal with this situation more generally in another paper.

Controller breakdown due to saturation

We have shown that the breakdown of the simple derepression m2 controllers (Figs 2 and 13) can be delayed or even circumvented by including a component C which is activated by derepression from E, but itself activates the generation of E via A (Fig 5). However, there is the question how saturation may affect the performance of the controllers. For example, although the controllers containing a first-order autocatalysis in C are apparently able to follow exponentially increasing k_1 values (Figs 9 and 17) they eventually will break down since neither the perturbation k_1 nor the increase in Ccan continue ad infinitum. In this respect, the models presented here need to be considered as idealizations. For

For this respect, the models presented here need to be considered as idealizations. For example, concerning the growth of k_1 and C, k_1 and C will eventually approach saturation levels. In the case of k_1 , the removal of A may be due to an enzyme or a transporter, which eventually will go into saturation. Likewise, C will be generated by corresponding enzymatic processes and will be subject to saturation (see for example Ref [18] (Supporting Material) and Ref [56]).

A brief overview over these breakdown scenarios are now given. When k_1 , i.e. the removal of A, goes into saturation before the production of C and before the C-signaling to the compensatory flux, the controller will be able to keep homeostasis of A at its set-point, at a high but constant level of k_1 . When C-signaling goes into saturation before k_1 and C become saturated, then the controller will break down due to an unbalanced exponential increase of C. Finally, when C production becomes 223

294

295

296

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

saturated, but the removal of A by k_1 and C-signaling are still operative, then breakdown of the controller occurs, because the kinetics of the C production are not able to oppose the rapid decrease of A and A levels will go to zero.

Example: The TOC1/PRR5-RVE8 negative feedback loop

A question natural to ask, is whether the above "A-E-C" regulatory circuits can be 328 found in physiology. Would the properties be the same as in the isolated case, i.e., as 329 studied here? Since circadian rhythms are based on transcriptional-translational 330 negative feedback loops, the Goodwin oscillator and the m2-scheme has served as a 331 basic model to describe circadian oscillations [49-51, 57, 58]. We have taken the plant 332 circadian clock organization and looked for the A-E-C motif (Fig 5) there. In plants the 333 circadian organization is complex [59,60] and consists of several interlocked negative 334 feedback loops where each of them can approximately be described by a basic 335 m2-scheme (Fig 13). Plants have a morning oscillator based on the genes LHY (Late 336 Elongated Hypocotyl) and CCA1 (Circadian Clock Associated 1) and an evening 337 complex (EC) which also contains an autoregulatory negative feedback loop. In 338 addition, there appear to be transcriptional-translational negative feedbacks in the 339 organization of pseudo-response regulators (PRR's). The PRR gene family consists of 340 five paralogue genes (PRR1, 3, 5, 7, and PRR9). PRR1 (also known as Timing of CAB 341 expression 1 (TOC1)) is presently one of the best characterized gene in the PRR family. 342 They are implicated in clock function and act as period controlling factors [61]. TOC1 343 and PRR5 are interlocked with the morning oscillator components LHY and CCA1344 and the evening complex. It is in the TOC1/PRR5 feedback structure including the 345 RVE8 (REVEILLE8) gene we find the A-E-C motif. 346



Fig 22. Part of the plant circadian clock involving the *TOC*1 m2-type feedback loop. The A-E-C motif (Fig 5) is outlined in red.

Fig 22 shows part of the plant circadian network including TOC1/PRR5 and RVE8. The TOC1 and RVE8 genes, when mutated, affect the period of the plant circadian clock. In addition, RVE8 has also an influence on the circadian period with respect to temperature (temperature compensation) [62]. RVE8 interacts with LHY, CCA1, and the EC, which in their turn also have an influence on the plant circadian rhythm. The TOC1/PRR5-mRNA's are part of a negative feedback loop where the proteins TOC1

324

325

326

and PRR5 feed negatively back on their transcription. RVE8 is a MYB-like 353 transcription factor and activates the transcription of TOC1/PRR5, but needs NIGHT 354 LIGHT-INDUCIBLE AND CLOCK-REGULATED 1 and 2 (LNK1 and LNK2) [63,64] 355 to do that. TOC1/PRR5 on their side inhibit the production of RVE8 possibly by 356 transcriptional repression [65]. Our results with the A-E-C motif suggests, in agreement 357 with experimental implications [62], that RVE8 may take part in the homeostasis of the 358 TOC1/PRR5 negative feedback loop, to stabilize homeostatic properties of the plant 359 circadian clock, including the period. For example, overexpression of RVE8 leads to a 360 shorter circadian period. In analogy, increase of k_5 in Fig 5 leads also to a shorter 361 period. However, the TOC1/PRR5 circuits are highly interlocked with other clock 362 components. Thus, other roles of the TOC1/PRR5-RVE8 loop may emerge when 363 detailed models of the plant circadian clock are considered. 364 While we started to find an improvement of the m2-regulatory loop, we arrived at the 365 366

A-E-C motif. We feel that this or similar feedback structures may be found in other physiological regulations, but more investigations are needed in this respect.

Supporting information	368
S1 Matlab. Matlab programs. A zip-file with Matlab programs showing results from Fig 4 $(n=4)$, Fig 6 $(n=4, i=1)$, Fig 8, Fig 10, Fig 14,	369 370
S1 Text. Controller performance towards step-wise changes in k_1 and linearly increasing k_1 values. Comparison between controllers (Fig 7) having first-order and second-order autocatalytic generation of C .	371 372 373
S2 Text. Hyperbolic growth by higher-order autocatalysis. It is shown that autocatalysis with an order larger than one shows hyperbolic growth.	374 375
References	376
 Langley, LL, editor. Homeostasis. Origins of the Concept. Stroudsbourg, Pennsylvania: Dowden, Hutchinson & Ross, Inc.; 1973. 	377 378
 Cannon W. Organization for Physiological Homeostatics. Physiol Rev. 1929;9:399–431. 	379 380
 Clancy J, McVicar A. Physiology and Anatomy. A Homeostatic Approach. Second edition. Arnold; 2002. 	381 382
4. Cannon W. The Wisdom of the Body. Revised and enlarged edition. New York: Norton; 1939.	383 384
5. Davis RC. The domain of homeostasis. Psychological Review. 1958;65(1):8–13.	385
6. Rosen R. Anticipatory Systems. Second edition. Springer; 2012.	386
 Rowland MA, Abdelzaher A, Ghosh P, Mayo ML. Crosstalk and the dynamical modularity of feed-forward loops in transcriptional regulatory networks. Biophys J. 2017;112(8):1539–1550. 	387 388 389
8. Leibiger IB, Leibiger B, Berggren PO. Insulin feedback action on pancreatic β -cell function. FEBS Letters. 2002;532(1-2):1–6.	390 391

9.	Jacques-Silva MC, Correa-Medina M, Cabrera O, Rodriguez-Diaz R, Makeeva N, Fachado A, et al. ATP-gated P2X3 receptors constitute a positive autocrine signal for insulin release in the human pancreatic β cell. PNAS. 2010;107(14):6465–6470.	392 393 394 395
10.	Wilkie J, Johnson M, Reza K. Control Engineering. An Introductory Course. New York: Palgrave; 2002.	396 397
11.	Lewis FL. Applied Optimal Control & Estimation. Englewood Cliffs, NJ: Prentice Hall; 1992.	398 399
12.	Ashby WR. An Introduction to Cybernetics. Chapman & Hall; 1961.	400
13.	Wiener N. Cybernetics: or Control and Communication in the Animal and the Machine. Second Edition. Cambridge, Massachusetts: The MIT Press; 1961.	401 402
14.	Cariani PA. The homeostat as embodiment of adaptive control. International Journal of General Systems. 2009;38(2):139–154.	403 404
15.	Milsum JH. Biological Control Systems Analysis. New York: McGraw-Hill; 1966.	405
16.	Milhorn HT. Application of Control Theory to Physiological Systems. Philadelphia: WB Saunders; 1966.	406 407
17.	Saunders PT, Koeslag JH, Wessels JA. Integral rein control in physiology. J Theor Biol. 1998;194(2):163–173.	408 409
18.	Drengstig T, Jolma I, Ni X, Thorsen K, Xu X, Ruoff P. A basic set of homeostatic controller motifs. Biophys J. 2012;103(9):2000–2010.	410 411
19.	Barkai N, Leibler S. Robustness in simple biochemical networks. Nature. 1997;387(6636):913–7.	412 413
20.	Yi TM, Huang Y, Simon MI, Doyle J. Robust perfect adaptation in bacterial chemotaxis through integral feedback control. PNAS. 2000;97:4649–53.	414 415
21.	El-Samad H, Goff JP, Khammash M. Calcium homeostasis and parturient hypocalcemia: an integral feedback perspective. J Theor Biol. 2002;214(1):17–29.	416 417
22.	Ni XY, Drengstig T, Ruoff P. The control of the controller: Molecular mechanisms for robust perfect adaptation and temperature compensation. Biophys J. 2009;97:1244–53.	418 419 420
23.	Jolma IW, Ni XY, Rensing L, Ruoff P. Harmonic oscillations in homeostatic controllers: Dynamics of the p53 regulatory system. Biophys J. 2010;98(5):743–52.	421 422 423
24.	Huang Y, Drengstig T, Ruoff P. Integrating fluctuating nitrate uptake and assimilation to robust homeostasis. Plant, Cell and Environment. 2011;35(5):917–928.	424 425 426
25.	Shoval O, Goentoro L, Hart Y, Mayo A, Sontag E, Alon U. Fold-change detection and scalar symmetry of sensory input fields. PNAS. 2010;(36):15995–16000.	427 428
26.	Drengstig T, Ni XY, Thorsen K, Jolma IW, Ruoff P. Robust Adaptation and Homeostasis by Autocatalysis. J Phys Chem B. 2012;116:5355–5363.	429 430
27.	Briat C, Zechner C, Khammash M. Design of a synthetic integral feedback circuit: dynamic analysis and DNA implementation. ACS Synthetic Biology. 2016;5(10):1108–1116.	431 432 433

28.	Briat C, Gupta A, Khammash M. Antithetic integral feedback ensures robust perfect adaptation in noisy biomolecular networks. Cell Systems. 2016;2(1):15–26.	434 435
29.	Aoki SK, Lillacci G, Gupta A, Baumschlager A, Schweingruber D, Khammash M. A universal biomolecular integral feedback controller for robust perfect adaptation. Nature. 2019;570:533–537.	436 437 438
30.	Seaton D, Krishnan J. Modular systems approach to understanding the interaction of adaptive and monostable and bistable threshold processes. IET Syst Biol. 2011;5(2):81–94.	439 440 441
31.	Krishnan J, Floros I. Adaptive information processing of network modules to dynamic and spatial stimuli. BMC Systems Biology. 2019;13(1):32.	442 443
32.	Fjeld G, Thorsen K, Drengstig T, Ruoff P. The performance of homeostatic controller motifs dealing with perturbations of rapid growth and depletion. J Phys Chem B. 2017;121:6097–6107.	444 445 446
33.	Ruoff P, Agafonov O, Tveit DM, Thorsen K, Drengstig T. Homeostatic controllers compensating for growth and perturbations. PLOS ONE. 2019;14(8):e0207831.	447 448 449
34.	Radhakrishnan K, Hindmarsh AC. Description and Use of LSODE, the Livermore Solver for Ordinary Differential Equations. NASA Reference Publication 1327, Lawrence Livermore National Laboratory Report UCRL-ID-113855. Cleveland, OH 44135-3191: National Aeronautics and Space Administration, Lewis Research Center; 1993.	450 451 452 453 454
35.	Segel IH. Enzyme Kinetics: Behavior and Analysis of Rapid Equilibrium and Steady State Enzyme Systems. New York: Wiley; 1975.	455 456
36.	Goldbeter A. Biochemical Oscillations and Cellular Rhythms. Cambridge: Cambridge University Press; 1996.	457 458
37.	Kruse K, Jülicher F. Oscillations in cell biology. Current Opinion in Cell Biology. 2005;17(1):20–26.	459 460
38.	Shabala S, Shabala L, Gradmann D, Chen Z, Newman I, Mancuso S. Oscillations in plant membrane transport: model predictions, experimental validation, and physiological implications. Journal of Experimental Botany. 2006;57(1):171–184.	461 462 463
39.	Novák B, Tyson JJ. Design principles of biochemical oscillators. Nat Rev Mol Cell Biol. 2008;9(12):981–991.	464 465
40.	Tyson JJ, Albert R, Goldbeter A, Ruoff P, Sible J. Biological switches and clocks. Journal of The Royal Society Interface. 2008;5(suppl_1):S1–S8.	466 467
41.	Bünning E. The Physiological Clock. Berlin: Springer-Verlag; 1963.	468
42.	Dunlap JC, Loros JJ, DeCoursey PJ. Chronobiology. Biological Timekeeping. Sunderland, MA: Sinauer; 2004.	469 470
43.	Moore-Ede MC. Physiology of the circadian timing system: predictive versus reactive homeostasis. American Journal of Physiology-Regulatory, Integrative and Comparative Physiology. 1986;250(5):R737–R752.	471 472 473
44.	Bonny O, Firsov D. Circadian clock and the concept of homeostasis. Cell Cycle. 2009;8(24):4015–14.	474 475

45.	Nakagawa H, Okumura N. Coordinated regulation of circadian rhythms and homeostasis by the suprachiasmatic nucleus. Proceedings of the Japan Academy, Series B. 2010;86(4):391–409.	476 477 478
46.	Solt LA, Kojetin DJ, Burris TP. The REV-ERBs and RORs: Molecular links between circadian rhythms and lipid homeostasis. Future Medicinal Chemistry. 2011;3(5):623–638.	479 480 481
47.	Davis GW. Homeostatic signaling and the stabilization of neural function. Neuron. 2013;80(3):718–728.	482 483
48.	Thorsen K, Agafonov O, Selstø CH, Jolma IW, Ni XY, Drengstig T, et al. Robust concentration and frequency control in oscillatory homeostats. PLOS ONE. 2014;9(9):e107766.	484 485 486
49.	Goodwin BC. Temporal Organization in Cells. Academic Press; 1963.	487
50.	Gonze D, Abou-Jaoudé W. The Goodwin model: Behind the Hill function. PLOS ONE. 2013;8(8):e69573.	488 489
51.	Gonze D, Ruoff P. The Goodwin oscillator and its legacy. Acta Biotheoretica. 2020; p. 1–18.	490 491
52.	Kurosawa G, Iwasa Y. Saturation of enzyme kinetics in circadian clock models. J Biol Rhythms. 2002;17(6):568–577.	492 493
53.	Goodwin BC. Oscillatory behavior in enzymatic control processes. In: Weber G, editor. Advances in Enzyme Regulation, Vol. 3. Oxford, UK: Pergamon Press; 1965. p. 425–438.	494 495 496
54.	Ruoff P, Vinsjevik M, Rensing L. Temperature compensation in biological oscillators: a challenge for joint experimental and theoretical analysis. Comments Theor Biol. 2000;5:361–382.	497 498 499
55.	Ruoff P, Behzadi A, Hauglid M, Vinsjevik M, Havås H. pH homeostasis of the circadian sporulation rhythm in clock mutants of Neurospora crassa. Chronobiology International. 2000;17(6):733–750.	500 501 502
56.	Ang J, McMillen DR. Physical constraints on biological integral control design for homeostasis and sensory adaptation. Biophys J. 2013;104(2):505–515.	503 504
57.	Ruoff P, Rensing L. The temperature-compensated Goodwin model simulates many circadian clock properties. J Theor Biol. 1996;179(4):275–285.	505 506
58.	Ruoff P, Vinsjevik M, Monnerjahn C, Rensing L. The Goodwin model: Simulating the effect of light pulses on the circadian sporulation rhythm of Neurospora crassa. J Theor Biol. 2001;209(1):29–42.	507 508 509
59.	McClung CR. The plant circadian oscillator. Biology. 2019;8(1):14.	510
60.	Nohales MA, Kay SA. Molecular mechanisms at the core of the plant circadian oscillator. Nat Struct & Mol Biol. 2016;23(12):1061.	511 512
61.	Nakamichi N, Kita M, Ito S, Yamashino T, Mizuno T. PSEUDO-RESPONSE REGULATORS, PRR9, PRR7 and PRR5, together play essential roles close to the circadian clock of Arabidopsis thaliana. Plant and Cell Physiology. 2005;46(5):686–698.	513 514 515 516

- 62. Rawat R, Takahashi N, Hsu PY, Jones MA, Schwartz J, Salemi MR, et al. REVEILLE8 and PSEUDO-REPONSE REGULATOR5 form a negative feedback loop within the Arabidopsis circadian clock. PLoS Genet. 2011;7(3):e1001350.
- Kie Q, Wang P, Liu X, Yuan L, Wang L, Zhang C, et al. LNK1 and LNK2 are transcriptional coactivators in the Arabidopsis circadian oscillator. The Plant Cell. 2014;26(7):2843–2857.
- 64. Xing H, Wang P, Cui X, Zhang C, Wang L, Liu X, et al. LNK1 and LNK2 recruitment to the evening element require morning expressed circadian related MYB-like transcription factors. Plant Signaling & Behavior. 2015;10(3):e1010888. 525
- 65. Nakamichi N, Kiba T, Henriques R, Mizuno T, Chua NH, Sakakibara H. PSEUDO-RESPONSE REGULATORS 9, 7, and 5 are transcriptional repressors in the Arabidopsis circadian clock. The Plant Cell. 2010;22(3):594–605.