# Utility of polygenic embryo screening for disease depends on the selection strategy

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#### Abstract

As of 2019, polygenic risk scores have been utilized to screen *in vitro* fertilization embryos for genetic liability to adult diseases, despite a lack of comprehensive modeling of expected outcomes. In this short report, we demonstrate that a strong determinant of the potential utility of such screening is the *selection strategy* employed, a factor that has not been previously studied. Minimal risk reduction is expected if only extremely high-scoring embryos are excluded, whereas risk reductions are substantially greater if the lowest-scoring embryo (for a given disease) is selected. We systematically examined the relative contributions of the variance explained by the score, the number of embryos, the disease prevalence, and parental scores and disease status on the utility of screening. We discuss the results in the context of relative vs absolute risk, as well as the potential ethical concerns raised by such procedures.

#### Introduction

Polygenic risk scores (PRS) have become increasingly well-powered, relying on findings from largescale genome-wide association studies for numerous diseases (Visscher et al., 2017; Wray et al., 2013). The predictive power of a PRS is usually represented by  $R^2$  (or, as we denote it below,  $r_{ps}^2$ ), the proportion of variance of the liability of the disease explained by the score (Dudbridge, 2013). However, for potential clinical applications, this statistical property needs to be translated into clinically actionable information (Torkamani et al., 2018). This translation requires careful consideration of the specific purposes for which the PRS will be used. For example, individuals with polygenic scores in the top percentiles for coronary artery disease (CAD) were shown to be as likely to have a heart attack as individuals heterozygous for a familial hypercholesterolemia mutation (Khera et al., 2018), and may therefore be good candidates for preventative treatment. By contrast, a wider range of high PRS scores (e.g, top quartile), may provide useful information as a part of a multimodal screening process for cancer risk (Callender et al., 2019; X. Zhang et al., 2018).

Another potential clinical application of PRSs is preimplantation screening of *in vitro* fertilization (IVF) embryos. Although currently in use (Treff, Eccles, et al., 2019) and fraught with ethical concerns (Lázaro-Muñoz et al., 2020), "polygenic embryo screening" (PES) has not yet received much research attention from either geneticists or ethicists. Understanding the statistical properties of PES forms a critical foundation to ethical consideration of the practice (Lázaro-Muñoz et al., 2020). For example, we have recently demonstrated that screening embryos on the basis of polygenic scores for quantitative traits (such as height or intelligence) has limited predictive power in most realistic scenarios (Karavani et al., 2019), and that  $r_{ps}^2$  is a more significant determinant of PES utility for quantitative traits compared with the number of available embryos (*n*). On the other hand, a series of four studies (Lello et al., 2020; Treff, Eccles, et al., 2019; Treff et al., 2020; Treff, Zimmerman, et al., 2019) conducted by a private company providing embryo screening services has suggested that PES for dichotomous disease risk may have significant clinical utility. However, these studies examined a relatively limited range of scenarios,

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primarily focusing on distinctions between sibling pairs discordant for illness, and did not provide a comprehensive examination of the potential utility of PES.

Here, we use statistical modeling to examine the potential utility of PES for disease risk (defined in terms of relative and absolute risk reduction), comparable to our prior study of PES for quantitative traits (Karavani et al., 2019), with an aim toward informing future ethical deliberations on the practice. We study a range of realistic scenarios, quantifying the role of  $\tau_{ps}^2$ , the number of embryos (*n*), the disease prevalence, and parental risk scores and disease status on the ability of PES to reduce disease risk when screening for a single disease (*Materials and Methods*). We utilize the liability threshold model (Falconer, 1967) to represent disease risk as a continuous liability, comprising genetic and environmental risk factors, under the assumption that individuals with liability exceeding a threshold are affected. The liability threshold model was shown to be consistent with data from family-based transmission studies (Wray & Goddard, 2010) and GWAS data (Visscher & Wray, 2015). Consequently, we define the disease risk of a given embryo probabilistically, as the chance that its liability will cross the threshold at any point after birth (**Figure 1A**).

In studies of potential clinical utility of PRSs in adults, attention has focused on those in the highest percentiles of risk, in which odds ratios become sufficiently large to be clinically meaningful (Chatterjee et al., 2016; Dai et al., 2019; Gibson, 2019; Khera et al., 2018; Mars et al., 2020; Mavaddat et al., 2019; Torkamani et al., 2018). To our knowledge, PES is currently being employed in a similar fashion, following a strategy of excluding embryos with extremely high (top 2-percentiles) PRS (Treff, Eccles, et al., 2019; Treff et al., 2020), which we term "*high-risk exclusion*" (HRE: **Figure 1B**, upper panel). In such a strategy, after high-risk embryos are set aside, an embryo is randomly selected for implantation among the remaining available embryos. (In the rare scenario that all embryos are high-risk, we assume a random embryo is selected among them.) We utilized both theory and simulations to study this strategy, given varying disease prevalence, strength of PRS, and embryo exclusion thresholds. As elaborated below, we demonstrate that the HRE strategy has very limited utility, but that a strategy of selecting the embryo with the lowest genetic risk can result in large risk reductions.

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#### **Results and Discussion**

We quantified the outcome of PES in terms of relative risk reduction (RRR), defined as RRR =  $\frac{K-P(\text{disease})}{K} = 1 - \frac{P(\text{disease})}{K}$ , where K is the prevalence. For example, if a disease has prevalence of 5% and the selected embryo has a probability of 3% to be affected, the relative risk reduction is 40%. The achievable risk reductions with the *high-risk exclusion* strategy (*Materials and Methods*) are plotted in **Figure 2** (upper row), showing strong dependence on the PRS exclusion threshold. When the 2-percentile threshold is applied (straight black lines), the reduction in risk is limited; RRR is <10% in all scenarios where  $r_{ps}^2 \leq 0.1$ . Currently,  $r_{ps}^2 \approx 0.1$  is the upper limit of predictive power (on the liability scale) of PRS for most complex diseases (Khera et al., 2018; Lambert et al., 2019), with the exception of a few disorders with large-effect common variants (such as Alzheimer's disease or type 1 diabetes) (Sharp et al., 2019; Q. Zhang et al., 2020).

In the future, more accurate PRSs are expected; however, it has been suggested that  $r_{ps}^2 = 0.3$ , which is at the top end of the common-variant SNP heritability for even the most heritable diseases such as schizophrenia and celiac disease (Holland et al., 2020; Y. Zhang et al., 2018), is the maximal realistic value for the foreseeable future (Wray et al., 2020). At this value, relative risk reduction would only be 20% for K = 0.01, 9% for K = 0.05, and 3% for K = 0.2. These small gains achieved with *high-risk exclusion* follow because the overwhelming majority of affected individuals do not have extreme scores (Murray et al., 2020; Wald & Old, 2019).

Risk reduction increases as the threshold for exclusion is expanded to include the top quartile of scores, and then reaches a maximum at  $\approx 25-50\%$  under a range of prevalence and  $r_{ps}^2$  values. For all of these simulations, we set the number of available embryos to n = 5, which is a reasonable estimate of the number of testable, viable embryos from a typical IVF cycle (Sunkara et al., 2011). Simulations show that these estimates do not change much with increasing the number of embryos (see **Figure 2 - Figure Supplement 1**). This holds especially at more extreme threshold values, since most batches of *n* embryos will not contain any embryos with a PRS within, e.g., the top 2-percentiles.

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The *high-risk exclusion* strategy treats non-high-risk embryos equally; however, other strategies are possible. For example, current research on optimizing IVF protocols focuses on ranking embryos for potential viability, on the basis of microscopy and time-lapse imaging (Bormann et al., 2020; Montag et al., 2013; Rhenman et al., 2015). Thus, it is readily conceivable that the ranking of embryos on the basis of disease PRS could also be attempted. We term the implantation of the embryo with the lowest PRS as "*lowest-risk-prioritization*" (LRP; **Figure 1b**, lower panel). As we show in **Figure 2** (lower panels), for LRP with n = 5 available embryos, RRR>20% across the entire range of prevalence and  $r_{ps}^2$  parameters considered, and can reach  $\approx 50\%$  for  $K \le 5\%$  and  $r_{ps}^2 = 0.1$ , and even  $\approx 80\%$  for K = 1% and  $r_{ps}^2 = 0.3$ . While RRR continues to increase as the number of available embryos increases, the gains are quickly diminishing after n = 5.

We also examined the effects of parental PRSs on the achievable risk reduction, given the possibility that families with high genetic risk for a given disease would be more likely to seek PES. **Figure 2** - **Figure Supplement 2** demonstrates that, as expected, the HRE strategy shows greater relative risk reduction as parental PRS increases, in particular when excluding only very high-scoring embryos. In contrast, the RRR for the LRP strategy is relatively stable across parental PRSs. Nevertheless, as expected, the RRR for the LRP strategy remains greater than that for the HRE strategy across all possible combinations of parameters. Similarly, it is possible that families may be more likely to seek PES when one or both prospective parents is affected by a given disease. **Figure 2 - Figure Supplement 3** illustrates that parental disease status has relatively little impact on the expected risk reduction, especially in comparison to conditioning on the parental polygenic risk scores. This is because, as long as  $r_{ps}^2 \ll 1$ , parental disease does not necessarily provide much information about parental PRS, and thus does not strongly constrain the number of risk alleles available to each embryo.

It is important to emphasize that all of the above results are presented in terms of *relative* risk reduction. The clinical interpretation of these changes in terms of *absolute risk* will vary based on the population prevalence of the disorder (or the baseline risk of specific parents), and can offer a very

different perspective on the magnitude of the effects (**Figure 2 - Figure Supplement 4**) (Gordis, 2014; Lázaro-Muñoz et al., 2020; Murray et al., 2020). Specifically, a large *relative* risk reduction may result in a very small change in *absolute* risk for a rare disease. As an example, schizophrenia is a highly heritable (Sullivan et al., 2003) serious mental illness with prevalence of at most 1% (Perälä et al., 2007). The most recent large-scale GWAS meta-analysis for schizophrenia (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2020) has reported that a PRS accounts for approximately 8% of the variance on the liability scale. Our model shows that a 52% RRR is attainable using the LRP strategy with n = 5 embryos. However, this translates to only  $\approx$ 0.5 percentage points reduction on the absolute scale: a randomly-selected embryo would have a 99% chance of not developing schizophrenia, compared to a 99.5% chance for an embryo selected according to LRP. In the case of a more common disease such as type 2 diabetes, with a lifetime prevalence in excess of 20% in the United States (Geiss et al., 2014), the RRR with n = 5 embryos (if the full SNP heritability of 17% (Y. Zhang et al., 2018) were achieved) is 43%, which would correspond to >8 percentage points reduction in absolute risk.

The results of the present study demonstrate that, contrary to our previous study of PES for quantitative traits (Karavani et al., 2019), substantial effects of PES for disease risk are attainable under certain conditions. Specifically, we observed that the *selection strategy* is a crucial determinant of risk reduction. The use of PRS in adults has focused on those at highest risk (Chatterjee et al., 2016; Dai et al., 2019; Gibson, 2019; Khera et al., 2018; Mars et al., 2020; Mavaddat et al., 2019; Torkamani et al., 2018), for whom there may be maximal clinical benefit of screening and intervention. However, as PRSs have relatively low sensitivity, such a strategy is relatively ineffective in reducing the overall population disease burden (Ala-Korpela & Holmes, 2020; Wald & Old, 2019). Similarly, in the context of PES, exclusion of high-risk embryos will result in relatively modest risk reductions.

The differential performance of PES across selection strategies and risk reduction metrics may be difficult to communicate to couples seeking assisted reproductive technologies (Cunningham et al., 2015; Wilkinson et al., 2019). These difficulties are expected to exacerbate the already profound ethical issues raised by PES (as we have recently reviewed (Lázaro-Muñoz et al., 2020)), which include stigmatization

(McCabe & McCabe, 2011), autonomy (including "choice overload" (Hadar & Sood, 2014)), and equity (Sueoka, 2016). In addition, the ever-present specter of eugenics (Lombardo, 2018) may be especially salient in the context of the LRP strategy. We thus call for urgent deliberations amongst key stakeholders (including researchers, clinicians, and patients) to address governance of PES and for the development of policy statements by professional societies. We hope that our statistical framework can provide an empirical foundation for these critical ethical and policy deliberations.

The present study has several limitations. First, our analytical modeling is based on several simplifying assumptions; as we discuss in the *Materials and Methods*, we do not expect these assumptions to substantially impact the risk reduction estimates, although our estimates likely represent an upper bound relative to real-world scenarios. For example, we did not model the role of family-specific environmental factors (Wang et al., 2017), including the influence of parental genetic factors on the child's environment (Kong et al., 2018; Morris et al., 2020; Mostafavi et al., 2020; Young et al., 2019). Additionally, given the fact that polygenic risk scores are often correlated across diseases (Watanabe et al., 2019; Zheng et al., 2017), selecting based on the PRS of one disease may increase or decrease risk for other diseases. In practice, couples may seek to profile an embryo on the basis of multiple disease PRSs simultaneously, which may affect the overall and per-disease outcomes. These more complex questions will be the subject of future work. Finally, for the purposes of our calculations, we assumed  $r_{ps}^2$  represents the realistic accuracy achievable (within-family) in a real-world setting in the target population. However, the accuracy of PRSs may be sub-optimal when applied in non-European populations, across different socio-economic groups, or within siblings (as compared to between unrelated individuals) (Duncan et al., 2019; Mostafavi et al., 2020). Moreover, currently constructed PRSs do not include rare or ultra-rare variants (including copy number variants), which may have greater penetrance than common variants. On the other hand, future developments in PRS construction will likely increase the realized  $r_{ps}^2$  for some diseases. Either way, the analyses presented in this paper cover a broad range of values of  $r_{ps}^2$  likely to be available at present and in the foreseeable future.

# **1** Materials and Methods

# 2 1 Methods summary

Our model for the polygenic risk scores of a batch of n IVF embryos is described in detail below. Briefly, we write 3 the polygenic scores of the embryos as  $(s_1, \ldots, s_n)$ , where  $s_i = x_i + c$ ,  $(x_1, \ldots, x_n)$  are embryo-specific independent 4 variables with distribution  $x_i \sim N(0, r_{ps}^2/2)$  (where  $r_{ps}^2$  is the proportion of variance in liability explained by the score), 5 and c is a shared component with distribution  $c \sim N(0, r_{ps}^2/2)$ , also representing the mean parental score. In each batch, 6 an embryo is selected according to the selection strategy. For lowest-risk prioritization, we select the embryo with the 7 lowest value of s. For high-risk exclusion, we select the first embryo with score  $s < z_q r_{ps}$ , where  $z_q$  is the (1 - q)-8 9 quantile of the standard normal distribution. (If no such embryo exists, we select the first embryo.) The liability of the selected embryo is computed as y = s + e, where  $e \sim N(0, 1 - r_{ps}^2)$ . We designate the embryo as affected if  $y > z_K$ , 10 where  $z_K$  is the (1 - K)-quantile of the standard normal distribution and K is the prevalence. For each parameter setting, 11 we compute the disease probability as the fraction of batches (out of 10<sup>6</sup> repeats) in which the selected embryo was 12 13 affected.

14 We also compute the disease probability analytically. For lowest-risk prioritization, we derive the disease risk based on the theory of order statistics. For high-risk exclusion, we first condition on the shared component c, and then 15 study separately the case when all embryos are high-risk (have score  $s > z_q r_{ps}$ ), in which the distribution of the unique 16 17 component of the selected embryo (x) is a normal variable truncated from below at  $z_q r_{ps}$ , and the case when at least one embryo has score  $s < z_q r_{ps}$ , in which x is a normal variable truncated from above. We solved the integrals in the final 18 19 expressions numerically in R. We also provide explicit results for the case when the family-specific component c is 20known. Finally, if the parental disease status is known, we integrate the disease probability of the selected embryo over the posterior distribution of the parental score and non-score genetic components. For full details and for an additional 21 discussion of previous work and limitations, see the sections below. 22

# 23 **2** The liability threshold model

The liability threshold model (LTM) is a classic model in quantitative genetics (Dempster and Lerner, 1950; Falconer, 1965; Lynch and Walsh, 1998), and is also commonly used to analyze modern data (e.g., (Wray and Goddard, 2010; So et al., 2011; Lee et al., 2011, 2012; Do et al., 2012; Hayeck et al., 2017; Weissbrod et al., 2018; Hujoel et al., 2020)).

The LTM assumes that a disease has an underlying "liability", which is normally distributed in the population, and is
 the sum of two components: genetic and non-genetic (the environment). Further, the LTM assumes an "infinitesimal",
 or "polygenic" genetic basis, under which a very large number of genetic variants of small effect combine to form the
 genetic component. An individual is affected if his/her total liability (genetic + environmental) exceeds a threshold.

5 Mathematically, if we denote the liability as *y*, the LTM can be written as

$$y = g + \mathcal{E},\tag{1}$$

6 where  $y \sim N(0,1)$  is a standard normal variable,  $g \sim N(0,h^2)$  is the genetic component, with variance equal to the 7 (narrow-sense) heritability  $h^2$ , and  $\varepsilon \sim N(0, 1 - h^2)$  is the non-genetic component. In practice, we cannot measure the 8 genetic component, but only estimate it imprecisely with a polygenic risk score, denoted *s*. Following previous work 9 (So et al., 2011; Lee et al., 2012; Treff et al., 2019; Karavani et al., 2019), we assume that the LTM can be written, 10 similarly to Eq. (1), as

$$y = s + e, \tag{2}$$

where  $y \sim N(0,1)$  as above,  $s \sim N(0, r_{ps}^2)$ , where  $r_{ps}^2$  is the proportion of the variance in liability explained by the score, and  $e \sim N(0, 1 - r_{ps}^2)$  is the residual of the regression of the liability on *s* (and is uncorrelated with *s*), representing environmental effects as well as genetic factors not accounted for by the score.

An individual is affected whenever his/her liability exceeds a threshold, i.e.,  $y > \tau$ . The threshold is selected such that the proportion of affected individuals is equal to the prevalence *K*, i.e.,  $\tau = z_K$ , where  $z_K$  is the (1 - K)-quantile of a standard normal variable. Thus,

$$P(\text{disease}) = P(y > z_K) = K.$$
(3)

17 The model is illustrated in Figure 1A of the main text.

# 18 **3** A model for the scores of *n* IVF embryos

19 Consider the polygenic risk scores (for a disease of interest) of n IVF embryos of given parents. We assume no 20 information is known about the parents, or, in other words, that the parents are randomly drawn from the population.

21 The scores of the embryos have a multivariate normal distribution,

$$\boldsymbol{s} = (s_1, \dots, s_n) = \mathrm{MVN}(\boldsymbol{0}_n, \boldsymbol{\Sigma}), \tag{4}$$

1 where the means form a vector of *n* zeros, and the  $n \times n$  covariance hbfrix is

$$\Sigma = r_{\rm ps}^2 \begin{pmatrix} 1 & \frac{1}{2} & \dots & \frac{1}{2} \\ \frac{1}{2} & 1 & \dots & \frac{1}{2} \\ \dots & \dots & \dots & \dots \\ \frac{1}{2} & \frac{1}{2} & \dots & 1 \end{pmatrix}.$$
(5)

2 The diagonal elements of the matrix are simply the variances of the individual scores of each embryo. The off3 diagonal elements represent the covariance between the scores of the embryos, who are genetically siblings. Based on
4 standard quantitative genetic theory (Lynch and Walsh, 1998) (see also our previous paper (Karavani et al., 2019)), the

- 5 covariance between the scores of two siblings is  $Cov(s_i, s_j) = \frac{1}{2}Var(s)$ , and hence the off-diagonal elements follow.
- 6 [The non-score components (the *e* terms in Eq. (2)) are also correlated, as they represent both genetic and environmental
- 7 factors. However, this does not affect the results of the present paper. See also Section 9 below.]

As we showed in our previous work (Karavani et al., 2019), the scores can be written as a sum of two independent multivariate normal variables, s = x + c, with

$$\boldsymbol{x} = (x_1, \dots, x_n) \sim \text{MVN}\left(\boldsymbol{0}_n, \frac{r_{\text{ps}}^2}{2}\boldsymbol{I}_n\right),$$
$$\boldsymbol{c} = (c_1, \dots, c_n) \sim \text{MVN}\left(\boldsymbol{0}_n, \frac{r_{\text{ps}}^2}{2}\boldsymbol{J}_n\right),$$
(6)

8 where  $\mathbf{0}_n$  is a vector of zeros of length *n*,  $\mathbf{I}_n$  is the  $n \times n$  identity matrix, and  $\mathbf{J}_n$  is the  $n \times n$  matrix of all ones. The 9  $x_i$ 's and  $c_i$ 's have the same marginal distribution, namely normal with mean zero and variance  $r_{ps}^2/2$  each. However, 10 the  $x_i$ 's are independent, whereas *c* has a constant covariance matrix, which means that the  $c_i$ 's are *n* identical copies 11 of the same random variable,

$$c_1 \sim N\left(0, \frac{r_{\rm ps}^2}{2}\right)$$
 and  $c_2 = c_3 = \dots = c_n = c_1 \equiv c.$  (7)

12 Thus, for each embryo i = 1, ..., n,

$$s_i = x_i + c. \tag{8}$$

# 3.1 An alternative interpretation: conditioning on the average parental scores

3 The representation of the score in Eq. (8) can also be interpreted as conditioning on the average score of the parents.

4 To see that, write the maternal score as  $s_m$  and the paternal score as  $s_f$ . The variables  $(s_i, s_m, s_f)$  have a multivariate

5 normal distribution, with

$$f(s_i, s_m, s_f) \sim \text{MVN}\left(\begin{pmatrix} 0\\0\\0\\0 \end{pmatrix}, \begin{pmatrix} r_{\text{ps}}^2 & \frac{r_{\text{ps}}^2}{2} & \frac{r_{\text{ps}}^2}{2}\\ \frac{r_{\text{ps}}^2}{2} & r_{\text{ps}}^2 & 0\\ \frac{r_{\text{ps}}^2}{2} & 0 & r_{\text{ps}}^2 \end{pmatrix}\right).$$
(9)

In the above equation, the variances of all scores are equal to  $r_{ps}^2$ . The covariance terms are  $Cov(s_i, s_m) = Cov(s_i, s_f) = \frac{1}{2}Var(s) = \frac{r_{ps}^2}{2}$ , as the relatedness between between parent and child is the same as for pairs of siblings. We assume no correlation between the scores of the parents (i.e., no assortative mating, see Section 10 for discussion). We are now interested in the density of  $s_i$  given  $s_m$  and  $s_f$ . Using standard results for multivariate normal distributions, the conditional density of  $s_i$  is  $N(\mu, \sigma^2)$ , where,

$$\mu = \Sigma_{12} \Sigma_{22}^{-1} \begin{pmatrix} s_m \\ s_f \end{pmatrix},$$
  
$$\sigma^2 = \Sigma_{11} - \Sigma_{12} \Sigma_{22}^{-1} \Sigma_{21},$$
 (10)

6 where

$$\Sigma_{11} = r_{\rm ps}^2, \Sigma_{12} = \begin{pmatrix} \frac{r_{\rm ps}^2}{2} & \frac{r_{\rm ps}^2}{2} \end{pmatrix}, \Sigma_{21} = \begin{pmatrix} \frac{r_{\rm ps}^2}{2} \\ \frac{r_{\rm ps}^2}{2} \end{pmatrix}, \Sigma_{22} = \begin{pmatrix} r_{\rm ps}^2 & 0 \\ 0 & r_{\rm ps}^2 \end{pmatrix}.$$
(11)

Carrying out the matrix calculations, we obtain

$$\mu = \frac{s_m + s_f}{2},$$
  
$$\sigma^2 = \frac{r_{\rm ps}^2}{2}.$$
 (12)

7 Thus,

$$s_i \sim N\left(\frac{s_m + s_f}{2}, \frac{r_{\rm ps}^2}{2}\right) \equiv N(c, r_{\rm ps}^2/2),$$
 (13)

1 where we defined the shared component  $c \equiv \frac{s_m + s_f}{2}$  as the average parental score. We therefore have the distribution 2 of the score of the child given the average parental score. The variance of *c* itself at the population is  $\operatorname{Var}\left(\frac{s_m + s_f}{2}\right) =$ 3  $\frac{2\operatorname{Var}(s)}{4} = r_{\mathrm{ps}}^2/2$ . Thus, across the population,  $c \sim N(0, r_{\mathrm{ps}}^2/2)$ . In a given family, *c* is the same across all embryos. Thus, 4 Eq. (13) is equivalent to  $s_i = c + x_i$ , with  $c \sim N(0, r_{\mathrm{ps}}^2/2)$  and  $x_i \sim N(0, r_{\mathrm{ps}}^2/2)$  is an embryo-specific component.

5 An analogous result holds for the total genetic component of the embryo  $g_i$ , simply by replacing the variance 6 explained by the heritability  $h^2$ . In other words, if  $g_m$  and  $g_f$  are the maternal and paternal genetic components, 7 respectively, then

$$g_i \sim N\left(\frac{g_m + g_f}{2}, \frac{h^2}{2}\right). \tag{14}$$

# 8 4 The disease risk when implanting the embryo with the lowest

9 risk

We assume next that we select for implantation the embryo with the lowest polygenic risk score for the disease of interest. Our goal will be to calculate the probability of that embryo to be affected. Since  $s_i = x_i + c$ , the score of the selected embryo satisfies

$$s_{\min} = \min(x_1 + c, \dots, x_n + c)$$
$$= \min(x_1, \dots, x_n) + c$$
$$= x_{\min} + c,$$
(15)

where we defined  $x_{\min} = \min(x_1, \dots, x_n)$ . Denote by  $i^*$  the index of the selected embryo  $(x_{i^*} = x_{\min})$ . The liability of the embryo with the lowest risk is thus

$$y_{i^*} = s_{\min} + e_{i^*}$$
$$= x_{\min} + c + e_{i^*}$$
$$= x_{\min} + \tilde{e},$$
(16)

10 where  $\tilde{e} = c + e_{i^*}$ . Consequently,

$$\operatorname{Var}\left(\tilde{e}\right) = \operatorname{Var}\left(c\right) + \operatorname{Var}\left(e_{i^*}\right) = \frac{r_{\mathrm{ps}}^2}{2} + (1 - r_{\mathrm{ps}}^2) = 1 - \frac{r_{\mathrm{ps}}^2}{2}.$$
(17)

- 1 Therefore, the liability of the selected embryo can be written as a sum of two (independent) variables:  $x_{\min}$ , which
- 2 is the minimum of *n* independent (zero mean) normal variables with variance  $r_{ps}^2/2$  each; and  $\tilde{e}$ , which is a normal
- 3 variable with (zero mean and) variance  $1 r_{ps}^2/2$ .

4 The distribution of  $x_{\min}$  can be computed based on the theory of order statistics,

$$P(x_{\min} > x_m) = [P(x > x_m)]^n = \left[1 - \Phi\left(\frac{x_m}{r_{\rm ps}/\sqrt{2}}\right)\right]^n.$$
(18)

- 5 In the above equation, the probability that the minimum of n variables is greater than  $x_m$  is the probability that all
- 6 variables are greater than  $x_m$ . The distribution of each x is normal with zero mean and variance  $r_{ps}^2/2$ , and hence
- 7  $P(x > x_m) = 1 \Phi\left(\frac{x_m}{r_{ps}/\sqrt{2}}\right)$ , where  $\Phi(\cdot)$  is the cumulative probability distribution (CDF) of a standard normal variable. We can now compute the probability of the selected embryo to be affected by the disease by demanding that the

total liability is greater than the threshold  $z_K$ . Conditional on  $\tilde{e}$ ,

$$P(\text{disease} | \tilde{e}) = P(y_{i^*} > z_K | \tilde{e})$$

$$= P(x_{\min} + \tilde{e} > z_K)$$

$$= P(x_{\min} > z_K - \tilde{e})$$

$$= \left[1 - \Phi\left(\frac{z_K - \tilde{e}}{r_{\text{ps}}/\sqrt{2}}\right)\right]^n, \qquad (19)$$

where in the fourth line we used Eq. (18). Next, denote by  $f(\tilde{e})$  the density of  $\tilde{e}$ , and by  $\phi(\cdot)$  for the probability density function of the standard normal variable. Given that  $\tilde{e} \sim N(0, 1 - r_{\rm ps}^2/2)$ ,

$$P(\text{disease}) = \int_{-\infty}^{\infty} P(\text{disease} \,|\, \tilde{e}) f(\tilde{e}) d\tilde{e}$$
  
$$= \int_{-\infty}^{\infty} \left[ 1 - \Phi\left(\frac{z_K - \tilde{e}}{r_{\text{ps}}/\sqrt{2}}\right) \right]^n \frac{1}{\sqrt{1 - r_{\text{ps}}^2/2}} \phi\left(\frac{\tilde{e}}{\sqrt{1 - r_{\text{ps}}^2/2}}\right) d\tilde{e}$$
  
$$= \int_{-\infty}^{\infty} \left[ 1 - \Phi\left(\frac{z_K - t\sqrt{1 - r_{\text{ps}}^2/2}}{r_{\text{ps}}/\sqrt{2}}\right) \right]^n \phi(t) dt.$$
(20)

8 In the third line, we changed variables:  $t = \tilde{e}/\sqrt{1 - r_{ps}^2/2}$ . Eq. (20) is our final expression for the probability of the 9 embryo with the lowest score to be affected.

#### 1 4.1 The risk reduction when conditioning on the mean parental score

Consider the case when c is given, or, in other words, that we know the mean parental polygenic score. Let us compute the disease risk in such a case. We start from Eq. (16),

$$y_{i^*} = s_{\min} + e_{i^*}$$
  
=  $x_{\min} + c + e_{i^*}$ . (21)

Then,

$$P(\text{disease} | c, e_{i^{*}}) = P(y_{i^{*}} > z_{K} | e_{i^{*}})$$
  
=  $P(x_{\min} + c + e_{i^{*}} > z_{K})$   
=  $P(x_{\min} > z_{K} - c - e_{i^{*}})$   
=  $\left[1 - \Phi\left(\frac{z_{K} - c - e_{i^{*}}}{r_{\text{ps}}/\sqrt{2}}\right)\right]^{n}$ , (22)

2 where in the last line we used Eq. (18).

Finally, recalling that  $e_{i^*} \sim N(0, 1 - r_{ps}^2)$ ,

$$P(\text{disease} | c) = \int_{-\infty}^{\infty} P(\text{disease} | c, e_{i^{*}}) f(e_{i^{*}}) de_{i^{*}}$$

$$= \int_{-\infty}^{\infty} \left[ 1 - \Phi\left(\frac{z_{K} - c - e_{i^{*}}}{r_{\text{ps}}/\sqrt{2}}\right) \right]^{n} \frac{1}{\sqrt{1 - r_{\text{ps}}^{2}}} \phi\left(\frac{e_{i^{*}}}{\sqrt{1 - r_{\text{ps}}^{2}}}\right) de_{i^{*}}$$

$$= \int_{-\infty}^{\infty} \left[ 1 - \Phi\left(\frac{z_{K} - c - t\sqrt{1 - r_{\text{ps}}^{2}}}{r_{\text{ps}}/\sqrt{2}}\right) \right]^{n} \phi(t) dt, \qquad (23)$$

3 where in the last line we changed variables,  $t = e_{i^*} / \sqrt{1 - r_{ps}^2}$ . Eq. (23) thus provides the probability of disease when 4 we are given the mean parental score *c*.

# **5 5 The disease risk when excluding high-risk embryos**

6 We now consider the selection strategy in which the embryo for implantation is selected at random, as long as its risk 7 score is not particularly high. Specifically, we assume that whenever possible, embryos at the top *q* risk percentiles are

1 excluded. When *all* embryos have high risk, we assume that a random embryo is selected. Let  $z_q$  be the (1-q)-quantile 2 of the standard normal distribution. The variance of the score is  $r_{ps}^2$ , and therefore, the score of the selected embryo 3 must be lower than  $z_q r_{ps}$ .

To compute the disease risk in this case, we first condition on the shared, family-specific component c. We later integrate over c to derive the risk across the population. Denote by  $x_s$  the value of x for the *selected* embryo, and for the moment, also condition on  $x_s$ . We have,

$$P(\text{disease} | x_s, c) = P(y > z_K | c)$$

$$= P(s + e > z_K)$$

$$= P(x_s + c + e > z_K)$$

$$= P(e > z_K - x_s - c)$$

$$= 1 - \Phi\left(\frac{z_K - x_s - c}{\sqrt{1 - r_{\text{ps}}^2}}\right), \qquad (24)$$

To obtain P(disease | c), we need to integrate over  $f(x_s)$ , the density of  $x_s$ . In fact,  $f(x_s)$  is a mixture of two distributions, depending on whether or not all embryos were high risk. Denote by H the event that all embryos are high risk, and let us first compute the probability of H. Recall that given c, the scores of all embryos,  $s_i = x_i + c$ , are independent. The event H is equivalent to the intersection of the *independent* events  $s_i > z_q r_{ps}$  for i = 1, ..., n. Thus, recalling that  $x_i \sim N(0, r_{ps}^2/2)$ ,

$$P(H) = \prod_{i=1}^{n} P(s_i > z_q r_{ps})$$
  
=  $\prod_{i=1}^{n} P(x_i + c > z_q r_{ps})$   
=  $\prod_{i=1}^{n} P(x_i > z_q r_{ps} - c)$   
=  $\left[1 - \Phi\left(\frac{z_q r_{ps} - c}{r_{ps}/\sqrt{2}}\right)\right]^n$ . (25)

Given *H*, we know that all scores were higher than the cutoff, i.e., that  $x_i > z_q r_{ps} - c$  for all i = 1, ..., n. An embryo is then selected at random, and again, we recall that the  $x_i$ 's are independent. Thus,  $x_s$ , the value of *x* of the selected embryo, is a realization of a normal random variable truncated from below. Specifically, if  $f_x(\cdot)$  is the unconditional

1 density of *x*, then for  $x_s > z_q r_{ps} - c$ ,

$$f(x_s|H) = \frac{f_x(x_s)}{P(x > z_q r_{\rm ps})} = \frac{\frac{1}{r_{\rm ps}/\sqrt{2}} \phi\left(\frac{x_s}{r_{\rm ps}/\sqrt{2}}\right)}{1 - \Phi\left(\frac{z_q r_{\rm ps} - c}{r_{\rm ps}/\sqrt{2}}\right)}.$$
(26)

In the case *H* did not occur, we select an embryo at random among embryos with score  $s_i < z_q r_{ps}$ , i.e.,  $x_i < z_q r_{ps} - c$ . The density of  $x_s$  is again, analogously to the above case, a realization of a normal random variable, but this time truncated from above. For  $x_s < z_q r_{ps} - c$ ,

$$f(x_s | \overline{H}) = \frac{f_x(x_s)}{P(x < z_q r_{\rm ps})} = \frac{\frac{1}{r_{\rm ps}/\sqrt{2}} \phi\left(\frac{x_s}{r_{\rm ps}/\sqrt{2}}\right)}{\Phi\left(\frac{z_q r_{\rm ps} - c}{r_{\rm ps}/\sqrt{2}}\right)}.$$
(27)

Using these results, we can write the unconditional density of  $x_s$ ,

$$f(x_s) = \begin{cases} f(x_s | H) P(H) + 0 \cdot P(\overline{H}) & \text{for } x_s > z_q r_{\text{ps}} - c \\ 0 \cdot P(H) + f(x_s | \overline{H}) P(\overline{H}) & \text{for } x_s < z_q r_{\text{ps}} - c \end{cases}$$
$$= \begin{cases} \frac{1}{\frac{r_{\text{ps}}/\sqrt{2}}{p_s}} \phi\left(\frac{x_s}{r_{\text{ps}}/\sqrt{2}}\right)}{1 - \Phi\left(\frac{z_q r_{\text{ps}} - c}{r_{\text{ps}}/\sqrt{2}}\right)} \left[ 1 - \Phi\left(\frac{z_q r_{\text{ps}} - c}{r_{\text{ps}}/\sqrt{2}}\right) \right]^n & \text{for } x_s > z_q r_{\text{ps}} - c \end{cases}$$
$$= \begin{cases} \frac{1}{\frac{r_{\text{ps}}/\sqrt{2}}{p_s}} \phi\left(\frac{x_s}{r_{\text{ps}}/\sqrt{2}}\right)}{1 - \Phi\left(\frac{z_q r_{\text{ps}} - c}{r_{\text{ps}}/\sqrt{2}}\right)} \left[ 1 - \Phi\left(\frac{z_q r_{\text{ps}} - c}{r_{\text{ps}}/\sqrt{2}}\right) \right]^n & \text{for } x_s > z_q r_{\text{ps}} - c \end{cases}$$
(28)

We can now integrate over all  $x_s$ , still conditioning on c, and using Eq. (24) and some algebra,

$$P(\text{disease} | c) = \int_{-\infty}^{\infty} f(x_s) P(\text{disease} | x_s, c) dx_s$$

$$= \int_{-\infty}^{z_q r_{\text{ps}}-c} \frac{\frac{1}{r_{\text{ps}}/\sqrt{2}} \phi\left(\frac{x_s}{r_{\text{ps}}/\sqrt{2}}\right)}{\Phi\left(\frac{z_q r_{\text{ps}}-c}{r_{\text{ps}}/\sqrt{2}}\right)} \left\{ 1 - \left[1 - \Phi\left(\frac{z_q r_{\text{ps}}-c}{r_{\text{ps}}/\sqrt{2}}\right)\right]^n \right\} \left[1 - \Phi\left(\frac{z_K - x_s - c}{\sqrt{1 - r_{\text{ps}}^2}}\right)\right] dx_s$$

$$+ \int_{z_q r_{\text{ps}}-c}^{\infty} \frac{\frac{1}{r_{\text{ps}}/\sqrt{2}} \phi\left(\frac{x_s}{r_{\text{ps}}/\sqrt{2}}\right)}{1 - \Phi\left(\frac{z_q r_{\text{ps}}-c}{r_{\text{ps}}/\sqrt{2}}\right)} \left[1 - \Phi\left(\frac{z_q r_{\text{ps}}-c}{r_{\text{ps}}/\sqrt{2}}\right)\right]^n \left[1 - \Phi\left(\frac{z_K - x_s - c}{\sqrt{1 - r_{\text{ps}}^2}}\right)\right] dx_s$$

$$= \int_{-\infty}^{\infty} \eta(t) \xi(t) dt, \qquad (29)$$

where we defined

$$\gamma = \sqrt{2}z_q - \frac{c}{r_{\rm ps}/\sqrt{2}},$$

$$\xi(t) = \phi(t) \left[ 1 - \Phi\left(\frac{z_K - tr_{\rm ps}/\sqrt{2} - c}{\sqrt{1 - r_{\rm ps}^2}}\right) \right], \text{ and}$$

$$\eta(t) = \begin{cases} \frac{1 - [1 - \Phi(\gamma)]^n}{\Phi(\gamma)} & \text{for } t < \gamma, \\ [1 - \Phi(\gamma)]^{n-1} & \text{for } t > \gamma. \end{cases}$$
(30)

1 Eq. (29) provides an expression for the probability of disease given the mean parental score c.

Finally, we can integrate over all c in order to obtain the probability of disease in the population. Recalling that  $c \sim N(0, r_{ps}^2/2)$  and denoting its density as f(c), and again after some algebra,

$$P(\text{disease}) = \int_{-\infty}^{\infty} P(\text{disease} | c) f(c) dc$$
$$= \int_{-\infty}^{\infty} \phi(u) \left[ \int_{-\infty}^{\infty} \Psi(u, t) \zeta(u, t) dt \right] du, \tag{31}$$

where we defined

$$\beta = \sqrt{2z_q} - u,$$

$$\zeta(u,t) = \phi(t) \left[ 1 - \Phi\left(\frac{z_K - (u+t)r_{\rm ps}/\sqrt{2}}{\sqrt{1 - r_{\rm ps}^2}}\right) \right], \text{ and}$$

$$\psi(u,t) = \begin{cases} \frac{1 - [1 - \Phi(\beta)]^n}{\Phi(\beta)} & \text{for } t < \beta, \\ [1 - \Phi(\beta)]^{n-1} & \text{for } t > \beta. \end{cases}$$
(32)

2 Eq. (31) is our final expression for the probability of an embryo selected randomly among non-high-risk embryos to3 be affected.

# 1 6 The relative risk reduction

2 Given the prevalence K and the probability of the selected embryo to be affected P(disease), we define the relative risk

3 reduction (RRR) as

$$RRR = \frac{K - P(\text{disease})}{K} = 1 - \frac{P(\text{disease})}{K}.$$
(33)

For example, if a disease has prevalence of 5% and the selected embryo has probability 3% to be affected, the relative
risk reduction is 40%. We use the relative risk reduction defined in Eq. (33) in the main text, where *P*(disease) is given
by Eq. (20) for the *lowest-risk prioritization* strategy, and by Eq. (31) for the *high-risk exclusion* strategy. We solve
the integrals in these equations numerically in R (see Section 11).

#### 8 6.1 The relative risk reduction conditional on the mean parental score

In Sections 4 and 5 we computed the probability of disease conditional on the mean parental score c for the two selection strategies (Eqs. (23) and (29)). Let us denote this probability as  $P_s$ (disease  $|c\rangle$ ). To compute the relative risk reduction in these cases, we also need the "baseline" risk, i.e., the probability of disease when a random embryo is selected (as in natural procreation). We denote this probability  $P_r$ (disease  $|c\rangle$ ). Let us write the liability of a random embryo as

$$y = s + e$$
  
=  $x + c + e$   
=  $\tilde{x} + c$ , (34)

where we defined  $\tilde{x} = x + e$ . Var  $(\tilde{x}) = \text{Var}(x) + \text{Var}(e) = r_{\text{ps}}^2/2 + 1 - r_{\text{ps}}^2 = 1 - r_{\text{ps}}^2/2$ , and thus,  $\tilde{x} \sim N(0, 1 - r_{\text{ps}}^2/2)$ . The conditional probability of disease is

$$P_{r}(\text{disease} | c) = P(y > z_{K} | c)$$

$$= P(\tilde{x} + c > z_{K})$$

$$= P(\tilde{x} > z_{K} - c)$$

$$= 1 - \Phi\left(\frac{z_{K} - c}{\sqrt{1 - r_{\text{ps}}^{2}/2}}\right).$$
(35)

1 The relative risk reduction conditional on the mean parental score can be computed as

$$RRR = \frac{P_r(\text{disease} | c) - P_s(\text{disease} | c)}{P_r(\text{disease} | c)} = 1 - \frac{P_s(\text{disease} | c)}{P_r(\text{disease} | c)}.$$
(36)

2 The absolute risk reduction is defined as  $P_r(\text{disease} | c) - P_s(\text{disease} | c)$ .

### 3 7 The risk reduction conditional on family history

#### 4 7.1 Model

5 Let us rewrite our model for the liability as

$$y = s + w + \varepsilon. \tag{37}$$

6 Here, *w* represents all genetic factors not included in the score. It would be important to keep track of this component 7 as it is also inherited, and hence, information on the disease status in the family will be informative on *w* for both 8 parents and children. As in Section 2, we assume *s*, *w*, and  $\varepsilon$  are independent,  $s \sim N(0, r_{ps}^2)$ ,  $\varepsilon \sim N(0, 1 - h^2)$ , and thus 9  $w \sim N(0, h^2 - r_{ps}^2)$ .

We derive the risk reductions in two main steps. First, we assume that the values of *s* and *w* are known for each parent, and compute the risk of the embryo under each selection strategy (*lowest-risk prioritization*, *high-risk exclusion*, or random selection). Then, we derive the posterior distribution of the parental genetic components given any specific configuration of family history, and integrate over these components to obtain the final risk estimate.

#### 14 7.2 The risk of the selected embryo given its score

- 15 Denote the maternal score as  $s_m$  and the paternal score as  $s_f$ , and similarly for  $w_m$  and  $w_f$  and assume they are given.
- 16 (Also denote  $g_m = s_m + w + m$  and  $g_f = s_f + w_f$ .) As we explained in Section 3.1, at any child *i* the distribution of the
- 17 score s is

$$s_i \sim N\left(\frac{s_m + s_f}{2}, \frac{r_{\rm ps}^2}{2}\right) \text{ or } s_i = c + x_i,$$
(38)

18 where  $c = (s_m + s_f)/2$  and  $x_i \sim N(0, r_{ps}^2/2)$ . Similarly, the distribution of the non-score genetic component is

$$w_i \sim N\left(\frac{w_m + w_f}{2}, \frac{h^2 - r_{\rm ps}^2}{2}\right) \text{ or } w_i = \frac{w_m + w_f}{2} + v_i,$$
(39)

1 where  $v_i \sim N(0, (h^2 - r_{\rm ps}^2)/2)$ .

Given the parental genetic components, we can write the liability of each embryo as, for i = 1, ..., n,

$$y_i = \frac{s_m + s_f}{2} + x_i + \frac{w_m + w_f}{2} + v_i + \varepsilon_i,$$
(40)

where  $\varepsilon_i \sim N(0, 1 - h^2)$ . All the three random variables in the above equation  $(x_i, v_i, \text{ and } \varepsilon_i)$  are independent, and  $x_i$  and  $v_i$  are independent across embryos. (It is not necessary to specify whether the  $\varepsilon_i$  are independent.) Denote the event that embryo *i* is affected as  $D_i$ , and condition on the value of  $x_i$  for that embryo. The probability of disease is

$$P(D_{i}|s_{m}, w_{m}, s_{f}, w_{f}, x_{i}) = P(y_{i} > z_{K})$$

$$= P\left(\frac{s_{m} + s_{f}}{2} + x_{i} + \frac{w_{m} + w_{f}}{2} + v_{i} + \varepsilon_{i} > z_{K}\right)$$

$$= P\left(v_{i} + \varepsilon_{i} > z_{K} - \frac{s_{m} + s_{f}}{2} - \frac{w_{m} + w_{f}}{2} - x_{i}\right)$$

$$= 1 - \Phi\left(\frac{z_{K} - \frac{s_{m} + s_{f}}{2} - \frac{w_{m} + w_{f}}{2} - x_{i}}{\sqrt{1 - h^{2}/2 - r_{ps}^{2}/2}}\right).$$
(41)

3 The last line holds because  $\operatorname{Var}(v_i + \varepsilon_i) = (h^2 - r_{\rm ps}^2)/2 + (1 - h^2) = 1 - h^2/2 - r_{\rm ps}^2/2$ .

We denote henceforth  $D_s$  as the event that the selected embryo is affected. In the next three subsections, we integrate the probability of the disease over all  $x_i$ , where the distribution of  $x_i$  will vary depending on the selection strategy. This will give us the disease risk given the parental genetic components.

#### 7 7.3 Selecting the lowest-risk embryo

8 Denote by  $x_{i^*}$  is the embryo-specific component of the embryo with the lowest such component, where these com-9 ponents are distributed across embryos as  $x_i \sim N(0, r_{ps}^2/2)$ . We can use the theory of order statistics, as in previous 10 sections, to compute the density of  $x_{i^*}$ .

$$f(x_{i^*}) = \frac{n}{r_{\rm ps}/\sqrt{2}} \phi\left(\frac{x_{i^*}}{r_{\rm ps}/\sqrt{2}}\right) \left[1 - \Phi\left(\frac{x_{i^*}}{r_{\rm ps}/\sqrt{2}}\right)\right]^{n-1}.$$
(42)

Eq. (41) can now be integrated over all  $x_{i*}$ . After changing variables  $t = x_{i*}/(r_{ps}/\sqrt{2})$ , we obtain

$$P\left(D_{s} \mid s_{m}, w_{m}, s_{f}, w_{f}\right) = \int_{-\infty}^{\infty} n\phi(t) \left[1 - \Phi(t)\right]^{n-1} \left[1 - \Phi\left(\frac{z_{K} - \frac{s_{m} + s_{f}}{2} - \frac{w_{m} + w_{f}}{2} - tr_{\text{ps}}/\sqrt{2}}{\sqrt{1 - h^{2}/2 - r_{\text{ps}}^{2}/2}}\right)\right] dt$$
$$= \int_{-\infty}^{\infty} n\phi(t) \left[1 - \Phi(t)\right]^{n-1} \left[1 - \Phi\left(\frac{z_{K} - \frac{g_{m} + g_{f}}{2} - tr_{\text{ps}}/\sqrt{2}}{\sqrt{1 - h^{2}/2 - r_{\text{ps}}^{2}/2}}\right)\right] dt.$$
(43)

11 Note that the final result depends only on  $g_m$  and  $g_f$ . Thus, Eq. (43) can be integrated over  $g_m$  and  $g_f$  (according to 1 their distribution given the family disease history) to provide the disease risk probability.

### 2 7.4 Excluding high-risk embryos

3 Here, the density of the score of the selected embryo is given by Eq. (28), which continues to hold, with  $c = (s_m + s_f)/2$ .

$$f(x_{s}) = \begin{cases} \frac{1}{\frac{r_{ps}/\sqrt{2}}{ps}} \phi\left(\frac{x_{s}}{r_{ps}/\sqrt{2}}\right)}{1 - \Phi\left(\frac{z_{q}r_{ps}-c}{r_{ps}/\sqrt{2}}\right)} \left[1 - \Phi\left(\frac{z_{q}r_{ps}-c}{r_{ps}/\sqrt{2}}\right)\right]^{n} & \text{for } x_{s} > z_{q}r_{ps} - c \\ \frac{1}{\frac{r_{ps}/\sqrt{2}}{ps}} \phi\left(\frac{x_{s}}{r_{ps}/\sqrt{2}}\right)}{\frac{\Phi\left(\frac{z_{q}r_{ps}-c}{r_{ps}/\sqrt{2}}\right)}{\Phi\left(\frac{z_{q}r_{ps}-c}{r_{ps}/\sqrt{2}}\right)} \left\{1 - \left[1 - \Phi\left(\frac{z_{q}r_{ps}-c}{r_{ps}/\sqrt{2}}\right)\right]^{n}\right\} & \text{for } x_{s} < z_{q}r_{ps} - c \end{cases}$$
(44)

4 Integrating over all  $x_s$ , following similar steps as in Section 5, we obtain, denoting by  $D_s$  the event that the selected

5 embryo is affected,

$$P(D_s|s_m, w_m, s_f, w_f) = \int_{-\infty}^{\infty} \eta(t)\xi(t)dt, \qquad (45)$$

where we defined

$$\begin{split} \gamma &= \sqrt{2} z_q - \frac{c}{r_{\rm ps}/\sqrt{2}}, \\ \xi(t) &= \phi(t) \left[ 1 - \Phi\left(\frac{z_K - tr_{\rm ps}/\sqrt{2} - \frac{s_m + s_f}{2} - \frac{w_m + w_f}{2}}{\sqrt{1 - h^2/2 - r_{\rm ps}^2/2}}\right) \right] \\ &= \phi(t) \left[ 1 - \Phi\left(\frac{z_K - tr_{\rm ps}/\sqrt{2} - \frac{g_m + g_f}{2}}{\sqrt{1 - h^2/2 - r_{\rm ps}^2/2}}\right) \right], \text{ and} \\ \eta(t) &= \begin{cases} \frac{1 - [1 - \Phi(\gamma)]^n}{\Phi(\gamma)} & \text{for } t < \gamma, \\ [1 - \Phi(\gamma)]^{n-1} & \text{for } t > \gamma. \end{cases} \end{split}$$
(46)

6 Here, Eq. (45) depends on  $c, g_m, g_f$ , and they must be integrated over to obtain the final disease probability.

#### 1 7.5 The baseline risk

To compute the relative risk reduction, we need the baseline risk, i.e., the risk when selecting a random embryo. Here we need not condition on  $x_s$ . Thus,

$$P(D_{s}|s_{m}, w_{m}, s_{f}, w_{f}) = P(y_{i} > z_{K})$$

$$= P\left(\frac{s_{m} + s_{f}}{2} + x_{i} + \frac{w_{m} + w_{f}}{2} + v_{i} + \varepsilon_{i} > z_{K}\right)$$

$$= P\left(x_{i} + v_{i} + \varepsilon_{i} > z_{K} - \frac{g_{m} + g_{f}}{2}\right)$$

$$= 1 - \Phi\left(\frac{z_{K} - \frac{g_{m} + g_{f}}{2}}{\sqrt{1 - h^{2}/2}}\right).$$
(47)

#### 2 7.6 The disease risk conditional on the parental disease status

3 In the subsections 7.3, 7.4, and 7.5, we computed the disease probability for the various strategies. For the baseline risk 4 and for *lowest-risk prioritization* strategy, the risk depended only on  $g_m$  and  $g_f$ . For the *high-risk exclusion* strategy, the 5 risk also depended on *c*. In this section, we compute the posterior probability of these genetic components conditional 6 on the disease status of the parents.

Denote by  $D_m$  the indicator variable that the mother is affected (i.e.,  $D_m = 1$  is the mother is affected and  $D_m = 0$  otherwise), and similarly define  $D_f$ . The risk of the selected embryo conditional on the parental disease status can be written as

$$P\left(D_{s} \mid D_{m}, D_{f}\right) = \iiint dg_{m} dg_{f} d_{c} P\left(D_{s} \mid g_{m}, g_{f}, c, D_{m}, D_{f}\right) f\left(g_{m}, g_{f}, c \mid D_{m}, D_{f}\right)$$
$$= \iiint dg_{m} dg_{f} dc P\left(D_{s} \mid g_{m}, g_{f}, c\right) f\left(c \mid g_{m}, g_{f}\right) f\left(g_{m}, g_{f} \mid D_{m}, D_{f}\right).$$
(48)

7 Eq. (48) consists of three terms. The first is  $P(D_s | g_m, g_f, c)$ , which was computed in the previous subsections for the 8 various selection strategies. Note that  $P(D_s | g_m, g_f, c, D_m, D_f) = P(D_s | g_m, g_f, c)$ , because, given the genetic compo-9 nents of the parents, their disease status does not provide additional information on the disease status of the children 10 (at least under a model where the environment is not shared). The second term is the density of *c*, which can be simi-11 larly written as  $f(c | g_m, g_f, D_m, D_f) = f(c | g_m, g_f)$ . The third term is the posterior distribution of  $g_m$  and  $g_f$  given the

- 12 parental disease status,  $f(g_m, g_f | D_m, D_f)$ . In the following, we derive the third term and then the second term.
- 1 Note that if  $P(D_s | g_m, g_f, c) = P(D_s | g_m, g_f)$ , as in the case of the baseline risk (Eq. 47) and the *lowest-risk*
- 2 prioritization (Eq. (43)), the risk of the selected embryo can be simplified by integrating over c,

$$P\left(D_s \mid D_m, D_f\right) = \iint dg_m dg_f P\left(D_s \mid g_m, g_f\right) f(g_m, g_f \mid D_m, D_f).$$

$$\tag{49}$$

#### 3 7.7 The distribution of the parental genetic factors given family history

- 4 First, it is reasonable to assume that given one parent's disease status, his/her genetic component is independent of the
- 5 spouse's disease status or genetic factors. Thus, the posterior distribution can be factored into

$$f(g_m, g_f | D_m, D_f) = f(g_m | D_m) f(g_f | D_f).$$
(50)

6 We next assume that the parents are equivalent, and thus, their posterior distributions are identical and we focus on a

7 single parent. To derive the posterior distribution  $f(g_m | D_m)$  we first need the prior,  $g_m \sim N(0, h^2)$ ,

$$f_{pr}(g_m) = \frac{1}{h}\phi\left(\frac{g_m}{h}\right).$$
(51)

Next, The likelihood that the mother is affected is

$$P(D_m = 1 | g_m) = P(y > z_K)$$
  
=  $P(g_m + \varepsilon > z_K)$   
=  $P(\varepsilon > z_K - g_m)$   
=  $1 - \Phi\left(\frac{z_K - g_m}{\sqrt{1 - h^2}}\right).$  (52)

8 Similarly,

$$P(D_m = 0 | g_m) = \Phi\left(\frac{z_K - g_m}{\sqrt{1 - h^2}}\right).$$
(53)

Using Bayes' theorem,

$$f(g_m | D_m = 1) = \frac{P(D_m = 1 | g_m) f_{pr}(g_m)}{P(D_m) = 1} = \frac{\left[1 - \Phi\left(\frac{z_K - g_m}{\sqrt{1 - h^2}}\right)\right] \frac{1}{h} \phi\left(\frac{g_m}{h}\right)}{K}.$$
(54)

Similarly,

$$f(g_m | D_m = 0) = \frac{P(D_m = 0 | g_m) f_{pr}(g_m)}{P(D_m) = 0}$$
$$= \frac{\Phi\left(\frac{z_K - g_m}{\sqrt{1 - h^2}}\right) \frac{1}{h} \phi\left(\frac{g_m}{h}\right)}{1 - K}.$$
(55)

9 We have thus specified the posterior distribution  $f(g_m, g_f | D_m, D_f)$ .

# 7.8 The distribution of the parental mean score given the parental genetic 2 factors

3 Let us now compute  $f(c | g_m, g_f)$ . To this end, we note that  $c, g_m$ , and  $g_f$  have a multivariate normal distribution,

$$f(c, g_m, g_f) \sim \text{MVN}\left(\begin{pmatrix} 0\\0\\0 \end{pmatrix}, \begin{pmatrix} \frac{r_{\text{ps}}^2}{2} & \frac{r_{\text{ps}}^2}{2} & \frac{r_{\text{ps}}^2}{2} \\ \frac{r_{\text{ps}}^2}{2} & h^2 & 0 \\ \frac{r_{\text{ps}}^2}{2} & 0 & h^2 \end{pmatrix}\right).$$
(56)

To explain the above equation, recall that  $\operatorname{Var}(c) = r_{ps}^2/2$  and  $\operatorname{Var}(g_m) = \operatorname{Var}(g_f) = h^2$ . Then,

$$\operatorname{Cov}(c,g_m) = \operatorname{Cov}\left(\frac{s_m + s_f}{2}, g_m\right) = \frac{1}{2}\operatorname{Cov}(s_m, g_m)$$
$$= \frac{1}{2}\operatorname{Cov}(s_m, s_m + w_m) = \frac{1}{2}\operatorname{Var}(s_m) = \frac{r_{\text{ps}}^2}{2}.$$
(57)

- 4 A similar result holds for the paternal genetic component. To compute the density of c given  $g_m$  and  $g_f$ , we use standard
- 5 theory for multivariate normal variables (as in Section 3.1). We can write

$$c \mid g_m, g_f \sim N(\mu, \sigma^2), \tag{58}$$

where

$$\mu = \Sigma_{12} \Sigma_{22}^{-1} \begin{pmatrix} g_m \\ g_f \end{pmatrix},$$
  
$$\sigma^2 = \Sigma_{11} - \Sigma_{12} \Sigma_{22}^{-1} \Sigma_{21}$$
(59)

/

6 and

$$\Sigma_{11} = \frac{r_{\rm ps}^2}{2}, \Sigma_{12} = \begin{pmatrix} \frac{r_{\rm ps}^2}{2} & \frac{r_{\rm ps}^2}{2} \end{pmatrix}, \Sigma_{21} = \begin{pmatrix} \frac{r_{\rm ps}^2}{2} \\ \frac{r_{\rm ps}^2}{2} \end{pmatrix}, \Sigma_{22} = \begin{pmatrix} h^2 & 0 \\ 0 & h^2 \end{pmatrix}.$$
(60)

/ \

Carrying out the matrix calculations, we obtain

$$\mu = \frac{r_{\rm ps}^2}{h^2} \left(\frac{g_m + g_f}{2}\right),$$
  
$$\sigma^2 = \frac{r_{\rm ps}^2}{2h^2} (h^2 - r_{\rm ps}^2).$$
 (61)

1 We have thus specified  $f(c | g_m, g_f)$ .

#### 2 7.9 Summary of the computation

3 In summary, for the *high-risk exclusion* strategy, the probability of disease of the selected embyro given the parental 4 disease status is given by Eq. (48), with  $P(D_s|g_m,g_f,c)$  given in Eq. (45) and  $f(c|g_m,g_f)$  in Eq. (58). The con-5 ditional probability of disease for the *lowest-risk prioritization* strategy and for random selection (the baseline risk) 6 is given by Eq. (49), with  $P(D_s|g_m,g_f)$  given in Eqs. (43) and Eq. (47), respectively. For all selection strategies, 7  $f(g_m,g_f|D_m,D_f)$  is given by Eqs. (50), (54), and (55), depending the particular family history.

8 Numerically, computing the baseline disease risk requires two integrals (over  $g_m$  and  $g_f$ ). Computing the risk for 9 the *lowest-risk prioritization* strategy requires three integrals (over  $g_m$ ,  $g_f$ , and t). Computing the risk for the *high-risk* 10 *exclusion* strategy requires four integrals (over  $g_m$ ,  $g_f$ , c, and t).

# 11 8 Comparison to previous work

12 In the "gwern" blog (https://www.gwern.net/Embryo-selection), the utility of embryo selection for 13 traits and/or diseases was investigated. For disease risk, a similar model to ours was studied, based on the liability threshold model. However, the model assumed that given the polygenic score, the distribution of the remaining contribution to the liability has unit variance, instead of  $1 - r_{ps}^2$  (the function liabilityThresholdValue therein). Moreover, only numerical results were provided, the *high-risk exclusion* strategy was not considered, and there was no analysis of the case when the parental scores or disease status are known. Treff et al. (Treff et al., 2019) also employed the liability threshold model to evaluate embryo selection for disease risk. However, they did not consider the *high-risk exclusion* strategy, and did not compute analytically the risk reduction. They only provided simulation results for the case when a parent is affected based on an approximate model.

# 7 9 Simulations

To simulate the outcomes of embryo selection, we used the representation  $s_i = x_i + c$ , where  $(x_1, \ldots, x_n)$  are independent 8 normals with zero means and variance  $r_{ps}^2/2$ , and  $c \sim N(0, r_{ps}^2/2)$  is shared across all embryos. Thus, for each "family", 9 we first drew *c*, then drew *n* independent normals  $(x_1, \ldots, x_n)$ , and then computed the score of embryo *i* as  $s_i = x_i + c$ , 10 for i = 1, ..., n. The score of the selected embryo was the lowest among the *n* embryos in the *lowest-risk prioritization* 11 strategy. For the high-risk exclusion strategy, we selected the first embryo with score  $s < z_q r_{ps}$ . If no such embryo 12 existed, we selected the first embryo. We then drew the residual of the liability as  $e \sim N(0, 1 - r_{ps}^2)$ , and computed the 13 liability as  $s^* + e$ , where  $s^*$  is the score of the selected embryo. If the liability exceeded the threshold  $z_K$ , we designated 14 the embryo as affected. We repeated over  $10^6$  sets of embryos, and computed the disease probability as the fraction of 15 "families" in which the selected embryo was affected. We compute the relative risk reduction using Eq. (33). 16

For given parental risk scores, we computed *c* as  $c = (s_m + s_f)/2$ . We specified the maternal score as a percentile  $p_m$ , such that the score itself was  $s_m = z_{p_m} r_{ps}$ , where  $z_{p_m}$  is the  $p_m$  percentile of the standard normal distribution. We similarly specified the paternal score. The remaining calculations were as above. For the baseline risk, we used the same data, with the first embryo in each family.

21 When conditioning on the parental disease status, we first drew the independent parental components:  $s_m$  and  $s_f$ as normal variables with zero mean and variance  $r_{ps}^2$ ;  $w_m$  and  $w_f$  with variance  $h^2 - r_{ps}^2$ ; and  $\varepsilon_m$  and  $\varepsilon_f$  with variance 22  $1-h^2$ . We computed the maternal liability as  $y_m = s_m + w_m + \varepsilon_m$ , and designated the mother as affected if  $y_m > z_K$ . 23 We similarly designated the paternal disease status. We then drew the score of each embyro as  $s_i = c + x_i$ , where 24  $c = (s_m + s_f)/2$  (using the already drawn parental scores) and  $x_i \sim N(0, r_{ps}^2/2)$ , for i = 1, ..., n, are independent across 25 embryos. We selected one embryo based on the selection strategy, as in the previous paragraph. If  $s^*$  is the score 26 of the selected embryo, we computed the liability of the selected embryo as  $s^* + (w_m + w_f)/2 + v + \varepsilon$ , where  $v \sim$ 27  $N(0, (h^2 - r_{ps}^2)/2)$  and  $\varepsilon \sim N(1 - h^2)$ . We designated the embryo as affected if its liability exceeded  $z_K$ . We tallied the 28

29 proportion of affected embryos for each number of affected parents (0,1,2). To compute the baseline risk, we again 1 used the first embryo in each family.

# 2 **10** Limitations of the model

Our model has a number of limitations. First, our results relies on several modeling assumptions. (1) We assumed 3 an infinitesimal genetic architecture for the disease, which will not be appropriate for oligogenic diseases or when 4 5 screening the embryos for variants of very large effect. (2) Our model also assumes no assortative mating, which seems reasonable given that for genetic disease risk, correlation between parents is weak (Rawlik et al., 2019), and given that 6 our previous study of traits showed no difference in the results between real and random couples (Karavani et al., 7 2019). (3) When conditioning on the parental disease status, we assumed independence between the environmental 8 9 component of the child and either genetic or environmental factors influencing the disease status of the parents. Familyspecific environmental factors were shown to be insignificant for complex diseases (Wang et al., 2017). The influence 10 of parental genetic factors on the child's environment is discussed in the next paragraph. Both of these influences, to 11 the extent that they are significant, are expected to reduce the degree of risk reduction. 12

Second, we assumed that the proportion of variance (on the liability scale) explained by the score is  $r_{ps}^2$ , but we 13 did not specify how to estimate it. Typically,  $r_{ps}^2$  is computed and reported by large GWASs based on an evaluation of 14 15 the score in a test set. However, the variance that will be explained by the score in other cohorts, and particularly, in other populations, can be substantially lower (Martin et al., 2019). Relatedly, the variance explained by the score, as 16 estimated in samples of unrelated individuals, is inflated due to population stratification, assortative mating, and indirect 17 18 parental effects ("genetic nurture") (Kong et al., 2018; Young et al., 2019; Morris et al., 2020; Mostafavi et al., 2020), where the latter refers to trait-modifying environmental effects induced by the parents based on their genotypes. These 19 effects do not contribute to prediction accuracy when comparing polygenic scores between siblings (as when screening 2021 IVF embryos), and thus, the variance explained by polygenic scores in this setting can be substantially reduced, in particular for cognitive traits. However, recent empirical work on within-family disease risk prediction showed that the 22 reduction in accuracy is at most modest (Lello et al., 2020). 23

Third, we did not model the process of IVF and the possible reasons for loss of embryos. Rather, we assumed that n viable embryos are available that would have led to live birth if implanted. The original number of fertilized oocytes would typically be greater than n (see, e.g., (Branwen) for more detailed modeling). Similarly, we did not model the age-dependence of the number of embryos; again, we rather assume n viable embryos are available.

Fourth, the residual *e* in Eq. (2) (y = s + e) has a complex pattern of correlation between siblings. As noted in Sec-

tion 3, *e* has contribution from both genetic and environmental factors. While the genetic covariance between siblings is straightforward to model (as in Section 3), the proportion of variance in liability explained by shared environment needs to be estimated and can be large (Lakhani et al., 2019). Further, embryos from the same IVF cycle (when only one is actually implanted) would have experienced the same early developmental environment, and are thus expected to share even more environmental factors, similarly to twins. In the current work, the correlation between the residuals across embryos does not enter our derivations. However, care must be taken in any future attempt to compare the predicted phenotypic outcomes across the embryos.

Finally, in this work, we modeled various scenarios for the ascertainment of the parents: either randomly, or based on their scores, or based on their disease status. In future work, it will be interesting to model other settings of family history, such as the presence of an affected child. Further, it is likely that parents would attempt to screen the embryos for more than one disease (Treff et al., 2020). A related important question for future studies is to what extent selecting for a lower risk of one disease (or a set of diseases) would increase risk for diseases that were not included in the screen.

# 13 **11** Code availability

14 The R code we used to compute the relative risk reduction for the *lowest-risk prioritization* strategy (for randomly

15 ascertained parents) is as follows.

```
library(MASS)
risk_reduction_lowest = function(r,K,n)
{
    zk = qnorm(K, lower.tail=F)
    integrand_lowest = function(t)
        return(dnorm(t)*pnorm((zk-t*sqrt(1-r^2/2)) / (r/sqrt(2)), lower.tail=F)^n)
    risk = integrate(integrand_lowest,-Inf,Inf)$value
    return((K-risk)/K)
}
```

16 The R code we used to compute the relative risk reduction for the *high-risk exclusion* (for randomly ascertained 17 parents) strategy is as follows.

```
risk_reduction_exclude = function(r,K,q,n)
```

```
{
 zk = qnorm(K, lower.tail=F)
 zq = qnorm(q, lower.tail=F)
 integrand_t = function(t,u)
   return(dnorm(t)*pnorm((zk-r/sqrt(2)*(u+t))/sqrt(1-r^2),lower.tail=F))
 integrand_u = function(us)
  {
   y = numeric(length(us))
    for (i in seq_along(us))
    {
     u = us[i]
     beta = zq*sqrt(2)-u
     internal_int1 = integrate(integrand_t,-Inf,beta,u)$value
     denom1 = pnorm(beta)
     if (denom1==0) {denom1=1e-300} # Avoid dividing by zero
     numer1 = 1-pnorm(beta,lower.tail=F)^n
     internal_int2 = integrate(integrand_t, beta, Inf, u) $value
     prefactor2 = pnorm(beta,lower.tail=F)^(n-1)
     y[i] = dnorm(u) * (numerl/denoml*internal_int1 + prefactor2*internal_int2)
    }
   return(y)
  }
 risk = integrate(integrand_u,-Inf,Inf)$value
 return((K-risk)/K)
}
```

18 The remaining code used to generate the figures of the main text, e.g., when conditioning on the parental scores of 1 disease status, can be found at https://github.com/scarmi/embryo\_selection.

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#### References

Ala-Korpela, M., & Holmes, M. V. (2020). Polygenic risk scores and the prediction of common diseases. *International Journal of Epidemiology*, 49(1), 1–3. https://doi.org/10.1093/ije/dyz254

Bormann, C. L., Kanakasabapathy, M. K., Thirumalaraju, P., Gupta, R., Pooniwala, R., Kandula, H.,
Hariton, E., Souter, I., Dimitriadis, I., Ramirez, L. B., Curchoe, C. L., Swain, J., Boehnlein, L.
M., & Shafiee, H. (2020). Performance of a deep learning based neural network in the selection of human blastocysts for implantation. *ELife*, *9*, e55301. https://doi.org/10.7554/eLife.55301

Branwen, Gwern. (n.d.). Embryo selection for intelligence. https://www.gwern.net/Embryo-selection

- Callender, T., Emberton, M., Morris, S., Eeles, R., Kote-Jarai, Z., Pharoah, P. D. P., & Pashayan, N. (2019). Polygenic risk-tailored screening for prostate cancer: A benefit-harm and cost-effectiveness modelling study. *PLoS Medicine*, *16*(12), e1002998. https://doi.org/10.1371/journal.pmed.1002998
- Chatterjee, N., Shi, J., & García-Closas, M. (2016). Developing and evaluating polygenic risk prediction models for stratified disease prevention. *Nature Reviews. Genetics*, 17(7), 392–406. https://doi.org/10.1038/nrg.2016.27
- Cunningham, J., Goldsmith, L., & Skirton, H. (2015). The evidence base regarding the experiences of and attitudes to preimplantation genetic diagnosis in prospective parents. *Midwifery*, *31*(2), 288–296. https://doi.org/10.1016/j.midw.2014.09.010
- Dai, J., Lv, J., Zhu, M., Wang, Y., Qin, N., Ma, H., He, Y.-Q., Zhang, R., Tan, W., Fan, J., Wang, T.,
  Zheng, H., Sun, Q., Wang, L., Huang, M., Ge, Z., Yu, C., Guo, Y., Wang, T.-M., ... Shen, H.
  (2019). Risk loci identification and polygenic risk score in prediction of lung cancer: a large-scale prospective cohort study in Chinese. *The Lancet. Respiratory Medicine*, *7*(10), 881–891. https://doi.org/10.1016/S2213-2600(19)30144-4

Dempster, E. R., & Lerner, I. M. (1950). Heritability of Threshold Characters. Genetics, 35(2), 212-236.

- Do, C. B., Hinds, D. A., Francke, U., & Eriksson, N. (2012). Comparison of family history and SNPs for predicting risk of complex disease. *PLoS Genetics*, 8(10), e1002973. https://doi.org/10.1371/journal.pgen.1002973
- Dudbridge, F. (2013). Power and predictive accuracy of polygenic risk scores. *PLoS Genetics*, *9*(3), e1003348. https://doi.org/10.1371/journal.pgen.1003348
- Duncan, L., Shen, H., Gelaye, B., Meijsen, J., Ressler, K., Feldman, M., Peterson, R., & Domingue, B. (2019). Analysis of polygenic risk score usage and performance in diverse human populations. *Nature Communications*, 10(1), 3328. https://doi.org/10.1038/s41467-019-11112-0
- Falconer, D. S. (1967). The inheritance of liability to diseases with variable age of onset, with particular reference to diabetes mellitus. *Annals of Human Genetics*, 31(1), 1–20. https://doi.org/10.1111/j.1469-1809.1967.tb01249.x
- Geiss, L. S., Wang, J., Cheng, Y. J., Thompson, T. J., Barker, L., Li, Y., Albright, A. L., & Gregg, E. W. (2014). Prevalence and incidence trends for diagnosed diabetes among adults aged 20 to 79 years, United States, 1980-2012. *JAMA*, *312*(12), 1218–1226. https://doi.org/10.1001/jama.2014.11494
- Gibson, G. (2019). On the utilization of polygenic risk scores for therapeutic targeting. *PLoS Genetics*, *15*(4), e1008060. https://doi.org/10.1371/journal.pgen.1008060
- Gordis, L. (2014). Epidemiology.
- Hadar, L., & Sood, S. (2014). When knowledge is demotivating: subjective knowledge and choice overload. *Psychological Science*, 25(9), 1739–1747. https://doi.org/10.1177/0956797614539165
- Hayeck, T. J., Loh, P.-R., Pollack, S., Gusev, A., Patterson, N., Zaitlen, N. A., & Price, A. L. (2017).
  Mixed Model Association with Family-Biased Case-Control Ascertainment. *American Journal of Human Genetics*, 100(1), 31–39. https://doi.org/10.1016/j.ajhg.2016.11.015
- Holland, D., Frei, O., Desikan, R., Fan, C.-C., Shadrin, A. A., Smeland, O. B., Sundar, V. S., Thompson,
  P., Andreassen, O. A., & Dale, A. M. (2020). Beyond SNP heritability: Polygenicity and
  discoverability of phenotypes estimated with a univariate Gaussian mixture model. *PLoS Genetics*, 16(5), e1008612. https://doi.org/10.1371/journal.pgen.1008612

- Hujoel, M. L. A., Gazal, S., Loh, P.-R., Patterson, N., & Price, A. L. (2020). Liability threshold modeling of case-control status and family history of disease increases association power. *Nature Genetics*, 52(5), 541–547. https://doi.org/10.1038/s41588-020-0613-6
- Karavani, E., Zuk, O., Zeevi, D., Barzilai, N., Stefanis, N. C., Hatzimanolis, A., Smyrnis, N.,
  Avramopoulos, D., Kruglyak, L., Atzmon, G., Lam, M., Lencz, T., & Carmi, S. (2019). Screening
  Human Embryos for Polygenic Traits Has Limited Utility. *Cell*, *179*(6), 1424-1435.e8.
  https://doi.org/10.1016/j.cell.2019.10.033
- Khera, A. V., Chaffin, M., Aragam, K. G., Haas, M. E., Roselli, C., Choi, S. H., Natarajan, P., Lander, E. S., Lubitz, S. A., Ellinor, P. T., & Kathiresan, S. (2018). Genome-wide polygenic scores for common diseases identify individuals with risk equivalent to monogenic mutations. *Nature Genetics*, 50(9), 1219–1224. https://doi.org/10.1038/s41588-018-0183-z
- Kong, A., Thorleifsson, G., Frigge, M. L., Vilhjalmsson, B. J., Young, A. I., Thorgeirsson, T. E., Benonisdottir, S., Oddsson, A., Halldorsson, B. V., Masson, G., Gudbjartsson, D. F., Helgason, A., Bjornsdottir, G., Thorsteinsdottir, U., & Stefansson, K. (2018). The nature of nurture: Effects of parental genotypes. *Science (New York, N.Y.)*, *359*(6374), 424–428. https://doi.org/10.1126/science.aan6877
- Lakhani, C. M., Tierney, B. T., Manrai, A. K., Yang, J., Visscher, P. M., & Patel, C. J. (2019).
  Repurposing large health insurance claims data to estimate genetic and environmental contributions in 560 phenotypes. *Nature Genetics*, *51*(2), 327–334.
  https://doi.org/10.1038/s41588-018-0313-7
- Lambert, S. A., Abraham, G., & Inouye, M. (2019). Towards clinical utility of polygenic risk scores. *Human Molecular Genetics*, 28(R2), R133–R142. https://doi.org/10.1093/hmg/ddz187
- Lázaro-Muñoz, G., Pereira, S., Carmi, S., & Lencz, T. (2020). Screening embryos for polygenic conditions and traits: ethical considerations for an emerging technology. *Genetics in Medicine: Official Journal of the American College of Medical Genetics*. https://doi.org/10.1038/s41436-020-01019-3

- Lee, S. H., Goddard, M. E., Wray, N. R., & Visscher, P. M. (2012). A better coefficient of determination for genetic profile analysis. *Genetic Epidemiology*, *36*(3), 214–224. https://doi.org/10.1002/gepi.21614
- Lee, S. H., Wray, N. R., Goddard, M. E., & Visscher, P. M. (2011). Estimating missing heritability for disease from genome-wide association studies. *American Journal of Human Genetics*, 88(3), 294–305. https://doi.org/10.1016/j.ajhg.2011.02.002
- Lello, L., Raben, T. G., & Hsu, S. D. H. (2020). Sibling validation of polygenic risk scores and complex trait prediction. *Scientific Reports*, *10*(1), 13190. https://doi.org/10.1038/s41598-020-69927-7
- Lombardo, P. A. (2018). The power of heredity and the relevance of eugenic history. *Genetics in Medicine*, 20(11), 1305–1311. https://doi.org/10.1038/s41436-018-0123-4
- Lynch, M., & Walsh, B. (1998). Genetics and analysis of quantitative traits. Sinauer.
- Mars, N., Koskela, J. T., Ripatti, P., Kiiskinen, T. T. J., Havulinna, A. S., Lindbohm, J. V., Ahola-Olli,
  A., Kurki, M., Karjalainen, J., Palta, P., FinnGen, Neale, B. M., Daly, M., Salomaa, V., Palotie,
  A., Widén, E., & Ripatti, S. (2020). Polygenic and clinical risk scores and their impact on age at onset and prediction of cardiometabolic diseases and common cancers. *Nature Medicine*, *26*(4), 549–557. https://doi.org/10.1038/s41591-020-0800-0
- Martin, A. R., Kanai, M., Kamatani, Y., Okada, Y., Neale, B. M., & Daly, M. J. (2019). Clinical use of current polygenic risk scores may exacerbate health disparities. *Nature Genetics*, 51(4), 584–591. https://doi.org/10.1038/s41588-019-0379-x
- Mavaddat, N., Michailidou, K., Dennis, J., Lush, M., Fachal, L., Lee, A., Tyrer, J. P., Chen, T.-H., Wang, Q., Bolla, M. K., Yang, X., Adank, M. A., Ahearn, T., Aittomäki, K., Allen, J., Andrulis, I. L., Anton-Culver, H., Antonenkova, N. N., Arndt, V., ... Easton, D. F. (2019). Polygenic Risk Scores for Prediction of Breast Cancer and Breast Cancer Subtypes. *American Journal of Human Genetics*, *104*(1), 21–34. https://doi.org/10.1016/j.ajhg.2018.11.002

- McCabe, L. L., & McCabe, E. R. B. (2011). Down syndrome: coercion and eugenics. Genetics in Medicine: Official Journal of the American College of Medical Genetics, 13(8), 708–710. https://doi.org/10.1097/GIM.0b013e318216db64
- Montag, M., Toth, B., & Strowitzki, T. (2013). New approaches to embryo selection. *Reproductive Biomedicine Online*, 27(5), 539–546. https://doi.org/10.1016/j.rbmo.2013.05.013
- Morris, T. T., Davies, N. M., Hemani, G., & Smith, G. D. (2020). Population phenomena inflate genetic associations of complex social traits. *Science Advances*, 6(16), eaay0328. https://doi.org/10.1126/sciadv.aay0328
- Mostafavi, H., Harpak, A., Agarwal, I., Conley, D., Pritchard, J. K., & Przeworski, M. (2020). Variable prediction accuracy of polygenic scores within an ancestry group. *ELife*, 9. https://doi.org/10.7554/eLife.48376
- Murray, G. K., Lin, T., Austin, J., McGrath, J. J., Hickie, I. B., & Wray, N. R. (2020). Could Polygenic Risk Scores Be Useful in Psychiatry?: A Review. JAMA Psychiatry. https://doi.org/10.1001/jamapsychiatry.2020.3042
- Perälä, J., Suvisaari, J., Saarni, S. I., Kuoppasalmi, K., Isometsä, E., Pirkola, S., Partonen, T., Tuulio-Henriksson, A., Hintikka, J., Kieseppä, T., Härkänen, T., Koskinen, S., & Lönnqvist, J. (2007).
  Lifetime prevalence of psychotic and bipolar I disorders in a general population. *Archives of General Psychiatry*, 64(1), 19–28. https://doi.org/10.1001/archpsyc.64.1.19
- Rawlik, K., Canela-Xandri, O., & Tenesa, A. (2019). Indirect assortative mating for human disease and longevity. *Heredity*, 123(2), 106–116. https://doi.org/10.1038/s41437-019-0185-3
- Rhenman, A., Berglund, L., Brodin, T., Olovsson, M., Milton, K., Hadziosmanovic, N., & Holte, J. (2015). Which set of embryo variables is most predictive for live birth? A prospective study in 6252 single embryo transfers to construct an embryo score for the ranking and selection of embryos. *Human Reproduction (Oxford, England)*, *30*(1), 28–36. https://doi.org/10.1093/humrep/deu295

- Schizophrenia Working Group of the Psychiatric Genomics, C., Ripke, S., Walters, J. T., & O'Donovan,
   M. C. (2020). Mapping genomic loci prioritises genes and implicates synaptic biology in
   schizophrenia. *MedRxiv*, 2020.09.12.20192922. https://doi.org/10.1101/2020.09.12.20192922
- Sharp, S. A., Rich, S. S., Wood, A. R., Jones, S. E., Beaumont, R. N., Harrison, J. W., Schneider, D. A., Locke, J. M., Tyrrell, J., Weedon, M. N., Hagopian, W. A., & Oram, R. A. (2019). Development and Standardization of an Improved Type 1 Diabetes Genetic Risk Score for Use in Newborn Screening and Incident Diagnosis. *Diabetes Care*, 42(2), 200–207. https://doi.org/10.2337/dc18-1785
- So, H.-C., Kwan, J. S. H., Cherny, S. S., & Sham, P. C. (2011). Risk prediction of complex diseases from family history and known susceptibility loci, with applications for cancer screening. *American Journal of Human Genetics*, 88(5), 548–565. https://doi.org/10.1016/j.ajhg.2011.04.001
- Sueoka, K. (2016). Preimplantation genetic diagnosis: an update on current technologies and ethical considerations. *Reproductive Medicine and Biology*, 15(2), 69–75. https://doi.org/10.1007/s12522-015-0224-6
- Sullivan, P. F., Kendler, K. S., & Neale, M. C. (2003). Schizophrenia as a complex trait: evidence from a meta-analysis of twin studies. *Archives of General Psychiatry*, 60(12), 1187–1192. https://doi.org/10.1001/archpsyc.60.12.1187
- Sunkara, S. K., Rittenberg, V., Raine-Fenning, N., Bhattacharya, S., Zamora, J., & Coomarasamy, A.
  (2011). Association between the number of eggs and live birth in IVF treatment: an analysis of 400 135 treatment cycles. *Human Reproduction (Oxford, England)*, 26(7), 1768–1774. https://doi.org/10.1093/humrep/der106
- Torkamani, A., Wineinger, N. E., & Topol, E. J. (2018). The personal and clinical utility of polygenic risk scores. *Nature Reviews. Genetics*, *19*(9), 581–590. https://doi.org/10.1038/s41576-018-0018-x
- Treff, N. R., Eccles, J., Lello, L., Bechor, E., Hsu, J., Plunkett, K., Zimmerman, R., Rana, B., Samoilenko, A., Hsu, S., & Tellier, L. C. A. M. (2019). Utility and First Clinical Application of

Screening Embryos for Polygenic Disease Risk Reduction. *Frontiers in Endocrinology*, *10*, 845. https://doi.org/10.3389/fendo.2019.00845

- Treff, N. R., Eccles, J., Marin, D., Messick, E., Lello, L., Gerber, J., Xu, J., & Tellier, L. C. A. M. (2020). Preimplantation Genetic Testing for Polygenic Disease Relative Risk Reduction: Evaluation of Genomic Index Performance in 11,883 Adult Sibling Pairs. *Genes*, 11(6), 648. https://doi.org/10.3390/genes11060648
- Treff, N. R., Zimmerman, R., Bechor, E., Hsu, J., Rana, B., Jensen, J., Li, J., Samoilenko, A., Mowrey,
  W., Van Alstine, J., Leondires, M., Miller, K., Paganetti, E., Lello, L., Avery, S., Hsu, S., &
  Melchior Tellier, L. C. A. (2019). Validation of concurrent preimplantation genetic testing for
  polygenic and monogenic disorders, structural rearrangements, and whole and segmental
  chromosome aneuploidy with a single universal platform. *European Journal of Medical Genetics*,
  62(8), 103647. https://doi.org/10.1016/j.ejmg.2019.04.004
- Visscher, P. M., & Wray, N. R. (2015). Concepts and Misconceptions about the Polygenic Additive Model Applied to Disease. *Human Heredity*, 80(4), 165–170. https://doi.org/10.1159/000446931
- Visscher, P. M., Wray, N. R., Zhang, Q., Sklar, P., McCarthy, M. I., Brown, M. A., & Yang, J. (2017). 10 Years of GWAS Discovery: Biology, Function, and Translation. *The American Journal of Human Genetics*, 101(1), 5–22. https://doi.org/10.1016/j.ajhg.2017.06.005
- Wald, N. J., & Old, R. (2019). The illusion of polygenic disease risk prediction. *Genetics in Medicine*, 21(8), 1705–1707. https://doi.org/10.1038/s41436-018-0418-5
- Wang, K., Gaitsch, H., Poon, H., Cox, N. J., & Rzhetsky, A. (2017). Classification of common human diseases derived from shared genetic and environmental determinants. *Nature Genetics*, 49(9), 1319–1325. https://doi.org/10.1038/ng.3931
- Watanabe, K., Stringer, S., Frei, O., Umićević Mirkov, M., de Leeuw, C., Polderman, T. J. C., van der Sluis, S., Andreassen, O. A., Neale, B. M., & Posthuma, D. (2019). A global overview of pleiotropy and genetic architecture in complex traits. *Nature Genetics*, 51(9), 1339–1348. https://doi.org/10.1038/s41588-019-0481-0

Weissbrod, O., Flint, J., & Rosset, S. (2018). Estimating SNP-Based Heritability and Genetic Correlation in Case-Control Studies Directly and with Summary Statistics. *American Journal of Human Genetics*, 103(1), 89–99. https://doi.org/10.1016/j.ajhg.2018.06.002

Wilkinson, J., Malpas, P., Hammarberg, K., Mahoney Tsigdinos, P., Lensen, S., Jackson, E., Harper, J., & Mol, B. W. (2019). Do à la carte menus serve infertility patients? The ethics and regulation of in vitro fertility add-ons. *Fertility and Sterility*, *112*(6), 973–977.
https://doi.org/10.1016/j.fertnstert.2019.09.028

- Wray, N. R., & Goddard, M. E. (2010). Multi-locus models of genetic risk of disease. *Genome Medicine*, 2(2), 10. https://doi.org/10.1186/gm131
- Wray, N. R., Lin, T., Austin, J., McGrath, J. J., Hickie, I. B., Murray, G. K., & Visscher, P. M. (2020). From Basic Science to Clinical Application of Polygenic Risk Scores: A Primer. JAMA Psychiatry. https://doi.org/10.1001/jamapsychiatry.2020.3049
- Wray, N. R., Yang, J., Hayes, B. J., Price, A. L., Goddard, M. E., & Visscher, P. M. (2013). Pitfalls of predicting complex traits from SNPs. *Nature Reviews. Genetics*, 14(7), 507–515. https://doi.org/10.1038/nrg3457
- Young, A. I., Benonisdottir, S., Przeworski, M., & Kong, A. (2019). Deconstructing the sources of genotype-phenotype associations in humans. *Science (New York, N.Y.)*, 365(6460), 1396–1400. https://doi.org/10.1126/science.aax3710
- Zhang, Q., Sidorenko, J., Couvy-Duchesne, B., Marioni, R. E., Wright, M. J., Goate, A. M., Marcora, E., Huang, K., Porter, T., Laws, S. M., Sachdev, P. S., Mather, K. A., Armstrong, N. J., Thalamuthu, A., Brodaty, H., Yengo, L., Yang, J., Wray, N. R., McRae, A. F., & Visscher, P. M. (2020). Risk prediction of late-onset Alzheimer's disease implies an oligogenic architecture. *Nature Communications*, *11*(1), 4799. https://doi.org/10.1038/s41467-020-18534-1
- Zhang, X., Rice, M., Tworoger, S. S., Rosner, B. A., Eliassen, A. H., Tamimi, R. M., Joshi, A. D.,Lindstrom, S., Qian, J., Colditz, G. A., Willett, W. C., Kraft, P., & Hankinson, S. E. (2018).Addition of a polygenic risk score, mammographic density, and endogenous hormones to existing

breast cancer risk prediction models: A nested case-control study. *PLoS Medicine*, *15*(9), e1002644. https://doi.org/10.1371/journal.pmed.1002644

Zhang, Y., Qi, G., Park, J.-H., & Chatterjee, N. (2018). Estimation of complex effect-size distributions using summary-level statistics from genome-wide association studies across 32 complex traits. *Nature Genetics*, 50(9), 1318–1326. https://doi.org/10.1038/s41588-018-0193-x

Zheng, J., Erzurumluoglu, A. M., Elsworth, B. L., Kemp, J. P., Howe, L., Haycock, P. C., Hemani, G., Tansey, K., Laurin, C., Early Genetics and Lifecourse Epidemiology (EAGLE) Eczema Consortium, Pourcain, B. S., Warrington, N. M., Finucane, H. K., Price, A. L., Bulik-Sullivan, B. K., Anttila, V., Paternoster, L., Gaunt, T. R., Evans, D. M., & Neale, B. M. (2017). LD Hub: a centralized database and web interface to perform LD score regression that maximizes the potential of summary level GWAS data for SNP heritability and genetic correlation analysis. *Bioinformatics (Oxford, England)*, *33*(2), 272–279. https://doi.org/10.1093/bioinformatics/btw613

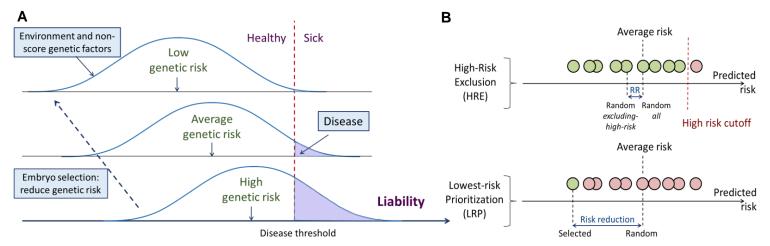


Figure 1. A schematic of the liability threshold model and polygenic embryo screening. (A) An illustration of the liability threshold model (LTM). Under the LTM, it is assumed that each disease has an underlying (unobserved) liability, and that an individual is affected if the total liability is above a threshold. The liability is composed of a genetic component and an environmental component, both assumed to be normally distributed in the population. For a given genetic risk (represented here by the polygenic risk score), the liability is the sum of that risk, plus a normally distributed residual component (environmental + genetic factors not captured by the PRS). Thus, for an individual with high genetic risk (bottom curve), even a modestly elevated (and thus, commonly-occurring) liabilityincreasing environment will lead to disease. For an individual with low genetic risk (top curve), only an extreme environment will push the liability beyond the disease threshold, making the disease less probable. Thus, disease risk reduction can be achieved with embryo screening by lowering the genetic risk of the implanted embryo. [Note that for the purpose of illustration, panel A displays three discrete levels of genetic risk, although in reality PRS is continuously distributed.] (B) An illustration of the embryo selection strategies considered in this report. In the figure, each embryo is shown as a filled circle, and embryos are sorted based on their predicted risk, i.e., their polygenic risk scores. Excluded embryos are shown in pink, and embryos that can be implanted in green. The risk reduction (RR) is indicated as the difference in risk between a randomly selected embryo (if no polygenic scoring was performed) and the embryo selected based on one of two strategies. In high-risk exclusion (HRE), the embryo selected for implantation is random, as long as its PRS is under a high-risk cutoff (usually the top few PRS percentiles). If all embryos are high-risk, a random embryo is selected. In lowest-risk prioritization (LRP), the embryo with the lowest PRS is selected for implantation. As we describe below, the LRP strategy yields much larger disease risk reductions.

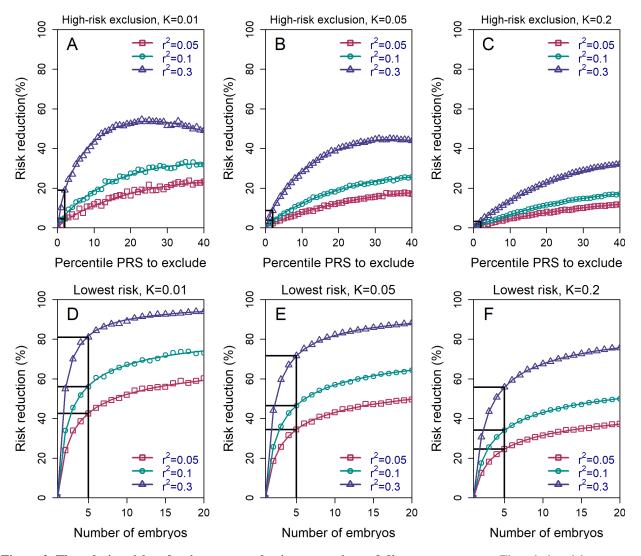


Figure 2. The relative risk reduction across selection strategies and disease parameters. The relative risk reduction (RRR) is defined as (K - P(disease))/K, where K is the disease prevalence, and P(disease) is the probability of the implanted embryo to become affected. The RRR is shown for the high-risk exclusion (HRE) strategy in the upper row (panels A-C), and for the lowest-risk prioritization (LRP) in the lower row (panels D-F). See Figure 1 for the definitions of the strategies. Results are shown for values of K = 1%, 5%, 20% in each panel, and within each panel, for variance explained by the PRS (on the liability scale)  $r_{ps}^2 = 5\%$ , 10%, 30% (legends). Symbols denote the results of Monte-Carlo simulations (Materials and Methods), where PRSs of embryos were drawn based on a multivariate normal distribution, assuming PRSs are standardized to have zero mean and variance  $r_{ps}^{2}$ , and accounting for the genetic similarity between siblings (Eq. (4) in *Materials and Methods*). In each simulated set of n sibling embryos (n = 5 for all simulations under HRE), one embryo was selected according to the selection strategy. The liability of the selected embryo was computed by adding a residual component (drawn from a normal distribution with zero mean and variance  $1 - r_{ps}^2$ ) to its standardized polygenic score. The embryo was considered affected if its liability exceeded  $z_K$ , the (upper) K-quantile of the standard normal distribution. We repeated the simulations over 10<sup>6</sup> sets of embryos and computed the disease risk. In each panel, curves correspond to theory: Eq. (31) in Materials and Methods for the HRE strategy, and Eq. (20) in Materials and Methods for the LRP strategy. Black straight lines correspond to the RRR achieved when excluding embryos at the top 2% of the PRS (for HRE, upper panels) or for selecting the lowest risk embryo out of n = 5 (for LRP, lower panels).

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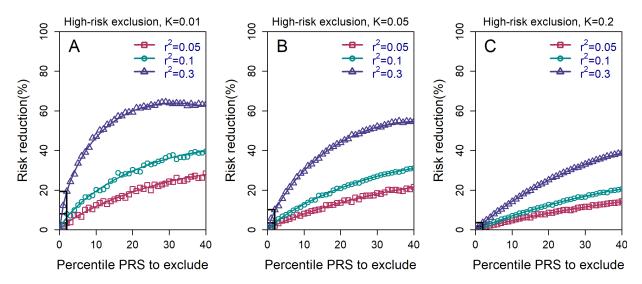


Figure 2 - Figure Supplement 1. The relative risk reduction for the *high-risk exclusion* strategy, with n = 10 available embryos. All details are exactly as in panels (A-C) in Figure 2 of the main text, except that we simulated n = 10 embryos.

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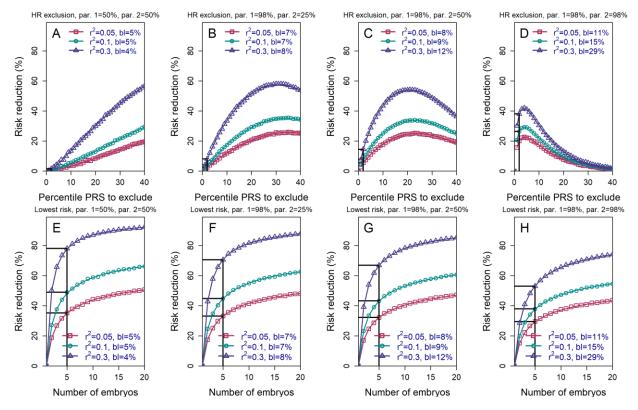


Figure 2 - Figure Supplement 2. The relative risk reduction when the polygenic risk scores of the parents are known. Panels (A)-(D) are for the high-risk exclusion (HRE) strategy, while panels (E)-(H) are for the lowest-risk prioritization (LRP) strategy. All details are as in Figure 2 of the main text, except the following. First, we fixed the prevalence at K = 5%. Second, in the simulations, we drew the PRS of each embryo as  $s_i = x_i + c$  (i = 1, ..., n), where  $x_i$  is an embryo-specific component (independent across embryos) and c is the shared component, also representing the mean parental PRS (Materials and Methods). This is so far as in Figure 2; however, here we assumed that c is given, equal to the average PRSs of the two parents. In each panel, we consider a different pair of PRSs for the parents. For example, in panels (A) and (E), both parents ("par. 1" and "par. 2") have PRS equal to the 50% percentile of the PRS distribution; in panels (B) and (F), one parent has PRS equal to the 98% percentile of the PRS distribution, while the other has PRS equal to the 25% percentile; and so on. Third, in the simulations, we computed the risk reduction (according to either strategy) relative to a baseline, obtained from the same sets of simulations, when we always selected the first embryo. The baseline risk is indicated in each legend as "bl". Note that the baseline risk depends on the variance explained by the PRS, because the parental PRSs are determined as percentiles of the population distribution of the score, which has variance  $r_{ps}^2$ . Finally, we computed the theoretical disease risk for the HRE strategy using Eq. (29) from *Materials and Methods*, the disease risk for the LRP strategy using Eq. (23), and the relative risk reduction (shown in curves) for both strategies using Eq. (36).

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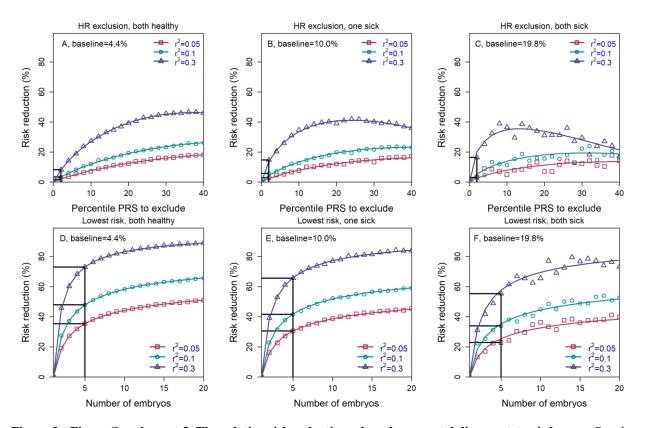
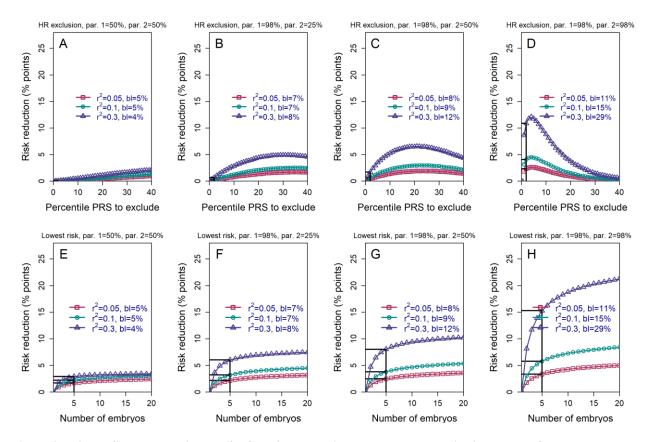


Figure 2 - Figure Supplement 3. The relative risk reduction when the parental disease status is known. Panels (A)-(C) are for the high-risk exclusion (HRE) strategy, while panels (D)-(F) are for the lowest-risk prioritization (LRP) strategy. The details are as in Figure 2 of the main text, except the following. First, we fixed the prevalence at K = 5% and the heritability to  $h^2 = 0.4$  (note that the heritability was not needed in previous figures). Second, in the simulations, we first drew the parental genetic components:  $s_m$  and  $w_m$  for the mother, and  $s_f$  and  $w_f$  for the father, where  $s_m \sim s_f \sim N(0, r_{ps}^2)$  are the polygenic scores and  $w_m \sim w_f \sim N(0, h^2 - r_{ps}^2)$  represent the non-score genetic factors (*Materials and Methods*). We drew the environmental component for each parent as  $\epsilon_m \sim \epsilon_f \sim$  $N(0,1-h^2)$  and computed the liability of each parent as  $s + w + \epsilon$ . If the liability of a parent exceeded  $z_k$  (the (1 - K)-quantile of the standard normal distribution), we designated that parent as affected. We then stratified the risk reduction results based on the number of affected parents: 0 (panels (A) and (D), 1 (panels (B) and (E)), and 2 (panels (C) and (F)). Note that as expected, the number of families in which both parents are affected is small, and thus, the results in panels (C) and (F) are noisier. For each set of parents, we drew the PRS of each embryo as  $s_i =$  $(s_m + s_f)/2 + x_i$  (i = 1, ..., n), where  $x_i \sim N(0, r_{ps}^2/2)$  is an embryo-specific component of the score (independent across embryos). We then selected one embryo from each family based on either selection strategy. We computed the liability of the selected embryo as  $s_i + (w_m + w_f)/2 + v_i + \epsilon_i$ , where  $v_i \sim N(0, (h^2 - r_{ps}^2)/2)$  is the embryo specific component of the non-score genetic factors, and  $\epsilon_i \sim N(0, 1 - h^2)$  is the environmental component of the embryo (Materials and Methods). The embryo was designated as affected or unaffected as described above for the parents. We computed the risk reduction (according to either strategy) relative to a baseline, obtained from the same sets of simulations when we always selected the first embryo. The baseline risk is indicated on top of each panel. We computed the theoretical relative risk reduction for the two strategies as summarized in Section 7.9 of the Materials and Methods.

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**Figure 2 - Figure Supplement 4. The** *absolute* **risk reduction when the polygenic risk scores of the parents are known.** All details are the same as in **Figure 2 - Figure Supplement 2**, except that the *absolute* (rather than the relative) risk reduction is shown. The absolute risk reduction is defined as the difference between the baseline disease risk (given the parental PRSs; legends) and the risk following either strategy of embryo selection. It is plotted as percentage points.